Longitudinal Auditory Pathophysiology Following Mild Blast

Induced Trauma

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Abstract

Blast-induced hearing difficulties affect thousands of veterans and civilians. The long-

term impact of even a mild blast exposure on the central auditory system is

hypothesized to contribute to lasting behavioral complaints associated with mild blast

traumatic brain injury (bTBI). Although recovery from mild blast has been studied

separately over brief or long time windows, few, if any, studies have investigated

recovery longitudinally over short-term and longer-term (months) time windows.

Specifically, many peripheral measures of auditory function either recover or exhibit

subclinical deficits, masking deficits in processing complex, real-world stimuli that may

recover differently. Thus, examining the acute time course and pattern of

neurophysiological impairment using appropriate stimuli is critical to better

understanding and intervention of bTBI-induced auditory system impairments. Here, we

compared auditory brainstem response, middle-latency auditory evoked potentials, and

envelope following responses. Stimuli were clicks, tone pips, amplitude modulated

tones in quiet and in noise, and speech-like stimuli (iterated rippled noise pitch

contours) in adult male rats subjected to mild blast and sham exposure over the course

of two months. We found that blast animals demonstrated drastic threshold increases

and auditory transmission deficits immediately after blast exposure, followed by

substantial recovery during the window of 7-14 days post-blast, though with some

deficits remaining even after two months. Challenging conditions and speech-like

stimuli can better elucidate mild bTBI-induced auditory deficit during this period. Our

results suggest multiphasic recovery and therefore potentially different time windows

for treatment, and deficits can be best observed using a small battery of sound stimuli.

New and Noteworthy

Few studies on blast-induced hearing deficits go beyond simple sounds or two times

post-exposure. Therefore, the recovery arc for potential therapies and real-world

listening is poorly understood. Evidence suggested multiple recovery phases over 2

months post-exposure. Hearing thresholds largely recovered within 14 days and partially

explained recovery. However, mid-latency responses, responses to AM in noise, and

speech-like pitch sweeps all exhibited extended changes, implying persistent central

auditory deficits and the importance of subclinical threshold shifts.

Introduction

Hearing loss stands out as one of the most commonly reported consequences following

blast injuries and can last for months or even years without significant external injury

(Cohen et al. 2002; Cave et al. 2007; Ritenour et al. 2008; Saunders et al. 2015).   Hearing

difficulties have been hypothesized to contribute to many behavioral complaints

associated with mild blast traumatic brain injury (Gallun et al. 2012b; Vander Werff 2012).

Most studies regarding blast-induced hearing loss have focused on damage in different

parts of the peripheral auditory system (PAS) (Kerr 1980; DePalma et al. 2005). However,

significant hearing difficulties can occur in the absence of peripheral diagnostic indicators

such as eardrum rupture or clinical threshold shifts (hearing loss >25 dB), indicating

potential disruptions further upstream (Remenschneider et al. 2014; Saunders et al. 2015;

Van Haesendonck et al. 2018). Increasing clinical (Berger et al. 1997; Cohen et al. 2002;

Cave et al. 2007; Ritenour et al. 2008; Lew et al. 2009; Gallun et al. 2012a) and laboratory

(Patterson and Hamernik 1997; Ewert et al. 2012; Cho et al. 2013b; Du et al. 2013; Masri

et al. 2018) evidence suggest that both peripheral and central auditory system (CAS) are

important blast-susceptible structures, where CAS includes the brainstem, midbrain,

thalamus, and cortex.

Subcortical CAS such as the auditory brainstem may be particularly vulnerable to blast

injury, including hemorrhages, intracranial deformation and blood-brain barrier (BBB)

permeability, glutamate excitotoxicity, elevated calcium, as well as elevated markers of

oxidative stress and neuroinflammation from near short-term (1-7 days) up to 2 weeks

(Knudsen and Øen 2003; Leung et al. 2008; Säljö et al. 2011; Cho et al. 2013a; Song et al.

2015; Walls et al. 2016). Functional changes, such as spontaneous hyperactivity in the

auditory brainstem (Luo et al. 2014a, 2014b), as well as structural changes in OHC loss

(Ewert et al. 2012), axonal integrity, white matter changes in the inferior colliculus (IC) and

auditory thalamus (Mao et al. 2012a), have been shown post-blast at various time points,

from 24 hours, 7 days up to 2-3 weeks. Most of these previous studies only assessed one

or two time points post-blast. Given the likelihood of multiple phases of primary and

secondary blast-induced damage and their corresponding anatomical and

electrophysiological changes, understanding the trajectory of post-blast recovery can

help to identify critical time points and to direct therapies that are matched to the

recovery mechanisms at those time points.

Clinical reports have suspected "hidden hearing loss" in blast-exposed veterans due to

deficits in suprathreshold auditory processing with minimal changes in auditory

thresholds (Gallun et al. 2012a; Saunders et al. 2015; Bressler et al. 2017). This occurs in

age-related or noise-induced sensorineural hearing loss and encompasses putative

changes resulting from hair cell or auditory nerve/synapse damage (Hickox and Liberman

2014; Plack et al. 2014; Bharadwaj et al. 2015; Kujawa and Liberman 2015; Viana et al.

2015; Liberman 2017). One consequence could be CAS adaptations in response to

peripheral deafferentation (Caspary et al. 2005, 2008; Wang et al. 2009), which has been

speculated to impair temporal processing (Walton 2010; Parthasarathy and Bartlett 2011,

2012; Rabang et al. 2012). Blast studies on human subjects often used speech and

complex temporally modulated stimuli to pin down "hidden" temporal processing losses

that were not apparent with simple audiological measures (Gallun et al. 2012b; Saunders

et al. 2015; Bressler et al. 2017; Kubli et al. 2018). However, blast studies in animals rarely

go beyond simple auditory stimuli such as clicks, tones and AM modulation in a quiet

background (Ewert et al. 2012; Race et al. 2017; Masri et al. 2018).

Using the same rat model as our previous publication (Race et al. 2017), our study

documents a detailed time course of blast-induced hearing loss recovery in the

subcortical CAS using auditory evoked potentials (AEPs). We specifically chose Iterated

Rippled Noise (IRN) with pitch sweep alongside Amplitude Modulation (AM) stimuli in

quiet and in modulated noise as temporally complex stimuli in assessing the processing

of temporal attributes. IRN was used in neurophysiological and behavioral studies in both

human (Krishnan et al. 2014, 2015; Peter et al. 2014; Wagner et al. 2017; Shearer et al.

2018) and animal models (Bendor and Wang 2005; Alsindi et al. 2018), creating "pseudo-

pitch" pitch contours with broadband carriers. IRN bypasses limitations posed by animal

frequency range as well as permitting easily adjustable pitch intelligibility by altering the

number of noise iterations and thus, the strength of temporal regularity and pitch salience

(Patterson et al. 1996).

Materials and Methods

Subject

Male Sprague-Dawley rats (3-4 months) were assigned into Sham group and Blast group

randomly. A total of 11 Sham animals and 13 blast animals were used in this study. All

animals were kept and raised in relatively quiet and standard laboratory animal housing

conditions. All protocols were approved by the Purdue Animals Care and Use Committee

(PACUC #1111000280).

Blast Exposure

Animals were anesthetized through intraperitoneal injection of a ketamine/xylazine

cocktail (80 mg/kg and 10 mg/kg, respectively). The absence of eye-blink and paw-

withdrawal reflexes was ensured prior to proceeding. Anesthetized animals were then

placed on a platform beneath an open-ended shock tube to be exposed to the blast

event, as described in our prior publications (Song et al. 2015; Walls et al. 2016; Race et

al. 2017).

For the Blast group, each rat's head was positioned beneath the open end of the shock

tube such that the dorsum of the skull was the incident surface exposed to a composite

blast (shock wave + blast wind). A custom plexiglass housing was temporarily placed over

the animal's torso for body protection to avoid cardiac or pulmonary effects of blast and

to simulate the protective effects of military body armor (Rafaels et al. 2011). The head

was fixed with a stereotaxic head frame with bite bar and ear bars (Kopf Instruments) to

prevent blast wind-induced head acceleration. The blast exposure exhibited a recorded

pressure profile with a rise to peak pressure within 0.3 msec, followed by overpressure

and underpressure periods as follows: side-on (static) 150 kPa maximum overpressure,

1.25 msec overpressure duration, and 20 kPa minimum underpressure; face on (dynamic)

160 kPa maximum overpressure, 1.75 msec overpressure duration, and 5 kPa minimum

underpressure. These conditions were the same as reported in our prior publications

(Song et al. 2015; Walls et al. 2016; Race et al. 2017). All but one blast animal survived the

exposure without displaying any external deficit during each animal's longitudinal follow-

up period.

Sham animals were placed equidistant from the blast source, but out of the path of the

shockwave, therefore only exposed to the blast noise. Tympanic membrane integrity was

verified for all Blast and Sham animals after injury using a surgical microscope.

(Insert Fig. 1 about here)

Auditory Evoked Potential Recordings

The animals underwent two-channel Auditory Evoked Potential (AEP) recordings at the

following time points: pre-exposure (baseline), 1 day, 4 days, 7 days, 10 days, 14 days, 1

month, and 2 months. While the animals were under 1.8-2% isoflurane anesthesia,

subdermal needle electrodes (Ambu) were inserted in the following locations (Fig. 1A):

Channel 1 positive electrode was placed along the midline of the head (mid-sagittal)

oriented Fz to Cz. Channel 2 positive electrode was positioned C3 to C4 along the

interaural line. The negative/inverting electrode (used with positive electrodes for both

channels 1 and 2) was placed under the mastoid of the right ear ipsilateral to the speaker.

A ground electrode was placed in the back of the animal. These configurations were

consistent with prior publications from our laboratory (Parthasarathy and Bartlett 2011,

2012; Parthasarathy et al. 2014; Lai and Bartlett 2015; Lai et al. 2017). Electrode

impedances were confirmed to be less than 1 kΩ using a low impedance amplifier (RA4LI,

TDT). After electrode placement, we subsequently sedated the animals by intramuscular

injection of 0.2-0.3 mg/kg dexmedetomidine (Dexdomitor). AEP recordings were

performed 10-15 min after removal from isoflurane to avoid anesthetic effects. The

animals could respond to pain and acoustic stimuli but tend sit calmly under

dexmedetomidine sedation, allowing about 3 hours of recording time.

Acoustic stimuli were presented free-field to the right ear (90^0 azimuth) of animals, with

directly in front of the animals' face as the reference for 0^0 azimuth, using a calibrated

speaker (Bowers and Wilkins) at a distance of 115 cm directly facing the right ear. The

measurements used in this study included auditory brainstem responses (ABRs), middle-

latency responses (MLRs), envelop-following responses (EFRs) using AM in noise stimuli,

and IRNs.

ABR and MLR

6 Sham animals and 10 Blast animals were used in ABR analysis. For ABR, rectangular

clicks (0.1 msec duration) and tone-pips (2 msec duration, 0.5 msec cos^2 rise-fall time)

with frequencies of 8 kHz and 16kHz were used. 8 kHz and 16 kHz were chosen based

on previous findings: with 6-16 kHz being the most sensitive hearing region of rats, 8 kHz

near the most sensitive region of normal rat audiogram (Parthasarathy et al. 2014) and

hearing of frequencies higher than 8 kHz being most vulnerable to blast injury (Race et

al. 2017). The sound levels of clicks and pips ranged from 90 to 10 dB peak SPL in 5-dB

steps. All stimuli were presented in alternating polarity at 26.6 per second with 1500

repetitions (750 at each polarity). A 20 msec acquisition window (0-20 msec) was used.

Data were processed with a 30 Hz high-pass (HP) filter and a 3000 Hz low-pass (LP) filter

prior to analysis. The ABR threshold was visually determined as the minimum sound level

that produced a distinct ABR waveform, with confirmation from two other researchers.

The ABR amplitudes of waves I and V from channel 2 were estimated as the differences

of each wave's amplitude, as seen in BioSigRP (TDT) and the baseline amplitude

(measured as an average of 2 msec waveform prior to the cochlear microphonic).

6 Sham animals and 8 Blast animals were used in MLR analysis. For MLR, similar

rectangular clicks and 8 kHz tone pips of alternating polarity as in ABR were used but

were presented at a slower rate (3.33/sec vs. 26.6/sec in ABRs) and with a recording

window of longer duration (100 msec vs. 20 msec in ABRs). This time window provides

enough time to capture the stimulus-evoked "middle-latency" neural responses from the

auditory midbrain, thalamus and cortex (Barth and Shi Di 1991; McGee et al. 1991; Di and

Barth 1992; McGee and Kraus 1996; Phillips et al. 2011; Šuta et al. 2011) alongside ABR.

Stimuli were presented at 80 dB sound pressure level (SPL) and 30 dB sensation level (SL,

30 dB above corresponding ABR thresholds), as determined in the previous ABR

recordings. 1500 repetitions were collected over an acquisition time window of 100 msec

to obtain an average response. Only one animal exhibited hearing threshold higher than

80 dB SPL at only one time point, for which MLR recording has been excluded for that

point.

Data were processed with HP (fc = 10 Hz) and LP (fc = 300 Hz) filters prior to analysis.

EFRs

EFRs were recorded during the same recording session following ABRs and MLRs using

the same electrode configurations with similar techniques to Lai and Bartlett (2018) and

Lai (Lai et al. 2017). The two channels were sensitive to a complementary range of

amplitude modulation frequencies (AMFs) (Parthasarathy and Bartlett 2012), with channel

1 (mid-sagittal) being more sensitive to higher AMFs (90-2048 Hz) while channel 2

(interaural) is more sensitive to lower AMFs (8-90 Hz). The AM stimuli used for EFRs were

sinusoidally amplitude-modulated (AM) sounds, with Gaussian noise, 8 kHz tone, or 16

kHz tone as carriers, and under 100% and 50% modulation depth with a stimulus duration

of 200 msec. The AMFs selected for this study are 10 Hz, 45 Hz, and 256 Hz, based on

the findings in Race at al. (Race et al. 2017), which found significant differences,

particularly at the lower AMFs. The acquisition window was 300 msec long, and each

response was an average of 200 repetitions. The stimuli were presented at 30 dB SL. For

animals that had a hearing threshold above 70 dB SPL, which usually happens on day 1

post-exposure, EFR was not collected at the time point due to the limitation of the

speaker and BiosigRP.

For AM in Noise stimuli, the same EFRs were used alongside a 71 Hz sinusoidally AM

masker of the same length and onset, with Gaussian noise as the carrier, similar to Lai

and Bartlett (Lai and Bartlett 2018). Noise AM maskers were presented at sound levels of

20dB SNR and 0SNR to the sound level of target AM. Prior to EFR amplitude analysis,

data were passed through an LP filter of 3000 Hz and a high-pass filter that was either

slightly below the AMF for AMFs <90 Hz, or 80 Hz for AMFs ≥ 90 Hz.

IRNs

For 6 Sham animals and 8 Blast animals, IRNs were recorded during the same recording

session following the previous stimuli using the same electrode configurations. The sound

level of presentation was 30 dB SL (above click hearing threshold). Data for animals with

a hearing threshold above 70 dB SPL were not collected at the time point.

IRN tone stimuli were created by sequential delay and add operations. Time-varying pitch

curves were created by applying polynomial equations to create delays constructed from

the fundamental frequencies of Chinese tone 2 and tone 4, delaying Gaussian noise (80

Hz-40 kHz) by the inversion of pitch and adding it back on itself in a recursive manner

(Yost 1996a). The core MATLAB program used for generating IRN was modified from

Krishnan et al. (Krishnan et al. 2014, 2015) This would generate dynamic, curvilinear pitch

patterns (Swaminathan et al. 2008) that preserves variations in pitch using a broadband

carrier. The number of iteration steps for these stimuli was 32, beyond which there is little

or no change in pitch salience (Yost 1996b).

IRN iteration (ite) stimuli were created with the same polynomial equations used for tone

2, but with different iterations to create an array of IRN stimuli with different pitch salience.

The numbers of iteration steps were 32, 16, 8, 4, and 2.

All IRN stimuli consisted of pairs of waveforms in original and inversed polarities to

compensate for envelope or fine structure response under different calculations and

cancel any microphonics. The stimulus duration was 250 msec, and the acquisition

window was 300 msec long. Each response was an average of 200 repetitions.

Statistics

Statistics were performed with statistics software JASP (Version 0.11, JASP Team, 2019).

All statistics for ABR and EFR utilized 2-way repeated measures ANOVA test ($\alpha = 0.05$) to

check the significance of each main effect and interaction, undergoing Greenhouse-

Geisser sphericity corrections (Greenhouse and Geisser 1959) and Tukey Post Hoc

corrections (Tukey 1949). For ABR statistics, Wave I (channel 2), III (channel 1) and V

(channel 2) were measured at each time point (Fig. 1B), corresponding to the auditory

nerve (Wave I), cochlear nucleus (Wave III), and rostral brainstem/IC sources (Wave V)

(Parthasarathy and Bartlett 2012; Simpson and Prendergast 2013). For EFR statistics,

responses were analyzed from channel 2 for 10 Hz and 45 Hz, and from channel 1 for

256 Hz (Parthasarathy and Bartlett 2012). Prior to statistical tests, EFR amplitudes at signal

frequencies were acquired through Fast Fourier Transformation (FFT) in MATLAB

(MathWorks) similar to (Lai and Bartlett 2018).

For MLR statistics, P1, N1, P2, and N2 (Fig. 5A) peaks were measured at each time point,

corresponding to subcortical (P1), thalamocortical (N1) and cortical sources (P2, N2)

274 (Simpson and Prendergast 2013). Peak amplitudes were normalized to the pre-blast

275 exposure baseline measurements for display in Fig. 5C, D. The peak amplitudes at each

276 time point were compared to the pre-stimulus baseline using a paired sign-rank test,

277 with a 0.05 significance criterion.

279 For IRN statistics, we performed moving-window autocorrelations in 25 msec moving

280 windows (5 msec steps) on each response waveform to simulate physiological tracking of

281 temporal periodicity. Peak autocorrelation frequency was defined by the inverse of the

282 time lag where peak autocorrelation value occurs in each window. This process yielded a

283 total of 51 peak frequencies that reflect the frequency representation of the IRN auditory

284 response. Of those, 45 occurred during the stimulus. The peak frequencies were then

285 compared to the "pseudopitches" of the IRN stimuli on corresponding time points. A

286 value within 5 Hz of absolute difference to corresponding "pseudopitch" was considered

287 "tracked." We used this number of "tracked" peak frequencies, or "pitch-tracking score,"

288 as a quantification for IRN performance. The significance of each main effect (time, blast

289 condition, and IRN iterations) and interaction was assessed using similar 2-way repeated

290 measures ANOVA test as ABR statistics ($\alpha = 0.05$). For response-to-response correlation

291 (Fig. 8D), the cross-correlation was measured between the response to the IRN stimuli

292 pre-exposure and the response to the same stimulus post-exposure. Blast versus sham

293 group was tested using the paired sign-rank test for this measure ($\alpha = 0.05$).

295 **Results**

          A.   ABR and MLR

   ABR Thresholds

   (Insert Fig. 2 about here)

   Click ABR recordings captured distinctive courses of threshold changes over the two

   months post-exposure for blast and sham animals (Fig. 2). A large, >30dB SPL maximum

   threshold increase was observed in post-blast-exposure animals (Fig. 2, red lines).

   Adjacent animals exposed only to blast noise (Sham) did not undergo significant

   threshold shifts (Fig. 2, blue lines). Thresholds for blast group animals showed clear

   recovery during the first two weeks, with the largest changes occurring between 4 days –

   10 days. Thresholds for blast-exposed animals remained significantly elevated (worse)

   than those of sham animals throughout the two months post-exposure that were

   measured (F=12.727, p=0.003). Significant main effects of both Group (F=61.943,

   p=<0.001, η^2_p=0.816) and Time Point (F=41.932, p<0.001, η^2_p=0.750), as well as a

   significant Group*Time Point interaction effect (F=23.503, p<0.001, η^2_p=0.627), were

   observed.

   Similar trends were observed with tone ABR recordings of 8 kHz and 16 kHz (Fig. 2), with

   a significant (p≤0.001) >30 dB increase in threshold within 48 hours post-blast-exposure

   and most prominent recovery between 4 days – 10 days. 8 kHz threshold differences

   between blast conditions became non-significant (F=3.151, p=0.098) at 10 days post-

   blast. At two weeks post-exposure, 16 kHz thresholds remained significantly elevated

   (F=16.527, p<0.001), after which point the thresholds for the two chosen tone frequencies

were no longer significantly different between Blast and Sham. Our rmANOVA analysis

using Group and Time Points as factors showed significant main effects of Group (8 kHz:

$F=10.847$, $p=0.005$, $\eta^2_p=0.437$; 16 kHz: $F=19.697$, $p<0.001$, $\eta^2_p=0.585$), Time (8 kHz:

$F=25.837$, $p<0.001$, $\eta^2_p=0.649$; 16 kHz: $F=20.181$, $p<0.001$, $\eta^2_p=0.590$) and Group*Time

Point interaction (8 kHz: $F=13.490$, $p<0.001$, $\eta^2_p=0.491$; 16 kHz: $F=15.860$, $p<0.001$, η^2

$_p=0.531$) for 8 kHz and 16 kHz threshold respectively. These results demonstrate that

broadband click thresholds remain significantly elevated over the 60 days measurement

window. 8 kHz thresholds largely returned to baseline (Day 30: 8 dB difference, $p=0.118$;

day 60: 4 dB difference, $p=0.965$) after two weeks, and 16 kHz thresholds remained

significantly elevated compared to pre-blast baseline according to post hoc analysis (Day

30: 15.5 dB difference, $p<0.001$; day 60: 14.5 dB difference, $p<0.001$), although the

difference between blast and Sham was not significant at these time points.

ABR Amplitudes

(Insert Fig. 3 about here)

For our ABR and MLR measurements, we used two sound levels: 80 dB SPL was chosen

because it is commonly used in auditory evoked potential studies in rat and human

studies (Simpson et al. 1985; Alvarado et al. 2012; Race et al. 2017), and it elicits clear ABR

responses in all except the most extreme cases of blast-exposure. In order to compensate

for changes in threshold induced by blast exposure, we also measured ABR amplitudes

at 30 dB SL above threshold (sensation level, or SL). This enabled us to separate changes

in ABR amplitudes due to audibility (threshold) versus those due to threshold-

independent changes in subcortical auditory signaling. Note that we did not attempt to

compare later ABR waves with equivalent wave I amplitudes, as in Lai et al. (2017).

ABR wave amplitudes were assessed for wave I (putative auditory nerve), III (putative

cochlear nuclei), and V (putative rostral brainstem and inferior colliculus) in response to

click stimuli at 80 dB SPL (Fig. 3) and 30 dB SL (Fig. 4). Repeated measures statistics for

80 dB SPL and 30dB SL are shown in Tables 1-4.

Wave I: Wave I amplitudes at 80 dB SPL for all ABR carriers at 80 dB SPL exhibited

significant main effects of Group, Time, and Group*Time interaction (Table 1). Compared

to pre-exposure responses, wave I amplitudes were significantly smaller at all time points

tested in blast animals for clicks, 8 kHz tones, and 16 kHz tones, indicating lasting

cochlear/auditory nerve damage (Table 4). No significant changes in wave I amplitudes

were observed in Sham exposed animals at any time point.

Wave III: Wave III amplitudes at 80 dB SPL for all ABR carriers at 80 dB SPL exhibited

significant main effects of Group, Time, and Group*Time interaction (Table 1), with Group

effects lasting for 14 days for Click and 16 kHz tones and 10 days for 8 kHz tones.

Compared to pre-exposure responses, wave III amplitudes were significantly smaller at

all time points tested in blast animals for clicks and 16 kHz tones and up to 30 days for 8

kHz tones, indicating lasting declines in cochlear nucleus excitation (Supplemental

Materials). No significant changes in wave III amplitudes were observed in Sham exposed

animals at any time point.

Wave V: Wave V amplitudes at 80 dB SPL for all ABR carriers at 80 dB SPL exhibited

significant main effects of Group, Time, and Group*Time interaction (Table 1), with Group

effects lasting for 14 days for Click and 16 kHz tones and 7 days for 8 kHz tones.

Compared to pre-exposure responses, wave V amplitudes were significantly smaller at all

time points tested in blast animals for clicks, indicating lasting declines in rostral

brainstem/IC excitation for brief, broadband clicks (Table 4). However, decreases in wave

V amplitudes persisted for only 7 days for 8 kHz tones and 14 days for 16 kHz tones,

suggesting that despite decreases in cochlear nucleus excitation (as represented by wave

III amplitude), rostral brainstem/IC responses compensated and restored their responses.

No significant changes in wave V amplitudes were observed in Sham exposed animals at

any time point except for a small decline for 16 kHz responses 60 days post Sham

exposure (Table 4).

(Insert Fig. 4 about here)

The effects on ABR waves were greatly diminished when responses to 30 dB SL sounds

were measured, as shown in Table 2 and Table 3. For Wave I, significant main effects of

Time (Click: $F=2.554$, $p=0.043$, $\eta^2_p=0.154$; 8 kHz: $F=3.146$, $p=0.018$, $\eta^2_p=0.183$; 16kHz:

$F=2.325$, $p=0.031$, $\eta^2_p=0.142$) but not Group (Click: $F=3.637$, $p=0.077$, $\eta^2_p=0.206$; 8 kHz:

$F<0.001$, $p=0.994$, $\eta^2_p<0.001$; 16kHz: $F=1.046$, $p=0.324$, $\eta^2_p=0.070$) were observed for

click, 8 kHz, and 16 kHz. Additionally, significant Group*Time interaction effects were only

observed for Click ($F=2.630$, $p=0.039$, $\eta^2_p=0.158$) and 16 kHz ($F=2.381$, $p=0.027$, $\eta^2_p=0.145$).

$_p=0.145$). Simple main effects of Group were only observed in Click (Table 3).

Compared to pre-exposure responses, wave I and V responses to clicks were significantly

reduced 1 day post-blast and wave III responses were significantly reduced days 1-4.

Otherwise, there were no significant declines in wave amplitudes in the Blast group, and

there were no significant amplitude changes in the Sham group.

(Insert Fig. 5 about here)

B.   MLR

In order to observe thalamocortical and cortical neural transmission in response to

acoustic transients, we recorded middle-latency auditory responses to click and 8 kHz

tone stimuli. These stimuli were identical to those used for ABR, but the presentation rate

was much slower, and the analysis window and filters were different (see Methods).

Measurements were made for the first four main peaks of the MLR. Here, P1 corresponds

to subcortical activity, largely encompassing the ABR. N1 corresponds to thalamocortical

transmission, while P2 and N2 are thought to correspond to primarily cortical activity

(Deiber et al. 1988; Liégeois-Chauvel et al. 1994; Tichko and Skoe 2017; Musiek and Nagle

2018).

80 dB SPL responses

In blast animals, all waves were decreased relative to pre-blast baseline for days 1-7 post-

blast (p<0.05, sign-rank test) in response to 80 dB SPL click stimuli. Grand average traces

are shown for MLR responses in this time window in Fig. 5A, relative to the pre-blast

waveform (thick blue line in A-D). Even after the blast, the morphology and timing of the

MLR waveform remained relatively intact, but the amplitudes were significantly

diminished. In Fig. 5E-H, wave amplitudes were normalized to the pre-blast waves and

measured. Between 7 and 10 days, the early P1 wave recovers to within 10-15% of its

baseline amplitude, whereas the later waves recovered more slowly (Fig. 5E). In particular,

the N1 wave, thought to reflect thalamocortical transmission (Barth and Shi Di 1991;

McGee et al. 1991, 1992; Di and Barth 1992; Brett et al. 1996; McGee and Kraus 1996;

Phillips et al. 2011; Šuta et al. 2011), remained significantly lower in blast animals even 60

days post-blast ($p < 0.05$, sign-rank test, Fig. 5E). By contrast, the MLR waves in sham

animals were largely stable across the measurement time (Fig. 5F). Although there was

some decline in the later waves for the last time window, this was not statistically

significant (Fig. 5B, D, F).

MLR responses to 8 kHz, 80 dB SPL tone pips largely mirrored the results to clicks, with

significant decreases for all waves for post-blast days 1-7 and a lasting decline in N1 for

the duration of measurements ($p < 0.05$, sign-rank test, traces not shown). Sham

responses did not show any significant changes in MLR waves in response to the 80 dB

SPL tone pips.

30 dB SL

MLR responses to clicks at 30 dB SL were reduced in Blast animals 1 day after the blast

but recovered to baseline levels afterwards. There was a tendency towards elevated P1

amplitudes, but this was not significant (Fig. 5G). Sham animals did not show any

significant changes, though there was a tendency towards an increase in wave amplitude

(Fig. 5H). Similar results were found for responses to tones at 30 dB SL (not shown).

C.   EFR and EFR in noise

Given the different time courses and extents of ABR threshold change for clicks and tones,

we measured the corresponding EFRs in response to Gaussian broadband noise (NAM),

8 kHz, and 16 kHz sinusoidal tone carriers. Considering that slow AM (<50 Hz) and faster

AM (>50 Hz) are differentially represented throughout cortical and subcortical auditory

nuclei (Joris et al. 2004; Wang et al. 2008), three representative AMFs (10, 45, and 256 Hz)

were selected from previous publications (Parthasarathy et al. 2010, 2014; Parthasarathy

and Bartlett 2011, 2012; Race et al. 2017) and tested in quiet at 100% and 50% modulation

depth. AM stimuli were also presented at 30 dB SL with a 71 Hz sinusoidally AM masker

of the same length and onset, with Gaussian noise as the carrier, at 20dB SNR and 0 SNR

relative to the sound level of target AM. For each carrier, simple main effects of all

conditions were analyzed.

(Insert Fig. 6 about here)

EFRs in quiet: For all three carriers in quiet, EFR amplitudes were similar at 10 and 256 Hz

across time points and AM modulation depths (Fig. 6). Overall, the NAM FFT amplitudes

were reduced in the Blast group in quiet ($F=9.629$, $p=0.0018$, $\eta^2_p=0.426$), with 45 Hz

being the most affected. Interestingly, in contrast to the lower FFT Amplitude found in

Blast AM at 80 dB SPL (Race et al. 2017), when the hearing threshold has been

compensated, FFT amplitude of 45 Hz NAM was higher in Blast than in Sham animals.

This difference was most salient on day 7 for 45 Hz NAM (p=0.006). For 8 kHz SAM and

16 kHz SAM, the slight elevation of AM FFT Amplitude in Blast animals was not significant.

Surprisingly, time did not have a significant interaction across repeated measures for AM

response with any carrier either.

EFR in noise: Not surprisingly, Noise level and Modulation Depth both had a significant

repeated measures effect on NAM (Noise level: $F=263.217$, $p<0.001$, $\eta^2_p=0.953$; Depth:

$F=455.655$, $p<0.001$, $\eta^2_p=0.972$), 8 kHz SAM (Noise level: $F=19.308$, $p<0.001$, $\eta^2_p=0.580$;

Depth: $F=72.031$, $p<0.001$, $\eta^2_p=0.837$) and 16 kHz SAM (Noise level: $F=16.691$, $p<0.001$,

$\eta^2_p=0.544$; Depth: $F=49.742$, $p<0.001$, $\eta^2_p=0.780$). Noise level and Depth also have a

significant interaction effect with Blast Groups for NAM overall (Noise level: $F=10.295$,

$p<0.001$, $\eta^2_p=0.442$; Depth: $F=6.057$, $p=0.029$, $\eta^2_p=0.318$), showing blast NAM responses

as less affected 20 SNR noise, but more sensitive amplitude declines for lower modulation

depth. Noise level also affects 8 kHz SAM differently between Groups ($F=5.696$, $p=0.008$,

$\eta^2_p=0.289$). These conditions do not have significant interaction effects with Blast Group

on 16 kHz SAM. Noise level had significant interaction effects with both Blast Group

($F=6.130$, $p=0.011$, $\eta^2_p=0.320$) and Depth ($F=19.438$, $p<0.001$, $\eta^2_p=0.599$) for NAM 45

Hz, while the effect of Time or Depth between Groups is not significantly different for any

modulation frequency.

(Insert Fig. 7 about here)

For 8 kHz SAM, the effects of Noise level were applied differently between Blast Groups,

as significant interaction effects were observed between Noise and Group for 10 Hz

(F=12.795, p=0.001, η^2_p=0.477) and 45 Hz (F=4.878, p=0.015, η^2_p=0.258) modulation

frequencies, though not for 256 Hz. Modulation Depth affects FFT amplitude without

regard to blast condition, with no significant interaction effects with Group observed. For

16 kHz SAM, none of the parameters tested had significantly different effects between

Groups at 30 dB SL (not shown).

D. IRN

Time-varying IRN stimuli were used to elicit frequency-following response (FFR)

mimicking Mandarin tone 2 (T2, rising) and tone 4 (T4, falling) pitch contours to measure

pitch-tracking ability using a broadband speech-like carrier at 30 dB SL, similar to what

has been measured in human studies of auditory learning and hearing loss (Anderson et

al. 2010, 2013; Skoe and Kraus 2010). We used autocorrelation interval contours that

simulated pitches similar to the forms of rising (T2) and falling (T4) pitch contours of the

Mandarin Chinese vowel /yi/ (Krishnan et al. 2014, 2015, 2017a, 2017b). IRN responses

were evaluated based on the pitch-tracking score, which measures the number of

windows where the dominant autocorrelation frequency of the response matches that of

the IRN stimulus autocorrelation frequency (see Methods). In general, we observed a loss

of pitch-tracking fidelity in Blast animals over the two months post-exposure. Even for

the most salient pitch (32 Iterations), blast exposure had a significant Group effect on

pitch-tracking scores in both Tone 2 (F=6.495, p=0.026, η^2_p=0.351) and Tone 4 (F=6.115,

p=0.029, η^2_p=0.338), with the largest mean differences on day 7-10. The interaction

effect between Time and Group was not significant.

Blast exposure significantly changed the neural response's morphology to IRN at 30 dB

SL (p=0.016, paired sign-rank test, Fig. 8D), such that the cross-correlation between the

pre-exposure response and the post-exposure response was much lower in the Blast

group up to 30 days post-blast.

(Insert Fig. 8 about here)

IRN iterations: As expected, reduced pitch salience, controlled by reducing iteration

number, affected pitch-tracking responses in animals (F=41.697, p<0.001, η^2_p=0.777),

also showing a significant interaction effect with Time post-exposure (F=1.722, p=0.031,

η^2_p=0.125). Specifically, pitch-tracking performances to 32 iterations and 16 iterations

worsened significantly up to 7-10 days post-exposure, with various degrees of recovery

over the following time course. Both the Blast and Sham group exhibited worse pitch

tracking with reduced iterations (salience) and to a similar degree. No significant

interaction effects with Group were observed for Time and Iterations (Fig 9).

(Insert Fig. 9 about here)

Discussion

The purpose of this study was to examine the time course of recovery from a single mild

blast injury using simple and complex auditory stimuli longitudinally over a two-month

period. Our study demonstrated drastic changes in high-frequency and click hearing

thresholds from ABR recordings and reductions in putative subcortical and

thalamocortical transmission from ABR and MLR recordings throughout the first two

weeks post-exposure. At 30-60 days post-blast, significant increases in click threshold,

decreases in wave I amplitude, decreases in MLR N1 amplitude, and declines in pitch-

tracking of speech-like IRN pitch trajectories were observed. Compensating for threshold

shift and using 30 dB sensation level for AM stimuli, we found that responses to sinusoidal

AM stimuli in quiet or noise recovered within 14 days. The 7-14 day window was

particularly rapid in the recovery of many auditory parameters, suggesting this as a

therapeutic time period when important changes take place.

ABR

We documented the rapid increase of threshold and slow recovery after trauma from

blast-exposure. Most notably, a >30 dB peak increase in threshold was observed for

click, 8 kHz, and 16 kHz (Fig. 2) during the first 4 days. This is consistent with the

description of IHC and OHC disturbances across a wide range of frequencies due to

blast overpressure as stated in multiple previous publications (Patterson and Hamernik

1997; Ewert et al. 2012; Race et al. 2017). Cho et al. (2013b) reported that ABRs exhibit

the least recovery at the highest and lowest frequencies. This broadband threshold shift

extended to the last time point at 60 days, though the ~10 dB difference would not be

considered clinically relevant.

Over the two-month observation, rapid improvements in ABR threshold and wave

amplitudes were observed in the 7-10 days recovery period for waves I, wave III, and

wave V (Fig. 3). Such rapid recovery could be explained by neuroplastic remodeling within

central auditory ABR generator regions (Mulders and Robertson 2009, 2011, 2013;

Mulders et al. 2011; Manzoor et al. 2013; Robertson et al. 2013) and recovery of hearing

thresholds. Notably, wave V amplitude recovered earlier than wave I, possibly indicating

the role of compensation in auditory midbrain as one of the post-blast recovery

mechanisms.

We hypothesized that the recovery of ABR threshold and wave I amplitude were the

results of a two-fold process, with persistent changes contributing to degraded speech

perception (Holmes and Griffiths 2019; Yeend et al. 2019). These parameters showed two

waves of post-blast changes: one between 1-10 days post-exposure, and one 10-30

days, as evidenced by Figs. 2 and 3. We hypothesize that these two waves of deficits

indicated the distinction of primary, physical impacts to multiple loci in PAS and CAS

(Garman et al. 2011; Ewert et al. 2012; Cho et al. 2013b; Song et al. 2015; Walls et al. 2016)

and a series of secondary, biochemical impacts surrounding CAS (Laplaca et al. 1997;

Knudsen and Øen 2003; Hamann et al. 2008; Garman et al. 2011; Säljö et al. 2011; Luo et

al. 2014a, 2014b; Song et al. 2015; Walls et al. 2016). Our observations of blast recovery

were mostly consistent with this hypothesis.

MLR

At 80 dB SPL, we found persistent deficits in thalamocortical and cortical transmission

based on the N1, P2 and N2 peaks (Fig. 5A vs. B, C vs. D), which were affected at 30 and

60 days, even after the early P1 response had fully recovered (Fig 5E). These deficits were

not present at 30 dB SL, suggesting that effects were at least partially due to shifts in

auditory thresholds (Fig. 5G), but the moderate, subclinical threshold shifts observed are

probably highly relevant functionally. In veterans and the general population with lifetime

noise exposure, MLR responses were shown to be smaller even when subjects had

clinically normal audiograms, and there was some evidence of increased cortical gain

(Valderrama et al. 2018; Bramhall et al. 2020). In another study with blast-exposed

veterans, most of the changes in auditory-evoked potentials were correlated with hearing

loss (Meehan et al. 2019). Together, these results suggest that hearing loss may be the

main contributor to MLR changes leading to declines in suprathreshold responses.

However, more work needs to be done for sustained sounds post-blast to examine the

relationships between early evoked potentials, representing subcortical neural activities,

and later potentials, representing cortical activities, since studies in aging and noise

exposure have suggested enhanced cortical responses (Syka and Rybalko 2000; Sun et al.

2012; Bidelman et al. 2014; Herrmann et al. 2019). These results also suggest that there

may be more pronounced hearing deficits from intense combined auditory and

mechanical trauma such as blast, compared to medium to intense level of auditory

exposure alone, such as recreational or occupational noise (Fulbright et al. 2017; Grinn et

al. 2017; Guest et al. 2018).

Amplitude Modulation EFRs

The current study extended an earlier study (Race et al. 2017) to include EFR responses

to more challenging auditory stimuli, including lower modulation depth (Fig 6) and in the

presence of modulated noise (Fig 7). The Race et al. (2017) study revealed differences in

AM processing at 80 dB SPL between Blast and Sham animals, such that blast animals

had lower AM FFR amplitudes, especially at lower modulation frequencies in the 20-50

Hz range. However, when the hearing threshold has been compensated, the differences

in AM FFR amplitude diminished and even changed sign (Fig. 6), suggesting that both

changes in audibility and changes in the gain of subcortical auditory system are critical

contributors to AM FFR deficits in the blast-exposed auditory system. There are

complicated interactions between the AM FFR amplitudes, blast exposure, and the

presence of noise, evident as a persistent Group*Noise Level interaction effect in both

NAM and 8 kHz SAM. AM responses consist of contributions from multiple generators

along the auditory neuraxis, with cortical generators contributing mainly to lower AMFs

<50 Hz, and higher frequency AM responses limited to nuclei lower in the auditory

neuraxis. The lack of blast-induced differences at higher AMFs distinguishes the blast-

induced damage from age-related changes, which are most prominent at higher

modulation frequencies (Parthasarathy et al. 2010, Parthasarathy and Bartlett 2012, Lai et

al. 2017).

The differences in low-middle AMFs were manifested in opposed directions under slow

(10 Hz) and middle (45 Hz) AMFs: notably, normalized FFT amplitudes of 8 kHz SAM in

noise is lower for Blast at 10 Hz modulation frequency, but higher for Blast at 45 Hz (Fig

7D). This dichotomy is ripe for further study since the 10 Hz and 45 Hz modulations

represent different temporal processing regimes and components of speech (Rosen

1992). In studies on age-related auditory nerve deficit models and AM-in-noise

responses in aged animals, decreased sensitivities to lower SNRs in AM-in-noise in

damaged auditory systems have been observed (Parthasarathy et al. 2016; Lai et al. 2017;

Lai and Bartlett 2018). This phenomenon could partly explain the changes seen in 45 Hz

AMF. It should be noted that the variance in AM responses post-blast exposure could

also be a result of post-blast increases in spontaneous activity for DCN and IC units

sensitive to tone versus noise carriers (Luo et al. 2014a, 2014b), which could either

desynchronize responses or strengthen AM response simply by heightened excitability.

Taken together, our results suggested that: 1) CAS neuroplasticity partially compensates

for hearing loss after blast exposure; and 2) surprisingly, at least for a single mild blast

with small lasting threshold shifts, our results suggest that sinusoidal AM processing in

quiet and noise largely recovers.

IRN EFRs

Bressler et al. (2017) have noted that perceptual and behavioral deficits persist in blast-

exposed veterans in the absence of major clinical threshold or ABR abnormalities.

Simple auditory tests such as ABRs and AM EFRs in a quiet background are limited

indicators of auditory challenges that blast-exposed individuals would face in daily life.

The present study investigated how noise affects responses post-blast as well as using

IRN stimuli that simulate the pitch trajectories and sound like simple speech sounds.

These stimuli may capture the more complex hearing deficits reported by humans who

have experienced blast injuries (Gallun et al. 2012a; Saunders et al. 2015; Kubli et al.

2018).

Complex temporal periodicity between 50 Hz and 500 Hz carries important speech

information such as voicing, stress and intonation (Rosen 1992). The present study

provided insights into blast-induced sound processing deficits through the use of an

IRN stimulus that simulates Chinese intonations and whose pitch and salience can be

reliably controlled, showing that IRN can be a useful diagnostic tool for neurotrauma.

We found out that even when click ABR thresholds have returned to subclinical

threshold shifts, the deficits in pitch-tracking response to IRN tone stimuli, lingered at

least 30 days post-exposure (Figs. 8D).

Decreased salience (fewer iterations), showed an overall reduction in pitch tracking in

Blast animals (Fig. 9), particularly for higher iterations which should be most salient, but

they were not differentially affected by the number of iterations. It is worth noting that a

previous study showed that increased IRN iterations improved auditory stream

segregation in normal hearing veterans more than hearing-impaired veterans (Shearer

et al. 2018). Our IRN data (Figs. 8-9) suggest that more dynamic and speech-like

modulation changes do not recover quickly or completely from even a single mild blast

exposure.

Limitations and Future Directions

This study has examined injuries elicited by a single dorsal blast exposure with body

shielding that did not result in tympanic membrane ruptures. Therefore, the deficits

observed may not be as drastic as that documented by some previous studies in which

the injuries were caused by more intense or multiple exposures (Cho et al. 2013b; Du et

al. 2013; Luo et al. 2014a, 2014b; Mahmood et al. 2014), often resulting in death or

tympanic membrane rupture. The distribution of injuries also differed from models in

which blast exposure comes from different orientations, as predicted in animals (Chavko

et al. 2011; Dal Cengio Leonardi et al. 2012) and computational studies (Hua et al. 2017;

Unnikrishnan et al. 2019). These differences in pressure wave amplitude, duration, and

propagation patterns would affect both the distribution and severity of damage across

the brain. Although these could change the potential mechanisms of recovery and

compensation, it is likely that all blast exposures undergo a multi-stage recovery process

similar to that observed in the present study. Our model utilized a top-down mild blast

model as described by Song et al. (2015), which exerts a blast impact from the top of the

skull. The overpressure blast wave passes through the entire rat brain, such that TBI can

be observed throughout the brain, including the frontal cortex and in multiple thalamic

regions (Walls et al. 2016), and it results in increased ventral BBB membrane permeation

and inflammation, encompassing many subcortical auditory nuclei and axonal tracts.

While overlapping loci of vulnerability were described in anatomical studies of animal

exposed to blast impacts of different or unknown orientations (Knudsen and Øen 2003;

Säljö et al. 2011; Mao et al. 2012b), additional injury mechanisms of different loci and

nature may be expected from blast impacts from a different angle. Moreover, non-

invasive physiological measurements of the auditory thalamus and cortex may be

indicators of more widespread blast damage in auditory and non-auditory brain regions.

Conclusion

Based on our chronic longitudinal AEP measurements over a two-month period, this

study documented the significant changes in auditory pathophysiology and a potential

multi-stage recovery process in animals exposed to a single mild blast trauma. We found

that hearing thresholds largely recovered except for broadband stimuli, and some aspects

of lasting deficits could be explained by threshold shifts. Recovery of temporal processing

of AM sounds was confined to lower modulation frequencies (<50 Hz), with responses

dependent on time point, sound level, noise masking, and AMF.   More complex, speech-

like stimuli (IRN) demonstrated lasting deficits, pointing towards more effective stimuli for

elucidating the behavioral difficulties in human and animal listeners.

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Riyi Shi is a co-founder of Neuro Vigor, a company developing novel drug treatments

and diagnostic approaches for neurodegenerative diseases and neurotrauma.

References

Alsindi S, Patterson RD, Sayles M, Winter IM. The responses of single units to simple and

complex sounds from the superior olivary complex of the Guinea pig. In: *Acta Acustica*

united with Acustica. S. Hirzel Verlag GmbH, 2018, p. 856–859.

Alvarado JC, Fuentes-Santamaría V, Jareño-Flores T, Blanco JL, Juiz JM. Normal

variations in the morphology of auditory brainstem response (ABR) waveforms: A study

in wistar rats. *Neurosci Res* 73: 302–311, 2012.

Anderson S, Parbery-Clark A, White-Schwoch T, Kraus N. Auditory brainstem response

to complex sounds predicts self-reported speech-in-noise performance. *J Speech, Lang*

Hear Res 56: 31–43, 2013.

Anderson S, Skoe E, Chandrasekaran B, Kraus N. Neural timing is linked to speech

perception in noise. *J Neurosci* 30: 4922–4926, 2010.

Barth DS, Shi Di. The functional anatomy of middle latency auditory evoked potentials.

Brain Res 565: 109–115, 1991.

Bendor D, Wang X. The neuronal representation of pitch in primate auditory cortex.

Nature 436: 1161–1165, 2005.

Berger G, Finkelstein Y, Avraham S, Himmelfarb M. Patterns of hearing loss in non-

explosive blast injury of the ear. *J Laryngol Otol* 111: 1137–1141, 1997.

Bharadwaj HM, Masud S, Mehraei G, Verhulst S, Shinn-Cunningham BG. Individual

differences reveal correlates of hidden hearing deficits. *J Neurosci* 35: 2161–2172, 2015.

Bidelman GM, Villafuerte JW, Moreno S, Alain C. Age-related changes in the

subcortical-cortical encoding and categorical perception of speech. *Neurobiol Aging*

35: 2526–2540, 2014.

Bramhall NF, **Niemczak CE**, **Kampel SD**, **Billings CJ**, **McMillan GP**. Evoked Potentials

Reveal Noise Exposure–Related Central Auditory Changes Despite Normal Audiograms.

Am J Audiol 29: 152–164, 2020.

Bressler S, **Goldberg H**, **Shinn-Cunningham B**. Sensory coding and cognitive processing

of sound in Veterans with blast exposure. *Hear Res* 349: 98–110, 2017.

Brett B, **Krishnan G**, **Barth DS**. The effects of subcortical lesions on evoked potentials and

spontaneous high frequency (gamma-band) oscillating potentials in rat auditory cortex.

Brain Res 721: 155–166, 1996.

Caspary DM, **Ling L**, **Turner JG**, **Hughes LF**. Inhibitory neurotransmission, plasticity and

aging in the mammalian central auditory system. *J Exp Biol* 211: 1781–1791, 2008.

Caspary DM, **Schatteman T a**, **Hughes LF**. Age-related changes in the inhibitory

response properties of dorsal cochlear nucleus output neurons: role of inhibitory inputs.

J Neurosci 25: 10952–9, 2005.

Cave K, **Cornish EM**, **Chandler DW**. Blast Injury of the Ear: Clinical Update from the

Global War on Terror. *Mil Med* 172: 726–730, 2007.

Chavko M, **Watanabe T**, **Adeeb S**, **Lankasky J**, **Ahlers ST**, **McCarron RM**. Relationship

between orientation to a blast and pressure wave propagation inside the rat brain. *J*

Neurosci Methods 195: 61–66, 2011.

Cho HJ, **Sajja VSSS**, **VandeVord PJ**, **Lee YW**. Blast induces oxidative stress, inflammation,

neuronal loss and subsequent short-term memory impairment in rats. *Neuroscience*

253: 9–20, 2013a.

Cho S II, Gao SS, Xia A, Wang R, Salles FT, Raphael PD, Abaya H, Wachtel J, Baek J,

Jacobs D, Rasband MN, Oghalai JS. Mechanisms of Hearing Loss after Blast Injury to the

Ear. *PLoS One* 8, 2013b.

Cohen JT, Ziv G, Bloom J, Zikk D, Rapoport Y, Himmelfarb MZ. Blast injury of the ear in a

confined space explosion: Auditory and vestibular evaluation. *Isr Med Assoc J* 4: 559–

562, 2002.

Dal Cengio Leonardi A, Keane NJ, Bir CA, Ryan AG, Xu L, VandeVord PJ. Head

orientation affects the intracranial pressure response resulting from shock wave loading

in the rat. *J Biomech* 45: 2595–2602, 2012.

Deiber MP, Ibañez V, Fischer C, Perrin F, Mauguière F. Sequential mapping favours the

hypothesis of distinct generators for Na and Pa middle latency auditory evoked

potentials. *Electroencephalogr Clin Neurophysiol Evoked Potentials* 71: 187–197, 1988.

DePalma RG, Burris DG, Champion HR, Hodgson MJ. Current concepts: Blast injuries. *N.

Engl. J. Med.* 352Massachusetts Medical Society: 1335–1342, 2005.

Di S, Barth DS. The functional anatomy of middle-latency auditory evoked potentials:

Thalamocortical connections. *J Neurophysiol* 68: 425–431, 1992.

Du X, Ewert DL, Cheng W, West MB, Lu J, Li W, Floyd RA, Kopke RD. Effects of

antioxidant treatment on blast-induced brain injury. *PLoS One* 8: e80138, 2013.

Ewert DL, Lu J, Li W, Du X, Floyd R, Kopke R. Antioxidant treatment reduces blast-

induced cochlear damage and hearing loss. *Hear Res* 285: 29–39, 2012.

Fulbright ANC, Le Prell CG, Griffiths SK, Lobarinas E. Effects of Recreational Noise on

Threshold and Suprathreshold Measures of Auditory Function. *Semin Hear* 38: 298–318,

2017.

Gallun FJ, Diedesch AC, Kubli LR, Walden TC, Folmer RL, Samantha Lewis S, McDermott

DJ, Fausti SA, Leek MR. Performance on tests of central auditory processing by

individuals exposed to high-intensity blasts. *J Rehabil Res Dev* 49: 1005–1024, 2012a.

Gallun FJ, Lewis MS, Folmer RL, Diedesch AC, Aud ;, Kubli LR, McDermott DJ, Walden

TC, Fausti S a, Lew HL, Leek MR. Implications of blast exposure for central auditory

function: a review. *J Rehabil Res Dev* 49: 1059–74, 2012b.

Garman RH, Jenkins LW, Switzer RC, Bauman RA, Tong LC, Swauger P V., Parks SA,

Ritzel D V., Dixon CE, Clark RSB, Bayır H, Kagan V, Jackson EK, Kochanek PM. Blast

Exposure in Rats with Body Shielding Is Characterized Primarily by Diffuse Axonal Injury.

J Neurotrauma 28: 947–959, 2011.

Greenhouse SW, Geisser S. On methods in the analysis of profile data. *Psychometrika*

24: 95–112, 1959.

Grinn SK, Wiseman KB, Baker JA, Le Prell CG. Hidden hearing loss? No effect of common

recreational noise exposure on cochlear nerve response amplitude in humans. *Front*

Neurosci 11: 465, 2017.

Guest H, Munro KJ, Prendergast G, Millman RE, Plack CJ. Impaired speech perception in

noise with a normal audiogram: No evidence for cochlear synaptopathy and no relation

to lifetime noise exposure. *Hear Res* 364: 142–151, 2018.

Van Haesendonck G, Van Rompaey V, Gilles A, Topsakal V, Van de Heyning P. Otologic

Outcomes After Blast Injury: The Brussels Bombing Experience. *Otol Neurotol* 39: 1250–

1255, 2018.

Hamann K, Durkes A, Ouyang H, Uchida K, Pond A, Shi R. Critical role of acrolein in

secondary injury following ex vivo spinal cord trauma. *J Neurochem* 107: 712–721, 2008.

Herrmann B, Buckland C, Johnsrude IS. Neural signatures of temporal regularity

processing in sounds differ between younger and older adults. *Neurobiol Aging* 83: 73–

85, 2019.

Hickox AE, Liberman MC. Is noise-induced cochlear neuropathy key to the generation

of hyperacusis or tinnitus? *J Neurophysiol* 111: 552–564, 2014.

Holmes E, Griffiths TD. 'Normal' hearing thresholds and fundamental auditory grouping

processes predict difficulties with speech-in-noise perception. *Sci Rep* 9: 1–11, 2019.

Hua Y, Wang Y, Gu L. Primary blast waves induced brain dynamics influenced by head

orientations. *Biomed Eng Lett* 7: 253–259, 2017.

Joris PX, Schreiner CE, Rees A. *Neural Processing of Amplitude-Modulated Sounds*.

American Physiological Society, 2004.

Kerr AG. The effects of blast on the ear. *J Laryngol Otol* 94: 107–110, 1980.

Knudsen SK, Øen EO. Blast-induced neurotrauma in whales. *Neurosci Res* 46: 377–386,

2003.

Krishnan A, Gandour JT, Suresh CH. Cortical pitch response components show

differential sensitivity to native and nonnative pitch contours. *Brain Lang* 138: 51–60,

2014.

Krishnan A, Gandour JT, Suresh CH. Pitch processing of dynamic lexical tones in the

auditory cortex is influenced by sensory and extrasensory processes. *Eur J Neurosci* 41:

1496–1504, 2015.

Krishnan A, **Gandour JT**, **Xu Y**, **Suresh CH**. Language-dependent changes in pitch-

relevant neural activity in the auditory cortex reflect differential weighting of temporal

attributes of pitch contours. *J Neurolinguistics* 41: 38–49, 2017a.

Krishnan A, **Suresh CH**, **Gandour JT**. Changes in pitch height elicit both language-

universal and language-dependent changes in neural representation of pitch in the

brainstem and auditory cortex. *Neuroscience* 346: 52–63, 2017b.

Kubli LR, **Brungart D**, **Northern J**. Effect of Blast Injury on Auditory Localization in Military

Service Members. *Ear Hear* 39: 457–469, 2018.

Kujawa SG, **Liberman MC**. Synaptopathy in the noise-exposed and aging cochlea:

Primary neural degeneration in acquired sensorineural hearing loss. *Hear Res* 330: 191–

199, 2015.

Lai J, **Bartlett EL**. Age-related shifts in distortion product otoacoustic emissions peak-

ratios and amplitude modulation spectra. *Hear Res* 327: 186–198, 2015.

Lai J, **Bartlett EL**. Masking Differentially Affects Envelope-following Responses in Young

and Aged Animals. *Neuroscience* 386: 150–165, 2018.

Lai J, **Sommer AL**, **Bartlett EL**. Age-related changes in envelope-following responses at

equalized peripheral or central activation. *Neurobiol Aging* 58: 191–200, 2017.

Laplaca MC, **Lee VMY**, **Thibault LE**. An in vitro model of traumatic neuronal injury:

Loading rate-dependent changes in acute cytosolic calcium and lactate dehydrogenase

release. *J Neurotrauma* 14: 355–368, 1997.

Leung LY, **VandeVord PJ**, **Dal Cengio AL**, **Bir C**, **Yang KH**, **King AI**. Blast related

neurotrauma: A review of cellular injury. *MCB Mol. Cell. Biomech.* 5: 155–168, 2008.

Lew HL, Garvert DW, Pogoda TK, Hsu P Te, Devine JM, White DK, Myers PJ, Goodrich

GL. Auditory and visual impairments in patients with blast-related traumatic brain injury:

Effect of dual sensory impairment on functional independence measure. *J Rehabil Res*

Dev 46: 819–826, 2009.

Liberman MC. Noise-induced and age-related hearing loss: New perspectives and

potential therapies. *F1000Research* 6: 1–11, 2017.

Liégeois-Chauvel C, Musolino A, Badier JM, Marquis P, Chauvel P. Evoked potentials

recorded from the auditory cortex in man: evaluation and topography of the middle

latency components. *Electroencephalogr Clin Neurophysiol Evoked Potentials* 92: 204–

214, 1994.

Luo H, Pace E, Zhang X, Zhang J. Blast-Induced tinnitus and spontaneous firing changes

in the rat dorsal cochlear nucleus. *J Neurosci Res* 92: 1466–1477, 2014a.

Luo H, Pace E, Zhang X, Zhang J. Blast-induced tinnitus and spontaneous activity

changes in the rat inferior colliculus. *Neurosci Lett* 580: 47–51, 2014b.

Mahmood G, Mei Z, Hojjat H, Pace E, Kallakuri S, Zhang JS. Therapeutic effect of

sildenafil on blast-induced tinnitus and auditory impairment. *Neuroscience* 269: 367–

382, 2014.

Manzoor NF, Gao Y, Licari F, Kaltenbach JA. Comparison and contrast of noise-induced

hyperactivity in the dorsal cochlear nucleus and inferior colliculus. *Hear Res* 295: 114–

123, 2013.

Mao JC, Pace E, Pierozynski P, Kou Z, Shen Y, Vandevord P, Haacke EM, Zhang X, Zhang

J. Blast-induced tinnitus and hearing loss in rats: Behavioral and imaging assays. *J*

Neurotrauma 29: 430–444, 2012a.

Mao JC, **Pace E**, **Pierozynski P**, **Kou Z**, **Shen Y**, **VandeVord P**, **Haacke EM**, **Zhang X**,

Zhang J. Blast-Induced Tinnitus and Hearing Loss in Rats: Behavioral and Imaging

Assays. *J Neurotrauma* 29: 430–444, 2012b.

Masri S, **Zhang LS**, **Luo H**, **Pace E**, **Zhang J**, **Bao S**. Blast Exposure Disrupts the Tonotopic

Frequency Map in the Primary Auditory Cortex. *Neuroscience* 379: 428–434, 2018.

McGee T, **Kraus N**. Auditory development reflected by middle latency response. *Ear*

Hear 17: 419–429, 1996.

McGee T, **Kraus N**, **Comperatore C**, **Nicol T**. Subcortical and cortical components of the

MLR generating system. *Brain Res* 544: 211–220, 1991.

McGee T, **Kraus N**, **Littman T**, **Nicol T**. Contributions of medial geniculate body

subdivisions to the middle latency response. *Hear Res* 61: 147–154, 1992.

Meehan A, **Hebert D**, **Deru K**, **Weaver LK**. Longitudinal study of hyperbaric oxygen

intervention on balance and affective symptoms in military service members with

persistent post-concussive symptoms. *J Vestib Res Equilib Orientat* 29: 205–219, 2019.

Mulders WHAM, **Ding D**, **Salvi R**, **Robertson D**. Relationship between auditory

thresholds, central spontaneous activity, and hair cell loss after acoustic trauma. *J Comp*

Neurol 519: 2637–2647, 2011.

Mulders WHAM, **Robertson D**. Hyperactivity in the auditory midbrain after acoustic

trauma: dependence on cochlear activity. *Neuroscience* 164: 733–746, 2009.

Mulders WHAM, **Robertson D**. Progressive centralization of midbrain hyperactivity after

acoustic trauma. *Neuroscience* 192: 753–760, 2011.

Mulders WHAM, **Robertson D**. Development of hyperactivity after acoustic trauma in

the guinea pig inferior colliculus. *Hear Res* 298: 104–108, 2013.

Musiek F, **Nagle S**. The middle latency response: A review of findings in various central

nervous system lesions. *J. Am. Acad. Audiol.* 29American Academy of Audiology: 855–

867, 2018.

Parthasarathy A, **Bartlett E**. Two-channel recording of auditory-evoked potentials to

detect age-related deficits in temporal processing. *Hear Res* 289: 52–62, 2012.

Parthasarathy A, **Bartlett EL**. Age-related auditory deficits in temporal processing in F-

344 rats. *Neuroscience* 192: 619–630, 2011.

Parthasarathy A, **Cunningham PA**, **Bartlett EL**. Age-related differences in auditory

processing as assessed by amplitude-modulation following responses in quiet and in

noise. *Front Aging Neurosci* 2: 152, 2010.

Parthasarathy A, **Datta J**, **Torres JAL**, **Hopkins C**, **Bartlett EL**. Age-related changes in the

relationship between auditory brainstem responses and envelope-following responses.

JARO - J Assoc Res Otolaryngol 15: 649–661, 2014.

Parthasarathy A, **Lai J**, **Bartlett EL**. Age-Related Changes in Processing Simultaneous

Amplitude Modulated Sounds Assessed Using Envelope Following Responses. *JARO - J*

Assoc Res Otolaryngol 17: 119–132, 2016.

Patterson JH, **Hamernik RP**. Blast overpressure induced structural and functional

changes in the auditory system. 1997.

Patterson RD, **Handel S**, **Yost WA**, **Jaysurya Datta A**. The relative strength of the tone

and noise components in iterated rippled noise. *J Acoust Soc Am* 100: 3286–3294, 1996.

Peter V, Wong K, Narne VK, Sharma M, Purdy SC, McMahon C. Assessing spectral and

temporal processing in children and adults using Temporal Modulation Transfer

Function (TMTF), Iterated Ripple Noise (IRN) perception, and Spectral Ripple

Discrimination (SRD). *J Am Acad Audiol* 25: 210–218, 2014.

Phillips DJ, Schei JL, Meighan PC, Rector DM. State-Dependent Changes in Cortical Gain

Control as Measured by Auditory Evoked Responses to Varying Intensity Stimuli. *Sleep*

34: 1527–1537, 2011.

Plack CJ, Barker D, Prendergast G. Perceptual consequences of "hidden" hearing loss.

Trends Hear 18: 233121651455062, 2014.

Rabang CF, Parthasarathy A, Venkataraman Y, Fisher ZL, Gardner SM, Bartlett EL. A

computational model of inferior colliculus responses to amplitude modulated sounds in

young and aged rats. *Front Neural Circuits* 6: 77, 2012.

Race N, Lai J, Shi R, Bartlett EL. Differences in postinjury auditory system

pathophysiology after mild blast and nonblast acute acoustic trauma. *J Neurophysiol*

118: 782–799, 2017.

Rafaels K, "Dale" Bass CR, Salzar RS, Panzer MB, Woods W, Feldman S, Cummings T,

Capehart B. Survival Risk Assessment for Primary Blast Exposures to the Head. *J*

Neurotrauma 28: 2319–2328, 2011.

Remenschneider AK, Lookabaugh S, Aliphas A, Brodsky JR, Devaiah AK, Dagher W,

Grundfast KM, Heman-Ackah SE, Rubin S, Sillman J, Tsai AC, Vecchiotti M, Kujawa SG,

Lee DJ, Quesnel AM. Otologic outcomes after blast injury: The Boston Marathon

experience. *Otol Neurotol* 35: 1825–1834, 2014.

Ritenour AE, Wickley A, Ritenour JS, Kriete BR, Blackbourne LH, Holcomb JB, Wade CE.

Tympanic membrane perforation and hearing loss from blast overpressure in Operation

Enduring Freedom and Operation Iraqi Freedom wounded. *J Trauma* 64, 2008.

Robertson D, Bester C, Vogler D, Mulders WHAM. Spontaneous hyperactivity in the

auditory midbrain: Relationship to afferent input. *Hear Res* 295: 124–129, 2013.

Rosen S. Temporal information in speech: acoustic, auditory and linguistic aspects.

Philos. Trans. R. Soc. Lond. B. Biol. Sci. 336The Royal Society London: 367–373, 1992.

Säljö A, Mayorga M, Bolouri H, Svensson B, Hamberger A. Mechanisms and

pathophysiology of the low-level blast brain injury in animal models. *Neuroimage* 54:

S83–S88, 2011.

Saunders GH, Frederick MT, Arnold M, Silverman S, Chisolm TH, Myers P. Auditory

difficulties in blast-exposed veterans with clinically normal hearing. *J Rehabil Res Dev*

52: 343–360, 2015.

Shearer DE, Molis MR, Bennett KO, Leek MR. impaired listeners by normal-hearing and

hearing-impaired listeners a). 378, 2018.

Simpson G V., Knight RT, Brailowsky S, Prospero-Garcia O, Scabini D. Altered peripheral

and brainstem auditory function in aged rats. *Brain Res* 348: 28–35, 1985.

Simpson MIG, Prendergast G. Auditory magnetic evoked responses. In: *Handbook of*

Clinical Neurophysiology. 2013, p. 253–270.

Skoe E, Kraus N. Auditory Brain Stem Response to Complex Sounds: A Tutorial. *Ear Hear*

31: 302–324, 2010.

Song S, Race NS, Kim A, Zhang T, Shi R, Ziaie B. A Wireless Intracranial Brain

Deformation Sensing System for Blast-Induced Traumatic Brain Injury. *Sci Rep* 5: 1–10,

2015.

Sun W, Deng A, Jayaram A, Gibson B. Noise exposure enhances auditory cortex

responses related to hyperacusis behavior. *Brain Res* 1485: 108–116, 2012.

Šuta D, Rybalko N, Pelánová J, Popelář J, Syka J. Age-related changes in auditory

temporal processing in the rat. *Exp Gerontol* 46: 739–746, 2011.

Swaminathan J, Krishnan A, Gandour JT. Pitch encoding in speech and nonspeech

contexts in the human auditory brainstem. *Neuroreport* 19: 1163–1167, 2008.

Syka J, Rybalko N. Threshold shifts and enhancement of cortical evoked responses after

noise exposure in rats. *Hear Res* 139: 59–68, 2000.

Tichko P, Skoe E. Frequency-dependent fine structure in the frequency-following

response: The byproduct of multiple generators. *Hear Res* 348: 1–15, 2017.

Tukey JW. Comparing Individual Means in the Analysis of Variance. *Biometrics* 5: 99,

1949.

Unnikrishnan G, Mao H, Sundaramurthy A, Bell ED, Yeoh S, Monson K, Reifman J. A 3-D

Rat Brain Model for Blast-Wave Exposure: Effects of Brain Vasculature and Material

Properties. *Ann Biomed Eng* 47: 2033–2044, 2019.

Valderrama JT, Beach EF, Yeend I, Sharma M, Van Dun B, Dillon H. Effects of lifetime

noise exposure on the middle-age human auditory brainstem response, tinnitus and

speech-in-noise intelligibility. *Hear Res* 365: 36–48, 2018.

Viana LM, O'Malley JT, Burgess BJ, Jones DD, Oliveira CACP, Santos F, Merchant SN,

Liberman LD, Liberman MC. Cochlear neuropathy in human presbycusis: Confocal

analysis of hidden hearing loss in post-mortem tissue. *Hear Res* 327: 78–88, 2015.

Wagner L, **Plontke SK**, **Rahne T**. Perception of Iterated Rippled Noise Periodicity in

Cochlear Implant Users. *Audiol Neurotol* 22: 104–115, 2017.

Walls MK, **Race N**, **Zheng L**, **Vega-Alvarez SM**, **Acosta G**, **Park J**, **Shi R**. Structural and

biochemical abnormalities in the absence of acute deficits in mild primary blast-induced

head trauma. *J Neurosurg* 124: 675–686, 2016.

Walton JP. Timing is everything: Temporal processing deficits in the aged auditory

brainstem. *Hear Res* 264: 63–69, 2010.

Wang H, **Turner JG**, **Ling L**, **Parrish JL**, **Hughes LF**, **Caspary DM**. Age-related changes in

glycine receptor subunit composition and binding in dorsal cochlear nucleus.

Neuroscience 160: 227–239, 2009.

Wang X, **Lu T**, **Bendor D**, **Bartlett E**. Neural coding of temporal information in auditory

thalamus and cortex. *Neuroscience* 154: 294–303, 2008.

Vander Werff KR. Auditory Dysfunction Among Long-Term Consequences of Mild

Traumatic Brain Injury (mTBI). *Perspect Hear Hear Disord Res Diagnostics* 16: 3, 2012.

Yeend I, **Beach EF**, **Sharma M**. Working Memory and Extended High-Frequency Hearing

in Adults: Diagnostic Predictors of Speech-in-Noise Perception. *Ear Hear* 40: 458–467,

2019.

Yost WA. Pitch of iterated rippled noise. *J Acoust Soc Am* 100: 511–518, 1996a.

Yost WA. Pitch strength of iterated rippled noise. *J Acoust Soc Am* 100: 3329–3335,

1996b.

Fig 1. Auditory evoked potential experiment setup and examples of ABR waveforms. A)

Electrode placement and channel configuration of the study's auditory evoked potential

experiment. B) Examples of ABR waveforms at 80 dB SPL and 30 dB SL, with relevant wave

peaks labeled. The waves for which amplitudes are measured are labeled with a black

triangle.

Fig 2. ABR threshold changes of Blast (N=10) and Sham (N=6) rats during the first two

months post-exposure. Blast animals demonstrated drastic increases (worse) of Click, 8

kHz, and 16 kHz thresholds (red lines) post-exposure as opposed to Sham animals (blue

lines). Significant main effects ($p \leq 0.001$) of Groups and Group*Time interactions were

observed in all carriers. Significant Simple Main Effect of single time points observed in

various carriers throughout the two months. For subsequent figures, red lines will denote

blast-exposed animals, and blue lines will denote sham-exposed animals.

***Blast threshold significantly higher than Sham in Click, 8kHz, and 16kHz, $p<0.05$;

**Blast threshold significantly higher only in Click and 16 kHz;

*Blast threshold significantly higher only in Click.

1009

Fig 3. ABR wave I, III, and V amplitudes of Blast (N=10) and Sham (N=6) rats during the

first two months post-exposure expose persistent blast-induced differences at 80 dB SPL.

Significant main Group*Time interaction effects ($p \leq 0.001$) observed in waves I (left

column), III (center column), and V (right column) for all carriers: A) Click ABR; B) 8 kHz

ABR; C) 16 kHz ABR. Click ABR revealed blast-induced reduction of ABR wave amplitudes

to a greater degree than both tone ABRs. Later waves (Wave III and V) showed earlier

recovery in Blast animals.

*Significant Simple Main Effect of Group in ABR Wave Amplitudes, p<0.05.

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1019   **Fig 4. ABR wave I, III, and V amplitudes of Blast (N=10) and Sham (N=6) rats during the**

1020   **first two months post-exposure at 30 dB SL.** Similar format to Fig. 3. Significant main

1021   Group*Time interaction effects only observed with Click ABR waves (Wave I: p=0.016;

1022   Wave III: p=0.04; Wave V: p=0.003) A) Click ABR; B) 8 kHz ABR; C) 16 kHz ABR.

1023   *Significant Simple Main Effect of Group in ABR Wave Amplitudes, p<0.05.

1024

Fig 5. MLR waveforms and peak amplitudes of Blast (N=8) and Sham (N=6) rats during

the first two months post-exposure at 80 dB SPL and at 30 dB SL (Thresh + 30 dB). Grand

average traces of Click MLR waveforms at 80 dB SPL: A) Blast, pre-blast to day 7.

Arrowheads indicate measured peaks in E-H; B) Sham, pre-blast to day 7; C) Blast, day

10 to day 60; D) Sham, day 10 to day 60. Normalized Click MLR wave amplitudes over

time: E) Blast, 80 dB SPL; F) Sham, 80 dB SPL; G) Blast, 30 dB SL; H) Blast, 30 dB SL.

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Fig 6. AM depth stimuli and EFR responses from Blast (N=10) and Sham (N=6) rats during

the first two months post-exposure at 30 dB above threshold, in quiet. A) AM depth

stimulus waveforms at 100% and 50% modulation depths; B) NAM FFT amplitudes at 10

Hz (left), 45 Hz (center), and 256 Hz (right). Significant Group effect at 45Hz (p=0.007);

Similar format in C and D. C) SAM 8 kHz FFT amplitudes at 45 Hz show a steady yet

insignificant increase in later short-term (day 7-14); D) SAM 16k FFT amplitudes found

no significant Group effect.

**Significant Simple Main Effect of Group in FFT Amplitudes in both 100% depth and 50%

depth

* Significant Simple Main Effect of Group in FFT Amplitudes only in 100% depth

1042

Fig 7. AM noise stimuli and EFR responses from Blast (N=10) and Sham (N=6) rats during

the first two months post-exposure at 30 dB above threshold. A) AM noise stimulus

composition and waveform. B) Amplitude modulated noise carrier. FFT amplitudes at

signal modulation frequency in quiet and with 71 Hz AM noise masker level of 20SNR or

0SNR (equal) show significant Noise * Group effect for: B) NAM noise at 45 Hz (p=0.011)

modulation frequency; C) SAM 8 kHz noise at 10 Hz (p=0.001) and 45 Hz (p=0.015)

modulation frequency.

* Significant Simple Main Effect of Group

1051

Fig 8. IRN Chinese Tone stimuli and responses from Blast (N=8) and Sham (N=6) rats

during the first two months post-exposure at 30 dB above threshold A) Example

waveform and spectrogram of IRN Tone 2 stimulus; B) Examples of Peak Frequency of

IRN Evoked Potential in Pre-blast and Post-blast Brain (day 10 post-blast) from an

individual animal; C) Significant effect of Group (*) was seen in IRN Tone 2 (top, p=0.026)

and Tone 4 (bottom, p=0.029) pitch-tracking score, though Simple Main Effect of is

limited for individual time points; D) Cross-correlation of post-blast IRN responses to

corresponding pre-blast responses. Significant differences (*) in correlation coefficients

to pre-blast responses between Blast and Sham were observed in two waves: day 1-10,

and day 30 (p<0.05, paired sign-rank).

1062

1063 **Fig 9. Pitch tracking scores of responses to IRN Tone 2 stimuli with pitch salience**

1064 **controlled by altering iteration number at different time points, at 30 dB above threshold.**

1065 Though the effect of Iterations on pitch-tracking score was significant (p<0.001), no

1066 significant Iteration * Group interaction was observed.

1067

1068 **Table 1. Summary of 80 dB SPL ABR Wave I, III and V repeated measure ABR statistics.**

1069

1070 **Table 2. Summary of 30 dB SL ABR Wave I, III and V repeated measure ABR statistics.**

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1072 **Table 3. Simple main effects of blast group on ABR wave amplitudes over time.** Post-blast

1073 ABR amplitudes of Blast (N=10) and Sham (N=6) groups are compared using rmANOVA

1074 at each time point recorded. A p<0.05 showed significant simple main effect of Group at

1075 that time point.

1076

1077 **Table 4. Summary of post hoc tests against pre-blast ABR amplitudes.** Post-blast ABR

1078 amplitudes of Blast (N=10) and Sham (N=6) are compared against pre-blast amplitudes

1079 of the same group to show blast impact and recovery.

Table 1

80dB ABR			F	p	η^2_p
Click	Wave I	Time	9.980	**< .001**	0.416
		Group	23.080	**< .001**	0.622
		Time * Group	11.685	**< .001**	0.455
	Wave III	Time	11.065	**< .001**	0.441
		Group	17.207	**< .001**	0.551
		Time * Group	8.871	**< .001**	0.388
	Wave V	Time	14.134	**< .001**	0.502
		Group	23.203	**< .001**	0.624
		Time * Group	10.990	**< .001**	0.440
8 kHz	Wave I	Time	8.102	**< .001**	0.367
		Group	14.409	**0.002**	0.507
		Time * Group	11.760	**< .001**	0.457
	Wave III	Time	14.084	**< .001**	0.501
		Group	6.266	**0.025**	0.309
		Time * Group	13.826	**< .001**	0.497
	Wave V	Time	8.545	**< .001**	0.379
		Group	9.859	**0.007**	0.413
		Time * Group	9.346	**< .001**	0.400

16 kHz	Wave I	Time	9.845	**< .001**	0.413
		Group	27.486	**< .001**	0.663
		Time * Group	14.328	**< .001**	0.506
	Wave III	Time	9.845	**< .001**	0.413
		Group	13.048	**0.003**	0.482
		Time * Group	14.328	**< .001**	0.506
	Wave V	Time	8.838	**< .001**	0.387
		Group	20.528	**< .001**	0.595
		Time * Group	10.066	**< .001**	0.418

30dB SL ABR			F	p	η^2_p
Click	Wave I	Time	2.554	**0.019**	0.154
		Group	3.637	0.077	0.206
		Time * Group	2.630	**0.016**	0.158
	Wave III	Time	2.544	0.052	0.154
		Group	1.811	0.200	0.115
		Time * Group	2.720	0.040	0.163
	Wave V	Time	2.568	**0.018**	0.155
		Group	1.479	0.244	0.096
		Time * Group	3.429	**0.003**	0.197
8 kHz	Wave I	Time	3.146	**0.018**	0.183
		Group	6.852e -5	0.994	0.000
		Time * Group	0.200	0.945	0.014
	Wave III	Time	2.432	0.069	0.148
		Group	0.020	0.889	0.001
		Time * Group	1.184	0.328	0.078
	Wave V	Time	1.837	0.136	0.116
		Group	0.361	0.558	0.025
		Time * Group	0.618	0.648	0.042
16 kHz	Wave I	Time	2.325	**0.031**	0.142

	Group	1.046	0.324	0.070
	Time * Group	2.381	**0.027**	0.145
Wave III	Time	0.562	0.651	0.039
	Group	1.149	0.302	0.076
	Time * Group	1.930	0.136	0.121
Wave V	Time	1.160	0.338	0.077
	Group	4.905e -4	0.983	0.000
	Time * Group	1.646	0.161	0.105

Simple Main Effects			80 dB SPL			30dB SL		
		Time	Mean square	F	p	Mean Square	F	p
Click	Wave I	day1	13.942	31.329	< .001	2.097	8.019	0.013
		day4	12.438	30.917	< .001	1.843	7.730	0.015
		day7	4.081	18.418	< .001	0.024	0.102	0.754
		day10	1.601	7.023	0.019	0.020	0.127	0.726
		day14	3.180	10.008	0.007	0.596	3.044	0.103
		day30	1.517	8.285	0.012	0.184	1.829	0.198
		day60	1.013	4.393	0.055	0.102	0.632	0.440
	Wave III	day1	9.708	30.148	< .001	1.977	6.629	0.022
		day4	5.754	23.070	< .001	0.692	3.416	0.086
		day7	3.725	33.967	< .001	0.470	3.917	0.068
		day10	1.690	11.712	0.004	0.093	0.665	0.429
		day14	0.854	8.587	0.011	0.039	0.474	0.502
		day30	0.634	4.714	0.048	0.005	0.088	0.771
		day60	0.064	0.315	0.584	0.029	0.219	0.647
	Wave V	day1	17.134	45.066	< .001	3.596	9.213	0.009
		day4	14.672	35.376	< .001	2.178	8.580	0.011
		day7	4.419	13.382	0.003	0.056	0.132	0.722
		day10	2.264	8.415	0.012	0.012	0.043	0.839
		day14	4.709	16.223	0.001	0.104	0.347	0.565
		day30	1.214	3.651	0.077	0.009	0.028	0.870

		day60	0.414	1.155	0.301	0.002	0.007	0.934
8 kHz	Wave I	day1	7.136	45.959	**< .001**	0.000	0.004	0.948
		day4	6.915	37.954	**< .001**	0.001	0.014	0.909
		day7	1.892	15.704	**0.001**	0.017	0.551	0.470
		day10	0.754	2.693	0.123	0.014	0.159	0.696
		day14	0.803	4.657	**0.049**	0.010	0.096	0.762
		day30	0.421	2.532	0.134	0.001	0.015	0.904
		day60	0.285	1.562	0.232	0.006	0.450	0.513
	Wave III	day1	5.977	37.898	**< .001**	0.084	0.838	0.375
		day4	4.139	36.832	**< .001**	0.000	0.000	0.984
		day7	3.544	23.802	**< .001**	0.034	0.703	0.416
		day10	0.990	5.847	**0.030**	0.017	0.232	0.637
		day14	0.167	0.851	0.372	0.055	1.274	0.278
		day30	0.022	0.131	0.722	0.041	0.301	0.592
		day60	0.167	0.640	0.437	0.062	1.420	0.253
	Wave V	day1	6.374	100.269	**< .001**	0.020	0.135	0.719
		day4	4.459	28.229	**< .001**	0.005	0.165	0.691
		day7	1.580	11.078	**0.005**	0.129	3.798	0.072
		day10	0.392	2.043	0.175	0.104	1.256	0.281
		day14	0.449	3.269	0.092	0.001	0.010	0.924
		day30	0.006	0.026	0.875	0.018	0.149	0.705
		day60	0.017	0.060	0.811	0.007	0.187	0.672

16 kHz	Wave I	day1		5.090	40.130	**< .001**	0.205	2.470	0.138
		day4		5.910	61.374	**< .001**	0.026	0.827	0.378
		day7		2.397	29.278	**< .001**	0.090	2.321	0.150
		day10		0.723	4.602	0.050	0.009	0.305	0.590
		day14		1.766	24.888	**< .001**	0.009	0.153	0.701
		day30		0.515	6.551	**0.023**	0.000	0.000	0.992
		day60		0.205	2.242	0.156	0.022	1.009	0.332
	Wave III	day1		3.592	39.592	**< .001**	0.205	2.470	0.138
		day4		2.919	38.728	**< .001**	0.026	0.827	0.378
		day7		2.507	35.072	**< .001**	0.090	2.321	0.150
		day10		0.965	11.997	**0.004**	0.009	0.305	0.590
		day14		0.676	8.374	**0.012**	0.009	0.153	0.701
		day30		0.253	3.337	0.089	0.000	0.000	0.992
		day60		0.006	0.056	0.816	0.022	1.009	0.332
	Wave V	day1		4.772	37.802	**< .001**	0.118	1.617	0.224
		day4		3.414	44.163	**< .001**	0.000	0.005	0.945
		day7		2.150	21.095	**< .001**	0.066	1.575	0.230
		day10		0.902	7.214	**0.018**	0.000	0.002	0.961
		day14		1.283	15.210	**0.002**	0.017	0.336	0.571
		day30		0.100	0.888	0.362	0.103	1.301	0.273
		day60		0.019	0.293	0.597	0.079	1.541	0.235

			80dB SPL							30dB SL						
			day1	day4	day7	day10	day14	day30	day60	day1	day4	day7	day10	day14	day30	day60
Click	Wave I	Blast	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.001	0.244	1.000	1.000	1.000	0.300	0.808
		Sham	1.000	1.000	1.000	1.000	1.000	0.984	0.999	1.000	1.000	1.000	1.000	1.000	0.997	1.000
	Wave III	Blast	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.002	0.003	0.015	0.780	0.285	0.934	0.528	0.797
		Sham	1.000	0.916	1.000	0.915	0.902	0.838	0.600	1.000	1.000	1.000	0.966	0.767	0.854	0.652
	Wave V	Blast	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.009	0.440	1.000	1.000	1.000	1.000	1.000
		Sham	1.000	1.000	1.000	1.000	1.000	0.990	0.997	1.000	1.000	1.000	1.000	1.000	0.967	1.000
8 kHz	Wave I	Blast	<0.001	<0.001	<0.001	0.129	0.525	0.575	0.008	0.840	0.996	1.000	1.000	1.000	1.000	0.943
		Sham	0.978	1.000	0.998	1.000	1.000	1.000	0.972	0.957	0.999	1.000	1.000	1.000	1.000	1.000

		m														
	Wave III	Blast	**<0.001**	**<0.001**	**<0.001**	**<0.001**	**<0.001**	**<0.023**	0.878	0.998	1.000	1.000	1.000	1.000	1.000	1.000
		Sham	1.000	0.996	1.000	1.000	0.968	1.000	0.999	0.995	1.000	1.000	0.998	0.932	1.000	0.932
	Wave V	Blast	**<0.001**	**<0.001**	**<0.001**	0.993	0.994	1.000	1.000	0.902	1.000	1.000	1.000	1.000	1.000	1.000
		Sham	1.000	1.000	0.991	1.000	1.000	1.000	0.974	1.000	1.000	0.998	1.000	1.000	1.000	0.999
16 kHz	Wave I	Blast	**<0.001**	**<0.001**	**<0.001**	**<0.001**	0.050	**0.020**	**<0.001**	1.000	1.000	1.000	1.000	1.000	1.000	0.999
		Sham	1.000	0.993	0.997	0.965	1.000	1.000	0.149	0.296	0.999	1.000	1.000	1.000	1.000	0.973
	Wave III	Blast	**<0.001**	**<0.001**	**<0.001**	**<0.001**	**<0.001**	**<0.001**	**<0.001**	1.000	1.000	1.000	1.000	0.993	1.000	0.930
		Sham	1.000	0.990	0.973	0.534	0.805	0.838	0.203	1.000	0.982	1.000	0.994	0.880	0.938	0.890

Wave V	Blast	**<0.001**	**<0.001**	**<0.001**	**<0.001**	**0.031**	0.879	0.063	1.000	1.000	1.000	1.000	0.998	1.000	1.000
	Sham	1.000	1.000	0.899	0.520	1.000	0.915	**0.031**	1.000	1.000	1.000	1.000	1.000	0.986	0.843