A putative *erm* gene is present in *ΔermB Clostridioides difficile* isolates showing high levels

of resistance to clindamycin

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Abstract:

Clostridioides difficile has become the leading cause of hospital-acquired diarrhea.

Clindamycin is a member of the macrolide-lincosamide-streptogramin B family of protein

synthesis inhibitors. Been the ribosomal methylation the most widespread mechanism of

resistance to these antibiotics in *C difficile erm* genes usually produce high levels of

resistance to these drugs. In this short report we present evidence about the presence of

an unreported putative *erm* gen in *C difficile* isolates that despite presenting a *ΔermB*

genotype maintain high levels of clindamycin resistance.

Key words: *Clostridiodes difficile*, *erm*, clindamycin.

Introduction:

Clostridiodes difficile is a Gram-positive, sporulating and anaerobic bacterium that has become the leading cause of hospital-acquired diarrhea (Banawas, 2018; Guery *et al*, 2019). *C difficile* infection (CDI) is associated with the exposure of the normal intestinal microbiota to antibiotics. The resulting disruption of this normal intestinal microflora allows the multiplication and establishment of *C difficile* in the large intestine causing the pathology (Banawas, 2018; Guery *et al*, 2019).

Antibiotic resistance is crucial in spreading CDI among hospitalized patients, particularly in the older ones (Banawas, 2018). Antibiotics used for treatment of any type of infection could potentially promote (CDI) and resistant to multiple agents may have a selective advantage for the bacteria (Spigaglia, 2016). Most of the antibiotics associated with CDI appearance, including clindamycin, remaining to be associated with the highest risk for CDI (Spigaglia, 2016; Banawas, 2018).

Clindamycin and erythromycin are members of the macrolide-lincosamide-streptogramin B (MLSB) family of protein synthesis inhibitors. Ribosomal methylation is the most widespread mechanism of resistance to these antibiotics in *C difficile*. *erm* genes usually produce high levels of resistance to these drugs this by modifying the ribosomal 23S rRNA (Spigaglia, 2016; Banawas, 2018).

The *ermB* gene can be located in the transposon Tn5398, a mobilizable nonconjugative

element of 9.6 kb found in *C difficile* genomes and contains two copies of *ermB* (Spigaglia,

2016; Banawas, 2018). Isolates whit a truncated version of Tn5398, containing only one

copy of *ermB*, are called Δ*ermB* isolates and fail in inducing the MLS$_B$ resistance phenotype

(Hussain *et al*, 2005).

In this short report we present evidence about the presence of an unreported putative *erm*

gen in *C difficile* isolates that despite presenting a Δ*ermB* genotype maintain high levels of

clindamycin resistance.

Results:

Figure 1. Alignment of *erm* genes. Marked in red boxes are the amino acids associated whit

essential ribosome methylation activities.

The gene product analyzed showed a 96% of coverage and 26% identity when compared to

ermB from Tn5398. In addition compared to ermB this putative erm protein presents 4 of 6

essential amino acids necessary for ribosome methylation activity already described

(Farrow *et al*, 2002).

Table 1. Features of analyzed C difficile isolates.

Isolate	*cfrC*	Δ*ermB*	CLI MIC (µg/ml) *
LIBA-5707	Yes	Yes	>256
LIBA-5701	No	Yes	>256

*CLI MIC: clindamycin minimum inhibitory concentration from Ramírez-Vargas *et al*, 2017.

Lacking *cfrC* gene (another ribosome methylating enzyme) and having Δ*ermB* genotype,

LIBA-5701 is reported to be resistant to clindamycin (MIC >256 µg/ml) according to CLSI

guides (Ramirez-Vargas *et al*, 2017; Stojković *et al*, 2019).

Discussion:

Here we report the presence of a putative *erm* gen in isolates from Costa Rican Hospitals

recovered from CDI patients in the 2000s. The isolates showed similarly pathologic

capabilities than other known *C difficile* epidemic isolates like NAP1 (Quesada-Gómez *et al*,

2015; López-Ureña *et al*, 2016).

This putative *erm* gen presents the key amino acids that has been associated to the

ribosome methylation activity essential for the high levels of resistance to for MLS$_B$ agents

such as clindamycin in other erm proteins (Farrow *et al*, 2002). Mutagenesis showed that

changes in these amino acids result in lower minimum inhibitory concentration for

erythromycin, another agent belonging to the MLS$_B$ group (Farrow *et al*, 2002).

In Δ*ermB C difficile* isolates the MLS$_B$ resistance phenotype is lost (Hussain *et al*, 2005). So,

other resistance mechanism should explain the resistance to clindamycin LIBA-5701. As

reported before (Ramirez-Vargas *et al*, 2017; Stojković *et al*, 2019) lack of the element

containing *cfrC* did not result in changes in the clindamycin resistance. Using the CARD

database (Alcock *et al*, 2020) no other resistance genes associated to MLS$_B$ resistance

genotype were found (data not shown). At this point we cannot exclude the possibility of

other new resistance mechanisms not detected here and the methylation activity of the

putative erm gen product must be probe.

Materials and methods:

Raw Illumina SRA files for LIBA-5701 and LIBA-5700 used in this work can be found under

the accession numbers ERR467550 and ERR467555 respectively at the SRA database. The

genomes were assembly using EDENA (Hernandez *et al*, 2008) and annotated using the

RAST platform (Aziz, 2008). The predicted erm protein (supplementary material 1) was

aligned against ermB from Tn5398 in *C difficle* 630 genome (accession number

NZ_CP019870.1) using Genius platform (Kearse *et al*, 2012). Similarly, they were compared

using pBLAST (https://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE=Proteins) for determination

of identity and coverage between the sequences.

Authors declare no conflict of interest.

References:

Alcock, B. P., Raphenya, A. R., Lau, T., Tsang, K. K., Bouchard, M., Edalatmand, A., Huynh,

W., Nguyen, A. V., Cheng, A. A., Liu, S., Min, S. Y., Miroshnichenko, A., Tran, H. K., Werfalli,

R. E., Nasir, J. A., Oloni, M., Speicher, D. J., Florescu, A., Singh, B., Faltyn, M., … McArthur, A.

G. (2020). CARD 2020: antibiotic resistome surveillance with the comprehensive antibiotic

resistance database. *Nucleic acids research*, *48*(D1), D517–D525.

https://doi.org/10.1093/nar/gkz935

Aziz, R. K., Bartels, D., Best, A. A., DeJongh, M., Disz, T., Edwards, R. A., Formsma, K., Gerdes,

S., Glass, E. M., Kubal, M., Meyer, F., Olsen, G. J., Olson, R., Osterman, A. L., Overbeek, R. A.,

McNeil, L. K., Paarmann, D., Paczian, T., Parrello, B., Pusch, G. D., … Zagnitko, O. (2008). The

RAST Server: rapid annotations using subsystems technology. *BMC genomics*, *9*, 75.

https://doi.org/10.1186/1471-2164-9-75

Banawas S. S. (2018). *Clostridium difficile* Infections: A Global Overview of Drug Sensitivity

and Resistance Mechanisms. *BioMed research international*, *2018*, 8414257.

https://doi.org/10.1155/2018/8414257

Farrow, K. A., Lyras, D., Polekhina, G., Koutsis, K., Parker, M. W., & Rood, J. I. (2002).

Identification of essential residues in the Erm(B) rRNA methyltransferase of Clostridium

perfringens. *Antimicrobial agents and chemotherapy*, *46*(5), 1253–1261.

https://doi.org/10.1128/aac.46.5.1253-1261.2002

Guery, B., Galperine, T., & Barbut, F. (2019). *Clostridioides difficile*: diagnosis and

treatments. *BMJ (Clinical research ed.)*, *366*, l4609. https://doi.org/10.1136/bmj.l4609

Hernandez, D., François, P., Farinelli, L., Osterås, M., & Schrenzel, J. (2008). De novo

bacterial genome sequencing: millions of very short reads assembled on a desktop

computer. *Genome research*, *18*(5), 802–809. https://doi.org/10.1101/gr.072033.107

Hussain, H. A., Roberts, A. P., & Mullany, P. (2005). Generation of an erythromycin-sensitive

derivative of Clostridium difficile strain 630 (630Deltaerm) and demonstration that the

conjugative transposon Tn916DeltaE enters the genome of this strain at multiple sites.

Journal of medical microbiology, *54*(Pt 2), 137–141. https://doi.org/10.1099/jmm.0.45790-

0

Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton, S.,

Cooper, A., Markowitz, S., Duran, C., Thierer, T., Ashton, B., Meintjes, P., & Drummond, A.

(2012). Geneious Basic: an integrated and extendable desktop software platform for the

organization and analysis of sequence data. *Bioinformatics (Oxford, England)*, *28*(12), 1647–

1649. https://doi.org/10.1093/bioinformatics/bts199

Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton, S.,

Cooper, A., Markowitz, S., Duran, C., Thierer, T., Ashton, B., Meintjes, P., & Drummond, A.

(2012). Geneious Basic: an integrated and extendable desktop software platform for the

organization and analysis of sequence data. *Bioinformatics (Oxford, England)*, *28*(12), 1647–

1649. https://doi.org/10.1093/bioinformatics/bts199

López-Ureña, D., Quesada-Gómez, C., Montoya-Ramírez, M., del Mar Gamboa-Coronado,

M., Somogyi, T., Rodríguez, C., & Rodríguez-Cavallini, E. (2016). Predominance and high

antibiotic resistance of the emerging Clostridium difficile genotypes NAPCR1 and NAP9 in a

Costa Rican hospital over a 2-year period without outbreaks. *Emerging microbes &*

infections, *5*(5), e42. https://doi.org/10.1038/emi.2016.38

Quesada-Gómez, C., López-Ureña, D., Acuña-Amador, L., Villalobos-Zúñiga, M., Du, T.,

Freire, R., Guzmán-Verri, C., del Mar Gamboa-Coronado, M., Lawley, T. D., Moreno, E.,

Mulvey, M. R., de Castro Brito, G. A., Rodríguez-Cavallini, E., Rodríguez, C., & Chaves-Olarte,

E. (2015). Emergence of an outbreak-associated Clostridium difficile variant with increased

virulence. *Journal of clinical microbiology*, *53*(4), 1216–1226.

https://doi.org/10.1128/JCM.03058-14

Ramírez-Vargas, G., Quesada-Gómez, C., Acuña-Amador, L., López-Ureña, D., Murillo, T., Del

Mar Gamboa-Coronado, M., Chaves-Olarte, E., Thomson, N., Rodríguez-Cavallini, E., &

Rodríguez, C. (2017). A Clostridium difficile Lineage Endemic to Costa Rican Hospitals Is

Multidrug Resistant by Acquisition of Chromosomal Mutations and Novel Mobile Genetic

Elements. *Antimicrobial agents and chemotherapy*, *61*(4), e02054-16.

https://doi.org/10.1128/AAC.02054-16

Spigaglia P. (2016). Recent advances in the understanding of antibiotic resistance in

Clostridium difficile infection. *Therapeutic advances in infectious disease*, *3*(1), 23–42.

https://doi.org/10.1177/2049936115622891

Stojković, V., Ulate, M. F., Hidalgo-Villeda, F., Aguilar, E., Monge-Cascante, C., Pizarro-

Guajardo, M., Tsai, K., Tzoc, E., Camorlinga, M., Paredes-Sabja, D., Quesada-Gómez, C.,

Fujimori, D. G., & Rodríguez, C. (2019). *cfr*(B), *cfr*(C), and a New *cfr*-Like Gene, *cfr*(E), in

Clostridium difficile Strains Recovered across Latin America. *Antimicrobial agents and*

chemotherapy, *64*(1), e01074-19. https://doi.org/10.1128/AAC.01074-19

Suplemnetary material 1. Putative erm protein sequence in this study

MKNLGYSQNFLVNKKLVERLISKSNIDVTDYVIEIGPGKGIITDVLSQHAGEVVAVEYDQELYNNLVRYH

SHDNVTYIFGDFLKYKLPLNRRYKIFSNIPFQITADIIRKLTDDVNPPSDINIIIQREAAKKNCGIPLQKYEG

FRAAIIKAQYKVEITHSFKRSDFFPSPNVDTVMLHMQLWDDRLSGDDLQNYKDLVAFFYTNIKGETAK

ERLSILFSNEQIKRLGKANRISLSNSYTLITAEQWMNIYYYSKIGLTDEKKKKFQDAYKKLQKMNKKLKKQ

NRTSLRKSSSNKKCRTRIDTSNGTKNR