

1 **RangeShifter 2.0: An extended and enhanced platform for modelling spatial eco-**
2 **evolutionary dynamics and species' responses to environmental changes.**

3 Greta Bocedi^{1*}, Stephen C. F. Palmer¹, Anne-Kathleen Malchow^{2,3}, Damaris Zurell^{2,3}, Kevin
4 Watts^{4,5}, Justin M. J. Travis¹.

5 ¹School of Biological Sciences, University of Aberdeen, Zoology Building, Tillydrone Avenue,
6 Aberdeen, AB24 2TZ, UK

7 ²University of Potsdam, Institute of Biochemistry and Biology, Potsdam, Germany

8 ³Humboldt-Universität zu Berlin, Geography Department, Berlin, Germany

9 ⁴Forest Research, Alice Holt Lodge, Farnham, Surrey GU10 4LH, UK

10 ⁵Biological and Environmental Sciences, University of Stirling, Stirling FK9 4LA, UK

11 [*greta.bocedi@abdn.ac.uk](mailto:greta.bocedi@abdn.ac.uk)

12 **Running title:** RangeShifter 2.0

13

14 **Abstract**

15 **1.** Process-based models are becoming increasingly used tools for understanding how
16 species are likely to respond to environmental changes and to potential management options.

17 RangeShifter is one such modelling platform, which has been used to address a range of
18 questions including identifying effective reintroduction strategies, understanding patterns of
19 range expansion and assessing population viability of species across complex landscapes.

20 **2.** Here we introduce a new version, RangeShifter 2.0, which incorporates important new
21 functionality. It is now possible to simulate dynamics over user-specified, temporally changing
22 landscapes. Additionally, the genetic and evolutionary capabilities have been strengthened,
23 notably by introducing an explicit genetic modelling architecture, which allows for simulation
24 of neutral and adaptive genetic processes. Furthermore, emigration, transfer and settlement
25 rules can now all evolve, allowing for sophisticated simulation of the evolution of dispersal. We
26 illustrate the potential application of RangeShifter 2.0's new functionality by two examples.
27 The first illustrates the range expansion of a virtual species across a dynamically changing UK
28 landscape. The second demonstrates how the software can be used to explore the concept
29 of evolving connectivity in response to land-use modification, by examining how movement
30 rules come under selection over landscapes of different structure and composition.

31 **3.** RangeShifter 2.0 is built using object-oriented C++ providing computationally efficient
32 simulation of complex individual-based, eco-evolutionary models. The code has been
33 redeveloped to enable use across operating systems, including on high performance
34 computing clusters, and the Windows GUI has been enhanced. Furthermore, the recoding of
35 the package has supported the development of a new version running under the R platform,
36 RangeShiftR.

37 **4.** RangeShifter 2.0 will facilitate the development of in-silico assessments of how species will
38 respond to environmental changes and to potential management options for conserving or

39 controlling them. By making the code available open source, we hope to inspire further
40 collaborations and extensions by the ecological community.

41 **Keywords**

42 Animal movement, dynamic landscapes, individual-based modelling, connectivity, population
43 viability, distribution modelling, process-based modelling.

44

45 **Introduction**

46 Faced with an accelerating global biodiversity crisis caused by multiple interacting and often
47 anthropogenic environmental changes (Ceballos et al., 2015; Urban, 2015; IPBES, 2019),
48 biologists are striving to understand and predict how species will respond, in both ecological
49 and evolutionary terms, to these threats and to management interventions (Urban et al., 2016;
50 Urban, 2019). Policy makers, conservation biologists and land managers are relying more and
51 more on such predictions to manage biodiversity on multiple fronts, including protecting
52 threatened species, limiting invasive species, and targeting habitat restoration efforts to both
53 enhance in-situ conservation and promoting range shifting (IPBES, 2019). Process-based
54 models, also called dynamic or mechanistic models, have become increasingly popular
55 following many calls urging the ecological community to move beyond correlative approaches
56 towards models that explicitly incorporate the key processes underpinning eco-evolutionary
57 responses to environmental changes (Franklin, 2010; Huntley et al., 2010; Schurr et al., 2012;
58 Evans et al., 2013; Thuiller et al., 2013; Urban et al., 2016; Cabral et al., 2017; Connolly et al.
59 2017; Briscoe et al., 2019; Peterson et al., 2019). Several models and platforms are actively
60 being developed (e.g. Lurgi et al. 2015; Landguth et al. 2017; Okamoto & Amarasekare, 2018;
61 Schumaker & Brookes, 2018; Cotto et al. 2020; Kearney & Porter, 2020; Visintin et al., 2020),
62 benefits and shortcomings scrutinised (Dormann et al., 2012; Singer et al., 2016; Zurell et al.,
63 2016; Fordham et al., 2018; Johnston et al., 2019), and a promising variety of applications is
64 emerging (e.g. Synes et al., 2016).

65 RangeShifter is a process-based models that we initially developed (Bocedi, Palmer,
66 et al., 2014), in response to the many calls for moving towards integrated dynamic modelling
67 approaches. The main objective was to provide an individual-based, spatially-explicit
68 modelling platform that integrated population dynamics with sophisticated dispersal behaviour,
69 and that could be used for a variety of applications, from theory development to in-silico testing
70 of management interventions. Indeed, since its release, RangeShifter has been used in
71 studies addressing a range of issues, including testing the effectiveness of alternative

72 management interventions to improve connectivity and population persistence (Aben et al.,
73 2016; Henry et al., 2017), facilitating range expansion (Synes et al., 2015, 2020), improving
74 reintroduction success (Heikkinen et al., 2015; Ovenden et al., 2019), investigating range
75 dynamics of invasive (Fraser et al., 2015; Dominguez Almela et al., 2020) and recovering
76 species (Sun et al., 2016) and theoretically investigating how different traits and processes
77 affect rate of range expansion (Bocedi, Zurell et al. 2014; Henry et al., 2014; Barros et al.,
78 2016; Santini et al., 2016). RangeShifter has also been coupled with CRAFTY (Murray-Rust
79 et al., 2014), an agent-based model designed to explore the impact of land managers'
80 behaviours on land-use change, showing that, in the example context of predicting interactions
81 between crops and their pollinators in a changing agricultural landscape, models that integrate
82 ecological processes with land managers' behaviours, together with their interactions and
83 feed-backs can reveal important dynamics in land use change which might otherwise be
84 missed (Synes et al., 2019; Willemen et al., 2019).

85 Here, we present the new RangeShifter 2.0, which, among various additions and
86 improvements, includes two major novelties: the option for implementing temporally dynamic
87 landscapes and a module for the explicit modelling of neutral and adaptive genetics
88 (controlling dispersal traits). RangeShifter is written in C++; it has been completely recoded
89 from its original release following object-oriented programming principles and is now open
90 source, thus facilitating wider usage and enhancements by the ecological community.
91 Additionally, we provide a dedicated website (<https://rangeshifter.github.io/>) and updated
92 tutorials for learning to use RangeShifter, and a forum page for more effective communication
93 among users. In addition to the original and improved Windows graphical user interface (GUI),
94 RangeShifter can now be compiled to run in batch-mode on Linux computer clusters. Below
95 we briefly describe, and illustrate with examples, the two major additions of dynamic
96 landscapes and explicit genetics, while we refer to the RangeShifter 2.0 User Manual
97 (<https://github.com/RangeShifter/RangeShifter-software-and-documentation>) for smaller
98 changes and new features.

99 **Model enhancements**

100 DYNAMIC LANDSCAPES

101 Considering dynamically changing landscapes is crucial for scenario-based simulations (e.g.
102 climate change or land-used change scenarios), for implementing landscape processes
103 through time (e.g. ongoing habitat fragmentation) and for testing dynamic management
104 interventions accounting for time lags from their deployment (e.g. creating a new woodland)
105 to the realization of their full potential (Watts et al., 2020). In RangeShifter 2.0, the landscape
106 may be changed any number of times during a simulation, but always at the start of the year,
107 i.e. prior to reproduction. The changes may comprise any of: alterations to the habitat
108 structure; addition, removal or changes of patches in a patch-based model; and modifications
109 of the cost map when using the stochastic movement simulator (SMS; Palmer, Coulon, &
110 Travis, 2011).

111 EXPLICIT GENETICS

112 A new module is provided to define the genetic architecture of a species in a flexible and
113 explicit way. Individuals may carry one or more chromosomes, to which neutral loci and
114 adaptive loci controlling dispersal traits are mapped. It is possible to model unlimited neutral
115 markers, thus allowing tracking of population structure and neutral diversity, as well as
116 simulating spatial genetic patterns emerging from the interaction between demographic and
117 spatial processes, e.g. for in-silico applications of landscape genetics (Manel et al., 2003). The
118 dispersal traits have been extended to cover density-dependent emigration and settlement
119 reaction norms, which may optionally differ between the sexes. Additionally, if SMS is selected
120 as the movement model in the transfer phase, the parameters controlling directional
121 persistence and the dispersal bias and its decay (see the User Manual) can be modelled as
122 evolving traits. Each dispersal or movement trait can be controlled by a separate single
123 chromosome, akin to RangeShifter v1 (Bocedi, Palmer, et al., 2014), or through a highly
124 flexible mapping of traits to chromosomes, which enables the degree of linkage between traits

125 to be controlled and, optionally, pleiotropy to be incorporated, thus allowing for complex
126 genetic architectures underlying evolution of dispersal strategies (Saastamoinen et al., 2018).
127 The whole genome of each individual may be output in a separate file if required, e.g. for the
128 calculation of landscape genetic indices.

129 **Example applications**

130 EFFECTIVENESS OF WOODLAND CREATION STRATEGIES TO FACILITATE RANGE 131 EXPANSION

132 We illustrate the application of dynamic landscapes using the example of woodland creation
133 in a real UK landscape introduced by Synes et al. (2015, 2020), who compared the effects of
134 various realistic management scenarios for improving functional connectivity for a range of
135 exemplar virtual woodland species on both species' persistence in existing patches and range
136 expansion ability. They compared persistence and expansion rates under the management
137 scenarios with a baseline rate for the original landscape. However, as the landscape changes
138 were 'instant', i.e. the new habitat was assumed to be immediately fully suitable, the
139 differences they observed could be over-estimated, as newly planted woodland would in reality
140 take many decades to develop into the equivalent of existing woodland in terms of its suitability
141 as breeding habitat for many species (Watts et al., 2020). Rather, newly planted areas might
142 be expected firstly to provide increased structure which might aid movement of woodland
143 species, and then gradually increase in quality as breeding habitat as canopy cover develops.

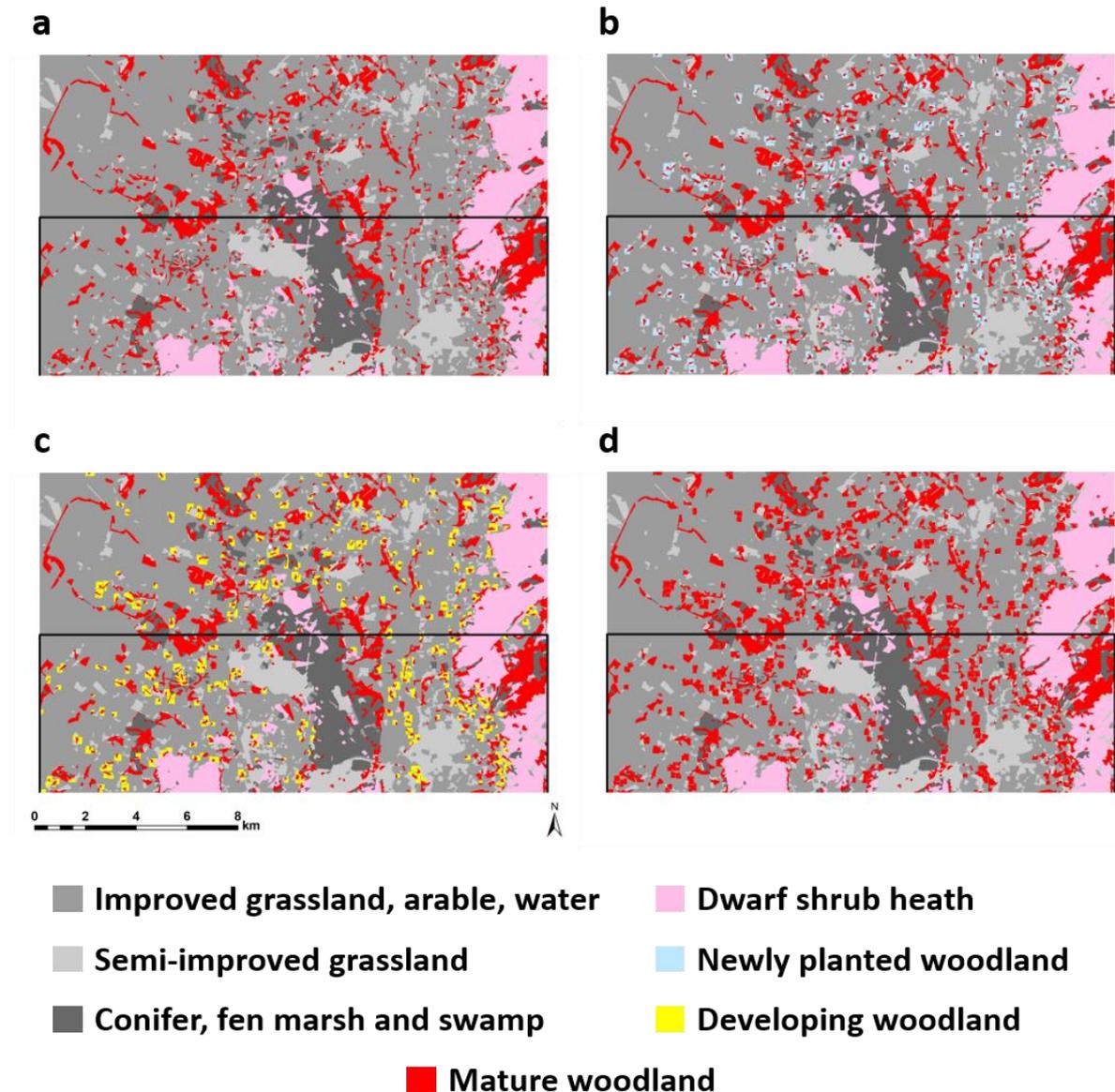
144 We assume as in Synes et al. (2015, 2020) that the locations of all new woodlands are
145 allocated immediately and on land previously used as improved grassland or arable, and that
146 planting of saplings occurs instantly in all locations. However, rather than instantly becoming
147 mature woodland habitat, planted areas develop gradually over a period of 40 years (Table 1,
148 Fig. 1). We compared the dynamic landscape approach with the 'instant' landscape approach
149 on the basis of the two most successful scenarios identified by Synes et al. (2015), namely
150 'CreateRandom' (new patches created anywhere) and 'CreateSmallAdjacent' (new planting to

151 increase the size of existing patches of under 3 ha), applied to 4% of the landscape. For
152 illustrative purposes we consider one virtual woodland species with simple sexual, stage-
153 structured demography and good dispersal abilities (Bird_D⁺P⁻S⁺ in Synes et al. 2015; see
154 Table S1 for the model parameters). We modelled dispersal movements through the
155 landscape with SMS. To ensure that the species was in equilibrium before management
156 commenced, we ran simulations for 50 years on the original landscape before applying the
157 first landscape change, and then continued for a further 100 years during which range
158 expansion was allowed to occur under the management scenario. Simulations were run on
159 the baseline landscape and on all the 10 replicate landscapes for each scenario generated by
160 Synes et al. (2015), and each simulation was replicated 10 times.

161 **Table 1.** Temporal development of newly planted woodland, where $1/b$ is the fecundity
 162 density-dependent coefficient (individuals / ha) (which largely determines the equilibrium
 163 density of the population), *Cost* is the perceived movement cost applied in modelling the
 164 transfer phase of dispersal by SMS and *Mort* is the per-step habitat-specific mortality
 165 probability.

Years	$1/b$	<i>Cost</i>	<i>Mort</i>	Description
0 - 4	0	100	0.05	Permeable habitat – the planting of new woodland provides enhanced movement potential (with lower movement costs). However, the limited canopy cover has no impact on mortality or breeding potential: <i>treat as similar to semi-improved grassland</i>
5 - 9	0	10	0.005	Permeable and safe habitat – as the new woodland develops, it provides increased structural cover for movement and reduced mortality, but it is still too young to provide suitable breeding habitat: <i>treat as dwarf shrub heath</i>
10 - 19	5	5	0.002	Low quality breeding habitat – the new woodland starts to provide a degree of reduced quality breeding habitat and settlement starts to occur, but there are still movement costs and a small mortality risk
20 - 29	10	2	0.001	Medium quality breeding habitat – movement and mortality risks decrease further as canopy cover develops and breeding quality is enhanced
30 - 39	15	1	0.0	High quality breeding habitat – further canopy closure removes movement costs and mortality risk and patch quality moves towards that of established woodland
40 - 100	21	1	0.0	Very high quality breeding habitat – optimal high quality woodland habitat is achieved in year 40 as previously Synes et al. (2015) implemented from year 0

166



167

168 **Figure 1.** Example of dynamic landscape development: (a) initial landscape, (b) after 5 years
169 when newly planted woodland adjacent to small patches is treated as dwarf shrub heath for
170 dispersal modelling, (c) after 20 years as canopy closure develops, (d) final landscape after
171 40 years when newly planted woodland is fully mature. The black line shows the northern limit
172 of the initial range.

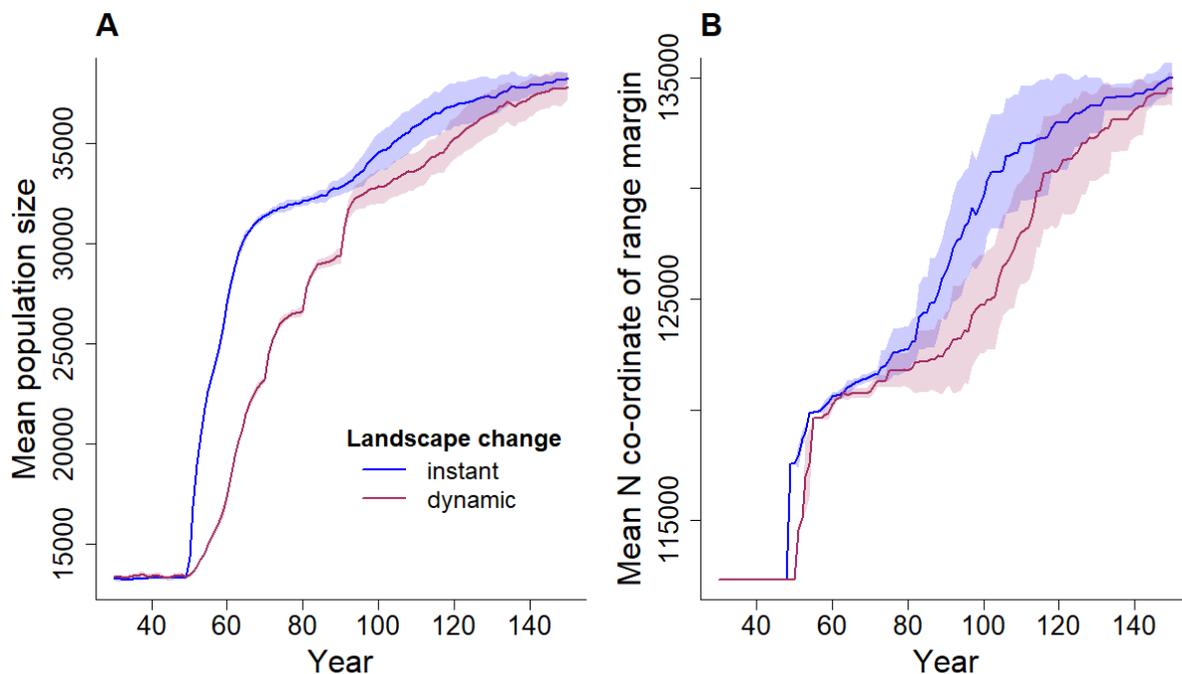
173

174

175

For the instant landscape approach (Synes et al. 2015), the mean rate of range expansion for the CreateRandom scenario was 187 m/year over 100 years (standard error s.e. 2.34 m/year), 2.0 times the rate on the baseline landscape. Similarly, for the

176 CreateSmallAdjacent scenario, the mean rate of range expansion was 201 m/year
177 (s.e. 2.90 m/year), 2.1 times faster than the baseline. By applying the dynamic landscape
178 approach to the CreateRandom scenario, the mean rate of range expansion was reduced
179 negligibly to 184 m/year (s.e. 2.64 m/year; relative reduction 1.6%). In contrast, for the
180 CreateSmallAdjacent scenario, the mean rate of range expansion was increased slightly to
181 216 m/year (s.e. 2.69 m/year; relative increase 7.5%). Despite rather similar total expansion
182 rates over a period of 100 years, the temporal trajectories differed considerably between the
183 instant and the dynamic landscape approach, as is illustrated for a single landscape replicate
184 of the CreateSmallAdjacent scenario (Fig. 2). The total population size on the dynamic
185 landscape lagged behind that on the instant landscape by up to 25% during the first 40 years
186 after planting (Fig. 2A), and the location of the northern range margin on the dynamic
187 landscape was up to 5 km further south during the succeeding 40 years (Fig. 2B).



188

189 **Figure 2.** Consideration of dynamic landscape restoration affects predictions on species'
190 range expansion dynamics. (A) Mean total population size and (B) mean location of species'
191 northern range margin for the instant (blue) and dynamic (red) landscape change methods

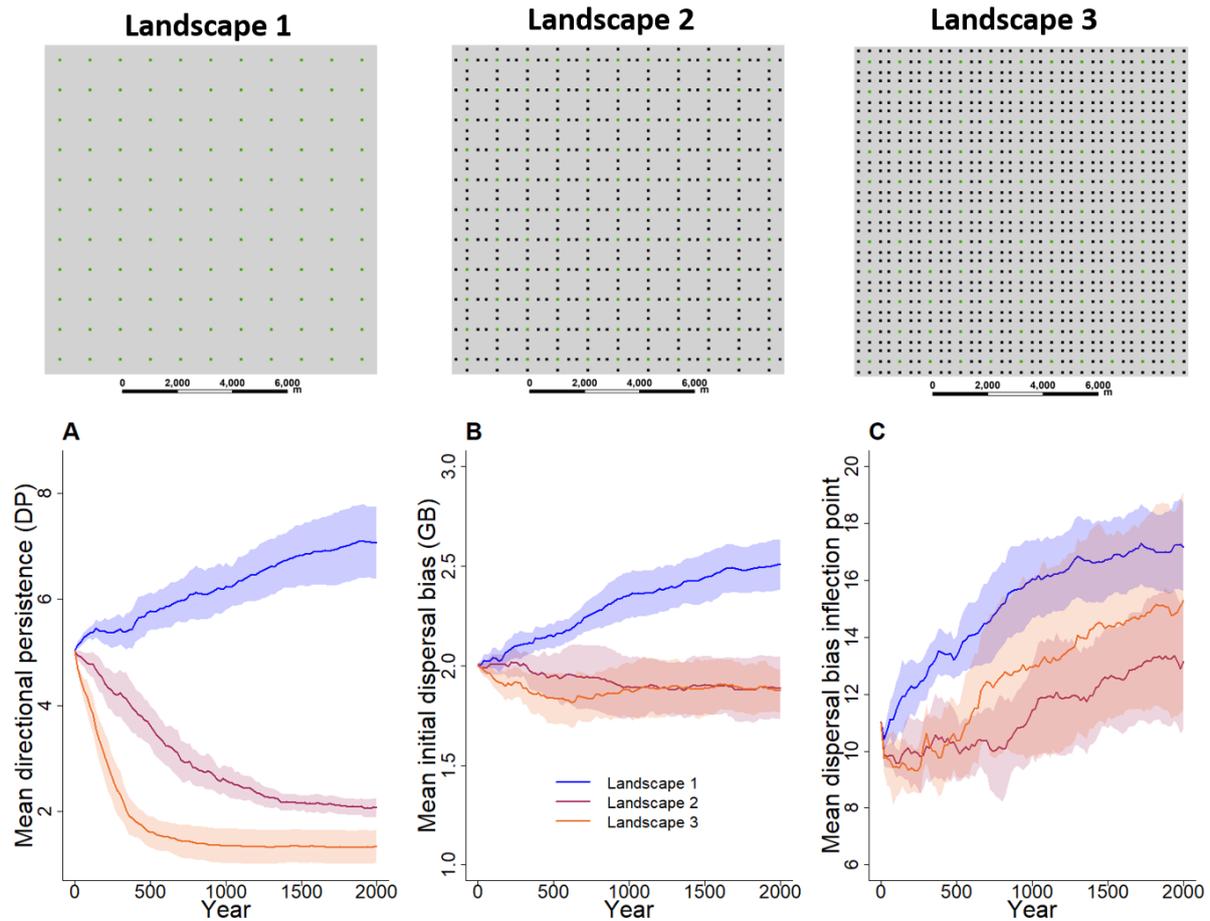
192 commencing at year 50 for a single landscape replicate of the CreateSmallAdjacent scenario.
193 Shades show 95% confidence intervals from 10 replicates.

194 EVOLUTION OF MULTIPLE DISPERSAL TRAITS

195 We illustrate how RangeShifter 2.0 can be used to model evolution of complex dispersal
196 strategies, which involve evolution of multiple traits defining all three phases of dispersal
197 (emigration, transfer through the landscape and settlement in a new habitat patch) on
198 landscapes that differ in their structure and composition. We modelled the evolution of
199 dispersal traits of an annual sexual species on a set of three stylised landscapes of 121 rows
200 x 121 columns differing in the degree to which movement was inhibited by the presence of
201 high-cost cells in the landscape (Fig. 3). Temporally uncorrelated local environmental
202 stochasticity was applied in two forms in order to promote dispersal evolution: as annual
203 variability in carrying capacity and as a small probability of local patch extinction. The
204 parameters controlling all three phases of dispersal, namely emigration (density-dependent),
205 transfer and settlement (density-dependent), evolved independently of one another, each trait
206 being determined by a separate autosome having three unlinked loci. Emigration and
207 settlement traits were modelled as sex-dependent, thus having sex-limited phenotypic
208 expression. Further, sex differences were assumed in settlement: while for both sexes
209 settlement probability could evolve density dependence, males had the additional fixed
210 settlement condition of requiring the presence of a mate in the patch.

211 The transfer phase of dispersal was modelled using SMS, which models the dispersal
212 trajectory on a stepwise basis whilst accounting for perceived movement costs and a tendency
213 to follow a correlated path, as determined by the directional persistence (DP) and the dispersal
214 bias parameters. The dispersal bias determines the tendency of moving in a straight line away
215 from the natal patch and is subject to a decay in strength as a function of the accumulated
216 number of steps taken (see Supplementary Information). DP and the parameters defining the
217 decay function of the dispersal bias were modelled as evolving traits, thus allowing for

218 evolution of movement rules. In the baseline Landscape 1 (Fig. 3), there were no inhibitory
219 cells in the matrix (Cost = 10; per-step mortality = 0.01), and therefore we would expect
220 relatively straight movement to evolve (Zollner & Lima, 1999). However, in Landscape 2, it is
221 much less clear what would be the best movement strategy, as the orthogonal paths between
222 patches are inhibited by high-cost cells (Cost = 1000; per-step mortality = 0.5), whereas the
223 diagonal movements are not. Finally, in Landscape 3, both orthogonal and diagonal paths are
224 impeded, and dispersers must therefore evolve strategies to reach a new patch whilst avoiding
225 as much as possible the high-risk inhibitory cells. We ran ten replicate simulations of 2000
226 years on each landscape. All model parameters and initialisation conditions are reported in
227 Table S2. Equations defining reaction norms are also reported in the Supplementary
228 Information.



229

230 **Figure 3.** Stylised landscapes used to model evolution of dispersal traits (upper panels).

231 Landscape 1 comprises evenly distributed breeding habitat patches of 100 m x 100 m (green)

232 set in a homogenous matrix (grey). In Landscape 2 high-cost cells inhibitory to movement

233 (black) are added orthogonally between the patches. In Landscape 3 additional inhibitory cells

234 are added to the diagonals between patches. **(A-C)** Evolution of mean transfer traits,

235 directional persistence (A), initial dispersal bias (B) and dispersal bias inflection point

236 (measured in steps taken; C), on the three landscapes. Phenotypic values are averaged over

237 all individuals and 10 replicate simulations.

238 As expected from the spatial and temporal configuration of the selective environments,

239 the dispersal strategies that evolved on the different landscapes differed mainly in their

240 movement rules (Fig. 3A-C), whereas they evolved similar reaction norms for the emigration

241 and settlement phases (Figs. S1-S2, S4-S5). In the absence of inhibitory features in

242 Landscape 1, very straight movement trajectories evolved (Fig. S6A): both mean directional
243 persistence (DP) and mean dispersal bias (the tendency to move in a direction away from the
244 natal patch) reached high values of ~ 7.0 and 2.5 respectively after 2000 years (Fig. 3A-B),
245 and indeed there was some indication that they were still increasing slightly. In contrast, when
246 orthogonal movement became inhibited in Landscape 2, much less direct movement evolved
247 (Fig. S6B), as determined by low mean DP (Fig. 3A). Mean dispersal bias initially remained
248 relatively high at around 2.0 (Fig. 3B), but its mean inflection point (the number of steps at
249 which dispersal bias decreases most rapidly) decreased from around 16 steps on
250 Landscape 1 to around 12 steps (Fig. 3C). Thus, dispersers having evolved in Landscape 2
251 would be following a much less straight path sooner after having left the natal patch compared
252 to dispersers having evolved in the more benign Landscape 1 (Fig. S3), thereby enabling them
253 to respond to the appearance of a suitable (low cost) cell within the perceptual range by
254 moving towards it. The addition of inhibitory features to diagonal movement further developed
255 this trend: dispersal bias altered little, but DP decreased to a very low level of around 1.3 on
256 average (Fig. 3A).

257 Emigration probability generally evolved to be male-biased. Mean male emigration
258 probability decreased as the occurrence of inhibitory cells in the landscape increased
259 (Landscapes 2 and 3) because the cost of dispersal effectively increased (Fig. S1-S2). Male-
260 biased emigration would be expected, given the loosely polygynous mating system (i.e. males
261 can mate with multiple females but each female mates only with one male) and the high
262 environmental and demographic stochasticity (Table S2), which increase between-patch
263 variance in male reproductive success (Henry et al., 2016; Li & Kokko, 2019). Density-
264 dependent settlement evolved similarly in the two sexes, so that individuals were likely to settle
265 at the first suitable patch encountered unless it was substantially above carrying capacity
266 (Fig. S4-S5).

267 **Discussion**

268 RangeShifter 2.0 provides enhancements and substantial extensions to the RangeShifter
269 software (Bocedi, Palmer, et al., 2014) expanding its potential range of applications. The
270 flexible and spatially-explicit demography and dispersal modules that are distinctive of this
271 platform (Lurgi et al., 2015) can be now combined with a flexible genetically-explicit
272 representation of neutral markers and/or multiple dispersal traits, allowing for diverse
273 applications focussed on combining population genetic processes with ecological and
274 environmental processes (Manel et al., 2003; Epperson et al., 2010) and accounting for
275 evolution of complex and multi-trait dispersal strategies (Cote et al., 2017; Legrand et al.,
276 2017; Saastamoinen et al., 2018). This is further combined with the ability of incorporating
277 dynamic landscapes, enabling applications that explicitly aim to predict species' genetic,
278 ecological and evolutionary responses to ongoing environmental changes. Such applications
279 include in-silico testing of management interventions which need to account for the occurrence
280 of ecological time-lags when targeting and evaluating conservation actions (Watts et al.,
281 2020).

282 Importantly, and in contrast with the previous release, RangeShifter 2.0 source code
283 is now open source (<https://github.com/rangeshifter>), published under the GNU general public
284 license (GPLv3). It is hence free for the wider community to use, modify and share.
285 Furthermore, RangeShifter 2.0 is also the core of the new package RangeShiftR (Malchow et
286 al., 2020), which allows running RangeShifter from the R environment (R Core Team, 2020)
287 while maintaining the high performance of the C++ code, and includes functions assisting with
288 the set-up of the simulations, the parameterisation and output analyses. RangeShiftR, in
289 addition to improving and broadening RangeShifter accessibility, makes it easily available for
290 multiple platforms, has access to R's infrastructure for parallel and cluster computing and
291 offers many opportunities for interoperation with other R packages.

292 RangeShifter 2.0 additionally comes with an enhanced Windows graphical user
293 interface (GUI) as freeware (<https://github.com/RangeShifter/RangeShifter-GUI>). From
294 current users, and from workshops that we are running worldwide, we are able to appreciate

295 the value of the RangeShifter GUI: it is particularly useful for non-modellers to explore eco-
296 evolutionary dynamics and their conservation implications, to recognise data gaps in empirical
297 systems, to communicate with stakeholders, and for teaching purposes across grades. It also
298 provides easily accessible and free software for countries with little funding for conservation
299 and research. Further, to improve accessibility, the User Manual has now been translated into
300 Spanish (<https://github.com/RangeShifter/RangeShifter-software-and-documentation>).

301 RangeShifter is in continuous development, and there are key areas for future
302 progress, which we hope, by making it open source and integrating it with R, will be addressed
303 by a common effort to move towards a fully-integrated dynamic platform that includes all the
304 key and necessary processes for predicting species' eco-evolutionary responses to global
305 changes. For example, RangeShifter 2.0 currently remains a single-species model, while
306 inter-specific interactions are often key in determining species' persistence to global changes
307 (Gilman et al., 2010; Norberg et al., 2012; Urban et al., 2012; Urban et al., 2019; Bocedi et al.,
308 2013; Svenning et al., 2014; Thompson & Fronhofer, 2019). Although we made a first
309 important step in including explicit genetics, and we are actively prioritising this area of
310 development, RangeShifter 2.0 does not yet include the level of sophistication that
311 characterises much forward-time population genetic software (Guillaume & Rougemont, 2006;
312 Haller & Messer, 2019), in terms of genetic processes, structures and outputs, and adaptive
313 traits. For example, the possibility of modelling adaptation to multiple environmental variables
314 will be a crucial addition. However, RangeShifter 2.0 holds an advantage in terms of the
315 ecological, demographic and dispersal complexity it can represent, which, combined with
316 explicit genetics, opens possibilities for sophisticated landscape genetics applications and for
317 fully accounting for evolution of dispersal behaviours (not just emigration rates) which are likely
318 to be critical for species' inhabiting or moving through complex, human-modified landscapes.

319 **Acknowledgments**

320 Development of RangeShifter 2.0 was supported by the project PROBIS funded by the
321 BiodivERsA European Joint Call 2012-2013. We thank R. L. Allgayer, A. Ponchon and N. W.
322 Synes for their help and contribution to the RangeShifter development and application. We
323 also thank the many users of RangeShifter and participants to workshops for their invaluable
324 feedback. GB was supported by a Royal Society University Research Fellowship. AKM and
325 DZ were supported by Deutsche Forschungsgemeinschaft (DFG) under grant agreement No.
326 ZU 361/1-1.

327 **Author's contributions**

328 GB, SCFP and JMJT mainly developed the model structure, implemented the C++ core code,
329 developed the GUI and wrote the documentation. AM contributed to code testing and cleaning,
330 and DZ developed the accompanying website. KW provided the landscape data for the first
331 example. GB, SCFP and JMJT wrote the initial manuscript and all authors contributed critically
332 to the drafts and gave final approval for publication.

333 **Data Availability**

334 RangeShifter C++ core code for the batch implementation is open source under the GNU
335 general public license (GPLv3). The code, as well as the compiled Windows package, the
336 User Manual and the data for the tutorials are available from GitHub
337 <https://github.com/RangeShifter>.

338 **References**

339 Aben, J., Bocedi, G., Palmer, S. C. F., Pellikka, P., Strubbe, D., Hallmann, C., ... Matthysen,
340 E. (2016). The importance of realistic dispersal models in conservation planning:
341 application of a novel modelling platform to evaluate management scenarios in an
342 Afrotropical biodiversity hotspot. *Journal of Applied Ecology*, 53(4), 1055–1065.
343 doi:10.1111/1365-2664.12643

- 344 Barros, C., Palmer, S. C. F., Bocedi, G., & Travis, J. M. J. (2016). Spread rates on
345 fragmented landscapes: the interacting roles of demography, dispersal and habitat
346 availability. *Diversity and Distributions*, 22(12), 1266–1275. doi:10.1111/ddi.12487
- 347 Bocedi, G., Atkins, K. E., Liao, J., Henry, R. C., Travis, J. M. J., & Hellmann, J. J. (2013).
348 Effects of local adaptation and interspecific competition on species' responses to
349 climate change. *Annals of the New York Academy of Sciences*, 1297, 83–97.
350 doi:10.1111/nyas.12211
- 351 Bocedi, G., Palmer, S. C. F., Pe'er, G., Heikkinen, R. K., Matsinos, Y. G., Watts, K., &
352 Travis, J. M. J. (2014). RangeShifter: a platform for modelling spatial eco-
353 evolutionary dynamics and species' responses to environmental changes. *Methods*
354 *in Ecology and Evolution*, 5(4), 388–396. doi:10.1111/2041-210X.12162
- 355 Bocedi, G., Zurell, D., Reineking, B., & Travis, J. M. J. (2014). Mechanistic modelling of
356 animal dispersal offers new insights into range expansion dynamics across
357 fragmented landscapes. *Ecography*, 37, 1240–1253. doi:10.1111/ecog.01041
- 358 Briscoe, N. J., Elith, J., Salguero-Gómez, R., Lahoz-Monfort, J. J., Camac, J. S., Giljohann,
359 K. M., ... Guillera-Aroita, G. (2019). Forecasting species range dynamics with
360 process-explicit models: matching methods to applications. *Ecology Letters*, 22(11),
361 1940–1956. doi:10.1111/ele.13348
- 362 Cabral, J. S., Valente, L., & Hartig, F. (2017). Mechanistic simulation models in
363 macroecology and biogeography: state-of-art and prospects. *Ecography*, 40(2).
364 doi:10.1111/ecog.02480
- 365 Ceballos, G., Ehrlich, P. R., Barnosky, A. D., García, A., Pringle, R. M., & Palmer, T. M.
366 (2015). Accelerated modern human-induced species losses: Entering the sixth mass
367 extinction. *Science Advances*, 1(5), e1400253. doi:10.1126/sciadv.1400253
- 368 Connolly, S. R., Keith, S. A., Colwell, R. K., & Rahbek, C. (2017). Process, mechanism, and
369 modeling in macroecology. *Trends in Ecology & Evolution*, 32(11), 835–844.
370 doi:10.1016/j.tree.2017.08.011

- 371 Cote, J., Bestion, E., Jacob, S., Travis, J., Legrand, D., & Baguette, M. (2017). Evolution of
372 dispersal strategies and dispersal syndromes in fragmented landscapes. *Ecography*,
373 *40*, 56–73. doi:10.1111/ecog.02538
- 374 Cotto, O., Schmid, M., & Guillaume, F. (2020). Nemo-age: Spatially explicit simulations of
375 eco-evolutionary dynamics in stage-structured populations under changing
376 environments. *Methods in Ecology and Evolution*. doi:10.1111/2041-210X.13460
- 377 Dominguez Almela, V., Palmer, S. C. F., Gillingham, P. K., Travis, J. M. J., & Britton, J. R.
378 (2020). Integrating an individual-based model with approximate Bayesian
379 computation to predict the invasion of a freshwater fish provides insights into
380 dispersal and range expansion dynamics. *Biological Invasions*, *22*(4), 1461–1480.
381 doi:10.1007/s10530-020-02197-6
- 382 Dormann, C. F., Schymanski, S. J., Cabral, J., Chuine, I., Graham, C., Hartig, F., ... Singer,
383 A. (2012). Correlation and process in species distribution models: bridging a
384 dichotomy. *Journal of Biogeography*, *39*(12), 2119–2131. doi:10.1111/j.1365-
385 2699.2011.02659.x
- 386 Epperson, B. K., McRae, B. H., Scribner, K., Cushman, S. A., Rosenberg, M. S., Fortin, M.-
387 J., ... Dale, M. R. T. (2010). Utility of computer simulations in landscape genetics.
388 *Molecular Ecology*, *19*(17), 3549–64. doi:10.1111/j.1365-294X.2010.04678.x
- 389 Evans, M. R., Bithell, M., Cornell, S. J., Dall, S. R. X., Díaz, S., Emmott, S., ... Morecroft, M.
390 (2013). Predictive systems ecology. *Proceedings of the Royal Society B*, *280*,
391 20131452.
- 392 Fordham, D. A., Bertelsmeier, C., Brook, B. W., Early, R., Neto, D., Brown, S. C., ... Araújo,
393 M. B. (2018). How complex should models be? Comparing correlative and
394 mechanistic range dynamics models. *Global Change Biology*, *24*(3), 1357–1370.
395 doi:10.1111/gcb.13935
- 396 Franklin, J. (2010). Moving beyond static species distribution models in support of
397 conservation biogeography. *Diversity and Distributions*, *16*(3), 321–330.
398 doi:10.1111/j.1472-4642.2010.00641.x

- 399 Fraser, E. J., Lambin, X., Travis, J. M. J., Harrington, L. A., Palmer, S. C. F., Bocedi, G., &
400 Macdonald, D. W. (2015). Range expansion of an invasive species through a
401 heterogeneous landscape - the case of American mink in Scotland. *Diversity and*
402 *Distributions*, 21(8), 888–900. doi:10.1111/ddi.12303
- 403 Gilman, S. E., Urban, M. C., Tewksbury, J., Gilchrist, G. W., & Holt, R. D. (2010). A
404 framework for community interactions under climate change. *Trends in Ecology &*
405 *Evolution*, 25(6), 325–31. doi:10.1016/j.tree.2010.03.002
- 406 Guillaume, F., & Rougemont, J. (2006). Nemo: an evolutionary and population genetics
407 programming framework. *Bioinformatics*, 22(20), 2556–7.
408 doi:10.1093/bioinformatics/btl415
- 409 Haller, B. C., & Messer, P. W. (2019). SLiM 3: Forward Genetic Simulations Beyond the
410 Wright-Fisher Model. *Molecular Biology and Evolution*, 36(3), 632–637.
411 doi:10.1093/molbev/msy228
- 412 Heikkinen, R. K., Pöyry, J., Virkkala, R., Bocedi, G., Kuussaari, M., Schweiger, O., ... Travis,
413 J. M. J. (2015). Modelling potential success of conservation translocations of a
414 specialist grassland butterfly. *Biological Conservation*, 192, 200–206.
415 doi:10.1016/j.biocon.2015.09.028
- 416 Henry, R. C., Bocedi, G., Dytham, C., & Travis, J. M. J. (2014). Inter-annual variability
417 influences the eco-evolutionary dynamics of range-shifting. *PeerJ*, 1, e228.
418 doi:10.7717/peerj.228
- 419 Henry, R. C., Coulon, A., & Travis, J. M. J. (2016). The evolution of male-biased dispersal
420 under the joint selective forces of inbreeding load and demographic and
421 environmental stochasticity. *The American Naturalist*, 188(4), 423–433.
422 doi:10.1086/688170
- 423 Henry, R. C., Palmer, S. C. F., Watts, K., Mitchell, R. J., Atkinson, N., & Travis, J. M. J.
424 (2017). Tree loss impacts on ecological connectivity: Developing models for
425 assessment. *Ecological Informatics*, 42, 90–99. doi:10.1016/j.ecoinf.2017.10.010

- 426 Huntley, B., Barnard, P., Altwegg, R., Chambers, L., Coetzee, B. W. T., Gibson, L., ... Willis,
427 S. G. (2010). Beyond bioclimatic envelopes: dynamic species' range and abundance
428 modelling in the context of climatic change. *Ecography*, 33(October 2009), 621–626.
429 doi:10.1111/j.1600-0587.2009.06023.x
- 430 IPBES. (2019). *IPBES (2019): Global assessment report on biodiversity and ecosystem*
431 *services of the Intergovernmental Science-Policy Platform on Biodiversity and*
432 *Ecosystem Services*. Bonn, Germany.
- 433 Johnston, A. S. A., Boyd, R. J., Watson, J. W., Paul, A., Evans, L. C., Gardner, E. L., &
434 Boulton, V. L. (2019). Predicting population responses to environmental change from
435 individual-level mechanisms: towards a standardized mechanistic approach.
436 *Proceedings of the Royal Society B: Biological Sciences*, 286, 20191916.
437 doi:10.1098/rspb.2019.1916
- 438 Kearney, M. R., & Porter, W. P. (2020). NicheMapR – an R package for biophysical
439 modelling: the ectotherm and Dynamic Energy Budget models. *Ecography*, 43(1),
440 85–96. doi:10.1111/ecog.04680
- 441 Landguth, E. L., Bearlin, A., Day, C. C., & Dunham, J. (2017). CDMetaPOP: an individual-
442 based, eco-evolutionary model for spatially explicit simulation of landscape
443 demogenetics. *Methods in Ecology and Evolution*, 8(1), 4–11. doi:10.1111/2041-
444 210X.12608
- 445 Legrand, D., Cote, J., Fronhofer, E. A., Holt, R. D., Ronce, O., Schtickzelle, N., ... Clobert, J.
446 (2017). Eco-evolutionary dynamics in fragmented landscapes. *Ecography*, 40(1), 9–
447 25. doi:10.1111/ecog.02537
- 448 Li, X. Y., & Kokko, H. (2019). Sex-biased dispersal: a review of the theory. *Biological*
449 *Reviews*, 94(2), 721–736. doi:10.1111/brv.12475
- 450 Lurgi, M., Brook, B. W., Saltré, F., & Fordham, D. A. (2015). Modelling range dynamics
451 under global change: Which framework and why? *Methods in Ecology and Evolution*,
452 6(3), 247–256. doi:10.1111/2041-210X.12315

- 453 Malchow, A.-K., Bocedi, G., Palmer, S. C. F., Travis, J. M. J., & Zurell, D. (2020).
454 RangeShiftR: an R package for individual-based simulation of spatial eco-
455 evolutionary dynamics and species' responses to environmental change. *BioRxiv*,
456 2020.11.17.384545. doi:10.1101/2020.11.17.384545
- 457 Manel, S., Schwartz, M. K., Luikart, G., & Taberlet, P. (2003). Landscape genetics:
458 Combining landscape ecology and population genetics. *Trends in Ecology and*
459 *Evolution*, 18(4), 189–197. doi:10.1016/S0169-5347(03)00008-9
- 460 Murray-Rust, D., Brown, C., van Vliet, J., Alam, S. J., Robinson, D. T., Verburg, P. H., &
461 Rounsevell, M. (2014). Combining agent functional types, capitals and services to
462 model land use dynamics. *Environmental Modelling and Software*, 59, 187–201.
463 doi:10.1016/j.envsoft.2014.05.019
- 464 Norberg, J., Urban, M. C., Vellend, M., Klausmeier, C. a., & Loeuille, N. (2012). Eco-
465 evolutionary responses of biodiversity to climate change. *Nature Climate Change*,
466 2(10), 747–751. doi:10.1038/nclimate1588
- 467 Okamoto, K. W., & Amarasekare, P. (2018). A framework for high-throughput eco-
468 evolutionary simulations integrating multilocus forward-time population genetics and
469 community ecology. *Methods in Ecology and Evolution*, 9(3), 525–534.
470 doi:10.1111/2041-210X.12889
- 471 Ovenden, T. S., Palmer, S. C. F., Travis, J. M. J., & Healey, J. R. (2019). Improving
472 reintroduction success in large carnivores through individual-based modelling: How
473 to reintroduce Eurasian lynx (*Lynx lynx*) to Scotland. *Biological Conservation*, 234,
474 140–153. doi:10.1016/j.biocon.2019.03.035
- 475 Palmer, S. C. F., Coulon, A., & Travis, J. M. J. (2011). Introducing a 'stochastic movement
476 simulator' for estimating habitat connectivity. *Methods in Ecology and Evolution*, 2(3),
477 258–268. doi:10.1111/j.2041-210X.2010.00073.x
- 478 Peterson, M. L., Doak, D. F., & Morris, W. F. (2019). Incorporating local adaptation into
479 forecasts of species' distribution and abundance under climate change. *Global*
480 *Change Biology*, 25(3), 775–793. doi:10.1111/gcb.14562

- 481 R Core Team. (2020). *R: “A language and environment for statistical computing. R*
482 *Foundation for Statistical Computing”*. Vienna, Austria. Retrieved from [https://www.R-](https://www.R-project.org/)
483 [project.org/](https://www.R-project.org/)
- 484 Saastamoinen, M., Bocedi, G., Cote, J., Legrand, D., Guillaume, F., Wheat, C. W., ... del
485 Mar Delgado, M. (2018). Genetics of dispersal. *Biological Reviews*, 93(1), 574–599.
486 doi:10.1111/brv.12356
- 487 Santini, L., Cornulier, T., Bullock, J. M., Palmer, S. C. F., White, S. M., Hodgson, J. A., ...
488 Travis, J. M. J. (2016). A trait-based approach for predicting species responses to
489 environmental change from sparse data: how well might terrestrial mammals track
490 climate change? *Global Change Biology*, 22(7), 2415–2424. doi:10.1111/gcb.13271
- 491 Schumaker, N. H., & Brookes, A. (2018). HexSim: a modeling environment for ecology and
492 conservation. *Landscape Ecology*, 33(2), 197–211. doi:10.1007/s10980-017-0605-9
- 493 Schurr, F. M., Pagel, J., Cabral, J. S., Groeneveld, J., Bykova, O., O’Hara, R. B., ...
494 Zimmermann, N. E. (2012). How to understand species’ niches and range dynamics:
495 a demographic research agenda for biogeography. *Journal of Biogeography*, 39(12),
496 2146–2162. doi:10.1111/j.1365-2699.2012.02737.x
- 497 Singer, A., Johst, K., Banitz, T., Fowler, M. S., Groeneveld, J., Gutiérrez, A. G., ... Travis, J.
498 M. J. (2016). Community dynamics under environmental change: How can next
499 generation mechanistic models improve projections of species distributions?
500 *Ecological Modelling*, 326, 63–74. doi:10.1016/j.ecolmodel.2015.11.007
- 501 Sun, Y., Wang, T., Skidmore, A. K., Palmer, S. C. F., Ye, X., Ding, C., & Wang, Q. (2016).
502 Predicting and understanding spatio-temporal dynamics of species recovery:
503 implications for Asian crested ibis *Nipponia nippon* conservation in China. *Diversity*
504 *and Distributions*, 22(8), 893–904. doi:<https://doi.org/10.1111/ddi.12460>
- 505 Svenning, J.-C., Gravel, D., Holt, R. D., Schurr, F. M., Thuiller, W., Münkemüller, T., ...
506 Normand, S. (2014). The influence of interspecific interactions on species range
507 expansion rates. *Ecography*, 37(12), 1198–1209. doi:10.1111/j.1600-
508 0587.2013.00574.x

- 509 Synes, N. W., Brown, C., Palmer, S. C. F., Bocedi, G., Osborne, P. E., Watts, K., ... Travis,
510 J. M. J. (2019). Coupled land use and ecological models reveal emergence and
511 feedbacks in socio-ecological systems. *Ecography*, *42*(4), 814–825.
512 doi:10.1111/ecog.04039
- 513 Synes, N. W., Brown, C., Watts, K., White, S. M., Gilbert, M. A., & Travis, J. M. J. (2016).
514 Emerging Opportunities for Landscape Ecological Modelling. *Current Landscape*
515 *Ecology Reports*, *1*(4), 146–167. doi:10.1007/s40823-016-0016-7
- 516 Synes, N. W., Ponchon, A., Palmer, S. C. F., Osborne, P. E., Bocedi, G., Travis, J. M. J., &
517 Watts, K. (2020). Prioritising conservation actions for biodiversity: Lessening the
518 impact from habitat fragmentation and climate change. *Biological Conservation*, *252*,
519 108819. doi:10.1016/j.biocon.2020.108819
- 520 Synes, N. W., Watts, K., Palmer, S. C. F., Bocedi, G., Bartoń, K. a., Osborne, P. E., &
521 Travis, J. M. J. (2015). A multi-species modelling approach to examine the impact of
522 alternative climate change adaptation strategies on range shifting ability in a
523 fragmented landscape. *Ecological Informatics*, *30*, 222–229.
524 doi:10.1016/j.ecoinf.2015.06.004
- 525 Thompson, P. L., & Fronhofer, E. A. (2019). The conflict between adaptation and dispersal
526 for maintaining biodiversity in changing environments. *Proceedings of the National*
527 *Academy of Sciences*, *116*(42), 21061–21067. doi:10.1073/pnas.1911796116
- 528 Thuiller, W., Münkemüller, T., Lavergne, S., Mouillot, D., Mouquet, N., Schiffers, K., &
529 Gravel, D. (2013). A road map for integrating eco-evolutionary processes into
530 biodiversity models. *Ecology Letters*, *16*, 94–105. doi:10.1111/ele.12104
- 531 Urban, M. C. (2015). Accelerating extinction risk from climate change. *Science*, *348*, 571–
532 573.
- 533 Urban, M. C. (2019). Projecting biological impacts from climate change like a climate
534 scientist. *WIREs Climate Change*, *10*(4), e585. doi:10.1002/wcc.585

- 535 Urban, M. C., Bocedi, G., Hendry, A. P., Mihoub, J.-B., Peer, G., Singer, A., ... Travis, J. M.
536 J. (2016). Improving the forecast for biodiversity under climate change. *Science*,
537 353(6304), aad8466–aad8466. doi:10.1126/science.aad8466
- 538 Urban, M. C., De Meester, L., Vellend, M., Stoks, R., & Vanoverbeke, J. (2012). A crucial
539 step toward realism: responses to climate change from an evolving metacommunity
540 perspective. *Evolutionary Applications*, 5(2), 154–167. doi:10.1111/j.1752-
541 4571.2011.00208.x
- 542 Urban, M. C., Scarpa, A., Travis, J. M. J., & Bocedi, G. (2019). Maladapted Prey Subsidize
543 Predators and Facilitate Range Expansion. *The American Naturalist*, 194(4), 590–
544 612. doi:10.1086/704780
- 545 Visintin, C., Briscoe, N. J., Woolley, S. N. C., Lentini, P. E., Tingley, R., Wintle, B. A., &
546 Golding, N. (2020). steps: Software for spatially and temporally explicit population
547 simulations. *Methods in Ecology and Evolution*, 11(4), 596–603. doi:10.1111/2041-
548 210X.13354
- 549 Watts, K., Whytock, R. C., Park, K. J., Fuentes-Montemayor, E., Macgregor, N. A., Duffield,
550 S., & McGowan, P. J. K. (2020). Ecological time lags and the journey towards
551 conservation success. *Nature Ecology and Evolution*, 4(3), 304–311.
552 doi:10.1038/s41559-019-1087-8
- 553 Willemen, L., Drakou, E. G., & Schwarz, N. (2019). Modelling how people and nature are
554 intertwined. *Ecography*, 42(11), 1874–1876. doi:10.1111/ecog.04747
- 555 Zollner, P., & Lima, S. (1999). Search strategies for landscape-level interpatch movements.
556 *Ecology*, 80(3), 1019–1030.
- 557 Zurell, D., Thuiller, W., Pagel, J., Cabral, J. S., Munkemuller, T., Gravel, D., ... Zimmermann,
558 N. E. (2016). Benchmarking novel approaches for modelling species range
559 dynamics. *Global Change Biology*, 22(8), 2651–2664. doi:10.1111/gcb.13251
560