# Can citizen science analysis of camera trap data be used to study reproduction? Lessons from Snapshot Serengeti program

Thel Lucie[1], Chamaillé-Jammes Simon[2, 3, 4], Keurinck Léa[1], Catala Maxime[1], Packer Craig[5], Huebner Sarah E.[5], Bonenfant Christophe[1, 4]

[1] Laboratoire de Biométrie et Biologie Évolutive, Unité Mixte de Recherche 5558, Bâtiment 711, Université Lyon I, 43 Boulevard du 11 novembre 1918, F-69622 Villeurbanne Cedex, France.

[2] CEFE, Univ Montpellier, CNRS, EPHE, IRD, Université Paul Valéry Montpellier 3, Montpellier, France.

[3] Mammal Research Institute, Department of Zoology & Entomology, University of Pretoria, Pretoria, South Africa.

[4] LTSER France, Zone Atelier "Hwange", Hwange National Park, Bag 62, Dete, Zimbabwe-CNRS HERD (Hwange Environmental Research Development) program.

[5] College of Biological Sciences, University of Minnesota, Saint Paul, MN 55108, U.S.A.

**Corresponding author:** lucie.mc.thel@gmail.com

**Abstract**

**Abstract**

1 - Ecologists increasingly rely on camera trap data to estimate a wide range of biological

parameters such as occupancy, population abundance or activity patterns. Because of the

huge amount of data collected, the assistance of non-scientists is often sought after, but an

assessment of the data quality is a prerequisite to their use.

2 - We tested whether citizen science data from one of the largest citizen science projects -

Snapshot Serengeti - could be used to study breeding phenology, an important life-history

trait. In particular, we tested whether the presence of juveniles (less than one or 12 months

old) of three ungulate species in the Serengeti: topi *Damaliscus jimela*, kongoni *Alcelaphus*

*buselaphus* and Grant's gazelle *Nanger granti* could be reliably detected by the "naive"

volunteers vs. trained observers. We expected a positive correlation between the proportion

of volunteers identifying juveniles and their effective presence within photographs, assessed

by the trained observers.

3 - We first checked the agreement between the trained observers for age classes and species

and found a good agreement between them (Fleiss' $\kappa > 0.61$ for juveniles of less than one and

12 month(s) old), suggesting that morphological criteria can be used successfully to

determine age. The relationship between the proportion of volunteers detecting juveniles less

than a month old and their actual presence plateaued at 0.45 for Grant's gazelle and reached

0.70 for topi and 0.56 for kongoni. The same relationships were however much stronger for

juveniles younger than 12 months, to the point that their presence was perfectly detected by

volunteers for topi and kongoni.

4 - Volunteers' classification allows a rough, moderately accurate, but quick, sorting of

photograph sequences with/without juveniles. Obtaining accurate data however appears more

difficult. We discuss the limitations of using citizen science camera traps data to study

breeding phenology, and the options to improve the detection of juveniles, such as the

addition of aging criteria on the online citizen science platforms, or the use of machine

learning.

**Keywords**: African ungulates, age determination, *Alcelaphus buselaphus*, camera trap,

citizen science, *Damaliscus jimela*, *Nanger granti,* reproduction

## Introduction

Camera trapping is increasingly used for ecological monitoring due to its low cost, relative

ease of use, and the variety of data it can supply (O'Connell, Nichols and Karanth 2010). For

instance, camera trap data are used to study species' occupancy and co-occurrence (Anderson

*et al.* 2016), population dynamics (Karanth, Nichols, Kumar and Hines 2006), or individual

behaviour (*e.g.* diel activity patterns: Luo *et al.* 2019, or vigilance behaviour: Chamaillé-

Jammes, Malcuit, Le Saout and Martin 2014). A potential drawback of camera traps is the

huge amounts of data that are generated (> 7 millions photographs for the Snapshot Serengeti

initiative alone). Ecologists have realized that the benefits of continuously collecting data in

the field can quickly be negated by the burden of database management, and visual inspection

and analysis of photographs to record the desired data (Wearn & Glover-Kapfer 2017).

To process such a massive amount of information in a reasonable time, scientists have

sought the help of non-specialists who perform diverse tasks like counting objects in

photographs, describing picture content or identifying animal and plant species (*e.g.* McShea,

Forrester, Costello, He and Kays 2015). Initially, part of the scientific community was

sceptical about citizen science, in particular questioning data quality (Riesch & Potter 2014).

However, volunteers have sometimes proven to be as efficient as experts, as for instance for

the identification of large herbivore species in savanna ecosystems (Swanson, Kosmala,

Lintott and Packer 2016). More recently, the advances in deep learning have led computers to

become as efficient as people at identifying species, and, sometimes, behaviour classification

problems (Norouzzadeh *et al.* 2018). However, human judgment is still valuable in particular

cases where too little data are available to train models (*e.g.* active learning, Joshi, Porikli and

Papanikolopoulos 2009), or when differences among the objects to be classified are subtle

and classification requires some subjectivity (Miele *et al.* 2020, *in prep.*). We believe this is

the case for age classification problems, for which to the best of our knowledge the number

of precisely labelled pictures is currently too small to allow an efficient and reliable

automatization of the process.

Classifying individuals into age-classes such as juveniles and adults would allow

estimates of key demographic parameters (*e.g.* reproductive rates) or life-history traits (*e.g.*

breeding phenology), which are essential in estimating population growth rates (Sibly &

Hone 2002). Furthermore, it would facilitate the study of the relationships between

population characteristics and their environments such as between birth phenology, diet and

food resource availability (Sinclair, Mduma and Arcese 2000), or their potential evolution in

the context of climate change (Visser & Both 2005). Until now, the study of those key

demographic parameters has been mainly conducted by direct field observations (*e.g.* Plard *et*

*al.* 2013 in roe deer; Côté & Festa-Bianchet 2001 in mountain goats). However, this

methodology still requires an intensive and often costly field effort. Identifying and counting

juveniles from camera traps could reduce this field effort, or allow larger-scale or longer-term

studies, as suggested by Hofmeester *et al.* 2019, but could also be time-consuming because of

tedious data processing. With the help of citizen science, data handling time could be

substantially reduced, but the accuracy of non-specialists in detecting juveniles of large

mammals from photographs has not yet been explored.

Here, we evaluate the usefulness of camera trap data annotated by citizen scientists

online to assess the presence of juveniles of large herbivores in the photographs. We use

photographs and citizen classifications from the Snapshot Serengeti project (Swanson *et al.*

2015), one of the world's largest citizen science programs, on a subset of the data. We focus

on the detection of juveniles in three species found in the Serengeti National Park, Tanzania

with contrasting social and neonatal behaviours: topi *Damaliscus jimela*, kongoni *Alcelaphus*

*cokii* and Grant's gazelle *Nanger granti*. We first evaluate the agreement between trained

observers from our research team, and then test the ability of the volunteers to detect

juveniles by comparing their classification with ours. We predict a better agreement between

trained observers for the youngest age class because determination criteria are clearer and

easier to identify than for older juveniles (*e.g.* absence of horns). Consequently, the level of

agreement should decrease for age classes that are based on more subjective or difficult-to-

assess criteria (*e.g.* shape of the horns). Under the hypothesis that volunteers could generally

identify juveniles correctly, we expect a positive relationship between the proportion of

volunteers reporting a juvenile on a photograph and the probability of the actual presence of a

juvenile, as determined by the trained observers. Again, we expect the correlation to be

stronger for the youngest age class of juveniles because they are easier to differentiate from

adults. Across species, we expect a higher agreement and correlation for topi and kongoni

than for Grant's gazelle because the former are larger, live in smaller groups and have similar

body growth rate between males and females (Wilson & Mittermeier 2011), hence reducing

the risk of confusion between young males and older females. Overall, our study details the

strengths and weaknesses of camera trap data, in particular when classified by citizen

scientists, for the study of reproductive traits such as reproductive rates or breeding

phenology.

**Materials and methods**

*Study site*

The surveyed area within the Serengeti National Park, Tanzania, is composed of open plains and savanna woodlands. Rains mostly occur between November and June (wet season), with mean annual rainfall increasing from 500 mm in the southeast to 1,100 mm in the northwest. This area harbours a rich community of large herbivores, composed of gregarious and migratory wildebeest *Connochaetes mearnsi*, zebra *Equus sp.* and Thomson's gazelle *Eudorcas nasalis*, but also resident populations such as Cape buffalo *Syncerus caffer* or warthog *Phacochoerus africanus* (Sinclair & Norton-Griffiths 1995). Community dynamics are driven both by herbivores, maintaining an open state of the grassland by intensive grazing (McNaughton 1985, Sinclair & Norton-Griffiths 1995) as well as large predators (*e.g.* lion *Panthera leo* and hyena *Crocuta crocuta*, Sinclair & Norton-Griffiths 1995).

*Camera trap data*

The Snapshot Serengeti camera trap grid was deployed in 2010 in Serengeti National Park, Tanzania, to monitor lions and their prey, though the bycatch of numerous other species has proven useful as well. Running continuously since 2010, the grid spans 1,125 km² in the center of the park. We used data provided by Snapshot Serengeti camera survey recorded between July 2010 and April 2013 (Appendix 1). The camera traps were set ~ 50 cm above ground in the centre of a 5 km² grid cell. The detection radius was approximately 45° and their field of view about 14 m (Swanson *et al.* 2015). Cameras took a rapid series of three pictures upon trigger of the motion and heat sensors ("capture event" in Swanson *et al.* 2015, hereafter called a "sequence" following Meek *et al.* 2014) in a few seconds interval, with a one-minute delay between sequences.

*Choice of studied species*

Among the many large herbivore species present in the study site, we selected topi, kongoni

and Grant's gazelle due to their contrasting biology and characteristics useful to assess the

age classes of individuals. The criteria considered were (1) number of available sequences,

(2) relatively small group size, (3) presence of horns in males and females, (4) relatively large

size of the young (young of larger species are larger, and therefore criteria like horns are

easier to detect), (5) contrasting anti-predator of behaviour of the young (Appendix 2).

*Assessing the presence or absence of juveniles in photographs*

All Snapshot Serengeti photographs have been uploaded to the online citizen science

platform "The Zooniverse" (www.zooniverse.org) to be classified by volunteers. Each

sequence was classified by as many as 25 volunteers (Swanson, Kosmala, Lintott and Packer

2016), who each identified what species was present in the sequence and recorded whether at

least one juvenile was present or not. Note that no formal definition of a juvenile was

provided to volunteers, nor were there any particular guidelines about how to identify a

juvenile. Volunteers simply searched and checked "young" in the Zooniverse interface when

they suspected the presence of non-adult individuals. For each sequence, the volunteers'

classifications were then compiled via a plurality algorithm to yield a consensus

classification, leading to a proportion of volunteers ($Pv$) who identified at least one juvenile

in each sequence (see details in Swanson *et al.* 2015). Here, we used all sequences where

volunteers identified topi, kongoni, and Grant's gazelle, with at least one volunteer ($Pv > 0\%$)

having annotated the presence of a juvenile. We could not analyse all sequences for which no

volunteer had reported a juvenile, as these were too many ($n = 2{,}018$, $11{,}141$ and $6{,}628$ for

topi, kongoni and Grant's gazelle respectively) to be reviewed individually. However, we

checked a subset of them ($n = 1{,}000$ for each species), and the chance that a trained observer

observed a juvenile was $< 6.5\%$ for all three species studied. We did not correct observations

for recaptures of the same individuals as we were only interested in the ability of volunteers

to detect the presence of juveniles on the sequences, but not the actual number of juveniles.

Three of us (LT, LK, MC), considered here as trained observers, searched all

sequences retained for juveniles, which were assigned to an age class when detected. We

used previously published morphological descriptions of the studied species (*e.g.* shape and

size of horns, size relating to the adult; Appendix 3) to identify and age individuals. We

distinguished between (1) juveniles < one month, (2) between one and six months, (3)

between six and 12 months, (4) between 12 and 24 months, termed yearling hereafter and (5)

individuals over 2 years old, termed adults hereafter. We defined age classes according to

biological characteristics relevant to juvenile identification for each species (*e.g.* very young

individuals for birth phenology identification, juveniles under one year for recruitment

estimation). We recorded observers' classifications with Aardwolf software (Krishnappa &

Turner 2014). Ultimately, we produced a dataset describing the presence or absence ($M_{i,s,j}$), in

each sequence of individuals of any of the five age categories *i*, for the species *s*, by trained

observer *j*.

*Statistical analyses*

We first evaluated the agreement between the three trained observers on the detection of

individuals assigned to each age class for each species. We measured this agreement with the

Fleiss' κ, implemented in the "*raters*" R package (Quatto & Ripamonti 2014). Fleiss' κ

(Fleiss 1971) is the comparison of agreement between 2+ judges and the level of agreement

expected by chance alone. It takes values between -1 and 1, values < 0 indicating an

agreement lower to what could be expected by chance, values > 0 indicating a greater

agreement than expected by chance, and values = 0 indicating an agreement close to random.

We tested for significance of the difference between the Fleiss' κ using a bootstrap procedure

following Vanbelle & Albert (2008) (Appendix 4).

We tested the relationship between the proportion of volunteers identifying at least

one juvenile ($Pv$) and the probability that trained observers had identified at least one juvenile

< 1 month (category $i = 1$). We also explored the same relationship with juveniles < 12

months (therefore including juveniles of categories 1 to 3 above). We fitted three generalized

estimating equation models: one linear (see equation 1 below) and two piecewise models.

The first piecewise model was characterized by a slope on both sides of the threshold

(equation 2), the second by a slope before and a plateau after the threshold (equation 3). We

fitted piecewise models to search for a potential "saturation" phenomenon, whereby beyond a

specific proportion of volunteers the probability to effectively observe a juvenile does not

increase anymore. We also fitted the null model for comparison. All the models were fitted

for the two age classes and for each species individually, using the *wgee* function

implemented in the *"wgeesel"* R package (Xu, Li and Wang 2018). We selected the best

model using the Quasi-likelihood under the Independence model Criterion QIC (thresholds

selected by comparison of QIC of the models for each species and age class as well). It is a

modification of the Akaike Information Criterion AICc, suitable when quasi-likelihood is

used instead of likelihood (Pan 2001), implemented in the *"MuMIn"* R package (Barton

2019). We used a logit link function and a binomial distribution of errors (Agresti 2002),

considering the proportion of volunteers identifying at least one juvenile as fixed effect, and

the identity of the sequence as clustering variable with an exchangeable correlation structure:

213
$$logit(M_{s,i,j}) = \mu + \beta_{s,i} \times Pv_s \qquad (\textit{eqn 1 – linear model})$$

214
<div align="center">and</div>

215
$$logit(M_{s,i,j}) = \begin{cases} \mu + \beta 1_{s,i} \times Pv_s \ if \ Pv_s \leq \theta \\ \\ \mu + \beta 2_{s,i} \times Pv_s \ if \ Pv_s > \theta \end{cases} \qquad (\textit{eqn 2 – piecewise model 1})$$

216
<div align="center">and</div>

$$\text{217} \quad logit(M_{s,i,j}) = \begin{cases} \mu + \beta 1_{s,i} \times Pv_s \ if \ Pv_s \leq \theta \\ \\ \mu 2 \ if \ Pv_s > \theta \end{cases} \qquad \textit{(eqn 3 – piecewise model 2)}$$

where $M_{s,i,j}$ denotes the presence or absence of at least one juvenile of the age class $i$ ($< 1$ or

12 months) for a given sequence, for the species $s$ and the observer $j$, $\mu$ is the expected mean

probability of actual presence of at least one juvenile of the age class $i$ when none of the

volunteers identified the presence of a juvenile, $Pv_s$ is the proportion of volunteers identifying

at least one juvenile for a given sequence, for the species $s$ and $\theta$ is the best threshold. We

performed all analyses using the R statistical software (R Core Development Team 2019).

## 225   **Results**

From the Snapshot Serengeti monitoring program, we obtained 281, 1,290 and 1,095

sequences for topi, kongoni and Grant's gazelle respectively (Fig. 1a) sorting step "Young

sorting 1"). Our selection process led to a dramatic drop in the number of usable sequences

(*i.e.* those containing at least one $< 1$ month old juvenile), with a tenfold reduction at the

species level: it only remained between 7.7% and 18.1% of the complete dataset available for

the species ($n = 59 \pm 9$ SD, $137 \pm 33$ SD and $58 \pm 3$ SD for each species respectively, see Fig.

1a) sorting step "young sorting 3"). The largest losses of sequences happened during the

phases of selection of sequences containing the studied species and then juveniles (see Fig.

1a) & Appendix 4).

*Figure 1: a) number of sequences available at each sorting step from the extraction of raw data to the selection*

*of all the independent sequences with at least one individual < 1 month old, for the three species of the study:*

*topi, kongoni, Grant's gazelle. Pictures from Serengeti Snapshot, Tanzania, between July 2010 and April 2013.*

*Raw data: all sequences produced during the survey; blank sorting: sequences with animals; species sorting 1:*

*sequences with study species according to the volunteers; young sorting 1: sequences with at least one volunteer*

*identifying "young"; species sorting 2: sequences with study species corrected by trained observers; young*

*sorting 2: sequences with at least one young according to the trained observers; young sorting 3: sequences*

*with at least one young < 1 month old according to the trained observers; resight sorting: independent*

*sequences once sequences taken less than 10 minutes after the previous one by the same camera trap and*

*presenting the same species have been removed (following Palmer, Swanson, Kosmala, Arnold and Packer*

*2018). Note the log scale for the ordinate axis, vertical bars represent the standard deviations. b) Number of*

*sequences available for all the large herbivore species present in the study site.*

The three trained observers identified 993 ± 49 SD, 3,020 ± 180 SD and 2,128 ± 224

SD individuals of any age class. On average, they could not assign an age class to ~11% (n =

118 ± 68 SD), ~20% (n = 775 ± 256 SD) and 32% (n = 1,026 ± 492 SD) of the individuals for

topi, kongoni and Grant's gazelle respectively, a significant between-species difference ($\chi^2$ =

262.83, $df = 2$, $p < 0.001$).

For all species, and as expected, the agreement between trained observers was highest

for the youngest age class, then declined with age until the yearling class was reached (Fig.

2). Yearlings were reasonably well classified in kongoni, but overly misclassified in topi and

Grant's gazelle (Fig. 2). As expected, the agreement among trained observers was the highest

for the youngest age class of juveniles, but also for the second age class and the adults, with

Fleiss' κ values almost always > 0.61 (denoting a substantial agreement, following Landis &

Koch 1977), except for Grant's gazelle aged between one and six months and adult topi.

Agreement among observers was the highest for topi aged < 1 month old (Fleiss' $\kappa = 0.78$

[0.72; 0.84]). In support of our prediction, we observed the lowest agreement for juveniles

aged six months and older, and more obviously so for Grant's gazelle. Agreement among

observers concerning the three first age classes pooled together (representing the juveniles)

was very good, with Fleiss' κ values largely > 0.61 for all the species. The same holds true

for juveniles between one and 12 months old when pooled together. Our results were globally

consistent among the three species studied, with the highest agreement for topi, and the least

for Grant's gazelle (Fig. 2). All estimated Fleiss' κ were significantly different from an

agreement obtained by chance (Appendix 5).

*Figure 2: Fleiss' κ denoting the level of agreement between the three trained observers (LT, LK, MC) in the*

*identification of presence or absence of at least one individual belonging to any of the five age classes (< 1*

*month, 1-6 months, 6-12 months, > 12 months, adults), for the three species of interest. Pictures from Serengeti*

*Snapshot Program, Tanzania, between July 2010 and April 2013. The two combined age classes "1-12 months"*

*and "< 12 months" are also presented. Light grey dots: topi, dark grey dots: kongoni, black dots: Grant's*

*gazelle. Vertical bars represent the confidence intervals. A Fleiss' κ near 1 denotes an almost perfect*

*agreement, whereas a value near or < 0 means a very poor agreement between raters.*

The model best describing our data for the age class < 1 month was the linear model

for topi, the piecewise model with slopes on both sides of the threshold for kongoni and the

piecewise model with slope before and plateau after the threshold for Grant's gazelle (Table

1). The model best describing our data for the age class < 12 months was the threshold model

with slopes on both sides of the threshold for topi, kongoni and Grant's gazelle (Table 1).

*Table 1: Statistics of models investigating the relationship between the proportion of volunteers identifying at*

| Age class | Species | Model type | θ | QIC | Quasi-likelihood | Estimates |
|---|---|---|---|---|---|---|
| < 1 month | Topi | Null | | 924.6 | -459.77 | *in.* = -1.509 ± 0.133 |
| | | **Linear** | | **630.9** | **-311.05** | **in. = -4.013 ± 0.342**<br>**β = 0.048 ± 0.005** |
| | | Piecewise - slope/plateau | 90 | 635.2 | -313.23 | *in.* = -4.102 ± 0.362<br>*β1* = 0.051 ± 0.005<br>*μ2* = 0.626 [0.523; 0.720] |
| | | Piecewise - slope/slope | 78 | 634.7 | -310.49 | *in.* = -8.863 ± 1.763<br>*β1* = 0.045 ± 0.007<br>*β2* = 0.064 ± 0.023 |
| | Kongoni | Null | | 2614.4 | -1304.83 | *in.* = -2.125 ± 0.081 |
| | | Linear | | 1993 | -992.05 | *in.* = -3.977 ± 0.174<br>*β* = 0.043 ± 0.003 |
| | | Piecewise - slope/plateau | 90 | 2005.8 | -998.42 | *in.* = -4.007 ± 0.178<br>*β1* = 0.044 ± 0.003<br>*μ2* = 0.498 [0.434; 0.563] |
| | | **Piecewise - slope/slope** | **10** | **1966.7** | **-977.58** | **in. = -10.522 ± 1.740**<br>**β1 = 0.687 ± 0.178**<br>**β2 = 0.039 ± 0.003** |
| | Grant's gazelle | Null | | 1236.2 | -615.91 | *in.* = -2.479 ± 0.117 |
| | | Linear | | 1016.6 | -501.81 | *in.* = -3.653 ± 0.181<br>*β* = 0.055 ± 0.007 |
| | | **Piecewise - slope/plateau** | **43** | **969.7** | **-480.75** | **in. = -4.355 ± 0.227**<br>**β1 = 0.097 ± 0.008**<br>**μ2 = 0.451 [0.353; 0.552]** |

| | | | | | | |
|---|---|---|---|---|---|---|
| | | Piecewise - slope/slope | 43 | 974.6 | -480.48 | *in.* = -4.573 ± 0.544<br>*β1* = 0.094 ± 0.010<br>*β2* = 0.006 ± 0.013 |
| < 12 months | Topi | Null | | 1241.3 | -618 | *in.* = 0.698 ± 0.111 |
| | | Linear | | 948 | -468.64 | *in.* = -0.866 ± 0.184<br>*β* = 0.059 ± 0.007 |
| | | Piecewise - slope/plateau | 60 | 964.6 | -477.23 | *in.* = -0.987 ± 0.191<br>*β1* = 0.069 ± 0.007<br>*μ2* = 0.958 [0.926; 0.977] |
| | | **Piecewise - slope/slope** | **30** | **946.5** | **-464.45** | ***in.* = -3.202 ± 0.624**<br>***β1* = 0.031 ± 0.017**<br>***β2* = 0.090 ± 0.022** |
| | Kongoni | Null | | 4992.3 | -2493.63 | *in.* = 0.609 ± 0.054 |
| | | Linear | | 3726.5 | -1858.95 | *in.* = -1.139 ± 0.108<br>*β* = 0.089 ± 0.006 |
| | | Piecewise - slope/plateau | 90 | 3726.6 | -1859.03 | *in.* = -1.140 ± 0.108<br>*β1* = 0.089 ± 0.006<br>*μ2* = 0.999 [0.997; 1] |
| | | **Piecewise - slope/slope** | **12** | **3684.8** | **-1835.45** | ***in.* = -3.041 ± 0.271**<br>***β1* = 0.213 ± 0.029**<br>***β2* = 0.066 ± 0.006** |
| | Grant's gazelle | Null | | 3127.4 | -1561.2 | *in.* = -0.154 ± 0.067 |
| | | Linear | | 2940.5 | -1463.22 | *in.* = -0.837 ± 0.140<br>*β* = 0.049 ± 0.010 |
| | | Piecewise - slope/plateau | 30 | 2866.2 | -1428.27 | *in.* = -1.285 ± 0.132<br>*β1* = 0.091 ± 0.009<br>*μ2* = 0.809 [0.748; 0.859] |
| | | **Piecewise - slope/slope** | **41** | **2866.4** | **-1425.59** | ***in.* = -0.009 ± 0.506**<br>***β1* = 0.083 ± 0.010**<br>***β2* = -0.029 ± 0.013** |

The probability of actually observing a juvenile when all volunteers reported one was near 1 only for juveniles < 12 months for topi and kongoni, but only reached 0.901 and above all, decreased beyond 41% of the volunteers for Grant's gazelle (Fig. 3 d), e), f)). When investigating the presence of juveniles < 1 month, the probability that a juvenile was actually present was never greater than 0.697 (Fig. 3 a), b), c)). On the other hand, when virtually no volunteers reported the presence of juveniles, there was indeed no juveniles < 1 month, but there was at least 9.6% chance to observe a juvenile < 12 months.

*Figure 3: relationship, as predicted from the best model (see text for details), between the proportion of volunteers identifying the presence of "young" and the probability of presence of at least one individual < 1 (a), b), c)) and < 12 (d), e), f)) months assessed by the trained observers in a given sequence (0 < proportion of volunteers identifying "young" ≤ 1), for the three species of interest: a) and d) topi, b) and e) kongoni and c) and f) Grant's gazelle. Pictures from Serengeti Snapshot Program, Tanzania, between July 2010 and April 2013. Light grey dots represent the probability of presence of at least one individual < selected age class in each sequence assessed by the three trained observers, dark grey dots represent the mean of those probabilities for each 10% volunteers interval, vertical bars represent 95% confidence intervals. Solid line represents predicted values from the best model. Shaded areas represent 95% confidence intervals.*

306      Contrary to our expectations, the piecewise model characterized by two slopes was

307    almost always selected as the best model for the three species and both age classes. This

308    denotes a sudden change in the rate of detection from a certain percentage of volunteers. The

309    detection rate decreases for kongoni and Grant's gazelle for young < 12 months beyond 12

310    and 41% of volunteers voting for the presence of young respectively.

## Discussion

Our study reveals the strength and weaknesses of using citizen-based assessment of age

classes on camera trap pictures. Clearly, citizen involvement through an online platform has

been critical for the classification of the millions of photographs collected by the Snapshot

Serengeti initiative. Also, despite leaving it to the volunteers to decide what a juvenile looks

like, volunteers' classification allows a rough, moderately accurate, but quick sorting of

sequences with/without juveniles.

Our study makes clear that, for the species studied and given the minimal guidelines

given to volunteers, the *absence* of very young (< 1 month) individuals on pictures can be

reliably assumed when virtually no volunteer reports a presence. This suggests that

volunteers rarely miss very young juveniles. This likely occurs because very young juveniles

are distinctively smaller than adults, and possibly also because very young mammals share

some physical characteristics such as relatively large eyes, long legs, short and rounded nose,

all belonging to *Kindchenschema*, known to be very attractive stimuli for humans (Brosch,

Sander and Scherer 2007; Golle, Lisibach, Mast and Lobmaier 2013). By contrast, the

*presence* of very young individuals appears more difficult to ascertain from volunteers' data,

and this apparently comes from the lack of guidelines given to volunteers. Indeed, consistent

with the idea that volunteers easily identify very young individuals, a large consensus among

volunteers around the presence of a very young juvenile could be a reliable indication of

actual presence, but only when no older juveniles are present (compare Fig. 3 a), b), c) &

Appendix 6 - Fig. A). Unfortunately, the absence or presence of older juveniles cannot

currently be known without a reassessment of the pictures because volunteers were not asked

to differentiate between juvenile age classes. Therefore, the presence of very young juveniles

remains difficult to ascertain. On the other hand, the *presence* of juveniles, irrespectively of

their age, can be reliably assumed when virtually all volunteers agree about a presence (Fig. 3

a), b), c)). This time, the *absence* of juveniles regardless of their age is less accurate, meaning

that they are missed quite often. As they grow, juveniles become increasingly similar to

adults, and are more likely to be labeled as such by volunteers. Anyhow, we emphasize that

detectability of juveniles is not equal between species( Fig. 3).

Among the three trained observers, we found the best agreement in the detection of

age classes for a given sequence for topi and kongoni, suggesting that they are the easiest

species to classify, and Grant's gazelle the hardest. The combination of small body size and

gregariousness could explain why the determination of age was more difficult and

heterogeneous for Grant's gazelle than for the two other species. Topi and kongoni are fairly

large, and small body size makes the detection of some inconspicuous age criteria

challenging (*e.g.* presence or absence of very small horn buds on the forehead of the

individuals). On the contrary, it is certainly easier to discriminate between adults and

juveniles in the largest herbivores, such as giraffe *Giraffa camelopardalis* or elephant

*Loxodonta africana* because of the marked size difference between them. Discrimination

between several juvenile age classes will certainly be difficult still as individuals are often

only partially spotted by camera traps and subtle aging criteria are not visible. More

generally, in species forming large herds such as wildebeest, young might be particularly

difficult to spot and are likely missed frequently. In large groups, the body of many

individuals overlaps, hampering our ability to see aging criteria correctly, and to age them

accurately. This could explain why in our study for instance, Grant's gazelle frequently

occurring in large herds reaching more than 30 individuals in some sequences, is also the one

with the lowest identification success of juveniles. Finally, a pronounced sexual dimorphism

in horn growth in Grant's gazelle likely led to confusions between the first two age classes as

male horns grow faster, a young male can look like an older female when using the length of

the horns as an aging criteria. This would also be the case for species with a sexual

dimorphism in which only males grow horns like impala *Aepyceros melampus* or waterbuck

*Kobus defassa*. Clearly, a description of the morphological changes that occur throughout the

development of young herbivores (*e.g.* Spinage 1976 in our species; Cunningham, Sandouka

and Wronski 2011 in other ungulates; but see also Dezeure *et al.* 2020 in other species) is of

great value and substantially helps at reaching consistent results among different observers.

Another limitation of camera trapping in the context of reproductive studies is the

level of effort required to obtain a sufficient sample size. Here, working with data from one

of the world's largest and long-running camera trap studies, we ended up with a small

number of sequences with juveniles < 1 month old after appropriate data selection. We

identified four main explanations for this important reduction in exploitable sequences. First

and foremost, our ability to determine an individual's age class depends strongly on the

photograph quality and particularly its framing and exposure. In many cases, individuals were

located too far from the camera or were only partly visible, or photographs were too blurry,

dark, or overexposed to be scrutinized, leading to potentially significant loss of reproductive

data and a high number of individuals of unknown or over-estimated age. Another potential

source of information loss was species misidentification by volunteers. In our study, about

30% of the sequences labelled with Grant's gazelle were misidentified because it greatly

resembles species such as impala and Thomson's gazelle which are also present in the study

site. Species abundance obviously directly impacts the number of sequences collected. The

abundance of the three studied species is low compared to other ungulates in the Serengeti

system. Sinclair reported 55,500 individuals topi, 20,700 individuals kongoni and 6,000

individuals Grant's gazelle in the 1970s, whereas the numbers of wildebeest and zebra were

720,000 and 240,000 individuals respectively (Sinclair & Norton-Griffiths 1995). Finally, the

anti-predator strategy of juvenile large herbivores, known as the hider-follower gradient

(Rutberg 1987; Lent 1974), could influence the number of sequences containing very young

individuals. While followers become active and stick with their mother just a few hours after

birth, hiders stay concealed in dense vegetation during their first weeks of life. The detection

probability of hiders from camera traps should then be much lower than of followers,

consistent with our observation of very young topi seen in a greater proportion than kongoni

and Grant's gazelle.

**Conclusions**

Volunteer classification provides information that can reliably be used to infer the *presence*

of juveniles < 12 months or the *absence* of juveniles < 1 month, as these annotations appear

robust. However, because volunteers seem able to discriminate between individuals of less

than one month old and the rest of the juveniles even in the absence of any stated criteria,

more precise results could be achieved by by asking them to differentiate between two age

classes of juveniles, such as "juvenile" and "newborn". Another way to improve results

generated via citizen science platforms could be the inclusion of detailed information about

what juvenile morphology looks like according to its species and age, as has already been

done for species identification purposes (Swanson *et al.* 2015). We also suggest evaluating

volunteers' classification skills by presenting them with images of individuals of known age

(captive or tagged animals for instance) and assessing their accuracy compared to labels

provided by experts. Volunteers could then be assigned classification tasks adapted to their

skills (*e.g.* species identification would belong to the easiest tasks, whereas age classification

would belong to the hardest). Snapshot Safari and Zooniverse continue to create new modes

of annotation that best leverage the public's interest in contributing to research, and this is a

logical next step for the Snapshot Safari initiative. Overall, we find that by closely

investigating the data already collected by volunteer-based programs, data collection

procedures can be adjusted to enhance the contributions of citizen scientists to scientific

research and conservation efforts.

**Declarations**

**Acknowledgments**: A PhD fellowship from the university Lyon 1 attributed to LT. This

work was performed using the computing facilities of the CC LBBE/PRABI. The authors

thank T. Michael Anderson for maintaining the camera traps in Tanzania, Zooniverse for

hosting "Snapshot Serengeti", >130,000 online volunteers for classifying the images, the

Tanzania Wildlife Research Institute and Tanzania National Parks for research permission,

and the Minnesota Supercomputing Institute (http://www.msi.umn.edu) for contributing to

data storage/processing and analysis.

**Funding**: A PhD fellowship from the University Lyon 1 attributed to LT.

**Authors' contributions**: LT, CB and SCJ conceived the ideas and designed methodology;

CP and SH organized data collection; LT, LK and MC analysed the data; LT, CB and SCJ led

the writing of the manuscript. All authors contributed critically to the drafts and gave final

approval for publication.

**Data availability:** Pictures available from the Labeled Image Library of Alexandria –

Biology and Conservation: http://lila.science/datasets/snapshot-serengeti and data from the

Dryad Data Repository: *to be completed*.

## References

Agresti, A. (2002). Categorical data analysis, 3rd edition (3rd ed.). Wiley, Hoboken, New Jersey.

Anderson, T. M., White, S., Davis, B., Erhardt, R., Palmer, M., Swanson, A., . . . Packer, C. (2016). The spatial distribution of african savannah herbivores: species associations and habitat occupancy in a landscape context. Philosophical Transactions of the Royal Society B: Biological Sciences, 371 (1703), 20150314. doi: 10.1098/rstb.2015.0314

Bartoń, K. (2019). Mumin: Multi-model inference [Computer software manual]. Retrieved from https://CRAN.R-project.org/package=MuMIn (R package version 1.43.10)

Brosch, T., Sander, D., & Scherer, K. R. (2007). That baby caught my eye… attention capture by infant faces. Emotion, 7(3), 685–689. doi: 10.1037/1528-3542.7.3.685

Chamaillé-Jammes, S., Malcuit, H., Le Saout, S., & Martin, J.-L. (2014). Innate threat-sensitive foraging: black-tailed deer remain more fearful of wolf than of the less dangerous black bear even after 100 years of wolf absence. Oecologia, 174 (4), 1151–1158. doi: 10.1007/s00442-013-2843-0

Côté, S. D., & Festa-Bianchet, M. (2001). Birthdate, mass and survival in mountain goat kids: effects of maternal characteristics and forage quality. Oecologia, 127 (2), 230–238. doi: 10.1007/s004420000584

Cunningham, P. L., Sandouka, M. A., & Wronski, T. (2011). Morphological characteristics and ageing criteria of sand gazelle (gazella subgutturosa marica thomas, 1897) and their use for wildlife management. European Journal of Wildlife Research, 57 (4), 865–872. doi: 10.1007/s10344-011-0498-x

Dezeure, J., Dagorrette, J., Baniel, A., Carter, A. J., Cowlishaw, G., Marshall, H. H., Martina, C., Raby, C. L. & Huchard, E. (2020). Developmental transitions in body color in chacma baboon infants: Implications to estimate age and developmental pace.

458    American Journal of Physical Anthropology, e24118. doi: 10.1002/ajpa.24118

459 Fleiss, J. L. (1971). Measuring nominal scale agreement among many raters. Psychological

460    bulletin, 76 (5), 378. doi: 10.1037/h0031619

461 Golle, J., Lisibach, S., Mast, F. W., & Lobmaier, J. S. (2013). Sweet puppies and cute babies:

462    Perceptual adaptation to babyfacedness transfers across species.PloS one,8(3),

463    e58248. doi: 10.1371/journal.pone.0058248

464 Hofmeester, T. R., Young, S., Juthberg, S., Singh, N. J., Widemo, F., Andrén, H., . . .

465    Cromsigt, J. P. (2019). Using by-catch data from wildlife surveys to quantify climatic

466    parameters and the timing of phenology for plants and animals using camera traps.

467    Remote Sensing in Ecology and Conservation. doi: 10.1002/rse2.136

468 Joshi, A. J., Porikli, F., & Papanikolopoulos, N. (2009). Multi-class active learning for image

469    classification. In 2009 ieee conference on computer vision and pattern recognition

470    (pp. 2372–2379). doi: 10.1109/cvpr.2009.5206627

471 Karanth, K. U., Nichols, J. D., Kumar, N. S., & Hines, J. E. (2006). Assessing tiger

472    population dynamics using photographic capture–recapture sampling. Ecology, 87

473    (11), 2925–2937. doi: 10.1890/0012-9658(2006)87[2925:atpdup]2.0.co;2

474 Krishnappa, Y. S., & Turner, W. C. (2014). Software for minimalistic data management in

475    large camera trap studies. Ecological informatics, 24 , 11–16. doi:

476    10.1016/j.ecoinf.2014.06.004

477 Landis, J. R., & Koch, G. G. (1977). The measurement of observer agreement for categorical

478    data. biometrics, 159–174. doi: 10.2307/2529310

479 Lent, P. C. (1974). Mother-infant relationships in ungulates. The behaviour of ungulates and

480    its relation to management, 1 , 14–55.

481 Luo, G., Yang, C., Zhou, H., Seitz, M., Wu, Y., & Ran, J. (2019). Habitat use and diel

482    activity pattern of the tibetan snowcock (tetraogallus tibetanus): a case study using

camera traps for surveying high-elevation bird species. Avian Research, 10 (1), 4. doi:

10.1186/s40657-019-0144-y

McNaughton, S. (1985). Ecology of a grazing ecosystem: the serengeti. Ecological

monographs, 55 (3), 259–294. doi: 10.2307/1942578.

McShea, W. J., Forrester, T., Costello, R., He, Z., & Kays, R. (2015). Volunteer-run cameras

as distributed sensors for macrosystem mammal research. Landscape Ecology, 31 (1),

55–66. doi: 10.1007/s10980-015-0262-9

Meek, P., Ballard, G., Claridge, A., Kays, R., Moseby, K., O'brien, T., . . . others (2014).

Recommended guiding principles for reporting on camera trapping research.

Biodiversity and conservation, 23 (9), 2321–2343. doi: 10.1007/s10531-014-0712-8

Miele, V., Dussert, G., Spataro, B., Chamaillé-Jammes, S., Allainé, D., & Bonenfant, C.

(2020). Revisiting giraffe photo-identification using deep learning and network

analysis. bioRxiv. doi: 10.1101/2020.03.25.007377

Norouzzadeh, M. S., Nguyen, A., Kosmala, M., Swanson, A., Palmer, M. S., Packer, C., &

Clune, J. (2018). Automatically identifying, counting, and describing wild animals in

camera-trap images with deep learning. Proceedings of the National Academy of

Sciences, 201719367. doi: 10.1073/pnas.1719367115

O'Connell, A. F., Nichols, J. D., & Karanth, K. U. (2010). Camera traps in animal ecology:

methods and analyses. Springer Science & Business Media.Palmer, M. S., Swanson,

A., Kosmala, M., Arnold, T., & Packer, C. (2018). Evaluating relative abundance

indices for terrestrial herbivores from large-scale camera trap surveys. African Journal

of Ecology, 56 (4), 791–803. doi: 10.1111/aje.12566

Palmer, M. S., Swanson, A., Kosmala, M., Arnold, T., & Packer, C. (2018). Evaluating

relative abundance indices for terrestrial herbivores from large-scale camera trap

surveys. African Journal of Ecology, 56(4), 791–803. doi:10.1111/aje.12566

Pan, W. (2001). Akaike's information criterion in generalized estimating equations.

Biometrics, 57 (1), 120–125. doi: 10.1111/j.0006-341x.2001.00120.x

Plard, F., Gaillard, J.-M., Bonenfant, C., Hewison, A. M., Delorme, D., Cargnelutti, B., . . .

Coulson, T. (2013). Parturition date for a given female is highly repeatable within five

roe deer populations. Biology Letters, 9 (1), 20120841. doi: 10.1098/rsbl.2012.0841

Quatto, P., & Ripamonti, E. (2014). raters: A modification of fleiss' kappa in case of nominal

and ordinal variables [Computer software manual]. Retrieved from https://CRAN.R-

project.org/package=raters (R package version 2.0.1).

Riesch, H., & Potter, C. (2014). Citizen science as seen by scientists: Methodological,

epistemological and ethical dimensions. Public understanding of science, 23 (1), 107–

120. doi: 10.1177/0963662513497324

Rutberg, A. T. (1987). Adaptive hypotheses of birth synchrony in ruminants: an interspecific

test. The American Naturalist, 130 (5), 692–710. doi: 10.1086/284739

Sibly, R. M., & Hone, J. (2002). Population growth rate and its determinants: an overview.

Philosophical Transactions of the Royal Society B: Biological Sciences, 357 (1425),

1153–1170. doi: 10.1017/cbo9780511615740.002

Sinclair, A., Mduma, S. A., & Arcese, P. (2000). What determines phenology and synchrony

of ungulate breeding in serengeti? Ecology, 81 (8), 2100–2111. doi: 10.1890/0012-

9658(2000)081[2100:wdpaso]2.0.co;2

Sinclair, A. R. E., & Norton-Griffiths, M. (1995). Serengeti: dynamics of an ecosystem.

University of Chicago Press.

Spinage, C. (1976). Age determination of the female grant's gazelle. African Journal of

Ecology, 14 (2), 121–134. doi: 10.1111/j.1365-2028.1976.tb00157.x

Swanson, A., Kosmala, M., Lintott, C., & Packer, C. (2016). A generalized approach for

producing, quantifying, and validating citizen science data from wildlife images.

Conservation Biology, 30 (3), 520–531. doi: 10.1111/cobi.12695

Swanson, A., Kosmala, M., Lintott, C., Simpson, R., Smith, A., & Packer, C. (2015).

Snapshot serengeti, high-frequency annotated camera trap images of 40 mammalian

species in an african savanna. Scientific data, 2 , 150026. doi: 10.1038/sdata.2015.26

Vanbelle, S., & Albert, A. (2008). A bootstrap method for comparing correlated kappa

coefficients. Journal of Statistical Computation and Simulation, 78 (11), 1009–1015.

doi: 10.1080/00949650701410249

Visser, M. E., & Both, C. (2005). Shifts in phenology due to global climate change: the need

for a yardstick. Proceedings of the Royal Society B: Biological Sciences, 272 (1581),

2561–2569. doi: 10.1098/rspb.2005.3356

Wearn, O., & Glover-Kapfer, P. (2017). Camera-trapping for conservation: a guide to best-

practices (Tech. Rep.). WWF-UK.

Wilson, R., D.E. & Mittermeier. (2011). Handbook of the mammals of the world. vol 2.

hoofed mammals. (Vol. 2; L. Edicions, Ed.). Lynx Edicions.

Xu, C., Li, Z., & Wang, M. (2018). wgeesel: Weighted generalized estimating equations and

model selection [Computer software manual]. Retrieved from https://CRAN.R-

project.org/package=wgeesel (R package version 1.5).