CellKb Immune: a manually curated database of mammalian hematopoietic marker gene sets for rapid cell type identification

Ajay Patil[1], Ashwini Patil[1*]

[1]Combinatics, Ichikawa-shi, Chiba, Japan.

* Correspondence:
Ashwini Patil
ashwini@combinatics.com

Keywords: single-cell, cell types, marker genes, immune cells, annotation, database, cell type
prediction

Abstract

Using single-cell RNA sequencing, it is now possible to get the detailed gene expression pattern in
each cell for millions of cells in a single experiment under varying conditions. Annotating cells by
assigning a type or state to them is important for correct interpretation of results. In spite of current
advances in technology, assigning cell types in single-cell datasets remains a bottleneck due to the
lack of a comprehensive reference database and an accurate search method in a single tool. CellKb
Immune is a knowledgebase of manually collected, curated and annotated marker gene sets
representing cell types in the immune response in mouse. It allows users to find matching cell types
in literature given a list of genes. We evaluated the contents and search methodology of CellKb
Immune using a leave-one-out approach. We further used CellKb Immune to annotate previously
defined marker gene sets from a bulk RNA-seq dataset to confirm its accuracy and coverage. CellKb
Immune provides an easy-to-use online tool with a reliable method to find matching cell types and
annotate cells in single-cell experiments. It is available at https://www.cellkb.com/immune.

1   Introduction

Single-cell RNA-seq is widely used to study transcriptional patterns of genes in individual cells.
Various computational tools have been developed for the annotation of cell types to cells in single-
cell experiments, given a reference database and a gene list (1,2). Several single cell reference
datasets or atlases have also been prepared (3–5), or compiled into aggregate reference databases of
cell types and marker gene sets (6–9). There are currently two primary methods used for annotating
cells in single-cell experiments. 1) Manually searching through scientific literature to find marker
genes associated with cell types being studied. This requires prior knowledge of the cell types
potentially present in the sample and fails to identify rare cell types without established markers. 2)
Selecting a single cell reference database/ dataset from literature, downloading and processing the
data, identifying and installing a computational method to assign cell types, and finally, writing a

program in R or Python to perform the cell type assignment. This method usually limits a researcher
to using a single, or a few datasets as a reference. Both these methods require significant time and
effort but fail to use the vast amount of cell type marker gene sets previously published. Reliable
annotations require a resource that contains a comprehensive reference database of cell types and
marker gene sets from literature along with a search method that allows users to find matching cell
types.

CellKb Immune addresses these issues by collecting and aggregating a large amount of immune-
related cell type signatures in mouse from literature and providing an online interface to find
matching annotated cell types easily, quickly and accurately.

2   Materials and Methods

2.1   Data collection and curation

The CellKb Immune database consists of marker gene sets of hematopoietic cell types with
associated annotations collected from literature. The marker genes and their cell type annotations are
taken primarily from published single-cell RNA-seq experiments. All publications related to single-
cell experiments are taken from PubMed and manually screened to select those identifying cell type
specific gene expression patterns.

Cell type specific marker gene sets are defined as those identified by authors based on their
significant change in expression within a group of cells as compared to all other cells in the
experiment. The marker gene sets are extracted from supplementary materials and raw, or processed,
data of publications describing these experiments. If calculated based on the raw/processed
expression data, cell type specific marker genes are identified using the methods described by the
authors in the publication, along with cell type annotations given for the single cells.

Marker gene sets are selected from published experimental studies if they satisfy the following
criteria: 1) Raw data related to the experiment is deposited in public databases, 2) The marker gene
sets are publicly available for download, 3) The experimental methods and conditions are adequately
described in the manuscript to allow detailed annotation, 4) The number of cells and samples studied
is clearly specified, 5) The computational methods used to normalize, filter and cluster cell types,
along with identification of cluster-specific genes is clearly described, 6) Associated values such as
average expression, fold change, statistical significance, raw UMI or read counts, are available for
marker genes (marker gene sets specified only as ranked gene lists without associated values are
included in some cases), 7) A majority of the gene identifiers in the marker gene set are valid and can
be mapped to the latest version of the Ensembl database (10). Single-cell experiments without cell
type annotations are not used to calculate cell type specific marker gene sets.

Detailed sample and treatment conditions, including sample counts and cell counts are also stored in
CellKb Immune. Information about disease state of the samples and the cell types enriched in disease
samples are also noted. All data in CellKb Immune is referenced and linked to the PubMed identifier
of the source publication.

CellKb Immune also contains marker genes extracted from select bulk RNA-seq and microarray
experiments from public databases. These include selected gene signatures from MSig-db (11) and

previously published bulk RNA-seq datasets of immune cell types (1,12). Cell-type specific
interaction networks are created using marker gene sets of all cell types and protein-protein
interactions with reliability scores from the HitPredict database (13).

2.2   Data harmonization and annotation

Data harmonization is essential for rapid search and retrieval of matching cell types. Given that there
is no standard format currently in place for sharing cell type specific marker gene sets, CellKb
Immune performs a number of operations to harmonize the data collected. Marker gene sets are
extensively curated to identify valid genes and cell types. Genes given in the published marker gene
sets are mapped to valid Ensembl identifiers by matching names, synonyms or identifiers. Assigning
a common gene identifier to the marker genes is an important part of data harmonization in CellKb
Immune.

Associated values given by authors with each signature are also stored in CellKb Immune. These
include, but are not limited to, rank, score, average expression, log fold change and
corrected/uncorrected p-values.

For each marker gene set up to 1,500 marker genes with corrected p-values less than or equal to 0.1
and log fold change greater than 0 compared to other cell types are selected. The marker genes are
ranked by CellKb Immune based on descending log fold change values. Marker gene sets that are
published in the form of ranked gene lists without associated values are also included in the database
if they match an existing marker gene set with the same cell type annotation. Unranked gene lists
without associated values are not included in CellKb Immune.

Each marker gene set is associated with a cell type, tissue/organ and disease condition given in the
source publication, which are mapped to standardized ontology terms using Cell Ontology (14),
Uberon Ontology (15) and Disease Ontology (16).

Each marker gene set is assigned a reliability score that gives an indication of how similar it is to
other marker gene sets of the same ontology. The higher the reliability score of a marker gene set, the
closer it is to other marker gene sets of the same cell type and, hence, the more likely it is to be
accurate. This helps identify marker gene sets that are incorrect, calculated from erroneous cell
cluster assignments by authors, or that have been calculated from mixtures of cell types.

Finally, all marker gene sets and their annotations are stored in a uniform format with indexed
identifiers enabling rapid search and retrieval of the data.

2.3   Data searching

In CellKb Immune, users can search for cell type specific marker gene sets by giving a list of
statistically significant query genes ranked in descending order of log fold change. Query genes given
by the user are compared with every marker gene set in the database and the best match is identified
using a rank-based match score.

The match score is based on the number of common genes between the query and a marker gene set,
their ranks, their rank differences and the total number of significant genes in the marker gene set.
This results in marker gene sets sharing highly ranked genes with the query being assigned higher
match scores. This method also accounts for differences in the sizes of gene lists between the query
and various marker gene sets, such those with fewer marker genes are not disregarded.

3   Results

3.1   Database contents

CellKb Immune contains author-defined mouse hematopoietic cell type marker gene sets manually
collected from publications describing mainly single-cell, and selected bulk RNA-seq or microarray
experiments (1,11) (Figure 1). Cell type marker gene sets are assigned a Cell Ontology term (14),
where available. Tissues, organs or body parts of the experimental sample are assigned Uberon
ontology terms (15). Disease ontology terms are provided where the marker gene sets are obtained
from a diseased sample (16). All marker gene sets reference their source publication with a link to its
PubMed identifier.

CellKb Immune contains:

• 2,395 marker gene sets from mouse
• 230 unique annotated hematopoietic cell types (Cell Ontology CL:0000988 and all its descendants).
• 104 tissues/organs
• 226 publications selected from approximately 10,000 studies published between 2013 and 2021.
• 40,122 protein-protein interactions from HitPredict (13)

3.2   Functionality

The CellKb Immune web interface allows users to search matching cell types and their marker gene
sets in mouse. It can be used to annotate cells from single-cell RNA-seq data, given a list of
statistically significant cluster-specific genes ranked by decreasing log fold change. Associated
values such as log fold changes and p-values are also accepted as input by CellKb Immune. Genes in
the user query are compared with every marker gene set in the database using a rank-based search
method and a match score is computed (see Methods). The user is notified if the query gene list
matches non-hematopoietic cell types in 5 out of top 10 marker gene set matches. Figure 2 shows the
CellKb Immune user interface.

3.3   Evaluation

To check the database quality and cell type matching performance of the CellKb Immune search
method, we used the leave-one-out approach. We matched each of the 2,223 mouse marker gene sets
annotated with cell ontologies in CellKb Immune with all others in the database. 1,568 (70.5%) of
gene sets were correctly matched to another gene set with the exact same ontology or its parent,
while for 427 (19.2%) gene sets, the ontology of the first hit was a sibling i.e. a descendant of the
same parent ontology (Table 1). Thus, 89.7% of marker gene sets were matched to another marker
gene set with the same ontology, or its parent or sibling (Table S1).

We further evaluated the performance of the CellKb Immune by checking the number of correct cell
types assigned to gene lists obtained from a bulk RNA-seq dataset of mouse immune cell types (12)
from Immgen (17). Using the top1,500 significantly upregulated genes for each cell type in the bulk
RNA-seq dataset, we searched the CellKb Immune database for matching marker gene sets. We then
compared the cell ontology of the top 10 hits with that of the ontology assigned to the gene lists in
the query dataset. Of the 98 marker gene sets representing 40 distinct cell types, CellKb Immune was
able to find the exact matching ontology or its parent as the first hit in 61 instances (62.2%). In
another 18 marker gene sets (18.4%), the ontology of the first hit was a sibling i.e. a descendant of

the same parent ontology as that in the query dataset (Table 1). Thus, a total of 79.5% of marker gene
sets in the bulk RNA-seq dataset had their first hit as the same ontology, its parent or sibling
ontology (Table S2).

These results show that 1) the CellKb Immune database includes high quality annotated marker gene
sets from single-cell experiments, and 2) the search methodology used by CellKb Immune to find
matching cell types is able to recapitulate the original cell types from within its own database and
other unseen bulk and single-cell datasets.

We next compared the functionality of CellKb Immune to two other similar databases, CellMarker
(7) and PanglaoDB (8) (Table 2). CellKb Immune is unique in its ability to search matching cell
types from published literature using ranked gene lists. CellMarker allows users to search only one
gene at a time, while PanglaoDB can search cell types using unranked gene lists. Additionally, both
CellMarker and PanglaoDB contain gene signatures that have been aggregated from multiple samples
and/or sources. In contrast, CellKb Immune contains cell type marker gene sets as described in
literature and gives the overlapping genes between the query and the matching cell types obtained in
a search result. Some other features that differentiate CellKb Immune from CellMarker and
PanglaoDB are deep annotations in the form of standardized ontologies and protein-protein
interaction networks. CellKb Immune also contains marker gene sets recently published in literature
and is regularly updated.

Thus, CellKb Immune provides complementary contents and functionality to other cell type marker
databases.

4   Discussion

CellKb Immune has been designed as a resource to find matching cell type specific marker gene sets
from literature to help assign cell types to cells in single-cell RNA-seq experiments. CellKb Immune
addresses several issues in existing single-cell reference databases and tools used for this purpose.

CellKb Immune consists of user-defined marker gene sets from literature. These carry significant
knowledge since the cell types in the form of cell clusters are often selected by authors based on
biological information. CellKb Immune is able to capture and aggregate this biological information.
On the other hand, several reference databases reanalyze the raw expression patterns using a common
pipeline while ignoring the methodology and cluster definitions used by the authors in the original
study, thus losing valuable biological information.

CellKb Immune provides deep annotations, in the form of extensive cell type information and
description, curated and validated marker genes ranked by significance, along with fold change and
significance values associated with gene expression. Additionally, CellKb Immune provides a web-
based interface to find matching cell types in published data sets given a user gene list, independent
of the experimental platform, analysis methods and size of the marker gene sets. Thus, users do not
need to spend time integrating reference data and search methods programmatically.

Unlike other computational methods which require the presence of associated expression or fold
change values and the same number of genes in all target cell types, the rank-based search method
used by CellKb Immune enables searching through marker gene sets of differing sizes, in the absence

of expression fold changes, and independent of experimental platform and pre-processing methods.
Searching of cell types happens by direct comparison of the query gene list with the marker genes of
every cell type in CellKb Immune using a fast and reliable method without any type of score
aggregation or machine learning. As a result, CellKb Immune provides information about the
matching cell type in the form of the publication, experimental details and the overlapping genes
between the query and matching marker gene set. This allows users to further explore the study to
which their cell type matches. In cases where the top matches to a marker gene set from single-cell
data are inconclusive, users can refer to the consensus of the top matching cell types or compare their
gene lists with gene sets from bulk RNA-seq or microarray experiments.

Since CellKb Immune contains marker gene sets from multiple publications for each cell type, it can
also be used to evaluate the validity of known marker genes for the cell type of interest. The
reliability score assigned to each cell type marker gene set can also be used to identify those that are
incorrectly annotated in publications. Additionally, comparing the query gene list with marker gene
sets from multiple publications helps assess the accuracy of the matching cell types found by CellKb
Immune.

Thus, CellKb Immune provides an easy-to-use reference database with a fast and reliable method to
find matching cell types and marker gene sets to help annotate cells in single-cell experiments in a
single tool.

214 5 Conflict of Interest

215 *Ashwini is CEO at Combinatics Inc., a privately-held company. Ajay and Ashwini are stock-holders*
216 *in Combinatics Inc.*

218 6 Author Contributions

219 Ashwini conceived of the project, collected the data and prepared the manuscript. Ajay and Ashwini
220 designed and developed the database, search method and web application. All authors read and
221 approved the final manuscript.

223 7 References

1.   Aran D, Looney AP, Liu L, Wu E, Fong V, Hsu A, Chak S, Naikawadi RP, Wolters PJ, Abate
AR, et al. Reference-based analysis of lung single-cell sequencing reveals a transitional
profibrotic macrophage. *Nat Immunol* (2019) doi:10.1038/s41590-018-0276-y

2.   Pliner HA, Shendure J, Trapnell C. Supervised classification enables rapid annotation of cell
atlases. *Nat Methods* (2019) doi:10.1038/s41592-019-0535-3

3.   Rozenblatt-Rosen O, Stubbington MJT, Regev A, Teichmann SA. The Human Cell Atlas:
From vision to reality. *Nature* (2017) doi:10.1038/550451a

4.   Han X, Wang R, Zhou Y, Fei L, Sun H, Lai S, Saadatpour A, Zhou Z, Chen H, Ye F, et al.
Mapping the Mouse Cell Atlas by Microwell-Seq. *Cell* (2018) doi:10.1016/j.cell.2018.02.001

5.   Schaum N, Karkanias J, Neff NF, May AP, Quake SR, Wyss-Coray T, Darmanis S, Batson J,
Botvinnik O, Chen MB, et al. Single-cell transcriptomics of 20 mouse organs creates a Tabula
Muris. *Nature* (2018) doi:10.1038/s41586-018-0590-4

6.   Ner-Gaon H, Melchior A, Golan N, Ben-Haim Y, Shay T. JingleBells: A Repository of
Immune-Related Single-Cell RNA–Sequencing Datasets. *J Immunol* (2017)
doi:10.4049/jimmunol.1700272

7.   Zhang X, Lan Y, Xu J, Quan F, Zhao E, Deng C, Luo T, Xu L, Liao G, Yan M, et al.
CellMarker: A manually curated resource of cell markers in human and mouse. *Nucleic Acids
Res* (2019) doi:10.1093/nar/gky900

8.   Franzén O, Gan LM, Björkegren JLM. PanglaoDB: A web server for exploration of mouse
and human single-cell RNA sequencing data. *Database* (2019) doi:10.1093/database/baz046

9.   Papatheodorou I, Moreno P, Manning J, Fuentes AMP, George N, Fexova S, Fonseca NA,
Füllgrabe A, Green M, Huang N, et al. Expression Atlas update: From tissues to single cells.
Nucleic Acids Res (2020) doi:10.1093/nar/gkz947

10. Howe KL, Achuthan P, Allen J, Allen J, Alvarez-Jarreta J, Amode MR, Armean IM, Azov
AG, Bennett R, Bhai J, et al. Ensembl 2021. *Nucleic Acids Res* (2021) **49**:D884–D891.
doi:10.1093/NAR/GKAA942

11. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, Paulovich A,
Pomeroy SL, Golub TR, Lander ES, et al. Gene set enrichment analysis: A knowledge-based
approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A* (2005)
doi:10.1073/pnas.0506580102

12. Yoshida H, Lareau CA, Ramirez RN, Rose SA, Maier B, Wroblewska A, Desland F,
Chudnovskiy A, Mortha A, Dominguez C, et al. The cis-Regulatory Atlas of the Mouse
Immune System. *Cell* (2019) **176**:897-912.e20. doi:10.1016/j.cell.2018.12.036

13. López Y, Nakai K, Patil A. HitPredict version 4: Comprehensive reliability scoring of physical
protein-protein interactions from more than 100 species. *Database* (2015) **2015**:
doi:10.1093/database/bav117

14. Bard J, Rhee SY, Ashburner M. An ontology for cell types. *Genome Biol* (2005)
doi:10.1186/gb-2005-6-2-r21

15. Mungall CJ, Torniai C, Gkoutos G V., Lewis SE, Haendel MA. Uberon, an integrative multi-
species anatomy ontology. *Genome Biol* (2012) doi:10.1186/gb-2012-13-1-r5

16. Schriml LM, Arze C, Nadendla S, Chang YWW, Mazaitis M, Felix V, Feng G, Kibbe WA.
Disease ontology: A backbone for disease semantic integration. *Nucleic Acids Res* (2012)
doi:10.1093/nar/gkr972

17. Aguilar SV, Aguilar O, Allan R, Amir EAD, Angeli V, Artyomov MN, Asinovski N, Astarita

J, Austen KF, Bajpai G, et al. ImmGen at 15. *Nat Immunol* (2020) doi:10.1038/s41590-020-
0687-4

8   Figures

272   Figure 1. CellKb Immune workflow – data collection, harmonization and search interface.

273   Figure 2. CellKb Immune user interface used to search matching cell types given a gene list.

275 **9   Tables**

276 *Table 1. Performance evaluation of CellKb Immune*

Evaluation database	Marker gene sets	CellKb Immune performance		Total ontologies matched
		Correct ontology match	**Parent ontology match**	
CellKb Immune Mouse (leave-one-out)	2,223	1,568 (70.5%)	427 (19.2%)	**1,995 (89.7%)**
Yoshida et al. (12) **Mouse (all-against-all)**	98	61 (62.2%)	18 (18.4%)	**79 (80.6%)**

279   *Table 2. Comparison of CellKb Immune functionality with other similar tools.*

Feature	CellKb Immune	CellMarker (7)	PanglaoDB (8)
Last update	10 February 2022	3 November 2018	27 March 2020
Manual curation	Yes	Yes	No
Ranked marker genes with associated values, eg. log fold change, FDR	Yes	No	Yes
Marker gene sets per cell type	Multiple (specified as given in literature)	Single (aggregated from multiple sources)	Single (aggregated from multiple sources)
Marker genes obtained from single-cell RNA-seq experiments	Yes	Yes	Yes
Processing of single-cell RNA-seq data to calculate marker genes	Using author-specified data processing pipeline	No	Using standardized data processing pipeline
Marker genes obtained from bulk RNA-seq/ microarray experiments	Yes	Yes	No
Disease information	All	Cancer only	All
Protein-protein interaction networks of marker genes	Yes	No	No
Search matching cell types with ranked gene lists	Yes	No	No
Cell/Uberon/Disease ontology annotations	Yes	Limited Cell Ontology annotations	No
Reliability score for marker gene sets	Yes	No	Yes

10   Supplementary Material

284   Table S1. Results of the leave-one-out evaluation in CellKb Immune shows the first hit of each cell
285   type marker gene set compared to all other marker gene sets within the species.

286   Table S2. First hit of each cell type marker gene set in a bulk RNA-seq dataset of immune cell types
287   compared to all marker gene sets in CellKb Immune.

11   Data Availability Statement

290   CellKb Immune is available at https://www.cellkb.com/immune. All data used in CellKb Immune is
291   publicly available and appropriately referenced in the web application.

Search marker gene sets using ranked gene list

View mapping of gene names to Ensembl IDs

View heatmap of matching marker gene sets and cell types

View cell type specific protein interaction network

View common genes in matching marker gene set

View table of matching marker gene sets with annotations