Sexual differentiation of neural mechanisms of stress sensitivity during puberty

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Abstract

Anxiety disorders are more common in women than men, and this difference arises during puberty. Increased secretion of gonadal hormones during puberty influences brain structure and function, but the extent to which hormones modulate anxiety-related brain circuits is unclear. The slow developing California mouse (*Peromyscus californicus*) is an ideal species for studying the effects of hormones on brain function during adolescence. In adults social defeat stress reduces social approach and increases vigilance in females but not males. Here we show this sex difference is absent in juvenile mice, and that prepubertal castration sensitizes adult males to social defeat. Since adult castration has no effect on stress sensitivity, our data show that gonadal hormones act during puberty to program behavioral responses to stress later in life. In adults, calcium imaging in the medioventral bed nucleus of the stria terminalis shows that threatening social contexts increase calcium transients. Furthermore, prepubertal castration generalizes these responses to less threatening social contexts. Prepubertal treatment with the non-aromatizable androgen dihydrotestosterone acts in males and females to reduce sensitivity to social defeat in adults. Together, these data indicate activation of androgen receptors during puberty are critical for programing behavioral responses to stress in adulthood, highlighting a possible mechanism contributing to sex differences in anxiety.

Significance Statement

Puberty is a key period when sex differences in anxiety emerges. Gonadal hormone release increases during this time but it is largely unknown how they impact brain circuits and behavior. We show that androgens play a key role in programming behavioral responses to social defeat stress. The bed nucleus of the stria terminalis responds to social threats and these responses are more generalized in males without gonadal hormone exposure during puberty. Our findings highlight the importance of pubertal androgens in determining adult behavioral responses to social stress.

Introduction

Adolescence is an important developmental period of cortical and sub-cortical re-organization (1–3) and a period when risk for anxiety disorders increases in girls (4–7). This increased risk is sustained into adulthood (8–13). The increased release of gonadal hormones may modulate these changes (14), as these hormones have important effects on brain structure (15–18) and function (19) by coordinating gene expression (20). Although human imaging studies demonstrate that neural circuits associated with anxiety are altered during pubertal development (21,22), little is known of how gonadal hormones alter stress-sensitivity of these circuits. This is important because stress exposure is an important risk factor for mental illness, and there is strong evidence in preclinical models that androgens can modulate behaviors related to anxiety or depression (23).

Social stressors exert powerful and sex-specific effects on anxiety-related behaviors in rodents (24,25). Female but not male adolescent rats exposed to a combination of social defeat and restraint stress showed reduced sucrose preference and increased passive coping in a forced swim test as adults (26). Adolescent social instability stress, in which rats are repeatedly housed with different cagemates, has stronger effects on anxiety-related behaviors in adult females than males (27,28). In adolescent mice, social isolation enhanced habit-related behaviors to a greater extent in females than males (29). A major outstanding question is whether pubertal hormones drive sex-specific behavioral and neurobiological responses to stress.

An important challenge for studying adolescent development in rodents is the relatively short temporal window. The most common domesticated mouse (30,31) and rat (32) lines are weaned at three weeks of age and are mature by five weeks of age. Here we introduce the California mouse (*Peromyscus californicus*) as a slow developing species that is ideal for studying adolescent development. The California mouse is an important model species for studying sex-specific effects of social stressors on anxiety-related behaviors (33). In adults social defeat increases the responsiveness of the bed nucleus of the stria terminalis (BNST) in adult females (34,35), which drives social avoidance and vigilance in novel social contexts (36–38). In adults these sex differences are independent of gonadal hormones (39,40). Here we use

a combination of pubertal hormone manipulations, calcium imaging, and

immunohistochemistry to demonstrate that androgens act during puberty to reduce behavioral

responses to social defeat stress. These identify a neuroendocrine mechanism acting during

puberty that directs developmental programing of stress-sensitive neural circuits.

Results

Late Onset of Pubertal Development in California mice. Complementary developmental

measures suggest a slower developmental trajectory in California mice (Fig. 1A) compared to

domesticated mice and rats. In males, testosterone levels increased with age (Fig. 1B,

$F_{5,28}=6.51$, $p<0.01$), but were not significantly elevated until post-natal day 90 (PN90, $p<0.001$,

Cohen's d=1.9). In females, progesterone levels increased with age (Fig. 1B, $F_{5.26}=2.56$, $p=0.05$)

but were not significantly elevated until PN70 ($p=0.02$, d=1.7). Other developmental measures

such as gonadal weight, external genitalia, body weight, and pelage molt followed a similar

trajectory (Fig. S1). Together, these data indicate that California mice have an extended period

of development, ideal for studying adolescence.

Juvenile Male and Female California Mice Reduce Social and Approach and Increase Vigilance

after Defeat. Juvenile mice were exposed to social defeat stress or control conditions after

weaning and tested in a social interaction test as PN50 adolescents (Fig. 1C). Social defeat

reduced social approach (Fig. 1D, $F_{1,27}=47.09$, $p<0.001$) in both males ($p<0.001$, Cohen's d=3.0)

and females ($p<0.001$, d=2.4). Defeated also increased social vigilance (Fig. 1E, Kruskal-

Wallis=16.33, $p<0.001$) in both males ($p=0.03$, d=1.6) and females ($p=0.02$, d=1.8). These results

contrast with observed sex-differences in adults, which show that social defeat decreases social

approach (35,39–41) and increases vigilance (42) in females but not males. During the

acclimation phase females spent more time investigating an empty cage than males (Fig. 1F, $F_{1,27}=6.64$, $p=0.016$) with no effects of stress. There were no differences in locomotor behavior in

the open field phase (Fig. 1G), time spent in the center (Fig. S2A) or vigilance behavior during

the acclimation phase (Fig S2B). Together with previous work, these results suggest that

developmental changes during puberty contribute to sex differences in stress responses in

adults.

Gonadal Hormones Reduce Susceptibility to Defeat in Adult Males. To assess the impact of

gonadal hormones during puberty, males were assigned to prepubertal castration, sham

surgery, or no surgery (Fig. 2A). Gonadectomy affected social approach only in males that were

exposed to social defeat as adults (Fig. 2B, trt*stress $F_{2,41}$=3.32, p<0.05). Castrated males

exposed to defeat had reduced social approach versus controls (p<0.0001, d=2.6) whereas

there was no effect of defeat in sham or no surgery males. Similarly, social defeat increased

vigilance in castrated (Fig. 2C, Mann-Whitney U=23.5, p<0.01, d=1.6) but not sham or no

surgery mice. There were no differences in behavior during the acclimation (Fig. 2D, S3A) or

open field phases (Fig. 2E, S3B). The results contrast sharply with previous results that showed

no effect of adult castration or ovariectomy on stress-induced social avoidance in California

mice (40), and point to gonadal hormones acting during adolescence as a key mechanism of

sexual differentiation. These data suggest that pubertal hormones may play a protective role in

organizing the brain to diminish the effects of social defeat on social approach and vigilance.

The BNST plays a key role in modulating stress-induced social avoidance and vigilance in males

and females (35,38,42). Since prepubertal castration did not affect behavior in unstressed mice,

we used fiber photometry to assess effects of castration on BNST neural activity in males

exposed to defeat.

Calcium imaging of the BNST activity in castrated and intact males. The BNST modulates social

approach and avoidance (34,42), so we performed calcium imaging of GCaMP6f in

medioventral BNST to determine how prepubertal castration impacted neural responses in

novel social contexts (Fig. 3A, 3B). Previous immediate early gene analyses show that social

stress increases activity within the BNST (43,44), but does not have the temporal resolution to

determine whether increased activity occurs during brief episodes of social approach or more

extended periods of avoidance. Our results indicated that GCaMP signals coincided with social

contact with target mice rather than avoidance, regardless of gonadal status. Using DeepLabCut

(Fig. 3C, 3D) we determined orientation (45) and distance between focal mice and non-

aggressive or aggressive target mice. Increased ΔF/F in BNST was observed when focal mice

were within one body length (8 cm) of target mice and within the central (Fig. 3D, 0°≥40°, β= -

0.438, z= -9.783, p>0.001) or peripheral visual field (40°≥100°, β= -0.486, z= -10.356, p>0.001),

but not if the focal mouse was facing away from target mice (100°≥180°). We then examined

ΔF/F in specific behavioral contexts.

Intriguingly, the most robust changes in ΔF/F occurred after close contact with an aggressive

target mouse. After engaging in nose-to-nose sniffing, both castrated and sham males showed a

delayed increase in BNST ΔF/F (Fig. 3E, β=10.22, z=2.05, p=0.04). In contrast, when engaging in

nose-to-nose sniffing with a non-aggressive target mouse, castrated males showed a larger

increase in BNST ΔF/F than sham males (Fig. 3E, β=13.2, z=2.23, p=0.03). These results suggest

that BNST ΔF/F is enhanced by social threats and that this response is more generalized in

prepubertally castrated males. We also examined BNST following bouts of freezing, which was

robustly induced by aggressive targets (Fig. S4) while non-aggressive targets induced more

freezing in castrated males than shams (t_{11}=2.43, p=0.03). There were no acute or delayed

changes in ΔF/F following bouts of freezing (Fig. 3F). We next examined bouts of anogenital

sniffing, which were less frequent and limited to non-aggressive target mice (Fig. S4). Similar to

nose-to-nose sniffing castrated mice showed a larger delayed increase in ΔF/F in BNST than

shams (Fig. 3G, β=6.12, z=2.06, p=0.04. Finally, when castrated males were attacked by

aggressive target mice, there were acute (Fig. 3H, β=5.71, z=2.56, p=0.01) and delayed (β=5.71,

z=2.67, p=0.008) increases in ΔF/F. Together these data suggest that neuronal activity in BNST

increases after potentially-threatening social situations, and is generalized to less threatening

contexts after prepubertal castration. To assess the extent to which prepubertal castration

impacted neuronal activity in other circuits modulating social approach, we used c-fos

immunohistochemistry.

Gonadal Hormones Reduce Neural Activity in PVN Oxytocin Neurons: Social defeat stress has

been demonstrated to elevated the activity of oxytocin releasing neurons in PVN (34), a

response that has been associated with heightened activation of stress-reactive neurons

expressing oxytocin receptors in the anteromedial BNST (42). We used oxytocin/c-fos

immunohistochemistry (Fig. 4A,B) to examine anterior and posterior PVN, which differ in

connectivity (46) and stress sensitivity (34). In anterior PVN, prepubertal castration increased

oxytocin/c-fos colocalizations when compared to sham (Fig. 4C', Mann-Whitney U=38, p=0.01)

in mice that had undergone social defeat. Oxytocin/c-fos colocalizations were negatively

correlated with social approach (Fig. 4C", Spearman ρ=-0.55, p<0.01) and positively correlated

with social vigilance (Fig. S5A, ρ=0.59, p<0.01). In the posterior PVN castration increased

oxytocin/c-fos colocalizations regardless of stress status (Fig. 4D', Mann-Whitney U=113.5,

p=0.04) and oxytocin/c-fos colocalizations was negatively correlated with social approach (Fig.

4D", ρ=-0.41, p=0.04) but not vigilance (Fig. S5B). We also examined the effects of prepubertal

castration on c-fos in the anteromedial BNST, where oxytocin induces social avoidance and

vigilance (38). Prepubertal castration increased the number of c-fos positive neurons in

BNSTam (Fig. 4E', S5D, $F_{1,21}$=4.51, p=0.046), regardless of stress status. The number of c-fos cells

was negatively correlated with social approach (Fig. 4E", ρ=-0.46, p=0.02) but not vigilance (Fig.

S5C).

We also tested whether defeat induced social avoidance and vigilance in adolescent mice

was dependent on oxytocin receptors (Fig. 4F) as in adults. In both males and females,

treatment with 5 mg/kg i.p. of the oxytocin receptor antagonist L-368,899 30 min before testing

increased social approach (Fig. 4G, main effect of dose $F_{2,42}$=8.42, p<0.001) and decreased

vigilance (Fig. 4H, Kruskal-Wallis=15.36, p<0.009). There were no differences in behavior during

the acclimation (Fig. S5E,S5F) or open field (Fig. S5G, S5H) phases. Together these results

suggest that stress-induced social avoidance and vigilance in both male and female juvenile

mice is dependent on oxytocin receptor activation, as seen in adult females. Additionally, the

results indicate that male pubertal hormones program these circuits to be less active in

adulthood.

Androgens Reverse the Effects of Prepubertal Castration on Behavior.

To test the impact of androgen replacement at puberty, juvenile males were prepubertally

castrated and randomly assigned to receive a silastic implant containing testosterone, the non-

aromatizable androgen dihydrotestosterone (DHT), or sealant only (Fig 5A). These implants

produce plasma hormone levels within physiological range of adult male California mice (47). At

PN90, mice were exposed to defeat stress and then tested in a social interaction test. Results

demonstrated that hormone replacement altered social approach (Fig. 5B, $F_{2,22}=6.2$, $p<0.01$)

and vigilance (Fig. 5C, Kruskal-Wallis=13.13, $p<0.001$). For social approach both testosterone

($p=0.002$, $d=1.7$) and DHT ($p=0.002$, $d=1.0$) treatment yielded significantly higher levels of social

approach compared to males treated with empty implants. Similarly for vigilance both

testosterone ($p<0.001$, $d=3.1$) and DHT ($P=0.049$, $d=1.1$) treated males had lower vigilance than

males treated with empty implants. There was a nonsignificant trend for testosterone treated

males to have lower vigilance than DHT treated males ($p=0.078$, $d=1.0$). During the acclimation

phase T and DHT treatment increased approach to an empty cage (Fig. S6A, $F_{2,22}=5.02$, $p<0.02$)

and there were no differences in behavior during the open field phase (Fig. S6B, S6C). We also

tested whether DHT treatment at puberty could impact female behavior (Fig. 5D). In social

interaction tests performed before stress exposure, there were no differences in social

approach (Fig. 5E), social vigilance (Fig. 5F), acclimation (Fig. S6D) or open field behavior (Fig.

S6E,F). Importantly, after social defeat, hormone treatment altered social approach (Fig. 5E,

$F_{2,23}=3.76$, $p= 0.04$) with DHT increasing social approach versus empty implants ($p=0.02$, $d=0.1$).

Effects on vigilance were weaker (Kruskal-Wallis=5.04, $p=0.08$), with a post-hoc test indicating

lower levels of vigilance in DHT treated mice versus empty implant (Mann-Whitney=11,

$p=0.028$, $d=1.2$). After stress exposure DHT treatment increased approach to an empty cage

(Fig. S6D, $F_{2,23}=5.32$, $p= 0.01$) while there were no differences in the open field phase (Fig. S6E,

S6F)

Discussion

Sex differences in stress-sensitivity emerge at puberty in both humans (48,49) and animals (26–

28), coinciding with an increased incidence of anxiety in women (4,6,7,12). However, little is

known about the underlying mechanisms. Using the California mouse, we found stress-induced

social avoidance and vigilance occurs in both sexes before puberty. Prepubertal castration

enhanced behavioral responses to stress in adult males. Gonadal hormones do not affect social

avoidance in adults (40,50), suggesting that puberty is a critical period for sexual differentiation.

Dihydrotestosterone replacement during puberty in castrated males and intact females

reduced stress-induced social avoidance in adulthood, highlighting an essential role for

androgen receptors. Using real-time calcium imaging we found that neurons in ventral BNST are

highly responsive to threatening social contexts, and that prepubertal castration generalized

these responses to non-aggressive mice. Our findings indicate that androgens play an

organizational role during puberty, which attenuates behavioral and neural responses to social

stress in adulthood.

BNST neurons respond to social contact. Immediate early gene analyses in rodents (34,51–54)

and neuroimaging studies in primates (55) and humans (56) show that stressful social contexts

increase neural activity within the BNST. However, these analyses lack the temporal resolution

to associate changes in neural activity with specific behaviors. Our calcium imaging data show

that ventral BNST calcium transients occurred after close contact with target mice. Specifically,

increased activity was observed following active engagement with aggressive target mice, such

as nose-to-nose sniffing or aggression. Intriguingly no increases occurred during freezing, a

behavior that does not involve active social engagement. While it has been hypothesized that

the BNST encodes more distant threats (57), our results are more consistent with recent

hypotheses that the BNST is highly responsive to proximate threats (58). The strength of

calcium transients in the presence of non-aggressive target mice was dependent on exposure to

pubertal hormones.

Pre-pubertally castrated males exhibited more freezing and heightened BNST calcium

transients in the presence of non-aggressive target mice. These results are consistent with our

findings showing that pre-pubertal castration leads to increased avoidance and vigilance

towards unfamiliar target mice, similar to stressed adult females (40,59). These findings suggest

that the absence of male gonadal hormones during puberty may alter valence assessment of

novel social contexts (60,61), potentially leading to an over-generalization of threat, which is a

key characteristic of anxiety disorders (62). Our results align with imaging studies in nonhuman

primates (63) and humans (64) reporting stronger BNST responses in individuals with elevated

anxiety-related behaviors. Intriguingly, androgens can reduce neuronal excitability in both adult

(65) and adolescent rodents (66–68). These findings are in line with our observations that

prepubertal castration led to increased c-fos/oxytocin colocalizations in the PVN and c-fos

expression in anteromedial BNST. Taken together, our results suggest that androgens may play

a role during puberty to reduce the excitability of circuits that are affected by social defeat.

The question remains as to which BNST cell types respond to social threats. This is further

complicated by the fact that the BNST exhibits heterogeneity in neural responses, even within

genetically defined cell populations (69–71). This variation could be explained by projection-

specific populations within a cell-type (72). This is supported by optogenetic studies showing

the projection-specific effects of BNST neurons on behavior (73–76). Future research

considering genetic and projection-specific populations during neuronal recording of activity in

different social contexts should be highly informative.

Activation of androgen receptors during puberty blunts stress induced social avoidance. In

adult hamsters, dominant males have more androgen receptor expression in the medial

amygdala (MEA) than subordinates and pharmacological inhibition of AR in the MEA of

dominant males increases sensitivity to social stress (77). Similarly testosterone can exert acute

anxiolytic effects in a variety of behavioral assays (78–80). Interestingly, in adult California mice,

androgens did not appear to affect social approach regardless of stress exposure (40). Here we

show that androgens act during puberty to influence how social behavior is impacted by stress,

suggesting that even in species where androgens do not have an overt role in regulating stress-

sensitivity in adults, androgens can program behavioral responses during adolescence. This may

be particularly relevant for humans, where the relationship between gonadal hormones and

stress responses in adults can be inconsistent (81–83). Although adolescent DHT and

testosterone treatment had similar effects on social approach in stressed males, the effect of

DHT on vigilance was weaker. Similar effects were observed in females. These results suggest

that estrogen receptors may play a complementary role in pubertal organization of stress-

induced behavior. Estrogens can enhance anxiety-related behaviors by activating *Esr1* or exert

anxiolytic effects via *Esr2* (84). Both receptors are present in the BNST, and future studies

should consider the possible developmental effects of these receptors on stress-induced social

vigilance.

Functional implications

Steroid hormones alter brain development during puberty (15,17,19), yet how these changes

affect anxiety-related behavior has been unclear. We showed that sex differences in social

stress effects on behavior are driven, in part, by pubertal androgen exposure in males. A lack of

pubertal hormone exposure increased reactivity of BNST neurons during social engagement.

Interestingly, the effects pubertal androgens on social behavior were only apparent following

exposure to social defeat. This demonstrates that organizational effects of pubertal hormones

can cause latent vulnerabilities that are only revealed after stressful social experiences. Our

results suggest that it will be worthwhile to consider whether testosterone levels during

adolescence predict behavioral or neural responses to stressors in adulthood. Overall, our

research sheds light on how androgens shape the complex interplay between brain circuits and

behavioral sensitivity to stress.

Methods

<u>Animals</u>

All studies were conducted with California mice (*Peromyscus californicus*) raised in a colony at

UC Davis. Mice were housed in same sex groups (2-4) in clear polypropylene cages with Sani-

Chip bedding (Harlan Laboratories, Indianapolis, IN, USA), Nestlets (Ancare, Bellmore, NY, USA)

and Enviro-Dri (Eco-bedding, Fibercore, Cleveland, OH, USA). Mice were kept on a 16L:8D light

cycle and had *ad libitum* access to food and water. All procedures were approved by the

Institutional Animal Care and Use Committee at the University of California, Davis and in

accordance with National Institutes of Health (NIH) guidelines.

Puberty Quantification

For measures of first vaginal opening, preputial separation, weight, and coat color mice were

briefly anesthetized (>1 minute isoflurane) before being weighed, photographed, and assessed

for vaginal opening/preputial separation. A separate set of mice were euthanized to determine

uterine or testes size. Trunk blood was collected and plasma obtained and frozen for hormone

analyses. Plasma was extracted and hormones quantified on a QTRAP 550 quadruple linear ion

trap mass spectrometer equipped with an atmospheric pressure chemical ionization source (LC-

MS/MS) as previously described (85). Quantitative results were recorded as multiple reaction

monitoring (MRM) area counts after determination for the response factor for each hormone

and internal standard. We also quantified the transition from the juvenile pelage (dark gray) to

adult (brown) adult pelage (86) by quantifying digital images in ImageJ as previously described

(87).

Juvenile Social Defeat

Male and female juveniles (PN34-36) underwent 3 consecutive days of social defeat stress

(or control handling) as described previously (50). During each episode of defeat the focal

mouse was placed in the home cage of a novel male-female adult resident pair. The opposite

sex resident mouse was removed from the cage prior to the start of the defeat session. Each

session of defeat lasted for 7 minutes, or until the test mouse received 7 bites. Control mice

were placed into a clean, empty cage for 7 minutes across 3 consecutive days.

On PN50 mice underwent a social interaction test. The social interaction test had 3 phases:

open field, acclimation, and interaction. In the open field phase, the mouse was placed into an

open area and allowed to explore it for 3 minutes. In the acclimation phase, an empty cage was

placed at one end of the arena and the mouse was allowed to explore for 3 minutes. In the

interaction phase, a caged, unknown adult conspecific replaced the empty cage at one end of

the arena, and the mouse was allowed to explore for 3 minutes. Time in the interaction zone (8

cm from the cage) is defined as "social approach" was scored with AnyMaze software (40).

Duration of vigilance behavior was hand scored from a video recording. Vigilance was defined

as any time the test mouse was sitting still, head oriented toward the target mouse, while

outside of the interaction zone (37).

<u>Prepubertal Castration</u>

Male juveniles were randomly assigned to be castrated, sham surgery, or no surgery control

between PN35 and PN40 (47). During castration mice were anesthetized with isoflurane and

treated with 0.1mg/kg buprenorphine and 5mg/kg of carprofen. Non-surgery controls were

included to control for possible effects of early life exposure to the anesthesia isoflurane (88).

These mice received no manipulations until the start of social defeat stress/control handling. At

PN90-92 all mice underwent 3 days of either social defeat or control handling. At PN106 mice

were tested in a social interaction test.

<u>Calcium imaging of the BNST activity in castrated and intact males</u>

Male juveniles were randomly assigned to castration or sham surgery after weaning. As

adults, all mice received an injection of AAV9.Syn.GCaMP6s at a rate of 100nL/min for a total

volume of 500nL in the ventral BNST (AP +0.45mm, ML +/-1.0mm, DV -5.6mm). The virus was

allowed to diffuse for 10 minutes before the needle was withdrawn. An optical fiber (Doric)

with 2.5 mm core and 0.66 NA threaded through a ceramic ferrule was implanted at the

injection site and the ferrule was secured to the skull with a layer of C&B Metabond (Parkell),

followed by a layer of dental cement to form a thick headcap. Mice were housed two per cage

with a clear, perforated acrylic divider that allowed auditory, tactile and olfactory contact.

Mice recovered for 1 week before undergoing 3 consecutive days of social defeat stress. Ten

days later mice began 3 consecutive days of patch cord habituation in which the patch cord was

gently coupled to their optical fiber implant. The mouse then explored a novel cage for 10 min.

The next day mice were tested in a social interaction test with freely moving target mice. The

focal mouse was placed into an empty area (51x25.4x76 cm) attached to a small box (13x10x18

cm) with a sliding door for 6 min. Photometry recording was performed with an isosbestic

channel of 405nm and an excitatory channel of 470nm, both set to 50μW output. Photometry

data from the acclimation period were excluded from analysis due to initial bleaching. Next, an

unfamiliar, nonaggressive adult male target mouse was introduced into the arena through the

sliding door. Mice were allowed to freely interact for 3 min. The target mouse was removed

and then a sexually experienced, aggressive male (from a previous social defeat episode) was

introduced into the arena for 3 min. After testing brains were collected to confirm viral

expression and placement of the fiber.

We used Deeplabcut to quantify distance and orientation of the focal mouse in relation to

target mice with precision that matched the high temporal resolution of photometry data (89–

91). Video was taken from the side view at a rate of 30fps. And training was performed to track

the nose, right ear, left ear, body midpoint, front leg, back leg, tail base, and tail tip for both the

test mouse and stimulus mouse. The tracking for the nose and body midpoint were used in

order to determine proximity, defined as times when the body midpoint of the two mice were

within 8 cm (the approximate length of a California mouse) of each other. We determined the

direction the test mouse was facing in relation to the stimulus mouse using their coordinate

points obtained from the deeplabcut tracking (test mouse nose, body midpoint, and target

mouse nose). We binned angles as within the test mouse's central vision (0°≥40°), peripheral

vision (40°≥100°), or facing away from the stimulus mouse (100°≥180°) (45). All scripts are

deposited at https://github.com/bctrainorlab/behavioral_quantification. Nose to nose sniffing,

anogenital sniffing, freezing and aggressive behavior directed towards focal mice were scored

by an observer without knowledge of the treatment group using BORIS.

<u>Pubertal hormone manipulation studies</u>

Male juveniles were castrated and received a subcutaneous silastic implant (i.d. 0.04 in, o.d.

0.085 in) containing 1mm crystalline testosterone, dihydrotestosterone, or Dowsil Sealant

(3145 RTV MIL-A-46146). These implants yield hormone levels that are within the physiological

range of intact male California mice (92). At adulthood mice underwent 3 days social defeat and

were tested in a social interaction test two weeks later.

Intact females were randomly assigned to receive a DHT or empty implant at PN35-PN40 as

described for males. At PN90 all mice were tested in a social interaction test. Three days later

females were exposed to 3 days of social defeat on consecutive days. One week later females

were tested again in a social interaction test. This within-subjects design produces behavioral

effects similar to the between-subjects design used for males (93).

Immunohistochemistry

Sections from castrated or intact males were stained for either oxytocin/c-fos (PVN) or c-fos

(anteromedial BNST) as previously described(34). Sections were blocked in 10% normal goat

serum for 20 minutes and then incubated in rabbit anti-cfos (Synaptic Systems, 1:K) in PBS 0.5%

TritonX overnight at room temperature. For oxytocin double staining, sections were washed in

PBS and then incubated in goat anti-rabbit AlexaFluor 555 (Molecular Probes, 1:500 in PBX-TX)

for 2 hrs. Sections were then incubated in mouse anti-oxytocin (Sigma-Aldrich MAB5296 1:2000

in PBS-TX) overnight at 4 C. Sections were then washed in PBS and incubated goat anti-mouse

AlexaFluor 488 (Molecular Probes, 1:500 in PBS-TX) for 2 hrs. For BNSTam sections, sections

were washed in PBS and incubated in biotinylated goat anti-rabbit (Vector labs, 1:500 in PBS-TX

for 2 hrs). Sections were washed in PBS and then incubated in avidin-conjugated peroxidase

(Vector labs) for 30 min. After washing in PBS sections were developed in nickel enhanced

diaminobenzidine (Vector labs) for 2 min.

Juvenile Social Defeat + OTA Injections

Male and female juveniles (PN34-36) underwent 3 consecutive days of social defeat stress.

At P50 mice were tested in a social interaction test. 30 minutes before the start of the social

interaction test, each mouse received an intraperitoneal (i.p.) injection of 5mg/kg or 10mg/kg

of the oxytocin antagonist (OTA) L-368,899 (37), or an injection of sterile phosphate buffered

saline (PBS).

Statistics

Statistical analyses for experiments (excluding fiber photometry experiments) were

performed using R statistical software. Normality was assessed via QQPlot. A Flinger-Killen test

was used to assess homogeneity of variance. Steroid hormone data were log transformed for

analysis. Hormone and most behavioral and cell count data were analyzed using ANOVA with

planned comparisons for comparing treatment groups with controls. Social vigilance and

oxytocin/c-fos colocalization data had heterogeneous variability, so non-parametric analyses

were used.

Photometry data was analyzed using a custom Python script, down sampled to 30 samples

per second to match the 30fps video framerate. 405nm was used as the isosbestic wavelength

and 470nm as the excitatory wavelength. To correct for motion artifact and bleaching of the

fluorophore, the 405nm signal was fit to a biexponential model and subtracted from the

excitatory output. Such that $\Delta F/F = [100*((470nm\ signal - fitted\ signal)/fitted\ signal)]$. Z-scores

were calculated using the formula: z-score = $((\Delta F/F$ - mean $\Delta F/F$ of baseline period)/standard

deviation of $\Delta F/F$ of baseline period). For behavior non-specific analysis, the baseline period

was 5 to 25 seconds after recording started. For behavior-specific analysis, the baseline period

was -10 to -5 seconds before the behavior onset. Area under the curve (AUC) was calculated

using the trapezoidal method for specified time points across behaviors. Linear mixed effects

models were used to determine the effects of surgery, social condition, and time point using

the statsmodels package (94) in Python using one of the following formulas:

• Z-Score=surgery condition + social condition + distance + surgery condition*social condition

+ surgery condition*distance + social condition*distance + surgery condition*social

condition*distance + (1|mouse)

• AUC=surgery condition + time point + surgery condition*time point + (1|mouse)

• AUC=social condition + time point + social condition*time point + (1|mouse)

• AUC=time point + (1|mouse)

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Figures and Legends

Figure 1: Delayed puberty in California mice is a period of sexual differentiation of behavioral stress responses. A. Summary of pubertal development in California mice **B.** In males testosterone levels are not increased until postnatal day (PN) 90 while in females progesterone levels increase at PN70 (n=4-5 per time point). **C.** Prepubertal California mice were exposed to social defeat or control conditions after weaning and tested as juveniles at PN50. **D,E.** Social defeat reduced social approach and increased social vigilance in both males and females (n=6-8 per group). **F.** Females investigated an empty cage during the acclimation phase more than males. **G.** There were no differences in locomotor behavior in the open field phase. *, **, *** p<0.05, p<0.01, p<0.001 vs. PN 35. *, **, *** p<0.05, p<0.01, p<0.001 vs same sex control. †p < 0.05 vs. males.

Figure 2: Prepubertal castration increases sensitivity to social defeat in adulthood. A. Timeline of prepubertal castration and behavioral testing in adults. **B,C.** Social defeat decreased social approach and increased social vigilance in castrated males but not sham or no surgery controls (n=7-8 per group). **D,E.** No differences were observed during the acclimation of open field phase. **F.** Representative heatmaps for the interaction phase showing reduced time spent in the interaction zone castrated males exposed to social defeat. **, *** p<0.01, p<0.001 vs control.

Figure 3: Prepubertal castration increases BNST neural activity in sensitivity to social defeat in adulthood. A. Experimental timeline for fiber photometry observations of GCaMP6f in the ventral BNST of prepubertally castrated or sham surgery California mice. **B.** Photomicrographs of GCaMP in BNST at low (B) and high (B') magnification. The magenta box indicates position of the fiber. **C.** Representative images showing DeepLabCut tracking of mice in proximate and distant conditions. **D.** DeepLabCut tracking of body midpoint and nose of the focal mouse was used to determine the orientation of the focal mouse to the target mouse nose (green triangle).

GCaMP6f signals were significantly stronger when the focal mouse was within 8 cm of the target mouse but only when the target mouse was in central vision (0°≥40°) or peripheral vision (40°≥100°). **E.** Nose-to-nose sniffing with a non-aggressive target mouse induced a delayed increase in ΔF/F in castrated but not sham males. In contract nose-to-nose sniffing with an aggressive target mouse increased ΔF/F in both sham and castrated males. **F.** There were no changes in ΔF/F following bouts of freezing. **G.** Anogenital sniffing of non-aggressive target mice increased ΔF/F in castrated but not sham males. **H.** When castrated males were attacked by aggressive target mice, increased ΔF/F was observed. Sham males were not attacked enough for analysis. * p< 0.05, **p<0.01 vs sham. *** p<0.001 vs proximate, †p<0.05 vs. baseline. ac= anterior commissure.

Figure 4: Sexual differentiation of oxytocin-dependent circuits of social avoidance occurs during puberty. A. Experimental timeline for immunohistochemistry analyses in prepubertally castrated or intact California mice. **B.** Overlay of oxytocin (green) and c-fos (magenta) immunostaining in the paraventricular nucleus. Arrows indicate colocalizations in oxytocin (B') and c-fos (B") images. **C.** In the anterior PVN social defeat increased colocalizations (n=6-7 per group) in prepubertally castrated males but not intact males(C') and that social approach was negatively correlated with oxytocin/c-fos colocalizations (C"). **D.** In the posterior PVN castration increased oxytocin/c-fos colocalizations regardless of stress status (D') and social approach was negatively correlated with colocalizations. E. In anteromedial BNST, where oxytocin receptors drive social avoidance, prepubertal castration increased c-fos immunoreactivity regardless of stress status (E') and c-fos positive cells were negatively correlated with social approach (E"). **F.** Experimental timeline for examining effects of oxytocin receptors on social behavior. **G,H.** In both males and females exposed to social defeat an i.p. injection of 5 mg/kg of the oxytocin receptor antagonist L368,899 increased social approach and decreased social vigilance (n=7-9 per group). *, ** p<0.05, control/vehicle. †p < 0.05 vs. intact.

Figure 5: Androgen treatment at puberty reduces effects of social defeat on social approach and vigilance in males and females. A. Experimental timeline for surgery, social defeat, and social interaction testing in males. **B**. Castrated males treated with dihydrotestosterone (DHT) or testosterone (T) implants had higher social approach than males treated with empty implants (n=7-8 per group). **C**. Treatment with DHT or T also lowered social vigilance. **D**. Experimental timeline for surgery, behavior testing, and social defeat in female California mice. **E**. Hormone treatment had no effect on social approach before stress exposure but after stress exposure DHT treated females showed more social approach than females treated with empty implants or no surgery controls (n=8-10 per group). **F**. Similarly there were no differences in social vigilance in before stress but after stress social vigilance increased in no surgery control and empty implant females but not DHT females. *p<0.05, ** p<0.01, *** p<0.001 vs. empty implant. & p<0.05 vs T. †p < 0.05 vs. control (pre-stress).