Lots of movement, little progress: A review of reptile home range literature

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Abstract

Reptiles are the most species-rich vertebrate group with a broad diversity of life history traits. Biotelemetry is an essential methodology for reptiles as it compensates for several limitations when studying their natural history. We evaluated trends in reptile spatial ecology studies focusing upon quantifying home ranges for the past twenty years. We assessed 290 English-language reptile home range studies published from 2000-2019 via a structured literature review investigating publications' study location, taxonomic group, methodology, reporting, and analytical techniques. Substantial biases remain in both location and taxonomic groups in the literature, with nearly half of all studies (45%) originating from the USA. Snakes were most often studied, and crocodiles were least often studied, while testudines tended to have the greatest within study sample sizes. More than half of all studies lacked critical methodological details, limiting the number of studies for inclusion in future meta-analyses (55% of studies lacked information on individual tracking durations, and 51% lacked sufficient information on the number of times researchers recorded positions). Studies continue to rely on outdated methods to quantify space-use (including Minimum Convex Polygons and Kernel Density Estimators), often failing to report subtleties on decisions that have substantial impact on home range area estimates. Moving forward researchers can select a suite of appropriate analytical techniques tailored to their research question (dynamic Brownian Bridge Movement Models for within sample interpolation, and autocorrelated Kernel Density Estimators for beyond sample extrapolation). Only 1.4% of all evaluated studies linked to available and usable telemetry data, further hindering scientific consensus. We ultimately implore

herpetologists to adopt transparent reporting practices and make liberal use of open data platforms to maximize progress in the field of reptile spatial ecology.

Keywords: home range, reptiles, open science, reproducibility, biotelemetry, space use, spatial ecology

Introduction

There are at least 11,242 described reptile species worldwide (Uetz, Freed & Hošek, 2020; accessed 2020-04). Reptiles typically have narrower niche requirements and smaller ranges than other vertebrates such as birds and mammals, leaving them increasingly susceptible to threats (Böhm et al. 2013). Nearly one in five reptilian species are threatened with extinction, and almost one in four assessed species are Data Deficient (Böhm et al. 2013). Data deficiency in reptiles (21%) is higher than that of bird and mammal species (<1% and 15%, respectively; Stattersfield, Bennun, & Jenkins, 2004; Schipper et al. 2008); in particular for tropical reptiles and those with fossorial habits. Recent collapses of snake diversity have been reported with rippling effects to the ecosystem (Zipkin et al. 2020), but baseline data is often unavailable to properly evaluate these events and is likely understating the cascading effect of disappearing species (Roll et al. 2017). Targeted conservation action requires information and knowledge that is lacking for many reptile species (Tingley, Meiri, Chapple, 2016; Etard, Morrill & Newbold, 2020).

Efforts to collect baseline data are frustrated by reptiles' nature history –often small, rare, and cryptic– limiting detection probabilities during visual count surveys. Telemetry studies can counteract low detection probability (or at least provide baseline detection estimates; Boback et al., 2020), as we know exactly the number, identity, and location of radio-marked individuals in the study site—with many potential applications (Refsnider et al., 2011). By tracking animal movement, we gain valuable insight into habitat requirements, foraging strategies, and behaviour (Kingsbury & Robinson 2016).

Radio-telemetry (VHF) is common in reptile research, whereas the use of GPS and other automated telemetry technology in terrestrial reptiles is still relatively rare (e.g., Hart et al. 2015, Smith et al. 2018) when compared to other taxa (Joo et al. 2020). Using novel technologies and the resulting increased data volume in telemetry studies should also increase the analytical method

complexity and encourage greater uptake of movement-based methods. However, the field of movement ecology has stagnated as the proportion of studies using movement-based methods is not matching the available software tools and methods (Joo et al. 2020).

The term "home range" is frequently and irrespectively applied to two distinct concepts: 1) the Burt (1943) home range definition, i.e., the area an animal uses for all of its lifetime activities, 2) *within sample* "space-use" (still commonly referred to as a "home range"), i.e., an area used by an animal throughout the study period duration. While both concepts have biological value, the chosen research question should govern choice of concept, and thus the space-use estimation methods researchers should use to answer their question. Researchers often use terms like "seasonal home range" to estimate animal space-use within the study period (Viana et al., 2018), delineating boundaries of interest based on season (Korbelová et al., 2016). Many studies improperly use the term home range (which by definition will include areas the animal will use outside the study period, i.e., *beyond sample*), when they intend to estimate overall space-use of their animals during the study period.

We treat reptile home range studies as any study intending to quantify space-use, regardless of whether the intent was to estimate areas used outside of the study or bounded by the study period. Initially, geometric methods such as the Minimum Convex Polygon (MCP) were the norm, but subsequently researchers have turned towards statistical techniques incorporating underlying probabilistic models, such as Kernel Density Estimators (KDE; Worton 1989). The autocorrelated nature of movement data violates traditional KDEs assumptions lead to the development of movement-based methods: autocorrelated KDEs (Fleming et al. 2015), and Brownian Bridge Movement Models (Horne et al. 2007; Kranstauber et al. 2012). Although researchers have continued to expand and develop analytical methods within movement and spatial ecology (Laver & Kelly 2008), the proportion of studies using movement-specific methods has not increased (Joo et al. 2020). Biotelemetry driven home range work has the potential to fill many gaps in reptile natural history information, and is a precursor to robust population estimates, but it is fraught with difficulty. Macartney, Gregory, & Larsen (1988) summarized the landscape of snake home range studies and suggested developing useful baseline data for comparative purposes requires longer-term studies and standardized data collection, analysis, and presentation.

In 1990, a general review found most studies focused on mammals and used MCPs to estimate home ranges (Harris et al. 1990). By 2008, the same patterns were still present, with mammalian and ornithological home range studies most prevalent, and with 96 out of 141 studies still utilizing MCPs (51% utilizing both MCPs and KDEs; Laver & Kelly 2008). Goldingay (2015) reviewed home-range studies for Australian terrestrial vertebrates between 2001-2012, and only 19% out of 150 papers pertained to reptiles, even though Australia has over 860 native reptile species; in comparison, 68% of studies were on mammals, which only correspond to 16% of Australian's land species. As for home range estimators, the MCPs appeared in 84% of these studies, followed by KDEs (45%), illustrating a lack of methodology advancement despite a growing field.

Here, we reviewed reptile telemetry literature to assess whether the field showed a similar trajectory of increasing data volume and shifts in collection methods (e.g., GPS and satellite tags), and a continuing stagnation in home range estimation techniques. We also sought to reveal underlying reptile home range study biases, both geographically and taxonomically, to determine future limitations in undertaking global syntheses and analyses. As most home range estimates are sensitive to study design and data collection protocol (e.g., number of locations and duration), we also evaluated reptile telemetry studies within the framework of open, reproducible, and comparable science, to determine the number of available datasets from our review. Finally, we make recommendations for improving reporting standards to aid in making reptile home range studies more broadly applicable and reproducible.

Survey methodology

We performed a comprehensive literature review by searching in Google Scholar, Web of Science, and Scopus on January 30th, 2020 for articles relating to reptile spatial ecology using the terms ("*reptile*" OR "*tortoise*" OR "*crocodile*" OR "*alligator*" OR "*snake*" OR "*lizard*") AND ("*home range*" OR "*home-range*" OR *"space use*" OR "*spatial ecology*"). We limited the search to papers from 2000-2019 published in peer-reviewed journals. Following Haddaway et al., (2015) we only included the first 300 results from Google Scholar.

Our aim was solely terrestrial/semi-terrestrial reptile home range studies, so we excluded studies on marine species (e.g., sea turtles, sea snakes). However, we did include studies from semi-aquatic or typically range-limited to waterway species (e.g., crocodilians, freshwater turtles). We excluded studies lacking home range or space-use estimates, such as those that only used

movement measurements. As multiple field sampling techniques can generate home ranges, we defined our inclusion criteria as only studies using an attached telemetric device (e.g., VHF transmitters, GPS). We further excluded clear re-analyses of previously published datasets to avoid pseudoreplication. In these cases, we included only the oldest published article returned from the systematic search for review. We did however include studies pooling previous data with newly collected data.

When studies included multiple species, we considered the overall methodology rather than for each species individually to avoid pseudoreplication. We only collected multiple values for a study's methodology if researchers used two different tracking devices (e.g., both VHF and GPS), as different tracking devices are subject to different limitations in data recording. We used the distinct biotransmitter type to review tracking protocols (e.g., sample frequency, number of locations) and trends in biotransmitter selection (e.g., VHF vs GPS), but used study level effort to review geographic and taxonomic patterns. From each included paper, we collected basic study information (country, year, species, number of individuals tracked) as well as more detailed information about the data sampling regime and home range estimation methods. To assess the field sampling protocols, we collected data concerning the reporting of tracking duration, number of locations, and tracking frequency (number of fixes per day) for studied individuals.

Regular temporal sampling is an assumption in several movement analyses, so we also identified whether studies conducted regular sampling. We defined two cases of regular temporal sampling: (1) whether individuals were located at least once per day consistently throughout the study, (2) whether sampling occurred at an equal hourly sampling rate (e.g., one fix every 2 hours, one fix every 15 min). We converted the reported tracking methodology into the number of tracks per day, recording both the minimum possible and maximum possible frequency. In cases where authors used ambiguous language (e.g., biweekly), or provided insufficient detail, we classified the tracking frequency as "not reported". We also documented whether the study used multiple regular tracking regimes (e.g., tracking once per day in the summer months, and only weekly during the winter).

We coded each article’s adherence to two key reporting characteristics that can impact space-use and home range estimates: tracking time duration, and number of fixes. Tracking time duration differs from study duration, and refers to the period of time over which researchers tracked an

individual. In contrast, study duration is the overall study period, and thus represents a study-level characteristic, while tracking duration represents an individual-level characteristic. We scored articles on a scale of zero to three. For example, zero indicated reporting only study duration/study-level number of fixes (e.g., "...tracked individuals from 2018-01-01 to 2018-09-23..." or "...collected a total of 356 fixes...") while failing to report the exact data quantity per individual (See Table 1 for details).

Table 1. Scoring category definitions for both number of location and study durations.

Data Field Type	Score of 0	Score of 1	Score of 2	Score of 3
Duration Reproducibility	*No reporting* – includes cases when authors claim at least ## days without a maximum	*Population-level reporting* – mean only *or* sum of duration (as number of days or number of weeks)	*Population-level reporting* – Mean only *or* sum of duration + a metric of spread (such as standard deviation or error)	*Individual-level reporting* – actual date ranges or number of days for each individual included in the paper.
Location Reproducibility	*No reporting* – includes cases when authors claim at least ## locations without a maximum	*Population-level reporting* – mean only *or* sum of number of locations.	*Population-level reporting* – Mean only *or* sum of number of locations + a metric of spread (such as standard deviation or error)	*Individual-level reporting* – actual number of times each individual was located during the study.

For each included study, we also recorded the method for estimating home range area. Kernel Density Estimation is a common technique but is highly dependent on the smoothing factor (*h*) selection method. To address this, we collected the method used to determine the *h*-value for KDEs (when reported). We recorded whether the authors reported a movement metric based on time (e.g., mean daily displacement), as field sampling regime can also affect such metrics. Finally, we recorded if the study attempted to "validate" the home range estimation —i.e., included any form of analysis that assessed the relationship between number of locations and the home range estimation (e.g., linear regressions, bootstrapped asymptotes).

Data and software availability

We use *R v.3.6.3* (R Core Team, 2020) and *RStudio v.1.4.1029* (RStudio Team, 2020) to summarise all data. We summarised data with *dplyr v.1.0.2* (Wickham et al., 2020), *raster v.3.4.5* (Hijmans, 2020), *forcats v.0.5.0* (Wickham, 2019a), *reshape2 v.1.4.4* (Wickham, 2007), and *stringr v.1.4.0* (Wickham, 2019b) packages, and visualised data with *cowplot v.1.1.0* (Wilke, 2019), *ggplot2 v.3.3.2* (Wickham, 2016), *ggpubr v.0.4.0* (Kassambara, 2018), *ggrepel v.0.8.2* (Slowikowski, 2018), *ggridges v.0.5.2* (Wilke, 2018), and *scico v.1.2.0* (Pedersen & Crameri, 2018). We have included all data and summary code to create manuscript materials at Zenodo (We have included all data and summary code to create manuscript materials at Zenodo (http://doi.org/10.5281/zenodo.4303644). Data file "reptileHRReview_References.csv" includes the results of the stages of systematic review alongside article information, the data file "reptileHRReview_LiteratureReview.csv" contains the raw data from the literature review process, metadata file "reptileHRReview_Metadata.csv" includes full descriptions of all columns in both main data files. An additional file from Reptile Database (Uetz, Freed & Hošek, 2020; accessed 2020-04) "reptileChecklist_2020_04.csv" includes the information used for genus- and clade-based summaries. We created the reptile diversity using Global Assessment of Reptile Distributions (GARD) data (Roll et al., 2017) and functionality from *sf v.0.9.6* (Pebesma, 2018) and *fasterize v.1.0.3* (Ross, 2020) packages. We counted terms used and produced the word cloud using *pdftools v.2.3.1* (Ooms, 2019) and *quanteda v.2.1.2* (Benoit et al., 2018).

Results

Data collection

From 1,028 unique articles returned from the literature searches (Figure S1), our exclusion criteria produced a final sample of 290 reptile spatial ecology studies consisting of 303 tracking subsets (accounting for multiple tracking protocols, i.e., GPS and VHF, within each study) involving 7,897 individual animals. However, one study failed to report the number of animals tracked. The majority of studies used VHF telemetry devices (278), with only 22 using GPS, and a further three instances of ultrasonic or satellite tracking. Regardless of the tracking method used, tracking frequency varied dramatically (Figure 1): ranging from 480 (24 if automated VHF is excluded) to 0.0328 tracks per day for VHF, and 144 to 0.143 per day for GPS. In other words, tracking 0.0328 times per day is equivalent to tracking approximately once a month (i.e., 1/30.5), and 0.143 times

per day is the equivalent to tracking once per week (i.e., 1/7). Ninety-two tracking subsets (30.4%) had consistent tracking frequencies throughout the study (i.e., minimum and maximum tracking frequency are the same, with no seasonal variation or multiple tracking regimes). The number of tracks/fixes per day was not always reported (*n* = 26), or reported ambiguous maximum and minimum number of tracks per day (e.g., "bi-weekly", "at least"); this number increases to 79 tracking subsets that failed to clearly report one extreme of the tracking frequency. Such reporting is key when measurements of movement capacity are calculated, and 189 out of 290 studies reported a movement metric.

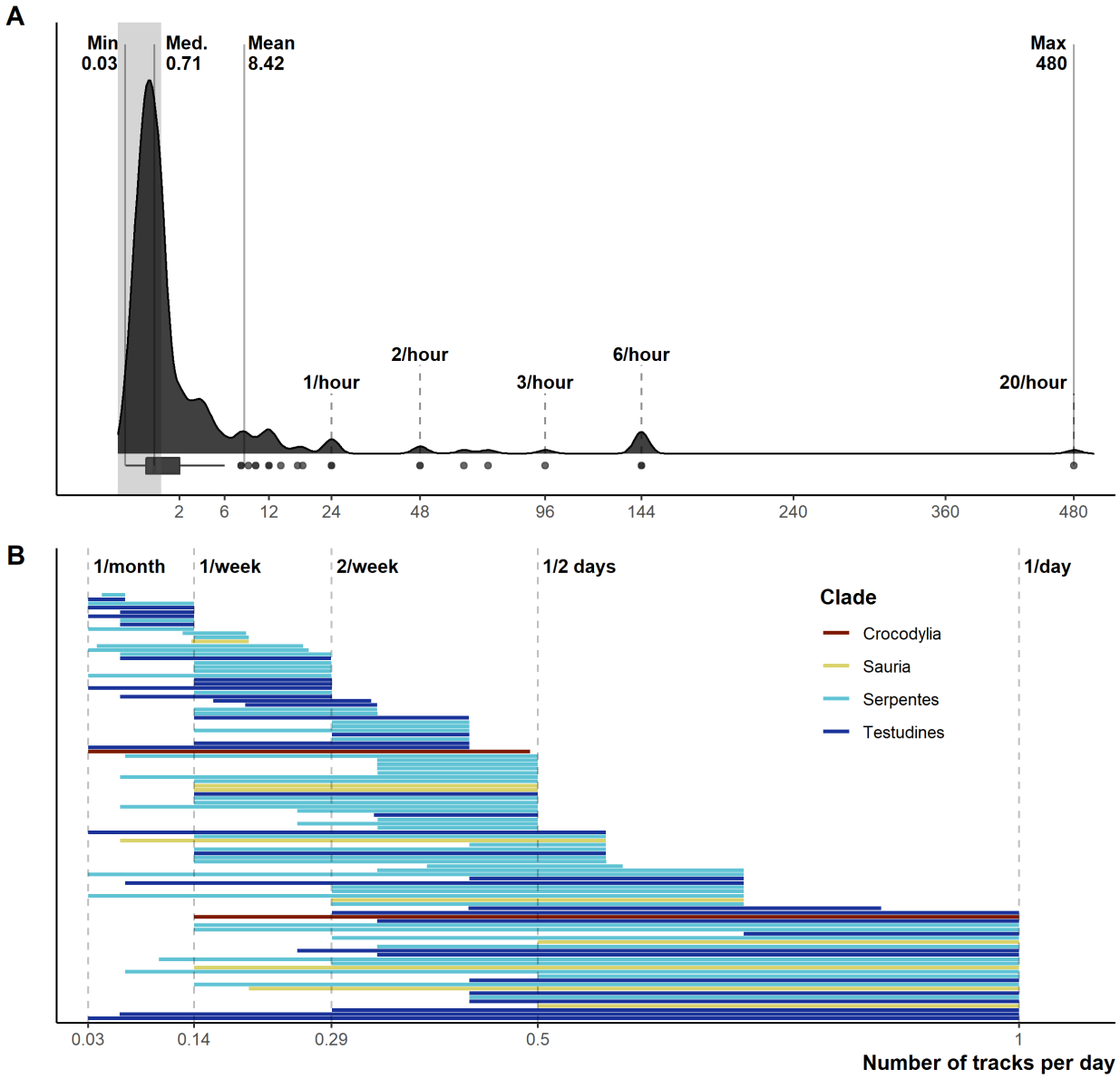

Figure 1. *Reported number of tracks per day.* ***A)*** *Density and boxplot plot showing the distribution of all reported tracking frequencies, x-axis is square-root transformed. Shaded grey area highlights instances of < 1 track per day. Text labels show the minimum, median, mean, and max numbers of tracks per day.* ***B)*** *The variation per study between the maximum and minimum tracking frequencies, provided both are < 1 per day. Both plots exclude all unreported tracking frequencies.*

Despite the extensive field effort expended tracking 7,897 animals, we identified serious gaps in basic reporting that undermine understanding basic study characteristics. In addition to the 26 instances of incomplete tracking frequency data, 162 studies provided very limited or missing descriptions of tracking duration (135 scored 0, 27 scored 1), and number of fixes obtained (95 scored 0, 52 scored 1, sum 147). Reporting standards of 2 and higher (i.e., likely sufficient to enable meta-analyses inclusion) were reached in 128 studies for durations and 143 for the number of fixes. Location reporting was further hindered by ambiguous terms, we found 34 different terms describing how many times an animal was tracked: studies largely used terms stemming from locat*, but even within a single study we often found multiple terms used to describe when researchers located animals (Figure S2). Providing raw data could mitigate reporting deficiencies; however, we found only 24 studies included links to external data and only 4 of those links led to raw tracking data.

Estimation methods

Between 2000-2019 the number of studies per year increased from 6 in 2000 to 18 in 2019, with a low of 4 in 2001 and a peak of 25 in 2017 (Figure 2A). Minimum Convex Polygons (MCP) and Kernel Density Estimations (KDE) use has dominated reptile home range studies for the past 20 years (272/290 studies; Figure 2B) and were present in over 75% of studies each year (Figure 2B). Frequently, studies include estimations from both methods, and rarely use KDEs without including MCPs (Figure S3). A minority of studies (n = 19) used “other” methods without pairing to estimations via MCPs and KDEs. These methods included: alpha-hull methods, harmonic means, linear home ranges, Brownian Bridge kernels among others (for full list see Table S1). Of all other methods listed, only dynamic (and standard) Brownian Bridge Movement Models directly incorporate movement to estimate space-use (i.e., movement models).

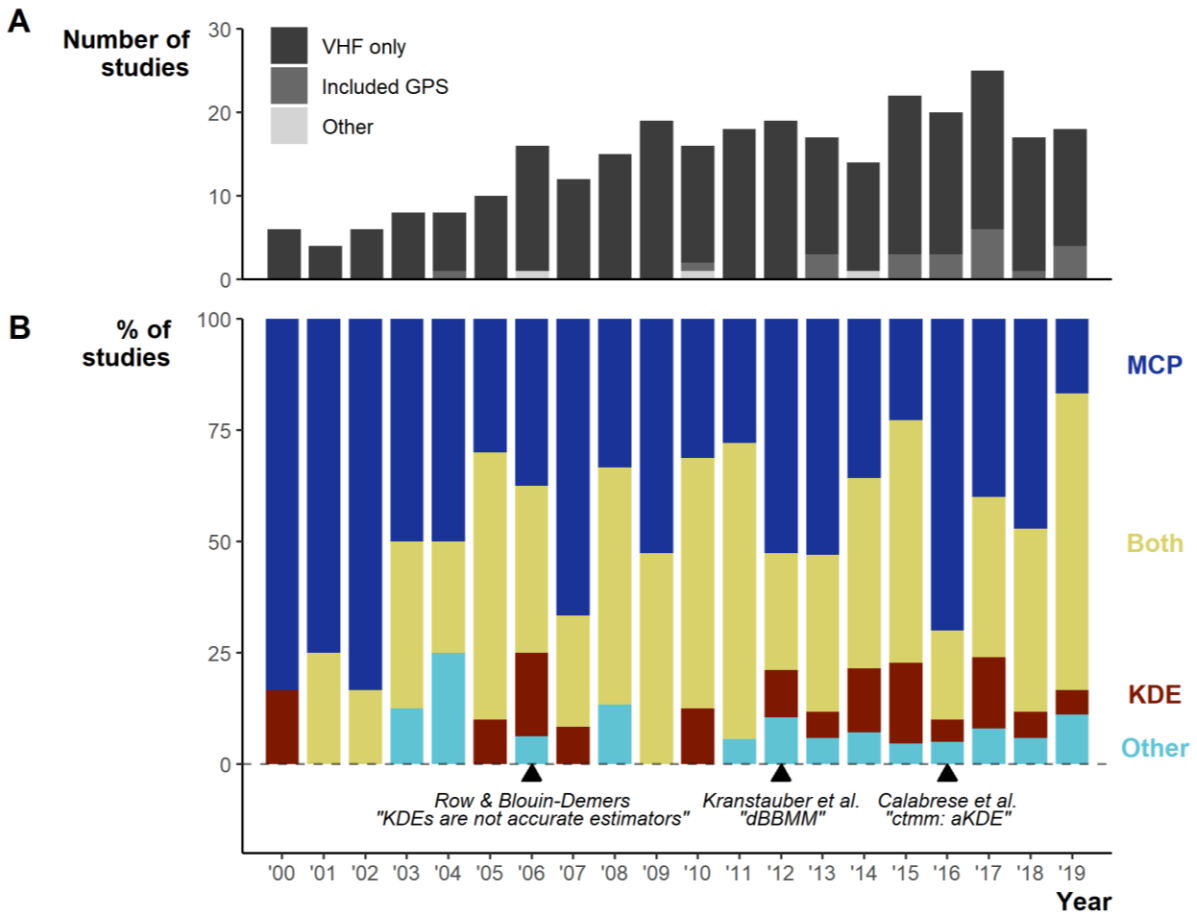

Figure 2. *Changes in the field from 2000 to 2019.* ***A)*** *Number of articles over per year and the telemetry devices used: dark grey = only VHF used, middle grey = GPS was used exclusively or in conjunction with VHF, light grey = other device used (ultrasonic and satellite).* ***B)*** *The percentage of studies using Minimum Convex Polygon (MCP), Kernel Density Estimations (KDE), both or other estimation methods. "Other" only includes studies that did not use either MCPs or KDEs. Lower text labels highlight the year select papers were published aiming to guide, or enable new, space-use estimation.*

Studies using MCPs largely made use of high % contours (100 and 95%; Figure S4). KDEs used a greater diversity of contour values (5 to 100%), but with clear concentration towards 95% and 50%. Studies more frequently (n = 97/270 studies using MCPs) failed to report the contour used with MCPs than other methods, potentially connected to the assumption that MCPs default to 100%.

For studies using KDEs, we found 14 smoothing factor selection methods, but researchers primarily used Least Squares Cross-validation (LSCV; 73/159; Figure S5). Similar to basic reporting, we show that 27 (17.4%) studies failed to report a smoothing factor, either by omission or by only stating the "default" for a software.

Geographic and taxonomic biases

The United States of America is a clear hotspot with 133 of 290 studies. All other countries are dramatically lower (<8 studies, 30 countries with a single study), with only Australia (35), Canada (22), and South Africa (12) breaking the trend. Despite high reptile diversity, Africa exhibited a dearth of reptile home range papers (Figure 3).

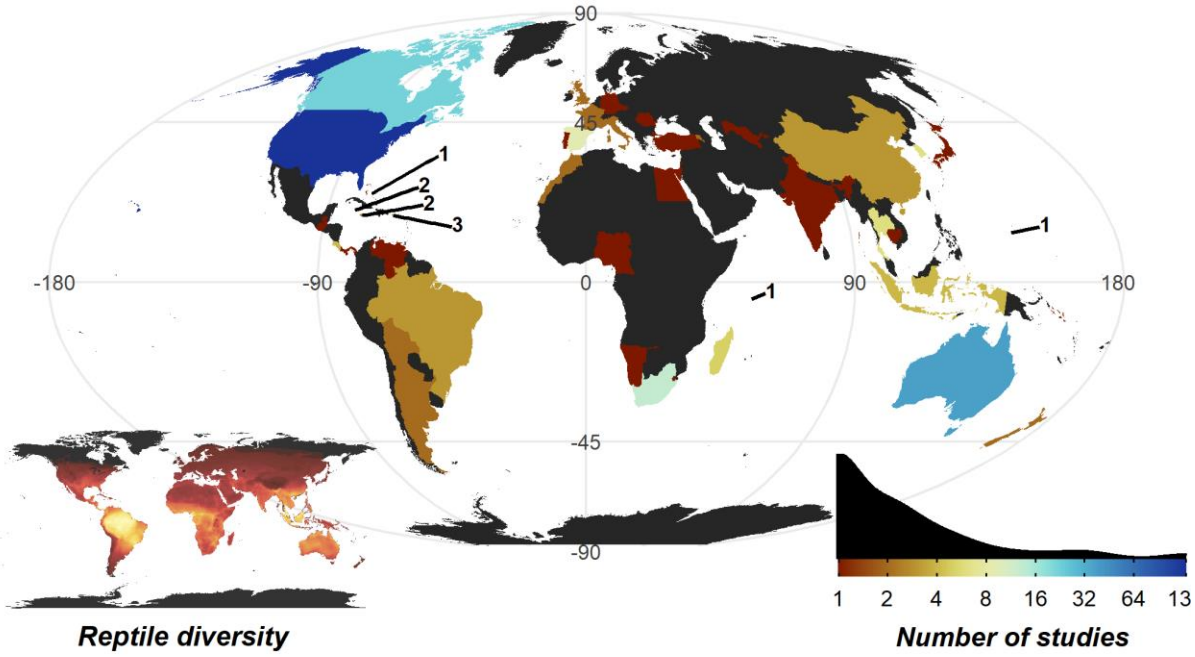

Figure 3. *Number of studies undertaken in each country. Insert map bottom left, shows the distribution of reptile species globally, ranging from zero species (black) to 182 species (yellow); the heatmap was generated using GARD data (Roll et al., 2017). Insert density plot bottom right, shows the distribution of per country study counts. Count of studies is shown on a log scale to help differentiate between countries with fewer studies. Smaller territories are highlighted with a label denoting the number of studies.*

The 7,897 tracked individuals, 303 tracking subsets, and tracking subset sample sizes were not split evenly across the major clades of Crocodylia (mean individuals per subset = 9.32 ± 1.48), Serpentes (22.3 ± 1.61), Sauria (28.4 ± 3.53), and Testudines (35.4 ± 6.80; Figure 5). Serpentes was the most studied clade and with the most tracked individuals, whereas Crocodylia was the lowest. However, in terms of percentage of genera studied, Crocodylia leads with 44.4% (4/9; Testudines 27/94, 28.7%; Serpentes 40/522, 7.66%; Sauria 28/564, 4.96%).

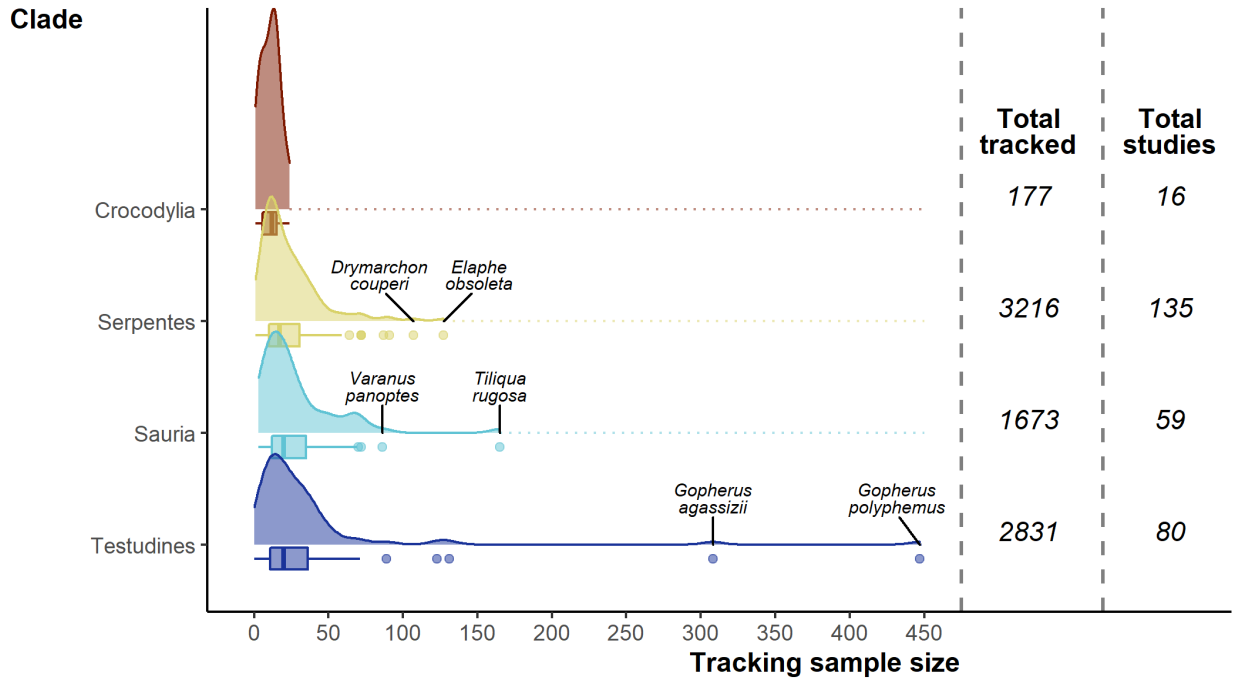

Figure 4. *Density and box plots showing the distribution of sample sizes (tracked individuals) per study by clade. Species names highlight the top two outlying sample sizes for clades other than Crocodylia.*

Overall, of the 1,210 reptile genera (Uetz, Freed & Hošek, 2020; accessed 2020-04), 99 (8%) have been tracked (but there are genera untrackable with current telemetry equipment, e.g., *Amphisbaena* and *Ramphotyphlops*). Two genera (*Crotalus* & *Gopherus*) stand out having been studied 22 and 23 times (Fig S6), whereas 45 genera had only a single study.

Discussion

We identified key issues limiting study comparability. Study design decisions on tracking frequency and duration are critical considerations when attempting to produce biologically relevant space-use estimates (Girard et al., 2002; Börger et al., 2006; Silva et al., 2020). These decisions determine total individual sampling effort; an individual with 12 locations over a single day is unequal to one with 12 locations over an entire year. Reporting solely the study duration (e.g., stating that tracking occurred between X and Y date) would then obscure individual variation—further hindering our ability to generalize across the study population. By reporting at the individual level, researchers can highlight potential sources of heterogeneity between studies (e.g., tracking an animal once every week while tracking others twice a week would hinder daily movement comparisons) (Alexander & Maritz, 2015; Riotte-Lambert & Matthiopoulos, 2019). As

we found a wide range of tracking frequencies throughout reptile spatial ecology studies, it limits our ability to conduct large inter-study comparisons and undermines their validity.

Our review also reveals major biases in the study of reptile home ranges. Geographically, nearly 50% of studies originated from a single country, the United States of America. We found a stark mismatch between reptile diversity and reptile home range study locations (Roll et al., 2017), reflecting similar gaps seen in reptile abundance studies (Doherty et al., 2020); in particular, the Middle East and Central Africa. Taxonomically, we observed less severe biases, but should still be considered in evaluating the patterns in the available data. Only 8% of genera have been studied and the groups studied are far from a random sample; a relatively small number of genera dominate the available reptile spatial ecology data (e.g., *Gopherus, Crotalus, Pituophis*). Efforts to synthesise reptile home range or movement must recognize that any results may be biased towards patterns in temperate western hemisphere species, rather than global trends. Global syntheses may be inhibited by the drastic differences in seasonal climate between well-studied temperate areas and neglected tropical regions, which is likely key to reptiles as ectotherms (Shine and Madsen, 1996).

Many of the issues that we revealed in the reptile spatial ecology literature can be mitigated with greater transparency, adopting open science and reproducible analyses (i.e., code-based analysis avoiding language ambiguities by being, and describing, the exact analytical procedure performed; Ince, Hatton & Graham-Cumming, 2012; Archmiller et al., 2020). Open science presents a vital resource for replication efforts and can facilitate better meta-analyses. It also benefits the original researchers by increasing citations, boosting publication chances, and creating more potential for collaborations (Piwowar & Vision, 2013; Markowetz, 2015; Allen & Mehler, 2018). The disparity between reptile data and other taxa on prominent movement data repositories re-emphasizes our review findings (when searching either "*reptilia*" or "*reptile*", 24 reptile studies have available movement data on MoveBank as of 12.02.2020, and only 11 of those studies focus on terrestrial species). Ecology journals (and herpetology journals especially; Marshall & Strine, 2020) should redouble efforts to enforce data availability statements (Roche et al., 2015), making data availability the default and refusing to accept "on request" statements (Aalbersberg et al., 2018). Researchers can make use of free data repositories (movement specific like MoveBank, or generic like Zenodo or OSF) to ease this process. We hope the opening of reptile movement data can

facilitate broader studies similar to those undertaken in avian and mammalian fields (e.g., Tucker et al., 2019; Noonan et al., 2020).

Researchers often justify using KDEs and/or MCPs to compare with the wider reptile spatial ecology literature. However, methodological choices in reptile space-use studies hinder inter-study comparisons, as KDEs and MCPs are sensitive to differences in sampling effort (e.g., number of locations, tracking duration and frequency) (Mitchell, White, & Arnold, 2019; Silva et al. 2020). Reptile studies also used a wide range of smoothing factors for KDEs, which can also result in considerable home range over- or underestimations (Bauder et al. 2015; Silva et al. 2020). For example, two widely used smoothing factors, h_{ref} and LSCV, produce dramatically different area estimations. Failure to report or account for smoothing factors is thus a major concern, as it would significantly alter meta-analysis patterns. Although Row & Blouin-Demers (2006) suggested MCPs over KDEs for home range size comparisons across groups or time periods, MCP and KDE comparability is unreliable rendering their use generally inappropriate for most ecological studies (Nilsen, Pedersen, & Linnell, 2008; Silva et al. 2020).

There is a growing body of work demonstrating the versatility of newer analytical methods (Noonan et al., 2018), and how they can be applied to the coarser and zero-inflated radio-telemetry data (Averill-Murray, Fleming, & Riedle, 2020; Hromada et al. 2020; Silva et al. 2020). Reptile spatial ecology so far has largely failed to capitalise on the wealth of analytical options available, namely integrating movement information explicitly into estimations of space-use. Unlike traditional estimation methods (KDEs and MCPs), movement-based models do not operate under the assumptions breached by tracking data (independence of points) and guard better against under- and overestimation (Fleming & Calabrese 2017; Silva et al. 2020). One of the common solutions to autocorrelation is the thinning of data; this procedure is inherently wasteful and inefficient, defeating the purpose of collecting high temporal-resolution data and reducing the biological significance of telemetry datasets (Fleming et al. 2015; Calabrese et al. 2020). With low temporal-resolution data, analytic approaches will not necessarily reveal the correct home range patterns and need to be applied with caution; in these cases, it may be necessary to reconsider our research questions or re-evaluate study design for additional data collection.

Researchers should aim to explore the estimation sensitivity choices (Signer & Fieberg, 2020), while ensuring that their method matches their question. Conceptualizing home range as *within*

sample versus *beyond sample* space-use requires distinguishing between *occurrence distribution* versus *range distribution* methods (Fleming et al. 2015; Horne et al. 2019; Figure 5). While *occurrence distributions* (e.g., dBBMMs) allow us to answer research questions regarding the actual movement trajectory of an animal (and its confidence region), *range distributions* (e.g., AKDEs) consider the processes underlying animal movements and long-term space-use (Horne et al. 2019). Some research questions investigated in the reptile home range literature are actually targeting *within sample* space-use, requiring no extrapolation beyond the sampling period. In many cases, the sampling duration was too short to confidently identify range stability, which is a prerequisite for *beyond sample* home range estimates. To help unify the terminology used in reptile spatial ecology studies we draw attention to definitions from existing literature and reiterate them in Figure 5.

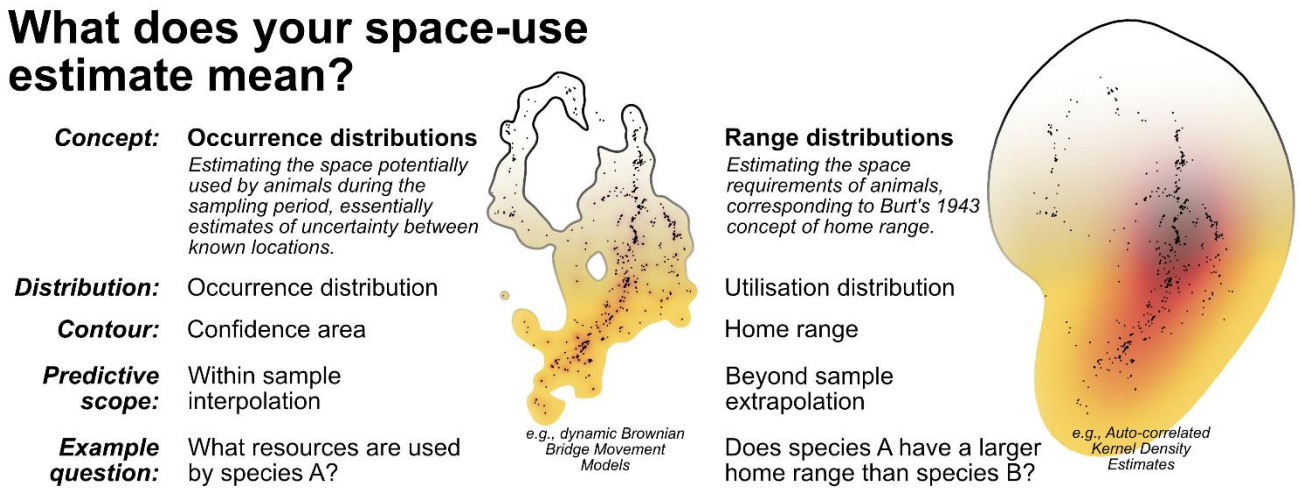

Figure 5. *A breakdown of the two complimentary conceptualisations of animal space-use.*

Ambiguous language further compounded reporting issues. Failing to report estimation methods (or reporting with ambiguous or ill-defined acronyms) and associated smoothing parameters completely undermines computational reproducibility and inter-study comparability. Relatively few studies failed to attempt reporting their tracking frequency; many of our 79 failures to determine minimum or maximum tracking frequency were a direct result of ambiguous language, such as using words with multiple definitions (e.g., “bi-weekly”) or using imprecise summaries (e.g., “at least”). We also found semantic ambiguity when describing locations. Studies used a wide range of terms to refer to locations, relocations, fixes, datapoints, etc., yet are selecting

contrasting or overlapping definitions for these terms. The key distinction for the definition of location is whether it refers each time researchers documented the animal's spatial position or whether it refers only to a unique spatial position (a movement from the previously recorded location) used by the study animal (often referred to as "relocations"). Standardising and unifying terminology is essential for creating widely useful methods and comparable databases (Schneider et al., 2019).

Answering specific questions requires appropriate protocols and, to draw broad inferences among a single study, those protocols must remain consistent. Between study comparisons also require consistency (or at least clear reporting on inconsistencies). The compound effect of geographical, taxonomical and methodological biases leads to inappropriate generalizations when ignored. Recent macroecological investigations have failed to account for varying tracking regimes and different estimation methods (Slavenko et al. 2016; Todd & Nowakowski 2020). To facilitate detailed reporting of tracking datasets, we have supplied an example report based on an existing tracking dataset (Marshall et al., 2020; Supp. File 1). This example aims to provide a foundation for transparent reporting of sample size, study duration, number of datapoints, as well as important aspects used to describe the tracking regime: namely plots that describe individual tracking durations (while highlighting deviation from proposed tracking protocols), and distribution of time lags between tracks. We have supplied the code (as an .Rmd file, Supp. File 2) and data (as a .csv, Supp. File 3) used to generate the report as supplementary material.

Conclusion

The past 20 years have seen a growing number of reptile home range studies and continued reliance on traditional but outdated methods, Kernel Density Estimations (KDEs) and Minimum Convex Polygons (MCPs), for home range and space-use estimations, despite more appropriate available methods. Scientific conventions can be slow to shift, and often require substantial interdisciplinary research to move towards better alternatives (Smaldino & O'Connor, 2020). We appeal to researchers focusing on reptiles to engage with quantitative ecologists to ease the uptake of more complex and appropriate statistical methods and thus maximize the value of hard-won field data. The best way to facilitate broader engagement is to adopt more transparent practices by sharing and fully reporting collected data.

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Author contributions

All authors contributed equally to the manuscript.

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References

Aalbersberg IJ, Appleyard T, Brookhart S, Carpenter T, Clarke M, Curry S, Dahl J, DeHaven AC, Eich E, Franko M, Freedman L, Graf C, Grant S, Hanson B, Joseph H, Kiermer V, Kramer B, Kraut A, Karn RK, Lee C, MacFarlane A, Martone M, Mayo-Wilson E, McNutt M, McPhail M, Mellor DT, Moher D, Mudditt A, Nosek BA, Orland B, Parker TH, Parsons M, Patterson M, Santos S, Shore C, Simons DJ, Spellman B, Spies JR, Spitzer M, Stodden V, Swaminathan S, Sweet D, Tsui A, Vazire S. 2018. Making Science Transparent By Default: Introducing the TOP Statement. OSF Preprints. DOI: 10.31219/osf.io/sm78t.

Alexander GJ, Maritz B. 2015. Sampling interval affects the estimation of movement parameters in four species of African snakes: Sampling interval affects estimation of movement. *Journal of Zoology* 297:309–318. DOI: 10.1111/jzo.12280.

Allen CPG, Mehler DMA. 2018. Open Science challenges, benefits and tips in early career and beyond. PsyArXiv. DOI: 10.31234/osf.io/3czyt.

Archmiller AA, Johnson AD, Nolan J, Edwards M, Elliott LH, Ferguson JM, Iannarilli F, Vélez J, Vitense K, Johnson DH, Fieberg J. 2020. Computational Reproducibility in The Wildlife Society's Flagship Journals. *The Journal of Wildlife Management*:jwmg.21855. DOI: 10.1002/jwmg.21855.

Averill-Murray ROYC, Fleming CH, & Riedle JD. 2020. Reptile home ranges revisited: a case study of space use of Sonoran Desert tortoises (*Gopherus morafkai*). *Herpetological Conservation and Biology* 15(2), 253-271.

Bauder JM, Breininger DR, Bolt MR, Legare ML, Jenkins CL, McGarigal K. 2015. The role of the bandwidth matrix in influencing kernel home range estimates for snakes using VHF telemetry data. *Wildlife Research*, 42:437–453.

Benoit K, Watanabe K, Wang H, Nulty P, Obeng A, Müller S, Matsuo A. 2018. quanteda: An R package for the quantitative analysis of textual data. *Journal of Open Source Software* 3:774. DOI: 10.21105/joss.00774.

Boback SM, Nafus MG, Yackel Adams AA, & Reed RN. 2020. Use of visual surveys and radiotelemetry reveals sources of detection bias for a cryptic snake at low densities. *Ecosphere* 11(1), e03000.

Böhm M, Collen B, Baillie JEM, Bowles P, Chanson J, Cox N, Hammerson G, Hoffmann M, Livingstone SR, Ram M, Rhodin AGJ, Stuart SN, van Dijk PP, Young BE, Afuang LE, Aghasyan A, García A, Aguilar C, Ajtic R, Akarsu F, Alencar LRV, Allison A, Ananjeva N, Anderson S, Andrén C, Ariano-Sánchez D, Arredondo JC, Auliya M, Austin CC, Avci A, Baker PJ, Barreto-Lima AF, Barrio-Amorós CL, Basu D, Bates MF, Batistella A, Bauer A, Bennett D, Böhme W, Broadley D, Brown R, Burgess J, Captain A, Carreira S, Castañeda M del R, Castro F, Catenazzi A, Cedeño-Vázquez JR, Chapple DG, Cheylan M, Cisneros-Heredia DF, Cogalniceanu D, Cogger H, Corti C, Costa GC, Couper PJ, Courtney T, Crnobrnja-Isailovic J, Crochet P-A, Crother B, Cruz F, Daltry JC, Daniels RJR, Das I, de Silva A, Diesmos AC, Dirksen L, Doan TM, Dodd CK, Doody JS, Dorcas ME, Duarte de Barros Filho J, Egan VT, El Mouden EH, Embert D, Espinoza RE, Fallabrino A, Feng X, Feng Z-J, Fitzgerald L, Flores-Villela O, França FGR, Frost D, Gadsden H, Gamble T, Ganesh SR, Garcia MA, García-Pérez JE, Gatus J, Gaulke M, Geniez P, Georges A, Gerlach J, Goldberg S, Gonzalez J-CT, Gower DJ, Grant T, Greenbaum E, Grieco C, Guo P, Hamilton AM, Hare K, Hedges SB, Heideman N, Hilton-Taylor C, Hitchmough R, Hollingsworth B, Hutchinson M, Ineich I, Iverson J, Jaksic FM, Jenkins R, Joger U, Jose R, Kaska Y, Kaya U, Keogh JS, Köhler G, Kuchling G, Kumlutaş Y, Kwet A, La Marca E, Lamar W, Lane A, Lardner B, Latta C, Latta G, Lau M, Lavin P, Lawson D, LeBreton M, Lehr E, Limpus D, Lipczynski N, Lobo AS, López-Luna MA, Luiselli L, Lukoschek V, Lundberg M, Lymberakis P, Macey R, Magnusson WE, Mahler DL, Malhotra A, Mariaux J, Maritz B, Marques OAV, Márquez R, Martins M, Masterson G, Mateo JA, Mathew R, Mathews N, Mayer G, McCranie JR, Measey GJ, Mendoza-Quijano F, Menegon M, Métrailler S, Milton DA, Montgomery C, Morato SAA, Mott T, Muñoz-Alonso A, Murphy J, Nguyen TQ, Nilson G, Nogueira C, Núñez H, Orlov N, Ota H, Ottenwalder J, Papenfuss T, Pasachnik S, Passos P, Pauwels OSG, Pérez-Buitrago N, Pérez-Mellado V, Pianka ER, Pleguezuelos J, Pollock C, Ponce-Campos P, Powell R, Pupin F, Quintero Díaz GE, Radder R, Ramer J, Rasmussen AR, Raxworthy C, Reynolds R, Richman N, Rico EL, Riservato E, Rivas G, da Rocha PLB, Rödel M-O, Rodríguez Schettino L, Roosenburg WM, Ross JP, Sadek R, Sanders K, Santos-Barrera G, Schleich HH, Schmidt BR, Schmitz A, Sharifi M, Shea G, Shi H-T, Shine R, Sindaco R, Slimani T, Somaweera R, Spawls S, Stafford P, Stuebing R, Sweet S, Sy E, Temple HJ, Tognelli

MF, Tolley K, Tolson PJ, Tuniyev B, Tuniyev S, Üzüm N, van Buurt G, Van Sluys M, Velasco A, Vences M, Veselý M, Vinke S, Vinke T, Vogel G, Vogrin M, Vogt RC, Wearn OR, Werner YL, Whiting MJ, Wiewandt T, Wilkinson J, Wilson B, Wren S, Zamin T, Zhou K, Zug G. 2013. The conservation status of the world's reptiles. *Biological Conservation* 157:372–385. DOI: 10.1016/j.biocon.2012.07.015.

Börger L, Franconi N, De Michele G, Gantz A, Meschi F, Manica A, Lovari S, Coulson T. 2006. Effects of sampling regime on the mean and variance of home range size estimates. *Journal of Animal Ecology* 75:1393–1405. DOI: 10.1111/j.1365-2656.2006.01164.x.

Burt WH. 1943. Territoriality and Home Range Concepts as Applied to Mammals. *Journal of Mammalogy* 24:346–352. DOI: 10.2307/1374834.

Calabrese JM, Fleming CH, Noonan MJ, & Dong X. 2020. ctmmweb: A graphical user interface for autocorrelation-informed home range estimation. BioRxiv.

Doherty TS, Balouch S, Bell K, Burns TJ, Feldman A, Fist C, Garvey TF, Jessop TS, Meiri S, Driscoll DA. 2020. Reptile responses to anthropogenic habitat modification: A global meta-analysis. *Global Ecology and Biogeography* 29:1265–1279. DOI: 10.1111/geb.13091.

Etard A, Morrill S, Newbold T. 2020. Global gaps in trait data for terrestrial vertebrates. *Global Ecology and Biogeography*:geb.13184. DOI: 10.1111/geb.13184.

Fleming CH, & Calabrese JM. 2017. A new kernel density estimator for accurate home-range and species-range area estimation. *Methods in Ecology and Evolution* 8(5), 571-579.

Fleming CH, Fagan WF, Mueller T, Olson KA, Leimgruber P, & Calabrese JM. 2015. Rigorous home range estimation with movement data: a new autocorrelated kernel density estimator. *Ecology* 96(5), 1182-1188.

Girard I, Ouellet J-P, Courtois R, Dussault C, Breton L. 2002. Effects of Sampling Effort Based on GPS Telemetry on Home-Range Size Estimations. *The Journal of Wildlife Management* 66:1290. DOI: 10.2307/3802962.

Goldingay RL. 2015. A review of home-range studies on Australian terrestrial vertebrates: adequacy of studies, testing of hypotheses, and relevance to conservation and international studies. *Australian Journal of Zoology* 63(2), 136-146.

Hijmans RJ. 2020. raster: Geographic data analysis and modeling.

Haddaway NR, Collins AM, Coughlin D, Kirk S. 2015. The Role of Google Scholar in Evidence Reviews and Its Applicability to Grey Literature Searching. *PLOS ONE* 10:e0138237. DOI: 10.1371/journal.pone.0138237.

Harris S, Cresswell WJ, Forde PG, Trewhella WJ, Woollard T, Wray S. 1990. Home-range analysis using radio-tracking data–a review of problems and techniques particularly as applied to the study of mammals. *Mammal Review* 20:97–123. DOI: 10.1111/j.1365-2907.1990.tb00106.x.

Hart KM, Cherkiss MS, Smith BJ, Mazzotti FJ, Fujisaki I, Snow RW, Dorcas ME. 2015. Home range, habitat use, and movement patterns of non-native Burmese pythons in Everglades National Park, Florida, USA. *Animal Biotelemetry* 3:8.

Horne JS, Garton EO, Krone SM, Lewis JS. 2007. Analyzing animal movements using Brownian bridges. *Ecology* 88(9), 2354-2363.

Horne EJS, Fieberg J, Börger L, Rachlow JL, Calabrese JM, Fleming CH. 2019. Animal Home Ranges. in Murray DL, & Sandercock BK (Eds.). 2020. *Population Ecology in Practice*. John Wiley & Sons.

Hromada SJ, Esque TC, Vandergast AG, Dutcher KE, Mitchell CI, Gray ME, Chang T, Dickson BG, Nussear KE. 2020. Using movement to inform conservation corridor design for Mojave desert tortoise. *Movement Ecology* 8(1), 1-18.

Ince DC, Hatton L, Graham-Cumming J. 2012. The case for open computer programs. *Nature* 482:485–488. DOI: 10.1038/nature10836.

Joo R, Picardi S, Boone ME, Clay TA, Patrick SC, Romero-Romero VS, & Basille M. 2020. A decade of movement ecology. arXiv preprint arXiv:2006.00110.

Kassambara A. 2018. ggpubr: “ggplot2” based publication ready plots.

Kingsbury BA, & Robinson NJ. 2016. Movement patterns and telemetry. Reptile Ecology and Conservation: A Handbook of Techniques, 110.

Kranstauber B, Kays R, LaPoint SD, Wikelski M, & Safi K. 2012. A dynamic Brownian bridge movement model to estimate utilization distributions for heterogeneous animal movement. *Journal of Animal Ecology* 81(4), 738-746.

Laver PN, & Kelly MJ. 2008. A critical review of home range studies. *The Journal of Wildlife Management* 72(1), 290-298.

Macartney JM, Gregory PT, & Larsen KW. 1988. A tabular survey of data on movements and home ranges of snakes. *Journal of Herpetology* 61-73.

Markowetz F. 2015. Five selfish reasons to work reproducibly. *Genome Biology* 16:274. DOI: 10.1186/s13059-015-0850-7.

Marshall BM, Strine CT. 2020. Make like a glass frog: In support of increased transparency in herpetology. OSF Preprints. DOI: 10.31219/osf.io/74frd.

Marshall BM, Crane M, Silva I, Strine CT, Jones MD, Hodges CW, Suwanwaree P, Artchawakom T, Waengsothorn S, Goode M. 2020. No room to roam: King Cobras reduce movement in agriculture. Movement Ecology 8:33. DOI: 10.1186/s40462-020-00219-5.

Mitchell LJ, White PC, & Arnold KE. 2019. The trade-off between fix rate and tracking duration on estimates of home range size and habitat selection for small vertebrates. *PloS One* 14(7), e0219357.

Nilsen EB, Pedersen S, & Linnell JD. 2008. Can minimum convex polygon home ranges be used to draw biologically meaningful conclusions?. *Ecological research* 23(3), 635-639.

Noonan MJ, Fleming CH, Tucker MA, Kays R, Harrison A, Crofoot MC, Abrahms B, Alberts SC, Ali AH, Altmann J, Antunes PC, Attias N, Belant JL, Beyer DE, Bidner LR, Blaum N, Boone RB, Caillaud D, Paula RC, la Torre JA, Dekker J, DePerno CS, Farhadinia M, Fennessy J, Fichtel C, Fischer C, Ford A, Goheen JR, Havmøller RW, Hirsch BT, Hurtado C, Isbell LA, Janssen R, Jeltsch F, Kaczensky P, Kaneko Y, Kappeler P, Katna A, Kauffman M, Koch F, Kulkarni A, LaPoint S, Leimgruber P, Macdonald DW, Markham AC, McMahon L, Mertes K, Moorman CE, Morato RG, Moßbrucker AM, Mourão G, O'Connor D, Oliveira-Santos LGR, Pastorini J, Patterson BD, Rachlow J, Ranglack DH, Reid N, Scantlebury DM, Scott DM, Selva N, Sergiel A, Songer M, Songsasen N, Stabach JA, Stacy-Dawes J, Swingen MB, Thompson JJ, Ullmann W, Vanak AT, Thaker M,

Wilson JW, Yamazaki K, Yarnell RW, Zieba F, Zwijacz-Kozica T, Fagan WF, Mueller T, Calabrese JM. 2020. Effects of body size on estimation of mammalian area requirements. *Conservation Biology:*cobi.13495. DOI: 10.1111/cobi.13495.

Ooms J. 2019. pdftools: Text Extraction, Rendering and Converting of PDF Documents.

Pebesma E. 2018. Simple Features for R: Standardized Support for Spatial Vector Data. *The R Journal.*

Pedersen TL, Crameri F. 2018. scico: Colour Palettes Based on the Scientific Colour-Maps.

Piwowar HA, Vision TJ. 2013. Data reuse and the open data citation advantage. *PeerJ* 1:e175. DOI: 10.7717/peerj.175.

R Core Team. 2020. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-project.org/.

Refsnider JM, Mitchell TS, Streby HM, Strickland JT, Warner DA, & Janzen FJ. 2011. A generalized method to determine detectability of rare and cryptic species using the ornate box turtle as a model. *Wildlife Society Bulletin* 35(2), 93-100.

Riotte-Lambert L, Matthiopoulos J. 2019. Environmental Predictability as a Cause and Consequence of Animal Movement. *Trends in Ecology & Evolution*: S0169534719302885. DOI: 10.1016/j.tree.2019.09.009.

Roche DG, Kruuk LEB, Lanfear R, Binning SA. 2015. Public Data Archiving in Ecology and Evolution: How Well Are We Doing? *PLOS Biology* 13:e1002295. DOI: 10.1371/journal.pbio.1002295.

Roll U, Feldman A, Novosolov M, Allison A, Bauer AM, Bernard R, Böhm M, Castro-Herrera F, Chirio L, Collen B, Colli GR, Dabool L, Das I, Doan TM, Grismer LL, Hoogmoed M, Itescu Y, Kraus F, LeBreton M, Lewin A, Martins M, Maza E, Meirte D, Nagy ZT, de C. Nogueira C, Pauwels OSG, Pincheira-Donoso D, Powney GD, Sindaco R, Tallowin OJS, Torres-Carvajal O, Trape J-F, Vidan E, Uetz P, Wagner P, Wang Y, Orme CDL, Grenyer R, Meiri S. 2017. The global distribution of tetrapods reveals a need for targeted reptile

conservation. *Nature Ecology & Evolution* 1:1677–1682. DOI: 10.1038/s41559-017-0332-2.

Ross N. 2020. fasterize: Fast polygon to raster conversion.

Row JR, & Blouin-Demers G. 2006. Kernels are not accurate estimators of home-range size for herpetofauna. *Copeia* 2006(4), 797-802.

RStudio Team (2020). RStudio: Integrated Development for R. RStudio, PBC, Boston, MA URL http://www.rstudio.com/.

Schipper J, Chanson JS, Chiozza F, Cox NA, Hoffmann M, Katariya V, Lamoreux J, Rodrigues ASL, Stuart SN, Temple HJ, Baillie J, Boitani L, Lacher TE, Mittermeier RA, Smith AT, Absolon D, Aguiar JM, Amori G, Bakkour N, Baldi R, Berridge RJ, Bielby J, Black PA, Blanc JJ, Brooks TM, Burton JA, Butynski TM, Catullo G, Chapman R, Cokeliss Z, Collen B, Conroy J, Cooke JG, da Fonseca GAB, Derocher AE, Dublin HT, Duckworth JW, Emmons L, Emslie RH, Festa-Bianchet M, Foster M, Foster S, Garshelis DL, Gates C, Gimenez-Dixon M, Gonzalez S, Gonzalez-Maya JF, Good TC, Hammerson G, Hammond PS, Happold D, Happold M, Hare J, Harris RB, Hawkins CE, Haywood M, Heaney LR, Hedges S, Helgen KM, Hilton-Taylor C, Hussain SA, Ishii N, Jefferson TA, Jenkins RKB, Johnston CH, Keith M, Kingdon J, Knox DH, Kovacs KM, Langhammer P, Leus K, Lewison R, Lichtenstein G, Lowry LF, Macavoy Z, Mace GM, Mallon DP, Masi M, McKnight MW, Medellin RA, Medici P, Mills G, Moehlman PD, Molur S, Mora A, Nowell K, Oates JF, Olech W, Oliver WRL, Oprea M, Patterson BD, Perrin WF, Polidoro BA, Pollock C, Powel A, Protas Y, Racey P, Ragle J, Ramani P, Rathbun G, Reeves RR, Reilly SB, Reynolds JE, Rondinini C, Rosell-Ambal RG, Rulli M, Rylands AB, Savini S, Schank CJ, Sechrest W, Self-Sullivan C, Shoemaker A, Sillero-Zubiri C, De Silva N, Smith DE, Srinivasulu C, Stephenson PJ, van Strien N, Talukdar BK, Taylor BL, Timmins R, Tirira DG, Tognelli MF, Tsytsulina K, Veiga LM, Vie J-C, Williamson EA, Wyatt SA, Xie Y, Young BE. 2008. The status of the world's land and marine mammals: diversity, threat, and knowledge. *Science* 322(5899), 225-230. DOI: 10.1126/science.1165115.

Schneider FD, Fichtmueller D, Gossner MM, Güntsch A, Jochum M, König-Ries B, Le Provost G, Manning P, Ostrowski A, Penone C, Simons NK. 2019. Towards an ecological trait-data standard. *Methods in Ecology and Evolution* 10:2006–2019. DOI: 10.1111/2041-210X.13288.

Shine R, & Madsen T. 1996. Is Thermoregulation Unimportant for Most Reptiles? An Example Using Water Pythons (*Liasis fuscus*) in Tropical Australia. *Physiological Zoology* 69:252–269.

Signer J, Fieberg J. 2020. A fresh look at an old concept: Home-range estimation in a tidy world. bioRxiv preprint. DOI: 10.1101/2020.08.19.256859.

Silva I, Crane M, Marshall BM, Strine CT. 2020. Reptiles on the wrong track? Moving beyond traditional estimators with dynamic Brownian Bridge Movement Models. *Movement Ecology 8*:1–13.

Slavenko A, Itescu Y, Ihlow F, & Meiri S. 2016. Home is where the shell is: predicting turtle home range sizes. *Journal of Animal Ecology* 85(1), 106-114.

Slowikowski K. 2018. ggrepel: Automatically Position Non-Overlapping Text Labels with "ggplot2."

Smaldino P, & O'Connor C. 2020. Interdisciplinarity Can Aid the Spread of Better Methods Between Scientific Communities. OSF Preprints. DOI: 10.31222/osf.io/cm5v3

Smith BJ, Hart KM, Mazzotti FJ, Basille M, & Romagosa CM. 2018. Evaluating GPS biologging technology for studying spatial ecology of large constricting snakes. *Animal Biotelemetry 6*(1), 1.

Stattersfield A, Bennun L, Jenkins M. 2004. State of the world's birds: indicators for our changing world. Birdlife International, Cambridge, United Kingdom.

Tingley R, Meiri S, Chapple DG. 2016. Addressing knowledge gaps in reptile conservation. *Biological Conservation* 204:1–5.

Todd BD, Nowakowski AJ. Ectothermy and the macroecology of home range scaling in snakes. *Global Ecology and Biogeography* n/a. DOI: https://doi.org/10.1111/geb.13225.

Tucker MA, Alexandrou O, Bierregaard RO, Bildstein KL, Böhning-Gaese K, Bracis C, Brzorad JN, Buechley ER, Cabot D, Calabrese JM, Carrapato C, Chiaradia A, Davenport LC, Davidson SC, Desholm M, DeSorbo CR, Domenech R, Enggist P, Fagan WF, Farwig N, Fiedler W, Fleming CH, Franke A, Fryxell JM, García-Ripollés C, Grémillet D, Griffin LR, Harel R, Kane A, Kays R, Kleyheeg E, Lacy AE, LaPoint S, Limiñana R, López-López P, Maccarone AD, Mellone U, Mojica EK, Nathan R, Newman SH, Noonan MJ, Oppel S, Prostor M, Rees EC, Ropert-Coudert Y, Rösner S, Sapir N, Schabo D, Schmidt M, Schulz H, Shariati M, Shreading A, Paulo Silva J, Skov H, Spiegel O, Takekawa JY, Teitelbaum CS, van Toor ML, Urios V, Vidal-Mateo J, Wang Q, Watts BD, Wikelski M, Wolter K, Žydelis R, Mueller T. 2019. Large birds travel farther in homogeneous environments. *Global Ecology and Biogeography* 28:576–587. DOI: 10.1111/geb.12875.

Uetz P, Freed P, Hošek J. (2020). Reptile Database. Available at http://www.reptile-database.org

Viana DS, Granados JE, Fandos P, Pérez JM, Cano-Manuel FJ, Burón D, Fandos G, Aguado MÁP, Figuerola J, Soriguer RC. 2018. Linking seasonal home range size with habitat selection and movement in a mountain ungulate. *Movement Ecology* 6:1. DOI: 10.1186/s40462-017-0119-8.

Wickham H. 2007. Reshaping Data with the reshape Package. *Journal of Statistical Software* 21:1–20.

Wickham H. 2016. ggplot2: Elegant graphics for data analysis. Springer-Verlag New York.

Wickham H. 2019a. forcats: Tools for Working with Categorical Variables (Factors).

Wickham H. 2019b. stringr: Simple, consistent wrappers for common string operations.

Wickham H, François R, Henry L, Müller K. 2020. dplyr: A grammar of data manipulation.

Wilke CO. 2018. ggridges: Ridgeline Plots in “ggplot2.”

Wilke CO. 2019. cowplot: Streamlined plot theme and plot annotations for “ggplot2.”

Worton BJ. 1989. Kernel methods for estimating the utilization distribution in home-range studies. *Ecology* 70(1), 164-168.

Zipkin EF, DiRenzo GV, Ray JM, Rossman S, & Lips KR. 2020. Tropical snake diversity collapses after widespread amphibian loss. *Science* 367(6479), 814-816.