

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32

Roles of hostplant availability and quality for the distribution and climate change response of a dietary specialist herbivore

A. Nalleli Carvajal Acosta^{1*}, and Kailen Mooney¹

¹ Department of Ecology and Evolutionary Biology, University of California-Irvine, Irvine, California, United States of America

* Corresponding Author

E-mail: ancarvaj@uci.edu (NC)

33 **Abstract**

34 Species distributions are recognized to be driven by abiotic factors, but the
35 importance of biotic interactions that provide critical resources is less well understood,
36 especially with respect to variation in critical resource quality. Disentangling the relative
37 importance of these factors – abiotic environment, presence of critical resources and their
38 quality– may be critical to predicting species response to climate change. We used species
39 distribution models (SDMs) to address these questions for the western monarch butterfly
40 (*Danaus plexippus*), a species that obligately feeds upon plants in the genus *Asclepias*, and
41 for which hostplant quality in this region varies among species by an order of magnitude. We
42 modeled the distribution of 24 *Asclepias* species to develop and compare three monarch
43 distribution models with increasing levels of ecological complexity: (i) a null model using
44 only environmental factors (a climate envelope model), (ii) a model using environmental
45 factors and *Asclepias* spp. distribution, (iii) and a model using environmental factors and
46 *Asclepias* spp. distribution weighted by hostplant quality assessed through a greenhouse
47 bioassays of larval performance. *Asclepias* models predicted that half of the *Asclepias* spp.
48 will both expand their ranges and shift their distribution towards higher latitudes while half
49 will contract within the study region. Our performance analysis of monarch models revealed
50 that the climate envelope model was the poorest performing. Adding hostplant distribution
51 produced the best performing model, while accounting for hostplant quality did not improved
52 model performance. The climate envelope model estimated more restrictive contemporary
53 and future monarch ranges compared to both hostplants models. Although all three models
54 predicted future monarch range expansions, the projected future distributions varied among
55 models. The climate envelope model predicted range expansions along the Pacific coast and

56 contractions inland while hostplants models predicted range expansions in both of these
57 regions and, as a result, estimated 14 and 19% increases in distribution relative to the climate
58 envelope model, respectively. These results suggest that information on biotic interactions
59 that provide critical resources is needed to predict future species distributions, but that
60 variation in the quality of those critical resources may be of secondary importance.

61

62 **Keywords:** *Danaus plexippus*; monarch, species distribution models; MaxEnt; climate change;
63 bioclimatic models; hostplant quality; Milkweeds; *Asclepias*; biotic interactions; specialized
64 herbivores

65 **Introduction**

66 Climate change is expected to alter the distribution of most species (Parmesan et al. 1999,
67 Crozier 2004, Bellard et al. 2012, Pauli et al. 2012) with many already experiencing range
68 contractions or facing extinctions (Sekercioglu et al. 2008, La Sorte and Jetz 2010, Bellard et al.
69 2012, Pauli et al. 2012). Understanding the underlying drivers is key to predicting such
70 distributional response and also critical if we are to mitigate these impacts. Species distributions
71 are presumed to be driven most strongly by abiotic factors, but biotic interactions can also play a
72 key role (Guisan and Thuiller 2005). Because species often respond differently to abiotic stress
73 (Schweiger et al. 2008, Van der Putten et al. 2010), producing accurate predictions necessitates
74 that we also account for climate change effects on interacting species. This is especially true for
75 species that engage in obligate interactions, as they depend on a few or even a single species to
76 survive, and such species may not be available in all areas that are otherwise climatically
77 suitable (Schweiger et al. 2008).

78 Herbivorous insects – the majority of multi-cellular species on earth (Lewinsohn et al.
79 2005) – are highly host-specific; thus, their response to climate change will likely depend
80 fundamentally on the responses of the plants upon which they are obligately dependent. Indeed,
81 most herbivorous insects feed on a single or a few plant families (Bernays 1989, Forister et al.
82 2015) with fewer than 10% feeding on plants belonging to more than three families (Price 1983).
83 Furthermore, it has long been recognized that hostplants demonstrate considerable intra- and
84 inter-specific variation in their resource quality to herbivores, and that resource quality is often
85 heterogeneously distributed across landscapes (Denno and McClure 1983, Hunter et al. 1992).
86 Intra- and inter-specific variation in host-quality can have large effects on herbivore performance
87 (Singer et al. 2012) and may also play a significant role in determining the spatial distribution of
88 host-specific herbivorous insects at local scales (Memmott et al. 1995, Mcmillin and Wagner
89 1998, Egan and Ott 2007). However, the role of hostplant quality as a driver of species
90 distribution at large spatial scales, and its implications for herbivore's distributional response to
91 climate change, are largely unknown.

92 In this study we investigated the importance of hostplant distribution and quality as
93 drivers of herbivore contemporary distribution and response to projected future climate change.
94 We use Species Distribution Models (SDMs), statistical tools that combine observations of
95 species occurrences with environmental covariates to estimate species distributions. These
96 models identify the factors driving contemporary species ranges and can also infer species
97 response to climate change based on projections for how those driving factors will change in the
98 future (Elith and Leathwick 2009). SDMs have most often assumed that species distributions are
99 defined by environmental factors alone. This so-called "climate envelope approach" are based on
100 the Eltonian noise hypothesis, which posits that biotic interactions may be a major driver of

101 abundance at smaller spatial resolutions, but at larger and coarser spatial resolutions the effects
102 of biotic interactions may average out, leaving abiotic factors as the principal drivers (Guisan
103 and Thuiller 2005, Soberon and Nakamura 2009, Elith and Leathwick 2009). Yet recent
104 modelling studies have identified biotic factors as important drivers of species distributions
105 (Dilts et al. n.d., Araújo and Luoto 2007, Preston et al. 2008, Schweiger et al. 2008, de Araújo et
106 al. 2014, Fraterrigo et al. 2014, Lemoine 2015, da Cunha et al. 2018) and SDMs predictions for
107 species response to climate change have yielded contrasting results based upon whether or not
108 biotic factors are included (Preston et al. 2008, Schweiger et al. 2008, Lemoine 2015).
109 Accordingly, climate envelope modeling may accurately define the potential niche of a species,
110 but the realized niche – defined in part by species interactions – may be substantially smaller.

111 Our aim in this study was to assess the importance of hostplant distribution and quality
112 for driving contemporary and future distributions of dietary specialist herbivores. To do so, we
113 studied the monarch butterfly (*Danaus plexippus*, Lepidoptera: Nymphalidae), the larvae of
114 which feed exclusively from plants in the *Asclepias* genus which varies greatly among species in
115 herbivore-defenses traits, nutrient content, and overall host quality (Agrawal and Fishbein 2006,
116 Pocius et al. 2017). Monarchs are well known for their migratory and overwintering behavior
117 (Pelton et al. 2019), and three previous studies have modelled their distribution. Lemoine
118 (Lemoine 2015) accounted for hostplant distribution in the eastern monarch population response
119 to climate change, predicting a poleward range expansion facilitated by *Asclepias* range
120 expansions. Steven and Frey (Stevens and Frey 2010), and more recently Dilts *et al* (Dilts et al.
121 n.d.), examined the role of hostplant availability and climate in determining the contemporary
122 western monarch distribution and their breeding grounds, again demonstrating the importance of
123 hostplants availability.

124 In the present study, we investigate the role of both hostplant distribution and quality in
125 driving contemporary and future distributions of the western monarch population. To do so, we
126 compared the performance of three species distribution models. In order of increasing
127 complexity, these three models were: (i) a model using only climatic variables as predictors
128 (hereafter, climate envelope model); (ii) a model using climatic variables and *Asclepias*
129 distribution as predictors (hereafter, hostplant-presence model); and (iii) a model that included
130 climatic variables, hostplants distribution and hostplant quality, which varied 10-fold among
131 species as assessed through bioassays of larval performance (hereafter, hostplant quality model).
132 We compared model performances and identified the variables determining the distribution of
133 the western monarch breeding ranges. These models were then used to project and estimate
134 changes in their distribution. Our study adds to past studies of this species and represents the first
135 to estimate the future breeding range of the western monarch population. More broadly, this
136 study is, to our knowledge, the first to explicitly test for the importance of hostplant quality of an
137 obligate resource in driving species contemporary and future distribution.

138 **Materials and methods**

139 *Study System*

140 Monarch butterflies occur world-wide and, in their larval stage, feed exclusively from
141 plants in the milkweed family (*Asclepias*, Apocynaceae: Asclepiadaceae). In North America,
142 there are two migratory populations that breed east and west of the Rocky Mountains, with each
143 of these regions being populated by multiple and largely unique sets of hostplant species (Ladner
144 and Altizer 2005). Despite its dramatic population decline (Pelton et al. 2019), the western
145 monarch population has been considerably understudied in comparison to the largest eastern
146 population and we know little about how this population will be affected by climate change.

147 Western monarchs breed west of the Rocky Mountains and overwinter along the Pacific
148 coast from Bodega Bay in northern California and as far south as Ensenada, Baja California,
149 Mexico (Stevens and Frey 2010). During the spring, monarchs leave their overwintering sites
150 and disperse throughout the western U.S. where they breed continuously during the summer. In
151 the fall, adult monarchs return to their overwintering grounds (Pelton et al. 2019). Within North
152 America, monarchs have been recorded feeding on 27 different plant species in the genus
153 *Asclepias* (Ladner and Altizer 2005); however, adult females may oviposit in any available
154 *Asclepias* species. Thus, monarchs may utilize multiple *Asclepias* species throughout their
155 migratory paths.

156 The genus *Asclepias*, commonly known as milkweeds, consists of over 140 different
157 species of which 130 are endemic to North America (Agrawal and Konno 2009). Milkweeds
158 vary in their herbivore defensive strategies, which variously include combinations of
159 cardenolides, latex, and trichomes, among others traits (Agrawal and Fishbein 2006). Inter-
160 specific variation in the quantity of plant defenses (Agrawal and Fishbein 2006) and nutrient
161 content (Pocius et al. 2017) have been associated with monarch larval mass, developmental rate,
162 and early instar survival (Zalucki et al. 2001). In this sense, the quality of the *Asclepias* species
163 may be important in determining monarch distributions.

164 ***Data Collection***

165 ***Occurrence data:*** We retrieved monarch and milkweed records for the United States
166 using R Studio (R Studio Team 2015) from multiple open source databases using the R
167 packages SPOCC, Ecoengine, rbison (Chamberlain et al. 2014, Karthik 2014, Chamberlain
168 2019) and by accessing species occurrences directly from GBIF and iNaturalist databases
169 (“GBIF Occurrence Download” 2019, “Naturalist [online]. Website” 2019). For monarchs,

170 we only selected eggs and larval records because they provide a direct index for the location
171 of the monarchs breeding grounds as opposed to adult records which may only indicate the
172 migratory path. Additional monarch larval records were provided by the Monarch Larvae
173 Monitor Program (MLMP) (Ries and Oberhauser 2015).

174 The occurrence data archived in open source databases originates mainly from citizen
175 scientist sightings and some from herbarium records. As opposed to formal survey methods, this
176 type of data has some limitations such as sampling biases, potential misidentification and
177 coordinate inaccuracies, and lack species absence records. We controlled for these limitations
178 whenever possible. For example, when permitted, we used filters that only retrieved records
179 confirmed by experts and/or records classified as of research quality and spatial filtering to
180 control for sampling biases.

181 To focus on the western monarch population, we selected Milkweeds and monarch larval
182 records from states corresponding to this region: California, Nevada, Colorado, Washington,
183 New Mexico, Arizona, Utah, Oregon, and Idaho. After removing duplicate records, incorrect (i.e.
184 over oceans) or inaccurate coordinates (>1000 meters uncertainty) and observations, the final
185 databases included 7,941 Milkweed records for 51 species (Data S1), and 904 monarch larval
186 records (Data S2). *A. fascicularis* and *A. speciosa* were the most common species with 22%
187 (2,541) and 12% (1,404) of total Milkweed records, respectively.

188 ***Environmental data and climate projections:*** Contemporary environmental
189 bioclimatic variables and projections for the year 2070 were downloaded in R from the
190 WorldClim website (Fick and Hijmans 2017) at 30-sec (approximately 1-km²) grid cells, the
191 finest spatial resolution available. The current bioclimatic variables represent averages of a
192 50-year period from 1950 to 2000. Climate change projections for the year 2070 represent

193 averages of a 30-year period from 2061 to 2080 based on the Hadley Centre Global
194 Environmental Model, version 2, Earth System (HadGEM2-ES) model. The HadGEM2-ES
195 model is recommended for ecological modeling as it accounts for ecologically-meaningful
196 processes such as dynamic vegetation cover (The HadGEM2 Development Team: G. M.
197 Martin et al. 2011). These projections are based on Representative Concentration Pathway
198 (RCP) 8.5. The RCP 8.5 represents the worst-case scenario for greenhouse gas (GHG)
199 concentrations, assuming that GHG emissions will continue to increase after the 21st century
200 in contrast to other scenarios that assume GHG will remain stable or will decline after the
201 21st century (Collins et al. n.d.). While a comparison of different projections for future
202 climate would provide a more nuanced prediction for the future distributions of milkweeds
203 and monarchs, using this single scenario met our primary purpose of evaluating the
204 importance of host plant information in predicting specialist herbivore distributions.

205 Environmental layers were cropped to include the states corresponding to range of the
206 western monarch population. To reduce multicollinearity among variables, we removed highly
207 correlated variables based on their Pearson correlation coefficients using a pairwise correlations
208 approach following Dormann *et al.* (Dormann et al. 2013) but with a less restrictive threshold of
209 0.85 as in Elith *et al.* (Elith et al. 2006). We first removed variables that were correlated with
210 multiple variables and, when only two variables were correlated, we selected the variable that
211 was less statistically derived. This process yielded 11 environmental predictors (Table 1).

212 ***Species distribution modeling***

213 Because species occurrences in these datasets are available in the form of presence-only
214 records, we used the maximum entropy method (hereafter MaxEnt) (Phillips et al. 2006) to
215 model the current and future distribution of *Asclepias* and monarch breeding ranges. The MaxEnt

216 algorithm is a presence-background modeling tool based on Bayesian and maximum likelihood
217 statistics (Elith et al. 2011). To estimate the probability of distribution of a species, MaxEnt uses
218 species presence records and a set of environmental predictors (e.g. precipitation, temperature)
219 across a pre-defined landscape that is divided into grid cells. From this landscape, background
220 points are randomly selected to represent the species environmental domain or background
221 environment. MaxEnt estimates the relative probability of occurrence for each grid cell by
222 maximizing the similarity between the environmental conditions of presence records and that of
223 the background environment, while constraining the prediction to have the same mean as the
224 presence records. The relative probabilities (raw output) are transformed to probability of
225 occurrence using post-logistic transformation (logistic output). Here we report the logistic output
226 which assigns a probability of presence between 0 and 1 to each grid cell, assuming that typical
227 presence localities have a probability of presence of 0.5. See Elith *et al.* (Elith et al. 2011) for a
228 comprehensive statistical explanation of MaxEnt.

229 Data collection, data processing, and modeling were performed in R studio (R Studio
230 Team 2015). Species distribution modeling was executed in MaxEnt using the ‘dismo’ package
231 (Hijmans et al. 2011).

232 ***Asclepias models:*** We developed models for individual *Asclepias* species and estimated
233 their distributions within an area restricted to the study region; therefore, our *Asclepias*
234 ranges do not represent their full distributions but only represent hostplant availability for the
235 western monarch. *Asclepias* species were modelled separately because their distributions
236 may be delimited by distinct environmental factors. We discarded records identified at the
237 genus level and species with fewer than 40 records as this limited number of observations
238 would not allow for an accurate estimation of their distributions. To correct for potential

239 sampling biases, we used a spatial filtering approach which consists on randomly selecting
240 one record per grid cell of a specified size (Kramer-Schadt et al. 2013). Spatial filtering was
241 performed individually for each *Asclepias* species. This allowed us to retain records for
242 multiple species co-occurring within a single grid cell as well as selecting the optimal spatial
243 resolution that maximizes sample size while correcting for sampling biases. For example,
244 species with a limited distribution (e.g. high-elevation species), were filtered at a finer spatial
245 resolution of 1 km² and more widely distributed *Asclepias* species were filtered at a 30-km²
246 resolution. An additional two *Asclepias* species, *A. viridiflora* and *A. curassavica*, were
247 discarded because their records were clearly subject to sample biases and spatial thinning
248 decreased their number of records to less than 40. The process of removing incorrect records
249 and rare species, and spatial filtering, resulted in 24 *Asclepias* species databases each with a
250 minimum of 40 records, totaling 3,549 *Asclepias* records (Table 2).

251 Spatially filtered data were randomly split into training and test data by withholding 25%
252 of the occurrences and the remaining 75% was used for model training. To select background
253 points, we first determined the *Asclepias* environmental domain, corresponding to an area of 50
254 km² surrounding *Asclepias* occurrences. The environmental domain was then divided into 1 km²
255 grid cells, and background points were randomly selected from within the monarch
256 environmental domain in a checkerboard fashion. Individual *Asclepias* species were modeled
257 using background points from the environmental domain represented by all *Asclepias* species.
258 This process yielded 9000 background points to model *Asclepias* species. The best-fitted models
259 for *Asclepias* with the highest AUC score were used to estimate their current and projected
260 distribution under climate change.

261 **Monarch models:** The monarch distribution was modeled using a similar approach to
262 *Asclepias*. As described above, we used spatial filtering to correct for sampling biases. Monarch
263 larval records were first filtered at a range of resolutions (1 to 55 km²) and the spatial resolution
264 yielding the highest AUC was then selected. The final dataset used to model monarch breeding
265 range was thinned using 30 km² grid cells (the best-fitted model) and included 110 observations.
266 As with *Asclepias*, we withhold 25% of the data for model testing and the remaining 75% was
267 used for model training. To determine monarch larvae environmental domain, we selected
268 4,000 background points following the same procedure described in the *Asclepias* modeling
269 section, although the number of background points was lower due to the more restricted
270 distribution of monarchs.

271 To test for the importance of hostplant availability, we first summarized the resulting
272 individual *Asclepias* distribution layers into a single predictor layer representing overall
273 *Asclepias* distribution under current and projected environmental conditions (Fig 1, A and B).
274 The values assigned to grid cell in the genus-level hostplant distribution layer were determined
275 by:

$$276 \quad P(Asc) = p(Asc1) + p(Asc2) + \dots p(Asc24)$$

277 Where ‘P’ represents the summarized genus-level probability and ‘p’ probability of
278 distribution of individual *Asclepias* species numbered from 1 to 24. Because the ranges of many
279 *Asclepias* species overlapped, forming the *Asclepias* distribution layer by summing probabilities
280 captures not only the mean probability of distribution but also reflects species richness. We
281 choose this approach under the assumption that higher species richness is associated with
282 increased milkweed abundance and thus higher habitat quality for monarchs. Although a direct
283 assessment of milkweed abundance would be preferable, no such data is readily available. This

284 process is mathematically equivalent to averaging species probabilities and then multiplying by
285 species richness.

286 To assess hostplant quality, we used the average monarch larval weight supported by
287 each *Asclepias* species grown under greenhouse conditions (Table 2). These protocols are
288 described in detail by Petschenka and Agrawal (Petschenka and Agrawal 2015). Briefly,
289 *Asclepias* plants were grown from seed in a greenhouse and after a period of 4-7 weeks neonate
290 monarch caterpillars were placed individually upon the leaves of potted plants and weighed after
291 5 days. Assessing hostplant quality under controlled greenhouse conditions controls for
292 extraneous factors such as natural predator, competition with other herbivores, induced plant
293 defenses and environmental variation that are necessarily associated with a field bioassay.

294 We weighted each *Asclepias* species distribution layer according to its host quality. The
295 Milkweed with the greatest larval weight (*A. sololana*) was given a value of 1, and all other
296 species were assigned values as proportions of this value, with the lowest quality weight being
297 0.078 (*A. asperula*) (Table 2). Five species with no information on larval weight were weighted
298 by the average host quality weight of 0.55. Weighted layers were then summarized into a single
299 predictive layer representing the hostplant probability of distribution and species richness
300 weighted by hostplant quality (Fig 1, C and D). The values assigned to grid cells of the overall
301 hostplant quality layer were calculated as follow:

$$302 \quad Pq(Asc) = [p(Asc1) * (q1)] + [p(Asc2) * (q2)] + \dots [p(Asc24) * (q24)]$$

303 Where 'Pq' represents the summarized genus-level probability of distribution weighted
304 by hostplant quality, 'p' the probability of distribution of individual *Asclepias* species numbered
305 from 1 to 24, and 'q' the host-quality weight estimated for each *Asclepias* species. This approach
306 is parallel to that used for the *Asclepias* distribution layer (above), capturing the effects of both

307 the mean probability of distribution and species richness for all co-existing *Asclepias* species, but
308 now weighting each species according to its relative hostplant quality.

309 Lastly, we used the summarized *Asclepias* layers as predictors to generate and compared
310 three models: a null model (climate envelope model) using only environmental factors as
311 predictors; a model using environmental factors and hostplant distribution represented by the
312 summarized *Asclepias* distribution (hostplant-presence model); and a second model using
313 environmental factors and *Asclepias* distribution weighted by host quality (hostplant-quality
314 model).

315 To identify the variables contributing more to each model, in addition to the “Analysis of
316 Variable Contribution” reported by MaxEnt, we performed a jackknife test of variable
317 importance. In a jackknife test, models are re-run using a single variable in isolation to identify
318 the variables that yield the highest model gain when used in isolation. This test also identifies
319 those variables that, when removed, decrease the model gain the most by re-running the models
320 excluding one variable at a time.

321 Since there is currently no consensus regarding a single most appropriate metric to
322 evaluate SDMs performance (Peterson et al. 2008, 2011, Warren and Seifert 2011), we evaluated
323 model performance based on several criteria. The area-under-the-curve (AUC) statistic provides
324 an estimate for the accuracy of predictions, with 0 indicating no predictive accuracy and 1
325 perfect predictive accuracy. An AUC score of 0.5 indicates that the model performs no better
326 than random. We also estimated performance metrics based on the Akaike Information Criterion
327 corrected for small sample sizes (AICc). The AICc metric have the advantage of balancing both,
328 model goodness-of-fit and model complexity. Furthermore, compared to AUC and BIC
329 (Bayesian-Information-Criterion) based methods, AICc evaluation methods have been shown to

330 favor models that more accurately estimate the relative importance of variables and habitat
331 suitability, both in the training region and when models are extrapolated to a different time
332 period (Warren and Seifert 2011). We calculated the AICc, delta AICc (Δ AICc), and Akaike
333 weights (wAICc) for each model using the ENMeval package (Muscarella et al. 2014). The
334 model with the lowest AICc value is considered the best model out of various candidate of
335 models. The Δ AICc is the difference between the best AICc and other candidate models. The
336 best candidate model has a Δ AICc of 0 and models with Δ AICc lower than 2 are generally
337 considered to have substantial support and should not be discarded (Muscarella et al. 2014).
338 Akaike weights (wAICc) represents the likelihood of a model given the data. The weights are
339 normalized to sum 1 and are interpreted as probabilities (Burnham and Anderson 2004).

340 Finally, we estimated suitable breeding area for monarchs and for *Asclepias* distribution
341 from polygons drawn around areas with grid cell values higher than 0.5 from the output logistic
342 layers projected from the final models.

343 **Results**

344 *Asclepias* models and estimated distribution

345 All *Asclepias* final models had AUC scores higher than 0.8, except for *A. speciosa* model
346 which yielded an AUC score of 0.74, indicating that these models are a good fit for the
347 observations (Table 2). The current estimated distributional ranges (Appendix S1, left panels.)
348 were consistent with *Asclepias* spp. distributions published by the Biota of North America
349 Program (BONAP) (Kartesz 2015).

350 Overall, within the study area, half of the *Asclepias* species are projected to expand their
351 ranges by a mean of 88% (i.e. nearly doubling their distributions) whereas the other half will
352 contract their ranges by a mean of 42% (i.e. more than halving their distributions) (Table 2 and

353 Appendix S1). Of the 24 *Asclepias* species, 19 species are predicted to shift their distributions to
354 higher latitudes (79%) both along the Pacific coast and inland, with 11 of these also expanding
355 their distributions. Of the 4 species not shifting their distributions northward, 3 will contract their
356 ranges.

357 ***Monarch models and estimated distribution***

358 The AUC scores did not differ considerably among the three models, but AUC values
359 were slightly higher for the hostplant-presence model (0.803) compared to both the hostplant-
360 quality (0.800) and climate envelope model (0.799). However, the AIC-based metrics preferred
361 the hostplant-presence model ($\Delta\text{AICc}=0$, $\text{wAICc}=1.00$) over the hostplant-quality
362 ($\Delta\text{AICc}=123.50$, $\text{wAICc}=1.515^{-27}$) and climate envelope models ($\Delta\text{AICc}=168.52$, $\text{wAICc}=2.545^{-$
363 37). The ΔAICc for the competing climate envelope and hostplant quality model was much larger
364 than 2 indicating that these two models had limited support. Likewise, the wAICc of the
365 hostplant model was nearly 1 suggesting that the likelihood of this model being the best-fitted
366 model was high (Table 3).

367 The environmental variables that contributed the most to the climate envelope model
368 were the “minimum temperature of the coldest month” (43.4% contribution; (Fick and Hijmans
369 2017) and “precipitation seasonality” (25.2% contribution, Fig. 3 A ; (Fick and Hijmans 2017)).
370 For both hostplant-presence and hostplant quality models, the hostplants variable was the second
371 most important factor for predicting the western monarch breeding range. The hostplants
372 variable contributed most to the hostplant distribution model (22.5%), after the “minimum
373 temperature of the coldest month” (33.1%) (Fig. 3 B and C). Although the hostplant quality
374 model did not produce the best-fit model, weighting the hostplant layer by host-quality increased
375 the contribution of the hostplant variable by 3% and decreased “minimum temperature of the

376 coldest month” variable contribution by 7% compared to the hostplant distribution model (Fig 3,
377 B and C). Both hostplants layers (weighted by host-quality and unweighted) exhibited the
378 highest gain (>0.40) in the jackknife test for variable importance in both hostplant models
379 (Appendix S2). This indicates that hostplants provided the most useful information for predicting
380 where monarch breeding grounds occur. For all three models the “average precipitation of the
381 warmest quarter” (Fick and Hijmans 2017) decreased model gain the most when omitted
382 suggesting that this environmental variable has the most information that is not present in other
383 variables (Appendix S2).

384 The process of weighting the *Asclepias* distribution layer by quality did not dramatically
385 altered the hostplant layer, and mainly rescaled the values of the layer (Fig 1, lower panels). This
386 was probably due to large range overlaps among *Asclepias* species as it can be observed by
387 overlaying the polygons corresponding to *Asclepias* suitable habitat (Appendix S3). Thus, a grid
388 cell occupied by multiple *Asclepias* with variable host-quality may have the same value as a grid
389 cell occupied by a few high-quality *Asclepias* species. The only area where weighting hostplants
390 by quality appeared to change the grid cell values of the hostplant quality layer was the
391 southwest region of Arizona and Utah which appeared to be occupied mostly by lower quality
392 species, predominantly by *A. asperula*, our lowest quality hostplant (Fig 1, lower panels and
393 Table 2).

394 The climate envelope model estimated more restricted ranges for the contemporary and
395 future monarch distributions. Both hostplants models estimated nearly identical contemporary
396 distributions for monarchs that were ~18% larger than the estimated by the climate envelope
397 model (Fig 2, left panels and Table 2). Although all three models predicted future range
398 expansions that nearly doubled their corresponding contemporary estimates, the hostplant-

399 presence and hostplant-quality models projected an increased in habitat suitability 14 and 19%
400 larger than that of the climate envelope model, respectively (Fig 2, right panels and Table 2).
401 This difference was primarily due to the fact that the climate envelope model predicted range
402 contractions inland whereas both hostplants models predicted range expansions in this region.
403 Finally, we detected some slight differences in the areas where hostplants models predicted that
404 such range expansions will occur. For example, the hostplant quality model predicted a smaller
405 range for monarchs in western New Mexico and a larger range in central Nevada, Utah and
406 western Colorado. (Fig 2, right panels).

407 **Discussion**

408 Predicting herbivore response to climate change requires incorporating future hostplant
409 availability, but hostplant quality may play a secondary role. While climate envelope projected a
410 more restrictive current monarch distributions than hostplant models, model comparisons
411 suggested that hostplant information provided superior predictive power. Furthermore, the three
412 models differed in their future monarch projections under climate change with models including
413 hostplant information predicting an increased in habitat suitability 14-19% larger than that of the
414 climate envelope model. Despite the importance of hostplant information, models including
415 hostplant quality did not prove superior to the model based on hostplant presence. Our study
416 suggests that information on critical biotic interactions is essential to predict future species
417 distributions under climate change.

418 The hostplant model was preferred by AIC-based metrics over a traditional climate
419 envelope model and hostplant quality model. Hostplant availability, together with the minimum
420 temperature of the coldest month, contributed over fifty percent to the hostplant model and over
421 forty percent to the model gain when used in isolation. This suggests that the western monarch

422 breeding ranges are co-limited by both cold temperatures and hostplant availability. These
423 findings are consistent with past work by Lemoine (Lemoine 2015) who found that models
424 incorporating hostplants and environmental factors most accurately estimated the eastern
425 monarch distribution. Additionally, our results are also supported by previous studies from
426 Steven and Frey (Stevens and Frey 2010) and Dilts *et al.* (Dilts et al. n.d.) who identified
427 *Asclepias* availability as well as climatic variables, including minimum temperature of the
428 coldest month, as key for structuring the western monarch breeding grounds. These findings add
429 to the increasing body of evidence suggesting that biotic interactions may govern species
430 distributions as strongly as environmental conditions (Dilts et al. n.d., Araújo and Luoto 2007,
431 Preston et al. 2008, Schweiger et al. 2008, de Araújo et al. 2014, Fraterrigo et al. 2014, Lemoine
432 2015, da Cunha et al. 2018).

433 Hostplant quality varied ten-fold among Milkweed species but did not have a large effect
434 on the overall estimates for the contemporary distribution of monarchs. We speculate this result
435 may be due to the fact that our genus-level distribution layer also reflected species richness.
436 *Asclepias* ranges exhibit substantial range overlaps in the American West (Appendix S3), thus,
437 adult monarchs may preferentially oviposit on higher quality milkweeds in areas with mixed
438 quality resources (Gripenberg et al. 2010), diminishing the influence of low-quality species.
439 Hostplants model projected similar monarch distributions under a climate change scenario;
440 however, their projections differed in some regions of the inland states of Utah, Nevada, New
441 Mexico and Colorado. This implies that the importance of hostplant quality in determining
442 herbivore distributions should not be discarded altogether as it may play a significant role in
443 instances where herbivores rely on hostplant with less geographic overlap, and therefore, fewer
444 food choices.

445 Our results demonstrate how climate envelope models that accurately represent current
446 distributions may provide poor prediction for the future. This can occur when critical
447 distributional drivers (e.g. hostplant distributions) correlate strongly with environmental factors
448 under contemporary conditions (Wharton and Kriticos 2004) but not under climate change.
449 These mechanistically-flawed models thus provide inaccurate predictions (Brewer and Gaston
450 2003, Soberon and Nakamura 2009). In our study, the climate envelope model– although more
451 restricted– estimated very similar monarch contemporary ranges than hostplant models (Fig 2,
452 left panels) but differed in their future projections (Fig 2, right panels). Specifically, the two
453 hostplant models predicted larger range expansions of monarchs than the climate envelope model
454 inland. This suggests that the climate envelope model over predicted monarch climatic
455 limitations due to contemporary correlations between climatic factors and milkweed
456 distributions, but that this correlation may not persist in the future. Accordingly, models based
457 solely on climatic factors may be adequate for estimating contemporary species distributions but
458 nevertheless produce misleading projections under novel circumstances where abiotic conditions
459 and biotic interactions do not respond in tandem to climate change.

460 The importance of incorporating the climatic response of hostplants into models is
461 underscore by the fact that only models including hostplant information predicted range
462 expansion inland while the climate-envelope model did not (Fig 2, right panels). The predicted
463 inland range expansions of the western monarch breeding range appeared to be driven by higher
464 hostplant availability in the regions of central Nevada, Utah and Colorado under future climatic
465 conditions, which was identified by our models as one of the most important factors delimiting
466 monarch distributions. Our results are congruent with previous findings by Lemoine (Lemoine

467 2015) whose study predicted northern range expansion of the eastern monarch population
468 resulting from projected *Asclepias* range expansions under future climate change scenarios.

469 Lastly, it is worth noting that our model projections do not consider factors that were
470 beyond the scope of our study but that may significantly impact monarch future distributions.
471 For example, pesticide and land-use practices, specially overwintering habitat loss to housing
472 development, is an existing threat to monarchs habitat (Pelton et al. 2019) that is likely to persist
473 in the upcoming years. Furthermore, dams and human-facilitated invasions, may alter riparian
474 areas potentially disrupting monarch migration patterns and monarch breeding grounds. Autumn
475 migrants often follow riparian corridors (Dingle et al. 2005) and riparian vegetation has been
476 associated with habitat suitability for some western Milkweed species (*A. subulata*, and *A.*
477 *asperula*) (Dilts et al. n.d.).

478 **Conclusions**

479 In summary, this study shows that accounting for biotic interactions– and their
480 distributional response to climate change– is required to predict the future distributions of
481 species obligately dependent on such interactions. A climate-envelope approach may be effective
482 for estimating contemporary species distributions but may produce misleading future projections
483 as climate change may uncouple suitable climate from essential biotic interactions. Hostplant
484 quality did not play a significant role in delimiting monarch distribution in the American West
485 where *Asclepias* ranges overlapped substantially. However, there were slight differences in some
486 regions suggesting that host-quality may still be important for predicting distributions of species
487 dependent on a fewer number of resources. These results are relevant, not only for most
488 herbivorous insects which are highly host-specific, but also for all organisms incurring in
489 obligate biotic interactions (e.g. parasitic or mutualistic interactions). Ultimately, accurate

490 projections for the future will require better incorporating inter-specific dynamics into our
491 models.

492 **Acknowledgements**

493 We thank Anurag Agrawal for providing hostplant quality data and Stijn Hantson,
494 Colleen Neil, and Will Petry for their advice on the data collection and modeling phase of this
495 project. We also thank the Monarch Larvae Monitoring Project (MLMP) for kindly providing
496 monarch larval records.

497

498

499

500

501

502

503

504

505

506

507

508 **Literature Cited**

- 509 Agrawal, A. A., and M. Fishbein. 2006. Plant defense syndromes. *Ecology* 87:S132–S149.
- 510 Agrawal, A. A., and K. Konno. 2009. Latex: A model for understanding mechanisms, ecology,
511 and evolution of plant defense against herbivory. *Annual Review of Ecology, Evolution,*
512 *and Systematics* 40:311–331.
- 513 de Araújo, C. B., L. O. Marcondes-Machado, and G. C. Costa. 2014. The importance of biotic
514 interactions in species distribution models: a test of the Eltonian noise hypothesis using
515 parrots. *Journal of Biogeography* 41:513–523.
- 516 Araújo, M. B., and M. Luoto. 2007. The importance of biotic interactions for modelling species
517 distributions under climate change. *Global Ecology and Biogeography* 16:743–753.
- 518 Bellard, C., C. Bertelsmeier, P. Leadley, W. Thuiller, and F. Courchamp. 2012. Impacts of
519 climate change on the future of biodiversity: Biodiversity and climate change. *Ecology*
520 *Letters* 15:365–377.
- 521 Bernays, E. A. 1989. Host range in phytophagous insects: the potential role of generalist
522 predators. *Evolutionary Ecology* 3:299–311.
- 523 Brewer, A. M., and K. J. Gaston. 2003. The geographical range structure of the holly leaf-miner.
524 II. Demographic rates. *Journal of Animal Ecology* 72:82–93.
- 525 Burnham, K. P., and D. R. Anderson. 2004. Multimodel inference: understanding AIC and BIC
526 in model selection 33:261–304.
- 527 Chamberlain, S. 2019. rbison:Interface to the “USGS” “BISON” API.
- 528 Chamberlain, S., K. Ram, and T. Hart. 2014. spocc: R interface to many species occurrence data
529 sources.

530 Collins, M., R. Knutti, J. Arblaster, J.-L. Dufresne, T. Fichefet, X. Gao, W. J. G. Jr, T. Johns, G.
531 Krinner, M. Shongwe, A. J. Weaver, M. Wehner, M. R. Allen, T. Andrews, U. Beyerle,
532 C. M. Bitz, S. Bony, B. B. Booth, H. E. Brooks, V. Brovkin, O. Browne, C. Brutel-
533 Vuilmet, M. Cane, R. Chadwick, E. Cook, K. H. Cook, M. Eby, J. Fasullo, C. E. Forest,
534 P. Forster, P. Good, H. Goosse, J. M. Gregory, G. C. Hegerl, P. J. Hezel, K. I. Hodges,
535 M. M. Holland, M. Huber, M. Joshi, V. Kharin, Y. Kushnir, D. M. Lawrence, R. W. Lee,
536 S. Liddicoat, C. Lucas, W. Lucht, J. Marotzke, F. Massonnet, H. D. Matthews, M.
537 Meinshausen, C. Morice, A. Otto, C. M. Patricola, G. Philippon, S. Rahmstorf, W. J.
538 Riley, O. Saenko, R. Seager, J. Sedláček, L. C. Shaffrey, D. Shindell, J. Sillmann, B.
539 Stevens, P. A. Stott, R. Webb, G. Zappa, K. Zickfeld, S. Joussaume, A. Mokssit, K.
540 Taylor, and S. Tett. (n.d.). Long-term climate change: projections, commitments and
541 irreversibility:108.

542 Crozier, L. 2004. Warmer winters drive butterfly range expansion by increasing survivorship.
543 Ecology 85:231–241.

544 da Cunha, H. F., É. D. Ferreira, G. Tessarolo, and J. C. Nabout. 2018. Host plant distributions
545 and climate interact to affect the predicted geographic distribution of a Neotropical
546 termite. *Biotropica* 50:625–632.

547 Denno, R. F., and M. S. McClurc. 1983. Variable plants and herbivores in natural and managed
548 systems. Academic Press, New York.

549 Dilts, T. E., M. O. Steele, J. E. Engler, E. M. Pelton, S. J. Jepsen, S. J. McKnight, A. R. Taylor,
550 C. A. Fallon, S. H. Black, E. E. Cruz, D. R. Cravier, and M. L. Forister. (n.d.). Host
551 plants and climate structure habitat associations of the western monarch butterfly.

- 552 Dingle, H., M. P. Zalucki, W. A. Rochester, and T. Armijo-Prewitt. 2005. Distribution of the
553 monarch butterfly, *Danaus plexippus* (L.) (Lepidoptera: Nymphalidae), in western North
554 America: Monarch Butterflies in Western North America. *Biological Journal of the*
555 *Linnean Society* 85:491–500.
- 556 Dormann, C. F., J. Elith, S. Bacher, C. Buchmann, G. Carl, G. Carré, J. R. G. Marquéz, B.
557 Gruber, B. Lafourcade, P. J. Leitão, T. Münkemüller, C. McClean, P. E. Osborne, B.
558 Reineking, B. Schröder, A. K. Skidmore, D. Zurell, and S. Lautenbach. 2013.
559 Collinearity: a review of methods to deal with it and a simulation study evaluating their
560 performance. *Ecography* 36:27–46.
- 561 Egan, S. P., and J. R. Ott. 2007. Host plant quality and local adaptation determine the distribution
562 of gall-forming herbivore. *Ecology* 88:2868–2879.
- 563 Elith, J., C. H. Graham, R. P. Anderson, M. Dudík, S. Ferrier, A. Guisan, R. J. Hijmans, F.
564 Huettmann, J. R. Leathwick, A. Lehmann, J. Li, L. G. Lohmann, B. A. Loiselle, G.
565 Manion, C. Moritz, M. Nakamura, Y. Nakazawa, J. McC. M. Overton, A. Townsend
566 Peterson, S. J. Phillips, K. Richardson, R. Scachetti-Pereira, R. E. Schapire, J. Soberón,
567 S. Williams, M. S. Wisz, and N. E. Zimmermann. 2006. Novel methods improve
568 prediction of species' distributions from occurrence data. *Ecography* 29:129–151.
- 569 Elith, J., and J. R. Leathwick. 2009. Species Distribution Models: Ecological Explanation and
570 Prediction Across Space and Time. *Annual Review of Ecology, Evolution, and*
571 *Systematics* 40:677–697.
- 572 Elith, J., S. J. Phillips, T. Hastie, M. Dudík, Y. E. Chee, and C. J. Yates. 2011. A statistical
573 explanation of MaxEnt for ecologists: Statistical explanation of MaxEnt. *Diversity and*
574 *Distributions* 17:43–57.

- 575 Fick, E. C., and R. J. Hijmans. 2017. WorldClim-Global Climate Data.
576 <http://www.worldclim.org>.
- 577 Forister, M. L., V. Novotny, A. K. Panorska, L. Baje, Y. Basset, P. T. Butterill, L. Cizek, P. D.
578 Coley, F. Dem, I. R. Diniz, P. Drozd, M. Fox, A. E. Glassmire, R. Hazen, J. Hrcek, J. P.
579 Jahner, O. Kaman, T. J. Kozubowski, T. A. Kursar, O. T. Lewis, J. Lill, R. J. Marquis, S.
580 E. Miller, H. C. Morais, M. Murakami, H. Nickel, N. A. Pardikes, R. E. Ricklefs, M. S.
581 Singer, A. M. Smilanich, J. O. Stireman, S. Villamarín-Cortez, S. Vodka, M. Volf, D. L.
582 Wagner, T. Walla, G. D. Weiblen, and L. A. Dyer. 2015. The global distribution of diet
583 breadth in insect herbivores. *Proceedings of the National Academy of Sciences* 112:442–
584 447.
- 585 Fraterrigo, J. M., S. Wagner, and R. J. Warren. 2014. Local-scale biotic interactions embedded in
586 macroscale climate drivers suggest Eltonian noise hypothesis distribution patterns for an
587 invasive grass. *Ecology Letters* 17:1447–1454.
- 588 GBIF.org. 2019. <https://doi.org/10.15468/dl.sjg2d2>.
- 589 Gripenberg, S., P. J. Mayhew, M. Parnell, and T. Roslin. 2010. A meta-analysis of preference-
590 performance relationships in phytophagous insects. *Ecology Letters* 13:383–393.
- 591 Guisan, A., and W. Thuiller. 2005. Predicting species distribution: offering more than simple
592 habitat models. *Ecology Letters* 8:993–1009.
- 593 Hijmans, R. J., S. Phillips, J. R. Leathwick, and J. Elith. 2011. Package ‘dismo.’
- 594 Hunter, M. D., T. Ohgushi, and P. W. Price. 1992. Effects of resource distribution on animal-
595 plant interactions. Academic Press, San Diego, CA.
- 596 Kartesz, J. T. 2015. Floristic Synthesis of North America. Taxonomic Data Center, Chapel Hill,
597 N.C. [maps generated from Kartesz, J.T. 2015.

- 598 Karthik, R. 2014. ecoengine: Programmatic interface to the API serving UC Berkeley's Natural
599 History Data.
- 600 Kramer-Schadt, S., J. Niedballa, J. D. Pilgrim, B. Schröder, J. Lindenborn, V. Reinfelder, M.
601 Stillfried, I. Heckmann, A. K. Scharf, D. M. Augeri, S. M. Cheyne, A. J. Hearn, J. Ross,
602 D. W. Macdonald, J. Mathai, J. Eaton, A. J. Marshall, G. Semiadi, R. Rustam, H.
603 Bernard, R. Alfred, H. Samejima, J. W. Duckworth, C. Breitenmoser-Wuersten, J. L.
604 Belant, H. Hofer, and A. Wilting. 2013. The importance of correcting for sampling bias
605 in MaxEnt species distribution models. *Diversity and Distributions* 19:1366–1379.
- 606 La Sorte, F. A., and W. Jetz. 2010. Projected range contractions of montane biodiversity under
607 global warming. *Proceedings of the Royal Society B: Biological Sciences* 277:3401–
608 3410.
- 609 Ladner, D. T., and S. Altizer. 2005. Oviposition preference and larval performance of North
610 American monarch butterflies on four *Asclepias* species. *Entomologia Experimentalis et*
611 *Applicata* 116:9–20.
- 612 Lemoine, N. P. 2015. Climate change may alter breeding ground distributions of eastern
613 migratory Monarchs (*Danaus plexippus*) via range expansion of *Asclepias* host plants.
614 *PLOS ONE* 10:e0118614.
- 615 Lewinsohn, T., V. Novotny, and Y. Basset. 2005. Insects on plants: diversity of herbivore
616 assemblages revisited 36:597–620.
- 617 Mcmillin, Joel. D., and M. R. Wagner. 1998. Influence of host plant vs. natural enemies on the
618 spatial distribution of a pine sawfly, *Neodiprion autumnalis*. *Ecological Entomology*
619 23:397–408.

- 620 Memmott, J., R. K. Day, and H. C. J. Godfray. 1995. Intraspecific variation in host plant quality:
621 the aphid *Cinara cupressi* on the Mexican cypress, *Cupressus lusitanica*. *Ecological*
622 *Entomology* 20:153–158.
- 623 Muscarella, R., P. J. Galante, M. Soley-Guardia, R. A. Boria, J. M. Kass, M. Uriarte, and R. P.
624 Anderson. 2014. ENMeval: An R package for conducting spatially independent
625 evaluations and estimating optimal model complexity for MAXENT ecological niche
626 models. *Methods in Ecology and Evolution* 5:1198–1205.
- 627 Naturalist [online]. Website. 2019, January. . <https://www.inaturalist.org>.
- 628 Parmesan, C., N. Ryrholm, C. Stefanescu, J. K. Hill, C. D. Thomas, H. Descimon, B. Huntley, L.
629 Kaila, J. Kullberg, T. Tammaru, W. J. Tennent, J. A. Thomas, and M. Warren. 1999.
630 Poleward shifts in geographical ranges of butterfly species associated with regional
631 warming. *Nature* 399:579–583.
- 632 Pauli, H., M. Gottfried, S. Dullinger, O. Abdaladze, M. Akhalkatsi, J. L. B. Alonso, G. Coldea, J.
633 Dick, B. Erschbamer, R. F. Calzado, D. Ghosn, J. I. Holten, R. Kanka, G. Kazakis, J.
634 Kollar, P. Larsson, P. Moiseev, D. Moiseev, U. Molau, J. M. Mesa, L. Nagy, G. Pelino,
635 M. Puscas, G. Rossi, A. Stanisci, A. O. Syverhuset, J.-P. Theurillat, M. Tomaselli, P.
636 Unterluggauer, L. Villar, P. Vittoz, and G. Grabherr. 2012. Recent plant diversity
637 changes on Europe’s mountain summits. *Science* 336:353–355.
- 638 Pelton, E. M., C. B. Schultz, S. J. Jepsen, S. H. Black, and E. E. Crone. 2019. Western Monarch
639 population plummets: status, probable causes, and recommended conservation actions.
640 *Frontiers in Ecology and Evolution* 7:258.

- 641 Peterson, A., J. Soberon, R. G. Pearson, R. P. Anderson, E. Martinez-Meyer, M. Nakamura, and
642 M. Araújo. 2011. Ecological niches and geographic distributions. Princeton University,
643 Press.
- 644 Peterson, A. T., M. Papeş, and J. Soberón. 2008. Rethinking receiver operating characteristic
645 analysis applications in ecological niche modeling. *Ecological Modelling* 213:63–72.
- 646 Petschenka, G., and A. A. Agrawal. 2015. Milkweed butterfly resistance to plant toxins is linked
647 to sequestration, not coping with a toxic diet. *Proceedings of the Royal Society B:
648 Biological Sciences* 282:20151865.
- 649 Phillips, S. J., R. P. Anderson, and R. E. Schapire. 2006. Maximum entropy modeling of species
650 geographic distributions. *Ecological Modelling* 190:231–259.
- 651 Pocius, V. M., D. M. Debinski, J. M. Pleasants, K. G. Bidne, R. L. Hellmich, and L. P. Brower.
652 2017. Milkweed matters: Monarch butterfly (Lepidoptera: Nymphalidae) survival and
653 development on nine midwestern Milkweed species. *Environmental Entomology*
654 46:1098–1105.
- 655 Preston, K. L., J. T. Rotenberry, R. A. Redak, and M. F. Allen. 2008. Habitat shifts of
656 endangered species under altered climate conditions: importance of biotic interactions.
657 *Global Change Biology*.
- 658 Price, P. W. 1983. Hypotheses on organization and evolution in herbivorous insect communities.
659 Pages 559–98 *Variable Plants and Herbivores in Natural and Managed Systems*.
660 Academic Press, New York.
- 661 R Studio Team. 2015. R Studio. Integrated Development for R. RStudio, Inc., Boston, MA.
- 662 Ries, L., and K. S. Oberhauser. 2015. A citizen army for science: quantifying the contributions of
663 citizen scientists to our understanding of monarch butterfly biology *65:419–430*.

- 664 Schweiger, O., J. Settele, O. Kudrna, S. Klotz, and I. Kühn. 2008. Climate change can cause
665 spatial mismatch of trophically interacting species. *Ecology* 89:3472–3479.
- 666 Sekercioglu, C. H., S. H. Schneider, J. P. Fay, and S. R. Loarie. 2008. Climate change,
667 elevational range shifts, and bird extinctions: *Elevation, Climate Change, and Bird*
668 *Extinctions*. *Conservation Biology* 22:140–150.
- 669 Singer, M. S., T. E. Farkas, C. M. Skorik, and K. A. Mooney. 2012. Tritrophic interactions at a
670 community level: effects of host plant species quality on bird predation of caterpillars
671 179:363–374.
- 672 Soberon, J., and M. Nakamura. 2009. Niches and distributional areas: Concepts, methods, and
673 assumptions. *Proceedings of the National Academy of Sciences* 106:19644–19650.
- 674 Stevens, S. R., and D. F. Frey. 2010. Host plant pattern and variation in climate predict the
675 location of natal grounds for migratory monarch butterflies in western North America.
676 *Journal of Insect Conservation* 14:731–744.
- 677 The HadGEM2 Development Team: G. M. Martin, N. Bellouin, W. J. Collins, I. D. Culverwell,
678 P. R. Halloran, S. C. Hardiman, T. J. Hinton, C. D. Jones, R. E. McDonald, A. J.
679 McLaren, F. M. O’Connor, M. J. Roberts, J. M. Rodriguez, S. Woodward, M.
680 J. Best, M. E. Brooks, A. R. Brown, N. Butchart, C. Dearden, S. H. Derbyshire, I.
681 Dharssi, M. Doutriaux-Boucher, J. M. Edwards, P. D. Falloon, N. Gedney, L. J. Gray, H.
682 T. Hewitt, M. Hobson, M. R. Huddleston, J. Hughes, S. Ineson, W. J. Ingram, P. M.
683 James, T. C. Johns, C. E. Johnson, A. Jones, C. P. Jones, M. M. Joshi, A. B. Keen, S.
684 Liddicoat, A. P. Lock, A. V. Maidens, J. C. Manners, S. F. Milton, J. G. L. Rae, J. K.
685 Ridley, A. Sellar, C. A. Senior, I. J. Totterdell, A. Verhoef, P. L. Vidale, and A.

686 Wiltshire. 2011. The HadGEM2 family of Met Office Unified Model climate
687 configurations. *Geoscientific Model Development* 4:723–757.

688 Van der Putten, W. H., M. Macel, and M. E. Visser. 2010. Predicting species distribution and
689 abundance responses to climate change: why it is essential to include biotic interactions
690 across trophic levels. *Philosophical Transactions of the Royal Society B: Biological
691 Sciences* 365:2025–2034.

692 Warren, D. L., and S. N. Seifert. 2011. Ecological niche modeling in Maxent: the importance of
693 model complexity and the performance of model selection criteria. *Ecological
694 Applications* 21:335–342.

695 Wharton, T. N., and D. J. Kriticos. 2004. Biodiversity: The fundamental and realized niche of the
696 Monterey Pine aphid, *Essigella*. *Diversity and Distributions*:10.

697 Zalucki, M., L. Brower, and A. Alonso. 2001. Detrimental effects of latex and cardiac glycosides
698 on survival and growth of first-instar monarch butterfly larvae *Danaus plexippus* feeding
699 on the sandhill milkweed *Asclepias humistrata* 26:212–224.

700

701

702

703

704

705

706

707

708

709

710

711

712

713

714 **Tables**

715

716

Table 1. Selected environmental variables

Worldclim Code	Environmental Variable*
Bio1	Annual Mean Temperature
Bio5	Max Temperature of Warmest Month
Bio6	Minimum Temperature of Coldest Month
Bio7	Temperature Annual Range
Bio8	Mean Temperature of Wettest Quarter
Bio9	Mean Temperature of Driest Quarter
Bio12	Annual Precipitation
Bio15	Precipitation Seasonality
Bio17	Precipitation of Driest Quarter
Bio18	Precipitation of Warmest Quarter
Bio19	Precipitation of Coldest Quarter

*Selected environmental variables with Pearson correlation coefficient of 0.85 or lower.

717

718

719

720

721

722

723

724

725

726

727

728

729

730

731

732

733 **Table 2. Milkweeds models summary and estimated habitat suitability**
 734

<i>Asclepias</i> species	Spatial filtering (Km ²)	Records		AUC best model	Estimated Habitat (Km ²)		Host quality weight ¹
		Total	Filtered		Current	Projected	
<i>A. albicans</i>	1	434	209	0.977	37,937	28,444	0.554
<i>A. asperula</i> *	30	586	244	0.815	614,502	678,420	0.078
<i>A. californica</i> *	1	607	277	0.945	84,794	124,187	0.306
<i>A. cordifolia</i> *	30	683	173	0.848	230,690	215,368	0.621
<i>A. cryptoceras</i>	30	210	128	0.839	416,872	246,545	0.218
<i>A. engelmanniana</i> *	30	90	70	0.915	277,368	230,811	0.502
<i>A. eriocarpa</i>	30	878	145	0.888	147,561	235,229	0.541
<i>A. erosa</i> *	30	524	145	0.885	285,506	466,099	0.671
<i>A. fascicularis</i> *	30	2259	399	0.859	306,140	512,078	0.545
<i>A. halli</i>	1	48	48	0.915	197,755	2,896	0.898
<i>A. incarnata</i> *	1	144	94	0.805	113,426	64,869	0.823
<i>A. labriformis</i>	1	68	57	0.945	22,126	4,863	0.554
<i>A. latifolia</i> *	30	162	102	0.858	415,185	412,476	0.394
<i>A. linaria</i> *	1	127	75	0.962	376,557	382,289	0.427
<i>A. macrosperma</i>	1	71	62	0.965	624,888	124,259	0.554
<i>A. macrotis</i> *	1	46	44	0.925	196,251	151,159	0.554
<i>A. nycatginifolia</i> *	1	154	100	0.958	112,228	233,621	0.554
<i>A. pumila</i> *	1	89	73	0.985	69,502	5,581	0.660
<i>A. soloanoana</i> *	1	132	68	0.995	17,704	65,588	1.00
<i>A. speciosa</i>	30	1359	478	0.746	958,479	937,320	0.768
<i>A. subulata</i> *	30	627	114	0.927	114,507	379,000	0.606
<i>A. subverticillata</i> *	30	545	286	0.809	594,885	674,988	0.505
<i>A. tuberosa</i> *	30	184	88	0.831	305,144	470,641	0.681
<i>A. vestita</i> *	1	159	70	0.983	34,449	80,731	0.286

735 *Species marked with an asterisk are projected to shift their distribution to higher latitudes (S1
 736 File.)

737 ¹Hostplant quality weight determined by the average weight of monarch larvae reared on 24
 738 *Asclepias* species within a 5-day period. Bold numbers indicate the average weight assigned to
 739 species with missing hostplant quality information.

740
 741
 742
 743
 744
 745
 746
 747
 748

749 **Table. 3 Monarch model performance comparison and estimated habitat suitability**

Model	n^1	AUC	AICc	Δ AICc	wAICc	Estimated habitat (km ²) *	
						Current	Projected (2070)
Climate envelope	60	0.799	25,694.99	168.52	2.545 ⁻³⁷	214,245	409,091
Hostplant-presence	61	0.803	25,526.46	0.000	1.00	252,464	466,306
Hostplant-quality	61	0.800	25,649.97	123.50	1.515 ⁻²⁷	252,465	486,200

750 *Estimated habitat was calculated by summarizing areas with probability of distribution higher
751 than 0.5 from the logistic output layers produced by each model. ¹ n gives the number of
752 parameters of each model.

753

754

755

756

757

758

759

760

761

762

763

764

765

766

767

768

769

770

771

772

773

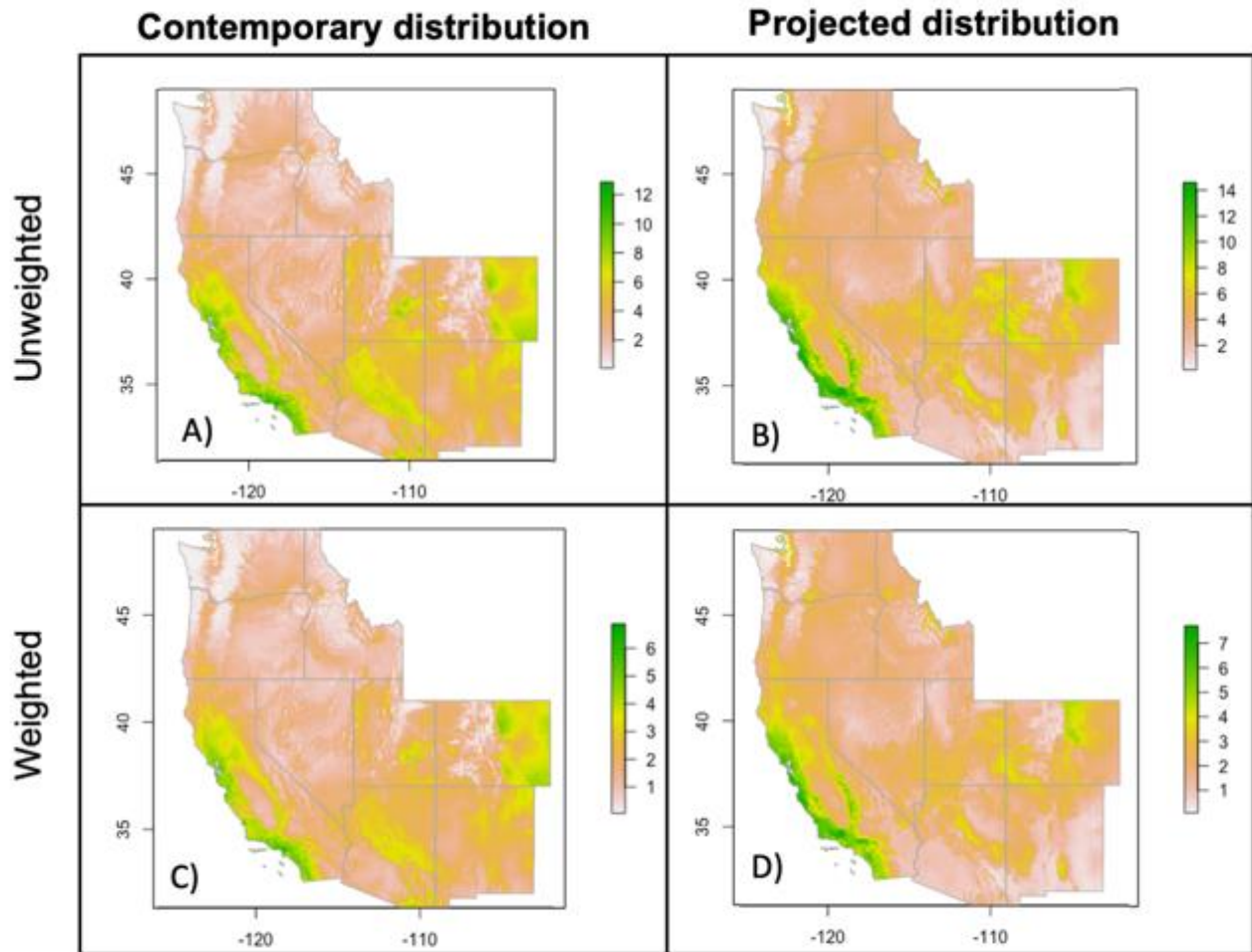
774

775

776 **Figures**

777

778 **Figure 1. Contemporary and projected genus-level milkweeds distribution**



779

780 **Fig 1.** Genus-level *Asclepias* predictor and projection layers used in the monarch hostplant
781 distribution and quality models. The unweighted layers reflect the summed probability of
782 occurrence of all *Asclepias* species, thus reflecting mean probability of occurrence and species
783 richness, while the weighted layer additionally weighs each species by hostplant quality. ‘Green’
784 indicate high probability of distribution, species richness and/or host-quality, and ‘white’ low
785 probability of distribution, species richness and/or low quality.

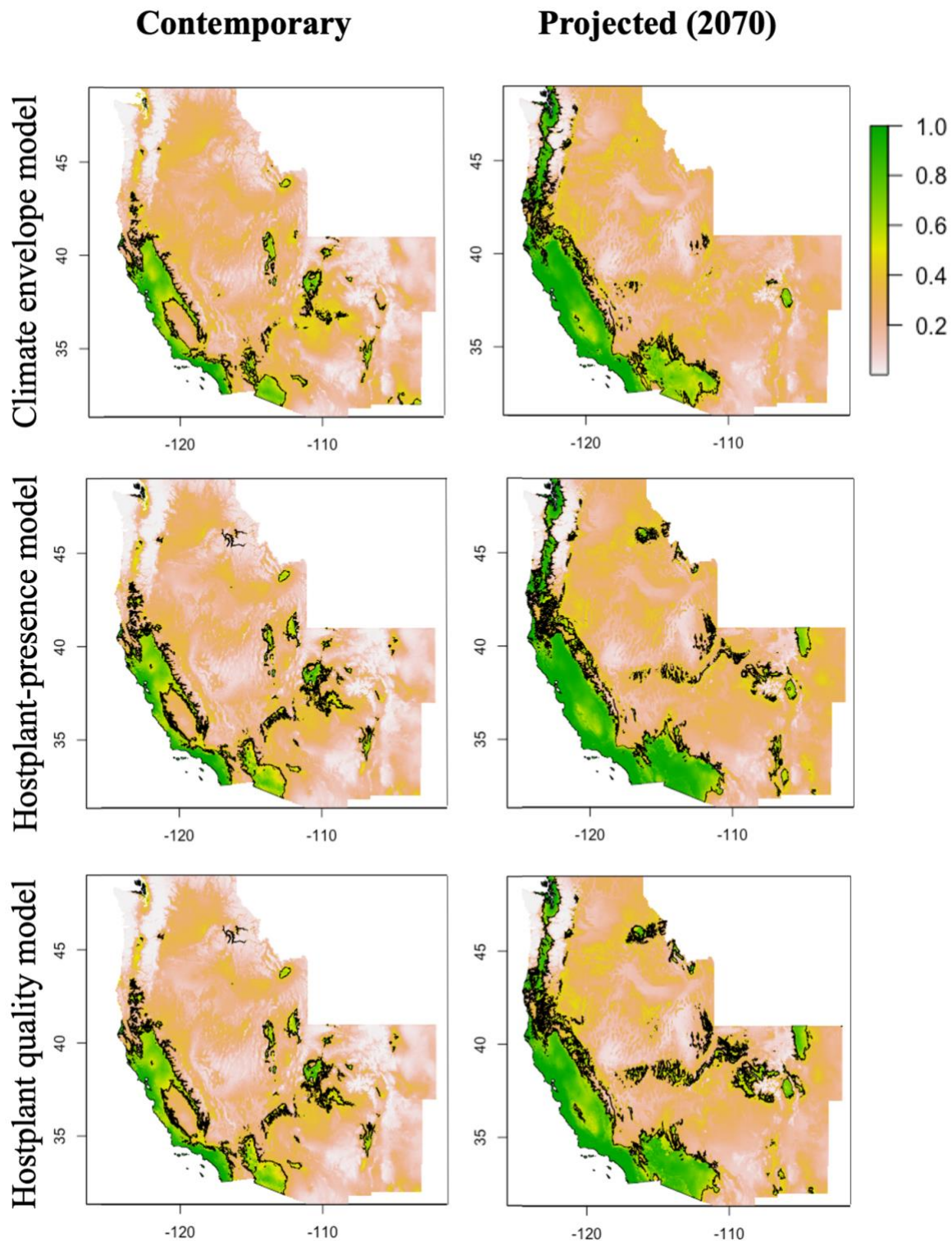
786

787

788

789

790 **Figure 2. Monarch contemporary and projected future breeding ranges**



791
792 **Fig 2.** Left panels (A, C, E) indicate the current probability of distribution of the monarch
793 breeding grounds estimated by the three models, with 'light yellow' representing low probability
794 and 'dark blue' high probability. Right panels (B, D, F) indicate the projected probability of

795 distribution of the monarch breeding grounds for the year 2070 estimated by the three models.
796 Suitable habitat for monarch breeding is delineated in black and represent areas with a
797 probability of distribution greater than 0.5.

798

799

800

801

802

803

804

805

806

807

808

809

810

811

812

813

814

815

816

817

818

819

820

821

822

823

824

825

826 **Figure 3. Analysis of variable importance**

827

828

829

830

831

832

833

834

835

836

837

838

839

840

841

842

843

844

845

846

847

848

849

850

851

852

853

854

855

856

857

858

859

860

861

862

863

864

865

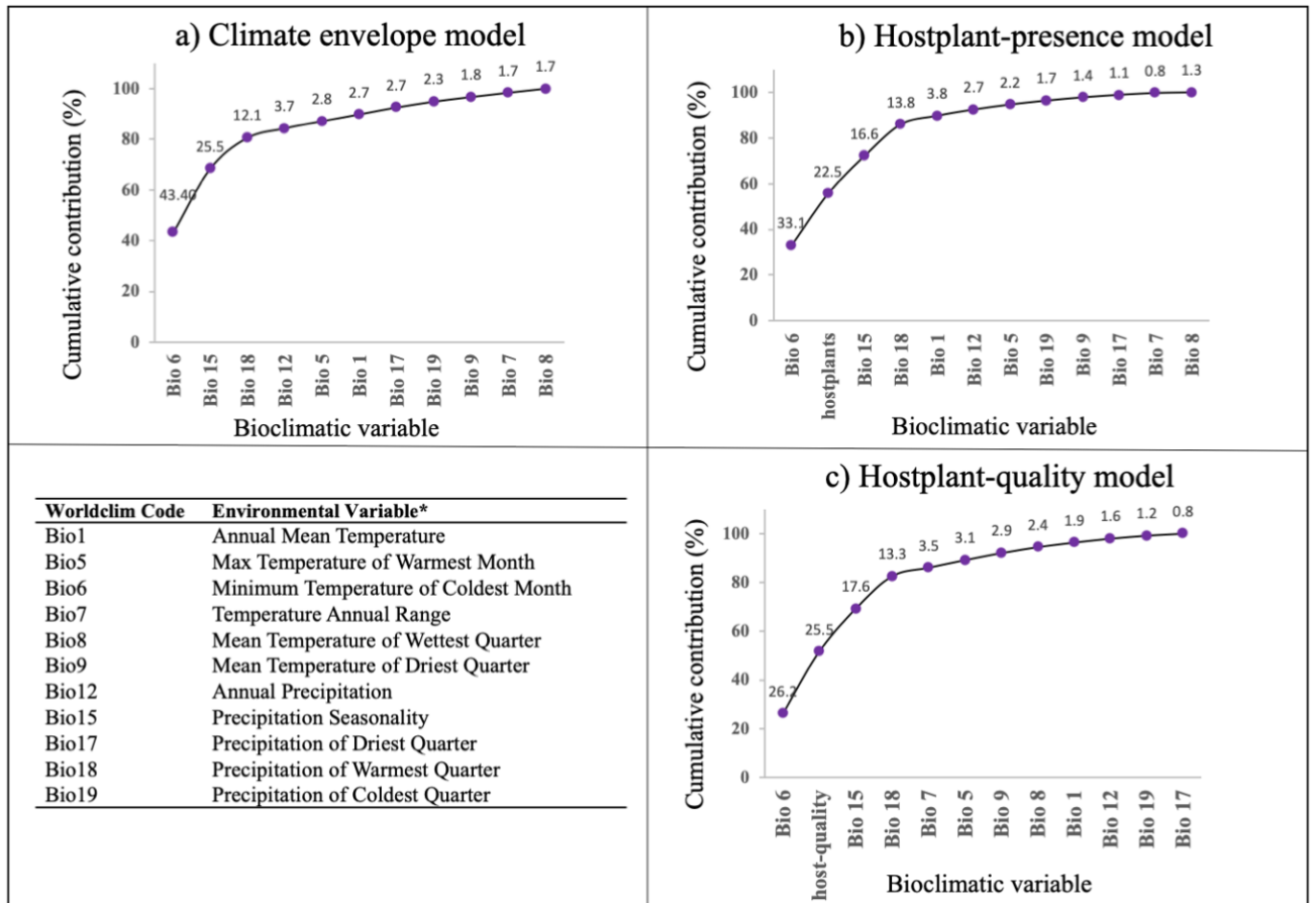


Fig. 3. Percent contribution that each variable contributes to the models in decreasing order from left to right.

866 **Appendices**

867

868 **Appendix S1. Milkweeds species contemporary and projected distributions.** Estimated
869 contemporary distribution of 24 species of *Asclepias* (left panels) and projections under a climate
870 change scenario for 2070 (right panels), with ‘green’ indicating high probability of distribution
871 and ‘white’ low probability. Suitable habitat for each *Asclepias* spp. is delineated in black and
872 represent areas with a probability of distribution greater than 0.5.

873 **Appendix S2. Jackknife Test of Variable Importance.** Jackknife test of variable importance.
874 Blue bars indicate model gain when each variable is used in isolation, turquoise bars represent
875 model gain when a single variable is excluded, and red bars represent model gain when all
876 variables are included.

877 **Appendix S3. Milkweeds range overlaps in the Western United States.** Each overlaid layer
878 represents the range of individual *Asclepias* species estimated by drawing a polygon around
879 areas with areas with a probability of distribution greater than 0.5. Regions in white represent
880 areas with no Milkweeds, ‘light green’ represent low range overlap, and ‘dark green’ high range
881 overlap.

882 **Metadata S1.** R code for monarch and milkweed species distributions modeling.

883 **Data S1. Milkweed species records** retrieved from various open source databases within the
884 study region.

885 **Data S2. Monarch larval records** retrieved from open source databases and the MLMP
886 (Monarch Larvae Monitoring Project) within the study region.

887

888

889

890

891

892

893

894

895

896

897

898

899

900

901

902

903

904

905

906

907

908

909

910