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Redox protein Memo1 coordinates FGF23-driven signaling and small Rho-GTPases in the mouse kidney

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Running title: FGF23 responses require Memo1-dependent redox signaling

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Abstract

Memo promotes receptor tyrosine kinase (RTK) signaling by unknown mechanisms. *Memo1* deletion in mice causes premature aging and unbalanced metabolism partially resembling *Fgf23* and *Klotho* loss-of-function animals. Here, we report a role for Memo's redox function in FGF23-driven RTK signaling in the kidney. Postnatally Memo-deficient (cKO) and floxed controls were treated with FGF23 or vehicle, followed by molecular and biochemical analyses. Findings were validated using cell culture and recombinant proteins. Memo cKO mice showed impaired renal ERK phosphorylation and transcriptional responses to FGF23. Redox proteomics revealed excessive thiols of Rho-GDP dissociation inhibitor 1 (Rho-GDI1). Renal RhoA abundance and activity were increased in Memo cKO. Immunoprecipitation analysis showed association between Memo and Rho-GDI1. We confirmed an interaction between the two proteins, with Memo-dependent irreversible dioxidation and trioxidation at Rho-GDI1 Cys79. Collectively, our findings reveal that redox protein Memo promotes renal FGF23 signaling together with oxidative modulation of the Rho-GTPase network.

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Introduction

Mediator of Cell Motility 1 (Memo) is an evolutionarily conserved protein associated with the cytoskeleton and receptor tyrosine kinases (Zaoui et al. 2008; Marone et al. 2004; Schotanus and Van Otterloo 2020; Jiang et al. 2013). In cultured cells, Memo promotes signaling and migration induced by fibroblast growth factors (FGFs) and assists in recruiting intracellular adaptor proteins to the FGF receptor (Marone et al. 2004; Haenzi et al. 2014). In mice, postnatally-inducible ubiquitous deletion of *Memo1* exon 2 (Memo cKO) resulted in a phenotype of premature aging and insulin hypersensitivity (Haenzi et al. 2014) which strikingly resembles *Fgf23* and *Klotho*-deficient mouse models (Kuro-o et al. 1997; Shimada et al. 2004). Four to eight weeks after *Memo1* recombination, Memo cKO animals developed kidney failure, elevated serum concentrations of FGF23, calcium and magnesium, and a bone and mineral disease (Haenzi et al. 2014; Moor et al. 2018; 2020). We hypothesized that Memo modulates renal FGF23-induced cellular signaling in the kidney and in the experiments presented here, we addressed the underlying mechanisms. We demonstrate that Memo regulates activity and abundance of small Rho-GTPases and proteomic oxidation that is required for physiological intracellular signaling responses to FGF23.

Results and discussion

We generated inducible Memo cKO and floxed control mice and maintained them on C57BL/6 background as previously described (Moor et al. 2018). We treated mice of both genotypes with tamoxifen to excise *Memo1* exon 2 in the whole body. We randomized mice of both genotypes to intraperitoneal FGF23 or vehicle injections as depicted in (Figure 1A) and studied all mice prior to onset of the previously described premature aging and lethal phenotype (Haenzi et al. 2014). Body weight (Figure 1B), serum creatinine (Figure 1 figure supplement 1A), calcium and phosphate (Figure 1 figure supplement 1B-C), were comparable between genotypes, and as expected Memo protein was absent in the kidney (Figure 1C). We assessed phosphorylated and total ERK protein in whole kidney and detected an increase in phospho-ERK in control mice treated with FGF23, but not in FGF23-treated Memo cKO animals (Figure 1D-E). Next, we analyzed expression of ERK downstream genes in kidney of mice in both genotype and treatment groups. After 1 hour of FGF23 treatment there were increases in *Egr1*, *cFos*, and *Cyp24a1* in control mice, but not in FGF23-treated cKO (Figure 1F-H), confirming a role for Memo in FGF23-driven cell signaling in the kidney. Transcripts of FGFR regulator SEF, encoded

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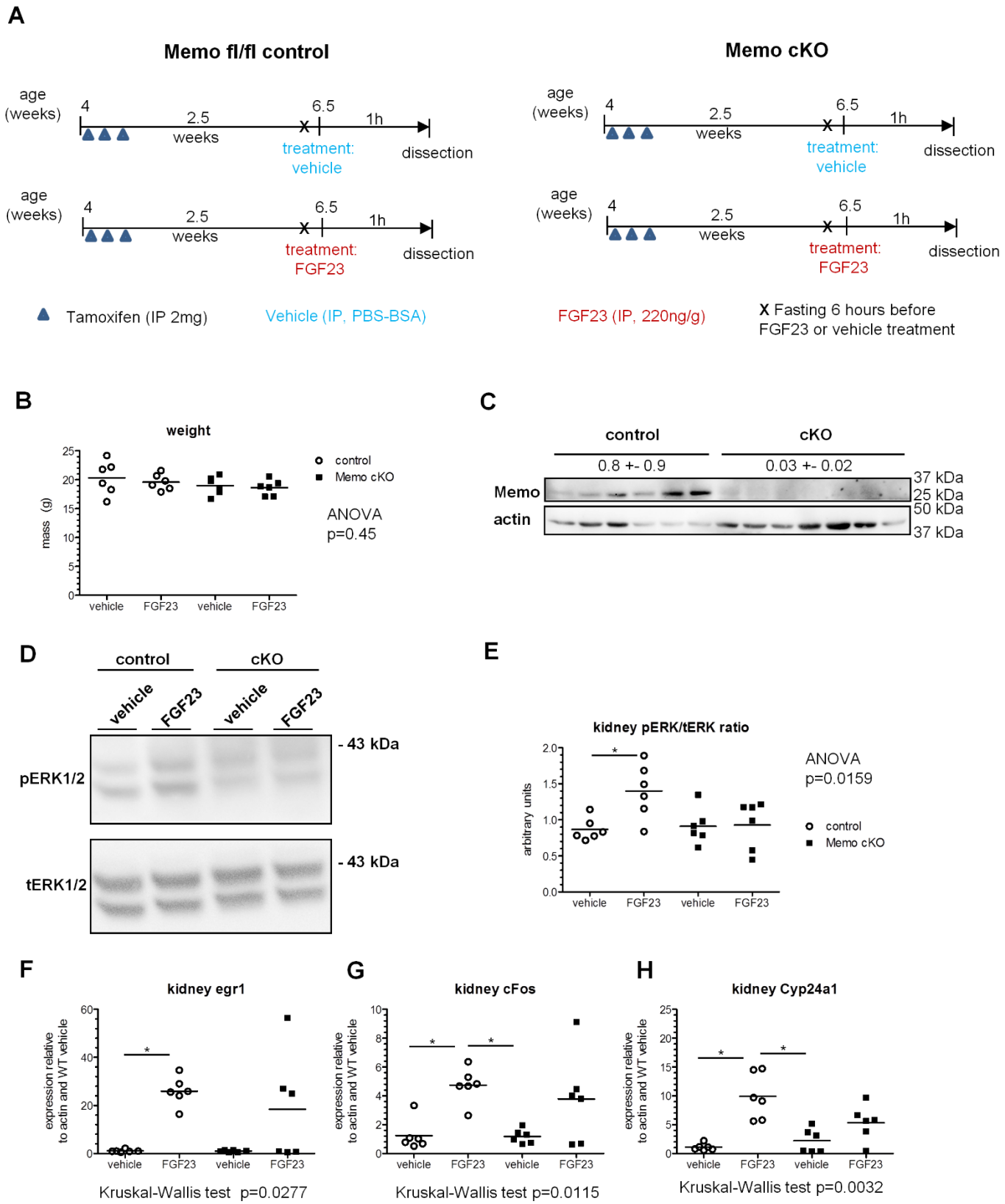

Figure 1. Impaired FGF23-induced renal signaling in whole-body Memo KO (cKO) mice. Mice were randomized to recombinant FGF23 or vehicle injection (A) at a time point prior to onset of phenotypes of premature aging with failure to thrive, shown by normal body weight (B) after successful ablation of Memo protein in kidney (C); densitometric quantification indicated (mean+-standard deviation), t-test p=0.046. *Figure 1 continued on next page.*

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Figure 1 continued.
Phosphorylated (p) versus total (t) renal ERK protein assessed by Western blot 1 hour after FGF23 or PBS vehicle injection (D) (representative of n=6 per condition). Densitometric quantification showed a blunted increase in pERK/tERK ratio in response to FGF23 in kidney of Memo cKO (E). Target gene responses of egr1 (F), cFos (G) and Cyp24a1 (H) were assessed by qPCR. Statistical analysis was obtained by ANOVA with Bonferroni's post-tests (B, E) or Kruskal-Wallis test with Dunn's post-test (F-H) between all groups, as appropriate. *, $p< 0.05$ in post-tests. n=6 per condition (B, D -H); n=6 controls vs. 7 Memo cKO (C).

by *Il17rd*, were unchanged across conditions (Figure 1 figure supplement 1D), as was renal protein abundance of the FGF23 co-receptor Klotho (Figure 1 figure supplement 1E).

To gain further insight into FGF23-dependent signaling pathways disturbed in Memo cKO, we assessed the transcriptome of pair-wise pooled kidney samples from FGF23-treated and vehicle-treated Memo cKO and control mice (Supplemental tables 1-5). In the acute response 1 hour after FGF23 treatment, a strong genotype effect but no overall treatment effect was visible in clustering and principal component analysis of 13362 protein-coding transcripts (Figure 1 figure supplement 2A-B). Thirteen transcripts were strongly increased by FGF23 in control genotypes but only 1 in Memo cKO (Figure 1 figure supplement 2C). Some transcripts showed a diverging FGF23 treatment effect across genotypes: *Csrnp3* encoding a regulatory subunit of protein phosphatase 1 was repressed by FGF23 in control mice but increased by FGF23 in Memo cKO (Figure 1 figure supplement 3A; Supplemental table 5). In contrast, a calcineurin-driven feedback regulator *Rcan1* (Gurda et al. 2010) was increased by FGF23 in controls but tended to decrease upon FGF23 treatment in Memo cKO (Figure 1 figure supplement 3B, Supplemental table 5). This suggests that there is an FGF23-driven activation of the FGFR4 - phospholipase Cγ - calcineurin signaling pathway in controls, but not in Memo cKO mice. *Klotho* transcripts were similar in all genotypes and treatment groups (Figure 1 figure supplement 3C, Supplemental tables 3-4). Gene ontology of transcripts was analyzed (Supplemental tables 1-5, 2nd to 4th sheet and Figure 1 figure supplements 4-6). Transcripts functionally annotated to mitogen-activated protein kinase (MAPK or ERK) phosphatase activity were significantly enriched by FGF23 treatment in Memo cKO (Figure 1 figure supplement 4 and Supplemental table 1), however FGF23-treated control mice showed much stronger enrichment (Figure 1 figure supplement 5, Supplemental table 2). Moreover, in interaction analysis of genotype and FGF23 treatment this effect remained significant (Figure 1 figure supplement 6, Supplemental table 5), indicating much stronger responses to FGF23 in Memo controls on global transcriptomic levels.

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We interpret the results as follows: The MAPK phosphatase-related transcripts which act as negative regulators on the MAPK cascade are quantitatively more affected by FGF23 in controls compared to in Memo cKO. However, the diverging alterations in *Csrnp3* and *Rcan1* rather represent a qualitative signaling alteration under Memo deletion.

Subsequently, we aimed at understanding the mechanisms of Memo by which it promotes FGF23 signaling. Memo protein itself has no phosphatase activity (Qiu et al. 2008) which could influence MAPK signaling. Importantly, however, RTK-dependent signal propagation depends on redox signaling: Protein tyrosine phosphatases that constitutively control kinase cascades are regulated by cysteine oxidation in defined spatial areas of the cell (Miki and Funato 2012; Truong and Carroll 2012), showing an increase in phosphatase activity in response to low-dose oxidants but a decrease at high-dose oxidants (Wright, Reiser, and Clanton 2009). For serine and threonine phosphatases such as calcineurin or PP1, the oxidation state of a Fe(II) atom determines their activity (Rusnak and Reiter 2000). Memo structurally resembles bacterial nonheme iron dioxygenases (Qiu et al. 2008) and has a copper-reducing redox function (MacDonald et al. 2014). Hence, we hypothesized that Memo's redox function drives FGF23-driven cellular signaling.

We used redox proteomics to determine the abundance of thiols and carbonyls as intermediates of oxidative reactions in kidney of FGF23 or vehicle-treated Memo cKO and controls. Total protein thiol content assessed by 5-iodoacetaminofluorescein (IAF) showed comparable levels between vehicle-treated control and Memo cKO mice (Figure 2A). IAF signals were 2.17-fold increased in FGF23-treated controls, but 2.54-fold in FGF23-treated Memo cKO, indicating that there are differences in FGF23-driven global cysteine oxidation between genotypes (Figure 2A). Total protein carbonyl content was 1.95-fold increased by FGF23 in controls, but only 1.45-fold in Memo cKO (Figure 2B), signifying that in Memo cKO kidneys are unable to normally process reactive intermediates (e.g. by peroxidation), which leads to uncontrolled protein thiolation.

We next determined the molecular identities of affected proteins process with altered oxidation states. We assessed the same kidney samples using 2-dimensional gel electrophoresis (2DGE) with separation by isoelectric point and mass, and used software-assisted determination of altered protein spots. Figure 2 figure supplement shows 27 protein spots that had Coomassie, IAF or 5-thiosemicarbazide (FTSC) signals which differed between controls and Memo cKO. Table 1 shows the corresponding protein

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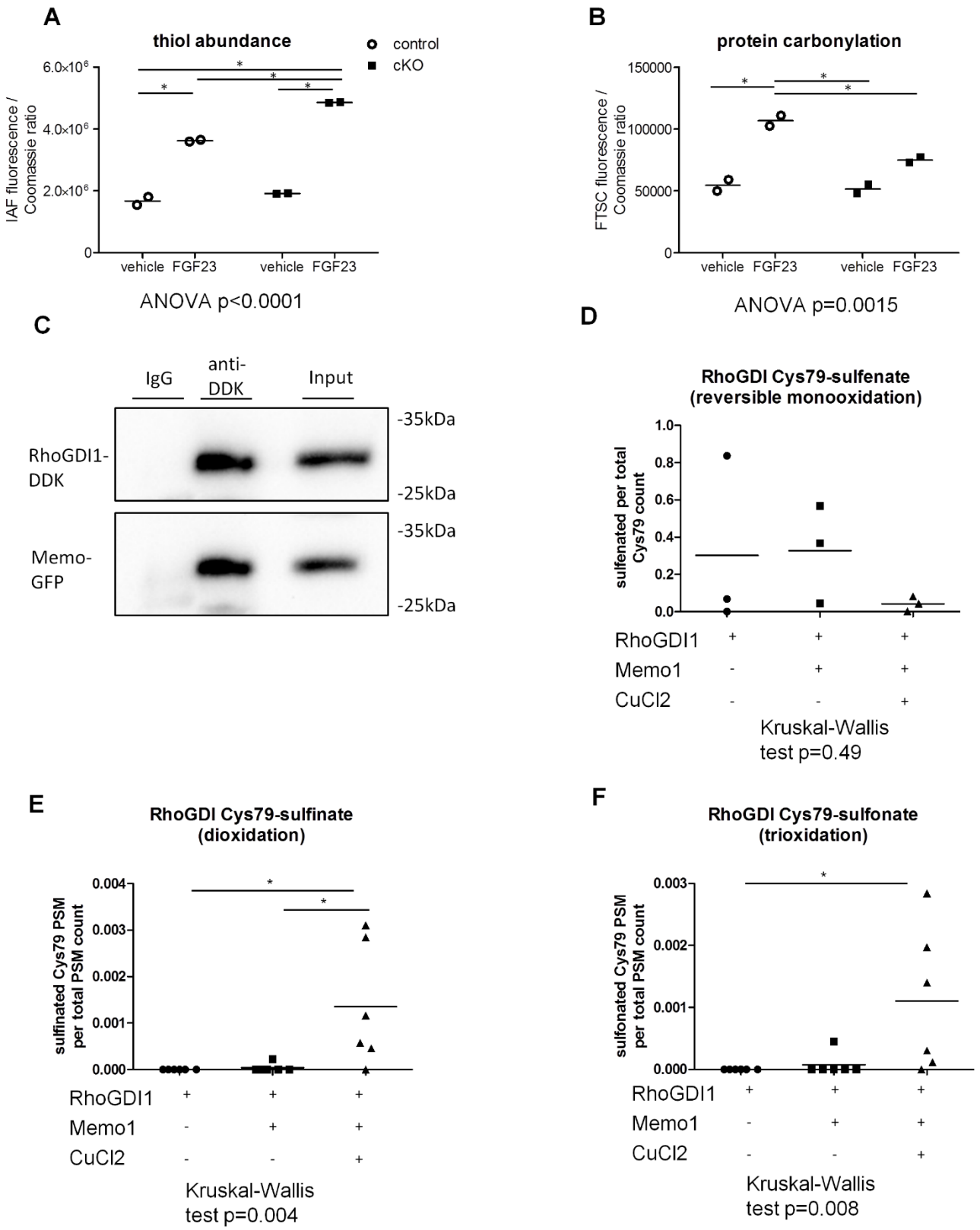

Figure 2. Redox proteomic analyses and validation of interaction between MEMO and Rho-GDI1. Quantification of the redox-sensitive thiol conjugate 5-iodoacetaminofluorescein (IAF) revealed that in response to FGF23 there was a moderate 2.17-fold increase in protein thiol content FGF23 in control mice, but a 2.54-fold increase in Memo cKO (A). *Figure 2 continued on next page.*

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Figure 2 continued.
Global protein carbonylation measured by fluorescein-5-thiosemicarbazide (FTSC) was increased 1.95-fold by FGF23 in controls, but only 1.45-fold in Memo cKO (B). Each data point represents means of 3 technical replicates of pooled kidney halves of 2 animals (n = 2x2 per condition). Measured intensities were normalized by Coomassie stained total protein. Panel C shows lysates of HEK293 cells transfected with DDK-Rho-GDI1 and GFP-Memo vectors that were immunoprecipitated with a DDK antiserum and probed for DDK-Rho-GDI1 and GFP-MEMO; representative of three independent experiments. Reversible sulfenylations (D) assessed using iodoTMT cysteine oxidation-specific tandem mass-spectroscopy label, dioxidized sulfinate (E) and trioxidized sulfonate (F) are shown in Cys79 of recombinant Rho-GDI1 alone, after incubation with MEMO or with copper-preloaded and dialyzed MEMO. Counts in D-E are normalized for total number of Peptide Scan Matches (PSM) of detected peptides. One outlier in the condition Rho-GD1 alone in E-F with values of 0.0035 (E) and 0.0059 (F) was considered implausible (e.g. contamination by H_2O_2) due to within-group difference of several orders of magnitude and was excluded from analysis; an additional sample was prepared and analyzed. Statistical analysis was obtained by ANOVA with Bonferroni's multiple testing correction (A-B) or Kruskal-Wallis test with Dunn's multiple testing correction (D-F) performed between all groups. *, $p< 0.05$ in post-tests. n=2x2 animals pooled per condition (A, B); n=3 independent experiments (C-D); n=6 independent experiments each (E-F).

identities as determined by MALDI-TOF/MS. In addition to differentially oxidized proteins associated with cytoskeleton and carbohydrate metabolism, we found one potential signaling-related protein, the Rho-GDP dissociation inhibitor 1 (Rho-GDI1), a chaperone protein of Rho-GTPases. The renal thiols of Rho-GDI1 were 1.69-fold increased in FGF23-treated Memo cKO compared to controls (Table 1).

We experimentally tested the possibility that Memo and Rho-GDI1 interact using two approaches: First, lysates from HEK293 cells transfected with vectors expressing GFP-tagged Memo1 and DDK-tagged Rho-GDI1 were immunoprecipitated with anti-DDK antibody, then probed for Memo-GFP. The results show a co-immunoprecipitation of both proteins (Figure 2C). Next, we determined if recombinant Memo can functionally affect Rho-GDI1 by oxidation of its single cysteine at position 79. We quantified ratios of oxidized and total Rho-GDI1 Cys79 using sulfhydryl-specific tandem mass tag iodoTMT in a cell-free environment. After pre-loading Memo protein with $CuCl_2$ as the putative cofactor copper, followed by dialysis to remove of free copper, we found that the reversible oxidation of Rho-GDI1 Cys79 to sulfenate was negligible (Figure 2D) but could measure irreversibly dioxidized and trioxidized Rho-GDI1 Cys79 with sulfinate (Figure 2E) or sulfonate groups (Figure 2F), respectively. These findings support the data from the 2DGE screen (Table 1) and reveal that the effects on Rho-GDI1 are similar to the previously reported effect of Memo on RhoA, its interaction partner RhoA. In this case, RhoA displayed reduced cysteines in Memo-deficient cells, but was directly oxidized by recombinant Memo (MacDonald et al. 2014). MacDonald et al. also demonstrated an increase of Memo oxidase activity upon copper loading (MacDonald et al. 2014), similar as the current findings of irreversible Cys79 oxidation. Overall, the interaction between Memo and Rho-GTPases results in differential regulation of multiprotein subunit recruitment of NADPH oxidases (MacDonald et al. 2014; Ewald et al. 2017), and the current data provide

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a novel link via Rho-GDI1 which has a known role in kidney diseases (Gee et al. 2013; Auguste et al. 2016, 1). Moreover, results from cysteine oxidation proteomics have recently highlighted extensive

Table 1: Identification of altered proteins in redox proteomics of the kidney

	Spot no.	Identified protein	Mw	Protein score	Total ion score	cover-age (%)	N of peptides identified	P Value	Average normalized volumes	
									Ctrl-V	Memo cKO-V
Coomassie	386	60 kDa heat shock protein, mitochondrial	61	666	447	36	19	0.008	1.68E+05	2.69E+05
	360	Succinyl-CoA:3-ketoacid coenzyme A transferase 1, mitochondrial	56	360	299	26	10	3.47E-05	2.67E+05	1.53E+05
	416	Alpha-1-antitrypsin 1-5	46	54	34	13	3	0.008	6.95E+04	3.41E+04
	628	Phosphatidylethanolamine-binding protein 1	21	131	110	25	2	0.038	4.00E+05	1.86E+05
	534	Aminoacylase-1	46	126	107	14	7	0.033	2.27E+05	1.27E+05
Thiols	175	Actin, cytoplasmic 1	42	375	246	32	13	0.047	2.49E+07	1.23E+07
	78	Heat shock cognate 71 kDa protein	71	433	250	22	10	0.024	3.39E+06	2.03E+06
	244	Pyruvate dehydrogenase E1 component subunit beta, mitochondrial	39	374	193	37	13	9.58E-04	7.69E+05	3.30E+06
	132	Glutathione synthetase	52	684	495	38	16	0.035	3.01E+06	1.28E+06
	202	L-lactate dehydrogenase B chain	37	611	334	52	19	0.001	6.05E+06	3.18E+06
	216	Malate dehydrogenase, cytoplasmic	37	321	296	31	10	0.024	1.02E+07	3.83E+06
Carbo-nyls	50	78 kDa glucose-regulated protein	72	641	415	28	15	0.047	2.33E+06	8.25E+05
	32	Serotransferrin	79	769	484	36	20	0.028	1.20E+06	2.72E+06
	117	Protein disulfide-isomerase A3	57	487	286	32	13	0.015	4.22E+05	1.94E+05
									Ctrl-F	Memo cKO-F
Coomas-sie	535	Aminoacylase-1	46	231	137	21	7	0.008	5.93E+05	3.46E+05
	507	Alpha-enolase	47	414	285	23	7	0.02	1.69E+05	3.07E+05
	735	Periaxin	148	36	28	6	10	0.013	4.32E+05	7.26E+05
Thiols	236	Selenium-binding protein 1	53	997	665	52	29	0.024	3.06E+06	6.32E+06
	267	Malate dehydrogenase, cytoplasmic	37	552	506	46	20	0.001	5.16E+05	3.19E+06
	378	Rho GDP-dissociation inhibitor 1	23	116	92	20	2	0.035	1.83E+06	3.10E+06
	212	Glutathione synthetase	52	185	106	28	8	0.008	6.58E+06	1.70E+06
	253	L-lactate dehydrogenase B chain	37	575	321	48	15	0.04	1.93E+06	1.18E+06
	305	Pyruvate dehydrogenase E1 component subunit beta, mitochondrial	39	366	237	25	7	0.009	5.09E+06	8.79E+05
	175	Actin, cytoplasmic 1	42	142	92	12	5	0.033	1.53E+07	3.94E+06
Carbonyls	67	ATP synthase subunit beta, mitochondrial	56	554	356	31	13	0.232	6.03E+06	1.11E+07
	146	Malate dehydrogenase, cytoplasmic	37	399	432	30	13	0.26	5.08E+05	9.52E+05
	96	Selenium-binding protein 1	53	691	394	46	22	0.156	1.19E+06	6.80E+05

Mw, molecular weight in kDa. Ctrl, control. V, vehicle. F, FGF23. P-Values determined by t-test.

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alterations in the oxidation of Rho-GTPase subfamilies RHO and RAC in response to epidermal growth factor-driven RTK signaling (Behring et al. 2020).

Protein abundance of Rho-GDI1 was comparable between genotypes and treatment groups (Figure 3A, quantification in Figure 3E). With total Rho-GDI1 unchanged, the less abundant Rho-GDI1 thiols in FGF23-treated control mice would correspond to transient oxidation of Rho-GDI1 Cys79 to sulfenate in order to liberate and activate a bound Rho-GTPase. This has previously been demonstrated for the RhoA – Rho-GDI1 complex (Kim et al. 2017). Memo is required for growth factor-driven RhoA relocalization to the membrane (Zaoui et al. 2008), and strikingly, RhoA protein abundance and activity were increased in the kidney of Memo cKO mice (Figure 3B, 3E-3F). The abundance of Rho-GTPase Rac1 was unchanged in all conditions (Figure 3C, quantification in Figure 3E). However, FGF23 treatment caused a trend to 10% increase in Rac1 activity in kidney of control mice (Figure 3F), similar to the previously reported effect of FGF2 treatment in cell culture (Shin et al. 2006; Barrios and Wieder 2009). In the kidney of Memo cKO animals, however, this trend was absent.

These findings suggest that the Rho-GTPases network is disturbed in the absence of Memo, which has implications for the well-characterized intracellular redox signaling by Rac1 leading to NADPH oxidase assembly (Flinder et al. 2011) and for RTK endocytosis which is regulated by both RhoA and Rac1 and is a cue to stop FGFR-mediated MAPK signaling (Lamaze et al. 1996; Grassart et al. 2008; Bryant, Wylie, and Stow 2005). Intriguingly, *Snap91* was the top upregulated transcript (Figure 3 figure supplement) in FGF23-treated Memo cKO compared to FGF23-treated controls. *Snap91* encodes a clathrin coat assembly protein that aids in recognizing membrane receptor cargo to drive endocytosis (Reider and Wendland 2011). Collectively, this raises exciting questions about the potential of Memo at the intersection of RTK signaling, redox biology and membrane trafficking of RTK.

To conclude, here we present results that shed light on the role of Memo in response to FGF23-driven RTK signaling in the kidney. Our data provide first insights into the *in vivo* physiological role and the underlying mechanisms showing how the evolutionarily conserved moonlighting protein Memo simultaneously modulates multiple RTK-dependent signaling pathways.

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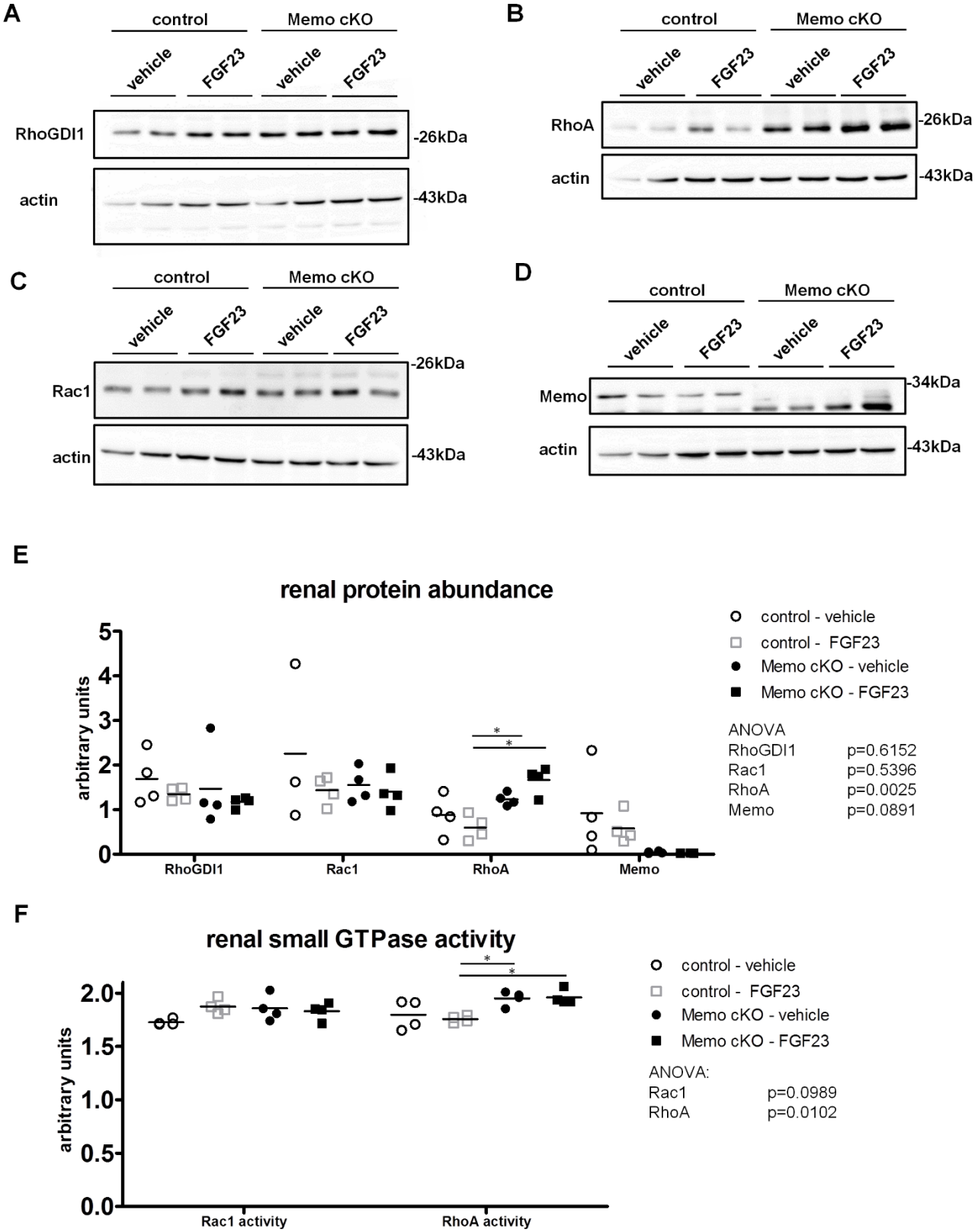

Figure 3. The Rho-GTPase network in kidneys from whole-body Memo cKO (cKO) mice is malfunctioning. Western blotting of renal lysates shows unchanged levels of Rho-GDI1 (A) and Rac1 (C) in Memo cKO compared to control mice; RhoA (B) was increased in Memo cKO mice. Protein band at 33kDa, specific for Memo protein, was absent in kidney from Memo cKO (D). Densitometric quantifications of Western blots A-D, relative to actin are shown in E. Renal Rac1 activity showed a trend to increase in control mice treated with FGF23, but was not significantly changed (ANOVA p=0.099). (F, left panel). *Figure 3 continued on next page.*

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Figure 3 continued.
Renal RhoA activity was significantly higher in Memo cKO (F, right panel). Statistical analyses in E-F were performed by ANOVA with Bonferroni correction for multiple testing (6 post-tests each). *, post-test p<0.05. n=4 per condition in A-F. n=4 per condition (A-F); except n=3 in E (Rac1: control vehicle) due to undetectable loading control.

Methods

Animal studies

All animal experimental protocols were approved by the veterinary service of the Canton de Vaud, Switzerland. Mice were fed standard chow (TS3242 Kliba Nafnag, Kaiseraugst, Switzerland) and were kept on 12/12 or 14/10 light-dark cycles. Mice floxed for exon 2 of the *Memo1* gene (Haenzi et al. 2014) backcrossed to C57BL/6J background over 10 generations were crossed with CreERTM transgenic mice (Hayashi and McMahon 2002) carrying a tamoxifen-inducible Cre recombinase controlled by a beta-actin promoter/enhancer. Genotypes were determined by PCR of ear punch biopsy DNA using primers: Memo forward 5'-*CCCTCTCATCTGGCTTGGTA*-3', Memo reverse 5'-*GCTGCATATGCTCACAAAGG*-3', Cre forward 5'-*AGGTTCGTGCACTCATGGA*-3', Cre reverse 5'-*TCACCAGTTTAGTTACCC*-3'. For the current study, all animals used were males, with the exception of the experiments reported in Figure 3 in which there was a sex ratio of 50:50 in each genotype. Loss of Memo was induced by 3 daily intraperitoneal injections with 2mg tamoxifen at age 4 weeks (T5648 Sigma-Aldrich, distributed through Merck, Buchs Switzerland). Memo$^{fl/fl}$ littermates without Cre but treated with tamoxifen served as controls. For FGF23 treatments, mice aged 6.5 weeks were starved for 6 hours and intraperitoneally injected with 220ng/g body weight of recombinant mouse FGF23 (2629-FG-025 R&D Systems, Minneapolis, MN, USA) in PBS-BSA 0.1% or PBS-BSA 0.1% vehicle with injection volume of 4µL/g body weight. One hour later mice were dissected, a protocol adapted from (Gattineni et al. 2014). Treatments were randomly allocated by flipping a coin. Animals were euthanized after 1h by terminal exsanguination under anesthesia.

Transfection

Hek293 cells were obtained from ATCC and were tested negative for mycoplasma using LookOut® PCR-based kit (MP0035-1KT Sigma-Aldrich). Cells were transfected with GFP-tagged Memo1 (MG204118 OriGene Technologies Inc., Rockville MD, USA) and Myc-DDK-tagged Rho-GDI1 (ARHGDIA) (MR202112 OriGene) using Lipofectamine 3000 (L3000015 Invitrogen, distributed through Thermo Fisher Scientific, Reinach, Switzerland).

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Redox proteomics

Kidney halves pooled from 2 mice per sample were homogenized in 10 mM Tris-HCl, pH 7.2, 250 mM sucrose, 1 mM EDTA, 150 mM KCl and 1 mM PMSF and spun down. Protein thiols and carbonyls were labelled with either 0.2 mM 5'-iodoacetamido fluorescein (IAF) or 1mM fluorescein-5-thiosemicarbazide (FTSC), respectively, and incubated for 150 min at 37 °C in the dark. Proteins were precipitated with 20 % TCA and centrifuged at 20,000 ×g (3 min, 4 °C). Protein pellets were resuspended and washed with 100% ethanol/ethyl acetate (1:1) and 96% acetone, respectively, for carbonyl and thiol groups. Pellets were resuspended in Tris-HCl 0.5 M pH 6.8, glycerol 10%, SDS 0.5% and bromophenol blue and applied to SDS-PAGE. Gels were scanned for fluorescence in a Typhoon Trio Scanner 9400 (Control v5.0 + variable Mode Imager-RA 501: PRT<I/06/004, GE Healthcare, Buckingshamshire, UK; excitation, 490-495 nm; emission, 515–520 nm). Protein-associated fluorescence intensities (arbitrary units, AU) were analyzed using Quantity One image analysis software (BioRad, Hercules, CA, USA). Gels were stained with Colloidal Coomassie Brilliant Blue G250 and scanned.

For two dimensional analysis, proteins were first separated by pI (first dimension: isoelectric focusing IEF), followed by orthogonal separation according to molecular weight (second dimension: SDS-PAGE). Proteins were rehydrated in 5 M urea, 2 M thiourea, 2% CHAPS, 4% ampholyte (Pharmalyte 3-10, Amersham-Pharmacia Biotech, Little Chalfont, Bucks, UK), 1% Destreak reagent (Amersham-Pharmacia Biotech, Buckinghamshire, UK), and trace amounts of bromophenol blue, and then immobilized in 7 cm IPG strips (pH 3-10 : 70×3×0.5 mm) and a linear gradient (NL) (GE Healthcare Immobiline™ Dry Strip IPG, GE17-6001-11 Bio-Sciences AB, Bio-Rad, Hercules, CA, USA). Proteins were focused at room temperature in a Protean IEF Cell (Bio-Rad) for at least 15 h, according to the following steps: (1) a linear voltage increase until 250 V for 15 min, (2) 10,000 V for 2 h (50 µA/ strip), (3) focusing at 20,000 V, and (4) hold at 500 V. Following IEF, strips were equilibrated for 20 min in equilibration buffer (6 M urea, 0.375 M Tris, pH 8.8, 2% SDS, 20% glycerol, containing 2% DTT) and then for 20 min in equilibration buffer containing 2.5% iodoacetamide. IPG strips were loaded onto 12% SDS-PAGE gels (PROTEAN Plus Dodeca Cell, BioRad). Gels were scanned for fluorescence as above and stained with Coomassie Blue followed by densitometry. Progenesis SameSpots Software (S/No.62605/3787; Nonlinear USA Inc, Durham, NC USA) was used to normalize FTSC/ IAF-labeled protein spots and Coomassie-staining intensity. Fluorescence spots were normalized to protein intensity for the same gel.

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All experiments were performed in triplicates. Images of 2D gels were subjected to landmarking alignment so that corresponding spots were matched with each other, based on 3D Gaussian distribution after raw image correction and background subtraction. Spot intensities were normalized. Differences between protein spots in 2D gel images were automatically determined.

Spots with differences of $p<0.05$ by ANOVA were manually excised from colloidal Coomassie-stained 2-DE gels. Proteins were extracted and enzymatically digested using a Perkin Elmer – Janus automated workstation. Following digestion, samples were reconstituted in 0.1% formic acid and analyzed using a Dionex U3000 Liquid Chromatography System (Dionex, Sunnyvale, CA, USA) and the Daltonics HCT Ion Trap Mass Spectrometer (Bruker, Glasgow, UK). The peptide fragment mass spectra were acquired in data-dependent AutoMS(2) mode with a scan range of 300–1500 m/z, and up to three precursor ions were selected from the MS scan (100–2200 m/z). Precursors were actively excluded within a 1.0-min window, and all singly charged ions were excluded. Following LC-MS/MS, Mascot Generic Files were created and the MASCOT database NCBInr was searched using Matrix Science webserver (www.matrixscience.com). The default search parameters were: enzyme = trypsin; maximum number of missed cleavages = 1; fixed modifications = carbamidomethyl (C); variable modifications = oxidation (M); peptide tolerance ±1.5 Da; MS/MS tolerance ± 0.5 Da; peptide charge = 2+ and 3+. Identified proteins were considered if a MASCOT score above 95% confidence was obtained ($p < 0.05$) and at least one peptide was identified with a score above 95% confidence ($p < 0.05$). This analysis was conducted at the Proteomics Core Facility of the University of Aberdeen, UK. All mass spectrometry proteomics data from these experiments have been deposited to the ProteomeXchange Consortium via the MassIVE repository with the dataset identifier PXD022342.

Cysteine oxidation analyses with recombinant protein

Amino acids 24-204 of human Rho-GDI1/ARHGDIA protein expressed in *E. coli* was obtained from Novusbio (NBP1-50861, Centennial, CO, USA NBP1-50861), with >95% purity by SDS-PAGE and supplied in 20mM Tris-HCl buffer (pH 8.0), 1mM DTT, 10% glycerol without preservatives. Recombinant human Memo expressed in *E. coli* with >95% purity, supplied in 20 mM Tris-HCl buffer, pH 8.0, 50% glycerol, 5 mM DTT, 300 mM NaCl, 2 mM EDTA was from Antibodies-online (ABIN2130536, Aachen, Germany). Copper-reducing oxidant activity of this batch of recombinant Memo protein is reported in (Moor et al. 2020).

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First, as a putative metal cofactor pre-loading step, 2µg Memo was incubated with RCM buffer with or without 10µM $CuCl_2$. After 30 minutes incubation at 4°C the free $CuCl_2$ was removed by Slide-A-Lyzer™ MINI Dialysis Device, 3.5K MWCO (69550 Thermo Fisher Scientific) against RCM buffer for 30 minutes at 4°C. Proteins were incubated for 15 minutes as 1000ng Rho-GDI1 alone; 1000ng Rho-GDI1 with 250ng Memo; 1000ng Rho-GDI1 with 250ng $CuCl_2$ pretreated Memo; and 1000ng Rho-GDI1 with 100µM of hydrogen peroxide as a positive control (H1009 Sigma-Aldrich), all in duplicates for each experiment, to individually label the total cysteine and the oxidized cysteine content of Rho-GDI1.

The samples for total cysteine content labeling were reduced with 5mM Tris (2-carboxyethyl) phosphine (TCEP) (646547 Sigma-Aldrich). Total cysteine content was labeled with iodoTMTsixplex™ (90102 Thermo Fisher Scientific) followed by quenching with 20mM 1DTT (43816 Sigma-Aldrich).

The samples for oxidized cysteine content labeling were treated with 500mM iodoacetamide (I6125 Sigma-Aldrich) for 30 minutes followed by quenching with 20mM DTT and reducing with 5mM TCEP. The oxidized cysteine content of the samples was labeled with iodoTMTsixplex™.

Samples were precipitated with the SDS-PAGE Clean-Up Kit (10074304 GE Healthcare Life Sciences, distributed through Thermo Fisher Scientific) and separated on SDS-PAGE, followed by Coomassie Blue staining and manual band excision. Proteins were digested by chymotrypsin at 50°C for 5 hours and analyzed by LC-MS/MS (PROXEON coupled to a QExactive HF mass spectrometer, Thermo Fisher Scientific) with two injections of 5µl digests. Peptides were trapped on a µPrecolumn C18 PepMap100 (5µm, 100 Å, 300 µm×5mm, Thermo Fisher Scientific) and separated by backflush on a C18 column (5µm, 100 Å, 75 µm×15 cm, C18) by applying a 20-minute gradient of 5% acetonitrile to 40% in water, 0.1% formic acid, at a flow rate of 350nl/min. The Full Scan method was set with resolution at 60,000 with an automatic gain control (AGC) target of 1E06 and maximum ion injection time of 50ms. The data-dependent method for precursor ion fragmentation was applied with the following settings: resolution 15,000, AGC of 1E05, maximum ion time of 110 milliseconds, mass window 1.6*m/z*, first mass 100m/z, collision energy 27, under fill ratio 1%, charge exclusion of unassigned and 1+ ions, and peptide match preferred, respectively. Spectra were interpreted with Proteome Discoverer 2.4.0.305, with chymotrypsin rules allowing up to 8 missed cleavages, using variable modification of carboamidomethylated (+57.021Da), dioxidation (+31.990Da), trioxidation (+47.985Da) and iodoTMT labelling (+324.216Da) on Cys, and variable modification of oxidation (+15.99Da) on Met, and acetylation (+42.011Da) on protein N-Term.

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Parent and fragment mass tolerances were set to 10ppm and 0.02Da, respectively. Strict target false discovery rate for highly confident peptide-spectrum matches was set to 0.01. Protein identifications were only accepted when two unique peptides fulfilling the 1% FDR criterium were identified. Results are displayed as PSM counts for each cysteine oxidation state normalized for total PSM counts. This analysis was performed at the Proteomics Mass Spectrometry Core Facility of University of Bern. All mass spectrometry proteomics data from recombinant protein analyses have been deposited to the ProteomeXchange Consortium via the PRIDE repository with the dataset identifier PXD022382.

Immunoprecipitation

For immunoprecipitation Protein G Sepharose beads (P3296 Millipore, distributed through Merck, Buchs Switzerland) were blocked with 1% BSA (A9647 Sigma-Aldrich) and incubated with anti-DDK Monoclonal Antibody (TA50011-100 OriGene) and Mouse IgG2a Isotype Control (02-6200 Invitrogen, distributed through Thermo Fisher Scientific) in 1:200 dilution for 45 minutes at 4°C. After washing with RIPA (R0278 Sigma-Aldrich), the cell lysate was added to the beads in 5:1 dilution and incubated overnight. Beads were resuspended in RIPA and 4x SDS Sample buffer, heated up to 95°C and spun down.

Western blot

For immunoblotting, immunoprecipitates or proteins of cells and tissues were lyzed in RIPA buffer or NP-40 buffer (150mM NaCl, 50 mM HEPES pH7.4, 25mM NaF, 5mM EGTA, 1mM EDTA, 1% Nonidet P-40, 2M Na ortho-vandate and 1mM DTT supplied with protease inhibitors leupeptin 10µg/L, aprotinin 10µg/L, and PMSF 1mM) were prepared and denatured. Proteins were separated by SDS-PAGE and transferred onto nitrocellulose or PVDF, stained by Ponceau S, blocked in dried nonfat milk 5%-TBST or bovine serum albumin (A9647 Sigma-Aldrich) 3% in TBST before incubation with primary antibodies against Memo (1:2000, produced in-house (Haenzi et al. 2014) or 1:1000, HPA042603 Sigma-Aldrich), GFP (1:1000, G1544 Sigma-Aldrich), or FLAG (1:1000, F7425 Millipore), pERK (1:1000, sc-7383 Santa Cruz, Dallas TX, USA), tERK (1:1000, sc-93 Santa Cruz), actin (1:2000, A2066 Sigma-Aldrich), Rho-GDI1 (1:500, ABIN969501 Antibodies-online), Rac1 (1:500, ab33186 Abcam, Cambridge UK), RhoA (1:500, NBP2-22529 NovusBio) and Klotho (1:1000, AF1819 R&D Systems). Membranes were incubated with anti-mouse or anti-rabbit horseradish peroxidase-conjugated secondary antibodies (Milian Analytica, Rheinfelden Switzerland or ImmunoResearch, distributed through LubioScience

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GmbH, Zürich, Switzerland) and imaged using Fusion Solo (Witec AG, Sursee Switzerland) or a ChemiDoc XRS+ System (BioRad). For reprobing, membranes were stripped using a low pH buffer (25mM glycine-HCl, pH2, 0.4% (w/v) SDS).

Biochemical analyses

Serum electrolytes were analyzed by the Lausanne University Hospital: Total calcium (NM-BAPTA method), phosphate (phosphomolybdate method), and creatinine (modified Jaffé method).

Rho-GTPase activity assays

For quantification of Rho-GTPase activity in kidney homogenates, colorimetric Rac1 and RhoA G-LISA Activation Assay Kits (BK128-S and BK124-S Cytoskeleton Inc., Denver CO, USA) were used according to the manufacturer's instructions.

RNA isolation

Frozen kidneys were homogenized with metal beads in a TissueLyser (Qiagen, Hombrechtikon, Switzerland). RNA was extracted using TRI Reagent Solution (AM9738 Ambion, Austin, TX, USA) for all downstream applications.

qPCR

One µg of RNA per kidney half was reverse-transcribed using PrimeScript RT Reagent kit (RR037 TAKARA, Shiga, Japan). Two µl of cDNA was used for quantitative real-time PCR to assess gene mRNA expression. Assays were performed using SYBR Green (Applied Biosystems, Foster City, CA UA) on a 7500 Fast machine (Applied Biosystems). Samples were run in triplicates in 20uL total volume for each gene, and actin or GAPDH was used for normalisation. Melting curves were obtained for every run. Program settings were: 95°C 20 seconds, 40 cycles (95°C 3 sec, 60°C 30 sec), and for melting curve stage: 95°C 15sec, 60°C 1min, rising at 1% ramp speed to 95°C (15sec), 60°C 15sec. Data were analyzed using the delta-delta CT method. Primers were ordered from Microsynth (Balgach, Switzerland), and sequences are shown in Supplemental table 6.

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RNAseq

For RNAseq, extracted RNA from mouse kidney halves was pairwise pooled to 3 individual pools for each of 4 experimental conditions (2 treatment states, 2 genotypes). Quality of mRNA was assessed and verified using a Fragment Analyzer (Advanced Analytical Technologies Inc.). Following quality validation, TruSeq Stranded RNA Sample Prep Kit (20020596 Illumina, San Diego CA, USA) was used with 1ug of total RNA for the construction of sequencing libraries. cDNA was sequenced using the Illumina HiSeq2500 platform and single-end reads (1x100). All sequencing data are accessible on National Center for Biotechnology Information, Sequence Reads Archive, Accession: PRJNA672305. Purity-filtered reads were trimmed with Cutadapt (v. 1.3, Martin 2011), filtered using seq_crumbs (v. 0.1.8) and aligned against the *Mus musculus* (Ensembl version GRCh38.82) transcriptome using STAR (v. 2.4.0f1). Data were normalized with the Trimmed Means of M-Values (TMM) method of the edgeR, and transformed with voom method of bioconductor package limma. Genes expressed at low levels were removed when none of the samples had more than 1 count per million reads. Only genes labeled as “protein coding” were retained for the statistical tests.

Statistical analyses

Analyses of parametric continuous variables were performed by t-test or by ANOVA, followed by post-tests using Bonferroni correction for multiple testing across all experimental groups. Analyses of nonparametric continuous variables were assessed by Kruskal Wallis test, followed by Dunn’s correction for multiple testing across all experimental groups. Analyses were performed in GraphPad Prism 5.0, and two-tailed p-values of <0.05 were considered significant.

Analyses of RNAseq data were performed with limma (R version 3.2.1, limma version 3.22.7). Because the experiment has a 2x2 factorial design with two mouse genotypes and two treatment states, linear models were generated with all 4 groups as factors. Subsequently, contrasts of interest were extracted using moderated t-tests for the following groups: 1. "KOFGF-KOV" (treatment effect in Memo cKO), 2. "WTFGF-WTV" (treatment effect in floxed control genotype), 3. "KOFGF-WTFGF" (KO genotype effect in FGF23-treated mice), 4. "KOV-WTV" (KO genotype effect in vehicle-treated mice), and 5. "(KOFGF-KOV)-(WTFGF-WTV)" for the interaction between genotype and treatment. P-values were adjusted by the Benjamini-Hochberg method, controlling for false discovery rate (FDR) across all 5 contrasts as a correction of multiple testing conditions. All sequencing data are available on National Center for Biotechnology Information, Sequencing Reads Archive under the identifier PRJNA672305. Gene

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ontology enrichment was assessed in GOrilla, accessed on Nov 17th 2020 (Eden et al. 2009) using differentially expressed genes sorted by adjusted p-values.

Accession codes

RNAseq data: NCBI SRA, Accession: PRJNA672305 (openly available)

LC-MS/MS kidney data: proteomeXchange accession PXD022342 (openly available)

LC-MS/MS recombinant protein data: proteomeXchange PXD022382 (available through username**:** reviewer_pxd022382@ebi.ac.uk Password**:** vnw1DyyI)

Raw data: Zenodo.org, DOI: 10.5281/zenodo.4293503 (openly available)

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Conflicts of interest

All authors declare that they have no conflict of interest to report.

Author participation

MBM and BH conceived the study. KB, SKR, SB, BH, DS, OB and MBM participated in experimental design. KB, SKR, SB, FD, AP, YZ, and MBM performed experiments. KB, SKR, DS, NEH, OB and MBM participated in data analysis and interpretation. DS and NEH provided crucial laboratory materials. OB

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and MBM acquired funding for the research. OB and MBM supervised the research. KB and MBM wrote the manuscript. All authors critically read and commented on the manuscript and agreed to manuscript submission.

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