Optogenetic activation of the neural circuit between dorsal raphe nucleus and the pre-Bötzinger complex contributes to inhibition of the seizure-induced respiratory arrest in the DBA/1 mouse SUDEP model

HaiXiang Ma[1,2], Qian Yu[1,2],, Yue Shen[1,2,3], XiTing Lian1, LeYuan Gu[1,2], Han lu[4], HaiTing Zhao[5], Chang Zeng[6], Kazuki Nagayasu[7], HongHai Zhang*[1,2,3]

[1]Department of Anesthesiology, Affiliated Hangzhou First People's Hospital, Zhejiang University School of Medicine, Hangzhou, 310006, China

[2]Department of Anesthesiology, the Fourth Clinical School of Medicine, Zhejiang Chinese Medical University, Hangzhou, 310006, China

[3]Department of Anesthesiology, Hangzhou First People's Hospital, Nanjing Medical University, Hangzhou, 310006, China

[4]Department of Anesthesiology, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China

[5]Department of Neurology, Xiangya Hospital, Central South University, Changsha 410008,

[6]China.Health Management Center，Xiangya Hospital，Central South University，Changsha，410008

[7]Department of Molecular Pharmacology, Graduate School of Pharmaceutical Sciences, 17 Kyoto University

HaiXiang Ma , Qian Yu and Yue Shen contributed equally to this work

Corresponding authors: HongHai Zhang

Email address: zhanghonghai_0902@163.com

Abstract

Sudden unexpected death in epilepsy (SUDEP) is the fatal cause leading to the death of epilepsy patients, especially those with anti-epileptic drug resistance. However, the underlying mechanism of SUDEP remains elusive. Our previous study demonstrated that enhancement of serotonin (5-HT) synthesis by intraperitoneal (IP) injection of 5-hydroxytryptophan (5-HTP) significantly reduced the incidence of seizure-induced respiratory arrest (S-IRA) in DBA/1 mice SUDEP models. Given that 5-HT2A receptor (5-HT2AR) plays an important role in mediating the respiration system in brain, we hypothesized that 5-HT2AR is of great significance to modulate S-IRA and SUDEP. To test

this hypothesis, we examined whether the decreased incidence of S-IRA evoked by either acoustic stimulation or either acoustic stimulation or pentylenetetrazole (PTZ) injection by administration with 5-HTP will be blocked by administration with ketanserin (KET), a selective antagonist of 5HT2AR, in DBA/1 mice SUDEP models to examine the role of 5-HT2AR in modulating S-IRA. Our results suggested that the decreased incidence of S-IRA by 5-Hydroxytryptophan (5-HTP), a precursor for central nervous system (CNS) serotonin (5-HT) synthesis, was significantly reversed by IP or intracerebroventricularly (ICV) injection of ketanserin in our models. Furthermore, given that 5-HT2AR locating in the pre-Bötzinger complex (PBC) which plays a key role in regulating the respiratory rhythm, we wonder whether the lower incidence of S-IRA by IP with 5-HTP was significantly reversed by KET which inhibits 5-HT2AR in the above models and acts on the 5-HT2AR in the pre-Bötzinger complex. To test our hypothesis, we activated the neural circuit between dorsal raphe nucleus and the pre-Bötzinger complex by optogenetics technology. Our results indicated that the lower incidence of S-IRA evoked by PTZ by stimulating the TPH2-ChETA, a variant of channelrhodopsin-2 (ChR2), neurons in the dorsal raphe nucleus (DR) in DBA/1 mice and the suppressant effects was significantly reversed by administration of KET in the bilateral pre-Bötzinger complex without changing electroencephalogram (EEG) activity. The neural circuit between DR and the pre-Bötzinger complex was confirmed by injection of cholera toxin subunit B (CTB)555, a nerve tracer, in DR or the pre-Bötzinger complex, respectively. The neural activity of calcium signal evoked by PTZ in the pre-Botzinger complex during seizures was significantly reduced by photostimulation of the DR by photometry recordings. Taken together, our findings suggested that targeting the neural circuit between the DR and the pre-Bötzinger complex in which 5-HT2AR plays a critical role in regulating this process would be promising to prevent SUDEP.

Keywords: sudden unexpected death in epilepsy (SUDEP); 5-Hydroxytryptophan (5-HTP); ketanserin (KET); pre-Bötzinger complex; dorsal raphe nucleus(DR); 5HT2A receptor; pentylenetetrazole (PTZ); GCaMP6f

1. INTRODUCTION

Growing studies have shown that sudden unexpected death in epilepsy (SUDEP) is the most important cause leading to the death of epilepsy patients, especially those who suffered from

antiepileptic drug resistance [1-4]. The latest advancements of the pathogenesis of SUDEP demonstrated that cardiopulmonary dysfunction plays an important role in the occurrence of SUDEP[1-5]. Our previous findings showed that seizure-induced respiratory arrest (S-IRA) can lead to the occurrence of SUDEP evoked by either acoustic stimulation or pentylenetetrazole (PTZ) in DBA/1 murine models [5-12], and the incidence of S-IRA by acoustic stimulation or PTZ was significantly reduced by 5-Hydroxytryptophan (5-HTP), a precursor for serotonin synthesis in two models of SUDEP [5]. However, the target of 5-HTP-mediated reduction of S-IRA and SUDEP in the brain remains unclear and still needs to be further explored. Due to that 5-HT2AR has emerged as a key factor in mediating respiratory function in brainstem[14-15], we wonder if the application of 5-HTP could reduce the incidence of S-IRA by targeting 5-HT2AR in the brain. Therefore, we hypothesized that the suppressant effects of 5-HTP on S-IRA would be reversed by ketanserin (KET), a selective antagonist for 5-HT2AR, in the DBA/1 mouse SUDEP models and that it may be a leading therapeutic target to prevent SUDEP.

To investigate the roles of 5-HTP and 5HT2AR in the pathogenesis of S-IRA and SUDEP, we continued to choose acoustic stimulation and PTZ injection DBA/1 mice SUDEP models to test the reversal effects of KET based on our previous study. The lower incidence of S-IRA by 5-HTP was significantly reversed by both intraperitoneal (IP) and intracerebroventricular (ICV) injection of KET in two models, respectively. Different from our previous study that atomoxetine reduced the incidence of S-IRA in DBA/1 mice by acoustic stimulation without affecting seizure behavior [8-9], administration of 5-HTP significantly reduced the incidence of S-IRA through its anticonvulsant effects in our models. The anticonvulsant effects of 5-HTP were basically consistent with other studies that activation of 5-HT neurons can reduce the severity of seizure [16-17]. Meanwhile, administration of KET reversed the lower incidence of S-IRA by 5-HTP with producing only partly anticonvulsant effects, which suggested that 5-HT2AR in brainstem may be a specific target to prevent S-IRA and SUDEP. Furthermore, considering that 5-HT2AR located in the pre-Bötzinger complex which plays a key role in modulating respiration rhythm[12], and that the incidence of S-IRA in DBA/1 mice can be significantly reduced by optogenetic activation of TPH2-ChR2 neurons in DR based on our previous findings, we further activated

TPH2-ChETA neurons in the DR of DBA/1 mice to observe whether the suppression of S-IRA in PTZ injection model by photostimulating the DR depends on activating 5-HT2AR located in the pre-Bötzinger complex or not. It turned out that the lower incidence of S-IRA evoked by PTZ by optogenetic activation of TPH2-ChETA neurons in the DR was remarkably reversed by injection of KET into the pre-Bötzinger complex. Meanwhile, the neural circuit between the DR and the pre-Bötzinger complex was confirmed by injection of tracer CTB-555 into the DR and the pre-Bötzinger complex, respectively. Thus, our findings suggested that activating the neural circuit between the DR and the pre-Bötzinger complex might contribute to preventing.

2. MATERIALS AND METHODS

2.1 Animals

All experimental procedures were in line with the National Institutes of Health Guidelines for the Care and Use of Laboratory Animals and approved by the Animal Advisory Committee of Zhejiang University. DBA/1 mice were housed and bred in the Animal Center of Zhejiang University School of Medicine and provided with rodent food and water ad libitum. In the acoustic stimulation murine model, DBA/1 mice were "primed" starting from postnatal day 26-28 to establish consistent susceptibility to audiogenic seizures and S-IRA. PTZ-evoked seizure model, PTZ was administered to non-primed DBA/1 mice of approximately 8 weeks of age. The two models were established just as we described previously [5-12]. TPH2-ChETA infected mice was established by viral delivery of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS with the control of promoter of TPH2 into the DR to express for 3 weeks and confirmed the expression of TPH2 -ChETA in DR by immunohistochemistry. The promoter of TPH2 by we used in the present study has been used to be significantly infected the 5-HT neurons in the DR from our group to modulate the balance between reward and aversion[13].

2.2 Seizure induction and resuscitation

S-IRA was evoked by acoustic stimulation or intraperitoneal (IP) administration of PTZ, as we previously described [5-12]. For the acoustic stimulation model, each mouse was placed in a cylindrical plexiglass chamber in a sound-isolated room, and audiogenic seizures (AGSZs) were evoked by an electric bell (96 dB SPL, Zhejiang People's Electronics, China). Acoustic

stimulation was given for a maximum duration of 60 s or until the onset of tonic seizures and S-IRA in most mice in each group. Mice with S-IRA were resuscitated within 5 s post the final respiratory gasp using a rodent respirator (YuYan Instruments, ShangHai, China). S-IRA was also evoked in all non-primed DBA/1 mice by IP administration of a single dose of PTZ (Cat # P6500; Sigma-Aldrich, St. Louis, MO) at a dose of 75 mg/kg. Mice with S-IRA were resuscitated by using a small animal respirator (YuYan Company, ShangHai, China).

2.3. Pharmacology experiment

2.3 .1 Effect of IP administration of KET on 5-HTP-mediated suppression of S-IRA evoked by acoustic stimulation

A vehicle or different doses of KET (Cat # 8006; Sigma-Aldrich) was administered to different groups of mice which were then acoustically stimulated. Susceptibility to S-IRA in primed DBA/1 mice was confirmed 24 h prior to treatment of 5-HTP (Cat # 107751; Sigma-Aldrich) or vehicle, which was followed by KET administration, respectively. 5-HTP (200mg/kg) or vehicle (saline) was administered i.p. once a day for 2 days and induction of S-IRA was performed 90 mins after the second administration. KET or vehicle (25% DMSO) was administered i.p. 30 mins before acoustic stimulation. The effects of 5-HTP and KET on S-IRA were examined and digitally recorded for offline analysis, respectively. Additionally, KET was administered by ICV injection 15 mins prior to PTZ injection as the selective antagonist of 5-HT2AR in DBA/1 mice in different groups from the same manner of 5-HTP administration. The incidences of S-IRA latency to AGSZs, duration of wild running, clonic seizures, tonic-clonic seizures, and seizure scores were videotaped for offline analysis [5-10], [17-18].

2.3.2 Effect of ICV administration of KET on 5-HTP-mediated suppression of S-IRA by PTZ

Aiming at the ICV surgery, an ICV guide cannula(O.D.I.41×I.D.O.0.25mm/M3.5,62004, RWD Life Science Inc.,China)was implanted into the right lateral ventricle (AP – 0.45 mm; ML – 1.0 mm; V-2.50 mm) to enable microinjections as we previously described [9,12]. 5-HTP (200mg/kg) or vehicle (saline) was administered i.p. once a day for 2 days and induction of S-IRA was performed 90 mins after the second administration. The groups to test as follows: 1) Saline (IP) was administered 75 mins prior to PTZ (75mg/kg, IP) and 25% DMSO at (2 μl,

at a rate of 0.5 μl/min ICV) 15 min prior to PTZ injection as controls. 2) 5-HTP (200mg/kg, IP) was administered 75 mins prior to PTZ (75mg/kg, IP) and 25% DMSO (2 μl, at a rate of 0.5 μl/min ICV) 15 min prior to PTZ injection. 3) 5-HTP (200mg/kg, IP) was administered 75 mins prior to PTZ (75mg/kg, IP), and KET (4.764 nmol and 9.528 nmol, dissolved in 2 μl 25% DMSO, at a rate of 0.5 μl/min ICV) 15 min prior to PTZ injection. The timing of 5-HTP administration in the above groups was in the manner of administering i.p. once a day for 2 days.

2.4 Optogenetics experiments

2.4.1 Stereotactic surgery

DBA/1 mice at 8 weeks of age were anesthetized with 3.5% chloral hydrate and head-fixed in a stereotaxic apparatus, (68018, RWD Life Science Inc., Shenzhen, China), as previously described [7]. During the entire process of surgery, the body temperature of anesthetized mice was kept constant at 37 °C using a heating pad. If the DBA/1 mice had a pain reflect in response with a paw pinch, then additional 10% of initial dosage of sodium pentobarbital was given to guarantee a painless state. Other procedures were basically the same with our previous methods concerning the optogenetics experiments. For optogenetic viral delivery of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS, microinjection (100nl,40nl/min) was performed using the following stereotaxic coordinates for the DR (AP−4.55mm, ML−0.44mm, DV−2.80mm,10°right) based on the mouse brain atlas. Viruses were delivered via a gauge needle by an Ultra Micro Pump (160494 F10E, WPI) over a period of 10 mins; the syringe was not removed until 15–20 min after the end of infusion to allow diffusion of the viruses. Then, the optical fiber (FOC-W-1.25-200-0.37-3.0, Inper, Hangzhou, China) was implanted above the area (AP−4.55mm, ML−0.44mm, DV−2.80mm,10°right) for 0.05 mm (AP−4.55mm, ML−0.44 m, DV−2.75 mm,10°right). For ICV surgery and an ICV guide cannula implantation was the same to the pharmacology experiment, and was implanted with a headstage for EEG in the same mice as previously described. For the microinjection of KET in the bilateral pre-Bötzinger complex, two sides of guide cannula (O.D.0.48×I.D.0.34mm/M3.5,62033,RWD Life Sience Inc.,Shenzhen, China) implantation were performed. CTB-555 100nl (1μg/μL, BrainVTA Technology Co.Ltd, Wuhan, China) was injected in the DR (AP−4.55mm, ML−0.44mm, DV−2.80mm,10°right)or the right side of the

pre-Bötzinger complex (AP−6.80mm, ML−1.25mm, DV−4.95mm) at a total content of ～100nL and we waited ～2 weeks to allow the retrograde labeling of projection neurons, respectively. For the photostimulating the bilateral pre-Bötzinger complex, viral delivery of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS was delivered in the bilateral pre-Bötzinger complex (AP−6.80mm, ML−1.25mm, DV−4.95mm) based on the mouse brain atlas to 3 weeks and the optical fiber with guide cannula (O.D.0.48×I.D.0.34mm/M3.5,62033,RWD Life (FOC-W-1.25-200-0.37-3.0, Inper, Hangzhou, China) over 0.05 mm was implanted, respectively. For the photometry recordings, viral delivery of AAV2/9-mCaMKIIa-GCaMP6f-WPRE-pA, microinjection (100nl,40nl/min) was performed using the following stereotaxic coordinates for the bilateral pre-Bötzinger complex (AP−6.80mm, ML−1.25mm, DV−4.95mm) based on the mouse brain atlas to 3 weeks and the optical fiber (FOC-W-1.25-200-0.37-3.0, Inper, Hangzhou, China) over 0.05 mm was implanted, respectively.

2.4.2 **The effects of ICV administration of KET on photostimulating of the DR-mediated suppression of S-IRA by PTZ and on EEG changes**

DBA/1 mice undergone by viral delivery of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS into the DR and implanted an ICV guide cannula with a headstage for EEG for 3 weeks will be used. Those DBA/1 mice were divided into 3 groups. For the control group without photostimulation of DR (n=7), ICV of the same concentration and volume of vehicle was given at the manner (25% DMSO, 2 μl, at a rate of 0.5 μl/min) prior to IP injection of PTZ (75mg/kg) for 30 mins. For the group treatment by photostimulation of DR without ICV of KET (n=6), ICV of the same concentration and volume of vehicle was given prior to the duration of photostimulation for 15mins and to IP injection of PTZ (75mg/kg) for 30 mins. For another group treatment by photostimulation of DR with ICV of KET(n=7), ICV of KET with the total content of 18.3 nmol was given prior to the duration of photostimulation for 15mins and to IP injection of PTZ (75mg/kg) for 30 mins. The statistical analyses of incidence of S-IRA was performed, respectively. EEG recordings of 3 groups started before PTZ injection for 5 mins and ended post PTZ for 30mins. The statistical analyses of EEG activities were performed as previously described. The parameter of photostimulating of the

DR (blue-light, 465 nm, 20Hz, 20ms Pulse Width, 15 mW, 20min) was used in the above mice.

2.4.3 The effects of microinjection of KET in the bilateral pre-Bötzinger complex administration of KET on photostimulatiing the DR-mediated suppression of S-IRA by PTZ

DBA/1 mice undergone by viral delivery of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS into the DR and with the bilateral pre-Bötzinger complex guide cannula implantation for 3 weeks will be used. For the group treated by photostimulation of DR without administration of KET (n=7), microinjection of the same concentration and volume of vehicle into every unilateralbilateral pre-Bötzinger complex was given in the manner (25% DMSO, 2 μl, at a rate of 0.5 μl/min for 200nl) prior to IP injection of PTZ (75mg/kg) for 25 mins, respectively. For another group treated by photostimulation of DR with administration of KET (n=7), microinjection of KET with 200 nl (containing of 0.4ug of KET) into every unilateralbilateral pre-Bötzinger complex prior to IP injection of PTZ (75mg/kg) for 25 mins, respectively. Both groups would be started laser stimulation after microinjection for 10 mins. Total content of KET 400 nl (containing of 0.8ug of KET) was given for one mouse of bilateral pre-Bötzinger complex. The parameter of photostimulating of the DR (blue-light, 465 nm, 20Hz, 20ms Pulse Width, 15 mW, 20min) was used in the above mice. For the laser stimulation, 465-nm blue-light (20Hz, 20ms Pulse Width, 15 mW, 20min) was delivered by the laser (B12124, Inper, Hangzhou, China) through a 200-μm optic fiber.

2.4.4 The effects of photo-stimulating the TPH2-ChETA neurons of **the bilateral pre-Bötzinger complex and microinjection of KET in the bilateral pre-Bötzinger complex on the incidence of of S-IRA by PTZ**

DBA/1 mice undergone by viral delivery of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS into the bilateral pre-Bötzinger complex for 3 weeks will be used. For the control group treated by without photostimulation, microinjection of the same concentration and volume of vehicle into every unilateralbilateral pre-Bötzinger complex was given in the manner (25% DMSO, 2 μl, at a rate of 0.5 μl/min for 200nl) prior to IP injection of PTZ (75mg/kg) for 25 mins (n=6), respectively. For the treated group by photostimulating the bilateral pre-Bötzinger complex, microinjection of the same concentration and volume of vehicle into every

unilateralbilateral pre-Bötzinger complex was given in the manner (25% DMSO, 2 μl, at a rate of 0.5 μl/min for 200nl) prior to IP injection of PTZ (75mg/kg) for 25 mins (n=7), respectively. For the KET-treated group photostimulating the bilateral pre-Bötzinger complex, microinjection of KET with 200 nl (containing of 0.4ug of KET) into every unilateralbilateral pre-Bötzinger complex prior to IP injection of PTZ (75mg/kg) for 25 mins, respectively (n=3). The parameter of photostimulating of the bilateral pre-Bötzinger complex (blue-light, 465 nm, 20Hz, 20ms Pulse Width, 15 mW, 20min) was used in the above mice. For the laser stimulation, 465-nm blue-light (20Hz, 20ms Pulse Width, 15 mW, 20min) was delivered by the laser (B12124, Inper, Hangzhou, China) through a 200-μm optic fiber.

2.4.5 The effects of photo-stimulating the TPH2-ChETA neurons of **the DR on the $Ca2^{+}$ activity of neurons in the bilateral pre-Bötzinger complex in the model of S-IRA by PTZ by photometry recordings**

DBA/1 mice undergone by viral delivery of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS or AAV2/9-mCaMKIIa-GCaMP6f-WPRE-pA (at a rate of 40 nl/min for 100nl) into the DR and into the bilateral pre-Bötzinger complex for 3 weeks will be used, respectively. For the control group treatment with the DR of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS without photostimulation and the bilateral pre-Bötzinger complex of AAV2/9-mCaMKIIa-GCaMP6f-WPRE-pA, the photometry recordings started 5 mins prior to PTZ (75mg/kg, i.p.) and ended after PTZ injection 60mns or until the duration of death of mice within 60 mins (n=3). For another group treatment with the DR of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS withphotostimulation and the bilateral pre-Bötzinger complex of AAV2/9-mCaMKIIa-GCaMP6f-WPRE-pA, the photometry recordings started 5 mins prior to IP injection of PTZ (75mg/kg) and ended after PTZ injection 60mns or until the duration of death of mice within 60 mins (n=3). The fiber photometry system (Inper, Hangzhou, China, C11946) used a 488 nm diode laser . We segmented the data based on individual trials of seizure duration and derived the values of fluorescence change (ΔF/F) by calculating (F − F0)/F0.

2.4.4 Immunohistochemistry and histology

The placement of the optical fiber cannula tip including ICV and microinjection of KET with bilateral pre-Bötzinger complex in each implanted mouse was verified by histology. After the

novelty-seeking test approximately 30mins, DBA/1 mice were sacrificed and perfused with PBS and 4% PFA. After saturated in 30% sucrose (24h), each brain was sectioned into 30μm thickness of coronal slices with a freezing microtome (CM30503, Leica Biosystems, Buffalo Grove, IL, USA), the sections were first washed in PBS 5mins for 3 times and then incubated in blocking solution containing 10% normal donkey serum (017-000-121, Jackson ImmunoResearch, West Grove, PA), 1% bovine serum albumen (A2153, Sigma-Aldrich, St. Louis, MO), 0.3% Triton X-100 in PBS for 1 h at room temperature. Then, for c-fos or TPH2 staining in DR, sections were incubated at 4°C overnight in rabbit anti-c-fos(1:1000 dilution, 2250T Rabbit mAb /74620 Mouse mAb, Cell Signaling Technolog, Danvers, Massachusetts, USA) primary antibody or mouse anti-TPH2 (1:100 dilution, T0678, Sigma-Aldrich, St. Louis, MO) primary antibody and donkey anti-mouse Alexa 488 secondary antibody (1:1000; A32766, Thermo Fisher Scientific, Waltham, MA, USA), donkey anti-mouse Alexa 546 (1:1000; A10036, Thermo Fisher Scientific, Waltham, MA, USA) secondary antibody or goat anti-rabbit cy5 (1:1000; A10523, Thermo Fisher Scientific, Waltham, MA, USA) secondary antibody for 2h at room temperature. Similarly, for neurokinin 1 receptor (NK1R) or SA-2A R (5-HT-2AR) staining in PBC, sections were incubated in rabbit anti-NK1(1:1000 dilution, SAB4502913, Sigma-Aldrich, St. Louis, MO) primary antibody or mouse anti-SA-2A R (5-HT-2A R) (1:100 dilution, sc-166775, Santa Cruz Biotechnology, Dallas, USA) primary antibody at 4°C overnight and donkey anti-rabbit Alexa 488 secondary antibody (1:1000; A32766, Thermo Fisher Scientific, Waltham, MA, USA), donkey anti-mouse Alexa 546 (1:1000; A10036, Thermo Fisher Scientific, Waltham, MA, USA) secondary antibody or goat anti-mouse cy5 (1:400; A10524, Thermo Fisher Scientific, Waltham, MA, USA) secondary antibody for 2h at room temperature. After washing with PBS 15mins for 3 times, the sections were mounted onto glass slides and incubated in DAPI (1:4000;Cat# C1002; Beyotime Biotechnology; Shanghai, China) 7mins at room temperature.Finally, the glass slides were sealed sheet by Anti-fluorescence attenuating tablet.All images were taken with Nikon A1 laser-scanning confocal microscope (Nikon, Tokyo, Japan).The numbers of immunopositive cells were counted and analyzed using ImageJ (NIH, Baltimore, MD). It is worth noting that the mouse of the implantation placement out of the target of brain structure will be abandoned in our experiments. The positive cells co-expression c-fos, ChETA and TPH2 were counted as

previously described[7].

2.4.5 Viral vectors

pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS (AVV2/8) (viral titers:$1x10^{13}$vg/ml), were purchased from Sheng BO, Co., Ltd (Sheng BO Co. Ltd, Shanghai, China). and the sequence of vectors was designed source by Kazuki Nagayasu (Department of Molecular Pharmacology Graduate School of Pharmaceutical Sciences, Kyoto University). CTB-555(1μg/μL) was purchased from BrainVTA Technology Co.Ltd (Wuhan, China). AAV2/9-mCaMKIIa-GCaMP6f-WPRE-pA (viral titers:$1x10^{14}$vg/ml) were purchased from Taitool Bioscience Co. Ltd (Shanghai Taitool Bioscience Co. Ltd, Shanghai, China)

2.5 Data analysis

All data are presented as the mean ± SEM. Statistical analyses were performed using SPSS 23 (SPSS Software Inc.,USA). The incidence of S-IRA in different groups was compared using Wilcoxon Signed Rank test. Seizure scores and the latency to AGSZs, the duration of wild running, clonic seizures, tonic-clonic seizures were evaluated using the one-way ANOVA tests or Mann Whitney U test or Kruskal-Wallis H test. ANCOVA was used to compare c-fos-positive cells in the DR of DBA/1 mice with and without photostimulation. Statistical significance was inferred if $p < 0.05$.

3. Results

3.1 5-HTP-mediated suppression of S-IRA evoked by acoustic stimulation was reversed by IP with KET

Compared with vehicle group in primed DBA/1 mice, the incidence of S-IRA evoked by acoustic stimulation was significantly reduced by 5-HTP with the dosage of 200 mg/kg, i.p ($p < 0.001$). Compared with vehicle group, the incidence of S-IRA in the group pre-treated with 5-HTP (200mg/kg, i.p) + KET (5,10, 25 mg/kg,i.p) was significantly decreased ($p < 0.01$, $p < 0.05$), instead, which indicated the these dosages of KET didn't significantly reversed the lower incidence of S-IRA by 5-HTP. By contrast, there was no difference between the vehicle group and the group with treatment with 5-HTP (200mg/kg, i.p) + KET (20 mg/kg,i.p) ($p > 0.05$) and the incidence of S-IRA was significantly reduced in the group (5-HTP +25% DMSO) compared with the group (KET, 20 mg/kg,i.p) ($p < 0.05$), on the contrary, the suppressant effects by 5-HTP was markedly reversed administration with KET (20 mg/kg, i.p).

(Figure 1)

3.2 5-HTP-mediated suppression of S-IRA evoked by PTZ was reversed by ICV with KET

Compared with vehicle control group, the incidence of S-IRA by PTZ was significantly reduced in the group treatment with 5-HTP and 25% DMSO (i.c.v) ($p < 0.05$). Compared with vehicle control group, the incidence of S-IRA by PTZ was not significantly reduced in the group treatment with 5-HTP and the KET (9.15 nmol, i.c.v) ($p > 0.05$), and compared with the group treated with 5-HTP and 25% DMSO (i.c.v), the incidence of -S-IRA in the group treated with 5-HTP and the KET (9.15 nmol, i.c.v) was significantly increased ($p < 0.05$), which suggested that the suppressant effects of S-IRA by 5-HTP was significantly reversed by KET with the dosage of 9.15 nmol (i.c.v). Furthermore, compared with vehicle control group, the incidence of S-IRA in the group with 5-HTP + KET (18.30 nmol, i.c.v) was not significantly reduced ($p > 0.05$) and the incidence of S-IRA by PTZ in the group treatment with 5-HTP and 25% DMSO (i.c.v) was significantly reduced compared with the group with 5-HTP and KET (18.30 nmol,i.c.v) ($p < 0.05$), instead, the suppressant effects of S-IRA by 5-HTP can be significantly reversed by the dosage of KET (18.30 nmol, i.c.v). There were no significant intergroup differences in latencies to AGSZs, durations of wild running, clonic seizures, tonic-clonic seizures and seizure scores (AGSZs latencies: $p = 0.763$, $F= 6$; durations of wild running: $p = 0.14$, $F= 6$; duration of tonic-clonic seizures: $p = 0.119$, $F= 6$; seizure scores: $p = 0.304$, $F= 6$) (Fig. 1). The seizure score of two models from different groups was no significantly difference ($p > 0.05$), no obvious intervention in seizure behavior of the revered effects by KET was to be fund. (Figure 2)

3.3 Selective expression of ChETA on 5-HT neurons in the DR of DBA/1 mice

The viral of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS, under the control of promoter of TPH2, was delivered into the DR of wild DBA/1 mice with 50 days to express for 3 weeks to create theTPH2-ChETA mice. We first examined the expression of ChETA and TPH2 in 5-HT neurons in the DR of DBA/1 mice using immunohistochemistry (n = 3 mice). The expression of GFP, a surrogate marker for ChETA, was predominantly localized on the membrane of cell body and axons of 5-HT neurons of the DR. TPH2 was predominantly confined to the cytosol of 5-HT neurons in the DR of DBA/1 mice. The merged expression ratio of ChETA and TPH2

was basically consistent with our previous data that the co-expression of ChR2 and TPH2 was located on the walls of 5-HT neurons in the DR of DBA/1 mice[7]. (Figure 3, Figure 4)

3.4 Activation of 5-HT neurons in the DR reduces S-IRA evoked by PTZ injection in DBA/1 mice and the lower incidence of S-IRA by photostimulating the DR was significantly reversed by ICV of KET

We examined the effect of selective enhancement of 5-HT neurotransmission on S-IRA by PTZ by applying photostimulation (blue light, 20 ms pulse duration, 20 Hz, 20 mins) to 5-HT neurons in the DR of noprimed DBA/1 mice. Comapred with the incidence of S-IRA by PTZ of DBA/1 mice without photostimulation (n=7), photostimulation of the DR for 20 mins significantly decreased the incidence of S-IRA in DBA/1 mice (n=6, $p < 0.05$). However, the incidence of S-IRA evoked by PTZ by photostimulation of the DR with the same parameter by ICV administration of KET was significantly (n=7, $p < 0.05$), which indicated the lower incidence of S-IRA by PTZ via photostimulating the DR was significantly reversed by blocked 5HT2A receptor in the brain. (Figure 5)

3.5 Activating 5-HT neurons in the DR to reduce S-IRA evoked by PTZ and produced obvious effects on EEG activity at the different stages

Based on the above findings in the same groups, we further examined the effect of activating 5-HT neurons in the DR to reduce incidence of S-IRA and the reversal effect by KET on EEG activity. Compared with the group without light, the EEG activity was significantly reduced. Analyzing the EEG wave, Delta wave of EEG was significantly reduced by light and reversed by KET ($p < 0.05$). There is no obvious changes of Theta, Alpha, Beta and Gamma waves in different treatment groups. The findings of EEG activity may reflect the specificity of 5-HTA receptor in brain modulating S-IRA and SUDEP. (Figure 6)

3.6 The reduction of S-IRA by photostimulating the DR is dependent on 5-HT2A receptor locating the pre-Bötzinger complex

Although the incidence of S-IRA can be reduced by 5-HTP and by photostimulating DR and was significantly reversed by both P and ICV injection of KET, it still was to determine whether 5-HT2A receptor in the pre-Bötzinger complex mediated the process of S-IRA and SUDEP. The incidence of S-IRA evoked by PTZ by photostimulating the DR was 14.28%. However, the incidence of S-IRA evoked by PTZ by photostimulating the DR with

microinjection of KET into bilateral pre-Bötzinger complex with total volume of 400 nl was 85.71%. Compared with the group by photostimulating DR without microinjection of KET, the incidence of S-IRA in the group by photostimulating DR with microinjection of KET was significantly reduced ($p < 0.01$). Subsequently, the existing of neural circuit between DR and the pre-Bötzinger complex was verified by using a nerve tracer named as CTB-555. Therefore, in the present study, optogenetic activation of the neural circuit between DR and the pre-Bötzinger complex contributes to inhibit the S-IRA and 5-HT2A receptor in the pre-Bötzinger complex may be a specific target to intervene for preventing SUDEP. (Figure 7-9)

3.7 Photostimulation increases c-fos expression in the 5-HT neurons in the DR

To investigate whether photostimulation increased the excitability of 5-HTneurons in DR, we examined the neuronal expression of c-fos, an immediate early gene that is widely accepted as a marker for neuronal activity in optogenetics studies in the DR of DBA/1 mice with photostimulation and those without photostimulation. Compared with the c-fos expression in 5-HT neurons in the DR of DBA/1 mice without photostimulation (n=2), the expression of c-fos in 5-HT neurons in the DR was significantly increased in the group with photostimulation (20 ms pulse duration, 20 Hz) at 15 mW for 20 min (n=2, $p < 0.05$), which showed the reduction of S-IRA by photostimulation via activating 5-HT neurons. (Figure 10)

3.8 Photostimulation of the bilateral pre-Bötzinger complex did not significantly reduce the incidence of S-IRA by PTZ

We investigated whether the incidence of S-IRA by PTZ by photostimulation of bilateral pre-Bötzinger complex can be independently reduced by photostimulation of TPH2-ChETA neurons in the bilateral pre-Bötzinger complex. Although the incidence of S-IRA by PTZ can be reduced by photostimulation of bilateral pre-Bötzinger complex, there was not significant difference between the control group treated by PTZ without photostimulation and the group treated by PTZ with photostimulation (n=6, n=7, $p > 0.05$). There was not significant difference between the control group treated by PTZ with photostimulation with micro-injection of vehicle and the group treated by PTZ with photostimulation with micro-injection of KET of 0.4ug (n=7,n=3, $p > 0.05$). The above finding showed that activation of the neural circuit between DRand the pre-Bötzinger complex can significantly

reduced the incidence of S-IRA rather than the pre-Bötzinger complex independently regulated S-IRA and SUDEP in our models.(Figure 11)

3.9 The neural activity of calcium signal evoked by PTZ in the pre-Bötzinger complex during seizures was significantly reduced by photostimulation of the DR by photometry recordings

To investigate of neural activity of calcium signal evoked by PTZ, the calcium strong signal from the bilateral pre-Bötzinger complex was recorded by photometry recordings in the mice infected with GCaMP6f in the the bilateral pre-Bötzinger complex without photostimulation of the DR during the clonic and tonic seizure phase evoked by PTZ respectively. The group without photostimulation of the DR, the activity wave of calcium signals from the bilateral pre-Bötzinger complex were strong during the phase of clonic and tonic seizures, respectively.However, the activity wave of calcium signals from the bilateral pre-Bötzinger complex were weaker in the group with photostimulation of the DR during the phase of clonic and tonic seizures, respectively. The data showed photostimulation of the DR can reduce the abnormal activity of calcium signals from the bilateral pre-Bötzinger complex to make the normal respiration rhythm to prevent S-IRA and SUDEP. (Figure 12)

4. DISCUSSION

SUDEP is a fatal complication for epilepsy patients. Although initial advancements about 5-HT nerve system have been made to identify the causes of SUDEP, the pathogenesis of SUDEP still seems elusive [1-4]. Currently, respiratory dysfunction during seizures has been regarded as a leading mechanism of SUDEP. Some studies indicated that several selective serotonin (5-HT) reuptake inhibitors (SSRIs) can prevent S-IRA in DBA/1 mice evoked by generalized audiogenic seizures (AGSz) by elevating 5-HT levels in the synaptic cleft [17-19]. However, due to the limitation of animal SUDEP models, the role of 5-HT synthesis and targeting 5-HT receptors in brain in modulating S-IRA and SUDEP remains unclear and still needs to be further explored.

It had been accepted that tryptophan hydroxylase-2 (TPH2) in brain could convert l-tryptophan to 5-HTP, which can be further converted to 5-HT by aromatic L-amino acid decarboxylase [22-24]. A previous study showed that TPH2 is the rate-limiting enzyme in 5-HT

synthesis in brain [22]. Although we only tested the protein changes of TPH2 in our previous study, another study from our group had tested the correlation between TPH2 protein and activity which showed that TPH2 protein varied consistently with TPH2 activity in the same model [11]. Thus, it is vital for TPH2 to make endogenous 5-HTP convert into serotonin synthesis to reduce the incidence of S-IRA and prevent SUDEP. However, it is unknown how 5-HT mediated S-IRA and SUDEP in our model.

Based on our previous study that administration of 5-HTP significantly reduced the incidence of -S-IRA in AGSz and PTZ SUDEP models, it is essential for us to further identify in which 5-HT2AR in the brain plays a key role in modulating the pathogenesis of S-IRA in our models. In our models, the lower incidence of S-IRA by 5-HTP was significantly reversed by IP or ICV injection of KET in the SUDEP models evoked by acoustic stimulation and PTZ administration, a chemoconvulsant that is widely used to model human generalized seizures. While KET reversed the suppressant effects of S-IRA by administration of 5-HTP in a dose-dependent manner in PTZ injection model, there was a ceiling effect of KET on reversing the lower incidence of S-IRA by 5-HTP in the acoustic stimulation SUDEP model.

Our previous study showed that administration of 5-HTP significantly reduced the incidence of S-IRA by anti-convulsant effects [5,7]. This indicated that most of DBA/1 mice avoided S-IRA without producing tonic-clonic seizures other than wild running and generalized clonic seizures, which remained the sensitivity for seizures. Other finding from other groups showed that enhance of 5-HT contents in the brain suppressed seizures by producing the anti-convulsant effects as well [23-24]. However, administration of KET increased the lower incidence of S-IRA by 5-HTP in both acoustic stimulation and PTZ injection models with producing partly wild running, clonic and/or tonic -clonic seizures, which demonstrated administration of IP of KET reversed the lower incidence of S-IRA by 5-HTP with only partly affecting seizure behaviors by acting on the neuron nucleus functioning in respiratory activity.

To further determine the effects of KET on the incidence of S-IRA by acting on the central nucleus in the brain, we administrated KET with ICV pathway and measured its effect on S-IRA in DBA/1 mice in both SUDEP models, respectively. The results of KET mediated the reversal effects of the S-IRA incidence by 5-HTP via IP injection in line with via ICV

injection, suggesting that the reversal effects by KET in our models are independent of SUDEP models.

Meanwhile, S-IRA in most of DBA/1 mice recovered at 24-72 hours between different treatment groups, which showed the recovery interval of S-IRA depended on the concentration of 5-HT in brain. In addition, we find that there is a dose-dependent effect on reversing the incidence of S-IRA by 5-HTP and a ceiling effect with KET 25mg/kg (i.p). Indeed, our previous study showed that the lower incidence of S-IRA by activating 5-HT neurons in the DR by optogenetics was significantly reversed by ondansetron, a specific antagonist for 5-HT3 receptor [7]. However, it can't rule out that 5-HT2A receptor mediated the pathogenesis of S-IRA and SUDEP by closely interacting with 5-HT3 receptor in the brain. Of course, it needs to be further tested in the future experiments.

Although an SSRI is also found effective in reducing S-IRA evoked by maximal electroshock (MES) in Lmx1b(f/f) mice on a primarily C57BL/6J background, a strain that is resistant to AGSz and depleting 5-HT neurons enhanced the seizure severity, which led to the S-IRA and can be prevented by 5-HT2AR activation through IP with (2,5-dimethoxy-4-iodophenylpropane hydrochloride, DOI), a selective agonist for 5-HT2AR, in MES model [18], it was unclear whether depleting 5-HT neurons itself led to S-IRA and the reversal effects by DOI targeted the peripheral or central 5-HT2AR. By contrast, based on our previous findings, we further explored the role of 5-HT neurotransmission and 5-HT2AR by peripheral and central intervention approaches with KET, a selective antagonist for 5-HT2AR, in different SUDEP models. Thus, according to our findings, the role of 5-HT2AR in modulating S-IRA supported by those from the MES model will further strengthen the understanding of the pathogenesis of S-IRA and SUDEP in the 5-HT nerve system, which will help design therapeutic strategies to prevent SUDEP.

What's more, based on 5-HT2AR located in the pre-Bötzinger complex which plays an important role in modulating respiration rhythm and on our previous finding that the incidence of S-IRA can be significantly reduced by activation of TPH2-ChR2 neurons in the DR, to further test the above pharmacology experiment concerning the exact mechanisms of the interaction between 5-HT and 5-HT2AR mediating S-IRA and SUDEP in the same models, we used the optogenetics methods to test whether activation of TPH2-ChR2 neurons

in the DR significantly reduced the incidence of S-IRA evoked by PTZ via activating 5-HT2AR in the pre-Bötzinger complex or not. In the present study, the lower incidence of S-IRA evoked by PTZ via optogenetic activation of TPH2-ChETA neurons in the DR was remarkably reversed by ICV injection of KET. Subsequently, the lower incidence of S-IRA evoked by PTZ by photostimulating the DR was significantly reversed by microinjection of KET into bilateral pre-Bötzinger complex, in turn, suggesting that photostimulating the DR remarkably reduced the incidence of S-IRA by activating 5-HT2AR in the pre-Bötzinger complex in our model. To further test the bridge between DR and the pre-Bötzinger complex to modulate S-IRA in our study, CTB-555, a nerve tracer, was used in our study to confirm and establish the neural circuit between the DR and the pre-Bötzinger complex. Thus, the lower incidence of S-IRA in PTZ evoked models by photostimulating DR stem from the neural circuit between the DR and the pre-Bötzinger complex. However, we only tested it using photostimulating the DR and direct inhibiting 5-HT2AR in the bilateral pre-Bötzinger complex in our study. Whether activating TPH2-neurons in the pre-Bötzinger complex could reduce the incidence of S-IRA still needs to be verified in the following experiments. Of course, other neural circuits between dorsal raphe nucleus and other brain structures involved in modulating S-IRA and SUDEP can't be excluded in our models. Nevertheless, pharmacologic and optogenetic findings suggested that the neural circuit between the DR and the pre-Bötzinger complex plays a key role in modulating S-IRA and SUDEP.

In addition, different from the pharmacology experiments, although S-IRA of DBA/1 mice was blocked in optogenetic experiments, most of DBA/1 mice remained clonic and tonic seizures. Analyzing of the cause for difference, the specificity of optogenetics and the specific injection of KET by both of ICV and pre-Bötzinger complex may be a cause. Meanwhile, the incidence of S-IRA evoked by PTZ was significantly reduced by activating the neural circuit between DR and the pre-Bötzinger complex without changing EEG activities, which contributed to reflecting the specificity of optogenetic experiments to inhibit S-IRA from another level as well.

What's more, our data further showed that it is insufficient for activating the TPH2-ChETA neurons in the PBC to significantly reduce the incidence of S-IRA by PTZ. Furthermore, the neural activity of calcium signal evoked by PTZ in the pre-Bötzinger

complex during seizures was significantly reduced by photostimulation of the DR by photometry recordings as well. Thus, activation of the neural circuit between DR and the pre-Bötzinger complex can specifically prevent S-IRA and SUDEP.

Conclusion

Our current data suggested that the lower incidence of S-IRA by administration of 5-HTP can be significantly reversed by KET. Furthermore, the suppressant effects of optogenetic activating the neural circuit between DR and the pre-Bötzinger complex on S-IRA were obviously reversed by KET in the DBA/1 mice as well. Therefore, 5-HT2AR in the pre-Bötzinger complex may be a specific and key target to intervene for preventing SUDEP

5. Funding

The work was supported by the National Natural Science Foundation of China (Grant.NO: 81771403 and 81974205); by the Natural Science Foundation of Zhejiang Province (LZ20H090001); by the Program of New Century 131 outstanding young talent plan top-level of Hang Zhou to HHZ, and by the National Natural Science Foundation of China (Grant.NO: 81,771,138) to Han Lu; by the National Natural Science Foundation of China (Grant.NO: 82001379) to HTZ, and by National Natural Science Foundation of China (No. 81501130, Dr. Chang Zeng), Natural Science Foundation of Hunan Province, China (No. 2021JJ31047, Dr. Chang Zeng).

6. ACKNOWLEDGEMENTS

We are grateful to Yi Shen and Yu Dong Zhou for developing the experimental design

7. DISCLOSURE

All authors declare no competing interests. We confirm that we have read the Journal's position on issues involved in ethical publication and affirm that this study is in accordance with those guidelines.

8. Figure legends

1

Figure1

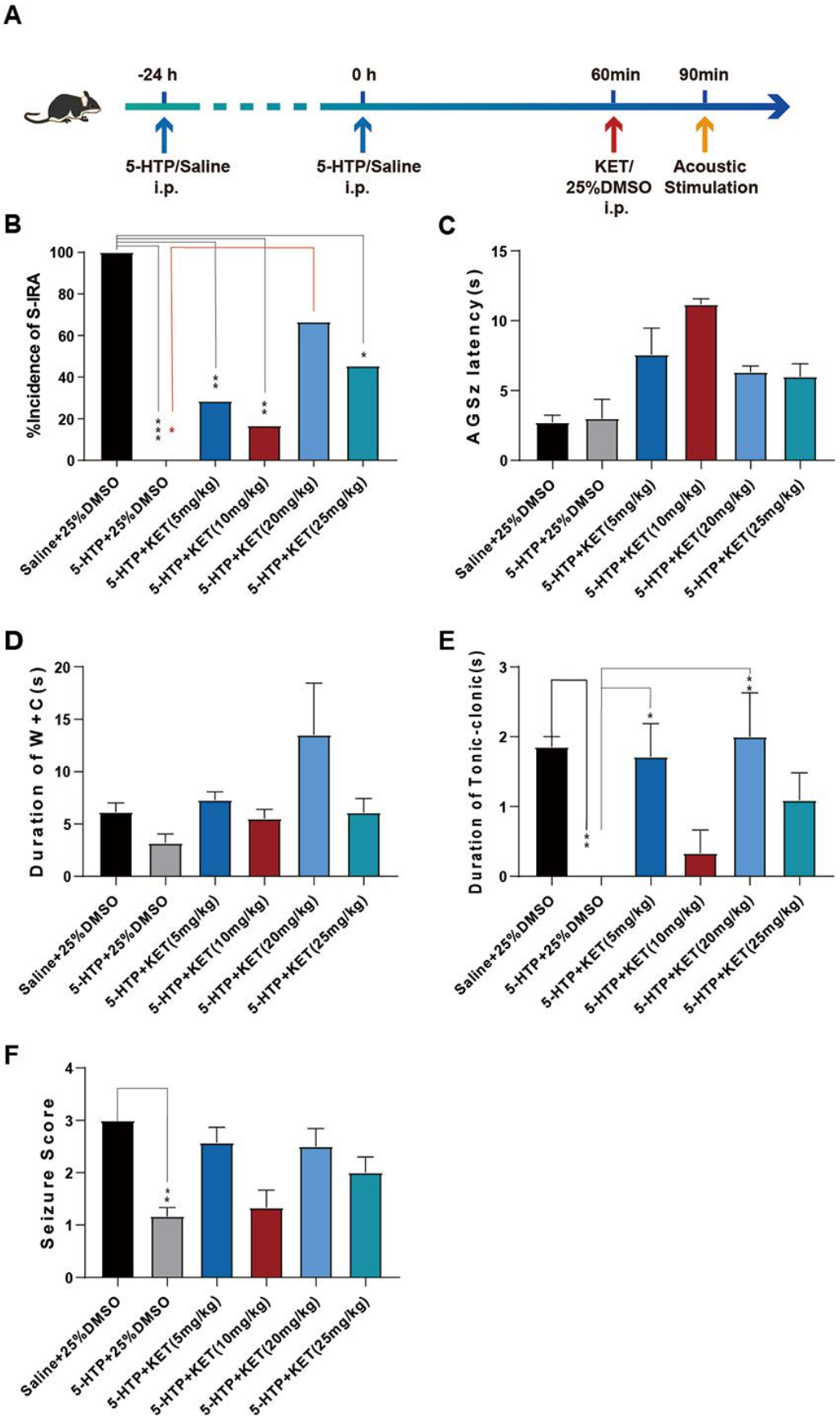
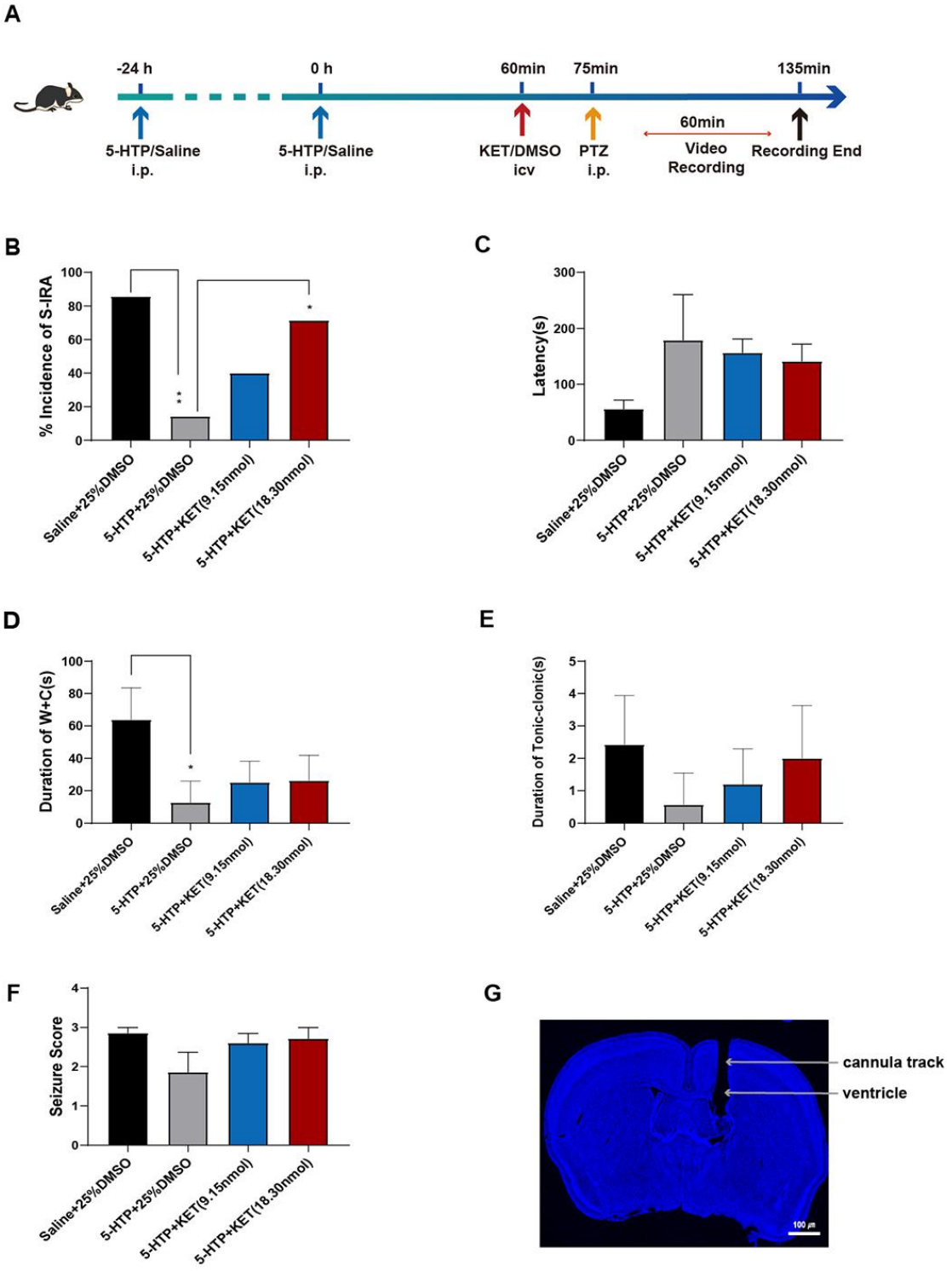

A
-24 h
0 h
60min
90min
5-HTP/Saline i.p.
5-HTP/Saline i.p.
KET/ 25%DMSO i.p.
Acoustic Stimulation
B
%Incidence of S-IRA
C
AGSz latency(s)
D
Duration of W+C(s)
E
Duration of Tonic-clonic(s)
F
Seizure Score
Saline+25%DMSO
5-HTP+25%DMSO
5-HTP+KET(5mg/kg)
5-HTP+KET(10mg/kg)
5-HTP+KET(20mg/kg)
5-HTP+KET(25mg/kg)

Figure 1 5-HTP-mediated reduction of S-IRA evoked by acoustic stimulation was significantly reversed by IP injection of KET.

B. Compared with the group treatment of saline and 25%DMSO, the incidence of S-IRA evoked by acoustic stimulation was markedly lower in groups treated with 5-HTP and 25% DMSO (n=7, n=6, * $p = 0.031$, $p < 0.05$). Compared to the control group treated with vehicle, the incidence of S-IRA was significantly reduced in the group treated with 5-HTP and KET (5-10 mg/kg) (n=7, n=6, $p < 0.01$). However, there was no difference between the control group and the group treated with 5-HTP and KET (20 mg/kg) (n=7, n=6, $p > 0.05$). Furthermore, compared with the group treated with 5-HTP and 25% DMSO, the incidence of S-IRA in the group treated with 5-HTP and KET (20 mg/kg) was significantly increased (n=7, n=6, *$p < 0.05$). However, compared with the group treated with 5-HTP and 25% DMSO, the incidence of S-IRA in the group treated with 5-HTP and KET (25 mg/kg) was not significantly increased (n=7, n=11, $p > 0.05$). C-F, There were no intergroup differences in latencies to AGSZs, durations of wild running plus clonic seizures (W+C), tonic-clonic seizures, and seizure scores ($p > 0.05$).

Figure2



C. Figure 2 5-HTP-mediated reduction of S-IRA evoked by PTZ was significantly reversed by ICV injection of KET

D. Compared to the control group, S-IRA evoked by PTZ was markedly lower in groups

treated with 5-HTP and DMSO. However, there is no difference between the control group and the groups treated with 5-HTP and KET (9.15 and 18.30 nmol). By contrast, the incidence of S-IRA was significantly reduced in the group treated with 5-HTP and i.c.v DMSO as compared with both groups treated with 5-HTP and KET (9.15 and 18.30 nmol). B. C-F. There were no intergroup differences in latencies to AGSZs, durations of wild running plus clonic seizures (W+C), tonic-clonic seizures, and seizure scores ($p > 0.05$). G, Representative images of implantation in a DBA/1 mice for ICV. S-IRA, seizure-induced respiratory arrest; AGSZs, audiogenic seizures; i.p., intraperitoneal injection; DMSO, dimethyl sulfoxide. (mean ± SEM).

Figure 3

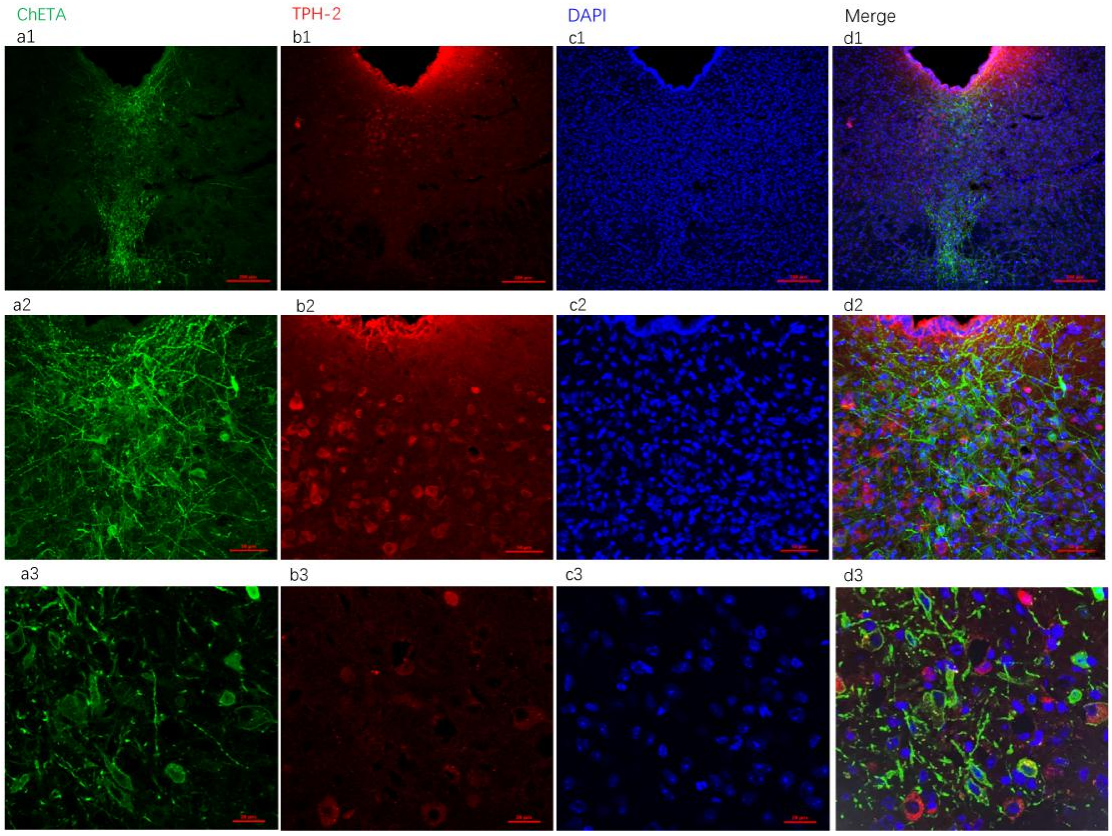


Figure 3 Selective expression of ChETA on 5-HT neurons in the DR of DBA/1 mice

a1, a2 and a3, neuronal immunostaining of GFP, a surrogate marker for ChETA, on 5-HT neurons in the DRof a coronal brain slice. b1, b2 and b3, immunostaining of TPH2, a key enzyme for 5-HT synthesis in the central nervous system (CNS). a1, b1, c3 and d1, merged images, showing the co-expression of TPH2 and GFP in 5-HT neurons. These data

demonstrate that ChETA is restrictively expressed on the surface of 5-HT neurons in the DR(n = 3 mice). Confocal image magnifications: a1–d1, 10x; a2–d2, 20x; a3–d3, 40x.

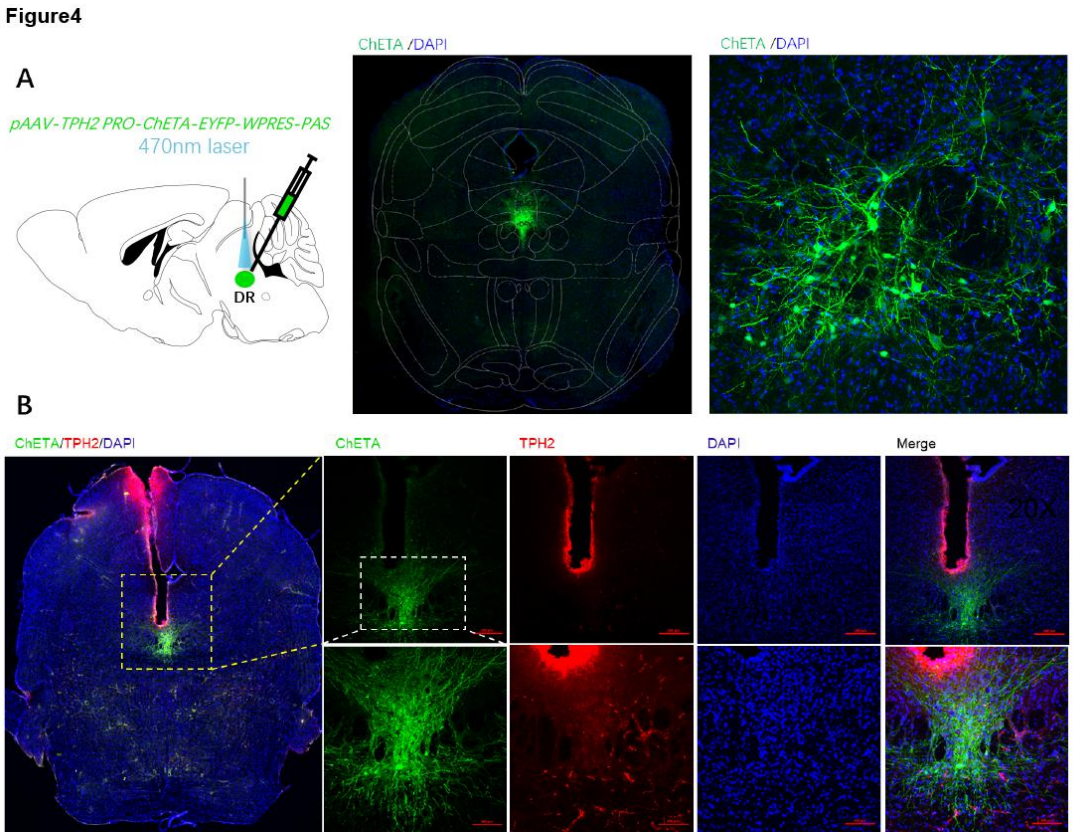


Figure 4 Placement of fiberoptic cannula tips in the DR

An example of coronal brain slice, showing the location of a optic fiber cannula tip and the expression of ChETA in the DR of a DBA/1 mouse, according to the mouse atlas of Paxinos and Franklin (4th Edition, Paxinos and Franklin,2013). B, the neuronal immunostaining of co-expression of ChETA and TPH2 in the DR of a DBA/1 mouse. No thermal injury due to photostimulation was observed in the area around the optic fiber tips.

Figure5

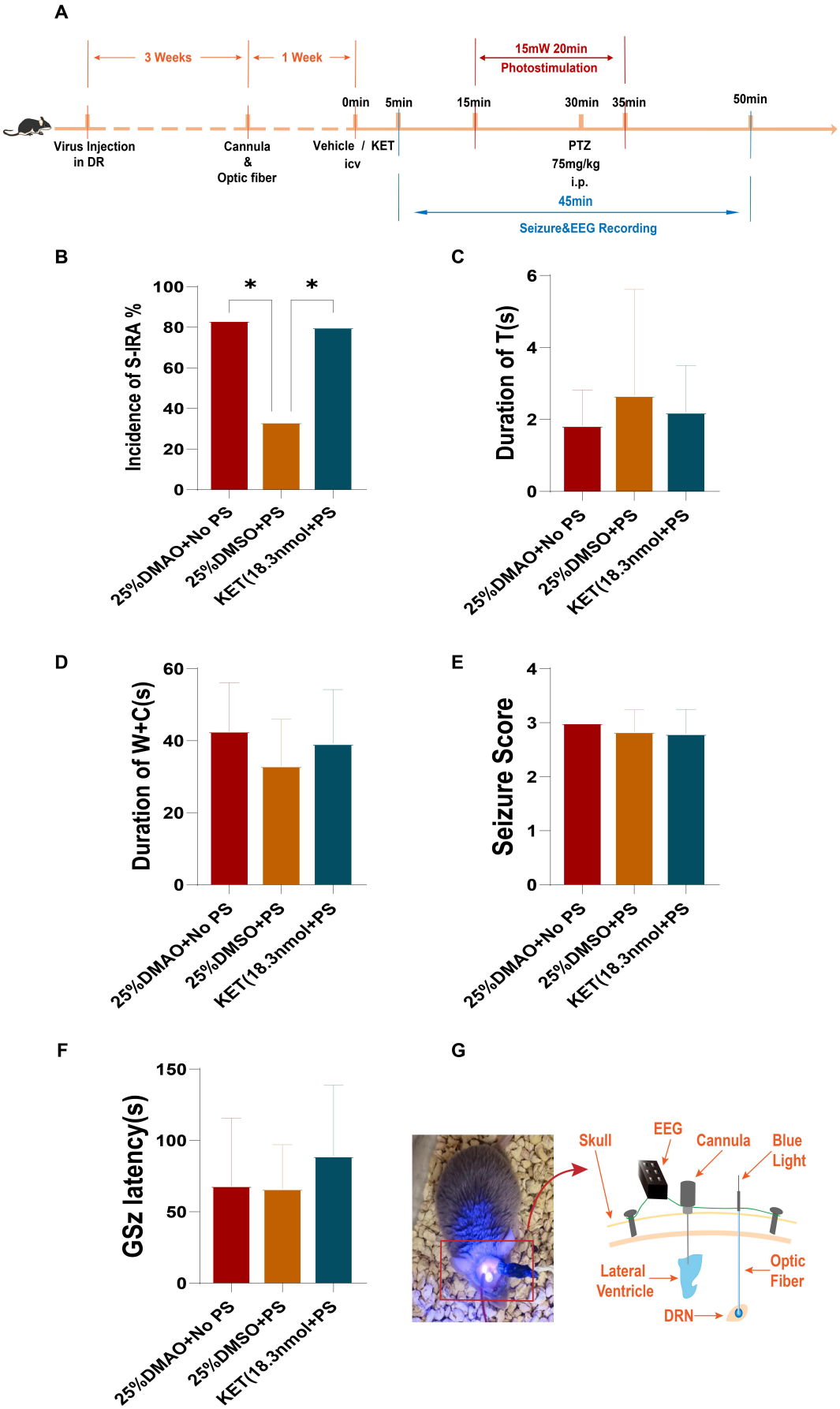

A
3 Weeks
1 Week
15mW 20min
Photostimulation
0min
5min
15min
30min
35min
50min
Virus Injection
in DR
Cannula
&
Optic fiber
Vehicle / KET
icv
PTZ
75mg/kg
i.p.
45min
Seizure&EEG Recording
B
Incidence of S-IRA %
100
80
60
40
20
0
*
*
25%DMAO+No PS
25%DMSO+PS
KET(18.3nmol+PS
C
Duration of T(s)
6
4
2
0
25%DMAO+No PS
25%DMSO+PS
KET(18.3nmol+PS
D
Duration of W+C(s)
60
40
20
0
25%DMAO+No PS
25%DMSO+PS
KET(18.3nmol+PS
E
Seizure Score
4
3
2
1
0
25%DMAO+No PS
25%DMSO+PS
KET(18.3nmol+PS
F
GSz latency(s)
150
100
50
0
25%DMAO+No PS
25%DMSO+PS
KET(18.3nmol+PS
G
Skull
EEG
Cannula
Blue
Light
Lateral
Ventricle
Optic
Fiber
DRN

Figure 5 Optogenetic activation of TPH2-ChETA neurons in DRN-mediated reduction of S-IRA evoked by PTZ was significantly reversed by ICV injection of KET without changing seizures behavior

B, Compared with the control group by treatment PTZ without photostimulation, the incidence of S-IRA by PTZ was significantly reduced by photostimulation (n=7, n=6, $p < 0.05$). However, the lower incidence of S-IRA by photostimulation was remarkably reversed by ICV injection of KET with the total content of 18.3 nmol (n=6, n=7, $p < 0.05$). C-F, No obvious difference between treatment groups in the analysis of seizure score, duration of wild ruing and clonic seizure, GSzs latency and duration of tonic seizure was to be found. G, Representative images of implantation devices in a DBA/1 mice for EEG, ICV and optic fiber. No PS = no photostimulation; PS = photostimulation

Figure6

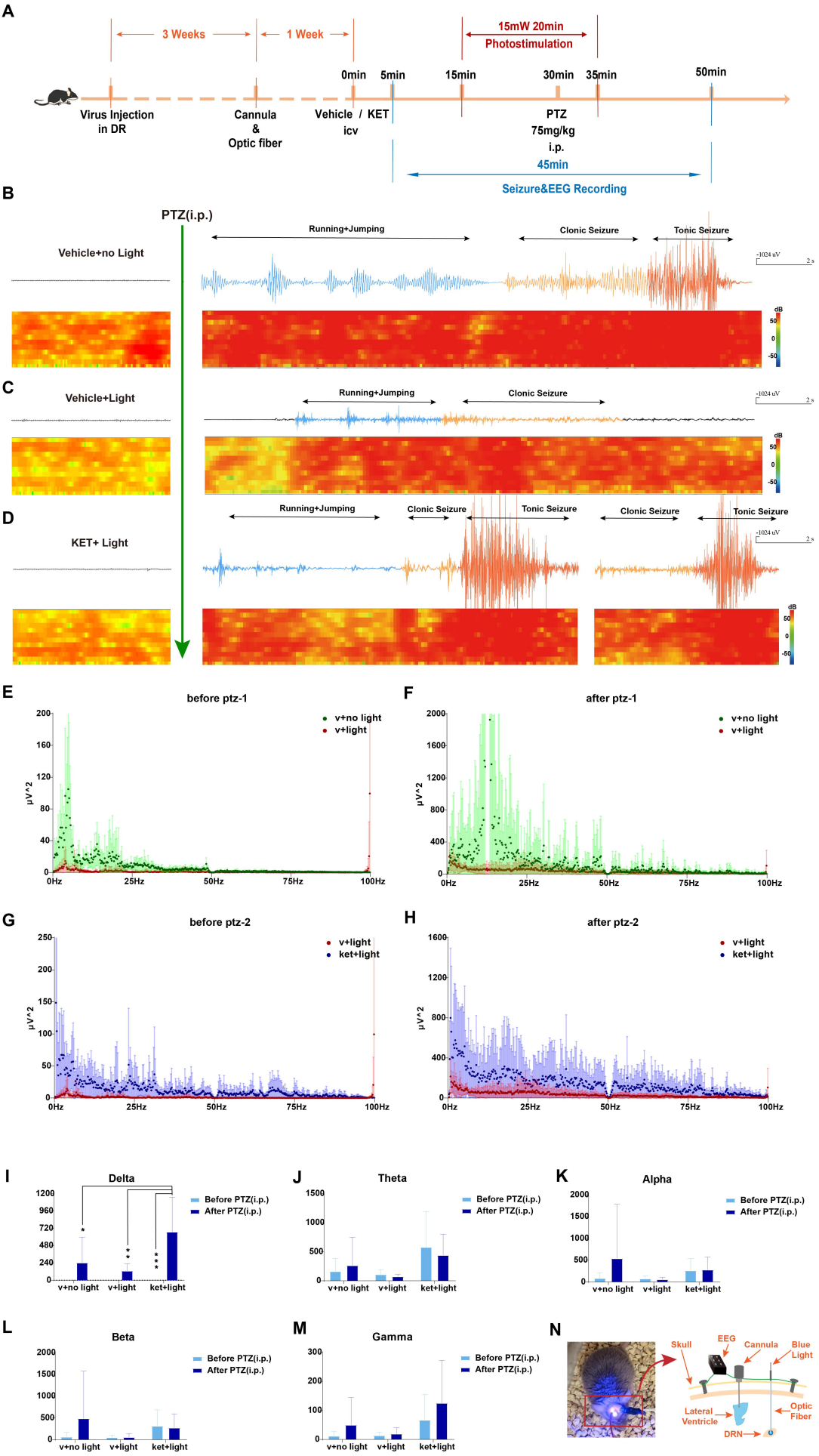

A
3 Weeks
1 Week
15mW 20min
Photostimulation
0min
5min
15min
30min
35min
50min
Virus Injection in DR
Cannula & Optic fiber
Vehicle / KET icv
PTZ 75mg/kg i.p.
45min
Seizure&EEG Recording
B
PTZ(i.p.)
Vehicle+no Light
Running+Jumping
Clonic Seizure
Tonic Seizure
C
Vehicle+Light
D
KET+ Light
E
before ptz-1
v+no light
v+light
F
after ptz-1
G
before ptz-2
ket+light
H
after ptz-2
μV^2
0Hz
25Hz
50Hz
75Hz
100Hz
I
Delta
J
Theta
K
Alpha
L
Beta
M
Gamma
Before PTZ(i.p.)
After PTZ(i.p.)
N
Skull
EEG
Cannula
Blue Light
Lateral Ventricle
Optic Fiber
DRN

Figure 6 The changing of EEG activity between optogenetic activation of TPH2-ChETA neurons in DRN-mediated reduction of S-IRA by PTZ and it was significantly reversed by ICV injection of KET

B-D, Compared with the control group by treatment PTZ without photostimulation, the EEG activity of during the clonic seizure and tonic seizure stage was significantly suppressed in the group treatment PTZ with photostimulation. However, the suppression of EEG activity was significantly increased in the group in the group treatment PTZ with photostimulation by microinjection ICV of KET. E-M, Delta wave of EEG activities was significantly reduced by light and reversed by KET. N, Representative images of implantation devices in a DBA/1 mice for EEG, ICV and optic fiber. Light = photostimulation; No light = no photostimulation

Figure7

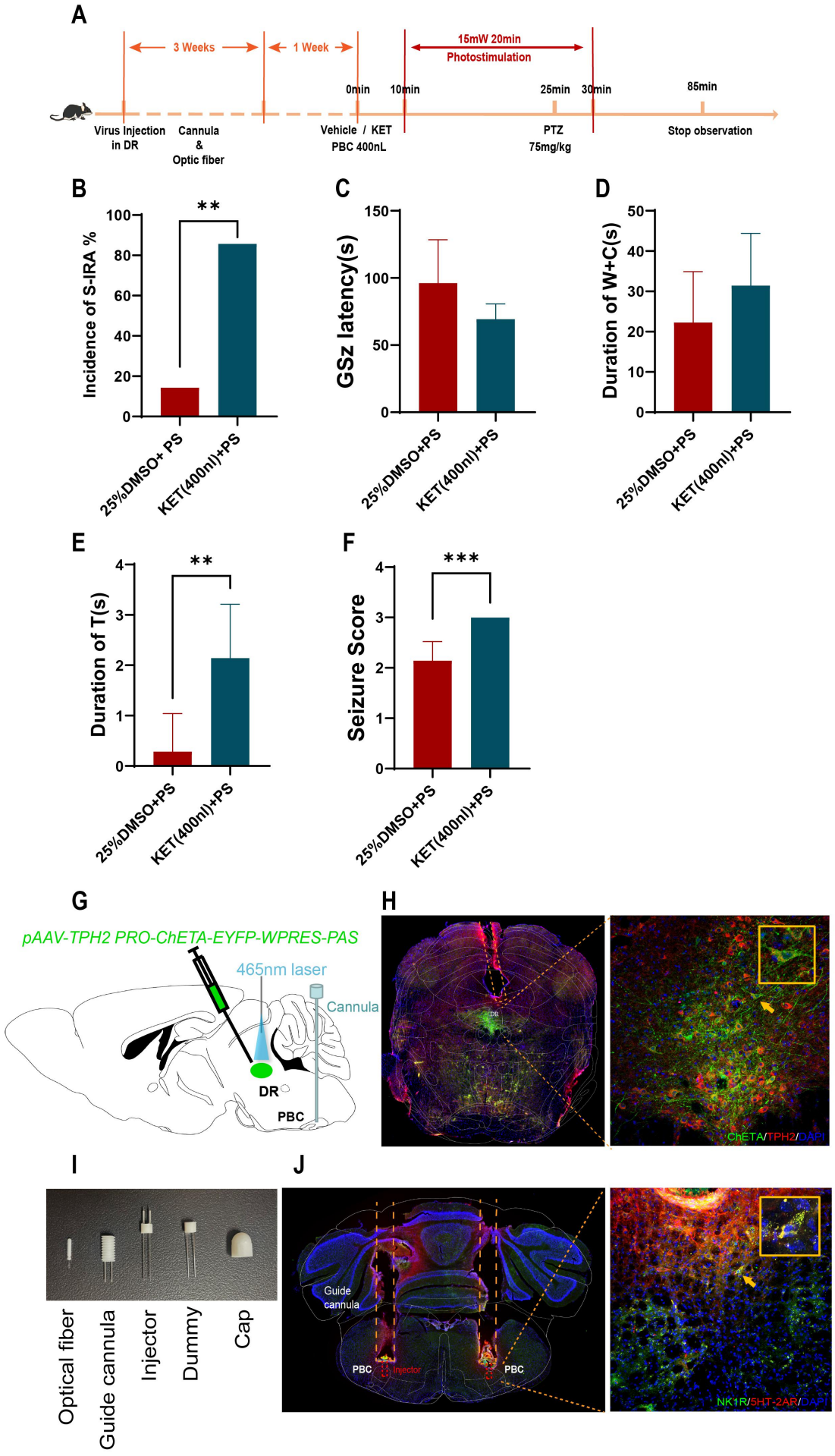

A
3 Weeks
1 Week
15mW 20min
Photostimulation
0min
10min
25min
30min
85min
Virus Injection in DR
Cannula & Optic fiber
Vehicle / KET PBC 400nL
PTZ 75mg/kg
Stop observation
B
Incidence of S-IRA %
**
25%DMSO+ PS
KET(400nl)+PS
C
GSz latency(s)
25%DMSO+PS
KET(400nl)+PS
D
Duration of W+C(s)
25%DMSO+PS
KET(400nl)+PS
E
Duration of T(s)
**
25%DMSO+PS
KET(400nl)+PS
F
Seizure Score
***
25%DMSO+PS
KET(400nl)+PS
G
pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS
465nm laser
Cannula
DR
PBC
H
ChETA/TPH2/DAPI
I
Optical fiber
Guide cannula
Injector
Dummy
Cap
J
Guide cannula
PBC
Injector
PBC
NK1R/5HT-2AR/DAPI

Figure 7 Optogenetic activation of TPH2-ChETA neurons in DR-mediated reduction of S-IRA evoked by PTZ was significantly reversed by injection of KET into the pre-Bötzinger complex

B, Compared with the control group by treatment PTZ with photostimulation, the incidence of S-IRA by PTZ was significantly reduced by photostimulation by microinjection of KET into the bilateral pre-Bötzinger complex with the total volume of 400 nl. C-D, No obvious difference between treatment groups in the analysis of seizure score, duration of wild ruing and clonic seizure, GSzs latency and duration of tonic seizure was to be found. E-F, Compared with the 25% DMSO + PS, duration of tonic seizure or seizure score were significantly increased, respectively.($p<0.05$, $p<0.01$) No PS = no photostimulation; PS = photostimulation. G-J, Representative images of the delivery of pAAV-TPH2 PRO-ChETA-EYFP-WPRES-PAS into the DR and of injection of KET into the bilateral pre-Bötzinger complex. H, The staining of ChETA, TPH2 and DAPI in the DR. I, All of implantation devices in a DBA/1 mice. J, The track of two sides injection of KET into the bilateral pre-Bötzinger complex and staining for NK1R, 5-HT2AR and DAPI in the bilateral pre-Bötzinger complex. No PS = no photostimulation; PS = photostimulation

Figure 8

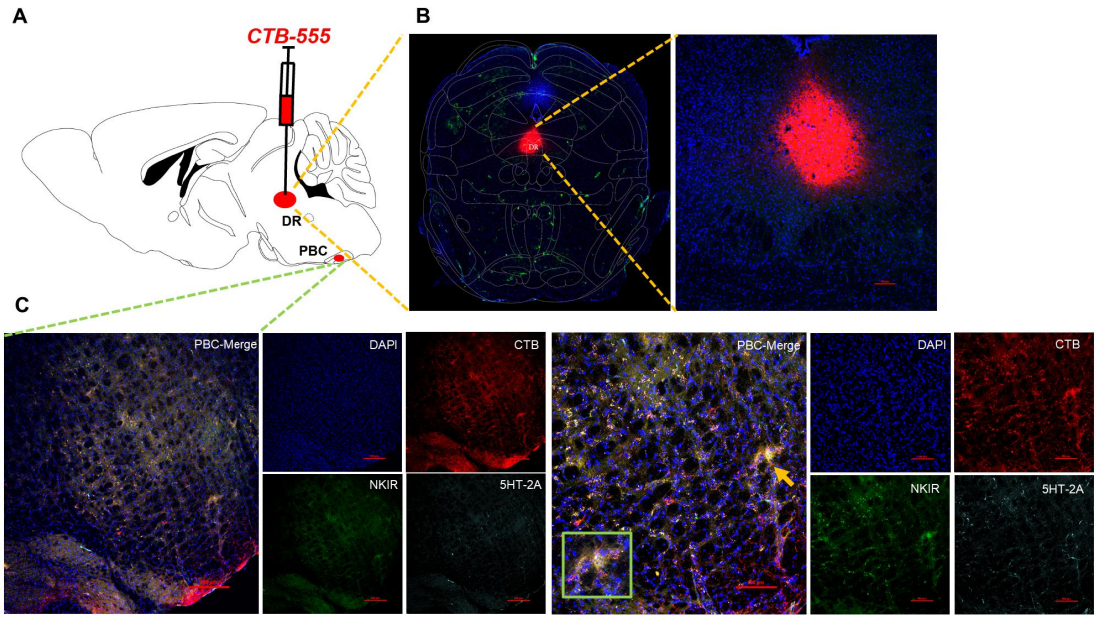


Figure 8 The neural projection from the DR to the pre-Bötzinger complex was

established by application of a nerve tracer of CTB555

A. An example of coronal brain slice, showing the location of a injection of CTB555 with the co-expression with TPH2 in DR. B. The projection of DR to pre-Bötzinger complex with the co-expression between CTB, NK1R and 5HT2AR.

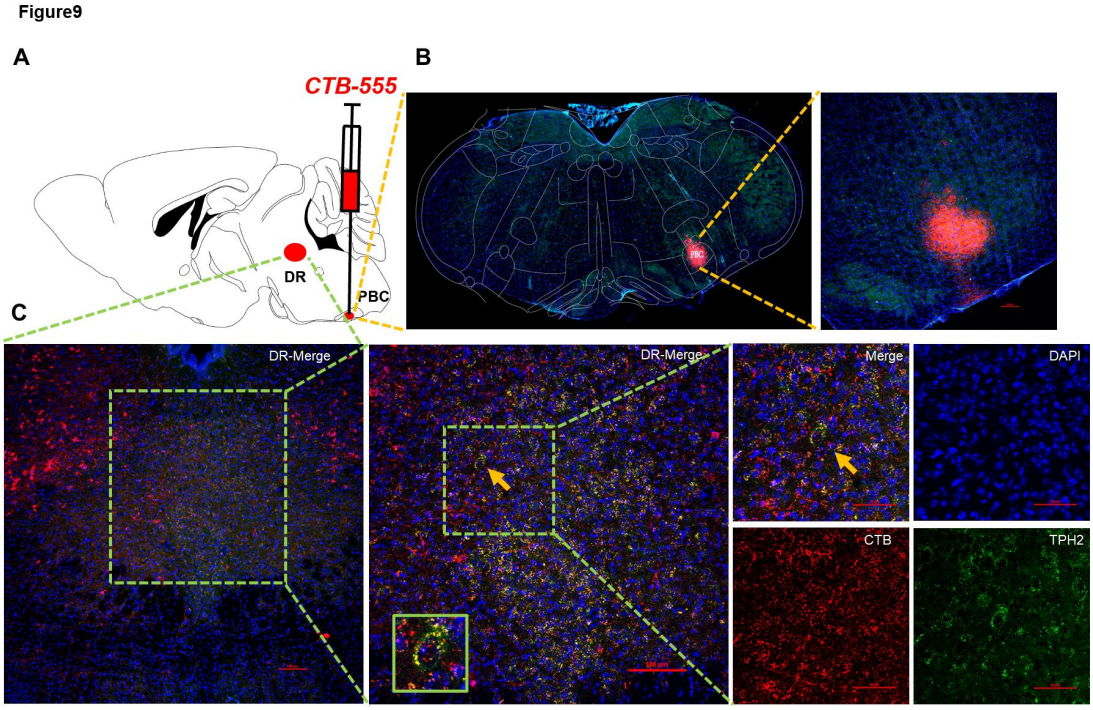
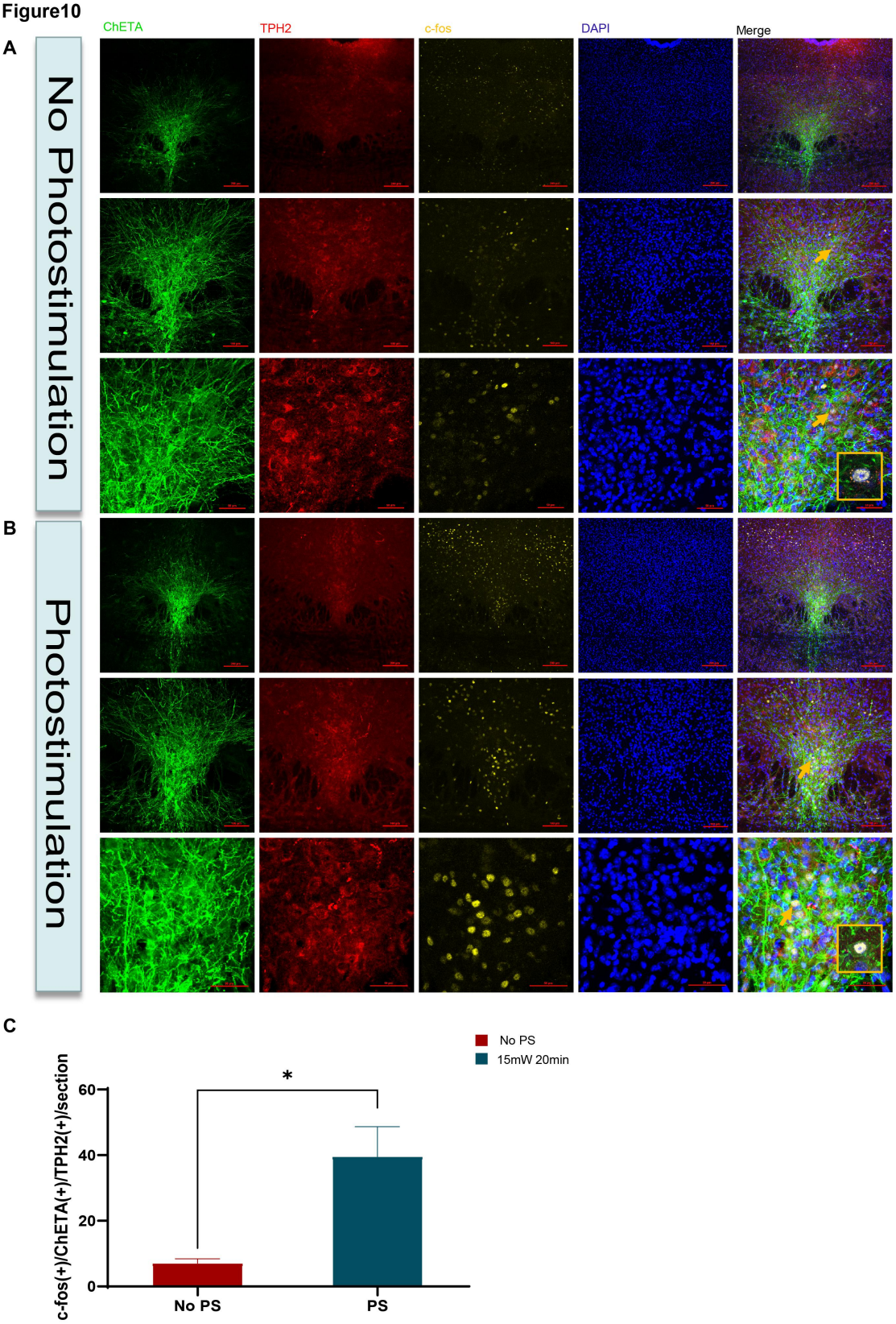


Figure 9 The neural projection from pre-Bötzinger complex to the DR was established by application of a nerve tracer of CTB555

A. An example of coronal brain slice, showing the location of a injection of CTB555 with the co-expression with NK1R and 5HT2AR in DRN. B, The projection to DR with the co-expression between TPH2 and CTB555.

Figure10



B. Figure 10 C-fos expression was significantly increased in DR by photostimulation of TPH2-ChETA neurons in DRin DBA/1 mice

A, neuronal immunostaining of c-fos, TPH2, GFP and co-localization c-fos, TPH2 and GFP in

a DBA/1 mouse that was implanted with fibe roptic cannula without photostimulation (n = 2 mice). B, immunostaining of c-fos, TPH2 GFP and co-localization c-fos, TPH2 and GFP, in an implanted DBA/1 mouse with photostimulation at 15 mW for 20 min (n = 2 mice). C, quantification of c-fos(+)/GFP(+)/TPH2(+) cells in implanted DBA/1 mice without photostimulation and those in implanted DBA/1 mice with photostimulation. The positive cells of c-fos(+)/GFP(+)/TPH2(+) in PS group significantly increased than the No PS group. ($p < 0.05$) No PS = no photostimulation; PS = photostimulation

Figure11

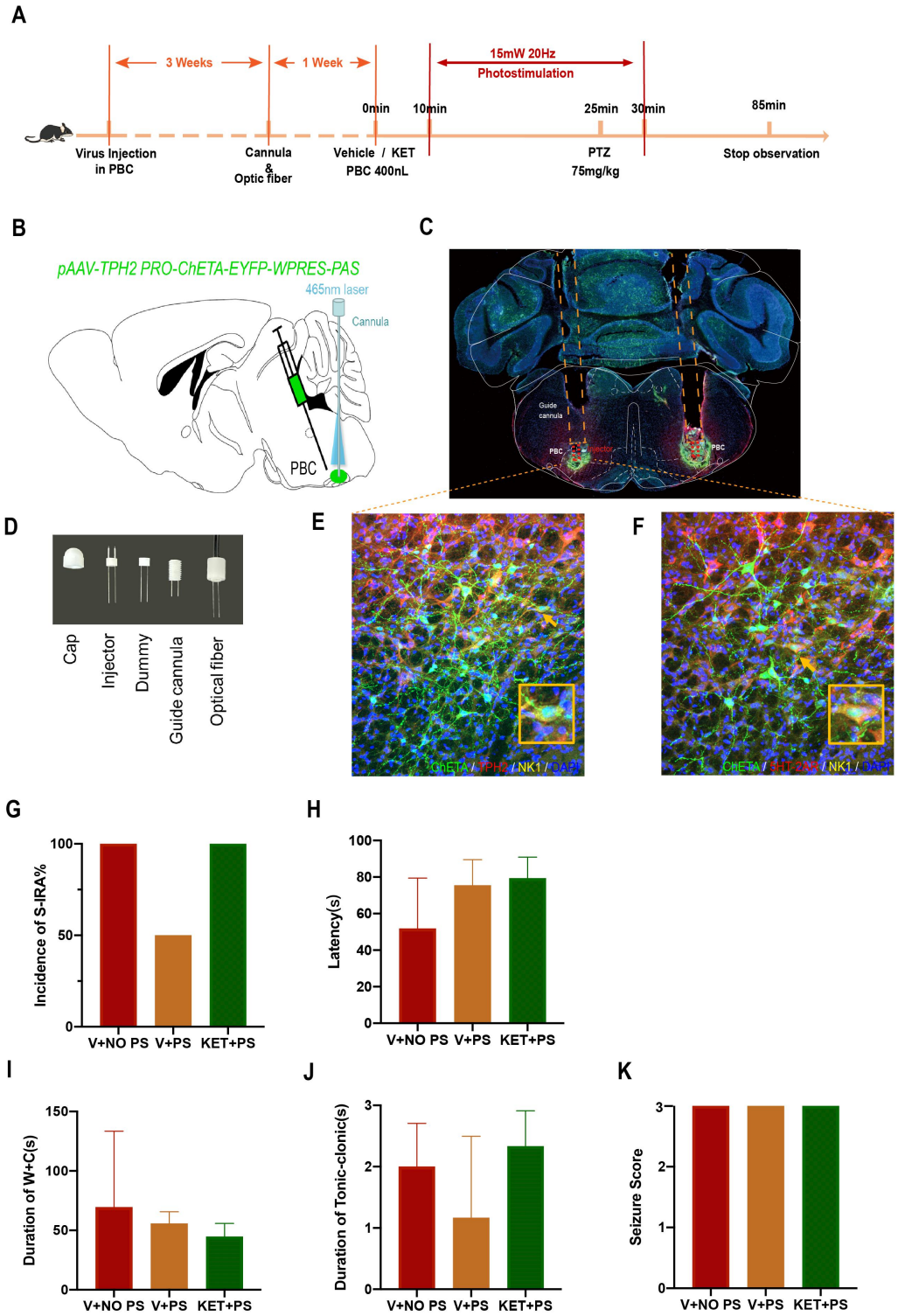


Figure 11 Activation of ChETA-TPH2 neurons in the PBC can suppress the S-IRA but it was insufficient to reduce the incidence of S-IRA of DBA/1 mice and block its death.

C-F, The track of two sides of optic fibers with cannula implanted into the bilateral pre-Bötzinger complex and staining for ChETA, TPH2, NK1R, 5-HT2AR and DAPI in the

bilateral pre-Bötzinger complex, respectively. G, Compared with the control group by treatment PTZ without photostimulation, the incidence of S-IRA by PTZ was not significantly reduced by photostimulation of the bilateral pre-Bötzinger complex. ($p > 0.05$). Furthermore, there was not significant difference between the group with photostimulation with microinjection of vehicle into the bilateral pre-Bötzinger complex and the group with photostimulation with microinjection of KET into the bilateral pre-Bötzinger complex.($p > 0.05$). H-K, No obvious difference between treatment groups in the analysis of seizure score, duration of wild ruing and clonic seizure, GSzs latency, duration of tonic seizure and seizure score was to be found. ($p > 0.05$). No PS = no photostimulation; PS = photostimulation

Figure12

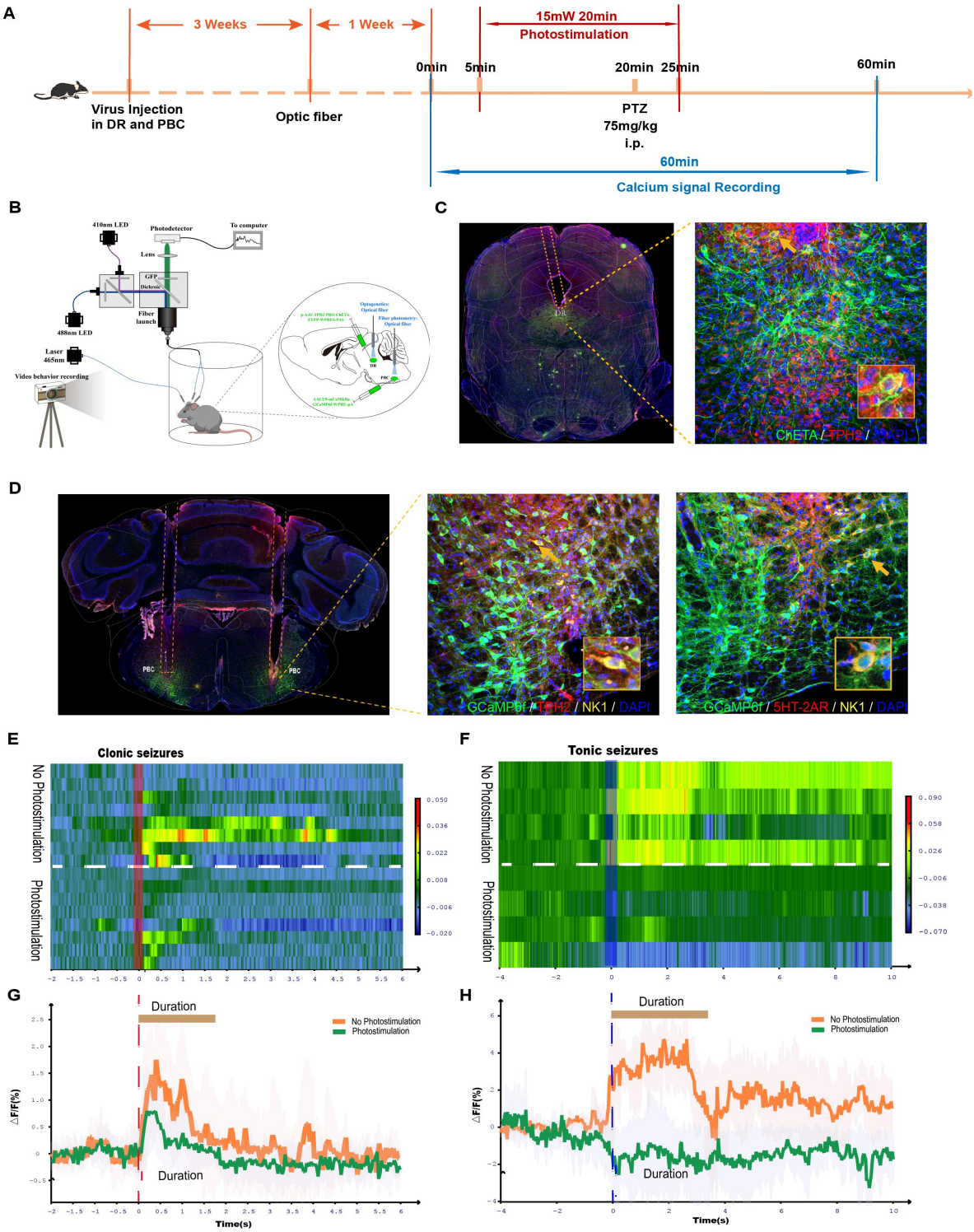


Figure 12 The changes of neural activity of calcium signal evoked by PTZ in the pre-Bötzinger complex during seizures and the effects of photostimulation of the DR on it by photometry recordings

A-D. All DBA/1 mice infected with ChETA and GCaMP6f in the DR and the bilateral pre-Bötzinger complex, respectively, and the neural activity of calcium signal from the bilateral pre-Bötzinger complex was recorded by photometry recordings. The track of optic fibers implanted into the DR and the bilateral pre-Bötzinger complex and staining for ChETA, TPH2 and GCaMP6f , NK1R, 5-HT2AR and DAPI in the DR and bilateral pre-Bötzinger complex, respectively. E-H, The group without photostimulation of the DR, the activity wave of calcium signals from the bilateral pre-Bötzinger complex were strong during the phase of clonic and tonic seizures evoked by PTZ, respectively. E-F, However, the activity wave of calcium signals were weaker in the group with photostimulation of the DR during the phase of clonic and tonic seizures, respectively

Figure13

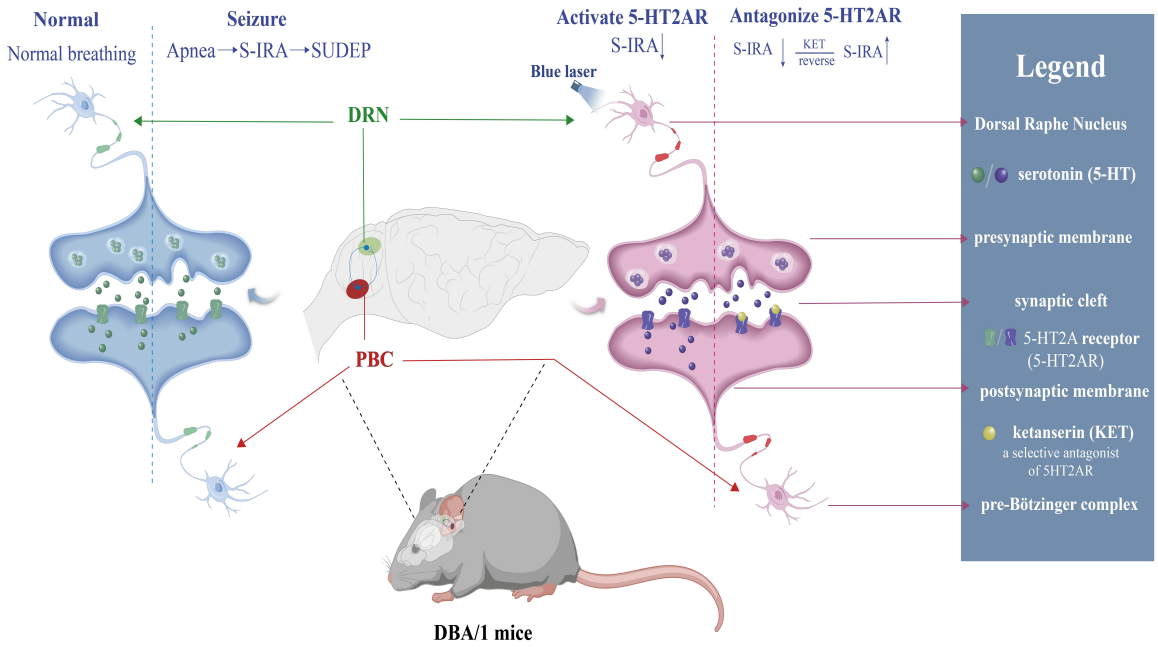


Figure 13 Mechanism of 5-HT neurons in brain modulates the S-IRA and SUDEP via the neural circuit between DRand the pre-Bötzinger complex

SUDEP: sudden unexpected death in epilepsy; S-IRA; seizure-induced respiratory arrest; KET: ketanserin; PBC; pre-Bötzinger complex; DR; dorsal raphe nucleus; 5HT2AR; 5HT2A receptor

9. References

1. Devinsky O, Hesdorffer DC, Thurman DJ, Lhatoo S, Richerson G. (2016). Sudden unexpected death in epilepsy: epidemiology, mechanisms, and prevention. Lancet Neurol (15), 1075–1088.
2. Philippe Ryvlin, Lina Nashef, Samden D Lhatoo, Lisa M Bateman, Jonathan Bird, Andrew Bleasel et al. Incidence and mechanisms of cardiorespiratory arrests in epilepsy monitoring units (MORTEMUS): a retrospective study.(2013). Lancet Neurol 12(10):966–977.
3. Lhatoo S, Noebels J, Whittemore V, NINDS Center for SUDEP Research. (2015). Sudden unexpected death in epilepsy: identifying risk and preventing mortality, Epilepsia (56), 1700–1706.
4. Massey CA, Sowers LP, Dlouhy BJ, Richerson GB. (2014). Mechanisms of sudden unexpected death in epilepsy: the pathway to prevention. Nat Rev Neurol (10), 271–82.
5. Zhang H, Zhao H, Yang X, Xue Q, Cotten JF, Feng HJ. (2016). 5-Hydroxytryptophan, a precursor for serotonin synthesis, reduces seizure-induced respiratory arrest. Epilepsia (57), 1228–1235.
6. Zhao H, Zhang H, Schoen FJ, Schachter SC, Feng HJ. (2019). Repeated generalized seizures can produce calcified cardiac lesions in DBA/1 mice. Epilepsy Behav 95 (4) 169–174.
7. Zhang H, Zhao H, Zeng C, Van Dort C, Faingold CL, Taylor NE, Ken Solt, Hua-Jun Feng. (2018). Optogenetic activation of 5-HT neurons in the dorsal raphe suppresses seizure-induced respiratory arrest and produces anticonvulsant effect in the DBA/1 mouse SUDEP model. Neurobiol Dis (110), 47–58.
8. Zhang H, Zhao H, Feng HJ. (2017). Atomoxetine, a norepinephrine reuptake inhibitor, reduces seizure-induced respiratory arrest. Epilepsy Behav 73:6–9.
9. Zhao H, Cotten JF, Long X, Feng HJ. (2017). The effect of atomoxetine, a selective norepinephrine reuptake inhibitor, on respiratory arrest and cardiorespiratory function in the DBA/1 mouse model of SUDEP. Epilepsy Res (137), 139–144.

10. Chang Zeng, Xiaoyan Long, Joseph F Cotten, Stuart A Forman, Ken Solt, Carl L Faingold, Hua-Jun Feng. (2015). Fluoxetine prevents respiratory arrest without enhancing ventilation in DBA/1 mice. Epilepsy Behav 2015, 45:1–7
11. Qinglan Chen, Fafa Tian, Qiang Yue, Qiong Zhan, Mian Wang, Bo Xiao, Chang Zeng. (2019). Decreased serotonin synthesis is involved in seizure-induced respiratory arrest in DBA/1 mice. Neuroreport. 14;30(12):842–846.
12. Yue Shen, Hai XiangMa, Han Lu, Hai Ting Zhao, Jian liang Sun, Yuan Cheng, HongHai Zhang. (2021). Central deficiency of norepinephrine synthesis and norepinephrinergic neurotransmission contributes to seizure-induced respiratory arrest. Biomedicine & Pharmacotherapy. 133, 111024.
13. kayama, Naoya Nishitani, Chihiro Andoh, Masashi Koda, Hisashi Shirakawa, Takayuki Nakagawa, Kazuki Nagayasu, Akihiro Yamanaka, Shuji Kaneko. (2020). The Role of Dorsal Raphe Serotonin Neurons in the Balance between Reward and Aversion Int J Mol Sci. 21(6):2160.
14. Tryba AK, Peña F, Ramirez JM. (2006). Gasping Activity In Vitro: A Rhythm Dependent on 5-HT2A Receptors. J. Neurosci. 26:2623–2634.
15. Peña F, Ramirez JM. (2002). Endogenous activation of serotonin-2A receptors is required for respiratory rhythm generation in vitro. J. Neurosci. 15:22:11055-11064.
16. Ozawa Y, Okado N. (2002). Alteration of serotonergic receptors in the brain stems of human patients with respiratory disorders. Neuropediatrics. 33:142–149.
17. Bateman LM, Li CS, Lin TC, Seyal M. (2010). Serotonin reuptake inhibitors are associated with reduced severity of ictal hypoxemia in medically refractory partial epilepsy. Epilepsia (51), 2211–2214.
18. Buchanan GF, Murray NM, Hajek MA, Richerson GB. Serotonin neurones have anti-convulsant effects and reduce seizure-induced mortality. J. Physiol. 2014;592:4395–4410.
19. Alexandra N Petrucci, Katelyn G Joyal, Benton S Purnell, Gordon F Buchanan. (2020). Serotonin and sudden unexpected death in epilepsy. Exp Neurol 325:113145.

20. .Zhang X, Beaulieu JM, Sotnikova TD, et al. (2004).Tryptophan hydroxylase-2 controls brain serotonin synthesis. Science 305:217-218.
21. Kulikov AV, Osipova DV, Naumenko VS, et al. (2005). Association between Tph2 gene polymorphism, brain tryptophan hydroxylase activity and aggressiveness in mouse strains. Genes Brain Behav 4;482–485.
22. Osipova DV, Kulikov AV, Mekada K, et al. (2010). Distribution of the C1473G polymorphism in tryptophan hydroxylase 2 gene in laboratory and wild mice. Genes Brain Behav 9:537–543.
23. E. Donato Di Paola, P. Gareri, A. Davoli, S. Gratteri, F. Scicchitano, C. Naccari, et al. (2007). Influence of levetiracetam on the anticonvulsant efficacy of conventional antiepileptic drugs against audiogenic seizures in DBA/2 mice, Epilepsy Res 75 112–121.
24. C. Soper, E. Wicker, C.V. Kulick, P. N'Gouemo, P.A. Forcelli. (2016). Optogenetic activation of superior colliculus neurons suppresses seizures originating in diverse brain networks, Neurobiol. Dis. (87) 102–115.Google Scholar