Genomic basis of drought resistance in *Fagus sylvatica*

Markus Pfenninger[1,2,3], Friederike Reuss[1], Angelika Kiebler[1], Philipp Schönnenbeck[4], Cosima Caliendo[1,4], Susanne Gerber[4], Berardino Cocchiararo[3,5], Sabrina Reuter[6], Nico Blüthgen[6], Karsten Mody[6,11], Bagdevi Mishra[7], Miklós Bálint[3,8,9], Marco Thines[3,7,10], Barbara Feldmeyer[1]

[1] Molecular Ecology, Senckenberg Biodiversity and Climate Research Centre, Frankfurt am Main, Germany

[2] Institute for Organismic and Molecular Evolution, Johannes Gutenberg University, Mainz, Germany

[3] LOEWE Centre for Translational Biodiversity Genomics, Frankfurt am Main, Germany

[4] Institute of Human Genetics, University Medical Center, Johannes Gutenberg University, Mainz, Germany

[5] Conservation Genetics Section, Senckenberg Research Institute and Natural History Museum Frankfurt, Gelnhausen, Germany

[6] Ecological Networks lab, Department of Biology, Technische Universität Darmstadt, Darmstadt, Germany

[7] Biological Archives, Senckenberg Biodiversity and Climate Research Centre, Frankfurt am Main, Germany

[8] Functional Environmental Genomics, Senckenberg Biodiversity and Climate Research Centre, Frankfurt am Main, Germany

[9] Agricultural Sciences, Nutritional Sciences, and Environmental Management, Universität Giessen, Giessen, Germany

[10] Institute for Ecology, Evolution and Diversity, Johann Wolfgang Goethe-University, Frankfurt am Main, Germany

[11] Department of Applied Ecology, Hochschule Geisenheim University, Geisenheim, Germany

Abstract

In the course of global climate change, central Europe is experiencing more frequent and prolonged periods of drought. The drought years 2018 and 2019 affected European beeches (*Fagus sylvatica* L.) differently: even in the same stand, drought damaged trees neighboured healthy trees, suggesting that the genotype rather than the environment was responsible for this conspicuous pattern. We used this natural experiment to study the genomic basis of drought resistance with Pool-GWAS. Contrasting the extreme phenotypes identified 106 significantly associated SNPs throughout the genome. Most annotated genes with associated SNPs (>70%) were previously implicated in the drought reaction of plants. Non-synonymous substitutions led either to a functional amino acid exchange or premature termination. A SNP-assay with 70 loci allowed predicting drought phenotype

38 in 98.6% of a validation sample of 92 trees. Drought resistance in European beech is a moderately
39 polygenic trait that should respond well to natural selection, selective management, and breeding.

Keywords
41 Genome-wide association study, genomic prediction, forest tree, Fagales, conservation genomics,
42 functional environmental genomics

Impact Statement
44 European beech harbours substantial genetic variation at genomic loci associated with drought
45 resistance and the loci identified in this study can help to accelerate and monitor adaptation to
46 climate change.

Introduction
Climate change comes in many different facets, amongst which are prolonged drought periods
(Christensen *et al.* 2007). The Central European droughts in the years 2018 and 2019 caused severe
water stress in many forest tree species, leading to the die-off of many trees (Schuldt *et al.* 2020a).
Among the suffering tree species was European beech, *Fagus sylvatica* L. As one of the most
common deciduous tree species in Central Europe, *F. sylvatica* is of great ecological importance:
beech forests are a habitat for more than 6,000 different animal and plant species (Brunet *et al.*
2010; Dorow *et al.* 2010). The forestry use of beech in 2017 generated a turnover of more than 1
billion € in Germany alone (Thünen_Institute 2020), without taking the economic and societal value
of the ecosystem services of woods into account (Elsasser *et al.* 2016). However, the drought years
2018 and 2019 severely impacted the beech trees in Germany (Paar & Dammann 2019). Official
reports on drought damage in beech recorded 62% of trees with rolled leaves and 20-30% of small
leaves, mainly in the crown, resulting in 7% of badly damaged or dead trees. As shown before
(Bressem 2008), most trees affected by drought stress were medium to old aged.

Under favourable conditions, beech is a competitive and shade tolerant tree species, dominating
mixed stands (Pretzsch *et al.* 2013). High genetic diversity within populations supports adaptation to
local conditions (Kreyling *et al.* 2012). Significant differences between local populations in tolerance
to various stress factors such as early frost (Czajkowski & Bolte 2006), drought (Cocozza *et al.* 2016;
Harter *et al.* 2015) or air pollution (Müller-Starck 1985) are known. The distribution of *F. sylvatica* is
mainly limited by water-availability, as the tree does not tolerate particularly wet or dry conditions
(Sutmöller *et al.* 2008). Therefore, it is quite conceivable that the species could suffer even more
under the predicted future climatic conditions than today (Sutmöller *et al.* 2008).

Despite the widespread, severe drought damage, a pattern observed in all beech forests was very
noticeable (personal observations). Using crown deteriation as significant indicator for drought
damage (Choat *et al.* 2018), not all trees in a beech stand were equally damaged or healthy. The
damage occurred rather in a mosaic-like pattern instead. Even though the extent of drought damage
varied among sites, apparently completely healthy trees immediately neighboured severely damaged
ones and *vice versa*. This observation gave rise to the hypothesis that not the local environmental
conditions might be decisive for the observed drought damage, but rather the genetic make-up of
the individual trees.

We decided to draw on this natural "experimental set-up" to infer the genomic basis underlying the
drought susceptibility in *F. sylvatica*. We identified more than 200 neighbouring pairs of trees with
extreme phenotypes and used a Pool-GWAS approach (Bastide *et al.* 2013) to infer associated SNP
loci by contrasting allele frequencies with replicated pools of drought susceptible and resistant
individuals. In addition, we individually re-sequenced a subset of 51 pairs of susceptible and resistant
trees. If the observed pattern indeed has a genetic basis, identifying the associated loci would enable

the genomic prediction of drought resistance (Stocks *et al.* 2019). Constructing a SNP assay from the
most highly phenotype associated SNPs, we validated 70 identified loci by predicting the drought
phenotype of an additional set of beech trees from their genotype at these loci using Linear
Discriminant Analysis and a new Machine Learning approach (Horenko 2020). These accurate
genomic prediction tools, e.g., the choice of drought resistant seed producing trees and selective
logging could help accelerate and monitor natural selection and thus harness beech forests against
climate change (Waldvogel *et al.* 2020).

Results

Sampling, climate development and phenotyping

Damaged and healthy beech tree pairs were sampled from woods in the lowland Rhein-Main plain,
the adjacent low mountain ranges of Odenwald and Taunus, and mountain ranges from Central and
Northern Hessen (Fig. 1A). When summarising the climatic conditions from 1950 to 2019 for the
sampling sites in a principal component analysis (PCA), the sites were divided into two groups by axis
1, a temperature gradient. The Taunus mountain sites grouped with those from the northern part of
Hessen, while the Rhein-Main plain clustered with the Odenwald sites (Fig. 1B). This grouping was
also used to construct the GWAS pools (see below). Comparing the climate from the 1950s, when
most of the trees sampled were already in place, with the decade from 2010-2019, showed that all
local conditions changed substantially and similarly in the direction and extent of warmer and drier
conditions (Fig. 1B). The steepest temperature increase occurred in the 1980s, while precipitation
patterns mainly changed in the last decade (Suppl. Fig. 1). A wide range of parameters, potentially
relevant as selection pressures changed drastically during this period: the mean January daily
minimum temperature at the sampling sites increased by 1.49°C from -2.64°C (s.d. 1.68°C) in the
1950s to -1.15°C (s.d. 2.50°C) during the last decade. The mean August daily maximum temperatures
increased even more by 2.37°C from 22.06°C (s.d. 1.95°C) to 24.43°C (s.d. 2.35°C). Simultaneously,
mean annual precipitation decreased by 40.5 mm or 5.5% from 741.2 mm (s.d. 85.8 mm) to 700.7
mm (s.d. 70.9 mm). Most of the precipitation loss (84%) occurred during the main growth period
between April and September, with a decrease of 33.9 mm from 410.4 mm (s.d. 36.1 mm) to 376.5
mm (s.d. 25.6 mm).

Mean monthly evaporation potential, available from 1991 onwards, showed that, compared to the
beginning of the 1990s, the main growth period of beech from April to September became
increasingly drier, with up to 30 mm more evaporation per month. The drought dynamics suggested
that the years 2018 and 2019 were not outliers, but rather part of a long-term, accelerating trend
(Fig. 1C), following the overall global pattern (Büntgen *et al.* 2021; Trenberth *et al.* 2014).

There was a strong negative correlation (r = 0.695) between the drought strength during the main
growth period (Apr- Sept) and a proxy for (green) leaf cover (leaf area index, LAI) for the sampled
plots in the years 2015-2019 (Suppl. Fig. 2). This observation suggested that leaf loss and dried leaves
are good indicators for drought stress.

The mean distance between paired trees was 5.1 m (s.d. 3.4 m, Suppl. Fig. 3). Phenotypic
measurements generally confirmed the study design and selection of trees: healthy and damaged
trees within each tree pair did not differ significantly in *trunk circumference*, *tree height*, *canopy*
closure and *competition index* (Fig. 2 A-D, Suppl. Table 2). Hence, these parameters were not
considered in further analyses. As expected, and confirming the assignment of damage status, the
quantity of *dried leaves* and *leaf loss* differed substantially between damaged and healthy ones (Fig.
2 E-F, Suppl. Table 2). A sample of photographs contrasting damaged and healthy paired trees can be
found in the Suppl. Fig. 4.

Linkage disequilibrium, population structure and genome-wide association study

For a subsample of 300 out of the 402 sampled beech trees we generated four DNA pools from two
climatically distinct regions (North and South Hessen, Fig. 1B), contrasting trees that were either
healthy or highly drought damaged respectively (Tab. S1). The "South" pools consisted of 100
individuals each, whereas the "North" pools contained 50 individuals each. We created ~50GB 150
bp-paired end reads with insert size 250-300 bp on an Illumina HiSeq 4000 system per pool. More
than 96% of the reads mapped against the repeat-masked chromosome level beech reference
genome (accession no. PRJNA450822). After filtering the alignment for quality and a coverage
between 15x and 70x, and removing indels, allele frequencies for 9.6 million SNPs were scored. All
100 individuals from the North population were additionally individually re-sequenced to ~20x
coverage each (for more details see M&M). This data was used to a) determine individual variability
in allele frequencies and b) to validate the information content of the candidate SNP-set.

Using all individually resequenced individuals, we inferred the extent of genome wide linkage
disequilibrium (LD;). The plot of LD r^2 against the distance from the focal SNP showed that LD fell to
$r^2 \sim 0.3$ within less than 120 bp, which means that genome positions such a distance apart are on
average effectively unlinked (Fig. 3A). The PCA on SNP variation of the individually re-sequenced
trees from the North population explained 12.3% of accumulated variation on the first two axes (Fig.
3B). Trees from the same sampling site (within the North population) did not tend to cluster together
(Fig. 3B). F_{ST} estimates among pools for non-overlapping 1 kb windows were virtually identical among
healthy/damaged pools within region as compared to between regions (Suppl. Fig. 5). Trees within a
phenotypic class were genomically not more similar than between classes (Suppl. Fig. 6, ANOSIM R =
-0.008, p = 0.76, 9,999 permutations).

Pool-GWAS analysis identified 106 SNPs significantly associated with the drought damage status
using a Cochran-Mantel-Haenszel test on the two pairs of damaged and healthy pools after false
discovery rate correction and a cut-off at 1×10^{-2} (Fig. 4A, Suppl. Fig. 7). Some of the 106 SNPs were
in close physical proximity (<120bp) and thus probably linked. Taking this into account, 80
independent genomic regions were associated with the drought damage status. None of the
significantly differentiated SNP loci was mutually fixed; the observed allele frequency differences
between healthy and damaged trees at associated loci ranged between 0.12 and 0.51 (Fig. 4B).

Associated genes and gene function

Of the 106 significant SNPs, 24 were found in 20 protein coding genes (Table 1). Forty-nine genes
were the closest genes to the remaining 82 SNPs. For 61 of these genes, the best BLAST hit was with
a tree, mainly from the Fagales genera *Quercus* and *Castanea* (Table 1, Suppl. Table 3). Among the 24
SNPs in genes, we observed 13 non-synonymous changes. In eleven of these changes, the alternate
allele was associated with the damaged phenotype and only in two cases with the healthy
phenotype. Three of the non-synonymous substitutions resulted in a stop codon. Of the remaining
ten, eight exchanges caused a major change in amino acid characteristics and thus probably in
protein folding or function (Table 1). One gene, a PB1 domain-containing protein tyrosine kinase,
contained four non-synonymous changes, suggesting that the allele version associated with the
damaged phenotype lost its function (Tab. 1). From the 20 genes with significant SNPs, functional
information could be obtained from the UniProt database for 14 (Suppl. Table 3). Of these, ten genes
were associated in previous studies with either environmental stress response (two) or specifically
with drought stress response (eight; Suppl. Table 3). Of the 49 predicted genes closest to the
remaining significant SNPs (Tab. 1), 16 could be reliably annotated (Suppl. Table 2). Twelve had been
directly related to drought in previous studies, while three were previously associated with other
environmental stress responses (Suppl. Tab. 3).

Genomic prediction

We furthermore set out to determine how many SNPs were needed to successfully predict the
drought susceptibility of individual trees, i.e. to develop a genotyping assay. All Pool-GWAS SNPs in
addition to the top 20 individual re-sequencing SNPs were used to create a SNP combination to reach
a genotyping success threshold of min. 90%. After excluding loci due to technical reasons and
filtering for genotyping success, seventy loci proved to be suitable for reliable genotyping with a SNP
assay. We genotyped only individuals sampled in 2019 that were not used to identify the SNPs in the
first place plus paired individuals sampled in Aug 2020. On average, each of the 95 individuals was
successfully genotyped at 67.7 loci (96.7%). We coded the genotypes as *0* for homozygous reference
allele, *1* for heterozygous and *2* for the homozygous alternate allele, thus assuming a linear effect
relationship. Figure 5 shows the genotypogram for the tested individuals.

Linear discriminant analysis (LDA) correctly predicted the observed phenotype from the genotype in
91 of 92 cases (98.9%). Prediction success decreased to 65% when successively removing loci from
the analysis (Suppl. Fig. 8). Nevertheless, ordering the individuals according to the LDA score of axis 1
revealed no clear genotype pattern that distinguished healthy from damaged trees (Fig. 5). Observed
heterozygosity at loci used in the SNP assay of individuals in the upper half of predictive values for a
healthy phenotype was not significantly different from heterozygosity of the lower half (Suppl. Fig.
9). Ordering the loci according to their squared loadings showed that loci's contribution to the
genomic prediction differed substantially (Fig. 5). As expected, the histogram of LDA scores showed
two peaks, corresponding to the two phenotypes (Suppl. Fig. 10).

To validate the results of the LDA prediction and to circumvent potential overfitting due to the small
sample size, we also applied a non-parametric Machine Learning algorithm for feature selection and
clustering that was especially designed for small sample sizes (Gerber2020, Horenko2020). The
Method identified the 20 most-significant SNPs allowing to make an almost 85% correct classification
that distinguished healthy from damaged trees (Suppl. Table 5).

Discussion
Over the last two decades, increasing drought periods caused severe damage to European forests
(Schuldt *et al.* 2020b; Etzold *et al.* 2019; Pretzsch *et al.* 2013). Conifers seem to suffer the most, but
also deciduous trees were strongly affected (Schuldt *et al.* 2020b). Weather data from our study area
from 1950 onwards suggested that the climatic conditions for beech trees in the area investigated
changed dramatically during this period. Roughly estimating the tree age from their trunk
circumference (Bošeľa *et al.* 2014), more than a third of the trees were already in place at the
beginning of this period. About 60% were recruited prior to the acceleration of temperature change
from the 1980s onwards. As a result, trees in the mountainous regions of the study area today
experience climatic conditions comparable to those experienced by low land trees in the 1950s,
which in turn now experience a climate that used to be typical for regions much further South. Given
the documented propensity of beech for local adaptation (Gárate-Escamilla *et al.* 2019; Pluess *et al.*
2016; Aranda *et al.* 2015), including drought (Bolte *et al.* 2016), it is therefore conceivable that
current conditions exceed the reaction norm of some previously locally well-adapted genotypes with
detrimental consequences for their fitness. If the trend of an increasingly drier vegetation period
persists, this will likely affect an even larger proportion of the currently growing beeches.

Evolutionary genomics will be indispensable to predict and manage the impact of global change on
biodiversity (Waldvogel *et al.* 2020). As already shown for other partially managed (tree) species
(Stocks *et al.* 2019), in particular pool-GWAS approaches (Endler *et al.* 2016) have proven to be useful
in guiding conservation management.

Our strictly pairwise sampling design avoided many pitfalls of GWAS studies, arising, e.g., from
cryptic population structure and shared ancestry (Hoban *et al.* 2016; Wellenreuther & Hansson
2016). Despite presented evidence from this and other studies (Schuldt et al. 2020) that the observed
crown damages in large parts of Central Europe used for phenotyping here are directly or indirectly
due to the severe drought years 2018 and 2019, we must acknowledge that we have no direct
physiological proof that the trees surveyed here indeed suffered from drought stress. In addition, the
observed diagnostic symptoms are not specific to drought stress. Nevertheless, an unknown
independent stressor would have needed to accidentally co-occur spatially and temporally with the
drought. The phenotypical drought response of individual trees may also be influenced by
microspatial variation (Carrière *et al.* 2020). In the present study, however, the mean distance
between sampled paired trees of about 5 m assured that their roots systems largely overlapped.
Thus, environmental variation in soil quality, rooting depth, water availability or other factors should
have been minimal. Please note that any phenotypical misclassification due to such microspatial
differences would have rather dissimulated the genotypic differences found in GWAS than enhanced
them artificially. Also he lumping of similar phenotypes induced by different stressors is unlikely to
have the same genomic basis and resulting in significant GWAs results.

As expected from previous studies (Rajendra *et al.* 2014), we found no population structure among
the sampling sites. Applying relatively strict significance thresholds, we found systematic genomic
differences between the healthy and damaged trees. In all cases, these differences were quantitative
and not categorical, i.e. we found allele frequency changes but no fixed SNPs between phenotypes.
Significant SNPs were mostly not clustered - we found on average 1.4 selected SNPs in a particular
region. These findings were in line with the observed very short average LD in *F. sylvatica,* indicating
that polymorphisms associated with the two phenotypes were likely old standing genetic variation
(Harris & Nielsen 2013). Moreover, such SNPs are mostly detached from the background in which
they arose And they are therefore often the actual causal variants. This observation is underlined by
the high proportion of non-synonymous significant SNPs within genes, which in most cases caused
substitution to amino acid with different properties or even premature termination. Such deviant
variants with likely substantial functional or conformational changes in the resulting proteins may be
selectively neutral or nearly neutral under ancestral benign conditions, but may become selectively
relevant under changing conditions (Paabi & Rockman 2014). Interestingly, most of the allelic
variants associated with a healthy phenotype were also the variants in the reference genome. This
might be due to the choice of the *F. sylvatica* individual from which the reference genome was
gained (Mishra *et al.* 2018). This more than 300-year-old individual is standing at a particularly dry
site on a rocky outcrop on the rim of a scarp where precipitation swiftly runs off. Trees at such sites
were likely selected for drought tolerance.

Even though the area sampled for this study was limited relative to the species distribution range, it
comprised its core area. In addition, the climatic variation covered by the sampling sites for this study
is representative for large parts of the species range (Baumbach *et al.* 2019). The relatively limited
population structure over large parts of the species range (Magri *et al.* 2006) together with the
propensity for long range gene- flow (Belmonte *et al.* 2008) suggested that the genomic variation
responsible for drought tolerance identified here is widely distributed (Lander *et al.* 2021).
Nevertheless, an assessment of the geographic distribution of the drought related genomic variants
over the entire distribution range would yield general insight into the species-wide architecture of
this important trait.

None of the genes found here was involved in a transcriptomic study on drought response in beech
saplings (Müller *et al.* 2017). However, most of the reliably annotated genes with or close to SNP loci
significantly associated with drought phenotypes had putative homologs in other plant species

previously shown to be involved in drought or different environmental stress response (for citations see Suppl. Table 2). This observation may be considered as post-hoc evidence that drought was indeed the most likely stressor causing the observed phenotypic responses. It remained unclear whether the remaining, not annotated genes had not yet been associated with drought before, or whether we were just unable to make this link them due to the lack of (ecological) annotation and standardised reporting. (Waldvogel *et al.* 2021) The involvement of in total 67 genes together with the relatively flat effect size distribution suggested that drought resistance in *F. sylvatica* is a moderately polygenic trait, which should respond well to artificial breeding attempts and natural selection. However, given the relatively strict threshold criteria, it is likely that more yet undetected loci contribute to the respective phenotypes. The low LD in beech predicts that an adaptation to drought will not compromise genome-wide genetic diversity and thus adaptation potential to other stressors. We achieved a high level of accuracy using genomic data to predict the drought phenotype from individuals not used to identify drought associated SNP loci. However, due to the small sample size, LDA might have resulted in overfitting (Hawkins 2004). We therefore also used a non-parametric machine learning algorithm that has been shown to produce more robust results, especially for small sample sizes (Horenko 2020).Both analyses confirmed that we mainly identified alleles widespread throughout the sampled range and not locally specific. Besides, we confirmed a considerable level of genetic variation in the sampled regions. The observation that trees with the highest predictive values showed no loss of heterozygosity indicated that there is still adaptive potential for drought adaptation in the species (Gienapp *et al.* 2017).With the SNP assay, we therefore created a tool that can i) support the choice of seed trees for reforestations, ii) provide decision guidance for selective logging and iii) monitor, whether natural selection on this quantitative trait is already acting in the species. The current study can also serve as a starting point for molecular and physiological research on how the identified loci or variants may, alone or in concert, confer resilience or tolerance to a range of drought stress symptoms.

Material and Methods

Sampling and phenotyping

In August/early September 2019, we sampled leaf tissue of 402 *Fagus sylvatica* trees from 32 locations in Hessen/Germany (set 1, Figure 1), of which 300 were used for the (pool)GWAS analysis. Forty three, plus additional 53 trees which were sampled in n August 2020, additional 52 trees from four sites were sampled (set 2, Figure 1) made up the confirmation set. The coordinates and characteristics of each site can be found in Suppl. Table 1. The sampling was performed in a strictly pairwise design. The pairs consisted of one tree with heavy drought damage of the crown (lost or rolled up, dried leaves) and one with an unaffected crown, respectively. This categorisation into least and most damaged trees was taken compared to the other trees in the respective forest patch. The pairs were *a priori* chosen such that the two trees were i) mutually the closest neighbours with contrasting damage status (i.e. no other tree in the direct sight-line), ii) free from apparent mechanical damage, fungal infestations or other signs of illness, similar iii) in tree height, iv) trunk circumference, v) light availability, and vi) canopy closure. In addition, each pair was situated at least 30 m from the closest forest edge. For each tree of the chosen pairs, we recorded the exact position, distance to the pair member and the estimated tree height (in 1 m increments), measured the trunk circumference at 150 cm height above the ground (in 10 cm increments), and estimated the leaf loss of the crown and the proportion of dried leaves (in 5% increments). We also recorded the estimated distance (in 1 m resolution) and the specific identity of the two closest neighbour trees for each pair member and calculated a competition index C as follows: $C = S_1/D_1 + S_2/D_2$, where S_1 and S_2 are the trunk diameter at 150 cm and D_1 and D_2 the distances of the nearest and second nearest neighbour

312 tree of the same size or larger than the focal tree. Photographs from the crown and the trunk were
313 taken from the trees sampled in 2019.

314 From each tree, we sampled 5 to 10 fully developed leaves from low branches. The leaves sampled
315 from each tree were placed in paper bags. After returning from the field, they were dried at 50°C for
316 30-90 min and then kept on salt until they could be stored at -80°C.

Climate and remote sensing data

Monthly daily mean minimum and maximum temperature values and precipitation data were
obtained for the 1 x 1 km grid cells harbouring the sampling sites for the period between 1950 and
2019. Data on the accumulated potential evapotranspiration during the growth season was obtained
for the same grid cells. The data is publicly available from
https://opendata.dwd.de/climate_environment/CDC/grids_germany/monthly/.

Leaf area index (LAI) data for the above grid cells was obtained from Copernicus remote sensing
(www.copernicus.eu) for the period 2014-2019, considering only the month of August. To see
whether drought conditions influenced leaf coverage of the woods at the sampling sites, we
calculated the relative annual deviation of LAI from the 2014 value. We correlated it to the relative
deviation of the cumulated potential evatransporation over the growth season from 2014. The year
2014 was used as a baseline, because of the significant drought increase since then (Büntgen *et al.*
2021). Please note that the absolute level of LAI depends on the wood coverage, vegetation density
and species composition of each plot. Changes in LAI are thus not exclusively due to drought
damages in beech.

DNA extraction, construction of GWAS pools and sequencing

DNA was extracted from 12.5 mm² of a single leaf from each tree following the NucleoMag Plant Kit
(Macherey Nagel, Düren, Germany) protocol. We set up four DNA pools for poolGWAS by pooling
equal amounts of DNA from each individual: damaged individuals from the Southern part (dSouth),
healthy individuals from the South (hSouth), damaged North (dNorth) and healthy North (hNorth).
The Southern pools consisted of 100 individuals each, the Northern pools of 50 individuals each. The
pools were sent to Novogene (Cambridge, UK) for library construction and 150bp paired end
sequencing with 350bp insert size with 25Gb data for the northern and 38Gb data for the southern
samples. The 100 individuals used to construct the Northern pools were also individually re-
sequenced. The exact composition of the genomic pools can be found in Supplemental Table 1. All
sequence information can be found on the European Nucleotide Archive (ENA) under project
accession number *PRJEB24056.*

Reference genome improvement

345 We used an improved version of the recently published reference genome for the European beech
346 (Mishra *et al.* 2018). Contiguity was improved to chromosome level using Hi-C reads with the help of
347 the allhic software after excluding the probable organelle backbones from the earlier assembly that
348 was generated from the Illumina-corrected PacBio reads using Canu assembler (Mishra *et al.* 2021)
349 Access. No. PRJNA450822.

Mapping and variant calling

Reads of pools and individual resequencing were trimmed using the wrapper tool autotrim v0.6.1
(Waldvogel *et al.* 2018) that integrates trimmomatic (Bolger *et al.* 2014) for trimming and fastQC
(Andrews 2010) for quality control. The trimmed reads were then mapped on the latest chromosome
level build of the *F. sylvatica* genome using the BWA mem algorithm v.0.7.17 (Li & Durbin 2009). Low

quality reads were subsequently filtered and SNPs were initially called using samtools v.1.10 (Li *et al.*
2009). A principal component analysis (PCA) was conducted on unlinked single nucleotide
polymorphisms (SNPs) using the R package Factoextra v.1.0.7 (Kassambara & Mundt 2017).

Pool GWAS and PLINK

The PoPoolation pipeline 2_2012 (Kofler *et al.* 2011a; Kofler *et al.* 2011b) was used to call SNPs and
remove indels from the four pools. Allele frequencies for all SNPs with a coverage between 15x and
100x with a minimum allele count of three were estimated with the R library PoolSeq v. 0.35 (Taus *et*
al. 2017).

The statistical test to detect significant allele frequency differences among damaged and healthy
trees was the Cochran-Mantel-Haenszel test. With this test, a 2x2 table was created for each variable
position and region with two phenotypes (healthy and damaged). The read counts of each allele for
each phenotype were treated as the dependent variables. We controlled for false discovery rate
using the Benjamini-Hochberg correction R package *p.adjust*.

For the individual resequencing data we followed the GATK-pipeline 4.1.3.0 (DePristo *et al.* 2011). In
short, Picard tools v.2.20.8 was used to mark duplicates. GVCF files were created with
HaplotypeCaller and genotyped with GEnotypeGVCFs. Since we did not have a standard SNP set we
hard filtered SNPs with VariantFiltration QD<2.0, MQ<50.0, MQRankSum<12.5,
ReadPosRankSum<8.0, FS>80.0, SOR>4.0 and QUAL<10.0. This conservative SNP-set was used for
base recalibration before running the HaplotypeCaller pipeline a second round. Finally, the
genotyped vcf-files were filtered using vcftools with --maf 0.03 --max-missing 0.9 --minQ 25 --min-
meanDP 10 --max-meanDP 50 --minDP 10 --maxDP 50. The detailed pipeline can be found in Suppl.
Info 2.

To conduct the GWAS association on the above generated SNP set with phenotypes being either
damaged or healthy and to generate a principal component analysis on the SNP positions of the
individually resequenced trees, we used PLINK 1.9 (Purcell *et al.* 2007). The detailed workflow can be
found in Suppl. Info 2. We calculated a non-parametric ANOSIM on an inter-individual Euclidean
distance matrix based on the first ten principal components to infer whether the trees within
phenotype groups are overall genetically more similar than within groups (9,999 permutations;
(Hammer *et al.* 2001).

Inference of Linkage Disequilibrium

The expected length of segregating haplotypes in a species depends on the recombination rate and
their age. The former can be approximated by an estimate of linkage disequilibrium (LD. To
determine LD decay based on individually re-sequenced data we used the software LDkit v 1.0.0
(Tang *et al.* 2020), in 1kb and 100kb windows.

Identification substitution type and gene function

We inferred whether significantly differentiated SNPs within genes lead to a (non-) synonymous
amino acid substitution using tbg-tools v0.2 (https://github.com/Croxa/tbg-tools) (Schoennenbeck *et*
al. 2021). The protein sequences of the identified genes were used in a blastp search against all non-
redundant GenBank CDS translations, PDB, SwissProt, PIR, PRF to infer potential gene functions. Only
the best BLAST-hits were considered.

Selection of SNP loci for SNPtype™ assay design

For the design of SNPtype™ assays we used the web-based D3 assay design tool (Fludigm corp.). We
aimed in first preference for the most significant SNPs of each genomic region identified by Pool-
GWAS (80 loci). If this was technically impossible and the region harboured more than a single
significant SNP, we opted for the second most significant SNP and so forth. This resulted finally in 76
suitable loci. The remaining 20 loci were recruited from the 20 most significant SNPs of the PLINK
analysis that were not scored in the Pool-GWAS.

SNP genotyping procedure

For validation of drought susceptibility associated SNPs, we conducted SNP genotyping on 96.96
Dynamic Arrays (Fluidigm) with integrated fluidic circuits (Wang *et al.* 2009, 2009). (N=96) to validate
the effectiveness of the identified SNPs in discriminating healthy from damaged trees. Prior to
genotyping PCR, DNA extracts were normalised to approximately 5-10 ng/µl. They underwent a pre-
amplification PCR (Specific Target Amplification, STA) according to the manufacturer's protocol to
enrich target loci. PCR products were diluted 1:10 with DNA suspension buffer (TEKnova, PN T0221)
before further use. Genotyping was performed according to the recommendations of manufacturer.
Four additional PCR cycles were added to accommodate for samples of lower quality or including
inhibitors (von Thaden, 2020). Fluorescent data were measured using the EP1 (Fluidigm) and
analysed with the SNP Genotyping Analysis Software version 4.1.2 (Fluidigm). The automated scoring
of the scatter plots was checked visually and if applicable, manually corrected.

Genomic prediction

To predict drought susceptibility from genotype data, we used a linear discriminant analysis (LDA) on
92 genotypes scored with the Fluidigm assay at 70 loci. Genotypes homozygous for the reference
allele were scored as 0, heterozygous as 1 and homozygous alternate alleles as 2. We used the LDA
option implemented in PAST v. 4.05. (Hammer, 2001).

We also used a non-parametric entropy-based Scalable Probabilistic Analysis framework (eSPA). This
method allows simultaneous solution of feature selection and clustering problems, meaning that
does not rely on a particular choice of user-defined parameters and has been shown to produce
more robust results, especially for small sample sizes (Gerber2020, Horenko2020). Following the
suggestion of the user manual, eSPA analysis was run 100 times with independent cross-validations
of the Area Under the Curve (AUC) on the validation data.

Acknowledgments

426 There was no external funding for this study. We thank the SBiK-F Data- and Modelling Centre (Aidin
427 Niamir) for support in obtaining climate and remote sensing data. CC acknowledges support from the
428 Emergent AI Center funded by the Carl-Zeiss-Stiftung.

Additional information

Author contributions

Markus Pfenninger: Conceptualisation, sampling, data analysis, writing – original draft. Friederike
Reuss, Angelika Kiebler: Laboratory support. Philipp Schönnenfeld, Cosima Caliendo, Susanne Gerber:
Bioinformatics support. Berardino Cocchiararo: SNP-assay. Sabrina Reuter, Nico Blüthgen, Karsten
Mody: Conceptualisation, sampling, data analysis. Miklós Bálint: Conceptualisation, data analysis.
Bagdevi Mishra, Marco Thines: Bioinformatics support. Barbara Feldmeyer: Conceptualisation,
labwork, data analysis. All authors read and approved the final version of the manuscript.

Author ORCIDs

Markus Pfenninger 0000-0002-1547-7245

Susanne Gerber 0000-0001-9513-0729

Berardino Cocchiararo 0000-0002-8427-7861

Karsten Mody 0000-0002-9251-0803

Marco Thines 0000-0002-9251-0803

Barbara Feldmeyer 0000-0002-0413-7245

Data availability

References

448   Andrews S (2010) FastQC: a quality control tool for high throughput sequence data. Babraham
449   Bioinformatics, Babraham Institute, Cambridge, United Kingdom.
450   Aranda I, Cano FJ, Gascó A*, et al.* (2015) Variation in photosynthetic performance and hydraulic
451   architecture across European beech (*Fagus sylvatica* L.) populations supports the case for
452   local adaptation to water stress. *Tree Physiology* **35**, 34-46.
453   Bastide H, Betancourt A, Nolte V*, et al.* (2013) A genome-wide, fine-scale map of natural
454   pigmentation variation in Drosophila melanogaster. *PLoS Genetics* **9**, e1003534.
455   Baumbach L, Niamir A, Hickler T, Yousefpour R (2019) Regional adaptation of European beech (*Fagus*
456   *sylvatica*) to drought in Central European conditions considering environmental suitability
457   and economic implications. *Regional Environmental Change* **19**, 1159-1174.
458   Belmonte J, Alarcón M, Avila A, Scialabba E, Pino D (2008) Long-range transport of beech (*Fagus*
459   *sylvatica* L.) pollen to Catalonia (north-eastern Spain). *International Journal Of*
460   *Biometeorology* **52**, 675-687.
461   Bolger AM, Lohse M, Usadel B (2014) Trimmomatic: a flexible trimmer for Illumina sequence data.
462   *Bioinformatics* **30**, 2114-2120.
463   Bolte A, Czajkowski T, Cocozza C*, et al.* (2016) Desiccation and mortality dynamics in seedlings of
464   different European beech (*Fagus sylvatica* L.) populations under extreme drought conditions.
465   *Frontiers in plant science* **7**, 751.
466   Bošeľa M, Sedmák R, Marušák R*, et al.* (2014) Evaluating similarity of radial increments around tree
467   stem circumference of European beech and Norway spruce from Central Europe.
468   *Geochronometria* **41**, 136-146.
469   Bressem U (2008) Komplexe Erkrankungen an Buche. Complex diseases in beech. *Ergebnisse*
470   *angewandter Forschung zur Buche* **3**, 87.
471   Brunet J, Fritz Ö, Richnau G (2010) Biodiversity in European beech forests-a review with
472   recommendations for sustainable forest management. *Ecological Bulletins*, 77-94.
473   Büntgen U, Urban O, Krusic PJ*, et al.* (2021) Recent European drought extremes beyond Common Era
474   background variability. *Nature Geoscience*, 1-7.
475   Carrière SD, Ruffault J, Cakpo CB*, et al.* (2020) Intra-specific variability in deep water extraction
476   between trees growing on a Mediterranean karst. *Journal of Hydrology* **590**, 125428.
477   Choat B, Brodribb TJ, Brodersen CR*, et al.* (2018) Triggers of tree mortality under drought. *Nature*
478   **558**, 531-539.
479   Christensen JH, Hewitson B, Busuioc A*, et al.* (2007) Regional climate projections. In: *Climate Change,*
480   *2007: The Physical Science Basis. Contribution of Working group I to the Fourth Assessment*
481   *Report of the Intergovernmental Panel on Climate Change, University Press, Cambridge,*
482   *Chapter 11*, pp. 847-940.
483   Cocozza C, De Miguel M, Pšidová E*, et al.* (2016) Variation in ecophysiological traits and drought
484   tolerance of beech (*Fagus sylvatica* L.) seedlings from different populations. *Frontiers in Plant*
485   *Science* **7**, 886.

Czajkowski T, Bolte A (2006) Frosttoleranz deutscher und polnischer Herkünfte der Buche (*Fagus*
sylvatica L.) und ihre Beeinflussung durch Trockenheit. *Archiv für Forstwesen und*
Landschaftsökologie **40**, 119-126.
DePristo MA, Banks E, Poplin R*, et al.* (2011) A framework for variation discovery and genotyping
using next-generation DNA sequencing data. *Nature Genetics* **43**, 491.
Dorow WH, Blick T, Kopelke J-P (2010) Zoologische Forschung in hessischen Naturwaldreservaten–
Exemplarische Ergebnisse und Perspektiven. *Forstarchiv* **81**, 61-68.
Elsasser P, Meyerhoff J, Weller P (2016) An updated bibliography and database on forest ecosystem
service valuation studies in Austria, Germany and Switzerland. Thünen Working Paper.
Endler L, Betancourt AJ, Nolte V, Schlötterer C (2016) Reconciling differences in pool-GWAS between
populations: a case study of female abdominal pigmentation in *Drosophila melanogaster*.
Genetics **202**, 843-855.
Etzold S, Ziemińska K, Rohner B*, et al.* (2019) One century of forest monitoring data in Switzerland
reveals species-and site-specific trends of climate-induced tree mortality. *Frontiers in Plant*
Science **10**, 307.
Gárate-Escamilla H, Hampe A, Vizcaíno-Palomar N, Robson TM, Benito Garzón M (2019) Range-wide
variation in local adaptation and phenotypic plasticity of fitness-related traits in *Fagus*
sylvatica and their implications under climate change. *Global Ecology and Biogeography* **28**,
1336-1350.
Gerber S, Pospisil L, Navandar M, Horenko I (2020). Low-cost scalable discretization, prediction, and
feature selection for complex systems. *Science Advances*, **6**, eaaw0961.
Gienapp P, Fior S, Guillaume F*, et al.* (2017) Genomic quantitative genetics to study evolution in the
wild. *Trends in Ecology & Evolution* **32**, 897-908.
Harris K, Nielsen R (2013) Inferring demographic history from a spectrum of shared haplotype
lengths. *PLoS Genet* **9**, e1003521.
Harter DE, Nagy L, Backhaus S*, et al.* (2015) A comparison of genetic diversity and phenotypic
plasticity among European beech (*Fagus sylvatica* L.) populations from Bulgaria and Germany
under drought and temperature manipulation. *International Journal of Plant Sciences* **176**,
232-244.
Hawkins DM (2004) The problem of overfitting. *Journal of chemical information and computer*
sciences **44**, 1-12.
Hoban S, Kelley JL, Lotterhos KE*, et al.* (2016) Finding the genomic basis of local adaptation: pitfalls,
practical solutions, and future directions. *The American Naturalist* **188**, 379-397.
Horenko I (2020) On a Scalable Entropic Breaching of the Overfitting Barrier for Small Data Problems
in Machine Learning. *Neural Computation* **32**, 1563-1579.
Kassambara A, Mundt F (2017) Factoextra: extract and visualize the results of multivariate data
analyses. *R package version* **1**, 337-354.
Kofler R, Orozco-terWengel P, De Maio N*, et al.* (2011a) PoPoolation: A Toolbox for Population
Genetic Analysis of Next Generation Sequencing Data from Pooled Individuals. *PLoS One* **6**,
e15925.
Kofler R, Pandey RV, Schlötterer C (2011b) PoPoolation2: identifying differentiation between
populations using sequencing of pooled DNA samples (Pool-Seq). *Bioinformatics* **27**, 3435-
3436.
Kreyling J, Thiel D, Nagy L*, et al.* (2012) Late frost sensitivity of juvenile *Fagus sylvatica* L. differs
between southern Germany and Bulgaria and depends on preceding air temperature.
European Journal of Forest Research **131**, 717-725.
Lander TA, Klein EK, Roig A, Oddou-Muratorio S (2021) Weak founder effects but significant spatial
genetic imprint of recent contraction and expansion of European beech populations.
Heredity **126**, 491-504.
Li H, Durbin R (2009) Fast and accurate short read alignment with Burrows-Wheeler Transform.
Bioinformatics **25**, 1754-1760.

Li H, Handsaker B, Wysoker A*, et al.* (2009) The Sequence alignment/map (SAM) format and
SAMtools. . *Bioinformatics* **25**, 2078-2079.
Magri D, Vendramin GG, Comps B*, et al.* (2006) A new scenario for the Quaternary history of
European beech populations: palaeobotanical evidence and genetic consequences. *New*
Phytologist **171**, 199-221.
Mishra B, Gupta DK, Pfenninger M*, et al.* (2018) A reference genome of the European beech (*Fagus*
sylvatica L.). *Gigascience* **7**, giy063.
Mishra B, Ulaszewski B, Meger J*, et al.* (2021) A chromosome-level genome assembly of the
European Beech (*Fagus sylvatica*) reveals anomalies for organelle DNA integration, repeat
content and distribution of SNPs. *bioRxiv https://doi.org/10.1101/2021.03.22.436437*.
Müller M, Seifert S, Lübbe T, Leuschner C, Finkeldey R (2017) De novo transcriptome assembly and
analysis of differential gene expression in response to drought in European beech. *PLoS One*
12, e0184167.
Paaby A, Rockman M (2014) Cryptic genetic variation: evolution's hidden substrate. *Nature Reviews*
Genetics **15**, 247–258.
Paar U, Dammann I (2019) Waldzustandsbericht Hessen 2019 (ed. Hessisches Ministerium für
Umwelt K, Landwirtschft und Verbrauscherschutz), Wiesbaden.
Pluess AR, Frank A, Heiri C*, et al.* (2016) Genome–environment association study suggests local
adaptation to climate at the regional scale in *Fagus sylvatica*. *New Phytologist* **210**, 589-601.
Pretzsch H, Schütze G, Uhl E (2013) Resistance of European tree species to drought stress in mixed
versus pure forests: evidence of stress release by inter-specific facilitation. *Plant Biology* **15**,
483-495.
Purcell S, Neale B, Todd-Brown K*, et al.* (2007) PLINK: a tool set for whole-genome association and
population-based linkage analyses. *The American Journal of Human Genetics* **81**, 559-575.
Rajendra K, Seifert S, Prinz K, Gailing O, Finkeldey R (2014) Subtle human impacts on neutral genetic
diversity and spatial patterns of genetic variation in European beech (*Fagus sylvatica*). *Forest*
Ecology and Management **319**, 138-149.
Schoennenbeck P, Schell T, Gerber S, Pfenninger M (2021) tbg-a new file format for genomic data.
bioRxiv https://doi.org/10.1101/2021.03.15.435393.
Schuldt B, Buras A, Arend M*, et al.* (2020a) A first assessment of the impact of the extreme 2018
summer drought on Central European forests. *Basic and Applied Ecology* **45**, 86-103.
Schuldt B, Buras A, Arend M*, et al.* (2020b) A first assessment of the impact of the extreme 2018
summer drought on Central European forests. *Basic and Applied Ecology*.
Stocks JJ, Metheringham CL, Plumb WJ*, et al.* (2019) Genomic basis of European ash tree resistance
to ash dieback fungus. *Nature Ecology & Evolution* **3**, 1686-1696.
Sutmöller J, Spellmann H, Fiebiger C, Albert M (2008) Der Klimawandel und seine Auswirkungen auf
die Buchenwälder in Deutschland The effects of climate change on beech forests in Germany.
Ergeb. Angew. Forsch. Buche **3**, 135.
Tang Y, Li Z, Wang C*, et al.* (2020) LDkit: a parallel computing toolkit for linkage disequilibrium
analysis. *BMC Bioinformatics* **21**, 1-8.
Taus T, Futschik A, Schlötterer C (2017) Quantifying selection with pool-seq time series data.
Molecular Biology and Evolution **34**, 3023-3034.
Thünen_Institute (2020) Clusterstatistik Forst & Holz.
Trenberth KE, Dai A, Van Der Schrier G*, et al.* (2014) Global warming and changes in drought. *Nature*
Climate Change **4**, 17-22.
Waldvogel AM, Feldmeyer B, Rolshausen G*, et al.* (2020) Evolutionary genomics can improve
prediction of species' responses to climate change. *Evolution Letters* **4**, 4-18.
Waldvogel A-M, Schreiber D, Pfenninger M, Feldmeyer B (2021) Climate Change Genomics Calls for
Standardized Data Reporting. *Coping with Climate Change: A Genomic Perspective on*
Thermal Adaptation.
Waldvogel AM, Wieser A, Schell T*, et al.* (2018) The genomic footprint of climate adaptation in
Chironomus riparius. *Molecular Ecology* **27**, 1439-1456.

Wang J, Lin M, Crenshaw A, *et al.* (2009) High-throughput single nucleotide polymorphism
genotyping using nanofluidic Dynamic Arrays. *BMC Genomics* **10**, 561.
Wellenreuther M, Hansson B (2016) Detecting polygenic evolution: problems, pitfalls, and promises.
Trends in Genetics **32**, 155-164.

Figures

Figure 1. A) Locations of sampling sites in Hessen, Germany. For abbreviations see Suppl. Table 1. The
sites where confirmation individuals were sampled are designated in grey.B) Principal Component
Analysis of monthly climate data 1950-2019, C) Development of main growth period drought
indicator from 1991-2019. Shown is the difference mean monthly evaporation potential in mm from
April to September relative to the 1991 level. Climate and drought data obtained from
https://opendata.dwd.de/climate_environment/CDC/grids_germany/monthly/.

603 Figure 2. Comparison of sampled beech pairs. A) trunk circumference, B) canopy closure, C) tree
604 height, D) competition index, E) dried leaves and F) leaf loss. Box-plots with indicated means, the
605 boxes represent one standard deviation, the whiskers are the 95% confidence intervals. Damaged
606 trees in ochre, healthy trees in green. Except for E and F, the difference of means among damaged
607 and healthy trees is insignificant between the groups.

Figure 3. Genome wide linkage disequilibrium and principal component analysis on genome-wide SNP data. A) Decay of genome wide linkage disequilibrium (LD), measured as r² on allele frequencies gained from individual resequencing, with distance from focal SNP in base pairs. B) Plot of the first two principal component axes of LD pruned SNP data from individually sequenced beech individuals

from the North population. Healthy trees are indicated by a green dot, damaged ones by ochre.
Individuals sampled from the same site are grouped by convex hulls, limited with dotted lines.

Figure 4. Significantly associated drought phenotype associated SNP marker. A) Manhattan plot of
false discovery rate (FDR) corrected –log10 probability values from CMH test. The black horizontal
line indicates the chosen significance threshold. SNPs on different chromosomes alternate in colour
(black and blue). B) Mean allele frequency difference at significantly associated SNP loci between
healthy and damaged phenotypes. The loci are ordered according to amount of change.

Figure 5. Combined results of SNP assay and discriminant analyses. The centre of the figure depicts the genotypogram of the SNP assay. Each column represents one of 70 loci, each row one of 92 beech individuals. The scored genotypes are colour-coded, with red squares = homozygous reference allele, light blue = homozygous alternate allele, white = heterozygous SNP, grey squares = locus could not be scored in the respective individual. The left bar indicates the observed phenotype for each tree individual with ochre rectangles for damaged, and green for healthy trees. Below the genotypogram, the relative contribution of each locus to the predictive model of the discriminant analysis is indicated, ordered from high to low. On the right side, first the genotype model scores for each individual are given, with the according predicted phenotype (ochre = damaged; green = healthy).

Tables

Table 1. Genes with significantly associated SNPs. Given are the chromosome number (CHR), nucleotide position (position), the gene ID for *Fagus sylvatica* (gene), the UniProt ID of the closest match (UniProt ID), the name of the gene (name), the nucleotide base in the reference (ref DNA base), and the alternate base (alt DNA base), if applicable, the amino acid of the reference (ref AA) and the alternate base (non-synonymous change), functional change (effect) and the phenotype associated with the alternate base.

CHR	position	gene	UniProt ID	name	ref DNA base	alt DNA base	ref AA	non-synoynmous change	effect	phenotype assoc. with alt base
1	40374762	1.g3851.t1	none		A	G	C	R	SH side chain > positive charge	healthy
10	32290645	10.g3914.t1	none		C	T	F	-	-	
11	20479628	11.g2467.t1	EXOS5_ORYSJ	Exosome complex exonuclease RRP46 homolog	T	C	T	A	polar > hydrophobic	damaged
11	23722307	11.g2832.t1	PCN_ARATH	WD repeat-containing protein PCN	A	G	I	V	hydrophobic > hydrophobic	damaged
12	13901034	12.g1695.t1	F4I5S1_ARATH	PB1 domain-containing protein tyrosine kinase	C	A	P	Q	hydrophobic > polar	damaged
	13901063				C	T	Q	stop	termination	damaged
	13901082				A	T	H	L	positive charge > hydrophobic	damaged
	13901094				T	A	I	N	hydrophobic > polar	damaged
2	43326571	2.g4736.t1	none		G	A	R	C	positive charge > SH side chain	damaged
3	31226940	3.g3590.t1	none		C	T	Q	stop	termination	healthy
4	34077017	4.g3980.t1	CKX1_ARATH	Cytokinin dehydrogenase 1	C	A	G	C	no side chain > SH side chain	damaged
5	16359587	5.g1807.t1	GDI2_ARATH	Guanosine nucleotide diphosphate dissociation inhibitor 2	T	C	P	-	-	
6	19865311	6.g2227.t1	NDUS7_ARATH	NADH dehydrogenase [ubiquinone] iron-sulfur protein 7, mitochondrial	T	C	D	-	-	
6	26383172	6.g2921.t1	none		T	C	A	-	-	
7	1493904	7.g177.t1	TLP10_ARATH	Tubby-like F-box protein 10	C	T	G	-	-	
7	20242023	7.g2350.t1	PRK4_ARATH	Pollen receptor-like kinase 4	A	G	P	-	-	
7	4504799	7.g1655			C	T	W	stop	termination	damaged

7	31456694	7.g3617.t1	LSH4_ARATH	Protein LIGHT-DEPENDENT SHORT HYPOCOTYLS 4	G	T	R	-	-	
7	33110000	7.g3816.t1	none		G	A	L	-	-	
7	4504813	7.g552.t1	none		G	A	G	-	-	
	4504831				C	T	L	-	-	
8	29295139	8.g3494.t1	VATC_ARATH	V-type proton ATPase subunit C	G	A	G	-	-	
9	25538827	9.g3080.t1	PPA14_ARATH	Probable inactive purple acid phosphatase 14	G	C	K	N	positive charge > polar	damaged
9	37955715	9.g4504.t1	TBL33_ARATH	Protein trichome birefringence-like 33	G	C	M	I	hydrophobic > hydrophobic	damaged