bioRxiv preprint doi: https://doi.org/10.1101/2020.12.06.412759; this version posted December 7, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

1 The new generation hDHODH inhibitor MEDS433 hinders the in vitro

2 replication of SARS-CoV-2

4	Arianna Calistri ^a , Anna Luganini ^b , Valeria Conciatori ^a , Claudia Del Vecchio ^a , Stefano
5	Sainas ^c , Donatella Boschi ^c , Marco Lucio Lolli ^c , Giorgio Gribaudo ^{b#} , Cristina Parolin ^{a#}
6	
7	
8	^a Department of Molecular Medicine, University of Padua, 35121 Padua, Italy.
9	^b Department of Life Sciences and Systems Biology, University of Turin, 10123 Turin, Italy.
10	^c Department of Sciences and Drug Technology, University of Turin, 10125 Turin, Italy.
11	
12	
13	[#] Corresponding Authors: Giorgio Gribaudo (giorgio.gribaudo@unito.it), Cristina Parolin
14	(cristina.parolin@unipd.it)
15	
16	
17	
18	Running title: MEDS433 inhibits SARS-CoV-2 replication
19	
20	
21	Word count: abstract 74; main text: 1013.
22	
23	
24	

- 25
- 26 Abstract

Identification and development of effective drugs active against SARS-CoV-2 are
urgently needed. Here, we report on the anti-SARS-CoV-2 activity of MEDS433, a novel
inhibitor of human dihydroorotate dehydrogenase (hDHODH), a key cellular enzyme of the *de novo* pyrimidines biosynthesis. MEDS433 inhibits *in vitro* virus replication in the low
nanomolar range, and through a mechanism that stems from its ability to block hDHODH
activity. MEDS433 thus represents an attractive candidate to develop novel anti-SARSCoV-2 agents.

34

35 Main text

36 The emergence of the novel Severe Acute Respiratory Syndrome Coronavirus 2 37 (SARS-CoV-2), and the rapid worldwide spreading of coronavirus disease 19 (COVID-19) 38 have produced a threat to global public health that calls for urgent deployment of effective 39 antiviral drugs (1-4). Among the therapeutic options that have been potentially considered, 40 small-molecules targeting host factors exploited by SARS-CoV-2 to replicate may 41 represent an alternative to direct acting agents prone to select drug resistant strains (5). 42 One of the cellular pathways that is attracting more attention for the advancement of host-43 targeting antivirals (HTA), is the *de novo* pyrimidines biosynthesis, essential for virus 44 replication in infected cells (6). In this pathway, the human dihydroorotate dehydrogenase 45 (hDHODH) catalyzes the rate-limiting step of dehydrogenation of dihydroorotate to orotate, 46 thus providing uridine and cytidine to fulfill nucleotides request (7-9). Given its critical role, 47 hDHODH is considered an emerging target of choice for the development of HTA against 48 SARS-CoV-2 (10). In this regard, two potent hDHODH inhibitors just entered in Phase II 49 clinical trials for COVID-19: brequinar (11) (NCT04425252), and PTC299 (12) 50 (NCT04439071). However, these drug candidates suffer of toxicity issues that slowed 51 down their earlier clinical pathway on other therapeutic applications (13, 14). Thus, new 52 safest hDHODH inhibitors are urgently needed.

53 To investigate the feasibility of targeting hDHODH to develop HTA against SARS-CoV-54 2, in this study we have characterized the *in vitro* antiviral activity of a new generation 55 hDHODH inhibitor produced by the rational modulation to brequinar (11), and 56 characterized by the presence of a 2-hydroxypyrazolo[1,5-a]pyridine moiety. This 57 compound, MEDS433 (Fig. 1A), is comparable to brequinar in inhibiting hDHODH activity, 58 while owing a better drug-like profile (15, 16). 59 The effect of MEDS433 on SARS-CoV-2 replication was evaluated in Vero E6 cells 60 infected with a clinical isolate of SARS-CoV-2 (2019-nCoV/Italy-INMI1) at a multiplicity of 61 infection (MOI) of 0.1, and then treated with 0.5 µM MEDS433. At 24 hours post-infection 62 (h p.i.), cells were fixed, permeabilized and stained with an anti-SARS-CoV-2 nucleocapsid 63 N protein mAb, and with DRAQ5 which stains cell nuclei, to count cell numbers. As shown 64 in Fig. 1B, confocal microscopy revealed that while about 85% of infected control cells 65 expressed the N protein, MEDS433 treatment completely abolished its accumulation, thus 66 indicating that N protein expression could be prevented by targeting the *de novo* 67 pyrimidine biosynthesis. 68 Next, a virus yield reduction assays (VRA) was performed in SARS-CoV-2-infected 69 Vero E6 cells treated with increasing concentrations of MEDS433. At 48 h p.i., cell 70 supernatants were harvested and titrated by plaque assay. A concentration-dependent 71 inhibition of SARS-CoV-2 replication was thus observed (Fig.1C), with EC_{50} and EC_{90} 72 values of 0.063 and 0.136 µM, respectively. Interestingly, MEDS433 was more effective 73 than brequinar (EC₅₀ 0.20, EC₉₀ 1 μ M) (Fig. 1C). In addition, the Cytotoxic Concentration 74 (CC_{50}) of MEDS433 as measured in uninfected Vero E6 cells by the MTT method (17), 75 was more than 500 μ M with a favorable Selective Index (SI) greater than 7,900, thus 76 indicating that its antiviral activity was not due to a reduced cell viability. 77 To get more insights into MEDS433 mechanism of action, time-of-addition 78 experiments were carried out. Briefly, Vero E6 cells were exposed to MEDS433 (0.5 μ M)

79 from -2 to - 1h prior to SARS-CoV-2 adsorption (MOI of 0.1) (pre-treatment); during 80 infection (adsorption stage, from -1 to 0 h) (co-treatment); or after viral adsorption (from 0 81 to 48 h p.i.) (post-treatment). Infectious SARS-CoV-2 particles were then quantified in cell 82 supernatants harvested at 48 h p.i. by plague assay. As depicted in Fig. 2A, MEDS433 did 83 not affect the initial attachment and entry phases of the SARS-CoV-2 life cycle, while it 84 produced a significant reduction of infectious virus production when added at a post-entry 85 stage, in agreement with its ability to block N protein accumulation (Fig. 1B). Immunoblot 86 analysis of total protein extracts prepared from the corresponding SARS-CoV-2-infected-87 and MEDS433-treated cells and fractionated through a 10% SDS-PAGE, confirmed a 88 reduction of N protein content only in the post-treatment sample (Fig. 2B), thus indicating 89 that MEDS433 interferes with a post-entry biosynthetic step in SARS-CoV-2 replication. 90 These results suggested a mechanism of the anti-SARS-CoV-2 activity of 91 MEDS433 consistent with the hypothesis of an interference with the *de novo* pyrimidine 92 biosynthesis. To verify this hypothesis, we investigated by plague reduction assays (PRA) 93 in Vero E6 cells whether the antiviral activity of MEDS433 could be overcome by 94 supplementing cell medium with increasing concentrations of exogenous uridine to bypass 95 the requirement of *de novo* pyrimidine biosynthesis. As shown in Fig. 3 (upper panel), the 96 anti-SARS-CoV-2 activity of 0.3 μ M MEDS443 was significantly reversed by a 100-fold 97 excess of uridine relative to MEDS433 concentration, and completely overturned by 98 greater uridine concentrations, thus confirming that the *de novo* pyrimidine pathway was 99 inhibited by MEDS433 in SARS-CoV-2-infected cells. Then, to conclusively prove that 100 hDHODH inhibition was responsible of MEDS433 antiviral effect, increasing 101 concentrations of the hDHODH substrate dihydroorotic acid or its product, orotic acid were 102 added to cell medium. In SARS-CoV-2-infected Vero E6 cells treated with MEDS433 (0.3 103 µM), the addition of orotic acid reversed in a dose dependent manner the antiviral effect of 104 MEDS433 (Fig. 3, lower panel), with complete reversion observed at the highest

concentration (1000 x the MEDS433 concentration) evaluated. In contrast, the supplement
of dihydroorotic acid, even at 1 mM (3,333 times more than MEDS433), did not affect
MEDS433 antiviral activity (Fig. 3, lower panel), thus indicating that MEDS433 inhibited a
step in the *de novo* pyrimidine biosynthesis pathway downstream from dihydroorotic acid.
Altogether, these results clearly confirmed that MEDS433 specifically targets hDHODH
activity in SARS-CoV-2-infected cells, and that this inhibition is responsible of its overall
antiviral activity.

In conclusion, our study while confirming along with others recent reports (12,18) hDHODH as a noteworthy target to inhibit SARS-CoV-2 infection (10,19), highlights MEDS433 as an attractive candidate to develop HTA for COVID-19. MEDS433 in fact performs better than brequinar for both antiviral potency and SI (this study and 16), and its safety profile is superior to that of PTC299 (12). Therefore, the potent *in vitro* anti-SARS-CoV-2 activity of MEDS433 and its valuable *drug-like* profile, support further studies to validate its therapeutic efficacy in preclinical animal models of COVID-19.

119

120 Acknowledgments

- 121 This work was supported by Italian Ministry for Universities and Scientific Research
- 122 (Research Programs of Significant National Interest, PRIN 2017–2020, Grant No.
- 123 2017HWPZZZ_002) to AL.; Ministero degli Affari Esteri e della Cooperazione
- 124 Internazionale (Grant number PGR01071 Italia/Svezia (MIUR/MAECI)) to D.B. and M.L.L.;
- 125 Associazione Italiana per la Ricerca sul Cancro (AIRC) Individual Grant 2019 (AIRC IG
- 126 2019 DIORAMA 23344) to D.B. and M.L.L.; the University of Torino (Ricerca Locale) to
- 127 D.B., G.G., M.L.L. and A.L.; the University of Padua (DOR) to A.C. and C.P.;
- 128 PARO_FINA20_01 to C.P.; and BIRD grant CALI_SID19_08 to A.C. This study was also
- 129 supported by the European Virus Archive goes Global (EVAg) project that has received

- 130 funding from the European Union's Horizon 2020 research and innovation programme
- 131 under grant agreement No 653316.
- 132

133 **References**

- 134 1. Salata C, Calistri A, Parolin C, Palù G. 2019. Coronaviruses: a paradigm of new
- emerging zoonotic diseases. Pathog Dis 77:ftaa006.
- 136 2. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, Zhao X, Huang B, Shi W, Lu R, Niu
- 137 P, Zhan F, Ma X, Wang D, Xu W, Wu G, Gao GF, Tan W, for the China Novel
- 138 Coronavirus Investigating and Research Team. 2020. A novel coronavirus from
- patients with pneumonia in China. N Engl J Med 382:727-733.
- 140 3. Hu B, Guo H, Zhou P, Shi Z-L. 2020. Characteristics of SARS-CoV-2 and COVID-19.
- 141 Nat Rev Microbiol 2020 Oct 6; 1-14 doi:10.1038/s41579-020-00459-7.
- 142 4. https://covid19.who.int.
- 143 5. Li G, De Clercq E. 2020. Therapeutic options for the 2019 novel coronavirus (2019-
- 144 nCoV). Nat Rev Drug Discov 19:149-150.
- 145 6. Okesli A, Khosla C, Bassik MC. 2017. Human pyrimidine nucleotide biosynthesis as a
- target for antiviral chemotherapy. Curr Op Biotech 48:127-134.
- 147 7. Reis RAG, Calil FA, Feliciano PR, Pinheiro MP, Nonato MC. 2017. The dihydroorotate
- dehydrogenases: past and present. Arch Biochem Biophys 632:175-191.
- 149 8. Loeffler M, Carrey EA, Knecht W. 2020. The pathway to pyrimidines: the essential
- 150 focus on dihydroorotate dehydrogenase, the mitochondrial enzyme coupled to the
- respiratory chain. Nucleosides Nucleotides Nucleic Acids 11:1-25.
- 152 9. Boschi D, Pippione AC, Sainas S, Lolli ML. 2019. Dihydroorotate dehydrogenase
- inhibitors in anti-infective drug research. Eur J Med Chem 183:111681.
- 154 10. Coehlo AR, Oliveira PJ. 2020. Dihydroorotate dehydrogenase inhibitors in SARS-CoV-
- 155 2 infection. Eur J Clin Invest 50:e13366.

156 11. Peters GJ. 2018. Re-evaluation of Brequinar sodium, a dihydroorotate dehydrogenase

- 157 inhibitor. Nucleosides Nucleotides Nucleic Acids 37:666-678.
- 158 12. Luban J, Sattler R, Mühlberger E, Graci JD, Cao L, Weetall M, Trotta C, Colacino JM,
- Bavari S, Strambio-De-Castillia C, Suder EL, Wang Y, Soloveva V, Cintron-Lue K,
- 160 Naryshkin NA, Pykett M, Welch EM, O'Keefe K, Kong R, Goodwin E, Jacobson A,
- 161 Paessler S, Peltz S. 2020. The DHODH inhibitor PTC299 arrests SARS-CoV-2
- replication and suppresses induction of inflammatory cytokines. Virus Res 2020 Nov.
- 163 26, 198246, doi:10.1016/j.virusres.2020.198246.
- 164 13. Natale R, Wheeler R, Moore M, Dallaire B, Lynch W, Carlson R, Grillo-Lopez A,
- 165 Gyves L. 1992. Multicenter phase II trial of brequinar sodium in patients with advanced
- 166 melanoma. Ann Oncol 3:659-660.
- 167 14. Emvododstat PTC Therapeutics. AdisInsight Drugs [Released 2006 May 17; Updated
- 168 2020 Aug. 11], https://adisinsight.springer.com/drugs/800024409
- 169 15. Sainas S, Pippione AC, Lupino E, Giorgis M, Circosta P, Gaidano V, Goyal P, Bonanni
- D, Rolando B, Cignetti A, Ducime A, Andersson M, Järvå M, Friemann R, Piccinini M,
- 171 Ramondetti C, Buccinnà B, Al-Karadaghi S, Boschi D, Saglio G, Lolli ML. 2018.
- 172 Targeting myeloid differentiation using potent 2-Hydroxypyrazolo [1,5- a] pyridine
- 173 scaffold-based human dihydroorotate dehydrogenase inhibitors. J Med Chem

174 61:6034-6055.

175 16. Sainas S, Giorgis M, Circosta P, Gaidano V, Bonanni D, Pippione AC, Bagnati R,

- 176 Passoni A, Qiu Y, Cojocaru CF, Canepa B, Bona A, Rolando B, Mishina M,
- 177 Ramondetti R, Buccinnà B, Piccinini M, Houshmand M, Cignetti A, Giraudo E, Al-
- 178 Karadaghi S, Boschi B, Saglio G, Lolli ML. 2020. Targeting acute myelogenous
- 179 leukemia using potent human dihydroorotate dehydrogenase inhibitors based on the
- 180 2-hydroxypyrazolo[1,5-a]pyridine scaffold: SAR of the biphenyl moiety. J Med Chem,
- in press.

182 17. Mosmann T. 1983. Rapid colorimetric assay for cellular growth and survival:

- application to proliferation and cytotoxicity assays. J Immunol Methods 65:55-63.
- 184 18. Xiong R, Zhang L, Li S, Sun Y, Ding M, Wang Y, Zhao Y, Wu Y, Shang W, Jiang X,
- 185 Shan J, Shen Z, Tong Y, Xu L, Chen Y, Liu Y, Zou G, Lavillete D, Zhao Z, Wang R,
- 186 Zhu L, Xiao G, Lan K, Li H, Xu K. 2020. Novel and potent inhibitors targeting DHODH
- are broad-spectrum antivirals against RNA viruses including newly-emerged
- 188 coronavirus SARS-CoV-2. Protein Cell 11:723-739.
- 189 19. Xu Y, Jiang H. 2020. Potential treatment of COVID-19 by inhibitors of human
- dihydroorotate dehydrogenase. Protein Cell 11:699-702.
- 191

192 Figure Legends

193 Figure 1. Antiviral activity of MEDS433 on SARS-CoV-2 replication. (A) Structure of

194 MEDS433. (B) Immunofluorescence analysis of SARS-CoV-2-infected cells. Vero E6 cells

195 were treated with vehicle (DMSO) or with 0.5 μM MEDS433 1 h prior to infection with

196 SARS-CoV-2 at an MOI of 0.1. At 24 h p.i., cells were fixed, permeabilized, and

197 immunostained with an anti-SARS-CoV-2 nucleocapsid protein (N) mAb, followed by Alexa

198 488-conjugated secondary antibody. Nuclei were stained with DRAQ5. Confocal laser

199 microscopy images acquired in the green (SARS-CoV-2 N) and the blue (DRAQ5)

channels are shown, as well as overlaid images (merge). (C) Dose dependent inhibition of

201 SARS-CoV-2 replication by MEDS433. Vero E6 cell monolayers were infected with SARS-

202 CoV-2 (50 PFU/well), and, where indicated, the cells were treated with vehicle (DMSO) or

- 203 increasing concentrations of MEDS433 or brequinar 1 h before, during virus adsorption,
- and throughout the experiment. At 48 h p.i., infectious SARS-CoV-2 in cell supernatants
- was titrated by plaque assay on Vero E6 cells. MEDS433 and brequinar concentrations
- producing 50 and 90% reductions in plaque formation (EC₅₀ and EC₉₀, respectively) were

207 determined as compared to control treatment (DMSO). The data shown represent means

± SD (error bars) of three independent experiments performed in triplicate.

209 Figure 2. MEDS433 targets a post-entry stage in the SARS-CoV-2 replicative cycle.

210 (A) Time-of-addition experiment. Vero E6 cells were incubated with vehicle (DMSO) or

with 0.5 μM MEDS433 from 2 to - 1h prior to SARS-CoV-2 infection (MOI of 0.1) (pre-

treatment, Pre-T); during infection (from -1 to 0 h) (co-treatment, Co-T); or after viral

infection (from 0 to 48 h p.i.) (post-treatment, Post-T). Thereafter, production of infectious

SARS-CoV-2 was measured by titrating cell supernatants by plaque assay on Vero E6

cells. The data shown represent means ± SD (error bars) of three independent

experiments performed in triplicate. Statistical significance was calculated by a one-way

ANOVA followed by Dunnett's multiple comparison test. *** (p < 0.0001) compared to the

218 calibrator sample (MEDS433 alone).

219 (B) Immunoblot analysis of SARS-CoV-2 nucleocapsid protein. Total protein extracts

220 prepared from SARS-CoV-2-infected Vero E6 cells monolayers treated as described in

221 (A), were fractionated through a 10% SDS-PAGE, and immunoblotted with an anti-N

222 protein mAb. Tubulin was immunodetected as protein loading control.

Figure 3. Uridine or orotic acid supplementation counteracts the anti-SARS-CoV2

activity of MEDS433. Vero E6 cell were treated with solvent (DMSO) or 0.3 μ M of

225 MEDS433 in the absence or presence of increasing concentrations of uridine (upper

226 panel), orotic acid or dihydroorotic acid (lower panel) before and during infection with

227 SARS-CoV-2 (30 PFU/well). Following virus adsorption, compounds were added to cell

228 monolayers and viral plaques were then stained and were microscopically counted at 48 h

p.i.. Plaque counts for each drug concentration were expressed as a percent of the mean

230 count of the control cultures treated with DMSO. The data shown represent means ± SD of

231 three independent experiments performed in triplicate. Statistical significance was

232 calculated by a one-way ANOVA followed by Dunnett's multiple comparison test. ** (p <

- 233 0.0001), ** (p < 0.001) and * (p < 0.05) compared to the calibrator sample (MEDS433
- 234 alone).

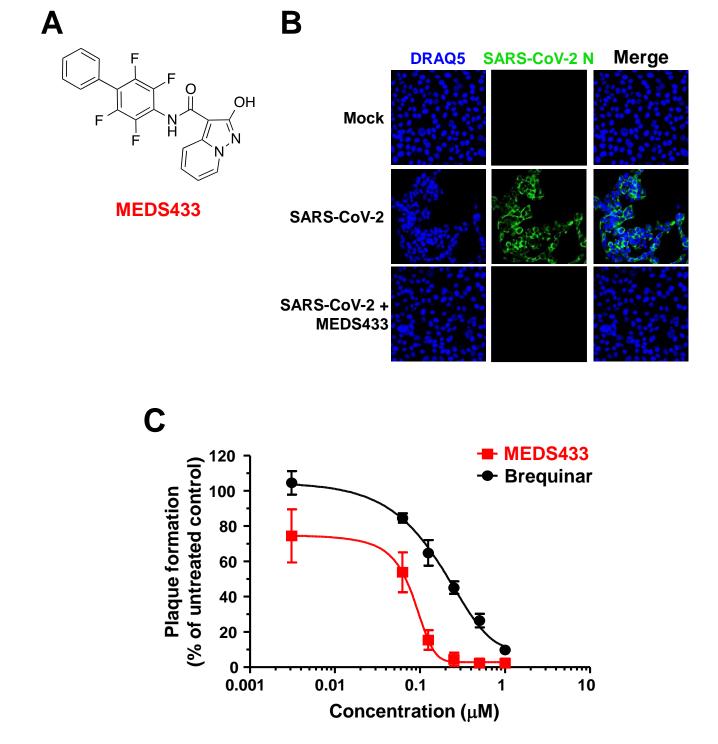


Figure 1. Antiviral activity of MEDS433 on SARS-CoV-2 replication. (A) Structure of MEDS433. (B) Immunofluorescence analysis of SARS-CoV-2-infected cells. Vero E6 cells were treated with vehicle (DMSO) or with 0.5 μ M MEDS433 1 h prior to infection with SARS-CoV-2 at an MOI of 0.1. At 24 h p.i., cells were fixed, permeabilized, and immunostained with an anti-SARS-CoV-2 nucleocapsid protein (N) mAb, followed by Alexa 488-conjugated secondary antibody. Nuclei were stained with DRAQ5. Confocal laser microscopy images acquired in the green (SARS-CoV-2 N) and the blue (DRAQ5) channels are shown, as well as overlaid images (merge). (C) Dose dependent inhibition of SARS-CoV-2 replication by MEDS433. Vero E6 cell monolayers were infected with SARS-CoV-2 (50 PFU/well), and, where indicated, the cells were treated with vehicle (DMSO) or increasing concentrations of MEDS433 or brequinar 1 h before, during virus adsorption, and throughout the experiment. At 48 h p.i., infectious SARS-CoV-2 in cell supernatants was titrated by plaque assay on Vero E6 cells. MEDS433 and brequinar concentrations producing 50 and 90% reductions in plaque formation (EC₅₀ and EC₉₀, respectively) were determined as compared to control treatment (DMSO). The data shown represent means ± SD (error bars) of three independent experiments performed in triplicate.

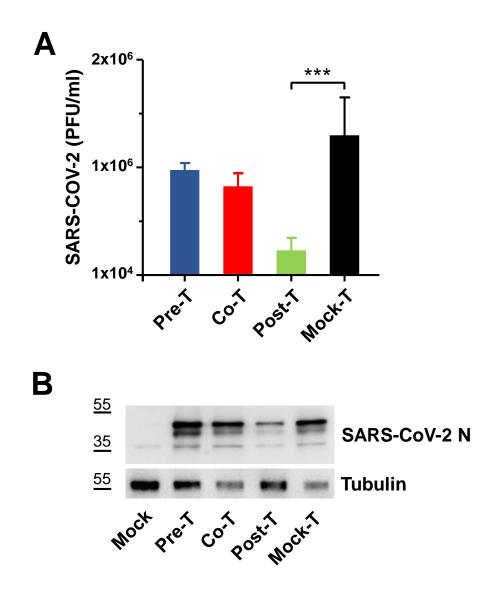


Figure 2. MEDS433 targets a post-entry stage in the SARS-CoV-2 replicative cycle. (A) Time-of-addition experiment. Vero E6 cells were incubated with vehicle (DMSO) or with 0.5 μ M MEDS433 from 2 to - 1h prior to SARS-CoV-2 infection (MOI of 0.1) (pre-treatment, Pre-T); during infection (from -1 to 0 h) (co-treatment, Co-T); or after viral infection (from 0 to 48 h p.i.) (post-treatment, Post-T). Thereafter, production of infectious SARS-CoV-2 was measured by titrating cell supernatants by plaque assay on Vero E6 cells. The data shown represent means ± SD (error bars) of three independent experiments performed in triplicate. Statistical significance was calculated by a one-way ANOVA followed by Dunnett's multiple comparison test. *** (p < 0.0001) compared to the calibrator sample (MEDS433 alone).

(B) Immunoblot analysis of SARS-CoV-2 nucleocapsid protein. Total protein extracts prepared from SARS-CoV-2infected Vero E6 cells monolayers treated as described in (A), were fractionated through a 10% SDS-PAGE, and immunoblotted with an anti-N protein mAb. Tubulin was immunodetected as protein loading control.

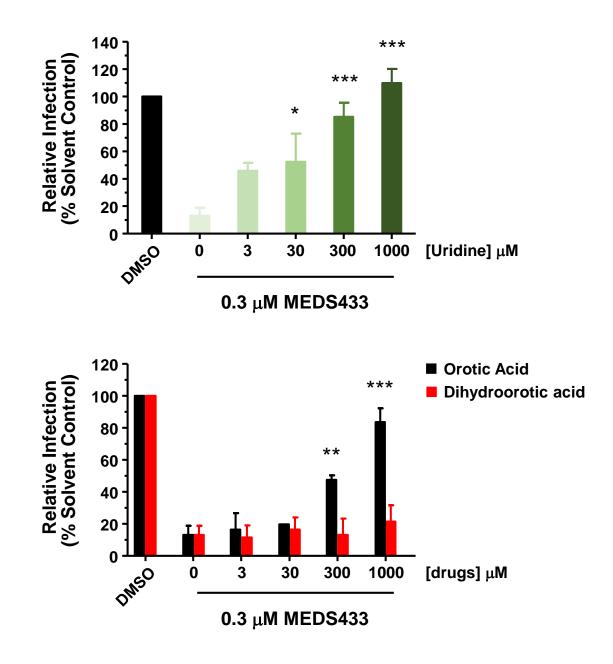


Figure 3. Uridine or orotic acid supplementation counteracts the anti-SARS-CoV2 activity of MEDS433. Vero E6 cell were treated with solvent (DMSO) or 0.3 μ M of MEDS433 in the absence or presence of increasing concentrations of uridine (upper panel), orotic acid or dihydroorotic acid (lower panel) before and during infection with SARS-CoV-2 (30 PFU/well). Following virus adsorption, compounds were added to cell monolayers and viral plaques were then stained and were microscopically counted at 48 h p.i.. Plaque counts for each drug concentration were expressed as a percent of the mean count of the control cultures treated with DMSO. The data shown represent means ± SD of three independent experiments performed in triplicate. Statistical significance was calculated by a one-way ANOVA followed by Dunnett's multiple comparison test. ** (p < 0.0001), ** (p < 0.001) and * (p < 0.05) compared to the calibrator sample (MEDS433 alone).