Easyreporting simplifies the implementation of Reproducible Research Layers in R software

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Abstract

During last years "irreproducibility" became a general problem in omics data analysis due to the use of sophisticated and poorly described computational procedures. For avoiding misleading results, it is necessary to inspect and reproduce the entire data analysis as a unified product. Reproducible Research (RR) provides general guidelines for public access to the analytic data and related analysis code combined with natural language documentation, allowing third-parties to reproduce the findings. We developed *easyreporting*, a novel R/Bioconductor package, to facilitate the implementation of an RR layer inside reports/tools without requiring any knowledge of the R Markdown language. We describe the main functionalities and illustrate how to create an analysis report using a typical case study concerning the analysis of RNA-seq data. Then, we also show how to trace R functions automatically. Thanks to this latter feature, *easyreporting* results beneficial for developers to implement procedures that automatically keep track of the analysis steps within Graphical User Interfaces (GUIs). *Easyreporting* can be useful in supporting the reproducibility of any data analysis project and the implementation of GUIs. It turns out to be very helpful in bioinformatics, where the complexity of the analyses makes it extremely difficult to trace all the steps and parameters used in the study.

Introduction

Due to accidental mistakes or misusage of sophisticated computational methods, many research findings in omics science are considered false (or partially incorrect) [1]. Moreover, in several cases, published results are not entirely reproducible due to the lack of information. For example, the analysis of the massive amount of omics data produced by high-throughput technologies requires combining several different methodologies from the preprocessing, data cleaning, and normalization to the downstream analysis. Therefore, it becomes challenging to trace all the steps and the parameters used within a complete analysis. Consequently, the lack of details, such as user-parameter or subtle data manipulation made with small code lines not reported in the material and methods sections of a manuscript, can lead to findings that are not reproducible. To prevent misleading results, several authors suggested adopting some best practices [2–4] that should help in publishing reproducible results. Nevertheless, the proposed approaches can be time-consuming and require significant effort by researchers. Therefore, to fully

exploit the advantages of Reproducible Research (RR), it is still necessary to provide tools that can trace all the details using automatic procedures [5,6].

Recently, the scientific community proposed several approaches to support RR by developing tools that require a lower cost in terms of time and efforts to be used [5–9]. Among the different approaches, one common idea is to describe the steps with an analysis report built up as a mixture of natural language sentences along with computational language and graphical outputs. This document should include: i) the analyzed data, ii) the Code Chunks (CCs), iii) results and intermediate outputs (as tables and figures), and iv) all information that can enhance the work comprehensibility and reproducibility. Using human-readable reports instead of other procedures, such as dockers, has the advantage that the final document can be easily understood by non-expert users, whereas dockers require computationally experienced users. Moreover, a human-readable report can be enriched with comments and favors knowledge transfer. Nevertheless, the two approaches are complementary and can be combined to achieve full reproducibility in terms of input/output of each algorithm/function and the possibility to re-create a computational environment that does not depend on specific user installations.

The R community proposed several solutions based on the literate statistical programming, like *sweave* [10], *knitr* and *rmarkdown* [11]. Within this framework, the authors can release a data analysis as a human-readable document that incorporates data, computational methods (including the short lines of code that are often omitted in a high-level description of the computational procedure), user-parameters, tables, and figures. Moreover, this report is automatically updated each time the analyst introduces some workflow changes to preserve complete reproducibility. R Studio (`https://rstudio.com`) already contains several functionalities that can help an analyst compiling detailed reports. Other R packages, such as Drake [12], go through the same directions.

Even though *rmarkdown* is very popular inside the R community, its usability when developing automated tools as Graphical User Interfaces (GUI) or packages is limited. Despite several efforts, incorporating a RR layer in other software and automatically tracing all the steps performed during a point-and-click analysis is still challenging. In the past, we proposed a solution with the RNASeqGUI [13,14] project. RNASeqGUI is a GUI for analyzing RNA-seq data that automatically traces the analysis steps and reports them in a unique report. Although very useful, this solution did not allow the user to add personal comments, a particularly relevant requisite for knowledge transfer. Moreover, its implementation was time-consuming.

In light of these reasons, we developed *easyreporting*, a R/Bioconductor package allowing us to construct HTML reports that automatically incorporate comments with data, code, plots, and tables. In this work, we describe the *easyreporting* class and its methods. Then, we show i) how *easyreporting* can be used to generate user analysis reports and ii) how *easyreporting* can be used to implement GUIs that automatically trace their functions and produce an analysis report.

Materials and methods

Implementation

Easyreporting is an open-source R/Bioconductor package aimed to 1) support analysts to speed up the compilation of their analysis reports (without learning the *rmarkdown* language) and 2) help developers to integrate a RR layer inside their software products (such as GUI). While the first aim can also be achieved using other similar tools, the latter constitutes one of the main advantages of our solution. In such a way, thanks to

minimal efforts on the developers' side, the end-user (that can be not necessarily aware of the R or the *rmarkdown* language) can obtain an *rmarkdown* file that incorporates the source code generated during the analysis with the user-friendly tools. Once compiled, this document can then be published as supplementary material of a scientific article, helping the interested community to reproduce the computational part of the work entirely, as suggested in [15]. Moreover, the document can be easily organized into sections, describing different analysis steps, and enriched with natural language comments, making the report more explainable to increase the knowledge transfer.

General Description and Initialization

Easyreporting is structured as an S4 class representing a schematic view of a *rmarkdown* file (see Fig 1). Thanks to *easyreporting*, an analysis report can be seen as a particular instance of the package class, where the attributes represent the report characteristics. Within this class, the available methods are useful for attribute manipulations and for inserting comments and organizing section titles inside the report.

Fig 1. The *easyreporting* class package is a representation of a *rmarkdown* file. The color codes indicate which attribute/method represents the same-color portion of the *rmarkdown* first, and the compiled final HTML report then.

When *easyreporting* is used to create a report for novel analysis, the analyst needs to initialize an instance of the *easyreporting* class with the *easyreporting()* constructor function, passing as mandatory arguments the path and the name of the report file accompanied by its title. Optionally, it is possible to specify an author. In this way, each analysis/project is uniquely associated with a specific *easyreporting* instance, and hence to the corresponding *rmarkdown* file. The initialization step can be transparent to any GUI user since the developers handle the tool's backend.

During the initialization, the class constructor automatically creates the report file inside the specified folder tree, setting up its header and declaring the general options for the *rmarkdown* file. As soon as the analyst or the user proceeds with his analysis, the *rmarkdown* file is updated with a new CC each time the analysis software performs a new analysis step. When the analysis is complete, it is possible to compile the report using the *compile* method, which produces the final report (in HTML format) and appends a final CC with the *sessionInfo* to trace all the packages versions used for the analysis.

General Exploitation

The *easyreporting* class is equipped with several methods for *rmarkdown* CC construction (see Table 1 for the full list). Once an *easyreporting* instance is available, it is possible to organize the report by inserting up to six levels of titles by using the *mkdTitle* method. It is also possible to add natural language comments with *mkdGeneralMsg*. The latter feature is particularly relevant to make the analysis more understandable.

For the implementation of the CCs creation, we suggest two main approaches based on the methods available in the class (see the examples in Listings 2 and 3 shown in the Results Section): i) The first approach builds a CC as a typical step-by-step process. It consists of opening a CC (*mkdCodeChunkSt*), adding variables assignments and/or function callings (*mkdVariableAssignement*), and finally closing the CC (*mkdCodeChunkEnd*). In this approach, it is possible to add comments with *mkdGeneralMsg* before closing the CC. ii) The second approach builds the CC in a single step by using the *mkdCodeChunkComplete* method. The method automatically

Table 1. Attributes and Methods of the *EasyReporting* class.

Attributes	Description
filenamePath	the report file name with the absolute path
title	the title of the report
author	the auhor
documentType	actually this is set to HTML
optionList	a list of R Markdown options

Methods	Description
mkdTitle	Inserts an R Markdown title inside the report
mkdGeneralMsg	appends a general message to the report
mkdGeneralTitledMsg	appends a title and a general message to the report
mkdVariableAssignment	includes a variable assignment into the report
mkdCodeChunkSt	creates a CC start
mkdCodeChunkEnd	creates a CC end
mkdCodeChunkComplete	creates a complete CC
mkdCodeChunkCommented	creates a complete CC with a previous comment
mkdCodeChunkTitledCommented	creates a complete CC with a previous comment and a t
mkdSourceFiles	includes a list of source files inside the CC
compile	prints sessionInfo and compiles the R Markdown file
setOptionsList	set an optionList to the class
getOptionsList	returns the optionList from the class
getReportFilename	returns the report filename with path

embraces the tracking code into a new CC, while the user has to take care of the variable   108
assignments and/or the function he/she wants to trace by passing it as an argument. In   109
this approach, the user can also add personal comments passing them as an additional   110
argument. Then, *easyreporting* automatically adds the comment before the new CC.   111

The first approach appears useful when we need to carry out several R commands in   112
a single CC. It is similar in the spirit to the functionalities offered by R-studio or other   113
development environments, however since it is entirely command-line, it can be easily   114
used on systems with limited development capabilities. By contrast, the second   115
approach is more appropriate for tracing a single function call automatically. In the   116
next section, we will show how this second possibility can be particularly useful to wrap   117
functions performing a specific step and trace their execution within GUIs.   118

Implementing automatically tracing functions and their usage   119
within GUIs   120

The previously described CCs creation approaches can be adapted to trace several steps   121
of an analysis pipeline and end-up with a nicely formatted and detailed analysis report.   122
However, they require the analysts to manually trace each step of the analysis (as he   123
could also do with other available tools). Consequently, the above approaches are useful   124
for generating analysis reports (that was the first aim of the *easyreporting* package).   125
However, they are not suited for the automatic tracing of the steps of an analysis   126
performed using point-and-click approaches thorough GUIs (the second aim of the   127
easyreporting package).   128

In the last decade, GUIs are becoming very popular in bioinformatics because they   129
simplify computational analysis allowing non-expert users to choose among several   130
computational procedures, algorithms, and parameter settings, see for   131
example [13, 14, 16–18]. In particular, the *shiny* (https://shiny.rstudio.com)   132
libraries simplified the development of GUIs that incorporate the power of the   133

statistical R language and the wide-amount of open-source packages available in 134
repositories such as Bioconductor (`https://www.bioconductor.org`). 135

Nevertheless, computational studies obtained from GUIs might lack reproducibility 136
since tracking all user choices is still challenging. To face this limit, the developers have 137
to implement a RR layer when designing the GUI's back-end so that the final users can 138
benefit from a better quality product. Moreover, the RR layer has to be transparent but 139
understandable to not-expert users. Ideally speaking, at the end of the analysis, the 140
user should have a human-readable report analogous to the one obtained using 141
command-line approaches. 142

Easyreporting methods can be easily adapted to support the automatic tracing of 143
any given function by combining a rendering function that performs the required step 144
with a wrapping function that traces its execution. The wrapper function (WF) needs 145
an *easyreporting* instance, and the arguments of the function to be traced (TF). Then, 146
the developer inserts the WF in the back-end of the interface (i.e., the server if the 147
context of a GUI implemented with the shiny library) in the TF place. The front-end of 148
the interface (i.e., the UI with the shiny library) remains unchanged. When the user 149
interacts with the interface to invoke the TF, the back-end will invoke the WF, which 150
will call both the TF function of interest and trace its usage with all parameters. In 151
brief, employing wrapper functions makes it possible to implement a reproducible 152
research layer within the GUI without implementing all the tracing *rmarkdown* code. 153
Listings 4-6 illustrate a specific case with a volcano plot, and Figure 2 schematically 154
represents the entire workflow of information. 155

Fig 2. Example of a Graphical User Interface working with *easyreporting* package. The
right panel shows the user side of the software with the user interactive GUI. The left
panel shows the RR layer transparent to the user, which traces the performed step as
organized by the developer. In this example, the GUI allows performing a Volcano Plot
starting from a DEGs data frame that includes Log Fold change and p-values as column
variables. With the p-value threshold selector (in blue), the user can set the threshold
for the significance of the p-values. Moreover, the user can add a personal comment (in
yellow) that wants to add alongside the traced code. The user has to press the perform
button (in red) to visualize the plot into the GUI. In the meantime, the back end will
trace the step into the *rmarkdown* file and will return the plot to the user interface.
The user can then compile the report through the compile report button (in green) and
visualize the HTML report. The back-end will execute the *compile()* function and
return the result to the interface. To make this possible, the developer needs to include
a RR layer into the GUI engine (i.e., the shiny server-side). For example, the developer
can define a traced function (TF) that renders the Volcano-plot and a wrapper function
(WF) that takes as input the input arguments from the graphical GUI (i.e., the shiny
UI side). The WF code can be structured to render the plot into the graphical GUI and
make a call to an *easyreporting* method, combining the input arguments with the
easyreporting class method required ones, to write a new CC into the analysis
rmarkdown. Additionally, the developer has to bind the *Compile Report* button with
the *compile easyreporting* method to generate the HTML final report.

Results 156

To illustrate the capabilities of *easyreporting* we first show its usage for generating an 157
analysis report in a case study concerning the analysis of RNA-seq data (see 158
Supplementary File 1 for details), then we illustrate how to implement a simple GUI 159
that automatically traces its usage and the choice of parameters to produce a report. 160

Easyreporting for the creation of analysis report

The RNA-seq data used in the example allows investigating the differences in CD8+ dendritic T-cells of the immune response of two different antibodies compared with control, see [19] for more details. We chose this illustrative example since it is well-known that the analysis of RNA-seq data can lack reproducibility [20]. The dataset contains the raw counts of 37991 genes and is composed of two replicates for each of the three conditions: DEC (fd-scaDEC-205 antibody samples); E2 (E2 antibody samples) and UNTR (control samples). For illustrative purposes, in our Supplementary file 1, we start the analysis from the raw count-matrix. Moreover, we released the raw counts as supplementary data with the *easyreporting* package, allowing the readers to reproduce our example. The naive pipeline will first load the data, perform some diagnostic plots, filter and normalize the raw counts, and visualize the principal component projection. It will then perform differential gene expression analysis and depict the results as a Venn diagram and MA-plots. A specific CC describes each phase.

In the following, we show the main fundamental steps that a user can adapt to any analysis, and we refer to the Supplementary File 1 for the detailed description of the remaining steps and to the Supplementary File 2 for the complete report.

Report Initialization

After loading the *easyreporting* package in the R environment, the analyst needs to initialize an analysis report by providing the file name (i.e., "rnaseq_report") and the title of the document (i.e., "RNA-seq Analysis Report"). It is also possible to specify an author (i.e., "Dario Righelli"). For simplicity, we set-up a project directory path starting from the working directory for our report, but the user can choose other locations by setting the *filenamepath* parameter. The initialization is carried out by using the *easyreporting()* function. Note that the *filenamepath* and *title* are mandatory parameters, while the author is optional. The following Listing 1 code illustrates the initialization of a report.

Listing 1. Initialization chunk

```
library("easyreporting")
 proj.path <- file.path(getwd(), "rnaseq_report")
 bioEr <- easyreporting(filenamepath=proj.path,
                         title="RNA-seq Analysis Report",
                         author=c("Dario Righelli"))
```

Creation of a chunk of code

Once the analyst has initialized the report, he/she can add a CC for each step of the analysis. As mentioned in the General Exploitation section, EasyReporting provides two main approaches for adding CCs within a report: 1) building up the CC step by step (as shown in Listing 2) and 2) using several kinds of wrapper functions (as shown in Listing 3).

As mentioned above, in the first case, the analyst has to use the *mkdCodeChunkSt* to open a new CC. Then, he/she needs to add the code to markdown, by using the *mkdVariableAssignment* and/or the *mkdGeneralMsg* functions, for tracking variables and functions. Finally, the analyst has to close the CC using the *mkdCodeChunkEnd* function. The following Listing 2 code illustrates a step-by-step CC for loading the counts matrix released with the package.

Listing 2. Step-by-step chunk construction

```
mkdTitle(bioEr, title="Loading Counts Data")                                    205
mkdCodeChunkSt(bioEr, sourceFilesList=system.file(                              206
          "script/importFunctions.R",                                          207
          package="easyreporting"),                                            208
          isComplete=TRUE)                                                      209
mkdVariableAssignment(bioEr, "geneCounts",                                      210
          paste("as.matrix(                                                    211
          importData(system.file('",                                           212
          "extdata/BMDC_counts_FeatureCounts.xlsx', ",                         213
          "package='easyreporting')))", sep="\n"),                             214
          show=FALSE)                                                          215
mkdGeneralMsg(bioEr, "head(geneCounts, 20)")                                    216
mkdCodeChunkEnd(bioEr)                                                          217
```

Although the first approach leaves complete freedom to the analyst, it can be tricky   218
for small CCs. The second approach can be more straightforward for small CCs. To this   219
purpose, the *mkdCodeChunkComplete* function allows tracing the steps through the   220
message parameter. The following Listing 3 code illustrates an example of a single step 221
CC. As for the above CC, we assume that the analyst wants to read the raw counts   222
using a user-defined function, here named "importData.R", that we stored into the   223
importFunctions.R file available in the package "script" folder.   224

Listing 3. One command chunk construction

```
mkdCodeChunkComplete(object=bioEr,                                              225
     message=paste("geneCounts <- ",                                           226
          "as.matrix(importData(system.file(",                                 227
          "'extdata/BMDC_counts_FeatureCounts.xlsx', ",                        228
          "package='easyreporting')))",                                        229
          "head(geneCounts, 20)", sep="\n"),                                    230
          sourceFilesList=system.file(                                         231
          "script/importFunctions.R",                                          232
          package="easyreporting"),                                            233
          optionList=makeOptionsList(evalFlag=FALSE))                          234
```

Note that the *mkdCodeChunkComplete* allows also to provide specific options for the   235
CC that we are creating. In particular, in this case, if we turn the *evalFlag* to *FALSE*, 236
the code will not be compiled during the final report construction.   237
It is possible to organize the report in both cases using the *mkdTitle* function. The   238
user has to repeat this operation for each step of the analysis, as shown in the   239
Supplementary File 1. At the end of the process, it is possible to compile the   240
easyreporting instance and obtain the analysis report as in Supplementary File 2 in   241
HTML format.   242

Implementing automatical tracing functions   243

This section shows a possible approach on how to encapsulate *easyreporting* methods   244
into third-parties functions to trace the analysis step and execute the code   245
automatically.   246
First of all, the developer has to write an R function that performs the analysis step   247
of interest (such as the MAedgeRMAPlotEx function for rendering an MA-plot, in our   248
example). Listings 4 shows a simple example of rendering function.   249

Listing 4. MA-plot rendering function

```
MAedgeRMAPlotEx <- function(degList)                                            250
```

```
{                                                                              251
    for (i in seq_along(degList)) {                                            252
    degenes <- degList[[i]]$FDR < 0.01                                         253
    with(degList[[i]], plot(logCPM, logFC,                                     254
        pch=16, cex=0.2, main=names(degList)[i]))                              255
    with(degList[[i]], points(logCPM[degenes],                                 256
        logFC[degenes], col='red', pch=16, cex=0.2))                           257
    }                                                                          258
}                                                                              259
```

Note that the developer does not require any extra effort at this stage. Moreover, ²⁶⁰
the rendering function could also be any function available from other packages. ²⁶¹

Then, the developer needs also to write a wrapper function (here ²⁶²
traceAndPlotMAPlot). The wrapper function should take as input the arguments of the ²⁶³
rendering function (here MAedgeRMAPlotEx), and a generic *easyreporting* object (here ²⁶⁴
er). Moreover, the wrapper function has to call the *mkdCodeChunkTitledCommented* ²⁶⁵
function of *easyreporting* (where we insert the rendering function call to be traced ²⁶⁶
(MAedgeRMAPlotEx) in the *codeMsg* argument) and the call to rendering function ²⁶⁷
(MAedgeRMAPlotEx). Listings 5 shows the wrapper function of our example. ²⁶⁸

Listing 5. Tracing wrapper function chunk

```
traceAndPlotMAPlot <- function(degList, er)                                    269
{                                                                              270
    mkdCodeChunkTitledCommented(er,                                            271
        title="Recursive Tracing Function", level=2,                          272
        codeMsg="MAedgeRMAPlotEx(degList='degList')")                         273
    MAedgeRMAPlotEx(degList=degList)                                           274
}                                                                              275
```

In this way, the wrapper function allows both to show the result and trace the ²⁷⁶
function in the same step. It is easy to place the *traceAndPlotMAPlot* function-call ²⁷⁷
wherever needed in the main code. ²⁷⁸

Listing code 5) shows how to use the wrapper function. In particular, we pass as ²⁷⁹
input i) the object required by the rendering function (an edgeR result class in this ²⁸⁰
particular example), ii) an *easyreporting* class instance (here it is the bioEr instantiated ²⁸¹
in the example). ²⁸²

Listing 6. Tracing function call chunk

```
traceAndPlotMAPlot(degList=degList, er=bioEr)                                  283
```

Note that writing the wrapper function is the only extra effort required to achieve ²⁸⁴
reproducibility. ²⁸⁵

easyreporting for GUI implementation   286

To better illustrate how to incorporate a RR layer into a GUI, *easyreporting* (since ²⁸⁷
version >= 1.3.1 released with Bioconductor 3.13) contains a simple ShinyApp example ²⁸⁸
for plotting a Volcano plot. The command *erGUIVolcano()* allows executing the app ²⁸⁹
and opens the user interface. The user interface allows the user to choose a threshold ²⁹⁰
for the P-value (i.e., the *P-value threshold* for detecting the significant genes in this ²⁹¹
example) and provide a text area for adding comments. In the interface, there are also ²⁹²
two buttons (*Perform Plot* and *Compile Report*) for executing the plot and compiling ²⁹³
the report, respectively. The *ui* function provides the code for the user interface. This ²⁹⁴
code does not need to be modified to allow reproducibility. Instead, the back-end of the ²⁹⁵

interface that executes the job has to incorporate a wrapper function (here   296
traceAndPlotVolcano), and the call to the wrapper function, respectively. In this   297
example, we also added the report's initialization. The *server* function provides the   298
code for the back-end interface.   299

Figure 2 right side shows a schematic representation of the user interface as in the *ui*   300
function. The left side illustrates the back-end as in the *server* function. By using the   301
Perform Plot button (red box), the user activates the WF into the server-side, which in   302
turn performs the plot and traces the executed function (the red cascade). In blue is   303
highlighted the argument value and how it is traced through the function cascade.   304

Additionally, when the user adds its comments (yellow box) to the performed   305
analysis step, the text is passed to the server.   306

Finally, the user can compile the report using the *Compile Report* button (green   307
box). In this way, the server executes the *compile()* function and produces the HTML   308
report that is automatically showed to the user.   309

This simple example can be generalized to complex interfaces in order to trace all   310
user interactions.   311

Conclusions   312

Easyreporting can be used to support RR in different analysis contexts. However, it is   313
particularly suited for analyzing omics data and developing software/GUIs, as we have   314
shown in this work. Compared to other previously proposed solutions such as [21], that   315
require not negligible commitments by the final user, potentially bringing him/her to   316
renounce to include RR inside the scripts, the implementation of a RR layer with our   317
approach is straightforward. Moreover, it leaves maximum freedom to the   318
developer/analyzer for automatically creating and storing an *rmarkdown* document and   319
providing methods for its compilation and adding comments in natural language.   320

Although several functionalities are already available, *easyreporting* can still benefit   321
from some extra features such as methods for file editing, graphical representation of   322
the analysis, and data caching. In particular, file editing can be useful for modifying   323
specific CCs, and the graphical representation of the analysis can be useful to provide   324
reports not only readable by third-party users but also graphically visualized as   325
workflows. On the other hand, even though a dedicated data caching infrastructure can   326
offer more manageability and share-ability of the data at the moment, it can be already   327
performed in *easyreporting* by *rmarkdown* CCs option flag.   328

Finally, thanks to its versatility, *easyreporting* can be ideally included in any   329
well-structured R project and the development of GUIs, helping to fulfill most of the   330
proposed rules in [2]. Moreover, if combined with other tools such as dockers (i.e.,   331
docker4seq [22]) it helps to create fully reproducible projects. *Easyreporting* naturally   332
complements dockers in terms of reproducibility, allowing both the preserve code lines   333
and user parameters and the computational environments and dependencies.   334

To conclude, our approach still requires the developer's effort to implement a RR   335
layer into their software, which makes us imagine possible future works in this area   336
where the code tracing is entirely left to the machine. The Java language provides a   337
well-known example that uses the Aspect-Oriented Programming (AOP) paradigm for   338
the software logging aspects. Unfortunately, this paradigm is still missing in the R   339
language, but possible future approaches in the Reproducible Research area inside R   340
could rely on implementing it, which, combined with *rmarkdown* or similar procedures,   341
can be used to trace ad-hoc tagged functions and to log them into the report file. In   342
such a way, reproducibility could be easier to implement and lesser subject to human   343
errors.   344

Supporting information

Supplementary File 1 provides an illustrative example for the creation of an analysis report.

Supplementary File 2 provides the report file obtained using the analysis steps described in the Supplementary File 1.

Author Contribution

Dario Righelli, Conceptualization, Data curation, Formal analysis, Methodology, Software, Writing – original draft and Claudia Angelini, Conceptualization, Funding acquisition, Methodology, Supervision, Writing – original draft

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