Molnupiravir (EIDD-2801) inhibits SARS-CoV2 replication in Syrian hamsters model

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Author Contributions

R.A., C.S.F. and J.N. designed research; R.A., C.S.F., S.J.F.K., X.Z. and L.L. performed research; R.A., C.S.F.

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Abstract

Since its emergence in Wuhan, China in December 2019, the severe acute respiratory syndrome

coronavirus 2 (SARS-CoV-2) has spread worldwide resulting in a global pandemic with >1.5 million

deaths until now. In the search for small molecule inhibitors of SARS-CoV-2, drug repurposing is being

extensively explored. Molnupiravir (EIDD-2801) is an orally bioavailable nucleoside analog that

possesses a relatively broad-spectrum antiviral activity including against coronaviruses. We here

studied the effect of EIDD-2801 in a well-established Syrian hamster SARS-CoV2 infection model.

Treatment of SARS-CoV-2-infected hamsters with 200 mg/kg BID of EIDD-2801 for four consecutive

days, starting from the day of infection, significantly reduced infectious virus titers and viral RNA loads

in the lungs and markedly improved lung histopathology. When onset of treatment was delayed until

1 or 2 days after infection, a very modest antiviral effect was observed. The potential of EIDD-2801 for

the treatment and or prevention of SARS-CoV2 deserves further attention.

Keywords

SARS-CoV-2; Antivirals; EIDD-2801; MK-4482; Molnupiravir; preclinical model.

 Main Text

The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is a β-coronavirus that was first

identified in Wuhan, China in December 2019 (1). Since then, the virus rapidly spread around the globe

with more than 66 million cases reported by the end of 2020, including >1.5 million deaths

[https://covid19.who.int/]. Infection with SARS-CoV-2 results in COVID-19 which is characterized by a

wide range of symptoms including fever, dry cough, muscle and/or joint pain, headache, decreased

sense of taste and smell and diarrhea. The disease can also progress into severe complications such as

acute respiratory distress syndrome (ARDS), respiratory failure, septic shock and multi-organ failure,

which are mainly attributed to a massive cytokine storm and exaggerated immune response (2).

To date, there are no approved, specific antiviral drugs or vaccines available to treat or prevent

infections with human coronaviruses. The use of potent antivirals against SARS-CoV-2 will reduce viral

loads and may hence reduce the chance to progress to a severe disease. In addition, such antiviral

drugs could be useful to protect for example health care workers and high-risk groups in a prophylactic

setting. Since the *de novo* development and approval of (a) specific, highly potent antiviral(s) for SARS-

CoV-2 may take years, the main focus for COVID-19 treatment in the current pandemic is to repurpose

drugs that have been approved or in clinical trials for other diseases (3).

The ribonucleoside analogue, N4-hydroxycytidine (NHC, EIDD-1931), has broad-spectrum antiviral

activity against multiple viruses belonging to different families of RNA viruses. Activity against SARS-

CoV and SARS-CoV-2 has been reported in cell lines and primary human airway epithelial cell cultures

(4). Acting through lethal mutagenesis, its incorporation into viral RNA results in the accumulation of

deleterious transition mutations beyond a permissible error threshold to sustain the virus population,

leading to error catastrophe (5). The orally bioavailable, pro-drug counterpart of NHC (6), Molnupiravir

(EIDD-2801, MK-4482) is currently being assessed for its potential as an antiviral treatment of SARS-

CoV-2 infection in Phase 2 clinical trials of infected patients (NCT04405570, NCT04405739). To our

knowledge, three very recent studies reported on activity of orally dosed EIDD-2801 in SARS-CoV-2

infected animals. Oral treatment of SARS-CoV2 infected Syrian hamsters with EIDD-2801 resulted in

marked reduction of viral loads when administered either in a pre- or post-exposure settings (7). In a

ferret model, EIDD-2801 significantly reduced virus load in a therapeutic setting and blocked SARS-

CoV-2 contact transmission (8), while in a humanized mouse model, EIDD-2801 prevented SARS-CoV-

2 infection in a pre-exposure prophylaxis setting (9).

Recently, we have established and characterized a Syrian Gold (SG) hamster model for the evaluation

of antiviral drugs against SARS-CoV-2 (10, 11). Using this model, we have shown that a high dose of the

influenza drug Favipiravir results in a pronounced antiviral activity in SARS-CoV-2 infected hamsters,

whereas hydroxychloroquine lacks antiviral activity in this model. Here, we use the same hamster

model to assess the anti-SARS-COV2 of EIDD-2801 against SARS-CoV-2 in infected hamsters.

First, we evaluated the effect when the drug was administered at a dose of either 75 or 200 mg/kg BID.

Briefly, 6-8 weeks female SG hamsters were treated with the intended dose of the compound or the

vehicle (i.e. the control group) for four consecutive days starting one hour before intranasal infection

with 50 µL containing 2×10^6 $TCID_{50}$ SARS-CoV-2 [BetaCov/Belgium/GHB-03021/2020 (EPI ISL 109

407976|2020-02-03)]. At day four post-infection (pi), the animals were euthanized and lungs were

collected for quantification of viral RNA, infectious virus titers and lung histopathology as described

previously (10) (Fig. 1A). Treatment of hamsters with 75 mg/kg BID EIDD-2801 resulted in 1.2 $\log_{10}$

reduction in the viral RNA copies per mg of lung tissue (P=0.01), compared to the vehicle-treated

infected hamsters (Fig. 1B). On the other hand, in the lungs of hamsters that had been treated with

200 mg/kg BID EIDD-2801 a 3$\log_{10}$ reduction in the viral RNA copies/mg of lung tissue was noted (P

<0.0001) (Fig. 1B). A similar pattern was observed for the infectious virus load in the lungs, the high

dose of 200 mg/kg, but not the 75 mg/kg dose, reduced infectious virus lung titers by 3.3 $\log_{10}$

(P<0.0001) $TCID_{50}$ per mg (Fig. 1C).

Fig.1. *In vivo* efficacy of EIDD-2801 against SARS-CoV2 at the time of infection. (A) Set-up of the study. (B) Viral RNA levels in the lungs of control (vehicle-treated) and EIDD-2801-treated SARS-CoV-2–infected hamsters at day 4 post-infection (pi) are expressed as $\log_{10}$ SARS-CoV2 RNA copies per mg lung tissue. The bars represent median values. (C) Infectious viral loads in the lungs of control (vehicle-treated) and EIDD-2801-treated SARS-CoV-2–infected hamsters at day 4 pi are expressed as $\log_{10}$ $TCID_{50}$ per mg lung tissue. The bars represent median values. (D) Cumulative severity score from H&E stained slides of lungs from control (vehicle-treated) and EIDD-2801-treated hamsters. The bars represent median values. (E) Weight change at day 4 pi in percentage, normalized to the body weight at the day of infection. Bars represent means ± SD. Data were analyzed with the Mann–Whitney U test. *$P < 0.05$; ***$P < 0.001$, ****$P < 0.0001$.

Importantly, treatment with 200 mg/kg EIDD-2801 BID significantly reduced the histological lung

disease score (P=0.001), whereas the lower dose resulted only in slight reduction in lung pathology

(P=0.05, ns, Fig. 1D). Both doses were well-tolerated without significant weight loss or any obvious

adverse effects (Fig. 1E).

We next explored whether the high dose can also be used in a post-exposure setting, i.e. when

treatment is started at 24 or 48 h after infection (Fig. 2A). Delaying the start of treatment with EIDD-

2801 (200 mg/kg, BID) by 1 or 2 days resulted in a loss of potency with only a rather slight reduction

of viral RNA copies/mg lung [0.3 to 0.4 $\log_{10}$ respectively] (Fig. 2B). Similarly, virus titrations of the lung

tissues also revealed no significant reduction of infectious virus load in both groups with delayed

treatment compared with the vehicle-treated group (Fig. 2C). However, a modest but significant

reduction of the histological lung disease score was observed in both the day 1 (p=0.016) and day 2

(p=0.01) delayed treatment groups (Fig. 2D). These results suggest that even if the delayed treatment

with the 200 mg/kg BID dose was not sufficient to efficiently stop the viral replication, it may still able

to delay the disease progression in the lung of infected hamsters.

In summary, EIDD-2801 is able to markedly reduce, albeit at a relatively high dose, SARS-CoV2 infection

and virus induced pathology in hamsters, in particular when given at the time of Infection. EIDD-2801

has been previously demonstrated to exert antiviral activity against MERS-CoV and SARS-CoV in cell

culture and mouse models. Very recently activity was also reported against SARS-CoV-2 (4, 8, 9). In

another SARS-CoV2 Syrian hamster model, 250 mg/kg of EIDD-2801 was given orally every 12 hours

starting either 12 hours pre-infection or 12 hours post-infection (7). Although a slightly higher

compound dose (250 mg/kg, BID) and a much lower virus inoculum (5×10^2 TCID$_{50}$) was used in their

hamster study, the observed antiviral effect was less pronounced (1 $\log_{10}$ reduction in viral RNA and 2

$\log_{10}$ reduction in infectious virus titers) than in our study (7).

Fig.2. *In vivo* efficacy of EIDD-2801 against SARS-CoV2 in a post-exposure setting. (A) Set-up of the study. (B) Viral RNA levels in the lungs of control (vehicle-treated) and EIDD-2801-treated SARS-CoV-2−infected hamsters at day 4 post-infection (pi) are expressed as log10 SARS-CoV2 RNA copies per mg lung tissue. The bars represent median values. (C) Infectious viral loads in the lungs of control (vehicle-treated) and EIDD-2801-treated SARS-CoV-2−infected hamsters at day 4 pi are expressed as log10 $TCID_{50}$ per mg lung tissue. The bars represent median values. (D) Cumulative severity score from H&E stained slides of lungs from control (vehicle-treated) and EIDD-2801-treated hamsters. The bars represent median values. Data were analyzed with the Mann−Whitney U test. *P < 0.05, ***P < 0.001, ****P < 0.0001.

EIDD-2801 has also been reported to be effective in SARS-CoV infected C57/BL6 mice either when

administered prophylactically (start of treatment 2 h before infection) or therapeutically (start of

treatment delayed until 48 hpi) at a dose of 500 mg/kg twice daily (Sheahan et al., 2020). In a

humanized mouse model of SARS-CoV-2 infection, pre-exposure (prophylaxis) (12 h before infection)

with 500 mg/kg of EIDD-2801 twice daily was efficacious in preventing SARS-CoV-2 infection, with a ~6

log_{10} reduction in virus lung titers (9). In a very recent report, EIDD-2801 was found to markedly reduce

SARS-CoV-2 viral titers in in the upper respiratory tract of ferrets when start of treatment was delayed

until 12 hpi to 36 hpi. Moreover, treating SARS-CoV2 infected ferrets with EIDD-2801 starting at 12 hpi

prevented contact transmission when they were co-housed with non-infected untreated contact

ferrets (8).

Our results are consistent with other recent studies (in hamster, mouse and ferret models) showing

that pre-emptive and early intervention with oral EIDD-2801 results in antiviral activity. However, at a

dose of 200 mg/kg we did not observe a reduction of virus in the lung when treatment was initiated

24 to 48 hpi in our Syrian hamster model (although some improvement in lung pathology was observed

in these groups). Further studies are ongoing to explore therapeutic (delayed start of treatment) effect

at higher doses.

The antiviral drug, Remdesivir (Veklury), is the first treatment for COVID-19 to receive FDA approval

for use in hospitalised patients, although the World Health Organisation has recently recommended

against its use (https://www.fda.gov/news-events/press-announcements/fda-approves-first-

treatment-covid-19; https://www.who.int/news-room/feature-stories/detail/who-recommends-

against-the-use-of-remdesivir-in-covid-19-patients). Both Remdesivir and EIDD-2801 are nucleoside

analogues acting on the viral RNA replication pathway, with Remdesivir resulting in chain termination

and EIDD-2801 in lethal mutagenesis (5, 12). Additionally, both have a high resistance barrier, with

resistant viral mutants having a loss in fitness (6, 13). However, Remdesivir needs to be administrated

intravenously, whereas EIDD-2801 can be dosed via the oral route. This is a significant advantage in

terms of the practicalities of treatment management, where we can envision that asymptomatic

patients who have had high-risk contact or patients presenting mild symptoms are prescribed EIDD-

2801 pills in a non-hospitalised setting.

By demonstrating the antiviral efficacy of the orally-bioavailable EIDD-2801 against SARS-CoV-2 in the

Syrian hamster model, we contribute to the pre-clinical profiling of EIDD-2801, and provide further

evidence in support of the ongoing Phase II clinical trials. The outcomes of these trials could play a

pivotal role in the control and management of this devastating pandemic.

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References

1.   N. Zhu, *et al.*, A Novel Coronavirus from Patients with Pneumonia in China, 2019. *N. Engl. J.*

Med. **382**, 727–733 (2020).

2.   M. Z. Tay, C. M. Poh, L. Rénia, P. A. MacAry, L. F. P. Ng, The trinity of COVID-19: immunity,

inflammation and intervention. *Nat. Rev. Immunol.* **20**, 363–374 (2020).

3.   L. Delang, J. Neyts, Medical treatment options for COVID-19. *Eur. Hear. J. Acute Cardiovasc.*

Care, 204887262092279 (2020).

4.   T. P. Sheahan, *et al.*, An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 in

human airway epithelial cell cultures and multiple coronaviruses in mice. *Sci. Transl. Med.* **12**

(2020).

5.   N. Urakova, *et al.*, β-D- N 4 -Hydroxycytidine Is a Potent Anti-alphavirus Compound That

Induces a High Level of Mutations in the Viral Genome . *J. Virol.* **92** (2017).

6.   M. Toots, *et al.*, Characterization of orally efficacious influenza drug with high resistance

barrier in ferrets and human airway epithelia. *Sci. Transl. Med.* **11** (2019).

7.   K. Rosenke, *et al.*, Orally delivered MK-4482 inhibits SARS-CoV-2 replication in the Syrian

hamster model. *Res. Sq.* (2020).

8.   R. M. Cox, J. D. Wolf, R. K. Plemper, Therapeutically administered ribonucleoside analogue

MK-4482/EIDD-2801 blocks SARS-CoV-2 transmission in ferrets. *Nat. Microbiol.* (2020).

9.   A. Wahl, *et al.*, Acute SARS-CoV-2 Infection is Highly Cytopathic, Elicits a Robust Innate

Immune Response and is Efficiently Prevented by EIDD-2801. *Res. Sq.* (2020).

10. S. J. F. Kaptein, *et al.*, Favipiravir at high doses has potent antiviral activity in SARS-CoV-

2–infected hamsters, whereas hydroxychloroquine lacks activity. *Proc. Natl. Acad. Sci. U. S. A.*

117, 26955–26965 (2020).

11.   R. Boudewijns, *et al.*, STAT2 signaling restricts viral dissemination but drives severe

199   pneumonia in SARS-CoV-2 infected hamsters. *Nat. Commun.* **11**, 5838 (2020).

200   12.   T. P. Sheahan, *et al.*, Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic

201   coronaviruses. *Sci. Transl. Med.* **9** (2017).

202   13.   M. L. Agostini, *et al.*, Coronavirus susceptibility to the antiviral remdesivir (GS-5734) is

203   mediated by the viral polymerase and the proofreading exoribonuclease. *MBio* **9** (2018).