Motor planning brings human primary somatosensory cortex into movement-specific preparatory states

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1 Abstract

2 Motor planning plays a critical role in producing fast and accurate movement. Yet, the neural 3 processes that occur in human primary motor and somatosensory cortex during planning, 4 and how they relate to those during movement execution, remain poorly understood. Here 5 we used 7T functional magnetic resonance imaging (fMRI) and a delayed movement 6 paradigm to study single finger movement planning and execution. The inclusion of no-go 7 trials and variable delays allowed us to separate what are typically overlapping planning and 8 execution brain responses. Although our univariate results show widespread deactivation 9 during finger planning, multivariate pattern analysis revealed finger-specific activity patterns 10 in contralateral primary somatosensory cortex (S1), which predicted the planned finger movements. Surprisingly, these activity patterns were similarly strong to those found in 11 12 contralateral primary motor cortex (M1). Control analyses ruled out the possibility that the 13 detected information was an artifact of subthreshold movements during the preparatory 14 delay. Furthermore, we observed that finger-specific activity patterns during planning were 15 highly correlated to those during movement execution. These findings reveal that motor 16 planning activates the specific S1 and M1 circuits that are engaged during the execution of 17 a finger movement – while activity in S1 and M1 is overall suppressed. We propose that 18 preparatory states in S1 may improve movement control through changes in sensory 19 processing or via direct influence of spinal motor neurons.

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21 Key words

22 Motor planning; Sensorimotor cortex; Finger control; fMRI; Representational geometry.

24 Significance statement

25 Motor planning is important for good motor performance, yet it is unclear which neural 26 processes underlie the preparation of the nervous system for an upcoming movement. Using 27 high-resolution functional neuroimaging, we investigated how the planning of finger 28 movements changes the activity state in primary motor (M1) and somatosensory (S1) cortex. 29 and how brain responses during planning and execution relate to each other. We show that 30 planning leads to finger-specific activation in both M1 and S1, which is highly similar to the 31 finger-specific activity patterns elicited during movement execution. Our findings suggest 32 that even S1 is being specifically prepared for an upcoming movement, either to actively 33 contribute to the outflowing motor command or to enable movement-specific sensory gating.

35 Introduction

36 Animals are capable of generating a wide variety of dexterous behaviors accurately and 37 effortlessly on a daily basis. This remarkable ability relies on the motor system reaching the appropriate preparatory state before each movement is initiated. At the level of behavior, the 38 process of motor programming, or planning, has long been shown to be beneficial to 39 40 performance (1-3), leading to faster reaction times (4-6) and more accurate response 41 selection (7–10). The behavioral study of motor planning has spurred the neurophysiological 42 investigation of what movement parameters are specified in the neuronal firing of a number 43 of cortical regions including the dorsal premotor cortex, PMd (11–13), the supplementary 44 motor area, SMA (14), and the posterior parietal cortex, PPC (15–17). Building on this work, 45 human neuroimaging studies have shown that activity in parieto-frontal brain regions during 46 planning of prehension movements can be used to decode several movement properties 47 such as grip type (18, 19), action order (20), and effector (21-24). At the level of neural population dynamics (25), motor planning can be understood as bringing the neuronal state 48 49 towards an ideal preparatory point. Once this state is reached and the execution is triggered, 50 the intrinsic dynamics of the system then let the movement unfold (26, 27). While the neuronal 51 correlates of movement planning have largely been studied in non-human primates using 52 upper limb movements, such as reaching and grasping, motor planning also plays a crucial 53 role in fine hand control (7).

Despite their importance for everyday dexterous behaviors such as typing, writing, or 54 55 tying a knot, finger movements have not been closely investigated at the single neuron level. 56 In humans, previous fMRI studies of finger movements have not separated planning from 57 execution (28–35). Therefore, we have an incomplete picture of how motor planning readies 58 the human sensorimotor system for the production of individuated finger movements. Based on previous work in reaching, we expected that the primary motor cortex (M1) should 59 60 represent movements during both planning and execution (36–38). What is unclear, however, 61 is whether the primary somatosensory cortex (S1) also receives information about the 62 planned movement before movement onset. Furthermore, we currently do not know how brain representations of planned finger movements relate to those during movement 63 execution. To address these gaps, we designed a high-field (7T) fMRI experiment to study 64 65 what brain regions underlie the planning of individual finger movements. We used no-go trials

- 66 and variable delays to temporally separate the evoked responses to movement planning and
- 67 execution, and advanced multivariate pattern analyses to examine the correspondence
- 68 between fMRI correlates of planned and executed finger movements.
- 69

70 Results

71 Deactivation in sensorimotor regions during planning of finger movements

We instructed 22 participants to plan and execute repeated keypresses with individual
fingers of their right hand on a keyboard device while being scanned with 7T fMRI. The key
to be pressed corresponded to one of three fingers (thumb, middle, and little) and was cued
during the preparation phase by numbers on the screen (1=thumb, 3=middle, 5=little, e.g.,
Fig. 1A).

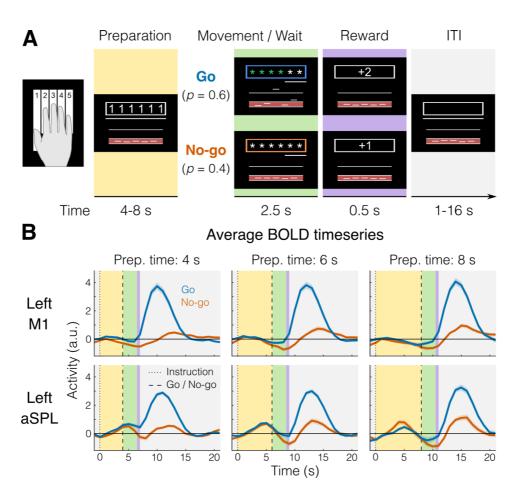


Figure 1 | fMRI task and BOLD responses. A. Example trial with timing information. Background colors indicate different experimental phases (yellow = preparation; green = movement (go) or wait (no-go); purple = reward; gray = inter-trial interval, ITI). B. Group-averaged BOLD response (N = 22) for go (blue) and no-go (orange) trials in a region that shows no planning evoked activity (Left M1, top), and one that shows some planning evoked activity (Left ASPL, bottom). Shaded areas indicate standard error of the mean (SEM). Background colors correspond to trial phases as in A.

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79 After a variable delay (4–8 seconds), participants received a color cue indicating whether to 80 press the planned finger (go trials), or whether to withhold the response (no-go trials). Upon 81 the go cue, participants had to initiate the correct response as fast as possible and make 6 82 presses of the designated finger, before receiving accuracy points for reward (see Methods). To control for involuntary overt movements during the preparation phase, we required 83 84 participants to maintain a steady force on all of the keys during the delay, which was closely 85 monitored online. To ensure that planning results would not be biased by the subsequent 86 execution, we restricted all our analyses to no-go trials without a subsequent movement (see 87 Methods). First, we asked which brain regions showed an evoked response during the planning of finger movements (e.g., Fig. 1B). We focused our analysis on the lateral aspect 88 89 of the contralateral (left) hemisphere (purple and white areas of the Fig. 2 inset) which 90 included premotor, primary motor, primary somatosensory, and anterior parietal cortical 91 regions. To examine brain activation during finger planning, we performed a univariate 92 contrast of the preparation phase (across the three fingers) versus the resting baseline (Fig. 93 2A). Overall, the instruction stimulus evoked little to no activation in our regions of interest 94 (ROIs, Fig. 2A). In fact, compared to resting baseline, we observed significant deactivation (Fig. 2E) in dorsal premotor cortex (PMd, t_{21} = -2.642, p = 0.015), primary motor cortex (M1, 95 96 $t_{21} = -7.592$, p = 1.887e-07), and primary somatosensory cortex (S1, $t_{21} = -6.618$, p = 1.491e-97 06). In comparison, movement execution strongly activated M1 and S1 (Fig. 2C), with activation being significant in all our ROIs (Fig. 2E, all t_{21} > 14.469, all p < 2.153e-12). 98

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100 *Planning induces informative patterns in primary motor and somatosensory cortex*

101 Although we found little univariate activation in our main ROIs, preparatory processes need 102 not increase the overall activation in a region. Rather, the region could converge to a specific 103 preparatory neural state (26), while activity increments and decrements within the region (i.e., 104 at a finer spatial scale) average each other out. In this case, information about planned 105 movements would be present in the multivoxel activity patterns in that region. To test this 106 idea, we calculated the crossnobis dissimilarity (see Methods) between activity patterns. 107 Systematically positive values of this dissimilarity measure indicate that the patterns reliably 108 differentiate between the different planned actions (39, 40). Indeed, a surface-based 109 searchlight approach (41) revealed reliably positive crossnobis dissimilarity between the

- activity patterns related to planning of individual finger movements (Fig. 2B) in both M1 (t_{21} =
- 111 2.734, p = 0.012) and S1 ($t_{21} = 2.987$, p = 0.007, Fig. 2F).

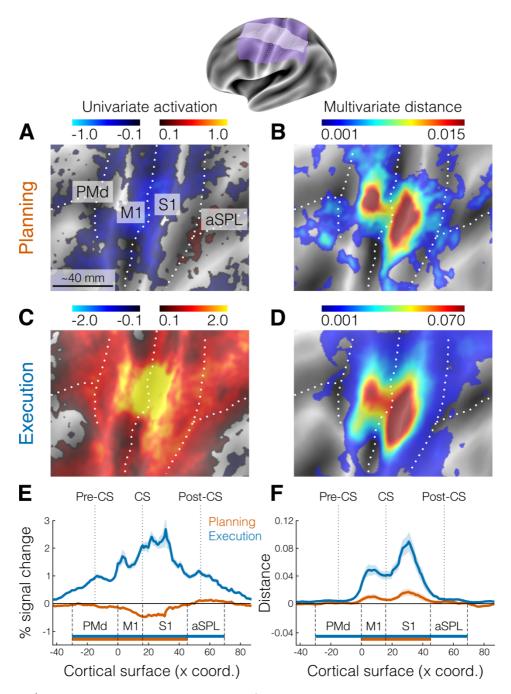


Figure 2 | Activation and distance analyses of movement planning and execution. The inset shows the inflated cortical surface of the contralateral (left) hemisphere, highlighting the area of interest (purple) and the strip used for cross-section analysis (white). Major sulci are indicated by white dotted lines. A. Univariate activation map (percent signal change) for the contrast planning>baseline (no-go trials only). B. Multivariate searchlight map of the mean crossnobis distance between the planning of the three fingers (no-go trials only). C. Same as A, but for the univariate contrast execution>baseline. D. Same as B, but for the mean crossnobis distance between fingers during movement execution. E. Cross-section analysis of the mean percent signal change (\pm SEM) during planning (orange) and execution (blue). Horizontal bars indicate significance (p < 0.05) in a 2-sided one-sample *t*-test vs zero for

selected ROIs. F. Same as E, but for the mean crossnobis distance (\pm SEM). Pre-CS = precentral sulcus; CS = central sulcus; Post-CS = postcentral sulcus; PMd = dorsal premotor cortex; M1 = primary motor cortex; S1 = primary somatosensory cortex; aSPL = anterior superior parietal lobule.

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114 Surprisingly, the distribution of these informative patterns was highly similar to the distribution 115 of information during movement execution (Fig. 2D). Visual inspection suggested that the 116 informative patterns during planning may be concentrated more dorsally in M1 and S1 relative to execution. However, a Hotelling T² test revealed no systematic difference in the 117 118 location of the peak vertex between planning and execution across subjects (M1: $T_{2,20}^2$ = 119 0.725, p = 0.712; S1: $T_{2,20}^2 = 2.424, p = 0.335$). Thus, our results show that information about 120 single finger movements is already represented during motor planning in the same parts of 121 the primary motor and somatosensory cortices that are engaged during execution of the movements. Given that we only used the activity estimates from no-go trials (~40% of total 122 123 trials), this information cannot be explained by a spill-over from subsequent execution-related 124 activity. An analysis using the estimates of planning activity from all trials yielded very similar 125 results (see Fig. S1), demonstrating that we could separate planning from execution-related 126 signals.

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128 Activity patterns are not caused by small movements during the preparation phase

129 The presence of planning-related information in primary sensorimotor regions was surprising, 130 especially in S1, where it had not previously been reported in comparable fMRI studies (18, 131 20). To ensure that these results were not caused by overt movement, participants were 132 instructed to maintain a steady force on the keyboard during the preparation phase, such 133 that we could monitor even the smallest involuntary preparatory movements. Inspection of the average force profiles (Fig. 3A) revealed that participants were successful in maintaining 134 135 a stable force between 0.2 and 0.4 N during preparation. However, averaging forces across trials may obscure small, idiosyncratic patterns visible during individual trials (Fig. 3B) that 136 137 could be used to distinguish the different movements. To test for the presence of such 138 patterns, we submitted both the mean and standard deviation of the force traces on each 139 finger to a multivariate dissimilarity analysis (see Methods).

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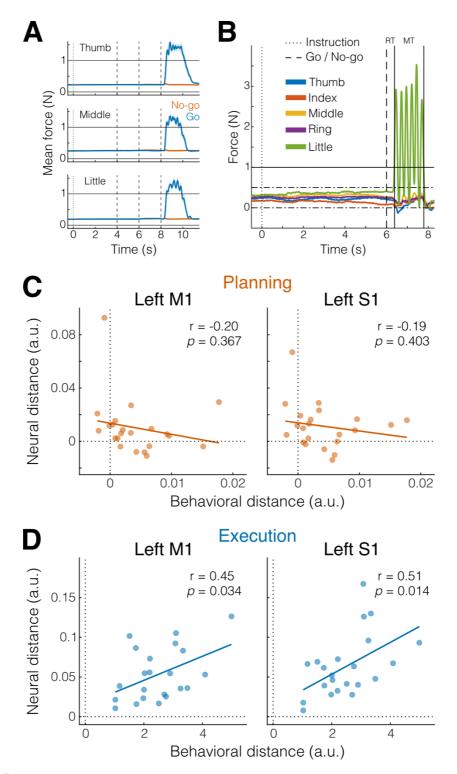


Figure 3 | Small involuntary movements do not explain preparatory activity patterns in M1 and S1. A. Mean finger force (\pm SEM) plotted in 10 ms bins, time-aligned to instruction onset (dotted vertical line) and end of the preparation phase (dashed vertical lines), separately for the three fingers and go (blue) and no-go (orange) trials. **B.** Example of an individual trial with a 6 s preparation phase, followed six presses of the little finger (green). Horizontal solid line denotes press threshold (1 N). Dash-dotted lines denote the boundaries of the finger pre-activation red area in Fig. 1A (see Methods). RT = reaction time; MT = movement time. **C.** Pearson's correlation (r) between behavioral and neural distances in M1 and S1 (see Methods) during the preparation phase (planning, orange). Each dot represents an individual participant (N = 22). Solid line shows linear regression, *p*-values refers to the slope of the line. **D.** Same as C, but during the movement phase (execution, blue).

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142 Indeed, this sensitive analysis revealed that some participants showed small movement 143 patterns predictive of the upcoming finger (Fig. 3C). These differences, however, were ~200-144 300 times smaller than the average differences during execution (Fig. 3D). More importantly, 145 the magnitude of the behavioral differences for the preparation phase was uncorrelated to 146 the amount of planning information present in sensory-motor regions (both p-values for the 147 slope of the linear fit > 0.3). Even after correcting for the influence behavioral patterns, the 148 informative patterns in M1 and S1 remained significant, as shown by a significantly positive 149 intercept in the linear fit in Fig. 3C (M1: $\rho = 0.032$; S1: $\rho = 0.007$). Thus, the finding of finger-150 specific activity patterns in M1 and S1 cannot be explained by small involuntary movements 151 during the preparation phase.

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153 Single finger activity patterns from planning to execution are positively correlated

154 How do planning-related activity patterns in M1 and S1 relate to the activity patterns observed 155 during execution? Neurophysiological experiments have suggested that patterns of 156 movement preparation are orthogonal - or uncorrelated - to the patterns underlying active 157 movement (42). This arrangement allows movement preparation to occur without causing 158 overt movement. When we compared the planning- and execution-related activity patterns 159 as measured with fMRI, a technique that samples neuronal activity at a much coarser spatial 160 resolution, we found the opposite result. Planning- and movement-related patterns for the 161 same finger were tightly related. Inspection of the representational dissimilarity matrices 162 (RDMs) for M1 and S1 (Fig. 4A), clearly shows that the biggest difference was between planning and execution patterns, which can also be seen in a 3D representation of the 163 representational geometry (PC1 in Fig. 4B). Within each phase, the pattern for the thumb was 164 165 more distinct than those for the other fingers, replicating previous results from execution 166 alone (28, 29).

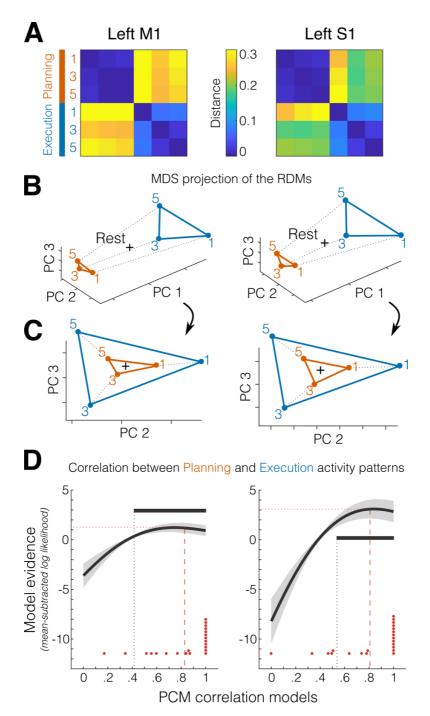


Figure 4 | Correlated representations of single fingers across planning and execution. A. Representational dissimilarity matrices (RDMs) of the activity patterns evoked by the three fingers during the preparation (no-go planning, orange) and movement (execution, blue) phases, separately for the two main ROIs (M1, left; S1, right). B. Multidimensional scaling (MDS) projection of the RDMs in A highlighting the first principal component (PC 1, difference between planning and execution). The black cross denotes resting baseline. C. Same as B, but rotated view to highlight the representational geometries for PC 2 and PC 3. D. Pattern component modelling (PCM) evaluation of models of different correlations between planning-and execution-related activity patterns (x-axis). Shown is the group-average of the individual log-likelihood (\pm SEM) curve expressed as a difference from the mean log-likelihood across models. Horizontal gray bars indicate models that perform statistically equivalently (p > 0.05) to the best fitting model (determined in a cross-validated fashion, see Methods). Red dots indicate points of individually best fitting correlations (N = 22). Red dashed lines denote the average winning models across participants. Dotted lines show the projections of horizontal bars and dashed lines on the respective axes.

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169 Importantly, however, a rotated view of the representational geometry (Fig. 4C) showed that 170 the finger patterns were arranged in a congruent way, with planning and execution related 171 activity patterns for the same finger being closer to one another. To more precisely quantify 172 the correspondence between planning and execution pattern for each finger, we used 173 Pattern Component Modeling (PCM, 43) to evaluate the likelihood of the data, assuming a 174 true correlation between 0 and 1. This approach automatically corrects for the biasing effect 175 of measurement noise, which would lead to simple empirical pattern correlations to be lower 176 than the true correlation. From the individual fits, we found that the average best correlation 177 model was at 0.83 (± 0.053 SEM) for M1 and at 0.81 (± 0.061 SEM) for S1 (Fig. 4D, dashed 178 lines). Using a cross-validated approach (see Methods), we compared the log-likelihoods to 179 test whether the best fitting model was significantly better than any of the other correlation 180 models. In both ROIs, the best fitting model was significantly better than the zero-correlation 181 model (i.e., activity patterns across planning and execution totally uncorrelated, both p < p182 0.005), but not better than the one-correlation model (i.e., activity patterns are scaled version 183 of each other, both p > 0.1). By applying this method to every other model, we have evidence that the true (i.e., noiseless) correlation between planning and execution finger-specific 184 activity pattern was between 0.41–1.0 in M1 and between 0.54–1.0 in S1 (Fig. 4D, horizontal 185 186 bars). In sum, at the resolution of fMRI, the process of movement planning seems to induce 187 a finger-specific pattern which very similar, and possibly identical, to the pattern activated 188 during movement execution.

189

190 Discussion

In the present study, we asked participants to produce repeated single finger movements while undergoing 7T fMRI. We used a variable preparatory delay and no-go trials to cleanly dissociate the brain responses to the consecutive preparation and movement phases. We found that information about planned finger movements is present in both S1 and M1 before movement onset, even though the overall level of activation in these regions was below resting baseline. Moreover, while execution elicited much higher brain activation, the finegrained, finger-specific activity patterns were highly similar across planning and execution.

198 Control analyses confirmed that the observed results were not caused by pre-movement199 finger activity.

200 Our finding that motor planning activates M1 in a finger-specific fashion was not 201 necessarily surprising given many neurophysiological studies reporting anticipatory activity 202 of M1 neurons related to movement intentions (37, 44, 45), as well as human neuroimaging 203 showing shared information between delayed and immediate movement plans (36). In 204 contrast, the robust activity patterns related to single finger planning in S1 were more 205 surprising, given that this region is often considered to be mostly concerned with processing 206 incoming sensory information from tactile and proprioceptive receptors arising after 207 movement onset (18, 20-22). So, what could then be the role of S1 during movement 208 planning?

First, it is worth noting that there are substantial projections from S1 (Brodmann area 3a) that terminate in the ventral horn of the cortico-spinal tract (46, 47). Although stimulation of area 3a in macaques typically fails to evoke overt movements (48), it has been suggested that this population of cortico-motoneurons specifically projects to gamma motoneurons that control the sensitivity of muscle spindle afferents (47). Thus, it is possible that S1 plays an active role in movement generation by preparing the spindle apparatus in advance of the movement.

Second, the finger-specific preparatory state in S1 may reflect the anticipation or prediction of the upcoming sensory stimulation, allowing for a movement-specific sensory gain control (49). Some sensory stimuli could become attenuated to maintain movement stability and filter out irrelevant or self-generated signals. Indeed, multiple studies have shown that both somatosensation and somatosensory-evoked potentials in S1 decrease during voluntary movement (50–53). Alternatively, sensory processing of the expected salient signals could be enhanced to improve movement execution and adaptation.

Very recently, a human electrocorticography (ECoG) study suggested a possible role for S1 in cognitive-motor imagery (54). The authors recorded neural activity from S1 while a tetraplegic participant imagined reaching movements and found that S1 neurons encoded movement direction during motor imagery in the absence of actual sensations. While the authors raise the concern that their results may be unique to individuals who have lost their main peripheral input, our finding of encoding of movement-related information in S1 before the onset of a movement suggests that movement-specific information in S1 without actual

movement is a general phenomenon in the intact human motor system. Our findings are also
consistent with a second recent ECoG study in non-human primates (55). During a delayed
reaching and grasping task, the authors showed movement-specific information in the ECoG
signals from S1 well before movement initiation, and only slightly later than in M1. Together,
these results suggest that the somatosensory system not only passively receives signals from
the external world but also actively processes them via interactions with anticipatory
information from the motor system.

One may wonder why such movement-specific encoding in S1 during planning has not been reported in previous human fMRI studies. One contributing reason may be that we had higher sensitivity to detect these signals than previous studies, as we used finger rather than arm movements (the former having more distinct cortical representation in S1), higher field strength (7T) and spatial resolution, and a more sensitive multivariate analysis method (crossnobis dissimilarity vs. pattern classification, Walther et al., 2016).

243 Our second main finding – the close correspondence between finger-specific activity 244 patterns across planning and execution – appears to be at odds with the idea that these two 245 processes occupy orthogonal neural subspaces to avoid overt movement during planning 246 (42, 56). We think that there are at least two possible explanations for this. First, the 247 divergence of results could be caused by the difference between spatially directed arm 248 movements and non-spatial finger movements. If for single finger movements even single-249 neuron activity patterns are highly correlated between planning and execution, then overt 250 movement during planning would need to be actively suppressed, for example through the 251 deactivation that we observed around the central sulcus. An alternative and perhaps more 252 likely explanation of the discrepancy lies in the different measurement modalities. While the 253 orthogonality was observed in electrophysiological recordings of individual neurons, the fMRI 254 measurements we employed here mainly reflect excitatory postsynaptic potentials (57) and average metabolic activity across hundreds of thousands of cortical neurons. Thus, it is 255 256 possible that planning pre-activates the specific cortical columns in M1 and S1 that are also 257 most active during movement of that finger. Within each of these cortical micro-circuits, 258 however, planning-related activity could still be orthogonal to the activity observed during 259 execution at the single neuron level (e.g., see Arbuckle et al., 2020, for a similar observation 260 for cortical representations of flexion and extension finger movements). This would suggest 261 a new hypothesis for the architecture of the sensory-motor system where movement planning

- 262 pre-activates the movement-specific circuits in M1 and S1. However, it does so in a fashion
- 263 that the induced planning-related activity is, in terms of the firing output of neurons,
- 264 orthogonal to the patterns during movement execution.
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266 Methods

267 *Participants*

268 Twenty-three right-handed neurologically healthy participants volunteered to take part in the 269 experiment (13 F, 10 M; age 20-31 years, mean 23.43 years, SD 4.08 years). Criteria for 270 inclusion were right-handedness and no prior history of psychiatric or neurological disorders. Handedness was assessed with the Edinburgh Handedness Inventory (mean 82.83, SD 271 272 9.75). All experimental procedures were approved by the Research Ethics Committee at 273 Western University. Participants provided written informed consent to procedures and data 274 usage and received monetary compensation for their participation. One participant withdrew 275 before study completion and was excluded from data analysis (final N = 22).

276

277 Apparatus

278 Repeated presses of right-hand finger movements were performed on a custom-made MRI-279 compatible keyboard device (Fig. 1A). The keys of the device did not move but force 280 transducers underneath each key measured isometric force production at an update rate of 281 2 ms (Honeywell FS series; dynamic range 0-25 N; sampling 200 Hz). A keypress/release 282 was detected when the force crossed a threshold of 1 N. The forces measured from the 283 keyboard were low-pass filtered to reduce noise induced by the MRI environment, amplified, 284 and sent to PC for online task control and data recording.

285

286 *Task*

We used a task in which participants produced repeated keypresses with their right-hand fingers in response to numerical cues appearing on the screen (white outline, Fig. 1A). On each trial, a string of 6 numbers (instruction) instructed which fingers to plan (1 = thumb, 3 = middle, 5 = little). After a variable delay (4, 6, or 8 s randomly sampled from a geometric distribution with p = 0.5; preparation phase, yellow background), participants received a color cue (go/no-go cue) indicating whether to perform the planned finger movements (blue 293 outline = go, p = 0.6), or not (orange outline = no-go, p = 0.4). The role of no-go trials was to 294 de-couple the hemodynamic response to the successive planning and execution events. To 295 encourage planning during the delay period, at the go/no-go cue the digits were masked 296 with asterisks, and participants had to perform the movements from memory (movement 297 phase, green background). Participants had 2.5 seconds to complete the movement phase, 298 and a vanishing white bar under the asterisks indicated how much time was left to complete 299 all of the keypresses. To limit and monitor unwanted movements during the preparation 300 phase, we required participants to pre-activate their fingers by maintaining a steady force of 301 around 0.2-0.3 N on all of the keys during the Preparation phase. As a visual aid, we 302 displayed a red area (between 0 and 0.5 N) and asked participants to remain within the 303 middle of that range with all the fingers (touching either boundary of the red area would incur 304 an error, counting as unwanted movement). In the case of a no-go trial, participants were 305 instructed to remain as still as possible maintaining the finger pre-activation until the end of 306 the movement phase (i.e., releasing any of the keys would incur an error). During the 307 movement phase participants also received online feedback on the correctness of each 308 press with asterisks turning either green, for a correct press, or red, for incorrect presses. 309 After the movement phase, participants received points based on their performance (reward 310 phase, 0.5 s, purple background). Participants were instructed to perform the movements as 311 accurately as possible. As long as they remained within task constraints (i.e., 6 keypresses 312 in less than 2.5 seconds), an exact movement speed was not enforced. On a trial-by-trial 313 basis, during the reward phase participants received points for their performance according 314 to the following scheme: -1 point in case of no-go error or go cue anticipation (timing errors); 315 0 points for pressing any wrong key (press error); 1 point in case of a correct no-go trial; and 316 2 points in case of a correct go trial. Inter-trial-intervals (ITIs) varied between 1 and 16 317 seconds within the domain $ITI = \{1, 2, 4, 8, 16\}$. To reduce the overlap in brain responses from one trial to the next, actual ITIs were randomly picked from a geometric distribution with 318 319 p = 0.5. This meant a higher probability of shorter intervals (1 and 2 s) and occasional very 320 long intervals (8 and 16 s). The design was optimized to minimize the variance inflation factor 321 (VIF):

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VIF = var(E) / var(X),

the ratio of the mean estimation variance of all the regression weights (planning- and execution-related regressors for each finger), and the mean estimation variance had these regressors been estimated in isolation. For our design, the average VIF was quite low, on average between 1 and 1.2, indicating that we could separate planning and execution related activity without a large loss of experimental power.

329

330 Design

331 Participants underwent one fMRI session consisting of 10 functional runs and 1 anatomical 332 scan. In an event-related design, we randomly interleaved 3 types of repeated single finger 333 movements involving the thumb (1), the middle (3), and the little (5) fingers (e.g., 111111 for 334 thumb movement, Fig. 1A) and 3 types of multi finger sequences (e.g., 135315). The day 335 before the fMRI scan, participants familiarized themselves with the experimental apparatus 336 and the go/no-go paradigm in a short behavioral session of practice outside the scanner (5 337 blocks, about 15-30 min in total). For the behavioral practice, inter-trial intervals were kept to 338 a fixed 1 s to speed up the task, and participants were presented with different sequences 339 from what they would see while in the scanner. These 6-item sequences were randomly 340 selected from a pool of all possible permutations of the numbers 1, 3, and 5, with the 341 exclusion of sequences that contained consecutive repetitions of the same number. Given 342 that the current paper is concerned with the relationship between representations of simple 343 planning and execution, here we will focus only on the results for single finger movements. 344 The results for multi finger sequences will be reported in a future paper. Each single finger 345 trial type (e.g., 111111) was repeated 5 times (2 no-go and 3 go trials), totalling 30 trials per 346 functional run. Two periods of 10 s rests were added at the beginning and at the end of each functional run to allow for signal relaxation and provide a better estimate of baseline 347 348 activation. Each functional run took about 5.5 minutes and the entire scanning session 349 (including the anatomical scan and setup time) lasted for about 75 minutes.

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351 Imaging data acquisition

High-field functional magnetic resonance imaging (fMRI) data were acquired on a 7T
Siemens Magnetom scanner with a 32-channel head coil at Western University (London
Ontario, Canada). The anatomical T1-weighted scan of each participant was acquired
halfway through the scanning session (after the first 5 functional runs) using a Magnetization-

356 Prepared Rapid Gradient Echo sequence (MPRAGE) with voxel size of 0.75x0.75x0.75 mm 357 isotropic (field of view = 208 x 157 x 110 mm [A-P; R-L; F-H], encoding direction coronal). To 358 measure the blood-oxygen-level dependent (BOLD) responses in human participants, each 359 functional scan (330 volumes) used the following sequence parameters: GRAPPA 3, multi-360 band acceleration factor 2, repetition time [TR] = 1.0 s, echo time [TE] = 20 ms, flip angle 361 [FA] = 30 deg, slice number: 44, voxel size: 2x2x2 mm isotropic. To estimate and correct for magnetic field inhomogeneities, we also acquired a gradient echo field map with the following 362 363 parameters: transversal orientation, field of view: 210 x 210 x 160 mm, 64 slices, 2.5 mm 364 thickness, TR = 475 ms, TE = 4.08 ms, FA = 35 deg.

365

366 *Preprocessing and univariate analysis*

367 Preprocessing of the functional data was performed using SPM12 (fil.ion.ucl.ac.uk/spm) and custom MATLAB code. This included correction for geometric distortions using the gradient 368 369 echo field map (58), and motion realignment to the first scan in the first run (3 translations: x, 370 y, z; 3 rotations: pitch, roll yaw). Due to the short TR, no slice timing corrections were applied. 371 The functional data were co-registered to the anatomical scan, but no normalization to a 372 standard template or smoothing was applied. To allow magnetization to reach equilibrium, 373 the first four volumes of each functional run were discarded. The pre-processed images were 374 analyzed with a general linear model (GLM). We defined separate regressors for each 375 combination of the 6 finger movements (single, multi) x 2 phases (preparation, movement). To control for the effect of potential spill-over of movement execution activity on the preceding 376 377 planning activity, we also estimated a separate GLM with separate regressors for the 378 preparation phases of go and no-go trials, resulting in a total of 18 regressors (12 go + 6 no-379 go), plus the intercept, for each run. Each regressor consisted of a boxcar function (on for 2 380 s of each phase duration and off otherwise) convolved with a two-gamma canonical hemodynamic response function with a peak onset at 5 s and a post-stimulus undershoot 381 382 minimum at 10 s (Fig. 1B). Given the relatively low error rates (single: 8.51 ± 1.52 %, multi: 383 17.21 ± 3.38 %; timing errors, single: 7.58 \pm 0. 62 %, multi: 10.23 \pm 0.85 %; press errors, 384 single: 1.18 ± 0.26 %, multi: 9.04 ± 1.03 %), all trials were included to estimate the regression 385 coefficients, regardless of whether the execution was correct or erroneous. Ultimately, the 386 first-level analysis resulted in activation images (beta maps) for each of the 18 conditions per 387 run, for each of the participants.

388

389 Surface reconstruction and ROI definition

390 Individual subject's cortical surfaces were reconstructed using Freesurfer (59). First, we 391 extracted the white-gray matter and pial surfaces from each participant's anatomical image. 392 Next, we inflated each surface into a sphere and aligned it using sulcal depth and curvature 393 information to the Freesurfer average atlas (Fischl et al., 1999). Both hemispheres in each 394 participant were then resampled into Workbench's 164k vertex grid. This allowed us to 395 compare similar areas of the cortical surface in each participant by selecting the 396 corresponding vertices on the group atlas. Anatomical regions of interest (ROIs) were 397 defined using a probabilistic cytoarchitectonic atlas (61) projected onto the common group 398 surface. Our main ROIs were defined bilaterally as follows: primary motor cortex (M1) was 399 defined by including nodes with the highest probability of belonging to Brodmann area (BA) 400 4 within 2 cm above and below the hand knob anatomical landmark (62); primary 401 somatosensory cortex (S1) was defined by the nodes related to BA 1, 2 and 3; dorsal 402 premotor cortex (PMd) was defined as the lateral part of the middle frontal gyrus; finally, the 403 anterior part of the superior parietal lobule (aSPL) included areas anterior, superior and ventral to the intraparietal sulcus (IPS). ROI definition was carried out separately in each 404 405 subject using FSL's subcortical segmentation. When resampling functional onto the surface, 406 to avoid contamination between M1 and S1 activities, we excluded voxels with more than 407 25% of their volume in the grey matter on the opposite side of the central sulcus.

408

409 *Multivariate distance analysis*

To detect single finger representations across the cortical surface, we used representational 410 411 similarity analysis (RSA; Diedrichsen and Kriegeskorte, 2017; Walther et al., 2016) with a 412 surface-based searchlight approach (64). For each node, we selected a region (the 413 searchlight) corresponding to 100 voxels (12 mm disc radius) in the gray matter and 414 computed cross-validated Mahalanobis (crossnobis, Walther et al., 2016) dissimilarities 415 between pairs of evoked activity patterns (beta estimates from first level GLM) of single finger 416 sequences, during both preparation and movement phases. Prior to calculating the dissimilarities, beta weights for each condition were spatially pre-whitened (i.e., weighted by 417 418 the matrix square root of the noise covariance matrix, as estimated from the residuals of the 419 GLM. The noise covariance matrix was slightly regularized towards a diagonal matrix (65).

420 Multivariate pre-whitening has been shown to increase the reliability of dissimilarity estimates 421 (39). The resulting analyses (one RDM per participant containing the dissimilarities between 422 the three single fingers during planning and execution: 6 conditions, 15 dissimilarity pairs) 423 were then assigned to the central node and the searchlight was moved across all nodes 424 across the surface sheet obtaining a cortical map (Fig. 2B-2D). Cross-validation ensures the distances estimates are unbiased, such that if two patterns differ only by measurement noise, 425 426 the mean of the estimated value would be zero. This also means that estimates can 427 sometimes become negative. Therefore, dissimilarities significantly larger than zero indicate 428 that two patterns are reliably distinct, similar to an above-chance performance in a cross-429 validated pattern-classification analysis. Additionally, to the searchlight analysis, the 430 multivariate analysis was conducted separately for each anatomically defined ROI (e.g., Fig. 431 4A).

432

433 Correlation between behavioral and neural distances

434 To ensure that our planning results were not contaminated by unwanted micro-movements 435 during the preparation phase, we calculated the behavioral distance between sequences on 436 the basis of keyboard force data and correlated behavioral and neural distances. For 437 behavioral distances, we first extracted force data (2 ms temporal resolution, smoothed with 438 a gaussian kernel of 9.42 full width at half maximum, FWHM) and binned it in 10 ms steps 439 (down sampling largely due to memory constraints) for both the preparation and movement 440 phases (Fig. 3A). Next, for each subject, we calculated the mean (5) and the standard 441 deviation (5) of the time-averaged force of each finger for each condition (3 sequences x 2 442 phases = 6) and block (10). These subject-specific finger force patterns (60 x 10) were multivariately pre-whitened using their covariance matrix. Finally, we calculated the cross-443 444 validated squared Euclidean distances for each condition (6 x 6 RDM) and averaged distances between the 3 finger movements for each phase (preparation, movement). These 445 446 mean finger force distances for each subject were correlated with the mean voxel activity 447 distances from the two phases for 2 ROIs (M1 and S1, Fig. 3C-3D).

448

449 Pattern component modelling correlation models

We used pattern component modelling (PCM) to quantify the correspondence of sequence-specific activity patterns across planning and execution (43). This method has been shown

452 to be advantageous in estimating correlations. In contrast to simple Pearson's or cross-453 validated correlation estimated from raw activity patterns, PCM separately models the noise 454 and signal components. We created 100 correlation models with correlations in the range [0-455 1] in equal step sizes and assessed the likelihood of the observed data from each participant 456 under each model. Fig. 4D shows average log-likelihoods for each model, relative to the 457 mean log-likelihood across models. Differences between the log-likelihoods can be 458 interpreted as log-Bayes factors. Group inferences were performed using a simple t-test on 459 log-likelihoods. To compare each model to the best fitting model, we had to correct for the 460 bias arising from picking the best model and testing it on the same data: We used n-1 461 subjects to determine the group winning model, and then chose the log-likelihood of this 462 model for the left-out subject (for whom this model may not be the best one) as the likelihood 463 for the "best" model. This was repeated across all subjects and a one-sided paired-sample 464 *E*test was performed on the recorded log-likelihood and every other model. This test revealed which of the correlation models were significantly worse (i.e., associated with a lower log-465 likelihood) than the winning model that was independently estimated via cross-validation. 466

468 References

469 1. S. W. Keele, J. J. Summers, "The Structure of Motor Programs" in *Motor Control*, 470 (Elsevier, 1976), pp. 109-142. S. W. Keele, Movement control in skilled motor performance. Psychological Bulletin 471 2. 472 70, 387–403 (1968). 473 3. D. A. Rosenbaum, Human movement initiation: Specification of arm, direction, and 474 extent. Journal of Experimental Psychology: General 109, 444–474 (1980). 475 4. S. T. Klapp, Motor Response Programming During Simple and Choice Reaction 476 Time: The Role of Practice. Journal of Experimental Psychology: Human Perception 477 and Performance 21, 1015–1027 (1995). 5. 478 S. T. Klapp, C. I. Erwin, Relation between programming time and duration of the response being programmed. Journal of Experimental Psychology: Human 479 480 *Perception and Performance* **2**, 591–598 (1976). A. M. Haith, J. Pakpoor, J. W. Krakauer, Independence of Movement Preparation and 481 6. 482 Movement Initiation. Journal of Neuroscience 36, 3007–3015 (2016). 483 7. G. Ariani, J. Diedrichsen, Sequence learning is driven by improvements in motor planning. Journal of Neurophysiology 121, 2088–2100 (2019). 484 8. 485 A. L. Wong, A. M. Haith, Motor planning flexibly optimizes performance under 486 uncertainty about task goals. Nature Communications 8, 14624 (2017). R. M. Hardwick, A. D. Forrence, J. W. Krakauer, A. M. Haith, Time-dependent 487 9. 488 competition between goal-directed and habitual response preparation. Nature 489 Human Behaviour 3, 1252–1262 (2019). 490 10. C. Ghez, et al., Discrete and continuous planning of hand movements and isometric 491 force trajectories. Experimental Brain Research 115, 217–233 (1997). 492 P. Cisek, J. F. Kalaska, Neural correlates of mental rehearsal in dorsal premotor 11. 493 cortex. Nature 431, 993-6 (2004). 494 12. P. Cisek, J. F. Kalaska, Neural mechanisms for interacting with a world full of action choices. Annual review of neuroscience 33, 269-298 (2010). 495 13. E. Hoshi, J. Tanji, Differential involvement of neurons in the dorsal and ventral 496 497 premotor cortex during processing of visual signals for action planning. Journal of 498 neurophysiology 95, 3596-616 (2006).

- 499 14. E. Hoshi, J. Tanji, Differential roles of neuronal activity in the supplementary and
 500 presupplementary motor areas: from information retrieval to motor planning and
 501 execution. *Journal of neurophysiology* 92, 3482–99 (2004).
- 502 15. R. A. Andersen, H. Cui, Intention, Action Planning, and Decision Making in Parietal503 Frontal Circuits. *Neuron* 63, 568–83 (2009).
- 504 16. H. Cui, R. A. Andersen, Posterior Parietal Cortex Encodes Autonomously Selected
 505 Motor Plans. *Neuron* 56, 552–559 (2007).
- 506 17. H. Cui, R. A. Andersen, Different Representations of Potential and Selected Motor
 507 Plans by Distinct Parietal Areas. *Journal of Neuroscience* 31, 18130–18136 (2011).
- 508 18. J. P. Gallivan, D. A. McLean, K. F. Valyear, C. E. Pettypiece, J. C. Culham, Decoding
- Action Intentions from Preparatory Brain Activity in Human Parieto-Frontal Networks. *Journal of Neuroscience* 31, 9599–9610 (2011).
- 511 19. G. Ariani, M. F. Wurm, A. Lingnau, Decoding Internally and Externally Driven
 512 Movement Plans. *Journal of Neuroscience* 35, 14160–14171 (2015).
- 513 20. J. P. Gallivan, I. S. Johnsrude, J. R. Flanagan, Planning Ahead: Object-Directed
 514 Sequential Actions Decoded from Human Frontoparietal and Occipitotemporal
 515 Networks. *Cerebral Cortex* 26, bhu302 (2015).
- 516 21. J. P. Gallivan, D. A. McLean, F. W. Smith, J. C. Culham, Decoding Effector-
- 517 Dependent and Effector-Independent Movement Intentions from Human Parieto-

518 Frontal Brain Activity. *Journal of Neuroscience* **31**, 17149–17168 (2011).

- 519 22. J. P. Gallivan, D. A. McLean, J. R. Flanagan, J. C. Culham, Where One Hand Meets
- 520 the Other: Limb-Specific and Action-Dependent Movement Plans Decoded from
- 521 Preparatory Signals in Single Human Frontoparietal Brain Areas. *Journal of*522 *Neuroscience* 33, 1991–2008 (2013).
- 523 23. L. Turella, *et al.*, Beta band modulations underlie action representations for
 524 movement planning. *NeuroImage* 136, 197–207 (2016).
- 525 24. F. T. M. Leoné, T. Heed, I. Toni, W. P. Medendorp, Understanding effector selectivity
- in human posterior parietal cortex by combining information patterns and activation
 measures. *The Journal of neuroscience* 34, 7102–12 (2014).
- 528 25. S. Vyas, M. D. Golub, D. Sussillo, K. V Shenoy, Computation Through Neural
- 529 Population Dynamics. *Annual Review of Neuroscience* **43**, 249–275 (2020).
- 530 26. M. M. Churchland, J. P. Cunningham, M. T. Kaufman, S. I. Ryu, K. V. Shenoy,

531		Cortical Preparatory Activity: Representation of Movement or First Cog in a
532		Dynamical Machine? <i>Neuron</i> 68 , 387–400 (2010).
533	27.	K. V Shenoy, M. Sahani, M. M. Churchland, Cortical control of arm movements: a
534	21.	dynamical systems perspective. <i>Annual Review of Neuroscience</i> 36 , 337–359
		(2013).
535	20	
536	28.	N. Ejaz, M. Hamada, J. Diedrichsen, Hand use predicts the structure of
537	00	representations in sensorimotor cortex. <i>Nature Neuroscience</i> 18 , 1034–1040 (2015).
538	29.	A. Yokoi, S. A. Arbuckle, J. Diedrichsen, The role of human primary motor cortex in
539		the production of skilled finger sequences. <i>Journal of Neuroscience</i> 38 , 1430–1442
540		(2018).
541	30.	J. Diedrichsen, T. Wiestler, J. W. Krakauer, Two distinct ipsilateral cortical
542		representations for individuated finger movements. Cerebral Cortex 23, 1362–1377
543		(2013).
544	31.	T. Wiestler, S. Waters-Metenier, J. Diedrichsen, Effector-independent motor
545		sequence representations exist in extrinsic and intrinsic reference frames. Journal of
546		<i>Neuroscience</i> 34 , 5054–5064 (2014).
547	32.	A. Yokoi, J. Diedrichsen, Neural Organization of Hierarchical Motor Sequence
548		Representations in the Human Neocortex. Neuron, 1–13 (2019).
549	33.	S. A. Arbuckle, et al., Structure of population activity in primary motor cortex for
550		single finger flexion and extension. The Journal of Neuroscience, JN-RM-0999-20
551		(2020).
552	34.	S. A. Arbuckle, A. Yokoi, J. A. Pruszynski, J. Diedrichsen, Stability of representational
553		geometry across a wide range of fMRI activity levels. Neurolmage 186, 155–163
554		(2019).
555	35.	L. Huber, et al., Sub-millimeter fMRI reveals multiple topographical digit
556		representations that form action maps in human motor cortex. NeuroImage 208,
557		116463 (2020).
558	36.	G. Ariani, N. N. Oosterhof, A. Lingnau, Time-resolved decoding of planned delayed
559		and immediate prehension movements. Cortex 99 (2018).
560	37.	J. Tanji, E. V. Evarts, Anticipatory activity of motor cortex neurons in relation to
561		direction of an intended movement. Journal of Neurophysiology 39, 1062–1068
562		(1976).

- 563 38. D. J. Crammond, J. F. Kalaska, Prior information in motor and premotor cortex:
- activity during the delay period and effect on pre-movement activity. *Journal of neurophysiology* 84, 986–1005 (2000).
- 566 39. A. Walther, *et al.*, Reliability of dissimilarity measures for multi-voxel pattern analysis.
 567 *NeuroImage* 137, 188–200 (2016).
- 568 40. J. Diedrichsen, *et al.*, Comparing representational geometries using whitened
 569 unbiased-distance-matrix similarity (2020).
- 570 41. N. N. Oosterhof, *et al.*, Surface-Based Information Mapping Reveals Crossmodal
- 571 Vision Action Representations in Human Parietal and Occipitotemporal Cortex
- 572 Surface-Based Information Mapping Reveals Crossmodal Vision Action
- 573 Representations in Human Parietal and Occipitotemporal. *Journal of neurophysiology*574 104, 1077–89 (2012).
- 575 42. M. T. Kaufman, M. M. Churchland, S. I. Ryu, K. V Shenoy, Cortical activity in the null
 576 space: permitting preparation without movement. *Nature Neuroscience* 17, 440–448
 577 (2014).
- 578 43. J. Diedrichsen, A. Yokoi, S. A. Arbuckle, Pattern component modeling: A flexible
 579 approach for understanding the representational structure of brain activity patterns.
 580 *NeuroImage* 180, 119–133 (2018).
- 581 44. G. E. Alexander, M. D. Crutcher, Neural representations of the target (goal) of
 582 visually guided arm movements in three motor areas of the monkey. *Journal of*583 *Neurophysiology* 64, 164–178 (1990).
- 584 45. A. Riehle, J. Requin, Monkey primary motor and premotor cortex: single-cell activity
 585 related to prior information about direction and extent of an intended movement.
 586 *Journal of neurophysiology* 61, 534–49 (1989).
- 587 46. J. D. Coulter, E. G. Jones, Differential distribution of corticospinal projections from
 588 individual cytoarchitectonic fields in the monkey. *Brain Research* 129, 335–340
 589 (1977).
- 590 47. J.-A. Rathelot, P. L. Strick, Muscle representation in the macaque motor cortex: An
 591 anatomical perspective. *Proceedings of the National Academy of Sciences* 103,
 592 8257 LP 8262 (2006).
- 593 48. G. L. Widener, P. D. Cheney, Effects on Muscle Activity From Microstimuli Applied to594 Somatosensory and Motor Cortex During Voluntary Movement in the Monkey. *Journal*

595		of Neurophysiology 77 , 2446–2465 (1997).
596	49.	E. Azim, K. Seki, Gain control in the sensorimotor system. Current Opinion in
597		<i>Physiology</i> 8 , 177–187 (2019).
598	50.	A. Starr, L. G. Cohen, 'Gating' of somatosensory evoked potentials begins before the
599		onset of voluntary movement in man. Brain Research 348, 183–186 (1985).
600	51.	C. E. Chapman, M. C. Bushnell, D. Miron, G. H. Duncan, J. P. Lund, Sensory
601		perception during movement in man. Experimental brain research 68, 516–524
602		(1987).
603	52.	W. Jiang, Y. Lamarre, C. E. Chapman, Modulation of cutaneous cortical evoked
604		potentials during isometric and isotonic contractions in the monkey. Brain Research
605		536 , 69–78 (1990).
606	53.	K. Seki, E. E. Fetz, Gating of sensory input at spinal and cortical levels during
607		preparation and execution of voluntary movement. Journal of Neuroscience 32, 890–
608		902 (2012).
609	54.	M. Jafari, et al., The human primary somatosensory cortex encodes imagined
610		movement in the absence of sensory information. Communications biology 3 , 1–7
611		(2020).
612	55.	T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about
612 613	55.	
	55. 56.	T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about
613		T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019).
613 614		 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham,
613 614 615		 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor
613 614 615 616	56.	 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor cortex. <i>Nature Communications</i>, 13239 (2016).
613 614 615 616 617	56.	 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor cortex. <i>Nature Communications</i>, 13239 (2016). N. K. Logothetis, J. Pauls, M. Augath, T. Trinath, A. Oeltermann, Neurophysiological
613 614 615 616 617 618	56. 57.	 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor cortex. <i>Nature Communications</i>, 13239 (2016). N. K. Logothetis, J. Pauls, M. Augath, T. Trinath, A. Oeltermann, Neurophysiological investigation of the basis of the fMRI signal. <i>Nature</i> 412, 150–7 (2001).
613 614 615 616 617 618 619	56. 57.	 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor cortex. <i>Nature Communications</i>, 13239 (2016). N. K. Logothetis, J. Pauls, M. Augath, T. Trinath, A. Oeltermann, Neurophysiological investigation of the basis of the fMRI signal. <i>Nature</i> 412, 150–7 (2001). C. Hutton, <i>et al.</i>, Image Distortion Correction in fMRI: A Quantitative Evaluation.
613 614 615 616 617 618 619 620	56. 57. 58.	 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor cortex. <i>Nature Communications</i>, 13239 (2016). N. K. Logothetis, J. Pauls, M. Augath, T. Trinath, A. Oeltermann, Neurophysiological investigation of the basis of the fMRI signal. <i>Nature</i> 412, 150–7 (2001). C. Hutton, <i>et al.</i>, Image Distortion Correction in fMRI: A Quantitative Evaluation. <i>NeuroImage</i> 16, 217–240 (2002).
 613 614 615 616 617 618 619 620 621 	56. 57. 58.	 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor cortex. <i>Nature Communications</i>, 13239 (2016). N. K. Logothetis, J. Pauls, M. Augath, T. Trinath, A. Oeltermann, Neurophysiological investigation of the basis of the fMRI signal. <i>Nature</i> 412, 150–7 (2001). C. Hutton, <i>et al.</i>, Image Distortion Correction in fMRI: A Quantitative Evaluation. <i>NeuroImage</i> 16, 217–240 (2002). A. M. Dale, B. Fischl, M. I. Sereno, Cortical Surface-Based Analysis: I. Segmentation
 613 614 615 616 617 618 619 620 621 622 	56. 57. 58.	 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor cortex. <i>Nature Communications</i>, 13239 (2016). N. K. Logothetis, J. Pauls, M. Augath, T. Trinath, A. Oeltermann, Neurophysiological investigation of the basis of the fMRI signal. <i>Nature</i> 412, 150–7 (2001). C. Hutton, <i>et al.</i>, Image Distortion Correction in fMRI: A Quantitative Evaluation. <i>NeuroImage</i> 16, 217–240 (2002). A. M. Dale, B. Fischl, M. I. Sereno, Cortical Surface-Based Analysis: I. Segmentation and Surface Reconstruction. <i>NeuroImage</i> 9, 179–194 (1999).
 613 614 615 616 617 618 619 620 621 622 623 	56. 57. 58.	 T. Umeda, T. Isa, Y. Nishimura, The somatosensory cortex receives information about motor output. <i>Science Advances</i> 5 (2019). G. F. Elsayed, A. H. Lara, M. T. Kaufman, M. M. Churchland, J. P. Cunningham, Reorganization between preparatory and movement population responses in motor cortex. <i>Nature Communications</i>, 13239 (2016). N. K. Logothetis, J. Pauls, M. Augath, T. Trinath, A. Oeltermann, Neurophysiological investigation of the basis of the fMRI signal. <i>Nature</i> 412, 150–7 (2001). C. Hutton, <i>et al.</i>, Image Distortion Correction in fMRI: A Quantitative Evaluation. <i>NeuroImage</i> 16, 217–240 (2002). A. M. Dale, B. Fischl, M. I. Sereno, Cortical Surface-Based Analysis: I. Segmentation and Surface Reconstruction. <i>NeuroImage</i> 9, 179–194 (1999). B. Fischl, M. I. Sereno, R. B. H. Tootell, A. M. Dale, High-resolution intersubject

627		<i>Cortex</i> 18 , 1973–1980 (2008).
628	62.	T. Yousry, Localization of the motor hand area to a knob on the precentral gyrus. A
629		new landmark. <i>Brain</i> 120 , 141–157 (1997).
630	63.	J. Diedrichsen, N. Kriegeskorte, Representational models: A common framework for
631		understanding encoding, pattern-component, and representational-similarity
632		analysis. PLOS Computational Biology 13, e1005508 (2017).
633	64.	N. N. Oosterhof, T. Wiestler, P. E. Downing, J. Diedrichsen, A comparison of volume-
634		based and surface-based multi-voxel pattern analysis. NeuroImage 56, 593–600
635		(2011).
636	65.	O. Ledoit, M. Wolf, Honey, I shrunk the sample covariance matrix. The Journal of
637		<i>Portfolio Management</i> 30 , 110–119 (2004).

639 Supplementary figures

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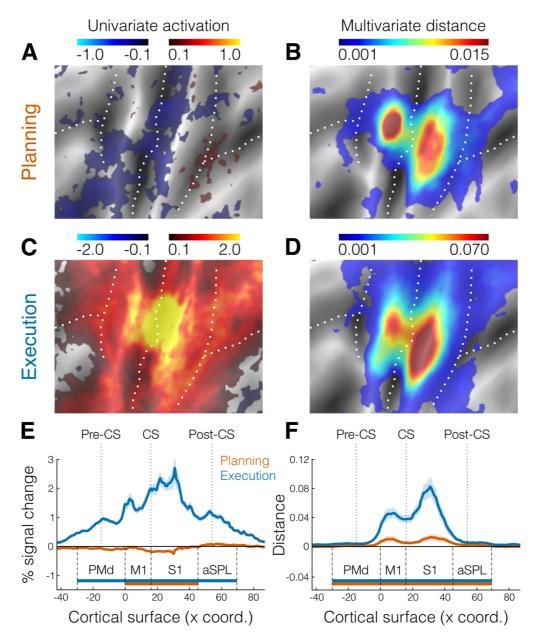


Figure S1 | Activation and distance analyses using planning of both go and no-go trials. A. Activation map (percent signal change) for the contrast planning>baseline. The selected area of interest is the same as shown in purple in the inset of Fig. 2A. B. Crossnobis distance searchlight map for movement planning. C. Same as A, but for the contrast execution>baseline. D. Same as B, but for movement execution. E-F. Cross-section analysis corresponding to the same area shown in white in the inset of Fig. 2A. E. Mean percent signal change (\pm SEM) during planning (orange) and execution (blue). F. mean crossnobis distance (\pm SEM). Horizontal bars indicate significance (p < 0.05) in a 2-sided one-sample *t*-test against zero. All other figure conventions are the same as in Fig. 2.