1   # Sensitive detection of DNA contamination in tumor samples via microhaplotypes

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Abstract

Low levels of sample contamination with other human DNAs can have disastrous effects on the accurate identification of somatic variation in tumor samples. Detection of sample contamination in DNA is often based on low frequency variants that indicate if more than a single source of DNA is present. This strategy works with standard DNA samples but can be problematic in solid tumor FFPE samples because there are often huge variations in allele frequency (AF) due to copy number changes arising from gains and losses across the genome. The variable AFs make detection of contamination challenging. To avoid this, we counted microhaplotypes to assess sample contamination. Microhaplotypes are sets of variants on the same sequencing read that can be unambiguously phased. Instead of measuring AF, the number of microhaplotypes is determined. Contamination detection becomes based on fundamental genomic properties, linkage disequilibrium (LD) and the diploid nature of human DNA, rather than variant frequencies. We optimized microhaplotype panel content and selected 164 SNV sets located in regions already being sequenced within a cancer panel. Thus, contamination detection uses existing sequence data. LD data from the 1000 Genomes Project is used to make the panel ancestry agnostic, providing the same sensitivity for contamination detection with samples from individuals of African, East Asian, and European ancestry. Detection of 1% contamination with no matching normal sample is possible. The methods described here can also be extended to other DNA mixtures such as forensic and non-invasive prenatal testing samples where DNA mixes can be similarly detected. The microhaplotype method allows sensitive detection of DNA contamination in FFPE tumor and other samples when deep coverage with Illumina or other high accuracy NGS is used.

Introduction

In many applications, DNA contamination is not a significant issue because small amounts of extraneous DNA do not affect the experimental outcome. When examining DNA from a single species, it is easy to filter out sequences from other species but within-species contamination is more challenging. When analyzing germ-line variants, a substantial fraction of human DNA from other sources is required to affect variant calls because non-reference calls will be approximately 0%, 50% or 100%. For example, concordance between variant calls made by SNP chips versus pure NGS for high depth samples with 5% contamination fell only from 99.7% to 99.2% (1). However, even with low-level DNA contamination, there can be a significant impact on some applications like detecting low frequency somatic variants in tumor DNA samples or deconvoluting complex mixtures in forensic samples. Clinical diagnostic samples are presumed to be pure when they are tested, but there is always a concern that unexpected contamination could affect results and escape standard detection methods. Knowledge of whether a sample is contaminated with another and, if so, the extent of that contamination, is critical for assessing whether somatic variant calls can be trusted.

Contamination detection is often carried out by measuring variable lengths of short tandem repeats (STRs) or by genotyping/sequencing single-nucleotide variants (SNVs). When unusual frequencies are observed, the degree of contamination is estimated by quantitating the frequency of the extra variants (2-5). However, accurate estimation becomes difficult at low variant frequency because the true signal can be obscured by the competing noise arising from

technical issues like STR stutter or genotyping/sequencing errors. This is especially problematic

with analog measurement systems that are often used with STR, genotyping, and classical

sequencing methods. In contrast, next-generation sequencing (NGS) methods produce a digital

signal, allowing greater sensitivity and accuracy at low contamination levels.

In addition to the method described here, there are informatic methods that make use of

different types of NGS read data to model whether contamination is likely. These methods are

often tied to particular data and/or sample types so may not extrapolate well to other systems

like clinical test samples. For example, ContEst (2) is highly sensitive but uses both whole

genome and microarray data or matched normal samples. This has recently been replaced by

CalculateContamination that relaxes copy number and matched sample requirements.

Verifybamid (6) can be used with just NGS data, but its accuracy is dependent on large numbers

of SNVs and knowledge of population SNV frequencies and cannot be used with DNA with large

copy number changes. It is also computationally intensive; but, VerifyBamID2 was recently

developed to address these issues (7). Conpair (8) is much less computationally intensive but

still requires large numbers of SNVs as well as tumor-normal pairs. It can be used with exomes

or large panels but not smaller panels. Sensitive detection of circulating transplant DNA has

been achieved with genome-wide SNVs (9) but this method is hampered when there is copy

number variation as is often found in tumor samples. Sehn et al. (10) have taken advantage of

closely spaced SNVs that are not in perfect LD, known as microhaplotypes (MH), to assess DNA

contamination to a level of 5%. Microhaplotypes can be defined as multiple variants on the

same sequence read such that the phasing of the variants is known and the haplotype can be

unambiguously determined.

Testing clinical tumor samples for somatic variation is complicated by the presence of large

regions of the tumor genome that may be highly amplified or deleted. When diagnostic

samples have large fractions of the genome with altered copy number, the allelic frequencies

(AFs) of SNVs can vary substantially and change from the typical 50-50 AF to much more

extreme values. These altered allele ratios can overlap the range of AFs for what is typically

seen with contamination and thus a sample that is 100% pure can mistakenly appear to be

significantly contaminated if only AF is examined. To avoid these problems, we have optimized

an MH counting method that is independent of genotype AF for determining contamination

status with tumor DNA and can be carried out with no matching normal sample.

For individuals with a normal set of chromosomes, each autosomal region of any size has two

sets of alleles. Those alleles may be the same (homozygous) or different (heterozygous); but,

under normal circumstances with a few specific exceptions, there should never be more than

two haplotypes in a genomic region in any individual. When DNA from two individuals is mixed,

accidentally or intentionally, there is a possibility for 1-4 MHs in each region. MHs have been

used for forensic purposes to detect contamination and identify individuals (11-17), to detect

transplant DNA in a host background and contamination in tumor samples (18), and to

determine ancestry (19-23). Use of MHs for routine detection and estimation of contamination

in clinical FFPE tumor samples is a powerful application of these methods.

Materials and Methods

Sequencing Samples

849 de-identified lung tumor samples and DNA from lung tumors were obtained from several

commercial and academic providers. These included BioIVT, Conversant Bio, Cureline, Duke,

Folio Bio, Fundacio Institute, Indivumed, iSpecimen, NCI, and Proteogenex. Samples were

received either as FFPE material or extracted DNA. If necessary, extraction of DNA from FFPE

specimens was carried out using standard kits. DNA was prepared for sequencing using

hybridization capture with panels directed primarily at exons in >500 genes implicated in cancer

(see https://www.personalgenome.com/assets/resources/elio-tissue-complete-brochure.pdf

for a listing of genes). For some genes, intronic and promoter DNA was also targeted. DNA was

sequenced using an Illumina NextSeq500. The PGDx elio™ tissue complete assay was used for

sample preparation and analysis (24, 25). 15 samples were sequenced on standard runs. For a

run to pass, at total of >90 GB sequence with >75% of reads at Q30 or above was required. For

an individual sample to pass, >90% of targeted regions were required to have >100x median

coverage. Individual reads were analyzed as described (26) with a base quality filter applied to

only include bases with a reported Phred quality score >30. Because erroneous reads could

artificially inflate the number of 3rd/4th MHs, MH calculations included 3rd MHs only if they

surpassed a minimum fraction of total calls. We usually set this threshold at >0.2% based on the

typical error rate for Illumina NGS (~0.1%). Higher values can be used to reduce noise but that is

accompanied by a loss in sensitivity.

Each accepted read covering the positions of all SNVs for each MH set was binned into the

appropriate haplotype and counted using Samtools mpileup. After read alignment, mpileup was

applied to the first region, the second region, and, if present, the third region. mpileup reads

are then combined and separated by basecalls. Each resultant set of base calls (MHs) is then

counted to determine MH frequency.

In silico read mixing

Artificially contaminated samples were made by *in silico* mixing of reads at defined

contamination levels. Contamination levels of x% were generated by combining randomly

selected 100-x% reads from the sample with x% random reads from the contaminant and

running the resultant reads through the standard analysis pipeline. It was found that the same

results were achieved if only the reads mapping to the MH regions being examined were

included in the mixing. These analyses involved much less data so ran on the pipeline much

faster. Thus, later experiments used only the truncated data set to speed analysis and allow

more conditions to be tested.

Results

DNA samples and MAF-based contamination detection

849 FFPE tumor DNA samples were examined, but most studies focused on the 51 samples

listed in Table 1. All samples that passed QC were amenable to contamination testing, but the

number of samples analyzed for mixing had to be limited due to the high compute

requirements for analyzing the many *in silico* mixtures. The mixing samples were chosen based

on being the most apparently pure and equally representing three of the five geographic

ancestral groups described by the 1000 genomes project (African, East Asian, European). DNA

quality was highly variable which primarily impacted mixing studies when two samples of much

different quality were mixed.

Prior to using MHs, we had assessed DNA contamination based on finding low level germline

SNVs that were assumed to come from contaminating DNA. However, FFPE tumor samples can

have high levels of copy number variation that can have a serious impact on germline SNV AFs

as shown in Fig 1. For 10 FFPE tumor samples, all variants in the VCF with a dbSNP number were

examined. Having an assigned dbSNP number was used as a surrogate for variation that is most

likely real since it has been observed previously. Variants were sorted by MAF into 10% bins.

Most samples had the expected distribution where >80% of the variants had 0-10% MAF with

this low-level variation arising from sequencing errors, FFPE artifacts or somatic variants. In

addition, there are also many with 40-50% MAF arising from heterozygous germline variants.

However, three samples had much more widely distributed MAFs including one sample

(AATF748T) where >20% of the variants had MAFs between 20 and 30%. These are germline

variants as confirmed by matching normal DNA samples that were also sequenced for these

samples. When we used a method for detecting DNA contamination based on low frequency

germline variants, these samples were discarded based on supposed high contamination levels

when, in fact, they are close to 100% pure based on MH analysis. The "contamination" signal is

actually copy number variation.

Figure 1 MAF for 10 FFPE tumor samples

All variants with dbSNP identifiers were binned by AF for the less common allele. Bins were 10%

ranges. These samples had 348-561 such variants in the VCF with the vast majority being

germline as confirmed by matching normal samples. The percentage of variants in each bin is

shown. Additional information about these samples is in Table 1.

Choice of variant sets for microhaplotype analysis

There are many MHs that have been characterized in the literature, and their usefulness for

distinguishing individuals and ancestries is well documented (13, 20, 23). However, most of the

well-characterized MHs are not in regions chosen for commercial cancer panel sequencing. In

addition, the criteria for selecting MHs for discriminating individuals and ancestry are not the

same as for detecting contamination where the goal is not distinguishing individuals but

maximizing the number of MHs with potentially >2 versions. Since high depth sequencing is

required for sensitive contamination detection using this approach, regions already targeted for

analysis were examined so that additional, non-productive sequencing in MH regions outside of

the pre-existing panel could be avoided. For gene panel sequencing, this means that literature

MH data is of limited value. Each novel MH set we chose was tested for behavior across

samples and ancestries. For these experiments, we have used a >500 gene, 2.23 Mb cancer

panel directed against FFPE tumor DNA

(https://www.personalgenome.com/assets/resources/elio-tissue-complete-brochure.pdf), but

any pre-existing panel can be examined in a similar manner to identify useful SNV sets. This

cancer panel included regions outside of exons and these regions were also inspected for

potentially informative SNV sets.

Candidate MH sets were chosen manually, but these steps can be automated if many or larger

panels are desired. The criteria for inclusion in the initial set of variants to be tested included

maximizing the frequency of 3rd MHs while minimizing the likelihood of sequencing errors in

those variants. This was achieved by avoiding regions with higher inherent sequencing error

rates such as insertions, deletions, and variants within homopolymeric regions and other

repetitive elements. The regions targeted for cancer panel sequencing were examined in the

gnomAD database (gnomad.broadinstitute.org/; 27) for multiple SNVs within 200 bp of the

targeted regions and of each other and with 3-97% AF. The desired final AF (5-95%) was higher,

but we wanted to ensure that MHs from populations underrepresented in gnomAD could still

be identified. The gnomAD database, at the time initially queried, had a higher proportion of

individuals of European ancestry (~60%) than individuals with other geographic ancestries.

Because of this, variation in European ancestry individuals had a larger impact on the initial

choice of SNVs than variation in other ancestries. Thus, there is a possibility that samples from

different ancestral backgrounds could respond differently when not filtered based on ancestry.

Once the candidate MH sets were identified, they were balanced for ancestry so that detection

of contamination in all groups would behave similarly.

Variants not in a segmental duplication or a low confidence region as defined by gnomAD were

examined for LD in the 1000 Genomes Project samples (ldlink.nci.nih.gov/?tab=ldhap; 28). This

site provides data on the frequency of each haplotype in each ancestry. Pairs and triplets of

SNVs with at least three haplotypes and the 3rd and greater haplotypes having a total frequency

of >5% in any individual ancestral grouping in the 1000 genomes project were advanced for

evaluation in real samples sequenced as described. SNV sets with median coverage >250x for all

relevant variants on individual reads were examined further. SNV sets with high frequency

3rd/4th MHs in many purportedly pure samples were eliminated because they might produce

high noise relative to signal. With these samples, SNV sets with more than 5 samples with >2

MHs were eliminated (26 of 264 evaluated SNV sets). SNV sets in close physical proximity to

each other in high LD would provide duplicate information. LD was considered too high if the

haplotype frequency was unchanged when the SNVs were combined and examined together.

SNV sets with shorter homopolymers, higher coverage, and fewer 3rd/4th MHs in pure samples

were included in the panel when high LD alternatives were considered. SNV sets that passed

these initial evaluations are shown in Supplementary Table 1. In addition to the genomic

coordinates and dbSNP identifiers, the frequency of MHs is shown for the five major ancestry

groupings defined by 1000 Genomes as well as the median coverage for the relevant SNVs

across 849 samples, the number of times the SNV set generated more than 2 MHs in "pure"

samples, and the DNA length separating the SNVs.

Once these criteria were met, SNV sets were balanced for ancestral frequency. While it would

have been desirable to use all population groupings from 1000 Genomes in this analysis, there

were not enough SNV sets to allow balancing of all five major ancestry groupings so only data

from African, East Asian, and European ancestry were assessed to achieve balanced variation.

SNV sets that adversely affected ancestry balance were eliminated. The balanced panel of 164

SNV sets is shown in Supplementary Table 1. The average frequency of 3rd/4th MHs was 13.9%

for each of the three balanced ancestries.

220   Assessing contamination and estimating levels

In order to determine how best to detect contamination, *in silico* mixing of reads from

individual pure samples was carried out so that the precise level of contamination would be

known and the sensitivity and accuracy of the MH assay could be determined. The initial

samples used for establishing contamination methods need to be as close to 100% pure as

possible for maximizing sensitivity. 51 samples were chosen for *in silico* mixing experiments to

examine this in more detail. Samples with self-declared ancestry were chosen to achieve

ancestral diversity and the fewest number of >2 MH sets. The performance of 51 reportedly

pure samples from individuals of known, self-declared ancestry is shown in Table 1.

The self-declared ancestry of samples was confirmed using Principle Component Analysis (PCA)

of the SNV sets based on 1000 Genome MH frequencies. The self-declared and PCA-determined

ancestries of all samples agreed. However, with some samples, the self-declared ancestry was

Asian and the PCA-determined ancestry was South Asian rather than East Asian. For analysis of

individuals who listed Asian ancestry, only samples with PCA-confirmed East Asian ancestry

were used in analyses. In addition to diverse ancestral histories, samples were selected for

mixing experiments based on having the fewest SNV sets with >2 MHs.

For these samples, the mean number of 3rd/4th MHs is over 20. Because these samples are

supposed to be pure, the 3rd/4th MHs are likely often due to artifactual false calls that arise due

to the high coverage depth for many SNV sets or FFPE-induced errors. This is supported by the

mean 3rd MH frequency which is only 0.003 for the panel. To minimize the number of artifactual

3rd MHs, we set a lower limit of 0.2% for counting 3rd/4th MHs based on the typical Illumina

error rate of ~0.1%. Using a threshold of >0.2% eliminates over 1/3 of 3rd MHs in these pure

samples, most of which are likely to be artifactual. When this threshold is employed, the

median 3rd MH frequency increases but remains less than 0.6% for all samples with >20 3rd

MHs. The need for a minimum threshold for the number of 3rd/4th MHs is highlighted by the

three examples from these 51 samples where the median 3rd MH frequency is >0.01. All three

have five or fewer 3rd MHs, suggesting that outlier artifactual MHs are driving the median

frequency when few real 3rd MHs are present. Based on this concern, we instituted a minimum

threshold of 25 SNV sets with >2 MHs with frequency >0.2%. 25 SNV sets with >2 MHs is not

intended to distinguish contaminated from pure samples but rather ensure that the 3rd MH

frequency is more reliable. 5 of the 51 pure DNAs in Table 1 have > 25 3rd/4th MHs (>0.2%

minimum threshold for inclusion as a 3rd MH). This threshold should never eliminate a truly

contaminated DNA but is intended only to ensure there are enough SNV sets to establish a

reliable 3rd MH median frequency. The low rate of 3rd/4th MHs across pure samples support the

concept that, at least at the sites analyzed, NGS and FFPE errors occur at a very low rate. All *in*

silico mixtures tested at the 0.5% contamination level using the samples in Table 1 had more

than 25 MHs. As seen in Fig 2, the average value for the number of 3rd MHs among all

ancestries with 1% contamination is >45 with this panel (range 29-69). Thus, this threshold

should not eliminate any samples contaminated at >0.5% and allows the 3rd MH frequency

value to be more reliable.

Precise levels of read mixing can be generated *in silico* but the functional level of contamination

was found to often differ significantly from the known mixing proportions. This issue was

particularly obvious when samples with divergent DNA qualities were mixed. The longer, higher

quality DNA appeared to have a higher concentration than what had been added to the mix.

This arises because longer reads map better and are more likely to contain all the variants being

examined and thus this DNA appears to be relatively more frequent than the shorter, more

degraded DNA. This is consistent with previous results that showed DNA with reduced

extraction yields tend to be lower quality and more susceptible to contamination (10). To

minimize the impact of this effect, all DNA mixes were examined in a pairwise fashion where

one sample was the primary sample and the second was the contaminant. These roles were

then switched and the results averaged. This allowed an over-contamination with one sample

set to be partially mitigated by the paired under-contamination of the opposite sample set.

15 DNA sets from each ancestry were mixed as sample/contaminant pairs with averaged results

shown in Fig 2. As expected, the East Asian fraction of >2 MHs was nearly identical to the

European results. The African American samples that we examined had a higher frequency of

>2 MHs than the other groupings, but nearly identical to the samples that were mixed across

ancestries. This is predicted by 1000 Genomes data. Their subgroupings of African ancestry

included seven different sub-populations, only one of which was US-based. When the African

American subgrouping is compared to the other African populations, it has a higher 3rd/4th MH

frequency (0.155 vs 0.139, Supplementary Table 1). The higher frequency in our population

reflects the fact our African ancestry samples are primarily from African American individuals.

Figure 2 Number of SNV sets with >2 MHs for individual samples

The number of 3rd MHs for pairs of individuals of East Asian, African American, and European

ancestry is shown for mixes of 0.1 – 40%. Mixing results from pairs of individuals with different

ancestry is included as "Mixed", all on a log scale.

Even at the lowest tested contamination level, 0.1%, the mean number of 3rd/4th MHs is over 30

compared to 13 (range 2-39) for the pure samples without mixing (Table 1). At 1%

contamination, the mean number of 3rd/4th MHs is over 45 for all groupings with a range for

individual mixes of 29-69. Because a small number of 3rd/4th MHs can cause a median values to

be impacted greatly by outliers, we have set a minimum threshold for the number of 3rd/4th

MHs before a contamination level is calculated. Based on these data, 25 is set as the minimum

number before a sample is considered potentially contaminated because it is less than all

samples mixed at 1% and high enough to minimize the impact of individual outliers.

While the number of 3rd MHs is relevant for assessing whether a sample is potentially

contaminated, it is less useful for estimating the level of contamination as it reaches a

maximum around 2% contamination with these coverage levels. In contrast, median 3rd MH

frequency changes as a function of contamination level so is more useful in this regard.

The *in silico* mixing studies provide an empirical calibration, but it is also useful to understand

the theoretical basis for those findings. As shown in Fig 3, the expected frequency of the

contaminating 3rd MH depends on the nature of the starting and incoming genotypes. Some

combinations will generate no 3rd MH, others will generate a 3rd MH with a frequency half the

level of incoming contaminant, and others will generate a genotype that is the same frequency

as the contaminant. Since most variant combinations will be at the 50% level, examination of

the median value for the 3rd MH should yield a value that is half of the true contamination level.

Figure 3 Expected frequencies for 3rd MHs based on genotype

A primary sample can be either homozygous for one MH or heterozygous for two versions. For

an incoming contaminating sample with at least one third MH, there are four different

possibilities for each starting primary sample as shown. Most of these generate a

contamination signal at half the frequency of the starting DNA. When the four alleles are all

different, the 3rd MH comes in at half the frequency while the combined 3rd/4th frequency is 1x.

If there is a 4th MH, it should have the same frequency as the 3rd MH as long as there are no

technical artifacts.

Figure 4 Contamination Detection as a function of ancestry

The median frequency of 3rd MHs for pairs of individuals of East Asian, African American, and

European ancestry is shown. Results from pairs of individuals with different ancestry is included

as "Mixed", all on a log scale for mixing levels of 0.1 – 40%.

The primary limitations on the sensitivity of contamination detection are the practical

sequencing depths for the SNV sets in question and the inherent error rates for sequencing. The

higher the sensitivity desired, the greater sequencing depth required. The usefulness of added

depth is limited by sequencing error rates that are typically on the order of 0.1% for Illumina. It

is possible to employ various error correction methods to improve sensitivity, but the need for

that is dependent on the sensitivity required for the application. In addition, there is no value in

sequencing a sample more deeply than the starting number of input molecules, which can be a

limitation in some situations. Based on these considerations, we have aimed for detection of

contamination at least as low as 1% in these samples.

As shown in Fig 4, the median 3rd MH level is independent of ancestry with this panel. All three

ancestries and mixed samples behave nearly identically. These data are combined in

Supplementary Table 2. 15 sample/contaminant pairs were tested for each ancestry with the

same number of each mixed ancestry pairs. When all samples are aggregated, 40% of samples

with 0.5% contamination were detected. If the threshold is set at 1%, all contaminated samples

were detected. The detection threshold can be set wherever required based on the detection

needs and the likely impact of contamination on final results. However, the core limitations of

sequencing accuracy, coverage depth, and input DNA molecules combine to ultimately limit

sensitivity.

In addition to detecting contamination, it is also possible to use this method to identify sources

of contamination. A partial set of genotypes for the contaminating DNA can be generated from

the MH data and then matched with samples that have been tested previously or

contemporaneously to determine if they are the source of contamination. In a contaminated

sample, MHs with 1 or 4 different haplotypes can be assigned their correct genotypes directly.

MHs with 2 or 3 different haplotypes can generally be assigned a partial or complete genotype

based on the frequency of the individual MHs. The frequency of the 2nd/3rd haplotypes will

provide at least one and sometimes two genotypes corresponding to the contaminant.

Comparison of the known and partial genotypes with other samples allows them to be ruled in

or out as potential sources of contamination. This procedure has been applied to multiple

contaminated samples to successfully determine the source in real clinical laboratory

situations. Often, the contamination source can be shown not to arise from any samples known

to have been handled in the laboratory so the contamination must have originated prior to

sample receipt.

Discussion

Cross-contamination of DNA samples is a well-known issue among clinical testing laboratories

and hence standard clinical testing guidelines generally suggest that precautions should be

taken to be prevent it. Two recent guidelines for somatic variant testing (29, 30) recommend

checking for handling-induced contamination and, suggest some possible methods for

detecting contamination (6, 10, 31). However, none of the methods mentioned is sensitive

enough to ensure that somatic variants (32) can be distinguished from sample contamination.

Thus, simple, sensitive methods for reliable detection of contamination are needed.

The degree of DNA contamination that causes problems varies depending on the application. In

some cases, the impact of contamination on results is obvious. With other applications, signals

arising from contamination can be easily confused with real signals. With diagnosis of somatic

variants in cancer, this is especially a problem because variant identification is attempted at and

below the very limits of the NGS technology. Being able to separate the tumor signal from any

contamination noise is critical for the proper diagnosis of tumors. Because tumor samples often

have highly unusual copy number patterns, use of simple genotype frequencies can cause

errors in contaminant detection. In our experience, many FFPE samples that had been

determined to be contaminated using raw genotype frequencies were found to be pure by MH

analysis (Fig 1). In contrast to FFPE tumor samples, circulating-free DNA (cfDNA) purified from

the plasma of cancer patients is not as prone to the same issues. Most cfDNA purified from

plasma is from normal cells so typical Mendelian ratios are observed with variants. MH analysis

in such samples is still valuable for sensitive contamination detection, but such samples are not

as likely to generate false positives as DNA from FFPE tumor samples where copy number

variation is much more significant because of the high tumor DNA content.

For most purposes, the approximation that the percent contamination is twice the median

frequency for the 3rd MH is sufficient. This relationship arises from the likelihood that a 3rd MH

is most likely heterozygous so is present on only one of the two incoming alleles. The

uncertainty around this probability introduces the potential for multiple sources of error and

those should be considered if a more exact measurement is necessary. At low coverage, two

confounding factors arise. First, low coverage leads to high AF variability due to the stochastic

nature of read accumulation in poorly covered SNV sets. Only SNV sets with 3rd MHs are

included in the median calculation rather than all SNV sets so, at low coverage, AF inflation

occurs among the variants that are counted. The observed AF is higher than reality because

there are not enough reads to generate the proper AF value. By requiring a large number of 3rd

MHs prior to calling a sample contaminated, this effect is mitigated. Superimposed on this

effect is the counting of homozygous 3rd MHs before heterozygous 3rd MHs as coverage

increases. If coverage is deep, this is not an issue but, at low coverage, homozygous variants will

appear first because they are being double counted. Further complicating the relationship is the

presence of low frequency technical artifacts and the copy number issues that can occur with

cancer samples. The copy number variations will make SNV sets appear at different fractional

rates depending on whether the 3rd MH is under- or over-represented. The more SNV sets used

in the calculations, the more likely it is that these effects can be averaged out.

The MH approach is perfectly suited for use when somatic variants are sought because such

studies use very high coverage in order to detect rare variants. The sensitivity of contamination

detection is limited by the coverage of the regions used in the analysis. Other panel-based and

exome NGS approaches can be made more reliable by using similar contamination detection

methods. A typical whole genome approach with less than 50x coverage would not be

amenable to sensitive detection using individual MH sets because the low coverage would

prevent the observation of infrequent 3rd MHs. To overcome this, combinations of MH sets

could be used. The simplest approach would be to combine SNV sets that are in high LD. Other

methods for pooling SNV sets could also be used.

The theoretical limitation on the sensitivity of the MH assay is the number of DNA molecules

available for study. The assay can only be as sensitive as the amount of input DNA allows. In

addition to that limitation (which is significant for some samples), there are also technology

limitations. When looking at a pair of SNVs, either one could be called incorrectly resulting in

false 3rd MHs. If panels with 3 or more SNVs in close proximity were available, this would

minimize such errors. However, there are other potential errors introduced in NGS processing.

It is not unusual for barcodes used for multiplexing samples in the same run to be

contaminated at 1% or more, resulting in misassignment of reads to samples. Index hopping

and chimeric PCR reads can also affect sensitivity (33, 34).

The same techniques used for these cancer samples can also be used with forensic samples to

detect contributors to a sample. Unlike tumor samples, copy number is not an issue. Like tumor

samples, the major and minor contributors to a sample may have different DNA qualities that

can affect the accuracy of MH frequencies, but this is less important in forensic situations.

Panels designed with a larger number of >4 potential MHs would be appropriate when forensic

samples are being examined to clarify the number of contributors.

Another useful application of the MH methods is for detection of CNV anomalies in fetal DNA in

Non-invasive prenatal testing (NIPT). MH methods are already in use for paternity testing (35-

38). For NIPT, panels can be designed to focus on genomic regions with the most likely copy

number variation (e. g. chr 5, 13, 18, or 21) and the frequency of 3rd MHs in targeted regions

compared to the rest of the genome. In genomic regions where the mother is heterozygous and

the father's contribution is a different allele, there will be a low frequency 3rd MH.

The methods described here highlight the factors that are important for examining DNA

mixtures in a variety of contexts. The use of MHs in these applications requires different

properties than other uses (39). Most early studies with MHs focus on distinguishing individuals

and assigning ancestry. The MH properties required in those situations are different with a

need for MHs specific for particular populations. With MHs used for contamination detection, it

is preferable to have MHs that are common to all populations so that detection is uniform

across individuals. The methods described herein allow the appropriate choice of MHs leading

to the ability to detect low levels of secondary DNA, which can be valuable in many

applications.

Abbreviations

AdAm: Admixed American

AF: Allele Frequency

AfAm: African American

Afri: African

cfDNA: circulating free DNA

EaAs: East Asian

Euro: European

FFPE: Formalin-fixed Paraffin-embedded

FP: False Positive

LD: Linkage Disequilibrium

MH: Microhaplotype

NGS: Next-generation Sequencing

NIPT: Non-invasive Prenatal Testing

PCA: Principle Component Analysis

SNV: Single Nucleotide Variant

SoAs: South Asian

STR: Short Tandem Repeat

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Table 1 Microhaplotypes and Coverage for Pure Samples

Performance of the 164 SNV set panel is shown with 51 pure samples with known ancestries. Comparisons include the median and

average coverage, the number of SNV sets with 1-4 MHs, and the median frequency of 3rd MHs among individuals of African (Afri),

East Asian (EaAs), and European (Euro) ancestry as defined by the 1000 Genomes project. The number and median 3rd MH

frequency for MHs with frequency >0.2% is shown. ND=Not Done; NA=Not Applicable

Sample	M/F	Ancestry	Number of MHs				Med 3rd MH	# 3rd MHs >0.002	Med 3rd >0.002	#	Med Cov	Ave Cov
			1	2	3	4						
AATF748T		Afri	52	87	21	3	0.0033	18	0.0081	163	313	620
AATF094T	F	Afri	36	90	36	0	0.0027	29	0.0031	162	421	533
AATF115T	F	Afri	67	82	12	0	0.0027	7	0.0059	161	202	286
AATF066T	F	Afri	48	92	21	0	0.0041	17	0.0044	161	305	337
AATF117T	F	Afri	37	##	22	1	0.0053	18	0.0065	163	439	605
AATF498T	F	Afri	68	76	19	0	0.0018	8	0.0037	163	674	959
AATF308T	M	Afri	42	96	24	0	0.0022	14	0.0033	162	531	683
AATF562T		Afri	46	##	14	1	0.0021	8	0.0029	163	574	706
AATF486T	F	Afri	39	88	36	0	0.0023	21	0.0034	163	709	924
PGRD0454T	M	Afri	58	90	14	0	0.0018	6	0.0027	162	797	930
AATF218T	F	Afri	42	88	33	1	0.0029	24	0.0034	164	405	555
AATF778T		Afri	60	91	11	1	0.0023	7	0.0055	163	462	520

AATF219T	M	Afri	33	95	34	1	0.0022	19	0.0039	163	553	668
AATF217T	M	Afri	57	90	15	0	0.0018	5	0.0029	162	609	996
AATF073T	F	EaAs	74	78	12	0	0.0021	7	0.0028	164	415	726
AATF503T	F	EaAs	62	76	24	1	0.0031	16	0.0037	163	426	545
AATF504T	M	EaAs	70	84	8	1	0.0042	9	0.0042	163	517	758
AATF027T	M	EaAs	58	92	13	1	0.0024	8	0.0052	164	331	443
AATF012T	F	EaAs	66	91	7	0	0.0029	5	0.0050	164	542	826
AATF029T	M	EaAs	42	74	46	1	0.0033	37	0.0035	163	379	590
AATF024T	F	EaAs	31	82	48	2	0.0037	39	0.0051	163	452	724
AATF023T	M	EaAs	33	91	36	3	0.0025	26	0.0028	163	500	765
AATF022T	M	EaAs	42	92	30	0	0.0023	19	0.0046	164	510	768
AATF021T	M	EaAs	56	78	29	0	0.0027	19	0.0041	163	528	792
AATF733T	F	EaAs	53	93	17	0	0.0047	12	0.0086	163	170	257
AATF595T	F	EaAs	53	84	27	0	0.0038	19	0.0098	164	201	390
AATF742T	F	EaAs	51	##	10	0	0.0020	5	0.0128	163	210	352
AATF734T	M	EaAs	60	89	13	1	0.0029	11	0.0032	163	344	431
AATF594T		EaAs	64	92	7	0	0.0028	5	0.0030	163	390	533
AATF735T	M	EaAs	68	88	6	1	0.0025	5	0.0027	163	399	614
AATF597T	F	EaAs	54	94	14	1	0.0021	8	0.0062	163	399	502
AATF606T	F	EaAs	62	85	15	0	0.0023	10	0.0029	162	443	659
AATF601T	F	EaAs	58	85	20	0	0.0022	11	0.0030	163	457	570
AATF599T	F	EaAs	58	84	21	1	0.0025	15	0.0042	164	487	648
AATF600T	F	EaAs	52	94	16	1	0.0024	10	0.0028	163	605	752
AATF417T	M	Euro	67	90	5	1	0.0142	5	0.0176	163	187	239
AATF391T	M	Euro	47	95	20	1	0.0019	10	0.0023	163	476	754

AATF375T	M	Euro	34	97	32	1	0.0052	30	0.0056	164	305	463
AATF088T	F	Euro	41	99	21	1	0.0029	16	0.0038	162	288	406
AATF389T	F	Euro	39	94	30	1	0.0021	16	0.0030	164	446	741
AATF567T	M	Euro	68	84	10	0	0.0035	8	0.0053	162	312	704
AATF107T	M	Euro	51	98	13	1	0.0034	9	0.0066	163	445	518
AATF079T	F	Euro	58	94	10	0	0.0016	4	0.0041	162	494	641
AATF496T	M	Euro	62	94	8	0	0.0018	2	0.0322	164	601	674
AATF036T	M	Euro	44	97	20	2	0.0029	15	0.0030	163	538	603
AATF042T	M	Euro	39	93	31	1	0.0020	16	0.0027	164	613	747
AATF039T	M	Euro	49	##	11	0	0.0020	5	0.0035	162	693	842
AATF038T	F	Euro	50	94	16	2	0.0015	7	0.0031	162	673	789
AATF731T	F	Euro	59	87	17	1	0.0024	12	0.0027	164	352	495
AATF378T	F	Mix	52	94	17	1	0.0028	12	0.0042	164	418	707
AATF097T	F	Mix	35	98	30	0	0.0035	22	0.0042	163	416	486
Median			52	91	17	1	0.0025	11	0.0038	163	445	641
Mean			52	90	20	1	0.0030	13	0.0052	163	450	623

Supplementary Table 1 Detailed characterization of MH sets

The genomic locations of all SNV sets tested are shown in column 1. Median coverage in 849 samples and the number of instances

of more than 2 MHs are shown in the next columns. The distance between SNVs and the dbSNP identifiers are listed in the next

columns. The final columns show the sum of frequencies for all 3rd and 4th MHs from the 1000 genome database and the reason for

why SNVs were removed from the final panel, if applicable.

Supplementary Table 2 Detection of in silico mixtures versus 3rd MH threshold

Pure samples were mixed in 15 different combinations from each ancestry group (Afri, EaAs, Euro) as well as 15 different mixed

ancestry combinations. For each threshold 3rd MH frequency, the percent of all samples that scored as contaminated at that level of

contamination are listed at each level of in silico mixing.

  Supplementary Table 1

Location	Final Panel	Med Cov	MH >2	SNV Len	SNV1	SNV2	SNV3	Afri 3+4	EaAs 3+4	Euro 3+4	AdAm 3+4	SoAs 3+4	AfAm 3+4	Reason for Omitting
chr1:23885498-23885599		692	25	102	rs11574	rs2067053		0.011	0.028	0.242	0.261	0.081	0.016	MH>2
chr1:46641827-46641837		296	1	11	rs12030928	rs12752892		0.017	0.000	0.125	0.071	0.080	0.057	Balance
chr1:120057158-120057246		689	3	89	rs6203	rs45609334		0.033	0.082	0.235	0.205	0.157	0.090	Balance
chr1:156846120-156846233	Yes	1526	2	114	rs1800880	rs6334		0.105	0.139	0.065	0.117	0.244	0.098	
chr1:226573364-226573402	Yes	2011	1	39	rs1805414	rs1805408		0.143	0.205	0.159	0.147	0.183	0.164	
chr1:226589833-226589958	Yes	361	2	126	rs1805407	rs1805404		0.115	0.251	0.154	0.147	0.100	0.164	
chr2:16042003-16042051	Yes	392	1	49	rs2693006	rs67056216		0.113	0.177	0.177	0.159	0.264	0.123	
chr2:16073257-16073263	Yes	1546	2	7	rs12986946	rs12986949		0.052	0.000	0.101	0.058	0.115	0.098	
chr2:16112814-16112828	Yes	835	1	15	rs16863159	rs6716344		0.022	0.276	0.088	0.244	0.131	0.148	
chr2:16113594-16113723	Yes	368	4	130	rs34339850	rs6741005		0.052	0.284	0.217	0.183	0.245	0.066	
chr2:29416366-29416481		677	2	116	rs1881421	rs1881420		0.240	0.000	0.150	0.127	0.027	0.312	Balance
chr2:29416481-29416615		750	15	135	rs1881420	rs56132472		0.078	0.000	0.123	0.065	0.024	0.090	MH>2
chr2:29446184-29446202	Yes	2130	0	19	rs2276550	rs4622670		0.259	0.054	0.236	0.222	0.203	0.033	
chr2:29446701-29446721	Yes	686	1	21	rs12619049	rs4665447		0.412	0.081	0.026	0.062	0.015	0.320	
chr2:29447108-29447253	Yes	448	1	146	rs4387740	rs6723311		0.390	0.141	0.254	0.232	0.173	0.426	
chr2:47800577-47800603	Yes	1072	0	27	rs56239373	rs3814360		0.077	0.154	0.042	0.065	0.086	0.057	
chr2:47852559-47852643	Yes	293	5	85	rs6722699	rs10165802		0.110	0.076	0.093	0.104	0.061	0.139	
chr2:47856243-47856328		572	8	86	rs11892597	rs11887626		0.209	0.078	0.273	0.203	0.261	0.205	MH>2
chr2:48010488-48010558		1461	2	71	rs1042821	rs1042820		0.020	0.000	0.175	0.114	0.065	0.033	Balance
chr2:112754828-112754880		366	1	53	rs3811632	rs3811633		0.103	0.106	0.287	0.189	0.230	0.156	Balance
chr2:112754943-112755001		747	3	59	rs3811634	rs2230515		0.104	0.106	0.287	0.189	0.230	0.156	Balance
chr2:113980645-113980736		550	0	92	rs12472361	rs56865224		0.017	0.073	0.110	0.117	0.129	0.057	
chr2:113982584-113982608		540	0	25	rs4849177	rs4849178		0.017	0.077	0.164	0.138	0.183	0.057	
chr2:113983937-113984033	Yes	776	1	97	rs3748915	rs3748916		0.203	0.086	0.163	0.135	0.229	0.213	
chr2:113984503-113984594	Yes	1400	0	92	rs2241975	rs67776659		0.142	0.013	0.110	0.087	0.038	0.074	

chr2:113985170-113985186		1536	2	17	rs4849179	rs4849180		0.020	0.077	0.163	0.138	0.183	0.066	LD, homopolymer
chr2:113989236-113989267	Yes	1009	2	32	rs2863242	rs2863243		0.017	0.074	0.163	0.138	0.183	0.057	
chr2:113990242-113990261		2923	0	20	rs2166421	rs7421852		0.011	0.074	0.162	0.135	0.183	0.041	LD, homopolymer
chr2:202122956-202122995	Yes	1337	0	40	rs3769824	rs3769823		0.000	0.000	0.047	0.114	0.043	0.016	
chr3:12641425-12641518		346	1	94	rs5746223	rs2246390		0.040	0.030	0.111	0.125	0.065	0.090	Balance
chr3:12642945-12643013		277	3	69	rs3773345	rs904464	rs904452	0.145	0.031	0.192	0.114	0.052	0.148	Balance
chr3:12649857-12649937	Yes	567	2	81	rs2055311	rs963959		0.225	0.028	0.164	0.310	0.125	0.221	
chr3:36986932-36986992	Yes	2760	4	61	rs2276809	rs2276808		0.073	0.077	0.115	0.160	0.216	0.090	
chr3:36986992-36987083		2802	6	92	rs2276808	rs2276807		0.210	0.000	0.048	0.078	0.000	0.303	MH>2
chr3:37022803-37022864		345	0	62	rs4678921	rs4678922		0.082	0.000	0.000	0.026	0.000	0.213	Balance
chr3:71247257-71247304	Yes	1098	0	48	rs939845	rs2037474		0.163	0.104	0.064	0.202	0.044	0.139	
chr3:138327951-138328016	Yes	634	1	66	rs61699523	rs111398337		0.167	0.020	0.028	0.071	0.110	0.295	
chr3:142277536-142277575	Yes	642	0	40	rs2227929	rs2227930		0.147	0.118	0.200	0.154	0.158	0.188	
chr3:178968634-178968660		1223	0	27	rs7645550	rs1170672		0.041	0.000	0.095	0.038	0.170	0.057	Balance
chr3:178984575-178984679	Yes	2320	2	105	rs7612684	rs7646600		0.302	0.011	0.177	0.131	0.132	0.213	
chr3:178986121-178986203	Yes	623	5	83	rs73188921	rs9830427	rs9830432	0.158	0.119	0.054	0.076	0.190	0.131	
chr3:178990402-178990462	Yes	1179	1	61	rs2864411	rs6443633		0.017	0.142	0.000	0.050	0.045	0.098	
chr3:183211906-183212026		536	2	121	rs1520101	rs2256061		0.128	0.000	0.182	0.123	0.162	0.238	Balance
chr4:1745492-1745500	Yes	4202	2	9	rs4865466	rs4865467		0.126	0.144	0.217	0.306	0.229	0.139	
chr4:1747971-1748069		1482	1	99	rs4572936	rs4558926		0.049	0.000	0.156	0.095	0.082	0.098	Balance
chr4:1748200-1748266		2245	8	67	rs4507432	rs4597899	rs62285133	0.328	0.159	0.392	0.311	0.273	0.336	MH>2
chr4:1750487-1750584	Yes	1702	3	98	rs7680647	rs73202803		0.042	0.161	0.235	0.180	0.121	0.057	
chr4:1785738-1785791		2874	2	54	rs1867926	rs61407096		0.063	0.000	0.158	0.157	0.087	0.074	Balance
chr4:1788994-1789044	Yes	678	4	51	rs11248077	rs11248078		0.249	0.233	0.383	0.346	0.377	0.254	
chr4:1796629-1796636	Yes	319	1	8	rs3135841	rs3135842		0.254	0.051	0.094	0.141	0.061	0.262	

chr4:1797741-1797852	Yes	995	4	112	rs3135848	rs743682		0.227	0.056	0.092	0.144	0.062	0.271	
chr4:54269096-54269173	Yes	557	1	78	rs10001201	rs62325166		0.050	0.133	0.140	0.105	0.046	0.082	
chr4:54526892-54526902		647	2	11	rs12649494	rs2668565		0.023	0.093	0.153	0.079	0.117	0.082	Balance
chr4:54589337-54589371		445	21	35	rs2590827	rs75219949		0.054	0.223	0.140	0.208	0.207	0.115	MH>2
chr4:54657737-54657790	Yes	288	5	54	rs28489910	rs4864823		0.233	0.111	0.209	0.226	0.148	0.254	
chr4:55049698-55049751		310	3	54	rs13131212	rs7659596		0.028	0.061	0.215	0.159	0.242	0.041	Balance
chr4:55208737-55208788	Yes	284	3	52	rs2412560	rs10018115	rs73234206	0.202	0.247	0.200	0.270	0.317	0.361	
chr4:55477000-55477073		991	4	74	rs17827038	rs2703460		0.020	0.072	0.134	0.098	0.195	0.090	Balance
chr4:55501109-55501195	Yes	357	5	87	rs6554196	rs6554197		0.110	0.110	0.200	0.163	0.223	0.230	
chr4:55582037-55582068	Yes	714	3	32	rs76272262	rs3134889		0.040	0.172	0.036	0.051	0.081	0.025	
chr4:55619846-55619859	Yes	892	3	14	rs11732442	rs4353958		0.125	0.109	0.109	0.069	0.212	0.139	
chr4:55916001-55916063		317	0	63	rs7664996	rs7665169		0.164	0.000	0.201	0.177	0.149	0.172	Balance
chr4:55977759-55977800		278	3	42	rs3943404	rs2034965		0.210	0.215	0.258	0.264	0.227	0.213	Balance
chr4:55982752-55982784	Yes	651	1	33	rs11133360	rs34945396		0.044	0.204	0.194	0.144	0.190	0.041	
chr4:56026865-56026914	Yes	565	1	50	rs4864958	rs75371420	rs34743464	0.216	0.200	0.284	0.180	0.453	0.238	
chr4:106196829-106196951	Yes	534	0	123	rs34402524	rs2454206		0.066	0.047	0.140	0.089	0.090	0.090	
chr4:143043340-143043404		351	0	65	rs2270658	rs13133767		0.016	0.075	0.082	0.105	0.089	0.057	Balance
chr4:187534362-187534375	Yes	2353	0	14	rs2249916	rs2249917		0.195	0.281	0.110	0.189	0.084	0.197	
chr4:187629497-187629538	Yes	1727	0	42	rs458021	rs3733413		0.128	0.085	0.070	0.091	0.031	0.098	
chr5:231111-231143	Yes	2366	1	33	rs1126417	rs2288459		0.164	0.058	0.111	0.241	0.079	0.139	
chr5:256472-256509		1282	9	38	rs6961	rs6962		0.269	0.059	0.105	0.108	0.075	0.197	MH>2
chr5:35861068-35861159	Yes	351	3	92	rs1494558	rs11567705	rs969128	0.328	0.191	0.413	0.349	0.239	0.443	
chr5:35871190-35871273	Yes	255	1	84	rs1494555	rs2228141		0.069	0.153	0.144	0.166	0.062	0.164	
chr5:56178111-56178217	Yes	473	0	107	rs3822625	rs832583		0.119	0.108	0.075	0.078	0.055	0.049	
chr5:57754808-57754851		359	2	44	rs697133	rs702722		0.230	0.105	0.104	0.069	0.098	0.205	Balance

chr5:67477132-67477234	Yes	371	0	103	rs34721946	rs34166422	rs73126524	0.017	0.247	0.035	0.105	0.072	0.033	
chr5:67492589-67492652	Yes	677	2	64	rs13188623	rs58409263		0.105	0.293	0.121	0.180	0.118	0.130	
chr5:67517563-67517646	Yes	275	1	84	rs6449959	rs831227		0.243	0.018	0.187	0.161	0.100	0.303	
chr5:67522722-67522851	Yes	262	1	130	rs706713	rs706714		0.130	0.051	0.012	0.029	0.060	0.098	
chr5:67534039-67534057	Yes	887	0	19	rs7709243	rs10940158	rs12652661	0.216	0.154	0.212	0.272	0.097	0.156	
chr5:67553771-67553827	Yes	584	1	57	rs6893676	rs34303		0.090	0.168	0.173	0.143	0.106	0.074	
chr5:79950497-79950512		1392	8	16	rs2250063	rs1105524		0.036	0.047	0.248	0.206	0.231	0.098	MH>2
chr5:149456772-149456811	Yes	1109	3	40	rs60844779	rs3829987		0.223	0.068	0.031	0.215	0.051	0.221	
chr5:176517326-176517461		652	3	136	rs422421	rs446382		0.169	0.000	0.078	0.040	0.033	0.148	Balance
chr5:176523562-176523597	Yes	1990	0	36	rs31777	rs31776		0.137	0.000	0.076	0.038	0.033	0.115	
chr5:176531772-176531857	Yes	284	3	86	rs7708357	rs165943		0.168	0.046	0.242	0.248	0.183	0.197	
chr5:176539212-176539286		922	0	75	rs244730	rs351864		0.031	0.027	0.121	0.140	0.207	0.066	Balance
chr5:176721198-176721272		1806	1	75	rs28580074	rs11740250		0.011	0.000	0.119	0.146	0.181	0.041	Balance
chr5:180046209-180046344		765	12	136	rs446003	rs448012		0.100	0.057	0.083	0.075	0.135	0.098	MH>2
chr5:180051003-180051118		2483	2	116	rs307826	rs728986		0.015	0.000	0.037	0.055	0.044	0.016	Balance
chr5:180057231-180057293		1518	0	63	rs3736061	rs34221241		0.000	0.000	0.081	0.033	0.041	0.016	Balance
chr6:26056549-26056708	Yes	524	2	160	rs10425	rs2230653	rs12204800	0.048	0.309	0.227	0.344	0.256	0.082	
chr6:30865115-30865204	Yes	461	5	90	rs2239517	rs2267641		0.120	0.244	0.038	0.063	0.094	0.098	
chr6:32188603-32188642	Yes	1185	1	40	rs520803	rs520692	rs520688	0.000	0.047	0.000	0.000	0.011	0.000	
chr6:32190390-32190484	Yes	2363	5	95	rs915894	rs8192569		0.330	0.232	0.102	0.141	0.205	0.328	
chr6:41924853-41924931	Yes	922	2	79	rs4623235	rs16895130		0.095	0.110	0.210	0.156	0.138	0.107	
chr6:41941980-41941993		1079	1	14	rs4415146	rs72853828		0.000	0.089	0.221	0.125	0.132	0.049	Balance
chr6:42013020-42013049	Yes	530	0	30	rs9381126	rs6919122	rs6942118	0.351	0.421	0.381	0.504	0.390	0.254	
chr6:42039487-42039542	Yes	651	3	56	rs9349215	rs66472208		0.023	0.245	0.020	0.048	0.127	0.057	
chr6:42039551-42039666	Yes	292	1	116	rs66489927	rs7763360	rs2492927	0.192	0.148	0.300	0.248	0.322	0.221	

chr6:42044204-42044273		717	1	70	rs11968793	rs9471750		0.184	0.057	0.046	0.045	0.014	0.221	Balance
chr6:42052577-42052667	Yes	305	0	91	rs9357387	rs2493841	rs9381136	0.050	0.163	0.176	0.161	0.139	0.123	
chr6:117725448-117725578		277	4	131	rs1998206	rs2243378		0.076	0.181	0.150	0.143	0.197	0.197	Balance
chr6:152382311-152382325		279	2	15	rs2273206	rs2273207		0.137	0.039	0.026	0.039	0.055	0.066	Balance
chr7:6026775-6026942		720	19	168	rs2228006	rs1805323		0.000	0.122	0.046	0.017	0.106	0.016	MH>2
chr7:6026942-6026988	Yes	3560	3	47	rs1805323	rs1805321		0.000	0.303	0.046	0.017	0.153	0.016	
chr7:55220177-55220202	Yes	1118	0	26	rs11506105	rs845561		0.115	0.265	0.254	0.304	0.413	0.148	
chr7:55236774-55236862		458	5	89	rs845550	rs1815156		0.096	0.000	0.114	0.062	0.042	0.123	Balance
chr7:55251541-55251648	Yes	672	4	108	rs2877261	rs13222385	rs11771471	0.200	0.076	0.233	0.183	0.090	0.189	
chr7:55306599-55306625		577	6	47	rs2037700	rs2037699	rs2037698	0.368	0.083	0.104	0.124	0.045	0.271	MH>2
chr7:100410597-100410657		1469	8	61	rs2230585	rs770657085		0.164	0.056	0.000	0.043	0.156	0.197	MH>2
chr7:100416139-100416250	Yes	1438	3	112	rs3857809	rs144173		0.185	0.059	0.000	0.301	0.173	0.172	
chr7:116336880-116336947	Yes	666	1	68	rs2237708	rs39749		0.036	0.209	0.257	0.228	0.242	0.115	
chr7:116471122-116471227	Yes	297	4	106	rs41773	rs62470772		0.129	0.093	0.206	0.115	0.148	0.131	
chr7:116480433-116480524		319	7	92	rs41777	rs35099490	rs41778	0.267	0.217	0.279	0.268	0.283	0.288	MH>2
chr8:30999122-30999123	Yes	554	3	2	rs3024239	rs2737335		0.149	0.024	0.060	0.032	0.085	0.246	
chr8:31024638-31024654	Yes	432	0	17	rs1801196	rs1346044		0.147	0.104	0.266	0.173	0.283	0.221	
chr8:38299624-38299715	Yes	1668	5	92	rs60527016	rs6987534		0.028	0.286	0.236	0.219	0.076	0.074	
chr8:38310910-38311001	Yes	1289	0	92	rs10958700	rs4733930		0.029	0.323	0.260	0.249	0.074	0.074	
chr8:38317398-38317476		1504	4	79	rs7012413	rs7388222	rs3758101	0.213	0.088	0.076	0.225	0.115	0.205	Balance
chr8:38350292-38350315	Yes	580	2	24	rs35305468	rs7830964		0.039	0.249	0.180	0.118	0.138	0.107	
chr8:38359468-38359529		1045	3	62	rs7845393	rs35804490		0.137	0.172	0.125	0.114	0.282	0.180	LD
chr8:38361379-38361430	Yes	1456	2	52	rs328294	rs328293		0.309	0.172	0.126	0.115	0.283	0.213	
chr8:128700175-128700233	Yes	496	2	59	rs13282849	rs7005394		0.208	0.179	0.063	0.084	0.201	0.287	
chr8:128713221-128713364	Yes	796	5	144	rs28548827	rs7820045		0.254	0.057	0.028	0.101	0.111	0.148	

Region														
chr8:128718068-128718102		457	2	35	rs9642880	rs78635199		0.000	0.035	0.129	0.062	0.171	0.041	Balance
chr8:128889285-128889371	Yes	1835	1	87	rs6470587	rs6470588		0.081	0.165	0.210	0.202	0.230	0.156	
chr8:145737636-145737816	Yes	485	0	181	rs4925828	rs4251691		0.000	0.203	0.000	0.072	0.000	0.000	
chr8:145741702-145741765		2058	6	64	rs4244612	rs4244613		0.081	0.027	0.108	0.094	0.129	0.148	MH>2
chr9:5408242-5408358	Yes	344	3	117	rs10758685	rs10975098	rs10975099	0.084	0.349	0.257	0.320	0.409	0.205	
chr9:5415025-5415111	Yes	372	3	87	rs78298180	rs10758687		0.104	0.161	0.054	0.052	0.199	0.066	
chr9:5420254-5420266	Yes	1180	1	13	rs10121219	rs11790878		0.064	0.227	0.222	0.248	0.218	0.172	
chr9:5456523-5456587		277	1	65	rs17742278	rs7023227	rs12002985	0.165	0.083	0.337	0.320	0.305	0.262	Balance
chr9:5458035-5458095	Yes	323	3	61	rs7042084	rs10481593		0.268	0.132	0.220	0.249	0.131	0.205	
chr9:5484100-5484203	Yes	395	4	104	rs11793113	rs11790610	rs10122509	0.139	0.151	0.094	0.084	0.167	0.156	
chr9:5505508-5505583		262	5	76	rs7025653	rs59338043	rs6476984	0.228	0.188	0.234	0.298	0.215	0.205	Balance
chr9:5514839-5514919		561	7	81	rs2381282	rs56381807		0.275	0.204	0.128	0.203	0.190	0.303	MH>2
chr9:87478135-87478172	Yes	1016	4	38	rs7048015	rs10780690		0.023	0.251	0.184	0.258	0.216	0.139	
chr9:93641175-93641199		693	2	25	rs2306041	rs2306040		0.062	0.000	0.064	0.058	0.020	0.066	Balance
chr9:98238358-98238379	Yes	3840	0	22	rs2066836	rs1805155		0.011	0.083	0.109	0.076	0.060	0.074	
chr9:139401504-139401577	Yes	1346	1	74	rs3124596	rs7870145	rs3829116	0.310	0.000	0.163	0.117	0.264	0.238	
chr9:139403268-139403280		500	1	13	rs3125000	rs11145765		0.046	0.000	0.095	0.059	0.238	0.025	Balance
chr9:139405093-139405261	Yes	626	3	169	rs36119806	rs3125001		0.150	0.012	0.102	0.065	0.184	0.156	
chr9:139410424-139410589	Yes	327	2	166	rs3125006	rs4880099		0.088	0.052	0.115	0.068	0.215	0.049	
chr9:139411714-139411880		428	5	167	rs11145767	rs9411254		0.209	0.000	0.000	0.025	0.000	0.213	Balance
chr10:43611708-43611865	Yes	629	2	158	rs741968	rs2256550		0.060	0.218	0.161	0.212	0.284	0.139	
chr10:43615505-43615633	Yes	463	5	129	rs2472737	rs1800863		0.105	0.121	0.193	0.187	0.160	0.115	
chr10:70332580-70332672	Yes	549	1	93	rs10823229	rs12773594		0.023	0.173	0.185	0.151	0.271	0.090	
chr10:104386934-104387019	Yes	250	0	86	rs17114803	rs12414407		0.224	0.250	0.093	0.238	0.240	0.197	
chr10:123194558-123194609	Yes	384	0	52	rs7911440	rs6585731		0.051	0.211	0.242	0.082	0.243	0.107	

Position														
chr10:123199092-123199095	Yes	1151	2	4	rs4752560	rs2114689		0.283	0.023	0.075	0.156	0.160	0.131	
chr10:123242724-123242780		415	2	57	rs3135811	rs3135810		0.154	0.000	0.064	0.030	0.000	0.148	Balance
chr10:123275662-123275666	Yes	320	1	5	rs2912761	rs2981453		0.211	0.000	0.000	0.050	0.000	0.197	
chr10:123335839-123335866	Yes	1055	1	28	rs45631611	rs10886946		0.017	0.113	0.071	0.055	0.114	0.041	
chr10:123346116-123346190	Yes	420	0	75	rs2981575	rs1219648		0.195	0.048	0.000	0.022	0.013	0.090	
chr10:123396636-123396715		259	5	80	rs10788193	rs2454809		0.059	0.176	0.195	0.192	0.246	0.139	Coverage
chr10:123396728-123396806	Yes	331	2	79	rs1909670	rs1614303		0.029	0.176	0.100	0.131	0.073	0.057	
chr10:123406645-123406663	Yes	699	4	19	rs10788194	rs7923788		0.084	0.227	0.151	0.192	0.125	0.139	
chr11:534197-534242	Yes	2026	1	46	rs41258054	rs12628		0.000	0.153	0.056	0.137	0.076	0.025	
chr11:8246326-8246343		287	6	18	rs34544683	rs3816490		0.022	0.098	0.125	0.219	0.133	0.123	MH>2
chr11:69412090-69412124	Yes	2968	1	35	rs79274134	rs7112989		0.254	0.232	0.000	0.127	0.031	0.254	
chr11:69446719-69446766		1480	2	48	rs609581	rs11826558		0.070	0.015	0.058	0.079	0.187	0.139	Balance
chr11:69448373-69448445		563	7	73	rs654240	rs654648		0.188	0.029	0.136	0.112	0.210	0.221	MH>2
chr11:69508082-69508159		574	3	78	rs7930020	rs10908198		0.000	0.041	0.125	0.267	0.045	0.066	Balance
chr12:4333159-4333203		1194	4	45	rs7955734	rs7970219	rs7954847	0.268	0.092	0.026	0.085	0.100	0.229	Balance
chr12:4346169-4346177	Yes	646	0	9	rs11063052	rs11832328		0.318	0.079	0.038	0.072	0.080	0.213	
chr12:4351884-4352027	Yes	468	5	144	rs7955545	rs4766223		0.051	0.113	0.033	0.076	0.092	0.139	
chr12:4376089-4376091	Yes	306	2	3	rs4238013	rs12818766		0.119	0.033	0.181	0.161	0.147	0.139	
chr12:4399036-4399054	Yes	1619	2	52	rs3217859	rs3217860	rs3217861	0.325	0.391	0.414	0.491	0.479	0.303	
chr12:4399917-4399970	Yes	892	2	54	rs3217867	rs3217868	rs3217869	0.173	0.041	0.220	0.133	0.188	0.213	
chr12:4411639-4411683	Yes	1376	1	45	rs3217925	rs3217926		0.127	0.068	0.253	0.172	0.227	0.188	
chr12:4417127-4417232	Yes	1224	1	106	rs7133323	rs9668504		0.449	0.324	0.237	0.282	0.142	0.418	
chr12:4463020-4463036		850	31	17	rs7297048	rs7296130	rs12823973	0.115	0.089	0.193	0.303	0.117	0.131	MH>2
chr12:12009741-12009874	Yes	379	2	134	rs2238126	rs743614		0.181	0.240	0.190	0.249	0.079	0.180	
chr12:12013572-12013612	Yes	647	3	41	rs2855708	rs6488463		0.232	0.196	0.211	0.347	0.146	0.262	

Location														
chr12:12016008-12016089	Yes	1488	3	82	rs2238130	rs2416944	rs2238131	0.125	0.248	0.144	0.216	0.104	0.197	
chr12:12020114-12020170	Yes	637	1	57	rs2723805	rs7973930		0.241	0.111	0.075	0.066	0.054	0.238	
chr12:12033536-12033570		2184	4	35	rs11832110	rs4763730		0.232	0.000	0.042	0.074	0.000	0.189	Balance
chr12:12035649-12035664	Yes	2052	1	16	rs2710310	rs2739085		0.126	0.271	0.194	0.251	0.159	0.180	
chr12:18656174-18656225		381	1	52	rs11044141	rs11044142		0.099	0.000	0.000	0.000	0.000	0.041	Balance
chr12:56494991-56494998		3387	6	8	rs2271189	rs773123		0.073	0.000	0.110	0.066	0.070	0.066	MH>2
chr12:69169222-69169316	Yes	404	3	95	rs6581833	rs73334654		0.256	0.016	0.059	0.078	0.000	0.328	
chr12:69265196-69265278	Yes	768	0	83	rs3817605	rs2293637		0.310	0.192	0.022	0.111	0.106	0.271	
chr12:69277127-69277165	Yes	773	1	39	rs10878875	rs1663588		0.126	0.162	0.124	0.133	0.215	0.180	
chr12:121416622-121416650	Yes	3076	2	29	rs1169289	rs1169288		0.082	0.049	0.132	0.112	0.151	0.131	
chr12:121431272-121431300	Yes	1774	0	29	rs2071190	rs1169301		0.118	0.255	0.236	0.272	0.182	0.189	
chr12:121435427-121435475		3503	1	49	rs2464196	rs2464195		0.014	0.000	0.062	0.046	0.092	0.049	Balance
chr12:121437114-121437221		1919	0	108	rs55834942	rs1169304		0.012	0.000	0.166	0.082	0.023	0.025	Balance
chr12:133208886-133208979	Yes	739	2	94	rs5745023	rs5745022		0.173	0.105	0.135	0.219	0.049	0.180	
chr12:133226159-133226196	Yes	587	2	38	rs4883613	rs4883537		0.105	0.107	0.135	0.222	0.050	0.180	
chr12:133253995-133254083	Yes	448	1	89	rs5744751	rs5744750		0.000	0.105	0.100	0.045	0.042	0.025	
chr13:21562832-21562948		1715	3	117	rs2770928	rs558614		0.175	0.000	0.080	0.087	0.153	0.156	Balance
chr13:32854998-32855018		333	1	21	rs17692070	rs11617975		0.129	0.000	0.098	0.128	0.122	0.123	Balance
chr13:32986219-32986340	Yes	313	0	122	rs206319	rs206320	rs615762	0.107	0.204	0.175	0.244	0.262	0.107	
chr14:35872792-35872926		643	1	135	rs2233415	rs1050851		0.020	0.019	0.213	0.130	0.146	0.033	Balance
chr14:102568296-102568367		969	0	72	rs10873531	rs8005905		0.278	0.049	0.017	0.068	0.123	0.303	Balance
chr14:104165753-104165927		765	4	175	rs861539	rs1799796		0.114	0.073	0.295	0.229	0.189	0.115	Balance
chr14:105239146-105239192	Yes	521	5	47	rs3803304	rs2494732		0.169	0.097	0.171	0.290	0.302	0.230	
chr14:105258892-105258893	Yes	737	1	2	rs2494748	rs2494749		0.120	0.122	0.092	0.231	0.245	0.123	
chr15:41857216-41857303		1528	2	88	rs11639399	rs2277536		0.096	0.012	0.308	0.172	0.236	0.139	Balance

chr15:41860411-41860490		860	2	80	rs7171675	rs12148316		0.095	0.011	0.134	0.131	0.110	0.139	Balance
chr15:67457335-67457485	Yes	475	4	151	rs1065080	rs2289261		0.133	0.238	0.139	0.087	0.220	0.156	
chr15:73996066-73996101		3299	3	36	rs11574483	rs10083681		0.147	0.000	0.000	0.017	0.000	0.148	Balance
chr15:88488326-88488428	Yes	1800	1	103	rs8042993	rs1369426		0.088	0.135	0.153	0.097	0.261	0.066	
chr15:88549118-88549151	Yes	1763	0	34	rs11073758	rs12324332		0.266	0.015	0.124	0.133	0.079	0.303	
chr15:88554878-88554886		2501	3	9	rs4989257	rs898707		0.040	0.014	0.124	0.128	0.082	0.164	Balance
chr15:88646922-88647038	Yes	975	1	117	rs16941255	rs76506232		0.110	0.132	0.000	0.010	0.000	0.115	
chr15:88667852-88667948	Yes	1099	0	97	rs3784411	rs3784410		0.192	0.100	0.217	0.225	0.151	0.172	
chr16:2138218-2138269		1617	14	52	rs1800718	rs1748		0.136	0.000	0.090	0.063	0.024	0.074	MH>2
chr16:2138269-2138398		941	4	130	rs1748	rs13332221		0.249	0.000	0.116	0.017	0.123	0.197	Balance
chr16:2138398-2138422	Yes	2026	0	25	rs13332221	rs13332222		0.118	0.000	0.000	0.013	0.000	0.090	
chr16:81819768-81819820	Yes	2558	1	53	rs1143685	rs4294811		0.140	0.141	0.282	0.271	0.126	0.148	
chr16:89806343-89806347	Yes	601	2	5	rs11647746	rs7195906		0.161	0.013	0.074	0.035	0.134	0.156	
chr16:89849480-89849629		275	2	150	rs2239359	rs12448860		0.032	0.013	0.064	0.370	0.352	0.319	Balance
chr16:89858505-89858525		698	3	21	rs6500452	rs1800287		0.177	0.012	0.073	0.043	0.133	0.221	Balance
chr17:1782952-1782957	Yes	1284	1	6	rs5030755	rs2230930		0.000	0.000	0.102	0.020	0.024	0.016	
chr17:37807698-37807707		1082	2	10	rs9972882	rs75397452		0.141	0.000	0.047	0.042	0.000	0.131	Balance
chr17:37832279-37832315	Yes	1408	1	37	rs1495100	rs2934953		0.194	0.000	0.016	0.062	0.053	0.188	
chr17:37833562-37833567		2351	2	6	rs11656146	rs907088		0.000	0.000	0.092	0.030	0.011	0.016	Balance
chr17:37834357-37834367		666	0	11	rs732084	rs732083		0.181	0.000	0.000	0.043	0.011	0.156	Balance
chr17:37834715-37834808	Yes	1558	5	94	rs12150603	rs72832915		0.042	0.153	0.308	0.196	0.235	0.131	
chr17:37853048-37853097		1655	1	50	rs2643194	rs2517951		0.188	0.000	0.017	0.085	0.081	0.303	Balance
chr17:37893458-37893484		499	8	27	rs67597968	rs4795393		0.209	0.000	0.053	0.069	0.098	0.221	MH>2
chr17:41616392-41616456	Yes	1646	1	65	rs76280498	rs7222604		0.000	0.150	0.106	0.110	0.181	0.033	
chr17:78820329-78820374	Yes	3252	0	46	rs3751945	rs2589156		0.082	0.000	0.107	0.078	0.115	0.066	

chr17:78865546-78865630	Yes	631	3	85	rs2289764	rs2289765		0.289	0.044	0.111	0.110	0.115	0.303	
chr17:78896488-78896529	Yes	2726	4	42	rs2271602	rs2271603		0.154	0.196	0.321	0.291	0.307	0.197	
chr17:78897547-78897561	Yes	1725	0	15	rs7217786	rs6565491		0.031	0.199	0.122	0.111	0.249	0.049	
chr17:78921117-78921211	Yes	1576	2	95	rs4969231	rs9912373		0.022	0.079	0.124	0.114	0.060	0.082	
chr19:2226676-2226772	Yes	2349	1	97	rs3815308	rs2302061		0.034	0.182	0.143	0.172	0.203	0.090	
chr19:3110349-3110361		2345	5	13	rs11085000	rs41276846		0.018	0.193	0.112	0.063	0.188	0.016	
chr19:3119184-3119239	Yes	1438	1	56	rs308046	rs4900		0.166	0.233	0.135	0.101	0.275	0.271	
chr19:3119405-3119406		884	33	2	rs370286	rs451778		0.000	0.233	0.135	0.087	0.280	0.025	MH>2
chr19:5210622-5210782		740	2	161	rs2302224	rs1143698		0.166	0.066	0.126	0.134	0.090	0.131	Balance
chr19:5210762-5210782		4185	0	21	rs1143699	rs1143698		0.222	0.000	0.099	0.081	0.056	0.205	Balance
chr19:5212380-5212482		1945	1	103	rs1064300	rs2230611		0.115	0.000	0.124	0.135	0.126	0.098	Balance
chr19:7163154-7163230	Yes	810	2	77	rs2963	rs2245648		0.186	0.025	0.065	0.068	0.141	0.148	
chr19:7166376-7166388	Yes	1028	2	13	rs2059806	rs2229429		0.179	0.065	0.191	0.144	0.262	0.197	
chr19:10267011-10267077	Yes	265	0	67	rs4804490	rs2228611		0.171	0.281	0.068	0.184	0.224	0.172	
chr19:17937758-17937786		1721	0	29	rs3212798	rs3212797		0.074	0.000	0.052	0.033	0.012	0.066	Balance
chr19:17955001-17955021	Yes	1946	1	21	rs3212713	rs3212712	rs3212711	0.197	0.000	0.000	0.022	0.000	0.139	
chr19:30253901-30253998	Yes	768	2	98	rs117342492	rs4805475		0.000	0.221	0.000	0.104	0.073	0.033	
chr19:30255068-30255090	Yes	495	2	23	rs8103966	rs8099838		0.043	0.310	0.250	0.232	0.252	0.082	
chr19:30290349-30290357	Yes	2732	1	9	rs1473201	rs111640872		0.085	0.106	0.247	0.180	0.213	0.066	
chr19:30319892-30319896		284	5	5	rs56285422	rs8106054		0.085	0.106	0.248	0.180	0.213	0.066	LD
chr19:30340381-30340412	Yes	593	3	32	rs929813	rs929814		0.216	0.087	0.121	0.293	0.263	0.271	
chr19:30361995-30362112	Yes	290	2	118	rs255270	rs255271		0.184	0.104	0.037	0.068	0.012	0.271	
chr19:41724820-41724885	Yes	2049	0	66	rs2301236	rs28364580		0.094	0.179	0.224	0.148	0.275	0.172	
chr19:41781493-41781579	Yes	1040	2	87	rs8103839	rs9304592		0.067	0.073	0.000	0.066	0.064	0.066	
chr19:50919797-50919828	Yes	2886	5	32	rs3218776	rs3218760		0.125	0.139	0.075	0.148	0.275	0.156	

Location														
chr20:9543622-9543681	Yes	813	5	60	rs2297345	rs2297346		0.122	0.214	0.088	0.174	0.059	0.139	
chr20:30729488-30729523	Yes	3150	2	36	rs6089193	rs6089194		0.206	0.085	0.026	0.137	0.053	0.189	
chr20:40714479-40714540	Yes	1095	1	62	rs2016647	rs1569548		0.114	0.074	0.242	0.167	0.138	0.066	
chr20:40714539-40714540		1134	12	2	rs1569547	rs1569548		0.000	0.073	0.231	0.150	0.120	0.016	MH>2
chr20:57478807-57478939		711	8	133	rs7121	rs3730168		0.186	0.091	0.286	0.120	0.169	0.205	MH>2
chr21:42845374-42845383	Yes	6069	0	10	rs2298659	rs17854725		0.173	0.115	0.230	0.218	0.189	0.220	
chr21:42876400-42876447	Yes	2128	0	48	rs7277080	rs395584		0.287	0.017	0.019	0.235	0.212	0.229	
chr21:45643008-45643052		288	31	45	rs1048710	rs2298564		0.069	0.237	0.045	0.140	0.043	0.066	MH>2
chr22:17640022-17640045	Yes	1258	0	24	rs11550530	rs7287672		0.125	0.035	0.086	0.130	0.058	0.066	
chr22:21337266-21337325	Yes	565	4	60	rs178280	rs13054014		0.116	0.200	0.259	0.223	0.234	0.189	
chr22:21348914-21349037		1246	25	124	rs4822790	rs178292		0.105	0.224	0.135	0.112	0.142	0.139	MH>2
chr22:24158895-24158899	Yes	713	2	5	rs9608192	rs2070457		0.098	0.059	0.115	0.071	0.153	0.090	
chr22:29690246-29690345	Yes	259	0	100	rs73156524	rs131189		0.032	0.281	0.086	0.053	0.034	0.090	
Final Panel	164	768	1.78	59				0.139	0.139	0.139	0.156	0.153	0.155	

573 Supplementary Table 2

		Threshold Median 3rd MH Frequency				
		0.5	1	1.5	2	2.5
In silico Mixing Levels	0.5	40	0	0	0	0
	1	100	0	0	0	0
	1.5	100	7	0	0	0
	2	100	63	0	0	0
	2.5	100	100	2	0	0
	3	100	100	35	0	0
	4	100	100	95	15	0
	5	100	100	100	88	7
	10	100	100	100	100	100