Coalescent-based species delimitation meets deep learning: Insights from a highly
fragmented cactus system

MANOLO F. PEREZ[1,2,¥], ISABEL A. S. BONATELLI[1,3,,¥], MONIQUE ROMEIRO-BRITO[1], FERNANDO

F. FRANCO[1], NIGEL P. TAYLOR[4], DANIELA C. ZAPPI[5], EVANDRO M. MORAES[1,*]

[1]*Departamento de Biologia, Universidade Federal de São Carlos, Rodovia João Leme dos*

Santos km 110, Sorocaba, SP, 18052780, Brazil;

[2]*Departamento de Genética e Evolução, Universidade Federal de São Carlos, Rodovia*

Washington Luiz km 235, C.P. 676, São Carlos, SP, 13565905, Brazil;

[3]*Departamento de Ecologia e Biologia Evolutiva, Universidade Federal de São Paulo,*

Diadema, SP 09972-270, Brazil;

[4]*University of Gibraltar, Gibraltar Botanic Gardens Campus, The Alameda, PO Box 843,*

Gibraltar;

[5]*Programa de Pós Graduação em Botânica, Instituto de Ciências Biológicas, Universidade*

de Brasília, PO Box 04457, Brasília, DF, 70910970, Brazil.

[¥] *Manolo F. Perez and Isabel A. S. Bonatelli contributed equally to this article.*

* Correspondence to be sent to Departamento de Biologia, Universidade Federal de São

Carlos, Rodovia João Leme dos Santos km 110, Sorocaba, SP, 18052780, Brazil;

E-mail: emarsola@ufscar.br

Abstract. - Delimiting species boundaries is a major goal in evolutionary biology. An

increasing body of literature has focused on the challenges of investigating cryptic diversity

within complex evolutionary scenarios of speciation, including gene flow and demographic

fluctuations. New methods based on model selection, such as approximate Bayesian

computation, approximate likelihood, and machine learning approaches, are promising tools

arising in this field. Here, we introduce a framework for species delimitation using the

multispecies coalescent model coupled with a deep learning algorithm based on

convolutional neural networks (CNNs). We compared this strategy with a similar ABC

approach. We applied both methods to test species boundary hypotheses based on current and

previous taxonomic delimitations as well as genetic data (sequences from 41 loci) in

Pilosocereus aurisetus, a cactus species with a sky-island distribution and taxonomic

uncertainty. To validate our proposed method, we also applied the same strategy on sequence

data from widely accepted species from the genus *Drosophila*. The results show that our

CNN approach has high capacity to distinguish among the simulated species delimitation

scenarios, with higher accuracy than the ABC procedure. For *Pilosocereus*, the delimitation

hypothesis based on a splitter taxonomic arrangement without migration showed the highest

probability in both CNN and ABC approaches. The splits observed within *P. aurisetus* agree

with previous taxonomic conjectures considering more taxonomic entities within currently

accepted species. Our results highlight the cryptic diversity within *P. aurisetus* and show that

CNNs are a promising approach for distinguishing divergent and complex evolutionary

histories, even outperforming the accuracy of other model-based approaches such as ABC.

Keywords: Species delimitation, fragmented systems, recent diversification, deep learning,

Convolutional Neural Networks, Approximate Bayesian Computation

INTRODUCTION

Recognizing species boundaries has long been a major challenge for biologists. The

major difficulty is to some degree related to the numerous existing species concepts. The use

of distinct concepts can lead to different strategies for identifying species boundaries in

empirical datasets (de Queiroz 2007; Carstens et al. 2013). However, different species

concepts can be considered elements related to diverse properties that are associated with the

dynamics of the speciation continuum. After the proposal of the unified species concept

(Queiroz, 2007), the roles of species concepts as a theory and species delimitation as a

methodology become apparent. The view of species as independent segments of a

metapopulation indicated lineage independence as the only criterion necessary for delimiting

species boundaries, avoiding any disagreement purely related to the species concept.

Selecting a suitable approach for species delimitation has been difficult, especially in

species complexes (Pinheiro et al. 2018). Identifying discontinuities among incipient stages

of divergence, such as those found in these systems, demands great effort and a

multidisciplinary approach to assess different sources of evidence supporting species limits

(Carstens et al. 2013). In this context, estimates based on independent sources of data such as

morphology, cytogenetics, anatomy, ecology, and genetics have been used to achieve a better

resolution in species delimitation (Domingos et al. 2014; Alvarado-Sizzo et al. 2018; Denham

et al. 2019). Some of these traits, such as phenotypes and geographic distributions, can be a

starting point for hypotheses about species circumscriptions (Solís-Lemus et al. 2015; Luo et

al. 2018).

Species delimitation methods based on multilocus data and the multispecies coalescent

model (MSC) compare the probability of species trees with different numbers of operational

taxonomic units (OTUs) to identify optimal partitions for the data (Ence and Carstens 2011).

Highly fragmented systems impose potential caveats on such methods, potentially resulting in

the oversplit of existing diversity, and thus, highly subdivided entities (Sukumaran and

Knowles 2017). Jackson et al. (2017) proposed the incorporation of the genealogical

divergence index (*gdi*) as an attempt to reduce the delimitation of population structure,

instead of species limits. Furthermore, the developers of the bayesian delimitation method

implemented in BPP (Yang and Rannala 2014) described an approach to integrate *gdi* on the

outputs of their software and showed that such a strategy was efficient in mitigating the effect

of over splitting (Leaché et al. 2018). However, a number of concerns on the use of *gdi* have

been raised. For instance, a large interval of *gdi* values (0.2 to 0.7) that corresponds to a 'grey

zone' where delimitation is ambiguous, and datasets containing putative species with

population sizes very different or extremely small (Rannala and Yang 2020).

Model selection methods using simulated datasets under competing delimitation

hypotheses are a promising tool for taxa that have potentially experienced a complex

evolutionary history, including recurrent gene flow and demographic fluctuations. These

methods also allow us to easily test for different assignments and topologies of the analyzed

samples into populations/species, a procedure that is not straightforward with other

approaches (but see Leaché et al. 2014). Examples of such approaches include approximate

Bayesian computation (ABC; Camargo et al. 2012), approximate likelihood analysis

(Morales et al. 2017), machine learning coupled with ABC (Smith and Carstens 2020) and

machine learning methods based on summary statistics (SuSt) (Pei et al. 2018). Among them,

the ABC represents a class of flexible likelihood-free algorithms for performing Bayesian

inference (Beaumont et al. 2002). The principle behind the method relies on the massive

simulation of genetic data using parameter values drawn from a prior distribution, followed

by the calculation of SuSt for each simulation. SuSt are values generated from the raw

genetic data, which are expected to capture important information to differentiate among the

simulated models. Simulations that produce genetic variation patterns (SuSt) close to the

observed data are retained to form an approximate sample from the posterior distribution.

A recently developed approach for demographic model selection, based on deep

learning image classification, uses convolutional neural networks (CNNs) to retrieve as much

information as possible from genetic datasets by converting them to images without requiring

user-specified SuSt (Flagel et al. 2019). Simulations have shown that CNNs maximize the

use of available information from the data and increase the ability to distinguish divergent

evolutionary histories, frequently overcoming the limitations of traditional approaches (Flagel

et al. 2019). Despite the recent use of CNNs in population genetics (Flagel et al. 2019;

Sanchez et al. 2020) and phylogeography (Souza et al. 2019; Oliveira et al. 2020; da Fonseca

et al. 2020), this approach has not previously been tested in the context of species

delimitation using genetic data.

Here, we introduce an approach for species delimitation that combines the use of MSC-

based methods with model selection via CNN. We compared our model selection method

with ABC to evaluate distinct species delimitation hypotheses in the cactus *Pilosocereus*

aurisetus. We chose this taxon because (1) it presents a controversial taxonomy, and (2) it has

a highly fragmented geographical distribution, comparable to other sky-island systems,

imposing recognized difficulties upon MSC-based methods (Leaché et al. 2018).

To gain better insights into the species limits in *P. aurisetus,* we used the following

protocol (Fig. 1): (1) apply the MSC approach implemented in BPP (Yang and Rannala 2014)

with sequence data and OTUs based on sampling localities, (2) apply the *gdi* index to the

BPP results, to obtain a hypothesis less prone to oversplitting, (3) simulate data under the

obtained BPP results (with and without *gdi*) and for each taxonomic hypothesis, with and

without gene flow among lineages (this step requires some previous knowledge about the

study group, such as divergence times, effective population size, and number of migrants per

generation, when applicable), (3) transform the simulated data into binary images, defining

the major and minor state for each site, (4) train a CNN to recognize simulations generated

from each model, (5) evaluate the accuracy of the trained CNN in comparison to an ABC

approach, by using cross-validation tests based on simulated data (i.e. independent

simulations not evaluated before), and (6) predict model probability with the trained network

(CNN) or by approximating the posterior (ABC). We also validated our method by following

this same procedure using a recently published dataset from two *Drosophila* species pairs

(Campillo et al. 2020), which have information about pre- and postzygotic reproductive

isolation and do not have controversial taxonomy.

MATERIAL AND METHODS

The Studied System

The taxon *P. aurisetus* is a puzzling system regarding species delimitation. The

taxonomy of *P. aurisetus* has been unstable over the years, which is a common attribute in

the family Cactaceae, probably related to the absence of clear diagnostic characters for some

groups due to convergent and parallel evolution (Hernández-Hernández et al. 2011; Copetti et

al. 2017). The morphological variation observed within this taxon has led to repeated

taxonomic evaluations, with species and subspecies being recurrently synonymized and re-

established (Zappi, 1994; Taylor and Zappi, 2004; Hunt et al., 2006). Another complicating

issue is the naturally fragmented distribution of *P. aurisetus*. The taxon is associated with the

Espinhaço Mountain Range campo rupestre landscapes in eastern Brazil, occurring on

mountaintops separated by valleys and lowland vegetation (Fig. 2). In line with this sky-

island distribution, *P. aurisetus* shows high intraspecific genetic structure (Bonatelli et al.

2014; Perez et al. 2016a, b). Currently, two subspecies are recognized based on differences in

the size and diameter of stems and the color and density of hairs on flower-bearing areoles:

the more widespread *P. aurisetus* subsp. *aurisetus* (Werderm.) Byles & Rowley, and *P.*

aurisetus subsp. *aurilanatus* (Ritter) Zappi, which occurs as few populations restricted to a

disjunct mountain (Serra do Cabral) west of the Espinhaço Mountain Range. We also

included in our study plants from three heterotypic synonyms of *P. aurisetus* subsp. *aurisetus*

(Fig. S1): *P. aurisetus* subsp. *densilanatus* (Ritter) Braun & Esteves, located in the

easternmost populations from the Serra Negra Mountains and distinguished by their densely

wooly stems, reaching up to 4 cm diameter; *P. aurisetus* subsp. *supthutianus* (Braun) Braun

& Esteves, a northern outlier of the species that also present densely hairy stems, which can

reach more than 5 cm diameter; and *P. aurisetus* subsp. *werdermannianus* (Buining &

Brederoo) Braun & Esteves, located in the central populations and showing more slender

stems, fewer ribs and sometimes green epidermis.

Sampling and DNA extraction

We sampled four individuals of *P. aurisetus* in each of nine localities in eastern Brazil,

totaling 36 sampled individuals and covering the entire known distribution of the species.

Hereafter, we refer to these sampled localities as populations justified by the clear limits of

the *P. aurisetus* habitat patches and the marked genetic structure in this taxon (Bonatelli et al.

2014). This sampling included representatives of all accepted and synonymized taxa

mentioned above (Fig. 2; Table 1). Root tissue was stored in silica gel and then transferred to

a -80°C freezer until DNA extraction. We maintained a distance of approximately 10 m

between sampled individuals to avoid collecting clones. Total DNA was extracted using the

Extract All kit (Applied Biosystems) and purified with 95% ethanol and 3 M sodium acetate

buffer. The DNA concentration was measured with a Qubit fluorometer (Invitrogen).

Microfluidic PCR and sequencing

Massive parallel target amplification of 41 loci in 36 samples was carried out in

microfluidic PCR reactors (Access Array System, Fluidigm). The selected loci included 26

anonymous nuclear markers developed for *P. aurisetus* (Perez et al. 2016c) and another five

nuclear, eight plastid and two mitochondrial genic regions selected from the available data in

GenBank (Supplementary Table S1). Development of these new markers was based on

available sequences from Cactaceae or from other related plant family species to detect

conserved regions for primer design. All primers were developed with PRIMER3 v4.0.0

(Untergrasser et al. 2012), with parameters suggested by the Fluidigm Assay Design Group

as follows: primer size from 18–23 bases, annealing temperature between 59°C and 61°C,

maximum poli-X of 3. The selected markers were synthesized according to Fluidigm

specifications and applied to a single 48×48 access array chip with reactions performed

according to the manufacturer's protocol. The obtained PCR products were pooled and

purified with 0.6X AMPure beads (Agencourt). The quality and amplification range were

visualized in a BioAnalyzer, and the concentration was estimated via real-time PCR using

KAPA qPCR (Kapa Biosystems). The samples were subjected to a MiSeq (Illumina) 300 bp

paired-end run along with 156 other samples from other projects.

After sequencing, a first filtering step was performed with cutadapt (Martin 2011), excluding

sequences smaller than 60 bp and removing 3' bases with a PHRED score <Q22. PHRED

quality for all reads was then visualized in FastQC

(http://www.bioinformatics.bbsrc.ac.uk/projects/fastqc) and graphically compiled in MultiQC

(http://multiqc.info/). Reference sequences for each marker were generated from the

alignments used to design the primers. Mapping reads for each marker was performed with

BWA-MEM (Li 2013), which is suited for pair-end long reads with indels, while SAMtools

(Li et al. 2009) was used to retain only sequences that were mapped with high fidelity. SNP

calling was carried out with GATK (McKenna et al. 2010) by transforming low-quality bases

in missing data, identifying possible indels with realignment, applying polymorphism filters

for quality and coverage, and generating FASTA files for each sample. To confirm the

obtained polymorphisms, each detected SNP was then mapped against reference files for

each marker that were built from reads from each analyzed sample. The phasing of SNPs in

nuclear markers was achieved with a Python script, modified from Harvey et al. (2016). The

aligned sequences from each marker were obtained with MAFFT v.7 (Katoh and Standley

2013) with default parameters. Recombination within each marker was tested with the DSS

method (McGuire and Wright 2000) implemented in TOPALi v2 (Milne et al. 2008) and

topological incongruences were tested in KDETrees (Weyenberg et al. 2014). The most likely

substitution model for each locus was obtained with PartitionFinder v.1.1.1 (Lanfear et al.

2012).

Species Tree and Coalescent Delimitation

The phylogenetic relationships among populations were recovered with a species tree in

BEAST v2.1.3 (Bouckaert et al. 2014), assigning the sequences from each population to a

single OTU. For all markers, we used the selected substitution model (Table S1) and a

Relaxed clock Log-Normal, with the mean sampled from a broad lognormal distribution

(according to Perez et al. 2016b). We performed three runs with 5×10^8 MCMC iterations

each, sampling trees every 10^4 generations with 75% burn-in. Convergence was observed in

TRACER 1.6 (Rambaut et al. 2014) and a Maximum Clade Credibility (MCC) tree with

median heights was obtained in TreeAnnotator (Drummond and Rambaut 2007).

We used the obtained species tree topology and the genomic sequences obtained from

microfluidic PCR as input to test the species limits in *P. aurisetus* with the method

implemented in BPP v3.4 (Yang and Rannala 2014). For this analysis, we concatenated all

mitochondrial (*cox1* and *cox2*) and chloroplast (*atpB-rbcL*, *petL-psbE* and *rpl16*) markers.

The BPP method uses a reversible jump Bayesian (rjMCMC) framework to estimate the

posterior probability of existence at each node of the phylogeny. To ensure convergence, five

independent runs were performed to estimate the best delimitation scheme, with parameters

selected according to the values suggested by Leaché et al. (2018) combined with previous

estimates for the species from Perez et al. (2016a). θ and τ priors followed an inverse gamma

(IG) distribution with α=3 and a β of 0.01 and 0.002, respectively (hereafter BPP 'specific

prior values'). We evaluated the potential effect of priors in the delimitation results by also

performing the same number of runs with broad prior values $\theta \sim IG(3, 0.002)$ and $\tau \sim IG(3,$

0.03) as suggested in the BPP manual (hereafter 'diffuse prior values'). We considered a node

as valid only if both analyses support its existence with a posterior probability (PP) of 0.99 or

higher. The runs were carried out for 10^5 iterations sampled every five steps after discarding

the first 10^4 iterations as burn-in. The obtained delimitations were mapped to the tips of the

species tree topology. As a more conservative alternative, we also evaluated our BPP results

using the *gdi* index, as suggested by Leaché et al. (2018). We considered the threshold values

suggested by Jackson et al. (2017) and Leaché et al. (2018), in which pairwise *gdi* values

smaller than 0.2 indicate a single species and above 0.7, two species.

Comparing Delimitation Hypotheses with Deep Learning and ABC

We adopted an integrative approach to establish putative species limits by considering

information on *P. aurisetus* from different sources and analyses, as suggested by Carstens et

al. (2013). For that, we compared the two delimitation hypotheses generated by the BPP

results (with and without the *gdi* index), as well as two morphological hypotheses based on

one lumper and one splitter taxonomic arrangement (Table 1; Fig. 1). The genetic hypotheses,

based on the BPP results, were composed of one species when *gdi* was considered and four

species when it was not incorporated (see results below); the 'splitter' morphological

hypothesis, considering both currently accepted and previously synonymized taxa, was

composed of four species; and the 'lumper' morphological hypothesis, considering only the

currently recognized subspecies, was composed of two species (Hunt et al. 2006). To test

these competing delimitation hypotheses, we considered scenarios including the number of

species and the potential migration between the delimited entities. Therefore, seven scenarios

(Fig. 1) were tested: (1) 'splitter', (2) 'splitter' combined with migration, (3) 'lumper', (4)

'lumper' with migration, (5) BPP combined with *gdi* (BPP_GDI), (6) BPP without *gdi*

(BPP_noGDI), and (7) BPP_noGDI with migration. As the BPP_GDI scenario consists of a

single species, an additional model including migration was not necessary. We performed

model comparison using both a deep learning approach based on CNN and a regular ABC

method. For these two strategies, we simulated genetic datasets under each scenario with a

modified version of the scripts from Perez et al. (2016a). To simplify our simulations and

avoid using an overly heterogeneous dataset, we included only nuclear markers and simulated

data mirroring sample sizes with the same number of segregating sites per loci of our

empirical dataset. For the deep learning approach, the simulated data were converted to

images that were used to train a CNN to recognize simulations generated by different

scenarios (for details on the simulation and training procedure, see Supplementary File 1).

The comparison of delimitation scenarios using regular ABC analysis (Fig. 1) was performed

following the approach of Perez et al. (2016a) with simulations conducted under the same

conditions used for the CNN (details in Supplementary File 1).

To validate our newly proposed deep learning method for species delimitation, we also

carried out the same analyses using a dataset recently published by Campillo et al. (2020).

These authors compared the results of reproductive isolation and coalescent-based (BPP)

delimitation methods to infer species boundaries in several species pairs of the genus

Drosophila, a classic model system in speciation research. We focused our approach on data

from two species pairs of the Melanogaster group: *D. melanogaster - D. sechellia* and *D.*

melanogaster - D. simulans. We chose these species because they are widely accepted and

corroborated by both BPP and reproductive isolation metrics (Campillo et al. 2020).

Furthermore, while the *D. melanogaster - D. sechellia* pair is allopatric, the *D. melanogaster*

- D. simulans pair is sympatric, allowing us to explore whether geographic context might

affect our ability to identify these species. Therefore, we tested a model with all specimens

belonging to the same species against a model in which the specimens were divided into two

species, which was the expected outcome according to the current classification. Again, our

simulations were based on empirical sample sizes and the number of segregating sites (details

in Supplementary File 1), with priors based on the values suggested by Campillo et al.

(2020).

RESULTS

Microfluidic PCR and sequencing

Four markers showed no amplification in any sample (*PaANL_46*, *petA*, *rpS16*, and

ycf1). To further minimize missing data and ensure that at least one individual was sequenced

from each population, we discarded 16 loci (*PaANL_8*, *PaANL_10*, *PaANL_50*, *PaANL_82*,

PaANL_123, *PaANL_134*, *PaANL_142*, *PaANL_155*, *PaANL_160*, *PaANL_187*,

PaANL_196, *its2*, *ppc*, *nhx1*, *psbA-psbB*, *psbC*). No recombination was detected at any of the

remaining loci, but kdetrees result suggested an incongruent topology in the *isi1* marker,

which was removed. Therefore, the final dataset included 20 markers that were considered in

the subsequent analyses for *P. aurisetus* (*PaANL_15*, *PaANL_17*, *PaANL_28*, *PaANL_35*,

PaANL_80, *PaANL_87*, *PaANL_96*, *PaANL_126*, *PaANL_140*, *PaANL_147*, *PaANL_164*,

PaANL_165, *PaANL_182*, *PaANL_205*, *apk1*, *atpB-rbcL*, *petL-psbE*, *rpl16*, *cox1* and *cox2*),

with a total of 8,908 bp.

Species Tree and Coalescent Delimitation

The species tree recovered most nodes with high support, especially the ones defining

previously and currently valid subspecies (Fig. 3). Population JFE of *P. aurisetus* subsp.

aurilanatus was the most external clade. Other populations were arranged in two subclades.

One of them contained populations in the center (INA and MEN, previously classified as *P. a.*

subsp. *werdermannianus*; ITA and PMN, formerly *P. a.* subsp. *densilanatus*) and south (COC)

of the *P. a.* subsp. *aurisetus* distribution. The second subclade contained populations from

northern distribution (EDB, BOV and ODA, formerly classified as *P. a.* subsp. *supthutianus*).

All BPP runs under each prior set showed the same results, therefore we provided the

mean posterior probability of each node in Fig. 3. Results with 'specific' and 'diffuse' priors

were similar, though the latter was less splitter. The congruent delimited species were *P. a.*

subsp. *aurisetus* (COC), *P. a.* subsp. *werdermannianus* (EDB, BOV and ODA) and the

currently recognized subspecies *P. aurisetus* subsp. *aurilanatus* (JFE). The only exception

was for the node separating the population ODA from the remaining *P. a.* subsp.

supthutianus populations (BOV and EDB), which was validated (PP=1) only in the 'specific'

but not (PP=0.95) in the 'diffuse' prior set. To be conservative, we decided to collapse this

node, considering all populations once recognized as *P. a.* subsp. *supthutianus* as a single

unit. Therefore, BPP resulted in four delimited species, that coincided with the 'splitter'

hypothesis, except for the OTUs composed by populations once recognized as *P. a.* subsp.

werdermannianus and *P. a.* subsp. *densilanatus*, which were collapsed (Fig. 3). When we

analyzed our BPP results with the *gdi* index, all estimates were below 0.7 and only three (*P.*

aurisetus subsp. *aurilanatus* - JFE; *P. a.* subsp. *aurisetus* - COC; and *P. a.* subsp.

supthutianus - ODA, BOV and EDB) had values between 0.2 and 0.7 (Fig. S2). Therefore, to

be conservative, we decided to collapse all populations in a single species for this hypothesis.

Delimitation Hypotheses Comparison

After 250 epochs, our CNN showed accuracies of 96.78% and 92.71% for the training

and validation sets, respectively. Some degree of overfitting was observed when we plotted

the accuracy of the training and validation sets throughout the epochs. However, this effect

decreased as the number of evaluated simulations increased (Fig. S3). Our cross-validation

procedure, using a test set of simulations not evaluated during training, also showed that

increasing the number of simulations resulted in accuracy improvement for all models (Fig.

4). It was also possible to observe that CNN presented a higher proportion of simulations that

were correctly predicted in relation to their generating model than the ABC results, with all

models being correctly assigned with more than 85% accuracy when 10,000 simulations per

model were used (Fig. 4 and S4). The percentages of correct predictions with the ABC

approach were all higher than 70% when 10,000 simulations per model were used, except for

the models with more delimited species that showed ~50% accuracy (Fig. 4). These two types

of models with lower accuracy in the ABC analysis ('splitter' and 'BPP_noGDI') showed the

most confusion with each other (Fig. S4). When the empirical data were submitted to the

trained CNN, the 'splitter' hypothesis was selected with a softmax probability of 99.8%

(Table 2). Our CNN approach showed consistent results, selecting the same delimitation

scenario even when the order of SNPs was shuffled (Supplementary File 1).The ABC

approach showed similar results, also recovering the 'splitter' hypothesis (PP=0.994). These

results are in agreement with what is observed when we plot the simulated and empirical

genotypes with a PCA (Fig. S5), showing that the empirical data are located inside the cloud

of simulations from the 'splitter' model .

The *Drosophila* dataset also pointed to a higher accuracy of CNN over ABC. The

cross-validation tests indicate a correct assignment of CNN simulations just above 78% and

81% in the *D. melanogaster-D simulans* and *D. melanogaster-D. sechellia* pairs, respectively

(Fig. S4). The accuracy of ABC was lower, reaching values slightly higher than 70% and

65% for the same species pairs (Fig. S4). When the empirical data were used, both methods

supported the hypothesis of two species for the *D. melanogaster-D simulans* pair, showing a

probability of 99.2% in CNN and 98.2% for ABC. In *D. melanogaster-D. sechellia,* the CNN

pointed to two species with a probability of 94.9%, while the ABC suggested a single species

with 89.4% PP (Table 2).

DISCUSSION

Deep learning in coalescent-based species delimitation

The advantages of the simulation approaches adopted here (CNN and ABC) over explicit statistical methods are that they allow testing models with complex demographic scenarios, such as migration events or size fluctuations. In addition to this use, these approaches allow to easily apply different assignments of samples into OTUs (but see Leaché et al. 2014) and even compare hypotheses relying on distinct topologies. Our comparison of these two approximate approaches suggests that the deep learning approach (CNN) showed a higher capacity to distinguish among the simulated demographic scenarios, outperforming the ABC procedure (Fig. 4 and S4). It is important to highlight that CNN achieved this performance with longer, though not impeditive, running times (~21 minutes in a Nvidia K80 GPU with an Intel Xeon 2.30GHz CPU) than ABC (~1 minute in an Intel Core i7 2.9GHz CPU). The results from the *Drosophila* dataset also supported a higher accuracy of CNN compared to ABC (Fig. S4). Moreover, when the empirical data was evaluated, only CNN recovered two species for the allopatric pair *D. melanogaster-D. sechellia* (Table 2). These results validated our CNN approach, as they are in agreement with our expectations based on Campillo et al. (2020).

Regarding the *P. aurisetus* dataset, both CNN and ABC approaches selected the 'splitter' model without migration as the most likely scenario with high PP (Table 2). The main difference between CNN and ABC functioning is the ability of the former to take information directly from SNP matrices without using SuSt, which is required by the latter. Moreover, while ABC relies on a rejection step that discards most of the simulations and leaves only a small proportion that are more similar to the empirical data (Csilléry et al. 2012), the CNN uses information from the whole set of simulations to learn how to

distinguish the concurrent scenarios (Schrider and Kern 2018). The inferior performance of

the ABC analyses could also be related to the choice of SuSt with overly high dimensionality

(Robert et al. 2011; but see Kousathanas et al. 2016) or with little information to distinguish

among the tested models. To reduce dimensionality, we adopted the suggestion of Perez et al.

(2016a), which consists of the transformation of the raw SuSt with a PCA and the retention of

only the axis necessary to ensure at least 99% of the variance. Other strategies usually applied

for this purpose (reviewed in Prangle 2015) include partial least squares regression

(Wegmann et al. 2009), linear regression (Fearnhead and Prangle 2012) or boosting

(Aeschbacher et al. 2012).

Although our CNN results were very encouraging, we note that care must be taken in

interpreting them, as deep learning techniques can suffer from overfitting (Nguyen et al.

2015; Ponti et al. 2017). To evaluate overfitting, we observed the history plot of the accuracy

throughout the epochs (Fig. S3) and adopted a cross-validation approach based on a test set,

which consisted of simulated data that were not evaluated during training. The results pointed

to high accuracy (Fig. 4 and Fig. S4), which is in agreement with the similarity of our

empirical and simulated data (Fig. S5). Another recent approach that could be coupled with

our model comparison strategy is based on unsupervised machine learning (Derkarabetian et

al. 2019). This strategy showed promising results and was particularly robust in

distinguishing population structure from species-level divergence.

Delimiting Species in P. aurisetus

Pilosocereus aurisetus populations diverged very recently in the last one million years

(Perez et al. 2016a). Delimiting species with recent diversification has been considered

challenging, and the main difficulty arises from the short time for species to accumulate

diagnostic characteristics that allow taxonomic distinction (Salicini et al. 2011). A similar

effect is observed in genetic data and is commonly referred to as incomplete lineage sorting

(ILS). ILS appears when ancestral genetic variants persist as polymorphisms across

successive diversification events. This phenomenon has been indicated as the primary cause

of discordance between gene trees and species trees in Cactaceae (Copetti et al. 2017).

Another issue that imposes additional challenges for species delimitation in *P. aurisetus*

is its sky-island distribution, leading to high population divergence (Bonatelli et al. 2014).

Topography-driven isolation in species of the Brazilian rupestrian highland habitats ("campos

rupestres"), such as *P. aurisetus*, has been considered an important factor driving speciation

and high microendemism in these landscapes (Vasconcelos et al. 2020; Zappi et al. 2017).

The major concern in this kind of system is that MSC may fail to discriminate between

genetic structure associated with population isolation and species boundaries (Jackson et al.

2017; Sukumaran and Knowles 2017; Leaché et al. 2018). Although *gdi* appears as a possible

solution for the overspliting tendency of MSC methods (Jackson et al. 2017), mainly in

highly structured systems, it is still not clear if it could lead to an opposite effect - over

lumping differentiated species. Another possibility for overcoming this bias is to incorporate

a priori knowledge to determine the structure associated with population-level processes and

related to the species level. An ideal scenario would incorporate data from a plethora of

classes, such as morphological, ecological, and genetic data, to recover more robust species

boundaries (Pinheiro et al. 2018). Since no single method can discriminate between these

processes, the results from species delimitation using multispecies coalescent models (MSC)

in sky-island systems should be treated as species hypotheses that need further confirmation

from multiple sources (Sukumaran and Knowles, 2017).

Taxonomic Implications for P. aurisetus

424  Our species delimitation investigation within *P. aurisetus* is discordant from the

425 currently recognized taxonomic diversity of this taxon (Hunt et al. 2006), as the 'splitter'

426 hypothesis has been recovered with higher probability with different methods. Convergent

427 evolution as well as the level of plasticity commonly found in cactus species (Guerrero et al.

428 2018) might be related to the disagreement in the number of species delimited by current

429 taxonomy and the detected genetic lineages, as observed within the genus *Pilosocereus*

430 (Bonatelli et al. 2014; Perez et al. 2016b). Therefore, our results should provide some

431 guidance for taxonomists interested in re-evaluating the taxonomy of *P. aurisetus*. In this

432 context, our results may help to fill gaps in the traditional taxonomy and provide

433 recommendations for a taxonomic treatment that better reflects the diversity of species,

434 leading us to a multidisciplinary approach for better understanding the boundaries within

435 species complexes considering their evolutionary history.

436  Previous genetic studies have identified well-differentiated lineages within *P. aurisetus*

437 and related species, with monophyletic groups more frequently being associated with the

438 geographic distribution than the proposed taxonomic boundaries (Fig. 2; Bonatelli et al. 2014;

439 Perez et al. 2016b; Calvente et al. 2016; Khan et al. 2018; Lavor et al. 2019). Different

440 taxonomic descriptions have been considered in the past for some of the recovered lineages

441 within *P. aurisetus* that agree with our results. For instance, the populations occurring in the

442 Serra Negra Mountains (ITA and PMN localities), in the center (MEN and INA) and in the

443 northern distribution of the species (EDB and BOV localities) were previously reported as *P.*

444 *aurisetus* subsp. *densilanatus* , *P. aurisetus* subsp. *werdermannianus*, and *P. aurisetus* subsp.

445 *supthutianus* (Braun and Esteves Pereira, 1995), respectively. These subspecies propositions

446 were based on some level of morphological variation that initially seemed rather distinctive,

447 such as the densely wooly stems observed in the ITA populations (Fig. S1). However, Taylor

448 and Zappi (2004) argued that such differences are not sufficiently important to merit

recognition as an additional subspecies. These authors reasoned that if this level of

morphological difference was to be recognized, the logical extension would be to give similar

status to other populations that show slightly divergent characteristics in terms of size, white-

wooly stems, and glaucousness.

CONCLUSIONS

Although the splits within species could be an artifact of the MSC, we might recognize

previous taxonomic issues that agree with the inferred species in this investigation.

Pilosocereus aurisetus exhibits many biological characteristics that make species delimitation

challenging, such as recent divergence, a sky-island distribution and unstable taxonomic

history. Here, we sought to investigate the species limits in this complex system and, for the

first time, incorporated a deep learning approach to select the best species hypothesis

according to our data and simulations. Finally, we stress that all the species hypotheses

reported here should be considered, as any of the taxonomic species in fact are hypotheses

prone to validation with other methods and sources of information.

SUPPLEMENTARY MATERIAL

All scripts and datasets used are available in GitHub:

https://github.com/manolofperez/CNN_spDelimitation_Piloso.

FUNDING

This work was supported by grants from the São Paulo Research Foundation (FAPESP)

(2015/06160-5 to EMM, 2012/22943-1 to MFP, and 2012/22857-8 to IASB); the National

Council for Scientific and Technological Development (CNPq) (03940/2019-0 to EMM,

305301/2018-7 to DCZ); and the Coordination for the Improvement of Higher Education

Personnel (CAPES) (Finance Code 001 to MRB).

ACKNOWLEDGMENTS

We thank Heidi Utsunomiya and Juliana de Fátima Martinez for laboratory assistance and

Gerardus Olsthoorn and Marlon Machado for sampling assistance.

REFERENCES

Aeschbacher S., Beaumont M.A., Futschik A. 2012. A novel approach for choosing summary

statistics in approximate Bayesian computation. Genetics 192:1027–1047.

Alvarado-Sizzo H., Casas A., Parra F., Arreola-Nava H.J., Terrazas T., Sánchez C. 2018.

Species delimitation in the *Stenocereus griseus* (Cactaceae) species complex reveals a

new species, *S. huastecorum*. PLoS One. 13:e0190385.

Beaumont M.A., Zhang W., Balding D.J. 2002. Approximate Bayesian computation in

population genetics. Genetics 162: 2025–2035.

Bonatelli I.A.S., Carstens B.C., Moraes E.M. 2015. Using next generation RAD sequencing

to isolate multispecies microsatellites for *Pilosocereus* (Cactaceae). PLoS One.

10:e0142602.

Bonatelli I.A.S., Perez M.F., Townsend A.P., Taylor N.P., Zappi D.C., Machado M.C., Koch

I., Pires A.H.C., Moraes E.M. 2014. Interglacial microrefugia and diversification of a

cactus species complex: Phylogeography and palaeodistributional reconstructions for

Pilosocereus aurisetus and allies. Mol. Ecol. 23:3044-3063.

Bouckaert R., Heled J., Kühnert D., Vaughan T., Wu C.H., Xie D., Suchard M.A., Rambaut

A., Drummond A.J. 2014. BEAST 2: a software platform for Bayesian evolutionary

analysis. PLoS Comput. Biol. 104:e1003537.

Braun P.J., Esteves Pereira E. 1995. Nieuwe combinaties en namen voor cactussen uit

Brazilië, Bolivia en Paraguay. Succulenta (Netherlands). 74: 134-135.

Carstens B.C., Pelletier T.A., Reid N.M., Satler J.D. 2013. How to fail at species

delimitation? Mol. Ecol. 22:4369–4383.

Calvente A., Moraes E.M., Lavor P., Bonatelli I.A.S., Nacaguma P., Versieux L.M., Taylor

N.P., Zappi D.C. 2017. Phylogenetic analyses of *Pilosocereus* (Cactaceae) inferred from

plastid and nuclear sequences. Bot. J. Linn. Soc. 183:25–38.

Camargo A., Morando M., Avila L.J., Sites Jr J.W. 2012. Species delimitation with ABC and

other coalescent-based methods: a test of accuracy with simulations and an empirical

example with lizards of the *Liolaemus darwinii* complex (Squamata: Liolaemidae).

Evolution 66:2834-2849.

Campillo, L.C., Barley, A.J., Thomson, R.C. 2020. Model-based species delimitation: are

coalescent species reproductively isolated? Syst. Biol. 69(4):708-721.

Copetti D., Búrquez A., Bustamante E., Charboneau J.L.M., Childs K.L., Eguiarte L.E., Lee

S., Liu T.L., McMahong M.M., Whiteman N.K., Winga R.A, Wojciechowskii M.F.,

Sanderson M.J. 2017. Extensive gene tree discordance and hemiplasy shaped the

genomes of North American columnar cacti. Proc. Natl. Acad. Sci. U S A. 114:12003-

12008.

Csilléry K., François O., Blum M.G.B. 2012. abc: an R package for approximate Bayesian

computation (ABC). Methods Ecol. Evol. 3:475–479.

de Queiroz K. 2007. Species concepts and species delimitation. Syst. Biol. 56: 879–886.

Denham S.S., Brignonea N.F., Johnson L.A., Pozner R.E. 2019. Using integrative taxonomy

and multispecies coalescent models for phylogeny reconstruction and species

delimitation within the "Nastanthus–Gamocarpha" clade (Calyceraceae). Mol.

Phylogenet. Evol. 130:211-226.

Derkarabetian S., Castillo S., Koo P.K., Ovchinnikov S., Hedin M. 2019. A demonstration of

unsupervised machine learning in species delimitation. Mol. Phylogenet. Evol.

139:106562.

Domingos F.M., Bosque R.J., Cassimiro J., Colli G.R., Rodrigues M.T., Santos M.G.,

Beheregaray L.B. 2014. Out of the deep: Cryptic speciation in a Neotropical gecko

(Squamata, Phyllodactylidae) revealed by species delimitation methods. Mol.

Phylogenet. Evol. 80:113–124.

Drummond A.J., Rambaut A. 2007. BEAST: Bayesian evolutionary analysis by sampling

trees. BMC Evol. Biol. 7:214.

Ence D.D., Carstens B.C. 2011. SpedeSTEM: a rapid and accurate method for species

delimitation. Mol. Ecol. Resour. 11:473–480.

Fearnhead P., Prangle D. 2012. Constructing summary statistics for approximate Bayesian

computation: Semi-automatic ABC. J. R. Statist. Soc. B. 74:419–474.

da Fonseca E.M., Colli G.R., Werneck F.P., Carstens B.C. 2020. Phylogeographic model

selection using convolutional neural networks. bioRxiv.2020.09.11.291856.

Flagel L., Brandvain Y., Schrider D.R. 2019. The unreasonable effectiveness of

convolutional neural networks in population genetic inference. Mol. Biol. Evol. 36:220-

238

Guerrero P.C., Majure L.C., Cornejo-Romero A., Hernández-Hernández T. 2018.

Phylogenetic relationships and evolutionary trends in the Cactus family. J. Hered. 110:4–

21.

Guillot G., Mortier F., Estoup A. 2005. GENELAND: a computer package for landscape

genetics. Mol. Ecol. Resour. 5:712–715.

Harvey M.G., Smith B.T., Glenn T.C., Faircloth B.C., Brumfield R.T. 2016. Sequence

capture versus restriction site associated DNA sequencing for shallow systematics. Syst.

Biol. 65:910–924.

Hernández-Hernández T., Hernández H.M., Arturo De-Nova J., Puente R., Eguiarte L.E.,

Magallón S. 2011. Phylogenetic relationships and evolution of growth form in Cactaceae

(Caryophyllales, Eudicotyledoneae). Am. J. Bot. 98:44–61.

Hunt D., Taylor N.P., Charles G. 2006. The new Cactus Lexicon: Atlas & Text. dh Books,

Milborne Port.

Jackson N.D., Carstens B.C., Morales A.E., O'Meara B.C. 2017. Species delimitation with

gene flow. Syst. Biol. 66:799–812.

Katoh K., Standley D. M. 2013. MAFFT multiple sequence alignment software version 7:

improvements in performance and usability. Mol. Biol. Evol. 30:772–780.

Khan G., Godoy M.O., Franco F.F., Perez M.F., Taylor N.P., Zappi D.Z., Machado M.C.,

Moraes E.M. 2018. Extreme population subdivision or cryptic speciation in the cactus

Pilosocereus jauruensis? A taxonomic challenge posed by a naturally fragmented

system. Syst. Biodivers. 16:188–199.

Kousathanas A., Leuenberger C., Helfer J., Quinodoz M., Foll M., Wegmann D. 2016.

Likelihood-Free Inference in High-Dimensional Models. Genetics 203:893–904.

Lanfear R., Calcott B., Ho S.Y., Guindon S. 2012. PartitionFinder: combined selection of

partitioning schemes and substitution models for phylogenetic analyses. Mol. Biol. Evol.

29(6): 1695-1701.

Lavor P., Calvente A., Versieux L. M., Sanmartin I. 2019. Bayesian spatio-temporal

reconstruction reveals rapid diversification and Pleistocene range expansion in the

widespread columnar cactus *Pilosocereus*. J. Biogeogr. 46:238–250.

Leaché, A.D., Fujita, M.K., Minin, V.N., Bouckaert, R.R. (2014). Species delimitation using
genome-wide SNP data. Syst. Biol. 63(4): 534-542.

Leaché A.D., Zhu T., Rannala B., Yang Z. 2018. The spectre of too many species. Syst. Biol.
68:168–181.

Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-
MEM. arXiv 1303.3997.

Li H., Handsaker B., Wysoker A., Fennell T., Ruan J., Homer N., Math G., Abecasis G.,
Durbin R. 2009. The sequence alignment/map format and SAMtools. Bioinformatics
25:2078–2079.

Luo A., Ling C., Ho S.Y.W., Zhu C. 2018. Comparison of methods for molecular species
delimitation across a range of speciation scenarios. Syst. Biol. 67:830–846.

Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing
reads. EMBnet. Journal 17:10–12.

McGuire G., Wright F. 2000. TOPAL 2.0: improved detection of mosaic sequences within
multiple alignments. Bioinformatics 16:130–134.

McKenna A., Hanna M., Banks E., Sivachenko A., Cibulskis K., Kernytsky A., Garimella K.,
Altshuler D., Gabriel S., Daly M., DePristo M.A. 2010. The Genome Analysis Toolkit: a
MapReduce framework for analyzing next-generation DNA sequencing data. Genome
Res. 20:1297–1303.

Milne I., Lindner D., Bayer M., Husmeier D., McGuire G., Marshall D.F., Wright F. 2008.
TOPALi v2: a rich graphical interface for evolutionary analyses of multiple alignments
on HPC clusters and multi-core desktops. Bioinformatics 25:126–127.

Morales A.E., Jackson N.D., Dewey T.A., O'Meara B.C., Carstens B.C. 2017 Speciation with
gene flow in North American Myotis bats. Syst. Biol. 66:440–452.

594 Nguyen A.,Yosinski J., Clune J. 2015. Deep neural networks are easily fooled: High

595  confidence predictions for unrecognizable images. In IEEE Conference on Computer

596  Vision and Pattern Recognition (CVPR), p. 427–436.

597 Oliveira, E.A.D., Perez, M.F., Bertollo, L.A.C., Gestich, C.C., Ráb, P., Ezaz, T., Souza,

598  F.H.S., Viana, P., Feldberg, E., Oliveira, E.H.C.D., Cioffi, M.B., 2020. Historical

599  demography and climate driven distributional changes in a widespread Neotropical

600  freshwater species with high economic importance. Ecography 43: 1291-1304.

601 Pei J., Chong C., Xin L., Bin L., Yufeng W. 2018. CLADES: A classification-based machine

602  learning method for species delimitation from population genetic data. Mol. Ecol.

603  Resour. 18:1144–1156.

604 Perez M.F., Bonatelli I.A.S., Moraes E.M, Carstens B.C. 2016a. Model-based analysis

605  supports interglacial refugia over long-dispersal events in the diversification of two

606  South American cactus species. Heredity 116:550–557.

607 Perez M.F., Carstens B.C., Rodrigues G.L., Moraes E.M., 2016b. Anonymous nuclear

608  markers reveal taxonomic incongruence and long-term disjunction in a cactus species

609  complex with continental-island distribution in South America. Mol. Phylogenetics Evol.

610  95: 11-19.

611 Perez M.F., Carstens B.C., Rodrigues G.L., Moraes E.M. 2016c. Anonymous nuclear

612  markers data supporting species tree phylogeny and divergence time estimates in a cactus

613  species complex in South America. Data Brief 6:456–460.

615 Petkova D., Novembre J., Stephens, M. 2016. Visualizing spatial population structure with

616  estimated effective migration surfaces. Nat. Genet. 48:94–100.

Pinheiro F., Dantas-Queiroz M.V., Palma-Silva C. 2018. Plant Species Complexes as Models

to Understand Speciation and Evolution: A Review of South American Studies. Crit.

Rev. Plant Sci. 37:54–80.

Ponti M., Ribeiro L., Nazare T., Bui T., Collomosse J. "Everything you wanted to know

about Deep Learning for Computer Vision but were afraid to ask". In: 30th SIBGRAPI

Conference on Graphics, Patterns and Images Tutorials (SIBGRAPI-T), 17–41. IEEE.

Prangle, D. 2015. Summary statistics in approximate Bayesian computation. arXiv preprint

arXiv:1512.05633.

Rambaut A., Suchard M.A., Xie D., Drummond A.J. 2014. Tracer v1.6. Available from: URL

http://beast.bio.ed.ac.uk/Tracer.

Rannala B., Yang Z. 2020. *Species Delimitation*. In: Scornavacca C., Delsuc F., Galtier, N.

Phylogenetics in the Genomic Era. No commercial publisher | Authors open access book,

5.5:1–18. ffhal-02536468

Robert C.P., Cornuet J.M., Marin J.M., Pillai N.S. 2011. Lack of confidence in approximate

Bayesian computation model choice. Proc. Natl Acad. Sci. U S A. 108:15112–15117.

Rosenberg N.A. 2004. DISTRUCT: a program for the graphical display of population

structure. Mol Ecol Notes. 4:137–138.

Salicini I., Ibáñez C., Juste J. 2011. Multilocus phylogeny and species delimitation within the

Natterer's bat species complex in the Western Palearctic. Mol. Phylogenet. Evol.

61:888–898.

Sanchez, T., Cury, J., Charpiat, G., Jay, F. 2020. Deep learning for population size history

inference: Design, comparison and combination with approximate Bayesian

computation. Mol. Ecol. Resour. 00: 1– 16.

Souza, F.H.S., Perez, M.F., Bertollo, L.A.C., Oliveira, E.A., Lavoué, S., Gestich, C.C., Ráb,

P., Ezaz, T., Liehr, T., Viana, P.F., Feldberg, E., Cioffi, M.B. 2019. Interspecific Genetic

Differences and Historical Demography in South American Arowanas

(Osteoglossiformes, Osteoglossidae, *Osteoglossum*). Genes 10: 693.

Sukumaran J., Knowles L.L. 2017. Multispecies coalescent delimits structure, not species.

Proc. Natl Acad. Sci. U S A. 114:1607–1612.

Schrider D.R., Kern A.D. 2018. Supervised Machine Learning for Population Genetics: A

New Paradigm. Trends Genet. 34:301–312.

Smith M.L., Carstens B.C. 2020. Disentangling the process of speciation using machine

learning. Evolution 74:216-229.

Solís-Lemus C., Knowles L.L., Ané C. 2015. Bayesian species delimitation combining

multiple genes and traits in a unified framework. Evolution 69:492–507.

Taylor N.P., Zappi D.C. 2004. Cacti of Eastern Brazil. Royal Botanic Gardens, Kew.

Untergrasser A., Cutcutache I., Koressaar T., Ye J., Faircloth B.C., Remm M., Rozen S.G.

2012. Primer3 – new capabilities and interfaces. Nucleic Acids Res. 40: e115.

Vasconcelos T.N.C., Alcantara S., Andrino C.O., Forest F., Reginato M., Simon M.F., Pirani

J.R. 2020. Fast diversification through a mosaic of evolutionary histories characterizes

the endemic flora of ancient Neotropical mountains. Proc. R. Soc. B 287:

20192933.Wegmann D., Leuenberger C., Excoffier L. 2009. Efficient approximate

Bayesian computation coupled with Markov chain Monte Carlo without likelihood.

Genetics 182:1207–1218.

Weyenberg G., Huggins P.M., Schardl C.L., Howe D.K., Yoshida R. 2014. kdetrees: non-

parametric estimation of phylogenetic tree distributions. Bioinformatics 30:2280–2287.

Yang Z., Rannala B. 2014. Unguided species delimitation using DNA sequence data from

multiple loci. Mol. Biol. Evol. 31:3125–3135.

Zappi D.C.1994. *Pilosocereus* (Cactaceae). The Genus in Brazil. Royal Botanic Gardens,

Kew.

Zappi D.C., Moro M.F., Meagher T.R., Lughadha E.N. 2017. Plant biodiversity drivers in

Brazilian campos rupestres: insights from phylogenetic structure. Front. Plant. Sci.

8:2141.

FIGURE CAPTIONS

FIGURE 1. A general protocol for species delimitation in *P. aurisetus*, comparing the adopted

CNN and ABC approaches. (1) Evaluate delimitation hypotheses based on taxonomic and

genetic information with the BPP (Yang and Rannala 2014); (2) simulate genetic data for

each delimitation hypothesis, with and without gene flow among lineages (+M models); (3)

transform the simulated data for the two approaches, CNN (convert to images, with black

pixels representing the major and white pixels representing the minor frequency alleles for

each segregating site) and ABC (extract SuSt averaged over loci); (4) train a neural network

(CNN) to recognize simulations generated from each model; (5) use independent simulations

to evaluate and compare the accuracy of CNN and ABC; (6) predict model probability, with

the trained network for the CNN approach or by approximating the posterior (retaining only

the 20% most similar simulations) for ABC.

FIGURE 2. (A) Geographic distribution of the sampled localities and elevational range in

eastern Brazil (codes according to Table 1). Symbols (triangles and circle) represent the

currently recognized *P. aurisetus* subspecies, and gray shades represent heterotypic

synonyms of *P. aurisetus* subsp. *aurisetus* (in parenthesis). The delimited species are shown

with lines colored according to the delimited entities in Fig. 3.

FIGURE 3. Species delimitation results are represented as bars at the tips of the topology

obtained in BPP. Values in parentheses above each bar present the number of species for each

hypothesis. The numbers at the nodes represent Beast PP/BPP 'specific'/and 'diffuse' prior

values. Symbols (triangles and circles) represent both currently recognized and synonymized

(in parenthesis) *P. aurisetus* taxa. A representative of each inferred species is shown, next to

the limits according to the 'splitter' hypothesis.

FIGURE 4. Cross-validation test to compare the performance of different model comparison

methods (CNN - filled lines and ABC - dashed lines) and numbers of simulations. Each point

represents the probability of choosing the "correct" model (i.e., the model from which the

data were simulated) over incorrect models. Each color represents a different scenario,

including (+M) or not migration between the OTUs.

APPENDICES

SUPPLEMENTARY FILE 1. Detailed simulation and model selection methods.

SUPPLEMENTARY TABLE S1. Primer sequence of each marker used in this study for

microfluidic amplification. The genomic region for each marker is coded as follows: nr =

nuclear, mt = mitochondrial, and cp = plastidial. Substitution models are provided for loci

used in the final analyses.

SUPPLEMENTARY FIGURE S1. Representatives of *P. aurisetus* subsp. *aurisetus* (A-D), and *P.*

aurisetus subsp. *aurilanatus* (E) sampled in this study. A. *P. aurisetus* subsp. *aurisetus*; B.

syn. *P. aurisetus* subsp. *werdermannianus*; C. syn. *P. aurisetus* subsp. *densilanatus*; D. syn.

P. aurisetus subsp. *supthutianus*; E. *P. aurisetus* subsp. *aurilanatus*. Photo credit: EMM.

SUPPLEMENTARY FIGURE S2. Posterior distribution of genealogical divergence index (*gdi*)

values generated by the BPP analysis. Color scheme according to branches from the tree

depicted in the upper right corner, with locality names following Fig. 2.

SUPPLEMENTARY FIGURE S3. Accuracy of the CNN along the training epochs with varying

numbers of simulations (500 in orange, 1,000 in yellow and 10,000 in green) for the training

(filled lines) and validation (dashed lines) sets.

SUPPLEMENTARY FIGURE S4. Confusion matrices showing the percentage of predicted classes

for each simulated scenario (model representation for *P. aurisetus* following Fig. 1) using test

data (independent simulations not evaluated in the training step). The results are shown for

both methods (CNN and ABC) in the three taxonomic systems analyzed (*P. aurisetus* and the

species pais *D. melanogaster-D. simulans* and *D. melanogaster-D. sechellia*.

SUPPLEMENTARY FIGURE S5. Principal coordinate analysis for simulated and empirical data in

P. aurisetus. Simulations from the 'splitter' model are shown following the same color

scheme in Fig. 1.

Table 1. Sampling localities for *P. aurisetus* and their assignment to operational taxonomic units according to three different species delimitation hypotheses ('splitter', 'lumper', 'BPP_noGDI' and 'BPP_GDI').

Taxon/Locality	Code	N	Longitude	Latitude	Delimitation hypothesis			
					'splitter'	'lumper'	'BPP_noGDI'	'BPP_GDI'
P. a. subsp. *aurisetus*								
Cocais-MG	COC	4	-43,450000	-19,866667	1	1	1	1
Itamarandiba-MG (a)	ITA	4	-42,935972	-18,001944	2	1	2	1
Pedra Menina-MG (a)	PMN	4	-43,044500	-18,136250	2	1	2	1
Olhos D'Água-MG (b)	ODA	4	-43,622361	-17,438833	3	1	3	1
Engenheiro Dolabela-MG (b)	EDB	4	-43,793667	-17,451417	3	1	3	1
Bocaiúva-MG (b)	BOV	4	-43,755833	-17,545833	3	1	3	1
Mendanha-MG (c)	MEN	4	-43,536583	-18,118917	4	1	2	1
Inhaí-MG (c)	INA	4	-43,605833	-17,146444	4	1	2	1
P. a. subsp. *aurilanatus*								
Joaquim Felício-MG	JFE	4	-44,178333	-17,757833	5	2	4	1

734 (a), (b) and (c) indicate the samples from the previously synonymized forms *P. a.* subsp. *densilanatus*, *P. a.*

735 subsp. *supthutianus* and *P. a.* subsp. *werdermannianus*, respectively.

Table 2. Model selection results for *P. aurisetus* and the two *Drosophila* species pairs using empirical data. CNN values are softmax probabilities for each scenario, while posterior probabilities are shown for ABC. The scenario with the highest probability for each method is shown in bold.

Scenario	CNN	ABC
P. aurisetus		
Splitter	**0.998**	**0.994**
Splitter+Migration	0.001	0.000
Lumper	0.000	0.000
Lumper+Migration	0.000	0.000
BPP+GDI	0.000	0.000
BPP_noGDI	0.001	0.006
BPP_noGDI+Migration	0.000	0.000
D. melanogaster_D.simulans		
One Species	0.008	0.018
Two Species	**0.992**	**0.982**
D. melanogaster_D. sechellia		
One Species	0.051	**0.894**
Two Species	**0.949**	0.106