Common anti-hemostatic medications increase the severity of uropathogenic *Escherichia coli* sepsis

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Abstract

Uropathogenic *Escherichia coli* (UPEC) causes urinary tract infections that can result in sepsis. Hemostasis is

protective in the pyelonephritis stage of ascending UPEC infection, the role of hemostasis but has not been

investigated during sepsis. Here we utilize a zebrafish-UPEC sepsis model to visualize infection-induced

coagulation and examine the effects of commonly prescribed anti-hemostatic medications on the infection

severity. Treatment of septic zebrafish with warfarin, aspirin, or ticagrelor reduced host survival, while

stabilization of clots with aminocaproic acid increased host survival. Our findings provide evidence that

commonly prescribed anti-hemostatic medications may worsen the outcome of severe UPEC infection.

Keywords

Hemostasis, coagulation, platelet, Extraintestinal *Escherichia coli*

Background

Uropathogenic *Escherichia coli* (UPEC) is the primary pathogen responsible for urinary tract infection (UTI), which is among the most common bacterial infections worldwide. UPEC most commonly initiates self-limiting cystitis inside the urinary tract, but can ascend the ureters to the kidneys as acute pyelonephritis and potentially progress into sepsis [1, 2].

Hemostasis plays an evolutionarily conserved roles in both immune control of infections and the pathogenesis of infectious complications from a wide range of pathogens [3, 4]. In the case of UPEC, treatment of rodent UPEC pyelonephritis models with the injectable anticoagulant heparin demonstrates an important role of coagulation in preventing the progression of pyelonephritis to sepsis via an α-hemolysin/renal epithelial cell CD147/Tissue Factor axis [5, 6].

Elderly patients undergoing long term catheterization are at high risk of UPEC infections. These patients are also likely to be taking anti-hemostatic medications for the management of chronic cardiovascular conditions which might compound their risk and severity of severe UPEC infections. Here, we have used a zebrafish embryo model of UPEC sepsis to study the interaction between commonly prescribed anti-hemostatic medications and the severity of UPEC sepsis [7].

Methods

Zebrafish husbandry

Adult zebrafish were housed at the Centenary Institute (Sydney Local Health District AWC Approval 2017-036). Zebrafish embryos were produced by natural spawning and raised at 28°C in E3 media.

Zebrafish lines

Wild type zebrafish are the AB background. Transgenic lines are: *Tg(fabp10a:fgb-EGFP)^{mi4001}* [8], *Tg(-6.0itga2b:eGFP)^{la2}* [9].

Infection of zebrafish embryos

Aliquots of single cell suspensions of midlog-phase *Escherichia coli* UTI89 carrying the pGI6 plasmid were

frozen at -80°C for use in infection experiments. Bacterial aliquots were thawed and diluted with phenol

red dye (0.5% w/v). 10-15 nL was injected into the caudal vein or trunk of M-222 (tricaine)-anaesthetized

30-48 hpf embryos resulting in a standard infectious dose of approximately 10,000 CFU. Embryos were

recovered into E3 and housed at 32°C.

Drug treatments

Embryos were treated with vehicle control (DMSO or water as appropriate), 10 µg/ml aspirin, 20 µg/ml

tirofiban, or 33 µM warfarin.

Imaging

Imaging was carried out on embryos anaesthetized in M-222 (Tricaine) mounted in 0.75% low melting point

agarose on a Deltavision Elite fluorescence microscope. Editing was carried out with Image J Software

Version 1.51j.

Statistics

Survival analyses were performed by Log-rank tests in GraphPad Prism. Fluorescent pixel count analyses

were performed by T-test in GraphPad Prism.

Results

Coagulation is targeted to bloodborne UPEC in zebrafish embryos

UPEC infection-induced clotting has observed in mouse explants [5, 6]. We infected *Tg(fabp10a:fgb-*

EGFP)^{mi4001} zebrafish embryos, where clots can be visualized by GFP fluorescence, with UTI89 UPEC carrying

the pGI6 plasmid, allowing visualization of red fluorescent UPEC, and performed timelapse microscopy. We

observed the progressive formation of both arterial and venous clots in infected embryos (Figure 1A).

Warfarin increases the severity of UPEC sepsis in zebrafish embryos

Inhibition of clotting with heparin has been reported to worsen UPEC infection in mice [5, 6]. We sought to

determine if commonly prescribed anti-coagulants had a similar effect in our zebrafish model using

warfarin, a vitamin K antagonist and the most commonly prescribed anti-coagulant.

We have previously demonstrated that warfarin reduced mycobacterial infection-induced coagulation in

zebrafish embryos [4], here we demonstrate that warfarin reduced clot formation at a sterile wound at the

embryo stage used in our UPEC infection experiments (Figure 1B). Treatment with warfarin decreased

embryo survival following systemic UPEC infection (Figure 1C). Conversely, clot stabilization with

aminocaproic acid increased embryo survival following systemic UPEC infection (Figure 1D).

Zebrafish thrombocytes interact with UPEC

Clotting and thrombosis are coordinated during infection-induced hemostasis [3, 4]. To determine if

zebrafish thrombocytes could play a role in UPEC sepsis, we infected *Tg(-6.0itga2b:eGFP)^{la2}* embryos, where

thrombocytes can be visualized by GFP expression, with UPEC- pGI6. We observed long lasting interactions

between zebrafish thrombocytes and clumps of UPEC but were unable to determine if the bacteria were

extracellular or intracellular (Figure 2A).

Aspirin and ticagrelor increase the severity of UPEC sepsis in zebrafish embryos

Having shown a negative effect of inhibiting clotting, we next investigated the effect of commonly

prescribed anti-platelet medications on systemic UPEC infection. Aspirin and ticagrelor reduced platelet

aggregation to a sterile wound (Figure 2B). Inhibition of platelets with either aspirin or ticagrelor reduced

survival of infected embryos (Figure 2C and 2D).

Discussion

We have found that common anti-hemostatic medications worsen the survival of UPEC-infected zebrafish.

Anti-hemostatic drug use is highly prevalent in the elderly as a preventative measure against heart attack

and stroke. Alongside this, the risk of urinary tract infections also increases from middle to old age and

there is elevated risk of recurrent urinary tract infections in aged care settings [10]. Our data illustrates a

negative interaction between preventative anti-hemostatic medication usage and the severity of UPEC

sepsis.

Our results add to a growing body of literature that hemostasis, and specifically clotting, is crucial to the

early containment of blood born UPEC [5, 6]. Clinical trials of anticoagulants to treat sepsis-induced

coagulopathy have delivered inconsistent results, however an emerging theme is that early coagulation is

host protective while late coagulation drives pathology [11]. Subsequently, administration of

anticoagulants at early stage can be detrimental to the clearance of pathogen by the native immune system

[12]. Additionally, there is little evidence on the beneficial effect of anticoagulants on the overall

population, however, is beneficial for critically ill subgroups such as sepsis-induced disseminated

intravascular coagulation [11, 13].

The negative effects of the antiplatelet drugs aspirin and ticagrelor demonstrate a host-protective role of

platelets during UPEC sepsis. Degranulation of activated platelets releases important inflammatory

mediators such as antimicrobial proteins, cytokines, and ADP/ATP which can directly kill pathogens and

activate cellular immunity [14]. Clinical trials with antiplatelets, especially aspirin, have demonstrated an

improvement in mortality of severe septic patients which, similar to anti-coagulant therapy, may reflect an

effect of timing [15].

Future studies should investigate the interaction between hemostasis and the natural course of UPEC

infection in both animal models and by retrospective clinical chart review. The potential for interaction

between aspirin and warfarin, the two most commonly prescribed anti-hemostatic medications, with

catherization-associated UPEC infections involves a significant proportion of aged care patients and may

represent an important cause of excess morbidity.

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Author contributions

E.H., and S.H.O designed the experiments. V.T., E.H., and S.H.O performed the experiments. V.T., E.H., and

S.H.O wrote the paper. E.H., W.J.B., and S.H.O. supervised the project.

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Figure Legends

Figure 1: Clotting is host-protective against systemic UPEC infection.

(A) Intravital microscopy of green fibrin deposition around UPEC pGI6 (red) in a *Tg(fabp10a:fgb-EGFP)*[mi4001]

embryo. Filled arrowhead indicates site of arterial clot formation, hollow arrowhead indicates site of

venous clot formation. Scale bar indicates 10 µm. (B) Fibrin-GFP fluorescent area 2 hours post wounding.

(C) Survival of UPEC-infected embryos treated with warfarin. n>=18 per group, representative of two

biological replicates. (D) Survival of UPEC-infected embryos treated with warfarin or ACA. n>=20 per group,

representative of two biological replicates.

Figure 2: Platelet activation is host-protective against systemic UPEC infection.

(A) Intravital microscopy of green thrombocytes interacting with UPEC pGI6 (red) in a *Tg(-6.0itga2b:eGFP)*[la2]

embryo. Filled arrowhead indicates first thrombocyte that interacts with clump of UPEC for 30 minutes,

hollow arrowhead indicates a second thrombocyte that interacts with the same clump of UPEC for 21

minutes. Unindicated green cells are thrombocytes in circulation. t0 is approximately 1 hour post infection.

Scale bar indicates 10 µm. (B) CD41 (thrombocyte) fluorescent area at 2 hours post wounding. (C) Survival

of UPEC-infected embryos treated with aspirin. n>=25 per group, representative of two biological

replicates. (D) Survival of UPEC-infected embryos treated with ticagrelor. n=25 per group, representative of

two biological replicates.

A *Tg(fabp10a:fgb-EGFP)^{mi4001}* UPEC-pGI6

10 h 11 h 12 h 13 h 14 h 15 h

16 h 17 h 18 h 19 h 20 h 21 h

B

Fibrin-GFP fluorescent area within 100 μm of wound (normalized)

P = 0.001

DMSO   Warfarin

C

Percent survival

P = 0.015

DMSO
Warfarin

Days post infection

D

Percent survival

P = 0.044

P = 0.0029

ACA
DMSO
Warfarin

Days post infection

A *Tg(-6.0itga2b:eGFP)^{la2}* UPEC-pGI6

0 | + 3 min | + 6 min | + 9 min | + 12 min | + 15 min

+ 18 min | + 21 min | + 24 min | + 27 min | + 30 min | + 33 min

B

P = 0.028

P = 0.084

CD41 fluorescent area within 100 μm of wound (px)

DMSO | Aspirin | Ticagrelor

C

Percent survival

P = 0.0002

— DMSO
--- Aspirin

Days post infection

D

Percent survival

P < 0.0001

— DMSO
--- Ticagrelor

Days post infection