Universal markers support a long inter-domain branch between Archaea and Bacteria

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Abstract

The tree of life is generally estimated from a core set of 16-56 genes coding for proteins predominantly involved in translation and other conserved informational and cellular processes. These markers represent only a fraction of the genes that were likely present in the last universal common ancestor (LUCA), but are useful for deep phylogenetic reconstructions because their mode of inheritance appears to be mainly vertical, which satisfies the assumptions of gene concatenation and supertree methods. Previous phylogenetic analyses of these genes recovered a long branch between Archaea and Bacteria. By contrast, a recent study made use of a greatly expanded set of 381 marker genes and recovered a much shorter branch length between Archaea and Bacteria, comparable to some divergences within the domains. These analyses suggest that the apparent deep split between Archaea and Bacteria may be the result of accelerated evolution of ribosomal genes. Here we re-evaluate the evolutionary history of the expanded marker gene set and show that substitutional saturation, inter-domain gene transfer, hidden paralogy, and poor model fit contribute to the inference of an artificially shortened inter-domain branch. Our results do not exclude a moderately faster rate of ribosomal gene evolution during the divergence of Archaea and Bacteria, but indicate that vertically-evolving marker genes across all functional categories support a major genetic divergence between the two primary domains of life.

Introduction

Much remains unknown about the earliest period of cellular evolution and the deepest divergences in the tree of life. Phylogenies encompassing both Archaea and Bacteria have been inferred from a "universal core" set of 16-56 genes encoding proteins involved in translation and other aspects of the genetic information processing machinery[1–10]. These genes are thought to evolve vertically and so more closely track an underlying tree of life compared to other genes[3,4,8,11]. In these analyses, the branch separating Archaea from Bacteria (hereafter, the AB branch) is often the longest internal branch in the tree[2,6,12–15]. In molecular phylogenetics, branch lengths are usually measured in expected numbers of substitutions per site, with a long branch corresponding to a greater degree of genetic change. Long branches can therefore result from high evolutionary rates, long periods of time, or a combination of the two. Molecular clock models can, in principle, disentangle the contributions of these effects, but only very few fossil calibrations[16] are currently available that are old enough to calibrate early divergences[17–20], and as a result the ages and evolutionary rates of the deepest branches of the tree remain highly uncertain.

Recently, Zhu et al.[21] inferred a phylogeny from 381 genes distributed across Archaea and Bacteria using the supertree method ASTRAL[22]. In addition to a large increase in the number of genes compared to other universal marker sets, the functional profile of these markers comprises not only proteins involved in information processing but also proteins affiliated with most other functional COG categories, including metabolic processes (Table S1). Subsequently, the genetic distance (branch length) between the domains[21] was estimated from a concatenation of the same marker genes, resulting in a much shorter AB branch length than observed with the core universal markers[2,6]. These analyses motivated the hypothesis[21] that the apparent deep divergence of Archaea and Bacteria might be the result of an accelerated evolutionary rate of genes encoding ribosomal proteins along the AB branch as compared to other genes. Interestingly, the same observation was made previously using a smaller set of 38 non-ribosomal marker proteins[5], although the difference in AB branch length between ribosomal and non-ribosomal markers in that analysis was reported to be substantially lower (roughly two-fold, compared to roughly ten-fold for the 381 protein set of Zhu et al.)[5,21].

A higher evolutionary rate of ribosomal genes might result from the accumulation of compensatory substitutions at the interaction surfaces among the protein subunits of the ribosome[5,23], or as a compensatory response to the addition or removal of ribosomal subunits early in evolution[5]. Alternatively, differences in the inferred AB branch length might result from varying rates or patterns of evolution between the traditional core genes[2,24] and the expanded set[21]. Substitutional saturation (multiple substitutions at the same site[25]) and across-site compositional heterogeneity can both impact the inference of tree topologies and branch lengths[26–30]. These difficulties are particularly significant for ancient divergences[31]. Failure to model site-specific amino acid preferences has previously been shown to lead to under-estimation of the AB branch length due to a failure to detect convergent changes [2,32], although the published analysis of the 381 marker set did not find evidence of a substantial impact of these features on the tree as a whole[21]. Those analyses also identified phylogenetic incongruence among the 381 markers, but did not determine the underlying cause[21].

This recent work[21] raises two important issues regarding the inference of the universal tree: first, that estimates of the genetic distance between Archaea and Bacteria from classic "core genes" may be unrepresentative of ancient genomes as a whole, and second, that there may be many more suitable genes to investigate early evolutionary history than generally recognized, providing an opportunity to improve the precision and accuracy of deep phylogenies. Here, we address these points by examining the evolutionary history of the 381 marker set (hereafter, the expanded marker gene set), and by evaluating the impact of orthology assignment, horizontal gene transfer (HGT), substitutional saturation and substitution model fit on inferences of the genetic divergence between Archaea and Bacteria based on gene concatenations.

Results and Discussion

Genes from the expanded marker set are not widely distributed in Archaea

The 381-gene set was originally derived from a larger set of 400 genes used to estimate the phylogenetic placement of new bacterial lineages as part of the PhyloPhlAn method[33]. Perhaps reflecting the focus on bacterial phylogeny in the original application, the phylogenetic distribution of the 381 marker genes in the expanded set varies substantially (Table S1), with many being poorly represented in Archaea. Indeed 25% of the published gene trees (https://biocore.github.io/wol/[21]) contain less than 0.5% archaeal homologues, with 21 (5%) and 69 (18%) of these trees containing no or less than 10 archaeal homologues, respectively. For the remaining 75% of the gene trees, archaeal homologs comprise 0.5%-13.4% of the dataset. While there are many more sequenced bacteria than archaea, 63% of the gene trees possessed genes from less than half of the 669 archaeal genomes included in the analysis , whereas only 22% of the gene trees possessed fewer than half of the total number of 9906 sampled bacterial genomes. These distributions suggest that many of these genes are not broadly present in both domains, and that some might be specific to Bacteria.

Conflicting evolutionary histories of individual marker genes and the inferred species tree

In the focal analysis of the 381 gene set, the tree topology was inferred using the supertree method ASTRAL[22], with branch lengths inferred on this fixed tree from a marker gene concatenation[21]. The topology inferred from this expanded marker set[21] is similar to published trees[6,34] and recovers Archaea and Bacteria as reciprocally monophyletic domains, albeit with a shorter AB branch than in earlier analyses. However, the individual gene trees[21] disagree regarding domain monophyly: Archaea and Bacteria are recovered as reciprocally monophyletic groups in only 24 of the 381 published[21] (Table S1) maximum likelihood (ML) gene trees for the expanded marker set.

Since single gene trees often fail to strongly resolve ancient relationships, we used approximately-unbiased (AU) tests[35] to evaluate whether the failure to recover domain monophyly in the published ML trees is statistically supported. For computational tractability,

we performed these analyses on a 1000-species subsample of the full dataset that was compiled in the original study[21]. For 79 of the 381 genes, we could not perform the test because the gene was not found on any of the 74 archaeal genomes present in the 1000-species subsample. For the remaining 302 genes, domain monophyly was rejected ($p < 0.05$) for 232 out of 302 (76.8%) genes. As a comparison, we performed the same test on several smaller marker sets used previously to infer a tree of life[2,5,36]; none of the markers in those sets rejected reciprocal domain monophyly (AU > 0.05 for all genes, Figure 1(a)). In what follows, we refer to four published marker gene sets: the Expanded set (381 genes[21]), the Core set (49 genes[2], encoding ribosomal proteins and other conserved information-processing functions), the Non-ribosomal set (38 genes, broadly distributed and explicitly selected to avoid genes encoding ribosomal proteins[5]), and the Bacterial set (29 genes used in a recent analysis of bacterial phylogeny[36]).

To investigate why 232 of the marker genes rejected the reciprocal monophyly of Archaea and Bacteria, we returned to the full 10,575-species dataset and annotated each sequence in each marker gene family by assigning proteins to KOs, Pfams, and Interpro domains, among others (Table S1, see Methods for details). We labelled the tips on each published marker gene phylogeny using the corresponding KO and PFAM annotations and descriptions (See Methods) and manually inspected the tree topologies for reciprocal domain monophyly of Archaea and Bacteria and the presence of paralogues (Table S1). This revealed that the major cause of domain polyphyly observed in gene trees was inter-domain gene transfer (in 357 out of 381 gene trees (93.7%)) and mixing of sequences from distinct paralogous families (in 246 out of 381 gene trees (64.6%)). For instance, marker genes encoding ABC-type transporters (p0131, p0151, p0159, p0174, p0181, p0287, p0306, po0364), tRNA synthetases (i.e. p0000, p0011, p0020, p0091, p0094, p0202), aminotransferases and dehydratases ((i.e. p0073/4-aminobutyrate aminotransferase; p0093/3-isopropylmalate dehydratase) often comprised a mixture of paralogues. For example, the phylogenetic tree comprising spermidine/putrescine import ATP-binding protein PotA (p0131) contains several paralogues families, such as different sugar or ion ATP-binding proteins. Further, dTDP-glucose 4,6-dehydratase (p0134) is representative of a gene tree that includes several paralogous families with different substrate specificities that include dehydrorhamnose and glucuronate.

Together, these analyses indicate that the evolutionary histories of the individual markers of the expanded set differ from each other and from the species tree. Zhu et al. acknowledged[21] the varying levels of congruence between the marker phylogenies and the species tree, but did not investigate the underlying causes. Our analyses establish the basis for these disagreements in terms of gene transfers and the mixing of orthologues and paralogues within and between domains. In principle, concatenation is based on the assumption that all of the genes in the supermatrix evolve on the same underlying tree; genes with different gene tree topologies should not be concatenated because the topological differences among sites are not modelled, and so the impact on inferred branch lengths is difficult to predict. In practice, it is often difficult to be certain that all of the markers in a concatenate share the same gene tree topology, and the analysis proceeds on the hypothesis that a small proportion of discordant genes are not expected to seriously impact the inferred tree. However, the concatenated tree inferred from the expanded marker set differs from previous trees in that the genetic distance between Bacteria and Archaea is greatly reduced, such that the AB branch length appears comparable to the distance between bacterial phyla[21]. An accurate estimate of the AB branch length is important because it has a major bearing on unanswered questions regarding the

root of the universal tree[31] and the deepest divisions among cellular life. We therefore evaluated the impact of the conflicting gene histories within the expanded marker set on inferred AB branch length.

The inferred branch length between Archaea and Bacteria is artifactually shortened by inter-domain gene transfer and hidden paralogy

To investigate the impact of gene transfers and mixed paralogy on the AB branch length inferred by gene concatenations[21], we compared branch lengths estimated from markers that rejected (AU < 0.05) or did not reject (AU > 0.05) the reciprocal monophyly of Bacteria and Archaea in the 381 marker set (Figure 1(a)). To estimate AB branch lengths for genes in which the domains were not monophyletic in the ML tree, we first performed a constrained ML search to find the best gene tree that was consistent with domain monophyly for each family under the LG+G4+F model in IQ-TREE 2[37]. While it may seem strained to estimate the length of a branch that does not appear in the ML tree, we reasoned that this approach would provide insight into the contribution of these genes to the AB branch length in the concatenation, in which they conflict with the overall topology. AB branch lengths were significantly (P = 2.159×10^{-12}, Wilcoxon rank sum test) shorter for markers that rejected domain monophyly (Figure 1(a); <0.05: mean AB branch length in expected substitutions/site 0.0130, >0.05: mean AB branch length 0.559). This result suggests that inter-domain gene transfers reduce the AB branch length when included in a concatenation. This behaviour might result from marker gene transfers reducing the number of fixed differences between the domains, so that the AB branch length in a tree in which Archaea and Bacteria are constrained to be reciprocally monophyletic will tend towards 0 as the number of transfers increases. Consistent with this hypothesis, we observed that ΔLL, the difference in log likelihood between the ML gene tree and the constrained ML tree consistent with domain monophyly (a simple proxy for marker gene verticality) correlates negatively with AB branch length (Figure 1(b)). Furthermore, AB branch length decreased as increasing numbers of low-verticality markers were added to the concatenate (Figure 1(c)). Taken together, these results indicate that the inclusion of genes that do not support the reciprocal monophyly of Archaea and Bacteria in the universal concatenate reduces the estimated AB branch length by homogenizing the genetic diversity of the two domains.

The inferred Archaea-Bacteria branch is artifactually shortened by unaccounted-for substitutional saturation

The longer AB branch length observed in concatenations of traditional core marker sets[2,6,24] compared to the expanded marker set[21] has been interpreted as evidence for ribosomal gene-specific accelerated evolution between domains[5,21], because ribosomal proteins and proteins that physically interact with the ribosome make up a large proportion (47.1%) of the core set[2]. An alternative hypothesis is that faster rates of evolution in the genes of the expanded set following the divergence of Archaea and Bacteria has resulted in substitutional saturation, overwriting some of the early changes and shortening the AB branch. To distinguish between these hypotheses, we compared AB branch lengths estimated from fast- and slow-evolving sites in the expanded concatenate, for the 1000 taxa subset[21]. We estimated site-specific rates of evolution using IQ-TREE 2[37], and constructed two new concatenates comprising the 25% fastest- and 25% slowest-evolving sites. As expected, the total tree length inferred from the slowest sites was shorter (105 substitutions/site compared to 1216 substitutions/site in the

25% fastest site concatenation). However, the relative length of the AB branch (that is, AB branch length as a proportion of total tree length) was 8.8-fold higher in the concatenation inferred from the 25% slowest sites compared to the fastest sites (Figure 2(a)). Considering only the top 5% of the expanded marker set in terms of **Δ**LL score, the relative difference increases to 11.1-fold. This analysis suggests that substitutional saturation has overwritten many of the changes that originally occurred during the divergence of Archaea and Bacteria at fast-evolving sites in the expanded gene set. Total tree lengths estimated for the expanded marker genes did not significantly differ (P = 0.5716) between genes that did or did not reject domain monophyly.

An earlier study[5] also recovered a longer AB branch from ribosomal compared to non-ribosomal genes, and none of the markers used in that study rejected domain monophyly (Figure 1, "Non-ribosomal"). While the non-ribosomal markers from that study[5] have similar total tree lengths to those in the core gene set (Figure 3(a)), they have ~2.4-fold shorter AB branches, Figure 3(b)). These results are consistent with the hypothesis that ribosomal proteins may have longer AB branches than other marker genes.

Failure to model across-site amino acid preferences reduces estimates of the AB branch length

Amino acid preferences vary across the sites of a sequence alignment, due to variation in the underlying functional constraints[26,29,30]. The consequence is that, at many alignment sites, only a subset of the twenty possible amino acids are tolerated by selection. Standard substitution models, such as LG+G+F, are site-homogeneous, and approximate the composition of all sites using the average composition across the entire alignment. Such models underestimate the rate of evolution at highly constrained sites because they cannot account for the high number of multiple substitutions that occur at such sites. The effect is that site-homogeneous models underestimate branch lengths when fit to site-heterogeneous data. The AB branch has previously been shown to be particularly susceptible to this effect[2]. Zhu et al.[21] investigated the impact of site heterogeneity on tree topology and branch lengths using the posterior mean site frequency (PMSF) model[38], and found that inferences under site-homogeneous and site-heterogeneous models were similar over the dataset as a whole[21]. To evaluate the effect of modelling site-heterogeneity on the AB branch length in particular, we inferred phylogenies from the concatenate constructed from the top 5% of genes scored by ΔLL under a series of models that account for site-heterogeneity using increasingly rich mixtures of site-specific composition profiles (C10-C60)[30]. Models that account for site heterogeneity support a branch that is 1.6-2.1-fold longer than using a site-homogeneous model; the best-fitting model among those we tested was LG+C60+G+F according to the BIC score, and this model inferred an AB branch length of 2.4; that is, twice as long as that inferred under the site-homogeneous model. Therefore, and consistent with previous work[2,32], substitution model fit has a significant effect on the inferred genetic proximity of Archaea and Bacteria (Figure 4). We also find that trimming poorly-aligned sites greatly increases the inferred length of the AB branch, likely because trimmed sites tend to be fast-evolving.

The age of the last universal common ancestor (LUCA) inferred from the expanded gene set is driven by variation in the AB branch length

Zhu et al.[21] argued that the expanded marker set is useful for deep phylogeny because estimates of the age of LUCA obtained by fitting molecular clocks to their dataset are in agreement with the geological record: a root (LUCA) age of 3.6-4.2 Gyr was inferred from the entire 381-gene dataset, consistent with the earliest fossil evidence for life, whereas estimates from ribosomal markers alone supported a root age of 7 Gya. This age might be considered unrealistic because it is much older than the age of the Earth (with the moon-forming impact occurring ~4.51Gya[42,43]). In the original analyses, the age of LUCA was estimated using a maximum likelihood approach, as well as a Bayesian molecular clock with a strict clock (assuming a constant evolutionary rate) or a relaxed clock with a single calibration. A strict clock model does not permit changes in evolutionary rate through time or across branches, and so a longer AB branch will lead to an older inferred LUCA age. Likewise, a relaxed clock model with a single calibration may fail to differentiate molecular distances and geological time. Given that the short AB branch in the expanded gene set results, in part, from phylogenetic incongruence among markers, we evaluated the age of LUCA inferred from the subset of the expanded gene set least affected by these issues. To do so, we analysed the top 5% of gene families according to their **Δ**LL score (including only 1 ribosomal protein) under the same clock model parameters as the original dataset. This analysis resulted in a significantly more ancient age estimate for LUCA (5.5-6.5 Ga), suggesting that, under these conditions, the inferred age for LUCA is driven by variation in the AB branch length, and is not in itself a reliable indicator of marker quality. In principle, more reliable estimates of LUCA's age might be obtained by using more calibrations. However, unambiguous calibrations remain elusive, particularly for the root and other deep branches of the tree. Despite advances in molecular clock methodology, such calibrations represent the only way to reliably capture the relationship between genetic distance and divergence time.

Conclusion

The apparent genetic proximity of Archaea and Bacteria inferred from a concatenation of the expanded gene set of Zhu et al. [21] results from substitutional saturation and inter-domain HGT. Saturation obscures evidence of substitutions along the AB branch, while undetected HGT acts to artifactually homogenize the genetic diversity of the domains, and leads to a reduction in the inferred AB branch when branch lengths are estimated on a fixed topology. Treatments of the data that improve estimation of the AB branch length - the use of better-fitting substitution models, or use of the markers least affected by inter-domain HGT - results in the inference of a ~15-fold longer AB branch that is similar to estimates from previously published datasets (Figure 6). These results emphasise the importance of the fit of the substitution model to the data, and the need to evaluate whether the data meet the assumptions of the methods used - in particular, the assumption that all genes evolve on the same underlying tree topology, which underlies analyses of gene concatenations. The violation of that assumption in branch length estimates from concatenated alignments of the expanded marker genes resulted in the recovery of an artifactually short AB branch.

A distinct question is whether the long AB branch inferred from congruent marker gene sets might be affected by accelerated ribosomal gene evolution. Comparison of AB branch length for sets of congruent "core" and non-ribosomal marker sets (Figure 3) are consistent with a higher relative rate of evolution along the AB branch for ribosomal genes, although the effect is small by comparison with the impact of phylogenetic congruence, site trimming and substitution model fit (Figure 6). In sum, all of our analyses support the conclusion that the split separating Archaea from Bacteria is by far the longest internal branch on the tree of the primary domains of life.

In the future, it may be useful to further evaluate the suitability of non-ribosomal markers for concatenated gene phylogenies. Furthermore, methods that explicitly model HGTs[44–46], while also calculating branch lengths in a manner that accounts for differences among the underlying gene trees, and that incorporate alternative means of dating information, such as gene transfers[17,47–49], may help to better constrain the divergence times of the deepest cellular lineages.

Methods

Data

We downloaded the individual alignments from [21] (https://github.com/biocore/wol/tree/master/data/), along with the genome metadata and the individual newick files. We checked each published tree for domain monophyly, and also performed approximately unbiased (AU)[35] tests to assess support for domain monophyly on the underlying sequence alignments using IQ-TREE 2[37]. The phylogenetic analyses were carried out using the 'reduced' subset of 1000 taxa outlined by the authors[21], for computational tractability. These markers were also trimmed according to the protocol in the original paper[21], i.e sites with >90% gaps were removed, followed by removal of sequences with >66% gaps.

We also downloaded the Williams et al.[2] ("core"), Petitjean et al.[5] ("non-ribosomal") and Coleman et al.[36] ("bacterial") datasets from their original publications.

Annotations

Proteins used for phylogenetic analyses by Zhu *et al*.[21], were annotated to investigate the selection of sequences comprising each of the marker gene families. To this end, we downloaded the protein sequences provided by the authors from the following repository: https://github.com/biocore/wol/tree/master/data/alignments/genes. To obtain reliable annotations, we analysed all sequences per gene family using several published databases, including the arCOGs (version from 2014)[50], KOs from the KEGG Automatic Annotation Server (KAAS; downloaded April 2019)[51], the Pfam database (Release 31.0)[52], the TIGRFAM database (Release 15.0)[53], the Carbohydrate-Active enZymes (CAZy) database (downloaded from dbCAN2 in September 2019)[54], the MEROPs database (Release 12.0)[55,56], the hydrogenase database (HydDB; downloaded in November 2018)[57], the NCBI- non-redundant (nr) database (downloaded in November 2018), and the NCBI COGs database (version from 2020). Additionally, all proteins were scanned for protein domains using InterProScan (v5.29-68.0; settings: --iprlookup --goterms)[58].

362 Individual database searches were conducted as follows: arCOGs were assigned using PSI-
363 BLAST v2.7.1+ (settings: -evalue 1e-4 -show_gis -outfmt 6 -max_target_seqs 1000 -dbsize
364 100000000 -comp_based_stats F -seg no)[59]. KOs (settings: -E 1e-5), PFAMs (settings: -E 1e-
365 10), TIGRFAMs (settings: -E 1e-20) and CAZymes (settings: -E 1e-20) were identified in all
366 archaeal genomes using hmmsearch v3.1b2[60]. The MEROPsand HydDB databases were
367 searched using BLASTp v2.7.1 (settings: -outfmt 6, -evalue 1e-20). Protein sequences were
368 searched against the NCBI_nr database using DIAMOND v0.9.22.123 (settings: –more-
369 sensitive –e-value 1e-5 –seq 100 –no-self-hits –taxonmap prot.accession2taxid.gz)[61]. For all
370 database searches the best hit for each protein was selected based on the highest e-value
371 and bitscore and all results are summarized in the Data Supplement,
372 Annotation_Tables/0_Annotation_tables_full/All_Zhu_marker_annotations_16-12-
373 2020.tsv.zip. For InterProScan we report multiple hits corresponding to the individual domains
374 of a protein using a custom script (parse_IPRdomains_vs2_GO_2.py).
375
376 Assigned sequence annotations were summarized and all distinct KOs and Pfams were
377 collected and counted for each marker gene. KOs and Pfams with their corresponding
378 descriptions were mapped to the marker gene file downloaded from the repository:
379 https://github.com/biocore/wol/blob/master/data/markers/metadata.xlsx and used in
380 summarization of the 381 marker gene protein trees (Table S1).
381
382 For manual inspection of single marker gene trees, KO and Pfam annotations were mapped
383 to the tips of the published marker protein trees, downloaded from the repository:
384 https://github.com/biocore/wol/tree/master/data/trees/genes. Briefly, the Genome ID, Pfam,
385 Pfam description, KO, KO description, and NCBI Taxonomy string were collected from each
386 marker gene annotation table and were used to generate mapping files unique to each marker
387 gene phylogeny, which links the Genome ID to the annotation information
388 (GenomeID|Domain|Pfam|Pfam Description|KO|KO Description). An in-house Perl script
389 replace_tree_names.pl
390 (https://github.com/ndombrowski/Phylogeny_tutorial/tree/main/Input_files/5_required_Scripts
391) was used to append the summarized protein annotations to the corresponding tips in each
392 marker gene tree. Annotated marker gene phylogenies were manually inspected using the
393 following criteria including: 1) retention of reciprocal domain monophyly (Archaea and
394 Bacteria) and 2) for the presence or absence of potential paralogous families. Paralogous
395 groups and misannotated families present in the gene trees were highlighted and violations of
396 search criteria were recorded in Table S1.

397 ***Phylogenetic analyses***

398 *Constraint analysis*

399 We performed a maximum likelihood free topology search using IQ-TREE 2[37] under the
400 LG+G+F model, with 10,000 bootstrap replicates. We also performed a constrained analysis
401 with the same model, in order to find the maximum likelihood tree in which Archaea and
402 Bacteria were reciprocally monophyletic. We then compared both trees using the
403 approximately unbiased (AU)[35] test in IQ-TREE 2[37] with 10,000 RELL[35] bootstrap replicates.
404 To evaluate the relationship between marker gene verticality and AB branch length, we
405 calculated the difference in log-likelihood between the constrained and unconstrained trees in
406 order to rank the genes from the expanded marker set, made concatenates comprised of the

top 20-100 (intervals of 5) of these marker genes, and inferred the tree length under LG+C10+G+F with 1000 bootstrap replicates.

Site and gene evolutionary rates

We inferred rates using the --rate option in IQ-TREE 2[37] for both the 381 marker concatenation from Zhu[21] and the top 5% of marker genes based on the results of difference in log-likelihood between the constrained tree and free-tree search in the constraint analysis (above). We built concatenates for the 25% slowest and 25% fastest sites, and inferred branch lengths from each of these concatenates using the tree inferred from the complete dataset as a fixed topology.

Individual markers

We inferred rates and trees (1000 bootstrap replicates) for each individual marker from Petitjean *et al.*[5], Williams *et al.*[2], Zhu *et al.*[21] under the LG+G+F and LG+C20+G+F models using IQ-TREE 2[37]. Mean rates per sequence were then calculated using a python script (see Data Supplement).

Model complexity

Model complexity tests were undertaken using the top 5% concatenate described above, with the alignment being trimmed with BMGE 1.12[41] with default settings (BLOSUM62, entropy 0.5) for all of the analyses except the 'untrimmed' LG+G+F run, other models on the trimmed alignment were LG+G+F, LG+R+F and LG+C10,20,30,40,50,60+G+F, with 1000 bootstrap replicates. Model fitting was done using ModelFinder[62] in IQ-TREE 2[37].

Molecular clock analyses

Molecular clock analyses were devised to test the effect of genetic distance on the inferred age of LUCA. Following the approach of Zhu et al[21], we subsampled the alignment to 100 species. Five alternative alignments were analysed, representing conserved sites across the entire alignment, randomly selected sites across the entire alignment, only ribosomal marker genes, the top 5% of marker genes according to ΔLL and the top 5% of marker genes further trimmed under default settings in BMGE 1.12[41]. Divergence time analyses were performed in MCMCTree[63] under a strict clock model. We used the normal approximation approach, with branch lengths estimated in codeml under the LG+G4 model. In each case, a fixed tree topology was used alongside a single calibration on the Cyanobacteria-Melainabacteria split. The calibration was modelled as a uniform prior distribution between 2.5 and 2.6 Ga, with a 2.5% probability that either bound could be exceeded. For each alignment, four independent MCMC chains were run for 2,000,000 generations to achieve convergence.

Plotting

Statistical analyses were performed using R 3.6.3[64], and data were plotted with ggplot2[65].

Figure legends

Figure 1: Expanded set genes in which Archaea and Bacteria are not monophyletic support a shorter AB branch. (A) Expanded set genes that reject domain monophyly (AU P < 0.05) support significantly shorter AB branch lengths when constrained to follow a domain monophyletic tree ($P = 2.159 \times 10^{-12}$, Wilcoxon rank-sum test). None of the marker genes from several other published analyses reject domain monophyly (AU p > 0.05 for all genes tested). (B) Marker gene verticality (ΔLL, see below) for the expanded gene set normalized by alignment length correlates negatively with the length of the AB branch between Archaea and Bacteria (R^2=0.03998, P = 0.0004731). (C) Concatenations of 20-100 markers of the expanded set markers ranked by marker gene verticality (ΔLL) show the same trend, with a reduction in AB branch length as markers with a greater ΔLL are added to the concatenate. ΔLL is the difference between the log likelihood of the ML gene family tree under a free topology search and the log likelihood of the best tree constrained to obey domain monophyly. The trendline is estimated using a LOESS regression.

Figure 2: Vertically-evolving genes and slow-evolving sites support a longer relative AB branch length. We estimated site-specific evolutionary rates for all marker genes in the expanded dataset (A-B), as well as for the 20 genes with the smallest ΔLL (top 5%) in that dataset (C-D). Concatenations based on the 25% slowest sites (A,C) and on the top 5% vertical genes (C,D) support a longer AB branch. This suggests that the inference of a short AB branch is impacted by both substitutional saturation and unmodelled inter-domain transfer of marker genes. Phylogenies were inferred under the LG+G4+F model in IQ-TREE 2[37]. Branch lengths are the expected number of substitutions per site, as indicated by the scale bars. Alignment lengths in amino acids: **A**: 36797, **B**: 67274, **C**: 2736, **D**: 3884.

Figure 3. Evidence for a faster rate of inter-domain ribosomal protein evolution. (A) Tree lengths (total number of substitutions/site for each gene family) for the core and non-ribosomal gene sets under the LG+G+F and LG+C20+G+F models. Modelling site heterogeneity increases tree lengths (the number of inferred substitutions) due to improved modelling of the site-specific features of the evolutionary process. Tree lengths are not significantly different between the core and non-ribosomal marker sets P = 0.4821/0.1651 (for LG+G+F and LG+C60+G+F respectively) (B) Non-ribosomal genes have moderately shorter AB branch lengths than core genes, consistent with a moderately faster rate of ribosomal gene evolution on the inter-domain branch. The difference is significant under both LG+G+F ($P = 8.78 * 10^{-9}$), and the better-fitting LG+C20+G+F model ($P = 2.237 * 10^{-7}$).

481 **Figure 4. The effect of modelling site heterogeneity on AB branch length.** Increasing the
482 number of protein mixture profiles, as well as trimming is associated with a change in AB
483 branch length on the expanded marker set[21]. All analyses used LG exchangeabilities, four rate
484 categories (Gamma-distributed or freely estimated), and included a general composition
485 vector containing the empirical amino acid frequencies (+F). Modelling of site heterogeneity
486 with the C10-C60 models increases the inferred AB branch length ~2-fold. Trimming poorly-
487 aligned sites slightly increases the AB branch estimation whereas relaxing the gamma rate
488 categories slightly decreases estimation of AB branch length. LG (LG substitution matrix), G
489 (four gamma rate categories), F (empirical site frequencies estimated from the data), C10-60
490 (number of protein mixture profiles used[30]) R (four free rate categories which relax the
491 assumption of a gamma distribution for rates[39,40], BMGE (trimming using Block Mapping and
492 Gathering with Entropy[41]). The trendline is estimated using a LOESS regression.
493
494 **Figure 5. Divergence time estimation of the Archaea-Bacteria split.** Violins represent
495 posterior age estimates from Bayesian molecular clock analyses of 1) Conserved sites as
496 estimated previously[21]; 2) Random sites[21] 3) Ribosomal genes[21] 4) The top 5% of marker gene
497 families according to their **Δ**LL score (including only 1 ribosomal protein) and 5) The same top
498 5% of marker genes trimmed using BMGE[41] to remove highly variable sites. In each case, a
499 strict molecular clock was applied, with the age of the Cyanobacteria-Melainabacteria split
500 constrained between 2.5 and 2.6 Ga.
501
502 **Figure 6. The impact of marker gene choice, phylogenetic congruence, alignment**
503 **trimming, and substitution model fit on estimates of the Archaea-Bacteria branch**
504 **length.** Analysis using a site-homogeneous model (LG+G+F) on the complete 381-gene
505 expanded set results in an AB branch substantially shorter than previous estimates. Removing
506 the genes most seriously affected by inter-domain gene transfer, trimming poorly-aligned sites
507 using BMGE[41], and using the best-fitting site-heterogeneous model available (LG+C60+G+F)
508 substantially increase the estimated AB length, such that it is comparable with published
509 estimates from non-ribosomal and core gene sets. Branch lengths measured in expected
510 number of substitutions/site.
511

512 ## Data and code availability

513 All of the data, including sequence alignments, trees, annotation files, and scripts associated
514 with this manuscript have been deposited in the FigShare repository at DOI:
515 10.6084/m9.figshare.13395470.
516

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681

Figure 1

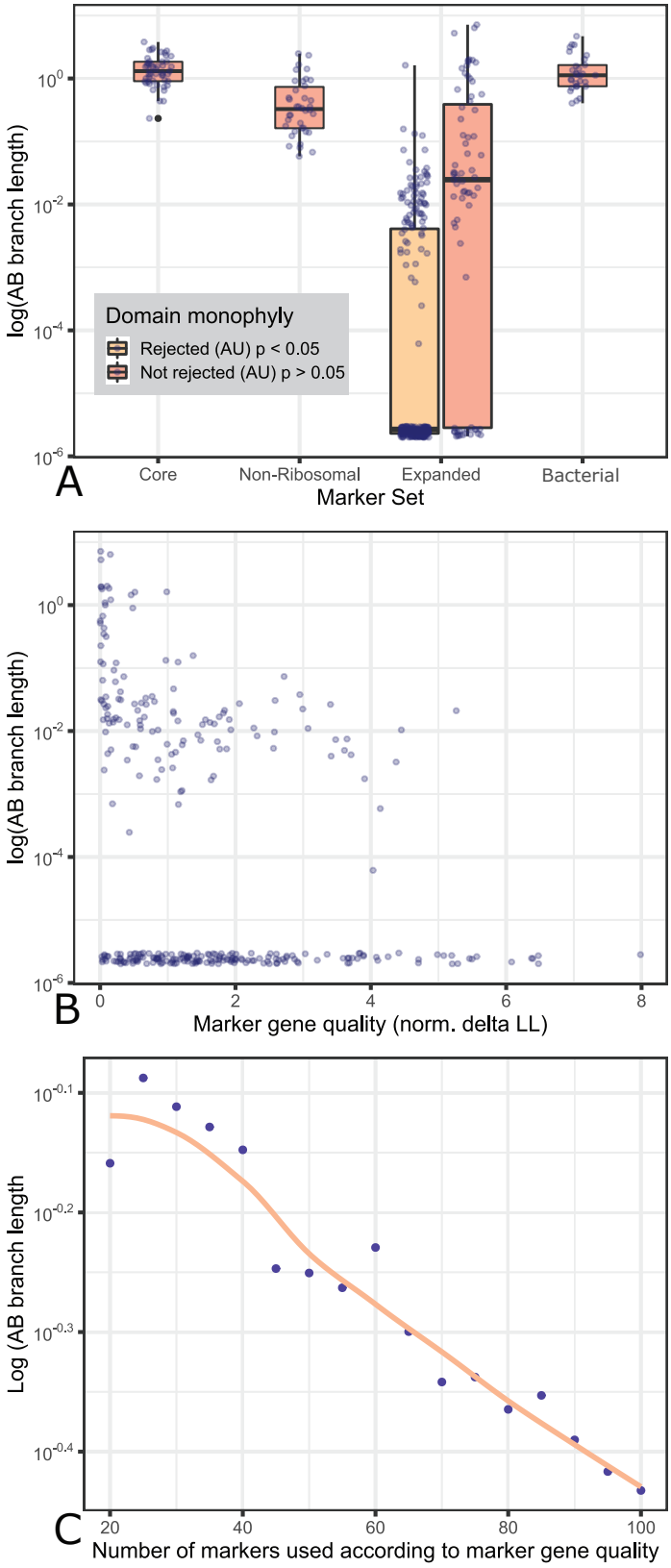

log(AB branch length)
Domain monophyly
Rejected (AU) p < 0.05
Not rejected (AU) p > 0.05
Core
Non-Ribosomal
Expanded
Bacterial
Marker Set
A
log(AB branch length)
Marker gene quality (norm. delta LL)
B
Log (AB branch length
Number of markers used according to marker gene quality
C

Figure 2

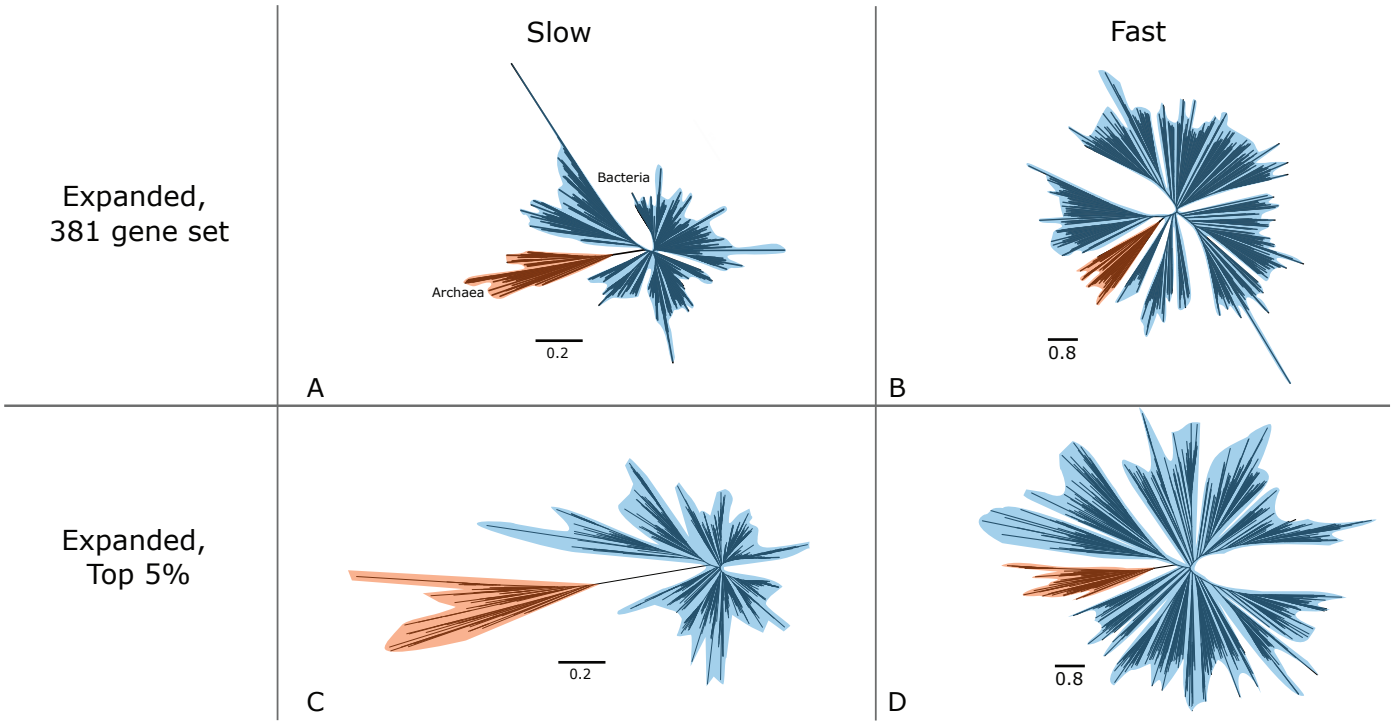

Slow
Fast
Expanded,
381 gene set
Expanded,
Top 5%
Bacteria
Archaea
0.2
0.8
0.2
0.8
A
B
C
D

Figure 3

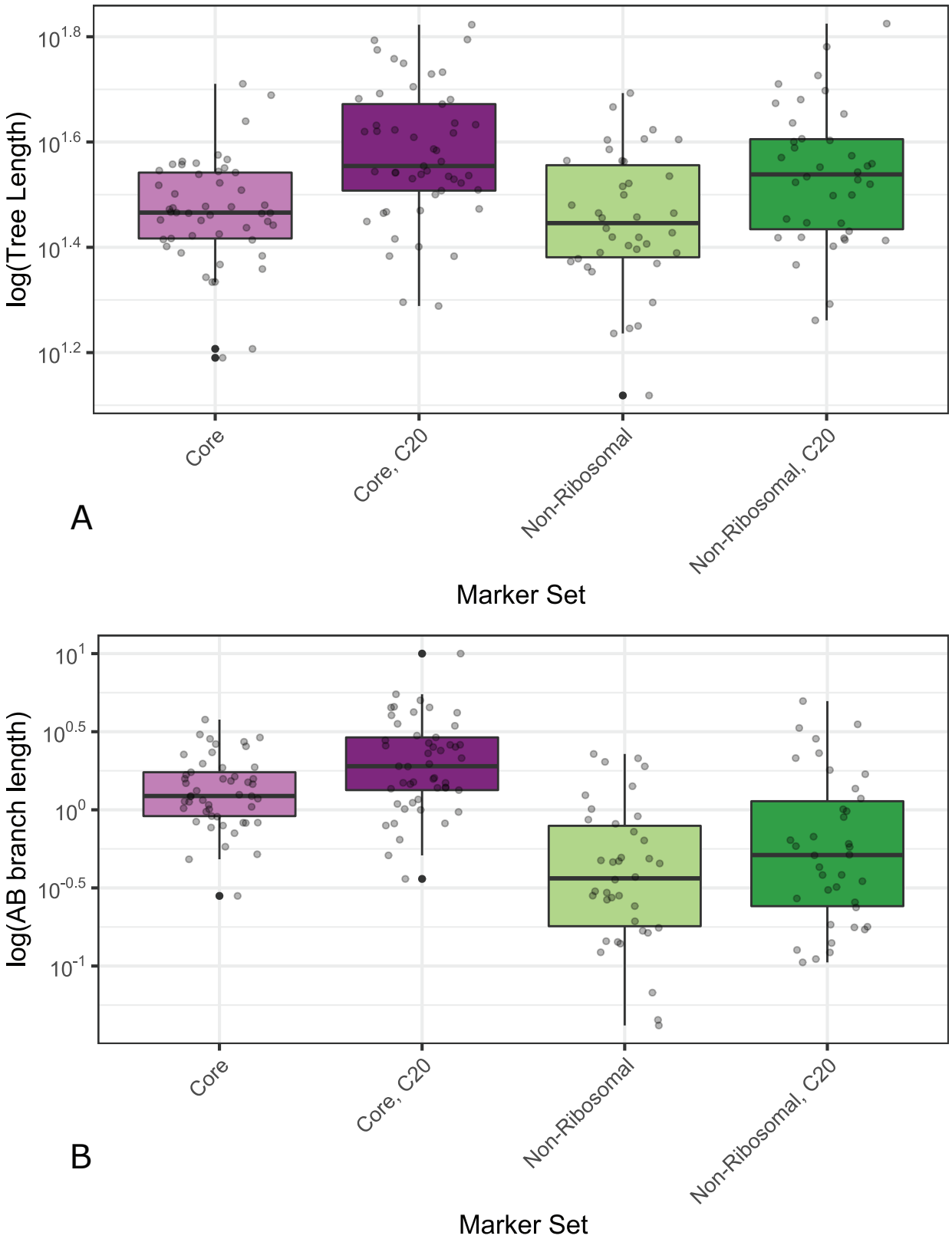

log(Tree Length)
$10^{1.8}$
$10^{1.6}$
$10^{1.4}$
$10^{1.2}$
Core
Core, C20
Non-Ribosomal
Non-Ribosomal, C20
A
Marker Set
log(AB branch length)
10^{1}
$10^{0.5}$
10^{0}
$10^{-0.5}$
10^{-1}
Core
Core, C20
Non-Ribosomal
Non-Ribosomal, C20
B
Marker Set

Figure 4

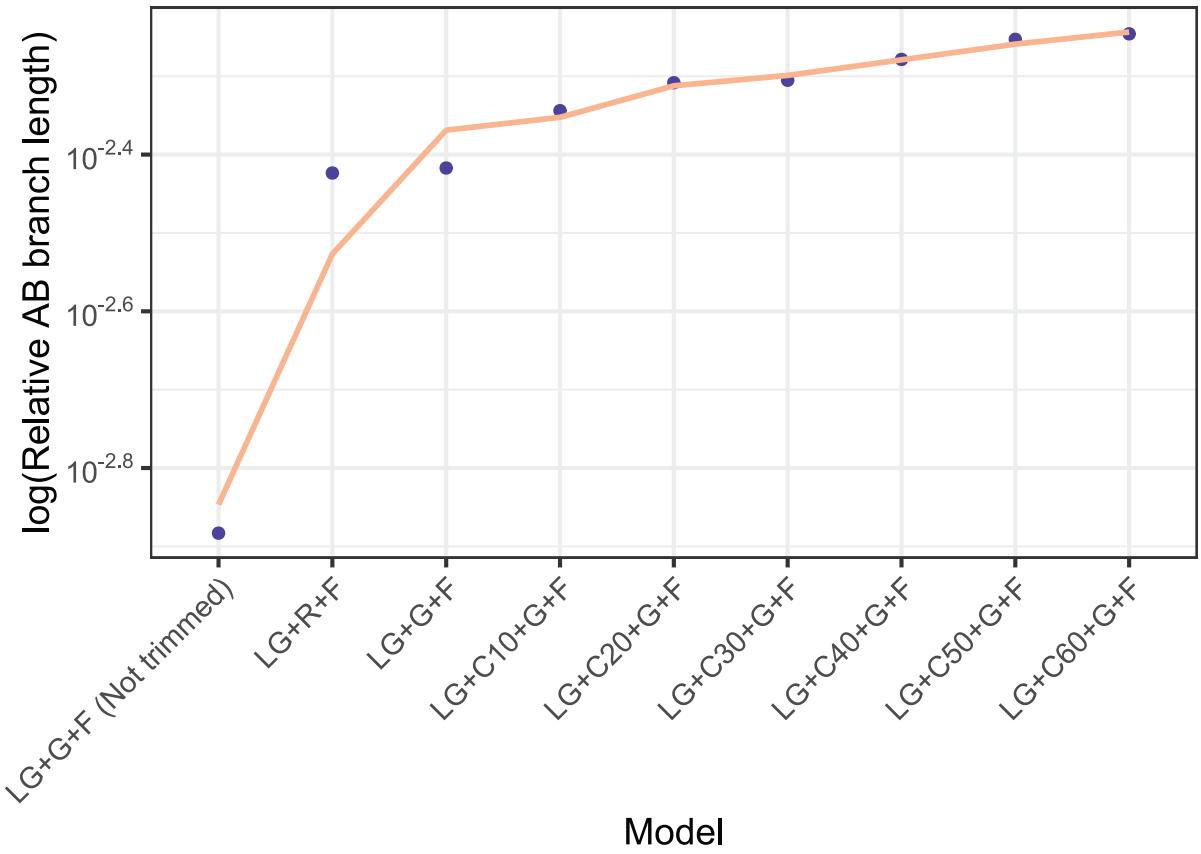

log(Relative AB branch length)
$10^{-2.4}$
$10^{-2.6}$
$10^{-2.8}$
LG+G+F (Not trimmed)
LG+R+F
LG+G+F
LG+C10+G+F
LG+C20+G+F
LG+C30+G+F
LG+C40+G+F
LG+C50+G+F
LG+C60+G+F
Model

Figure 5

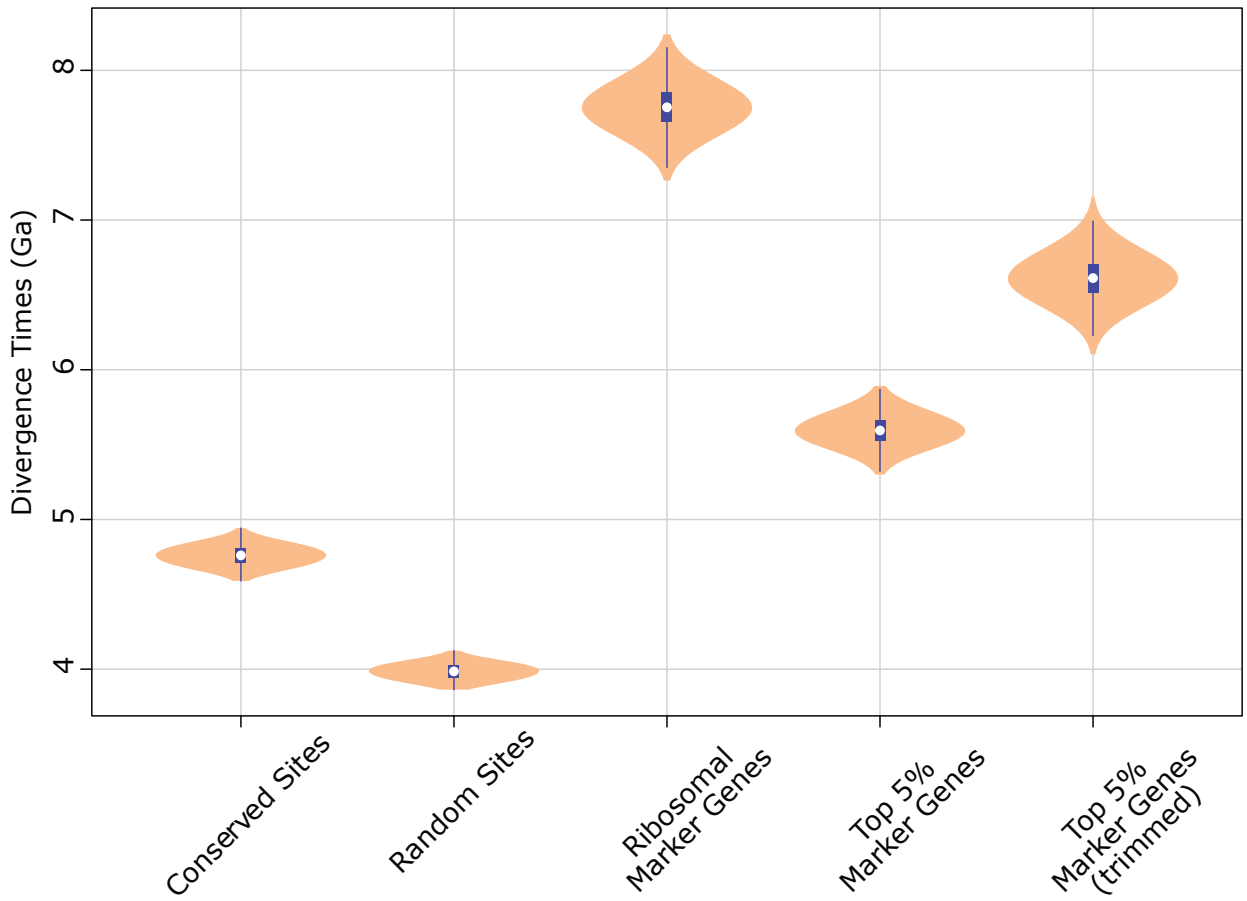
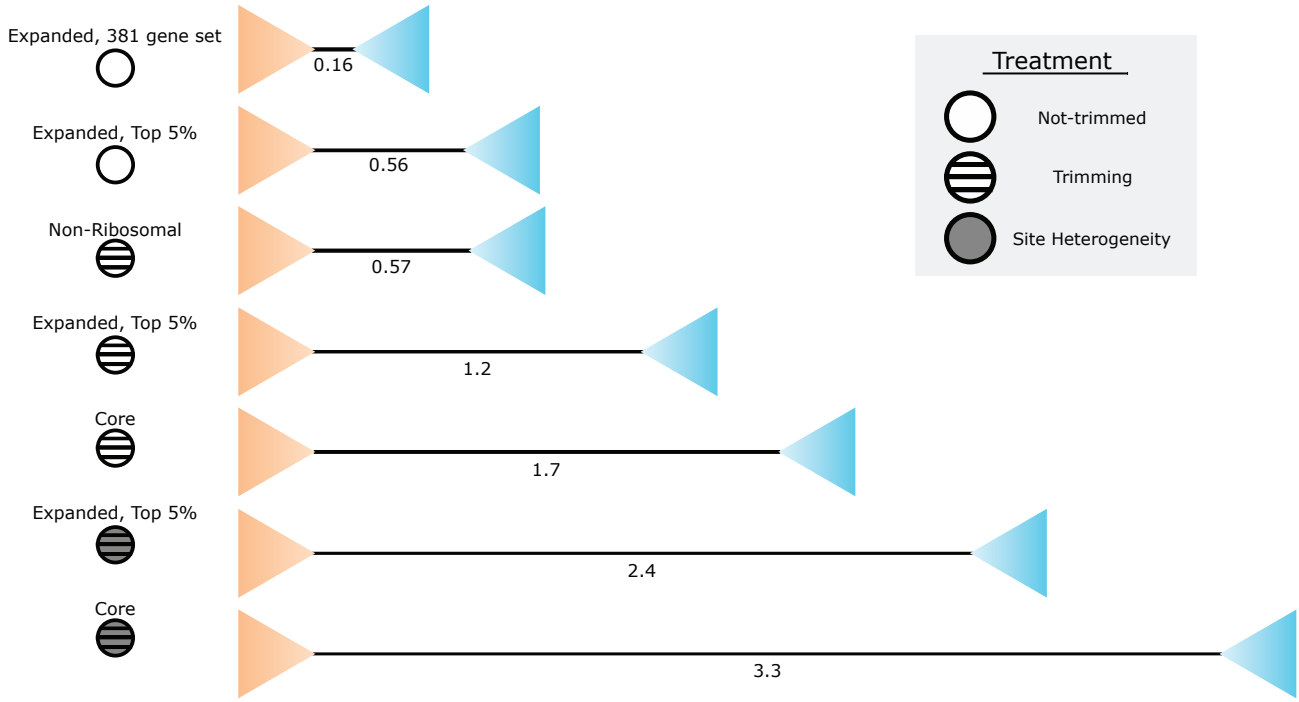

Divergence Times (Ga)
4
5
6
7
8
Conserved Sites
Random Sites
Ribosomal Marker Genes
Top 5% Marker Genes
Top 5% Marker Genes (trimmed)

Figure 6


Expanded, 381 gene set
0.16
Expanded, Top 5%
0.56
Non-Ribosomal
0.57
Expanded, Top 5%
1.2
Core
1.7
Expanded, Top 5%
2.4
Core
3.3
Treatment
Not-trimmed
Trimming
Site Heterogeneity