Improvements in the Sequencing and Assembly of Plant Genomes

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Abstract

Background Advances in DNA sequencing have reduced the difficulty of sequencing and assembling plant genomes. A range of methods for long read sequencing and assembly have been recently compared and we now extend the earlier study and report a comparison with more recent methods.

Results Updated Oxford Nanopore Technology software supported improved assemblies. The use of more accurate sequences produced by repeated sequencing of the same molecule (PacBio HiFi) resulted in much less fragmented assembly of sequencing reads. The use of more data to give increased genome coverage resulted in longer contigs (higher N50) but reduced the total length of the assemblies and improved genome completeness (BUSCO). The original model species, *Macadamia jansenii*, a basal eudicot, was also compared with the 3 other *Macadamia* species and with avocado (*Persea americana*), a magnoliid, and jojoba (*Simmondsia chinensis*) a core eudicot. In these phylogenetically diverse angiosperms, increasing sequence data volumes also caused a highly linear increase in contig size, decreased assembly length and further improved already high completeness. Differences in genome size and sequence complexity apparently influenced the success of assembly from these different species.

Conclusions Advances in long read sequencing technology have continued to significantly improve the results of sequencing and assembly of plant genomes. However, results were consistently improved by greater genome coverage (using an increased number of reads) with the amount needed to achieve a particular level of assembly being species dependant.

Keywords: long read sequencing, assembly, plant, Pacific Biosciences, HiFi reads, Oxford Nanopore Technology

Background

Recent advances in DNA sequencing technology have facilitated the sequencing and assembly of plant genomes with a rapid growth in reports of high quality chromosome level assemblies [1]. A basal eudicot, *Macadamia jansenii*, was used to compare the range of long read sequencing and assembly technologies available in 2019[2]. The Pac Bio (Sequel), Oxford Nanopore Technology (PromethION) and BGI (single-tube Long Fragment Read) platforms were applied to the analysis of the same sample. Assembly tools were evaluated for these data sets and the contribution of short reads to improving assemblies assessed [2]. Technology improvements had delivered ongoing increases in the length and quality of sequence reads delivered by these platforms. Since the original study, significant further advances have been made with the use of repeated sequencing of the same molecule to greatly increase sequence accuracy for long reads. This technology allows the generation of long reads (10-25kb) with greater than 99.5% accuracy [3]. Comparison of long read technologies demonstrates the pros and cons of different platforms in relation to contiguity, accuracy of sequence and data analysis time [4]. We now update the earlier study to demonstrate the impact of these improvements on genome assemblies. Factors such as the volume of data (bp) used in the assembly were explored for the *Macadamia* genome, related species and other diverse species with similar sized genomes.

Long read versus HiFi assemblies

Comparison of assemblies based upon long reads [5] and circular consensus sequence (CCS) reads from HiFi [3], showed the greater accuracy of the CCS reads resulted in greatly improved assemblies for the *Macadamia jansenii* genome (Table 1).

Table 1 Improvement in long read sequencing (Pac Bio) for *Macadamia jansenii* when using higher accuracy sequencing.

	Long reads*[2]	HIFi
Total data	65.2Gb	28Gb
Contig N50	1.57Mb	4.49Mb
Assembly length	758Mb	738Mb
Number of contigs	762	284
BUSCO	97%	98%

*phased Falcon assembly

The assembly with the high quality HiFi reads was less fragmented with slightly reduced total genome length and improved completeness (BUSCO). The use of around 20Gb of high quality (HiFi) data gave N50 values of 4 Mb and resulted in assemblies with fewer than 300 contigs required to cover the genome. This represents a significant advance over the assemblies possible when this sample was used previously to compare different long read platforms and assembly tools, many of which required long computing times to assemble contigs [2]. The high quality IPA assembly had a run time of 20 h with 120 Gb peak memory on the FlashLite computer cluster. This analysis requirement compares favourably with the

results for a large number of earlier assembly tools reported for the same sample[2], but provides a much higher quality assembly.

Results for other *Macadamia* species *Macadami jansenii* is an endangered species and is one of four species in the *Macadamia* genus. Sequences of all four species were obtained using the same techniques and all gave similar high quality outcomes when assembled (Table 2).

Table 2 Comparison of assemblies of *Macadamia* species

	M. jansenii	*M. integrifolia*	*M. tetraphylla*	*M. ternifolia*
Contig N50	4.5Mb	5.3Mb	10.0Mb	6.4Mb
Longest Contig	16.6Mb	26.4Mb	32.1Mb	21.2Mb
Assembly Length	738Mb	742Mb	707Mb	716Mb
Number of contigs	284	249	153	211
BUSCO	98%	98%	97%	98%

Results for other diverse plant species Methods for sequencing plant genomes need to be applied to genomes with a wide range of sizes and complexities. Macadamia is a basal eudicot. Other diverse flowering plant genomes were sequenced to determine how widely applicable the results of this study would be in plant genome sequencing. Jojoba (*Simmondsia chinensis*), a core eudicot from the Caryopyllales, and avocado, a magnoliid, were compared. The three diverse genomes were all similar in size (700-1000Mb). Many important plant genomes are in or near this size range [6]. *M. jansenii* is an endangered

species with relatively low heterozygosity, avocado has much greater heterozygosity [7] and jojoba is reported to be a tetraploid[8]. Heterozygosity and polyploidy both complicate assembly [9,10]. The quality of the assemblies was more contiguous with less data in each of these cases when HiFi reads were used instead of the earlier continuous long reads (Table 3).

Table 3 Long read versus HiFi sequencing of other diverse species

	Long reads		HiFi	
	Jojoba	Avocado	Jojoba	Avocado
Total data	152Gb	159Gb	41.4Gb	44Gb
Contig N50	4.73Mb	6.7Mb	4.89Mb	4.3Mb
Assembly length	1260Mb	787Mb	780Mb	749Mb
Number of contigs	762	308	284	298
BUSCO	99%	99%	98%	98%

The length of the contigs assembled (Figure1) was directly related to the volume of sequence data used. Analysis of four related Macadamia species gave a similar linear relationship between data volume and contig N50 for input of between 10 and 40 times genome coverage. The size of the contigs assembled showed a similar dependence upon the amount of sequence data (genome coverage) across species with the slope of the relationship varying for different species (Figure 2). The macadamia genomes could be assembled with lower coverage. This may be a function of genome size with their smaller genomes requiring less coverage to

achieve a given N50. The larger genomes may contain a higher proportion of repetitive sequences that are difficult to assemble.

Assemblies based upon more data were slightly shorter in total length (Figure 3). This reduction was probably due to removal of duplicated end sequences as contigs were joined. The high quality of these assemblies was confirmed by BUSCO values of more than 95%. Genome completeness was high in all cases but increased slightly when more data was used in the assembly (Figure 4).

These results were confirmed when applying these methods to sequencing the other phylogenetically diverse plant genomes with slightly larger genomes with greater genome complexity. In each case N50 and completeness increased with data volume and genome size declined.

The length of sequence reads was also expected to influence the assembly. Examination of size distribution of the 6 species showed that the length of the sequences varied slightly within the expected range around 15kb for HiFi data. The minor differences in mean read length and numbers of longer reads did not explain the differences in the size of the contigs assembled (Supplementary Figure 1). This suggest that the different amounts of sequence data required to drive assembly to a particular level may be associated more with the complexity of the sequence. The close relationship between sequence volume and N50 for the four *Macadamia* species may reflect the similar sequence complexity of the species in this group. The jojoba and avocado required more sequence data to reach the same level of assembly. The slightly larger genome size of these two species may be enough to explain this difference due to the likely higher proportion of repetitive sequence in the somewhat larger genomes.

Oxford Nanopore Technologies updates

Oxford Nanopore Technologies (ONT) regularly releases updated basecalling software to convert the raw electrical signal into sequence data. We repeated the basecalling the ONT raw data of *M. jansenii* using different versions of the Guppy basecaller released in March 2019 (v2.3.7), April 2019 (v3.0.3) and June 2020 (v4.0.11). The assembly quality improved as shown by an increase in the assembly contiguity and in the number of complete BUSCOs before any polishing (Table 4). The assembly size decreased from 817 Mb to 798 Mb. Two versions of the Flye assembler were applied to the same basecalled sequence dataset, which resulted in a significant increase in genome contiguity and completeness as well as a reduced genome assembly size.

Table 4 ONT genome assembly statistics of *M. jansenii* using the Flye assembler, the pass reads and different Guppy basecallers versions

Basecaller	Guppy v2.3.7	Guppy v3.0.3		Guppy v4.0.11
Assembler	Flye v2.5	Flye v2.4.2	Flye v2.5	Flye v2.5
Number of reads	1,597,353	1,592,919		1,594,802
Contig N50 (Mb)	1.44	0.94	1.51	1.79
Assembly length (Mb)	817	845	811	798
Number of contigs	2,996	4,242	2,855	2,841
Number of contigs (>10 kb)	2,300	3,275	2,088	1,913
BUSCO complete (%)	66.8	51.4	75.1	79.1

Conclusions

These assemblies represent significant advances over the highly fragmented genomes previously reported for these species [11-14]. Advances in long read sequencing using different platforms provide improving options for plant genome sequencing and assembly [15]. A recent comparison of these methods applied to rice genome sequencing showed strengths and weaknesses of both with greater sequence accuracy in the Pac Bio assemblies and more contiguity in the ONT assemblies [4]. The resulting genome sequences can be evaluated for the best combination of sequence and assembly accuracy [16]. The results presented here show that contig size can be increased by adding more sequence reads to achieve a linear increase in N50. This extra data will result in slightly shorter total assembly lengths and improved completeness of the genomes. The improved methods when combined with higher level assembly tools[17] will support routine, rapid and efficient generation of highly accurate chromosome level genome sequences of plant species[18].

Methods

DNA extraction

All local, national and international guidelines and legislation was observed in obtaining samples for this study. *Macadamia jansenii* DNA was prepared as described earlier[19]. Three other Macadamia species (*M. tetraphylla*, *M. ternifolia* and *M. integrifolia*) and Jojoba (*Simmondsia chinensis*) were also extracted using this method with minor modifications where phenol was excluded from the extraction method[20]. Avocado (*Persea americana*) DNA was isolated by a modified CTAB (cetyl-trimethyl ammonium bromide) DNA extraction protocol [21,22]. Leaf tissue (0.2 g) was ground and added to 15 ml of 2% CTAB buffer, pH 8.0 followed by 15 min incubation at 65 °C. The supernatant after centrifugation

at 10 g for 15 min was treated with RNAse A (10ng/µl) and incubated at 37°C for 30 min. Chloroform: Isoamyl alcohol (24:1) washes were performed followed by precipitation with isopropanol and 70% ethanol washes. The DNA was resuspended in ultrapure DNAse and RNAse free water for sequencing.

DNA sequencing and assembly

Long read sequencing was as previously described[19]. HiFi gDNA libraries were prepared using sheared genomic DNA (~15-20 kb) was sequenced on a PacBio Sequel II (software/chemistry v9.0.0) following diffusion loading. Sequence data was processed to generate CCS reads using the default settings of the CCS application (v4.2.0) in SMRT Link (v9.0.0); minimum parameters for passes (3), accuracy (0.99), CCS read length (10) and maximum CCS read length (50000). CCS reads were assembled using the Improved Phased Assembly (IPA) method (PacBio). The IPA assembly tool (https://github.com/PacificBiosciences/pbbioconda/wiki/Improved-Phased-Assembler) uses the HiFi sequencing reads (high-quality consensus reads) and generates phased assembly. This produces a primary contig folder, including the main assembly and an associated contig, containing haplotigs and duplications. The CCS reads (bam) were assembled in separate conda environments for each species. For all assemblies, 24 CPUs and 120Gb of memory was employed.

Assessment of completeness

The completeness of genome assemblies was evaluated using benchmarking universal single-copy orthologues (BUSCO) analysis (v4.1.2), using genome mode and lineage Eukaryota_odb10 dataset.

Availability of data

Sequence data from Pac Bio (Sequel), Oxford Nanopore Technology (PromethION) and BGI (single-tube Long Fragment Read) analysis of *M. jansenii* was described by Murigneux et al [2]. BGI, PacBio, ONT, and Illumina sequencing data generated in that study were deposited in the SRA under BioProject PRJNA609013and BioSample SAMN14217788. Accession numbers are as follows: BGI (SRR11191908), PacBio (SRR11191909), ONT PromethION (SRR11191910), ONT MinION (SRR11191911), and Illumina (SRR11191912). Assemblies and other supporting data are available from the *GigaScience* GigaDB repository [25]. Pac Bio HiFi reads described in this paper are deposited as CCS reads under NCBI Sequence Read Archive (SRA) submission number: SUB8909497 for macadamia; SUB8909619 for jojoba and SUB8909729 for avocado.

Additional files

Figure S1:Size distribution of reads sequenced

Competing interests

The authors declare that they have no competing interests.

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Author's contributions

Contributions of authors were as follows. RH, AF, VM, AM, IA Conceptualization; PS, OA, BA, ON, VM, AF Data curation; PS, OA, BA,ON, GM, VM ,AM, AF Formal Analysis; RH, AF, IA, AM Funding acquisition; PS, OA, BA, ON ,NM, VM, AM, AF,RH Investigation; IA, BT, Resources; IA, BT, NM, RH, AM, AF Supervision; RH, PS, ON, AM Writing – original draft; All authors Writing – review & editing

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Figure legends

Figure1 Influence of data volume on assembly for Macadamia species

N50 of contigs is plotted against the genome coverage. Genome sizes used to calculate coverage were; *M.integrifolia*, 895Mb [12]; *M janseni*, 780Mb [2]; *M. tetraphylla*, 758Mb [23] and *M. ternifolia*, 758Mb (not known but assumed the same as *M. tetraphylla* due to similar assembly size).

Figure 2 Influence of data volume on assembly for diverse species. N50 of contigs is plotted against the genome coverage. Genome sizes used to calculate coverage were, jojoba, 1003Mb[14]; avocado, 920Mb [24] and as in figure 1 for *Macadamia* species

Figure 3 Decrease in length of total assembly as more genome coverage is used in the assembly

Figure 4 Improvement in genome completeness (BUSCO%) with genome coverage

References

1. Michael, T.P.; VanBuren, R. Building near-complete plant genomes. *Current Opinion in Plant Biology* **2020**, *54*, 26-33, doi:10.1016/j.pbi.2019.12.009.
2. Murigneux, V.; Rai, S.K.; Furtado, A.; Bruxner, T.J.C.; Tian, W.; Harliwong, I.; Wei, H.; Yang, B.; Ye, Q.; Anderson, E., et al. Comparison of long-read methods for sequencing and assembly of a plant genome. *Gigascience* **2020**, *9*, doi:10.1093/gigascience/giaa146.
3. Hon, T.; Mars, K.; Young, G.; Tsai, Y.C.; Karalius, J.W.; Landolin, J.M.; Maurer, N.; Kudrna, D.; Hardigan, M.A.; Steiner, C.C., et al. Highly accurate long-read HiFi sequencing data for five complex genomes. *Sci Data* **2020**, *7*, doi:ARTN 399

10.1038/s41597-020-00743-4.

4. Lang, D.; Zhang, S.; Ren, P.; Liang, F.; Sun, Z.; Meng, G.; Tan, Y.; Li, X.; Lai, Q.; Han, L., et al. Comparison of the two up-to-date sequencing technologies for genome assembly: HiFi reads of Pacific Biosciences Sequel II system and ultralong reads of Oxford Nanopore. *Gigascience* **2020**, *9*, doi:10.1093/gigascience/giaa123.
5. Amarasinghe, S.L.; Su, S.; Dong, X.Y.; Zappia, L.; Ritchie, M.E.; Gouil, Q. Opportunities and challenges in long-read sequencing data analysis. *Genome Biology* **2020**, *21*, doi:ARTN 30

10.1186/s13059-020-1935-5.

6. Wendel, J.F.; Jackson, S.A.; Meyers, B.C.; Wing, R.A. Evolution of plant genome architecture. *Genome Biology* **2016**, *17*, doi:ARTN 37

10.1186/s13059-016-0908-1.

7. Juma, I.; Geleta, M.; Nyomora, A.; Saripella, G.V.; Hovmalm, H.P.; Carlsson, A.S.; Fatih, M.; Ortiz, R. Genetic diversity of avocado from the southern highlands of Tanzania as revealed by microsatellite markers. *Hereditas* **2020**, *157*, 40, doi:10.1186/s41065-020-00150-0.
8. Tobe, H.; Yasuda, S.; Oginuma, K. Seed Coat Anatomy, Karyomorphology, and Relationships of Simmondsia (Simmondsiaceae). *Bot Mag Tokyo* **1992**, *105*, 529-538, doi:Doi 10.1007/Bf02489427.
9. Kyriakidou, M.; Anglin, N.L.; Ellis, D.; Tai, H.H.; Stromvik, M.V. Genome assembly of six polyploid potato genomes. *Sci Data* **2020**, *7*, doi:ARTN 88

10.1038/s41597-020-0428-4.
10. Kyriakidou, M.; Tai, H.H.; Anglin, N.L.; Ellis, D.; Stromvik, M.V. Current Strategies of Polyploid Plant Genome Sequence Assembly. *Front Plant Sci* **2018**, *9*, doi:ARTN 1660

10.3389/fpls.2018.01660.
11. Nock, C.J.; Baten, A.; Barkla, B.J.; Furtado, A.; Henry, R.J.; King, G.J. Genome and transcriptome sequencing characterises the gene space of Macadamia integrifolia (Proteaceae). *Bmc Genomics* **2016**, *17*, doi:ARTN 937

10.1186/s12864-016-3272-3.
12. Nock, C.J.; Baten, A.; Mauleon, R.; Langdon, K.S.; Topp, B.; Hardner, C.; Furtado, A.; Henry, R.J.; King, G.J. Chromosome-Scale Assembly and Annotation of the Macadamia Genome (Macadamia integrifoliaHAES 741). *G3-Genes Genomes Genetics* **2020**, *10*, 3497-3504, doi:10.1534/g3.120.401326.
13. Rendon-Anaya, M.; Ibarra-Laclette, E.; Mendez-Bravo, A.; Lan, T.Y.; Zheng, C.F.; Carretero-Paulet, L.; Perez-Torres, C.A.; Chacon-Lopez, A.; Hernandez-Guzman, G.; Chang, T.H., et al. The avocado genome informs deep angiosperm phylogeny, highlights introgressive hybridization, and reveals pathogen-influenced gene space adaptation. *Proceedings of the National Academy of Sciences of the United States of America* **2019**, *116*, 17081-17089, doi:10.1073/pnas.1822129116.
14. Sturtevant, D.; Lu, S.P.; Zhou, Z.W.; Shen, Y.; Wang, S.; Song, J.M.; Zhong, J.S.; Burks, D.J.; Yang, Z.Q.; Yang, Q.Y., et al. The genome of jojoba (Simmondsia chinensis): A taxonomically isolated species that directs wax ester accumulation in its seeds. *Sci Adv* **2020**, *6*, doi:ARTN eaay3240

10.1126/sciadv.aay3240.
15. Belser, C.; Istace, B.; Denis, E.; Dubarry, M.; Baurens, F.C.; Falentin, C.; Genete, M.; Berrabah, W.; Chevre, A.M.; Delourme, R., et al. Chromosome-scale assemblies of plant genomes using nanopore long reads and optical maps. *Nat Plants* **2018**, *4*, 879-+, doi:10.1038/s41477-018-0289-4.
16. Wang, W.W.; Das, A.; Kainer, D.; Schalamun, M.; Morales-Suarez, A.; Schwessinger, B.; Lanfear, R. The draft nuclear genome assembly of Eucalyptus pauciflora: a pipeline for comparing de novo assemblies. *Gigascience* **2020**, *9*, doi:ARTN giz160

10.1093/gigascience/giz160.
17. Monat, C.; Padmarasu, S.; Lux, T.; Wicker, T.; Gundlach, H.; Himmelbach, A.; Ens, J.; Li, C.D.; Muehlbauer, G.J.; Schulman, A.H., et al. TRITEX: chromosome-scale sequence assembly of Triticeae genomes with open-source tools. *Genome Biology* **2019**, *20*, doi:ARTN 284

10.1186/s13059-019-1899-5.
18. Li, F.W.; Harkess, A. A guide to sequence your favorite plant genomes. *Appl Plant Sci* **2018**, *6*, doi:ARTN e1030

10.1002/aps3.1030.

325 19. Cheng, B.; Furtado, A.; Henry, R.J. Long-read sequencing of the coffee bean transcriptome
326 reveals the diversity of full-length transcripts. *Gigascience* **2017**, *6*,
327 doi:10.1093/gigascience/gix086.
328 20. Furtado, A. DNA Extraction from Vegetative Tissue for Next-Generation Sequencing. *Cereal*
329 *Genomics: Methods and Protocols* **2014**, *1099*, 1-5, doi:10.1007/978-1-62703-715-0_1.
330 21. Zou, Y.; Mason, M.G.; Wang, Y.; Wee, E.; Turni, C.; Blackall, P.J.; Trau, M.; Botella, J.R. Nucleic
331 acid purification from plants, animals and microbes in under 30 seconds. *PLoS Biol* **2017**, *15*,
332 e2003916, doi:10.1371/journal.pbio.2003916.
333 22. Bienvenue, J.M.; Duncalf, N.; Marchiarullo, D.; Ferrance, J.P.; Landers, J.P. Microchip-based
334 cell lysis and DNA extraction from sperm cells for application to forensic analysis. *J Forensic*
335 *Sci* **2006**, *51*, 266-273, doi:10.1111/j.1556-4029.2006.00054.x.
336 23. Niu, Y.-F.; Li, G.-H.; Ni, S.-B.; He, X.-Y.; Zheng, C.; Liu, Z.Y.; Gong, L.D.; Kong, G.H.; Liu, J.
337 Genome assembly and annotation of *Macadamia tetraphylla. bioXriv* **2020**
338 doi: https://doi.org/10.1101/2020.03.11.987057
339 24. Talavera, A.; Soorni, A.; Bombarely, A.; Matas, A.J.; Hormaza, J.I. Genome-Wide SNP
340 discovery and genomic characterization in avocado (Persea americana Mill.). *Scientific*
341 *Reports* **2019**, *9*, doi:ARTN 20137

342 10.1038/s41598-019-56526-4.

343 25. Murigneux V, Rai SK, Furtado A, et al. Supporting data for "Comparison of long-read
344 methods for sequencing and assembly of a plant genome." GigaScience Database 2020.
345 http://dx.doi.org/10.5524/100812.

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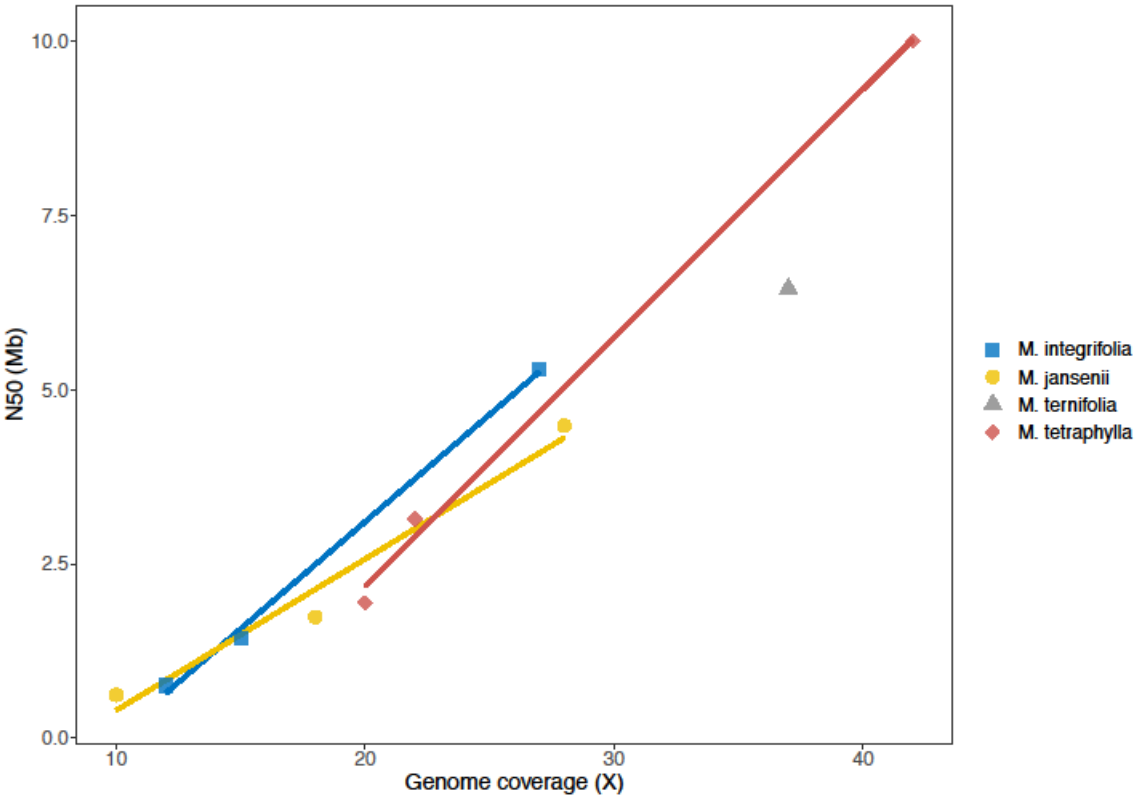
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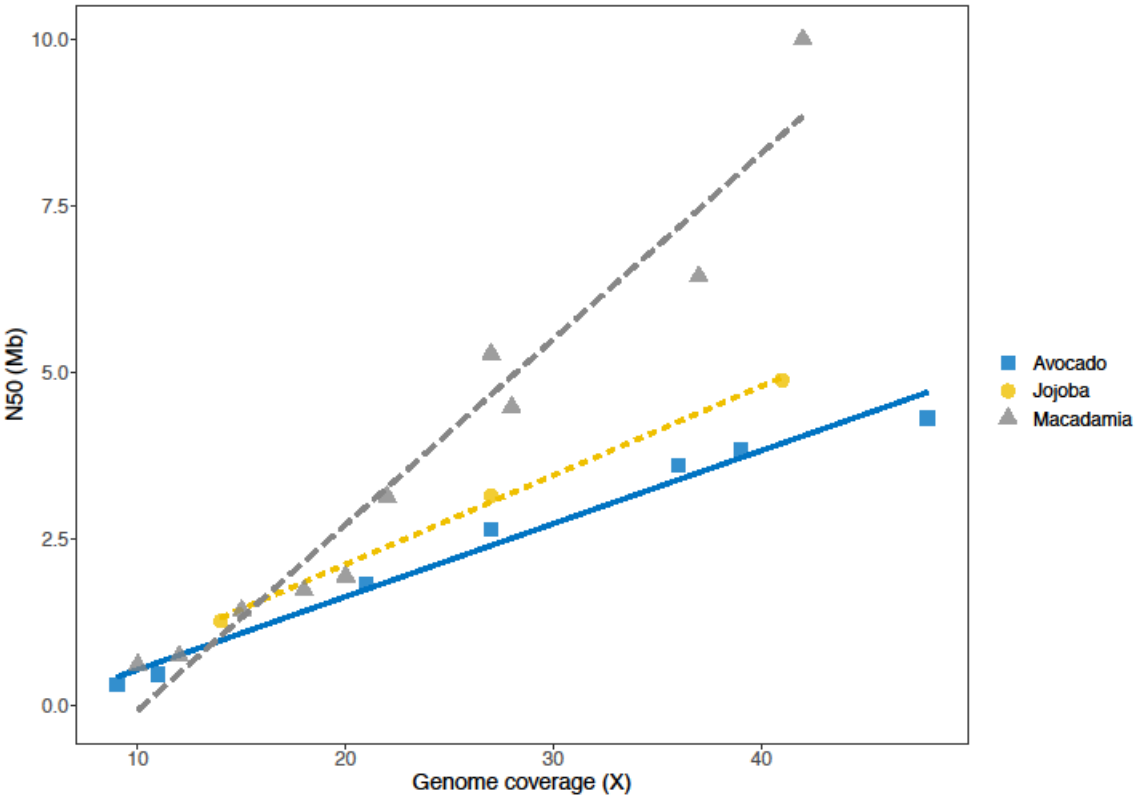
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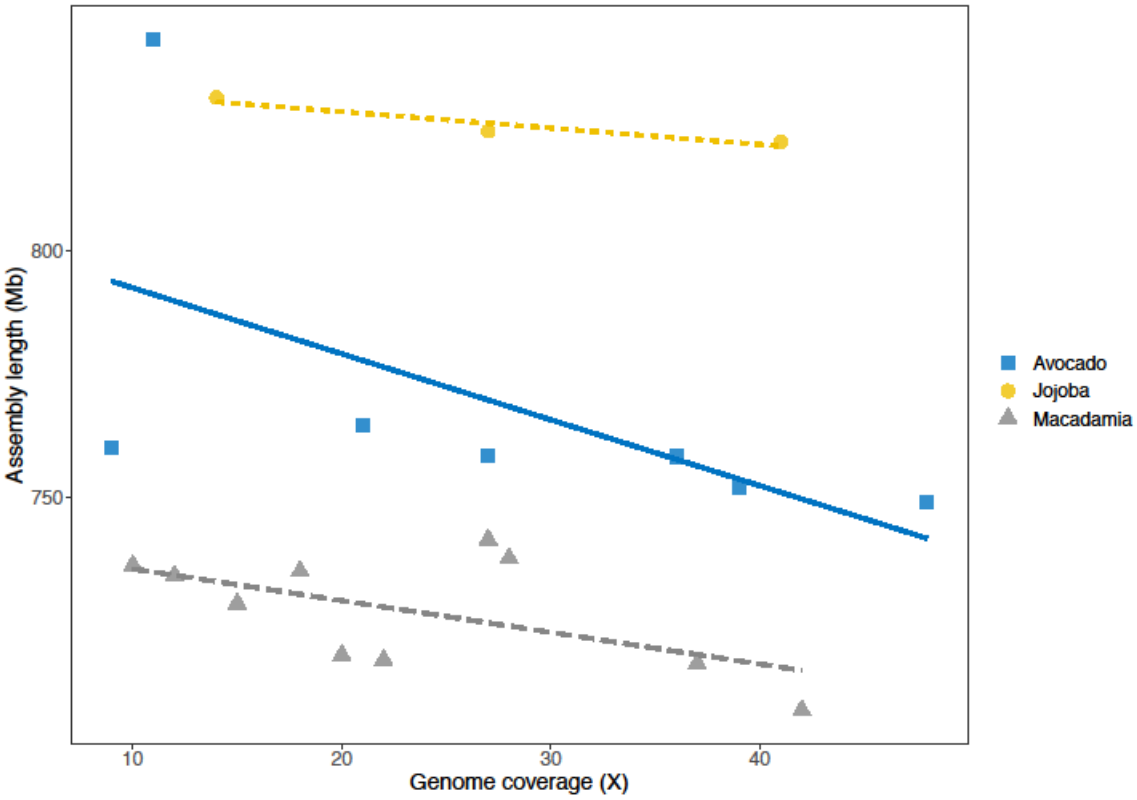
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Jojoba
Macadamia

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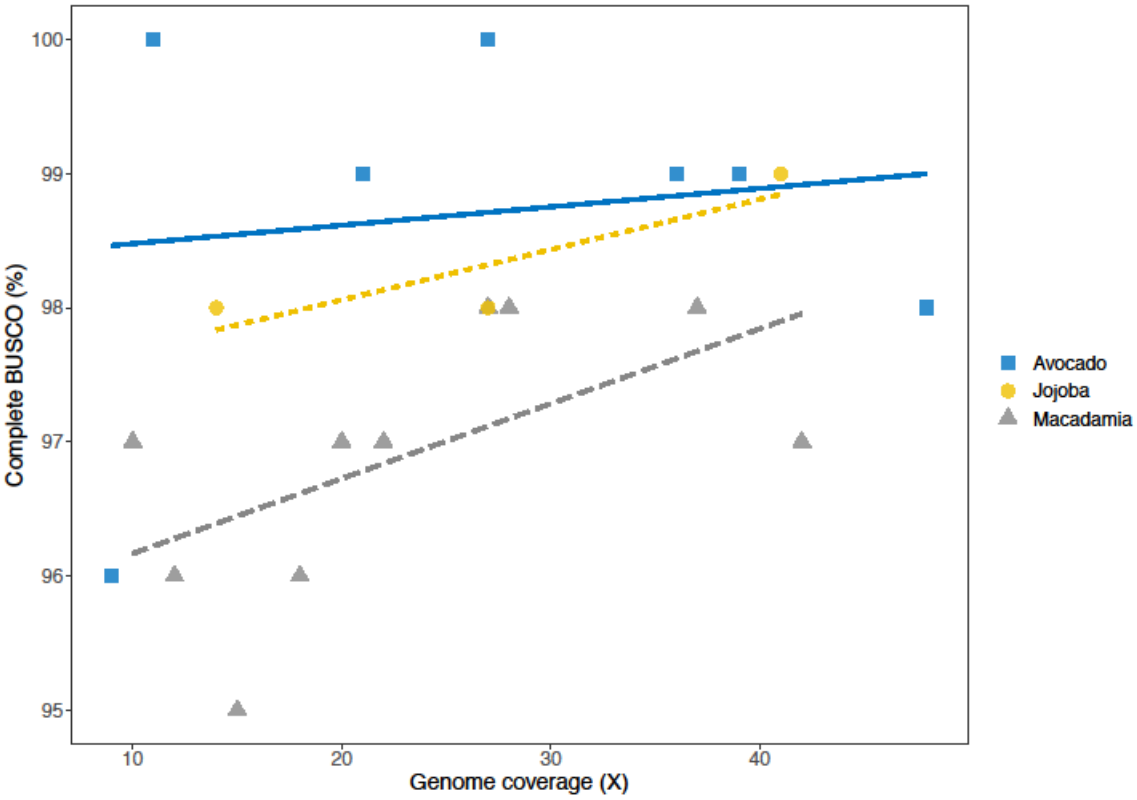
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Genome coverage (X)
Avocado
Jojoba
Macadamia

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364 Figure S1: Size distribution of reads sequenced

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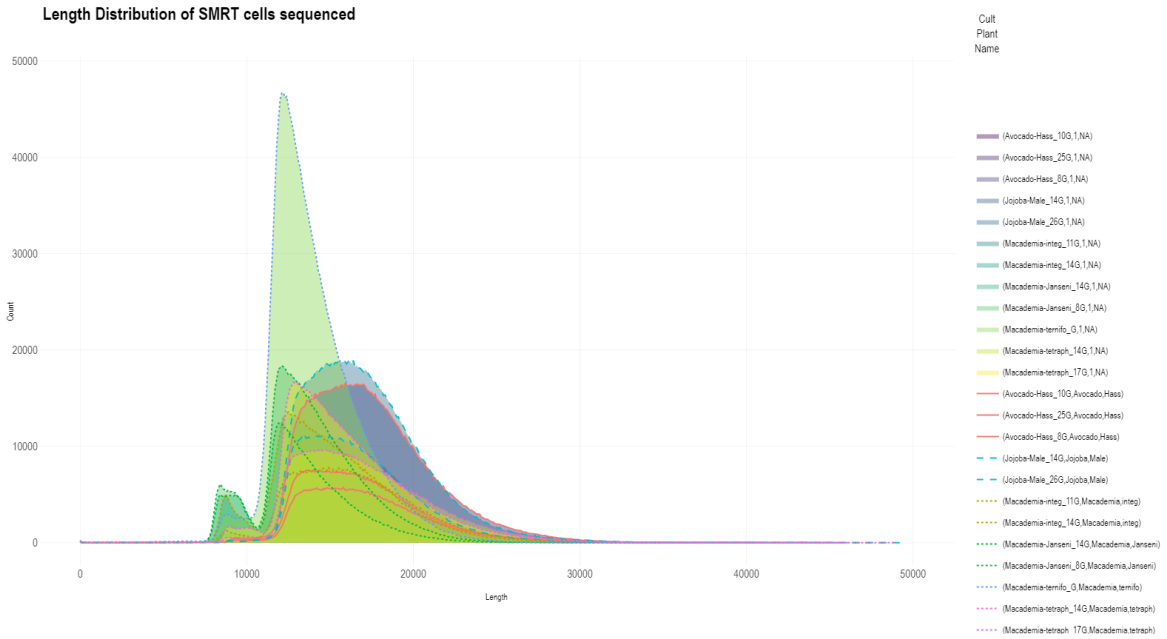

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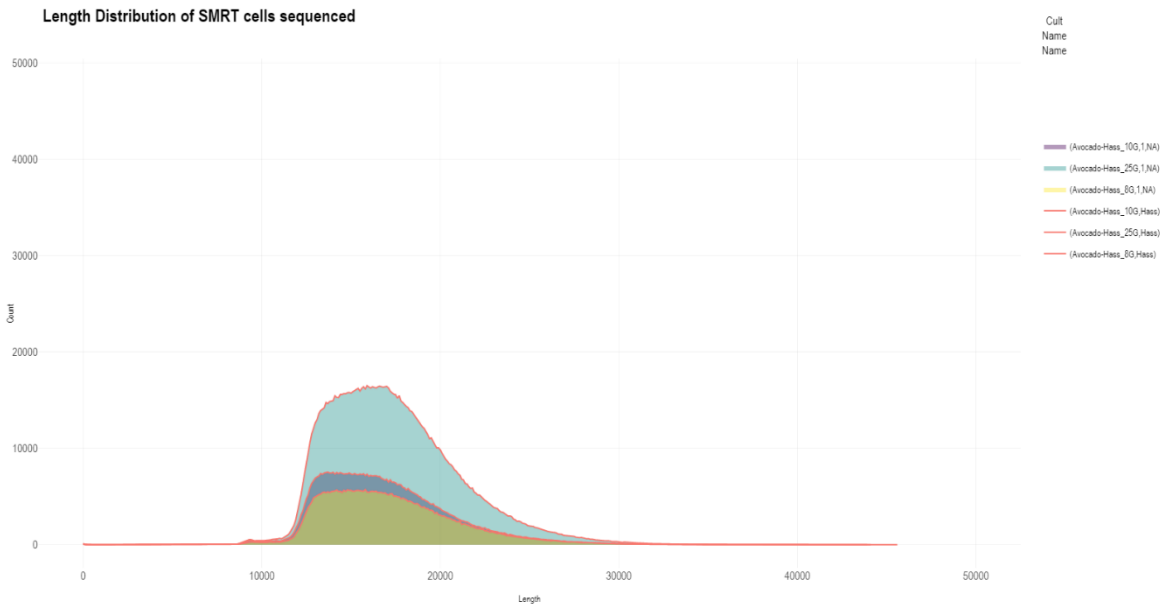

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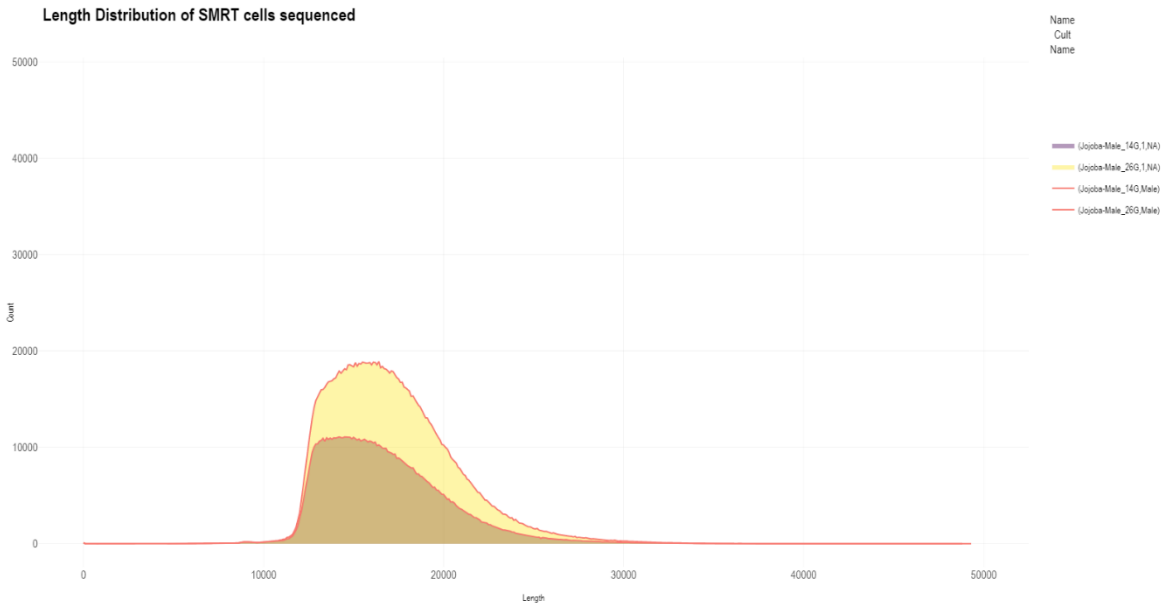

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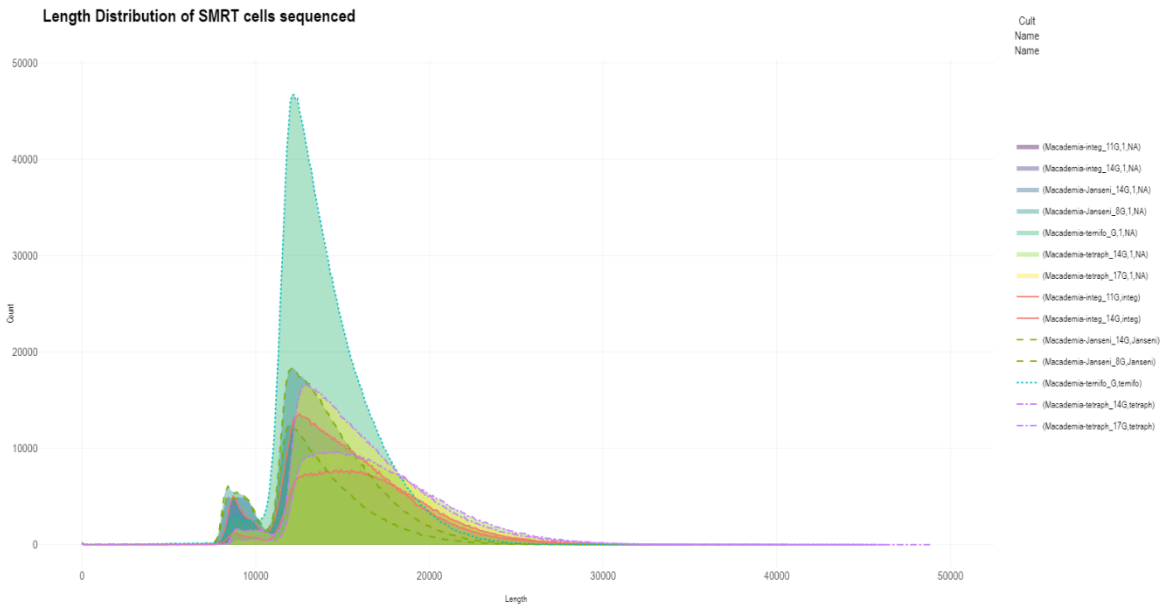
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(Macademia-integ_14G,integ)
(Macademia-Janseni_14G,Janseni)
(Macademia-Janseni_8G,Janseni)
(Macademia-temifo_G,temifo)
(Macademia-tetraph_14G,tetraph)
(Macademia-tetraph_17G,tetraph)

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