An auxin-inducible, GAL4-compatible, gene expression system for *Drosophila*

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Abstract

The ability to control transgene expression, both spatially and temporally, is essential for studying model organisms. In *Drosophila*, spatial control is primarily provided by the GAL4/UAS system, whilst temporal control relies on a temperature-sensitive GAL80 (which inhibits GAL4) and drug-inducible systems. However, these are not ideal. Shifting temperature can impact on many physiological and behavioural traits, and the current drug-inducible systems are either leaky, toxic, incompatible with existing GAL4-driver lines, or do not generate effective levels of expression. Here we describe the Auxin-inducible Gene Expression System (AGES). AGES relies on the auxin-dependent degradation of a ubiquitously expressed GAL80, and therefore, is compatible with existing GAL4-driver lines. Water-soluble auxin is added to fly food at a low, non-lethal, concentration, which induces expression comparable to uninhibited GAL4 expression. The system works in both larvae and adults, providing a stringent, non-lethal, cost-effective, and convenient method for temporally controlling GAL4 activity in *Drosophila*.

Introduction

The ability to manipulate the expression of specific genes in model organisms has been the cornerstone of genetics research over the last 50 years. It is from such studies that geneticists have been able to elucidate fundamental biological process, such as those underlying neurodegenerative diseases (Lu and Vogel, 2009), oncogenic mechanisms (Villegas, 2019), and neural circuitry involved in memory formation (Cognigni et al., 2018).

The fruit fly, *Drosophila melanogaster*, has been at the forefront of genetics since its cultivation in the lab in the early 1900s, by geneticist Thomas Morgan (Morgan, 1910). In the 100 years since, a plethora of genetic tools have been developed for this model organism, enabling geneticists to tightly control and manipulate gene expression, allowing them to study the roles of genes in development, physiology, and behaviour. One of the most prominent developments is the GAL4/UAS system, which has been an incredibly powerful tool for the spatial control of transgenes in *Drosophila* (Brand and Perrimon, 1993). This system relies on a cloned

or endogenous promoter to drive the expression of the yeast transcription factor GAL4, which can then bind to the UAS (upstream activating sequence) sites upstream of a transgene and activate its expression. Since the development of this system, *Drosophila* researchers have created thousands of genetically distinct lines to enable expression of transgenes in specific populations of cells (for example (Jenett et al., 2012; Robie et al., 2017)). The majority of these lines are publicly available at the Bloomington Drosophila Stock Center (BDSC) and Vienna Drosophila Resource Center (VDRC).

GAL4 activity can be regulated by GAL80, a protein which antagonises GAL4 by inhibiting its activation domain (Lue et al., 1987), and thus the ability of the GAL4 to induce transcript expression (Lee and Luo, 1999; Suster et al., 2004). The generation of a fly line that ubiquitously expressed a temperature-sensitive version of GAL80 ($GAL80^{ts}$) (Matsumoto et al., 1978) was a significant advance for the field, as it allowed temporal control of GAL4-induced transgene expression (McGuire et al., 2003). Flies can be kept at temperatures permissive of transcript-expression (29°C) or temperatures that inhibit expression (18°C), offering researchers to control not only where, but when transgenes are expressed. The versatility of this system enabled researchers to determine time-specific effects of gene function, and is a system that is widely employed within the *Drosophila* community.

Although the impact of this technology on the field cannot be understated, it is becoming increasingly evident that using a temperature shift to induce gene expression is far from optimal for some experiments, especially in fields such as behaviour, neuropathy, and ageing. Flies, as ectotherms, are highly dependent on their ambient temperature, which affects a wide range of physiological and behavioural traits (Abram et al., 2017). These include sleep (Beckwith and French, 2019), territorial success (Zamudio et al., 1995), lifespan (McCabe and Partridge, 1997), development rate (Danjuma et al., 2014), immunity (Hunt et al., 2016), metabolism (Schou et al., 2017) and even their microbiome (Moghadam et al., 2018). Therefore, the use of a significant temperature shift to induce gene expression should be avoided in these contexts.

As an alternative approach, a range of drug-inducible systems have been developed, where the drug can be administered to flies in their diet to activate transcript expression (Barwell et al., 2017; Kogenaru and Isalan, 2018; Osterwalder et al., 2001; Potter et al., 2010; Sethi and Wang, 2017). These systems each have advantages and disadvantages (see Table 1) and are not often compatible with available GAL4 lines, thus requiring the creation (and optimisation) of their own drivers. In addition, the systems that are compatible with existing GAL4 lines can affect the fly's physiology or survival, thereby limiting their use, and the certainty of any findings when employed. For example, the GeneSwitch (GS) system (Osterwalder et al., 2001) is not compatible with existing GAL4 lines and requires the user to generate a new modified GAL4 line. Furthermore, the system shows leaky expression in the absence of the drug (RU-486) (Poirier et al., 2008; Scialo et al., 2016), and RU-486 can cause behavioural changes (Li and Stavropoulos, 2016) and is dangerous to handle for pregnant women as it can cause termination of the pregnancy (Avrech et al., 1991). Another drug-inducible system utilised in *Drosophila* is the QF system (Potter et al., 2010). Here, QF is a transcriptional activator that can be inhibited by QS (analogous to GAL4 and GAL80). Quinic acid inhibits the repressive

73 action of QS, therefore, can switch on transgene expression. Quinic acid is non-toxic for flies, however, the
74 drawback of this system is that it is not compatible with the vast collection of existing GAL4 lines.

EXPRESSION SYSTEM	REFERENCE	ADVANTAGES	DISADVANTAGES
GENESWITCH	(Osterwalder et al., 2001)	Non-toxic for adult flies	Not compatible with existing GAL4 lines Leaky expression (Poirier et al., 2008; Scialo et al., 2016) Larval exposure impacts on adult sleep patterns (Li and Stavropoulos, 2016) Drug is unsafe to handle for female researchers Drug is expensive
QF SYSTEM	(Potter et al., 2010)	Non-toxic for flies	Not compatible with existing GAL4 lines
TET-OFF GAL80	(Barwell et al., 2017)	GAL4/UAS compatible	Long induction time (> 5 days) Requires x2 copies of the GAL80 transgene Affects microbiota
TMP INDUCIBLE	(Kogenaru and Isalan, 2018; Sethi and Wang, 2017)	GAL4/UAS compatible	Currently not compatible for general use with existing GAL4 lines. TMP drug is only dissolvable in DMSO (affects survival of larvae) or has to be added as a dry powder to food.

75 **Table 1 –** Main advantages and disadvantages of the most common drug-inducible gene expression systems currently
76 available to the *Drosophila* community.

77 The auxin-degron system, first identified in *Arabidopsis*, involves the auxin-dependent ubiquitination, and
78 subsequent degradation, of proteins that are tagged with a specific auxin-inducible degron (AID) sequence
79 (Dharmasiri et al., 2005; Dharmasiri and Estelle, 2002; Li et al., 2019). Its natural function allows for the
80 prompt elimination of Aux/IAA transcription factors in plants and has been adapted for use in other species,
81 most notably *C. elegans*, to artificially target proteins for rapid degradation (Zhang et al., 2015). In this
82 system, the F-box protein TIR1, an auxin-receptor, binds with conserved proteins Skp1 and Cullin to form a
83 Skp1–Cul1–F-box (SCF) E3 ligase complex which ubiquitinates proteins tagged with an AID sequence in an
84 auxin-dependent manner. The specificity and versatility of the system has made it an effective tool in multiple
85 systems including mice (Yesbolatova et al., 2020), and human cell culture (Li et al., 2019).
86 Importantly, the auxin-degron system has been successfully employed in *Drosophila* (Chen et al., 2018; Trost
87 et al., 2016). Trost and colleagues showed that AID-tagged EYFP and AID-tagged Rux protein levels could be
88 knocked down in S2 cell lines and transgenic *Drosophila*, respectively. While Chen and colleagues knocked
89 down AID-tagged PERIOD protein the adult brain. Inspired by these studies, we have applied the auxin-
90 degron system to enable both temporal and spatial control of transgene expression when combined with the
91 GAL4/UAS system in *Drosophila*. This was achieved by creating a ubiquitously expressed GAL80 fused with
92 AID tags, such that GAL80 is degraded in the presence of the auxin phytohormone (Figure 1). We demonstrate
93 that this system works in both larvae (5 mM auxin) and adult flies (10 mM), which are concentrations that do
94 not affect fly survival, development, or locomotion (larval crawling and adult climbing). This Auxin-inducible
95 Gene Expression System (AGES) is safe to handle, cheap, easy to prepare, provides tuneable and stringent

(i.e. not leaky) expression, and most importantly, is compatible with the majority of existing GAL4-driver lines developed by the *Drosophila* community.

Results

Generation of an auxin-degradable GAL80 *Drosophila* line

To generate an auxin inducible system for control of GAL4 activity in *Drosophila,* we designed a transgene that would ubiquitously express two proteins, TIR1, and AID-tagged GAL80. (Figure 1). We included shorter AID sequences (minimal degron of IAA17) than previously used in *Drosophila* (Trost et al., 2016). Fusion of this minimal AID to luciferase resulted in a fusion protein with short half-life in plants (~ 10 min) (Dreher et al., 2006). We also instead used *Arabidopsis thaliana* TIR1 (AtTIR1), which has two point mutations (improving affinity and auxin sensitivity) that can deplete nuclear and cytoplasmic proteins in *C. elegans* (Zhang et al., 2015). The P2A sequence between AtTIR1 and AID-tagged GAL80 encodes a self-cleaving peptide that is known to work efficiently in *Drosophila* cell culture (Daniels et al., 2014). The *AtTIR1-T2A-AID-GAL80-AID* sequence was codon optimized, synthesized, and cloned downstream of the *αTub84B* promoter in *pattB* (Figure 1 and Figure 1 – figure supplement 1). The *αTub84B* (tubulin) promoter drives expression of the bicistronic sequence in all cells, all of the time, and (in the absence of auxin) will inhibit any GAL4 activity. When present, auxin will tether the TIR1 to the AID sequences, triggering the degradation of GAL80 and the release of GAL4 inhibition (Figure 1).

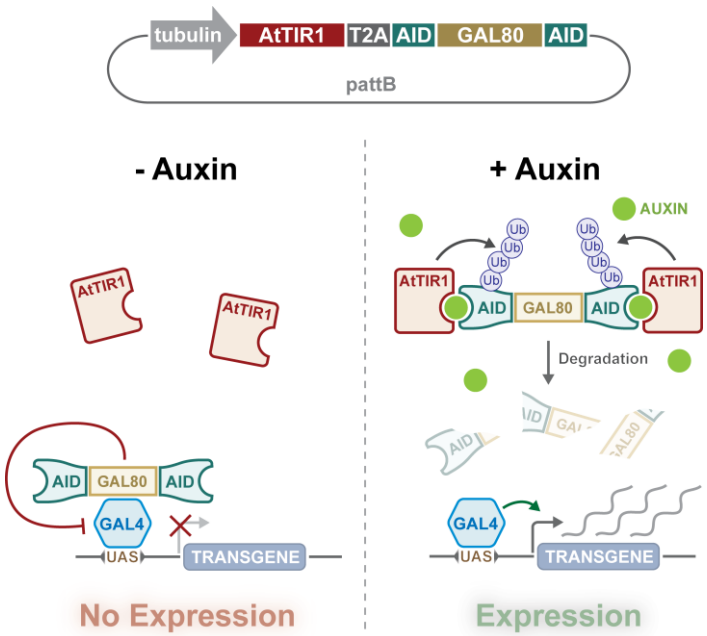


Figure 1. Design and action of AGES. A tubulin promoter ubiquitously expresses the bicistronic TIR1 and AGES cassette. AID-tagged GAL80 is cleaved from the TIR1 due to the presence of the T2A sequence. In the absence of auxin, GAL80 can inhibit GAL4 activity, whilst the presence of auxin induces degradation of GAL80, allowing GAL4 to drive the expression of *UAS*-transgene(s).

AGES allows induction of GAL4 activity in the adult fat body

We combined the AGES system with *c564*-GAL4 to see if we could get inducible expression in fat body. A nuclear localised GFP (nls-GFP) was used as a reporter of GAL4 activity. We first examined adult female flies that were placed on food containing different concentrations of K-NAA auxin, which has shown tobe more water soluble than NAA (Martinez et al., 2020). A negative control consisting of fly food without auxin (0 mM

auxin) was included to investigate whether there was any leaky GAL4 activity in absence of auxin. After 24 hours, GFP fluorescence in the abdomen were assayed (note that there is some naturally occurring autofluorescence in the fly abdomen). At 5 mM and 10 mM, GFP fluorescence is clearly detectable in the abdomen of live female flies (Figure 2A). Based on this assay, 10 mM for 24 hours results in ~60% of the GFP fluorescence levels seen in the positive control (Figure 2B). At 1 mM there is no detectable GFP above background fluorescence, and the levels are indistinguishable from both the 0 mM and the negative control (lacking both the GAL4 driver and the *UAS*-nls-GFP). To provide an alternative and more direct assay of transgene induction, quantitative PCR (qPCR) was used to measure the levels of *GFP* mRNA (Figure 2C). Here 5 mM and 10 mM show levels of mRNA equivalent to the positive control, while 0 mM showed no significant difference when compared with the negative control.

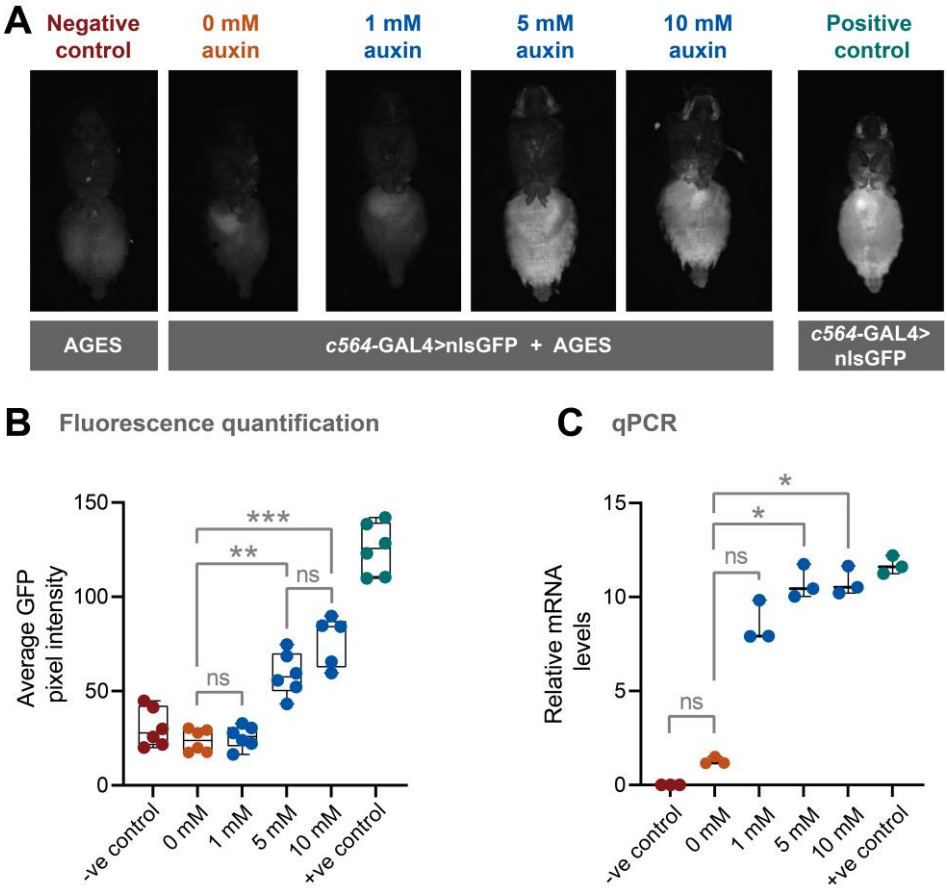


Figure 2. AGES effectively induces GAL4 activity in *Drosophila* adults. A) Ventral images of live females that express GAL4 in fat body tissue. Ingestion of food containing auxin (>= 5 mM for 24 hours) induces GAL4 activity and the expression of GFP. B) Quantification of GFP levels. Pixel intensity thresholding was performed to isolate abdomens as regions of interest. The average pixel intensities from 6 replicates were quantified and analysed using Kruskal-Wallis test with Dunn pair-wise comparison (***, $p < 0.001$ and **, $p < 0.01$). C) qPCR data for *GFP* mRNA levels using different concentrations of auxin (3 biological replicates). Values were normalised to housekeeping gene *RpL4* (*Ribosomal Protein L4*) and relative expression levels (compared to the negative control) were calculated using the ΔΔCt method. Y-axis displaying ΔΔCt values and statistics done using Kruskal-Wallis test with Dunn pair-wise comparison (*, $p < 0.05$). See Figure 2 - source data 1 for raw data.

Adult males eat less than females (Wong et al., 2009), therefore, this will impact on auxin ingestion and possibly on transgene induction. As with females, we examined both GFP fluorescence levels in the abdomen,

and GFP mRNA levels in male flies (Figure 2 – figure supplement 1). Although reduced compared to females, there is still a robust induction of GFP expression.

To assess how quickly transgene expression is turned off after removal of auxin from the food, we performed a time course experiment using adult females and 10 mM auxin. After 24 hours, flies were shifted to food without auxin and GFP fluorescence levels in the abdomen measured every 24 hours. At 48 hours (24 hours without auxin) GFP levels were reduced (Figure 2 – Figure Supplement 2A). At 72 and 96 hours, the levels were not significantly different to 0 hours. Furthermore, we tested how stable auxin is in prepared food kept at 4°C. Even after 15 weeks, the auxin food could induce GFP levels to that of newly prepared food (Figure 2 – Figure Supplement 2B).

AGES allows induction of GAL4 activity in *Drosophila* larvae

In addition to the need for temporal control of GAL4 activity in the adult, researchers might also want to induce expression at specific points during larval development. To test if AGES works in larvae, we used the *c564*-GAL4 fat body driver and tested multiple auxin concentrations (Figure 3A, C). Here we observed that expression could be induced at lower concentrations of auxin. When using food containing no auxin, there is no detectable GFP, whereas with 1 mM auxin, GFP is expressed, although at lower levels than with higher concentrations of auxin (Figure 3A, C). 5 mM auxin is an optimal concentration, as concentrations above that do not increase GFP levels. 5 mM auxin can induce low level GFP expression after 6 hours (Figure 3B, D). However, 18 hours is required to provide maximum levels of expression.

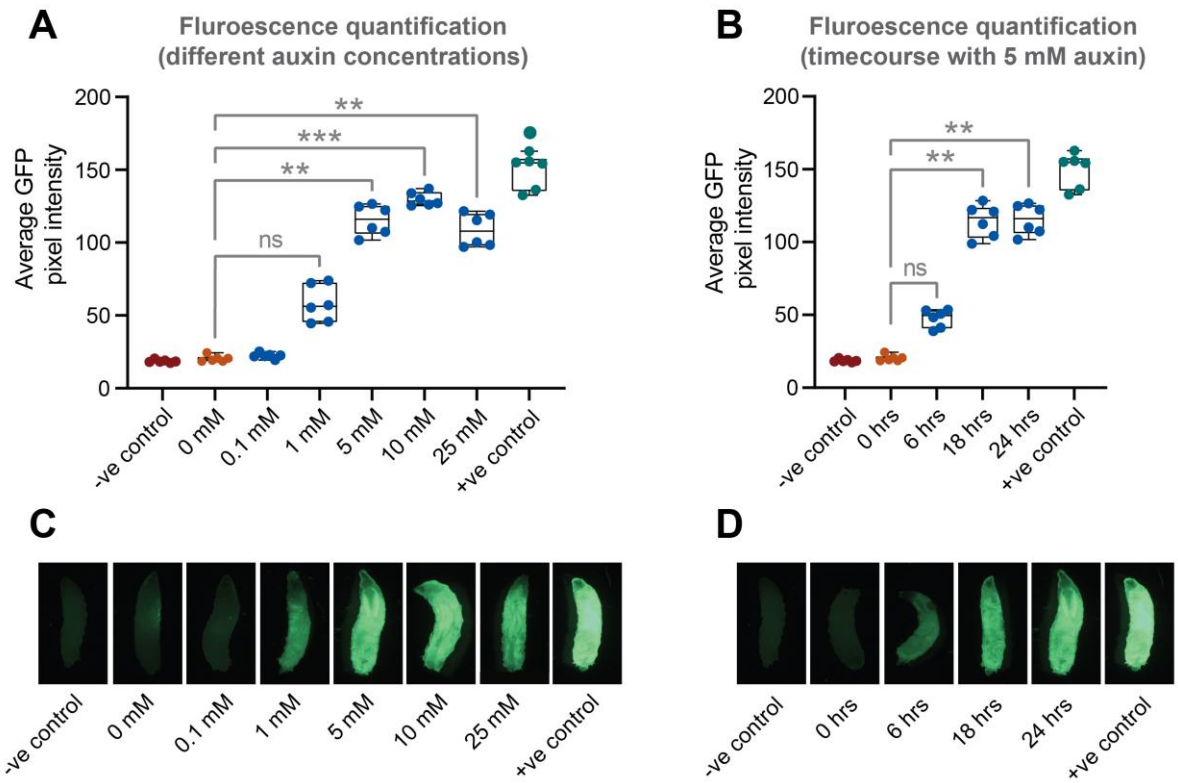
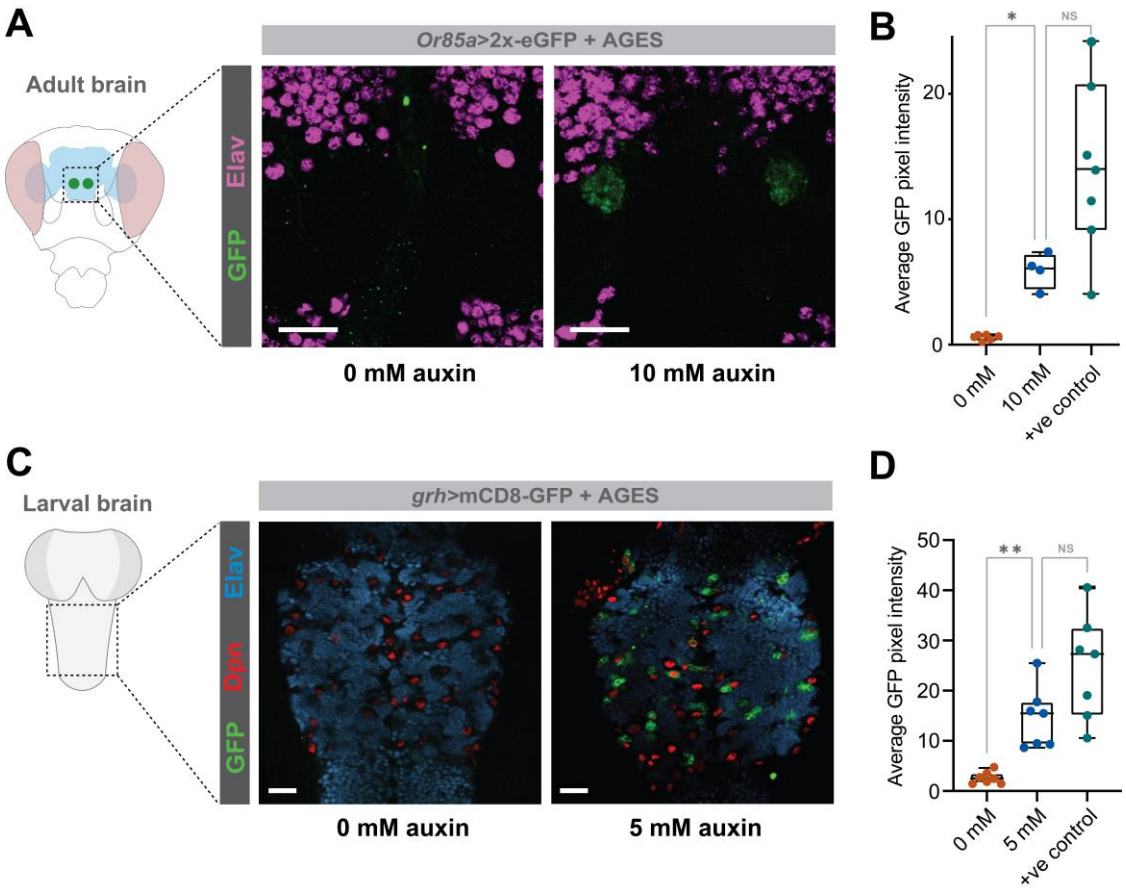


Figure 3. AGES allows induction of GAL4 activity in *Drosophila* larvae. A) GFP fluorescence quantification in fat body tissue after induction (for 24 hours) on food with different concentrations of auxin. The average pixel intensities from 6 larvae were quantified and analysed using Kruskal-Wallis test with Dunn pair-wise comparison (***, $p < 0.001$ and **, $p < 0.01$). B) Time course of GFP expression in fat body tissue when using 5 mM auxin. The average pixel intensities from 6 larvae were quantified and analysed using Kruskal-Wallis test with Dunn pair-wise comparison (**, $p < 0.01$) C) Representative images of larvae fed on different concentrations of auxin. D) Representative images of larvae imaged at each time interval since induction. See Figure 3 - source data 1 for raw data.

157 **Use of AGES for controlling cell specific transgene expression in the larval and adult nervous**
158 **system**

159 For AGES to work with GAL4 lines expressed in the central nervous system, auxin must be able to pass the
160 selectively-permeable glial membrane, akin to the mammalian blood-brain barrier (Limmer et al., 2014). NAA
161 can cross the blood-brain barrier in *Drosophila* (Chen et al., 2018). To verify this using K-NAA, we first used
162 *elav-GAL4* (expressed in all neurons) and examined the expression of GFP in the adult brain. Similar to the
163 fat body *c564*-GAL4 driver, we observe robust expression of GFP, when flies are fed auxin food (10 mM) for
164 24 hours (Figure 4 – Figure Supplement 1). This demonstrates that AGES works in the central nervous system
165 and that K-NAA auxin can cross the blood brain barrier in *Drosophila*.

166 We then tested more restricted GAL4 driver lines in the larval ventral nerve cord (*grh*-GAL4) and in the adult
167 (*Or85a*-GAL4). In both cases there was no detectable GAL4 activity in the absence of auxin, while in the
168 presence of auxin (10 mM for adults and 5 mM for larvae) GFP in a pattern consistent with the GAL4-driver
169 is observed (Figure 4A, C) at levels significantly above background fluorescence levels (Figure 4B, D).



170

171 **Figure 4. AGES allows induction of GAL4 activity in the *Drosophila* adult and larval brain.** A) GFP
172 fluorescence driven by *Or85a*-GAL4 in the antennal lobe, with and without auxin. B) GFP fluorescence
quantification in the antennal lobe 24 hours after induction (4-6 replicates). Statistics performed using
Kruskal-Wallis test with Dunn pair-wise comparison (*, $p < 0.05$). C) GFP fluorescence driven by *grh*-
173 GAL4 in the larval ventral nerve cord, with and without auxin. D) GFP fluorescence quantification in
the ventral nerve cord 24 hours after induction (6 replicates). Statistics performed using Kruskal-Wallis
test with Dunn pair-wise comparison (**, $p < 0.01$). See Figure 4 - source data 1 for raw data.

Effects of auxin on development, adult survival and behaviour

For a universally applicable drug-inducible system, the drug should not impact on development or mortality of the flies. We tested whether continuous exposure to different concentrations of auxin (K-NAA) would impact on developmental timing. Time to pupation (from egg-laying) is unaffected for concentrations up to 5 mM, however, 10 mM caused a delay of approximately one day (Figure 5A). 10 mM auxin causes some developmental delay, although it has no impact on the survival of the flies through these developmental stages (compared to a no auxin control) (Figure 5B). These data indicate that 10 mM auxin should be avoided for larval induction, however, this is not an issue as > 1 mM is sufficient for robust induction of expression (Figure 5A) and 5 mM does not cause any developmental delay. Concentrations of 5 mM and 10 mM auxin can induce expression in adults (Figure 2 and Figure 2 – figure supplement 1). Survival assays on both male and female adults show that concentrations of 5 mM and 10 mM have no effect on survival and lifespan (Figure 5D and 5E). Therefore, in summary, up to 5 mM auxin is optimal for use in larvae, whilst up to 10 mM auxin is best for use in adults. We also tested whether auxin affects locomotion of larvae (crawling assay) and adults (climbing assay) and see no significant effects with 5 mM and 10 mM auxin, respectively (Figure 5 – figure supplement 1).

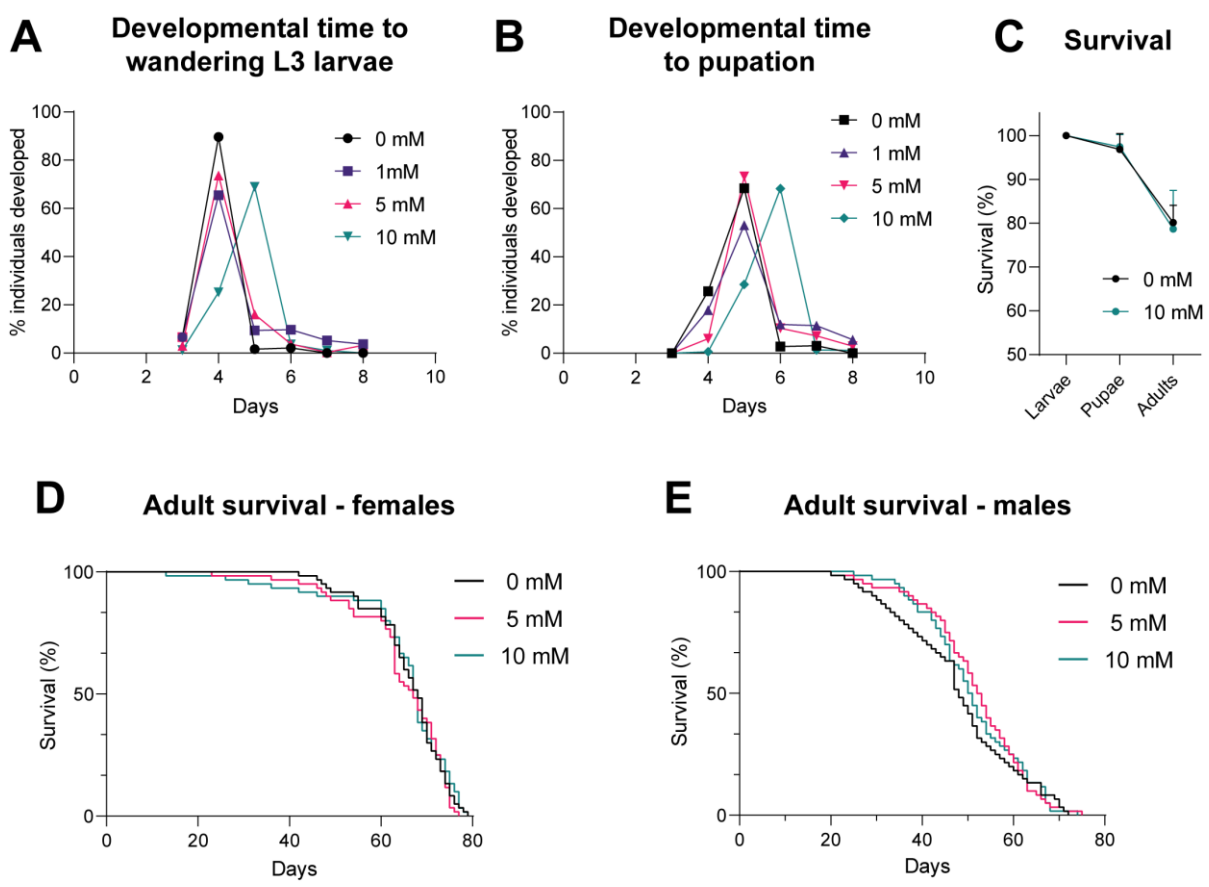


Figure 5. Impact of auxin on developmental timing and survival. A) Time taken from egg-laying to wandering L3 larvae. B) Time taken from egg-laying to pupation. C) Survival across developmental stages (larval to pupal and pupal to adult) with 10 mM auxin. D) Survival of adult females during continuous exposure to auxin. E) Survival of adult males during continuous exposure to auxin. Logrank test and weighted Gehan-Breslow-Wilcoxon model (ns) were used for the adult survival assays. See Figure 5 - source data 1 for raw data.

AGES allows inducible manipulation of *Drosophila* adult circadian locomotor rhythms.

To determine whether AGES is suitable for acute adult behavioural manipulation, we recapitulated a classic GeneSwitch manipulation of clock neuron excitability using AGES (Depetris-Chauvin et al., 2011). A key circuit for control of Drosophila rhythmic behaviour is the ventrolateral neuron (LNv) cluster, of which the small ventrolateral neurons (sLNvs) are both necessary and sufficient for maintenance of free-running locomotor rhythms (Grima et al., 2004; Renn et al., 1999; Stoleru et al., 2004). The four large LNvs (lLNvs) and four out of five small LNvs (sLNvs) express the neuropeptide pigment dispersing factor (PDF) (Helfrich-Forster, 1995; Renn et al., 1999). PDF is a key clock output neuropeotide required for synchronization of clock neuron groups, and loss of PDF function results in arrythmicity, desynchronization of clock oscillators and altered period length (Lear et al., 2009; Lin et al., 2004; Peng et al., 2003; Sheeba et al., 2008). Adult-specific silencing of *PDF*+ LNvs by expression of the inwardly rectifying potassium channel Kir2.1 using *PDF*-GAL4-GeneSwitch is sufficient to nearly ablate circadian locomotor rhythms in constant conditions without resetting the molecular clock (Depetris-Chauvin et al., 2011).

To recapitulate this behavioural experiment, we combined the *PDF*-GAL4 line with the AGES system to allow auxin-inducible control of Kir2.1 expression in *PDF*+ neurons. We simultaneously replicated GeneSwitch driven Kir2.1 expression in *PDF*+ neurons. After eclosion and entrainment to a 12-hour light: 12-hour dark (12:12 LD) schedule on standard fly food, flies were individually loaded into activity tubes and placed in the Drosophila Activity Monitoring (DAM) system (Trikinetics) for three days in 12:12 LD, followed by eight days in constant darkness (DD). Control activity tubes contained standard DAM food (2% agar, 5% sucrose) while experimental food contained RU-486 (200 mg/mL) or NAA (2 mM or 10 mM). In contrast to the previous experiments, NAA auxin was used here instead of K-NAA auxin. Analysis of the amplitude of the behavioural locomotor rhythm in constant darkness by fast Fourier transform (Plautz et al., 1997) showed robust inhibition of locomotor rhythms in both male and female experimental *PDF*-GAL4; AGES>UAS-Kir2.1 flies on 2 mM or 10 mM NAA (Figure 6A-D, Figure 6 – supplement 1A,B). 2 mM NAA was sufficient to completely ablate circadian locomotor rhythms in both male and female experimental flies (Figure 6C,D), without a significant effect on average 24-hour locomotor activity of parental controls (Figure 6 - supplement 2, A-D). 2 mM NAA feeding significantly lengthened the behavioural period of male *PDF*-GAL4/+; AGES/+ parental controls by 19 minutes, with no effect on females of the same genotype. In agreement with the classic study by Depetris-Chauvin *et al.* (Depetris-Chauvin et al., 2011), 200 μg/mL RU-486 feeding also reduced circadian locomotor rhythms in both sexes of flies with *PDF*-GAL4-GeneSwitch driven expression of Kir2.1 (Figure 6E,F), though the effect was not as strong as 2 mM NAA feeding in the AGES flies. In addition, RU-486 feeding resulted in a broader distribution of locomotor rhythm strengths and a net increase in rhythmicity in male *PDF*-GAL4-GeneSwitch parental control flies (Figure 6E), which was accompanied by a 51-minute increase period length and increased average 24-hour locomotor activity (Figure 6 – supplement 2G). RU-486 also increased the period of female PDF-Gal4-GeneSwitch parental controls by 46 minutes, with no effect on total

activity or rhythm strength (Figure 6F, Figure 6 – supplement 2F,H). In summary, we find that 2 mM NAA feeding is sufficient for AGES-induced expression of Kir2.1 in adult *PDF*+ clock neurons to ablate circadian locomotor behaviour. Indeed, we observed a stronger ablation of locomotor rhythmicity in the AGES system compared to GeneSwitch, with fewer off-target behavioural effects of NAA feeding of parental control flies.

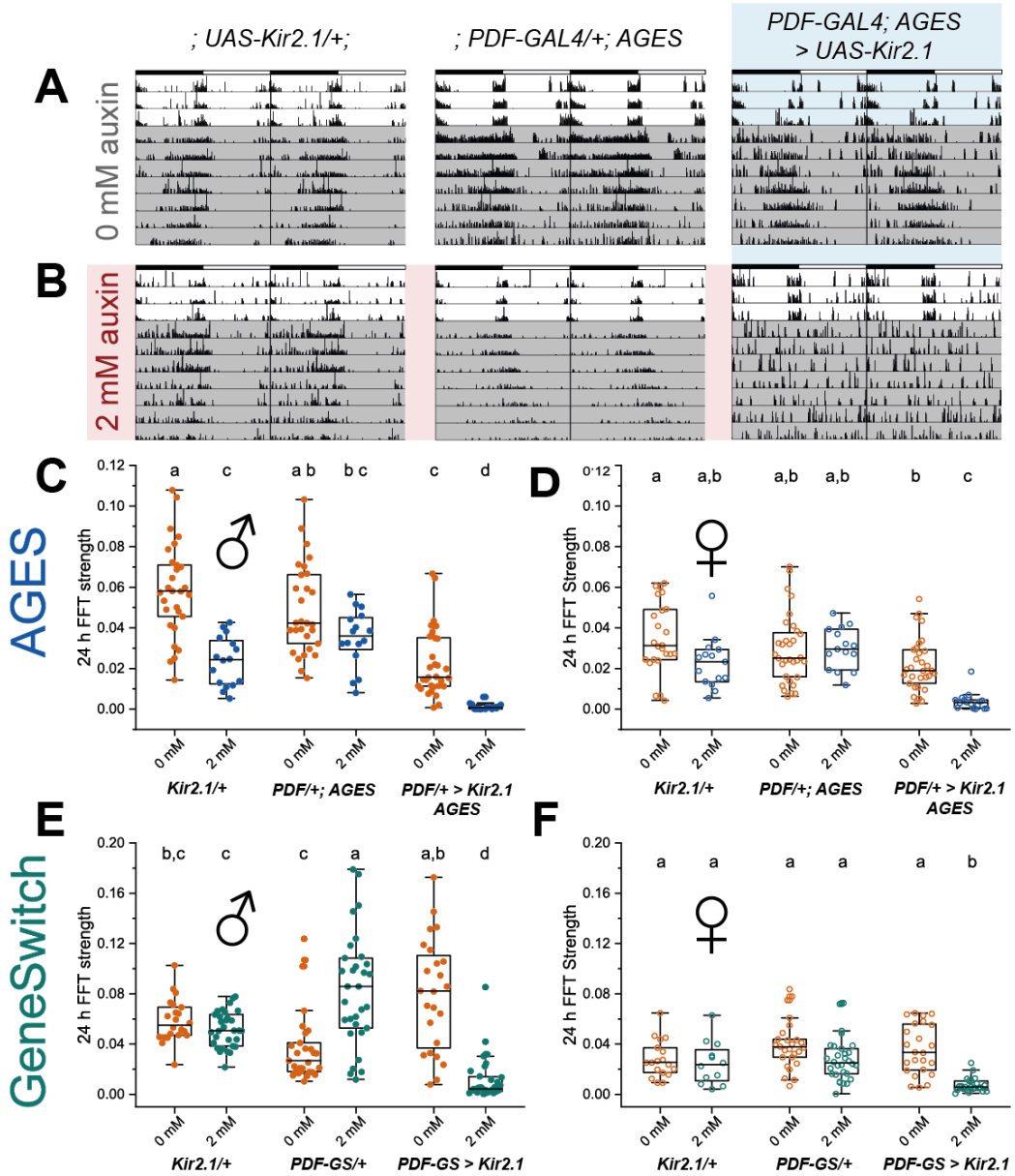


Figure 6. AGES induced expression of Kir2.1 in PDF+ clock neurons ablates circadian locomotor rhythms. A, B) Representative double-plotted actograms for 5-10 day old male flies maintained on standard food (A) or food supplemented with 2 mm NAA (B) for three days in 12:12 LD and 7 days in DD. Left: parental control *UAS*-Kir2.1, centre: parental control *PDF*-GAL4; AGES, right: experimental *PDF*-GAL4/*UAS*-Kir2.1; AGES/+. Bars indicate LD cycle, grey shaded days indicate constant darkness. C) Amplitude of circadian rest:activity rhythms on DD days 2-8 represented by FFT power at 24 h for male *PDF*-GAL4;AGES> *UAS*-Kir2.1 flies and their parental controls on standard food (orange) and food supplemented with 2 mM NAA (blue). Points represent individual flies, box shows 25-75% confidence interval, median line and outliers. D) 24 h FFT power as in (C) for female *PDF*-GAL4;AGES> *UAS*-Kir2.1 flies and their parental controls. E) 24 h FFT power for as in (C) male *PDF*-GAL4-Geneswitch>*UAS*-Kir2.1 flies and their

250 parental controls maintained on vehicle control food (orange) and food supplemented with 466 mM RU-486 (red). F)
251 24 h FFT power as in (E) for female *PDF*-GAL4-Geneswitch>*UAS*-Kir2.1 flies and their parental controls. For all panels,
252 means were compared by two-way ANOVA by genotype and food substrate, see Figure 6 – figure supplement 3 for *p*-
253 values. Means sharing the same letter are not significantly different from one another by Tukey's *post hoc* test ($p >$
254 0.05). See Figure 6 - source data 1 for raw data.

255 Discussion

256 The GAL4/UAS/GAL80^{ts} systems enable *Drosophila* researchers to control the expression of transgenes both
257 spatially and temporally. However, the impact of the required temperature shift on physiology and behaviour
258 to activate/deactivate the system, can affect the experimental results and subsequent conclusions (Abram
259 et al., 2017). Drug inducible systems have been developed as an alternative to the GAL80^{ts} method, however,
260 these are not ideal, limited by issues such as leaky expression, long induction times or incompatibility with
261 existing GAL4 lines. Here we present AGES, an auxin-inducible gene expression system, which eliminates the
262 need for temperature shifts to manipulate temporal expression and lacks many of the drawbacks of the
263 current drug-inducible systems.

264 A significant benefit of AGES over existing systems is its compatibility with most existing GAL4-driver lines.
265 Currently, the most prevalent drug-inducible expression systems, i.e. the GeneSwitch (Osterwalder et al.,
266 2001) and the Q-system (Potter et al., 2010), require the generation of their own driver lines. AGES uses a
267 GAL80 fusion protein, which allows the use of the system alongside the majority of existing GAL4-driver lines,
268 whereby suppression of the GAL4-*UAS* activation is achieved in all cells where auxin is absent. Notable
269 exceptions are split-GAL4 lines that do not use the GAL4 activation domain. As with all GAL80-based methods
270 of GAL4 control, the GAL4 line must contain the standard GAL4 activation domain for GAL80 to be affective.
271 There are split-GAL4 lines that use the GAL4 activation domain (Luan et al., 2006; Pfeiffer et al., 2010),
272 however, many use a p65 activator domain (Dionne et al., 2018; Tirian and Dickson, 2017), or a VP16 activator
273 domain (Luan et al., 2006), which will not work with AGES. We have demonstrated the system using multiple
274 driver lines, including *c564*-GAL4, *elav*-GAL4, *grh*-GAL4, *Or85a*-GAL4 and *PDF*-GAL4. These were, in part, used
275 to assess whether the auxin hormone used (K-NAA) could freely access various cell-types within the fruit fly,
276 particularly the central nervous system where auxin must pass the selectively-permeable glial membrane,
277 akin to the mammalian blood-brain barrier (Limmer et al., 2014). Our results, using one fat body GAL4 driver
278 and four different central nervous system GAL4 drivers, demonstrate that auxin can transverse the blood-
279 brain barrier, and that this system should, in theory, be compatible with all other GAL4 lines that contain a
280 GAL4 activation domain.

281 Successful gene driver systems require negligible expression when un-induced. While some current systems
282 are known to exhibit leaky expression (Scialo et al., 2016), AGES demonstrates an almost complete repression
283 of GAL4 activity when un-induced. Using GFP fluorescence as a readout, 0 mM auxin does not show any
284 greater signal than the negative control (Figure 2B, Figure 4). Also, the level of *GFP* mRNA without auxin

285 showed no significant difference when compared to the negative control (Figure 2C). The AID tags on GAL80
286 allow the rapid, and tuneable, degradation of the GAL80 protein, providing users with the option to
287 determine the strength of expression by adjusting the concentration of auxin in the food, or by limiting the
288 exposure time of the flies to auxin.

289 Temporal control of gene expression is essential for many behavioural experiments in *Drosophila*. We
290 demonstrate that AGES can elicit strong behavioural changes through activating expression of Kir2.1 (an
291 inward-rectifier potassium ion channel) in *PDF*-GAL4 expressing adult neurons (Figure 6). Kir2.1 inhibits the
292 activity of PDF neurons resulting in ablation of circadian rhythms. The strong loss of circadian rhythm was
293 observed in both males and females, and even worked at the lower concentration of 2 mM auxin. This is an
294 important development for the *Drosophila* behaviour field, as it allows experiments to be done with existing
295 GAL4 lines, and without the need of a temperature shift or having to use GeneSwitch (Osterwalder et al.,
296 2001). While 2 mM auxin food had no effect on the circadian rhythm of female flies (Figure 6D) or the *PDF*-
297 GAL4/+ control male flies (Figure 6C), it did impact on the *UAS*-Kir2.1/+ control male flies. We currently have
298 no explanation for this. However, it does emphasise the requirement to always perform the appropriate
299 controls. Also, in this context, 10 mM auxin food did have an effect on both male control lines (Figure 6 –
300 Supplement 1A). So, while 10 mM auxin had no impact on adult climbing (Figure 5 – figure supplement 1B),
301 it did impact on circadian rhythms. It should be noted that NAA was used for the circadian rhythm
302 experiment, whereas K-NAA (more water soluble) was used for all the other experiments. Therefore, K-NAA,
303 may have less of an effect on the controls as was seen with NAA.

304 AGES is also a system that is safe for both humans and flies alike. Auxin hormone carries no hazardous
305 concerns for human handling, and no lethal effects were identified for the effective concentrations of 10 mM
306 or below for both adult and larval stages (Figure 5). Continuous exposure of larvae to 10 mM auxin does
307 result in some developmental delay; however, it does not affect survival. Higher concentrations of auxin
308 result in increased mortality rates (data not shown) and should be avoided. Users are encouraged to consider
309 this if attempting to optimise exposure conditions for specific experiments, or GAL4 driver lines that have
310 not been assessed in this paper. Overall, based on our data, we recommend the use of up to 5 mM auxin and
311 up to 10 mM for optimal induction in larvae and adults, respectively.

312 It is also paramount that inducible systems are easy and cheap to apply, to facilitate their practical use in
313 diverse experimental settings. While expenses of the GeneSwitch system average £0.31 (GBP) per vial, AGES
314 costs ~120x less at £0.0025 per vial (authors' estimate). Furthermore, the water-soluble auxin (K-NAA) can
315 be easily added to the food immediately before the food is poured into vials or bottles, requiring no additional
316 safety measures, enabling ease of production. In our experience, auxin-containing food can be stored 4 °C
317 for up to 15 weeks (Figure 2 – figure supplement 2B) where the hormone's potency still persists. Further
318 studies are required to assess the stability and effectiveness of auxin when stored for longer periods of time,
319 or in differing conditions.

320 The sensitivity of the system is directly related to the amount of auxin present within the tissue, and thus
321 concerns are raised regarding the fly's age and sex, as these factors impact feeding rates (Carey et al., 2006;
322 Wong et al., 2009). As all experimental adult flies were aged to 5 days post-eclosion, the effects of age on the
323 system are yet to be determined, although lower expression is to be expected in older flies. To address these
324 concerns, alterations to the auxin application could be made, such as a longer exposure (> 24 hours), which
325 as we have shown does not impact on survival (Figure 5D and 5E).

326 Despite the various advantages that AGES offers the fly community, there are still limitations to consider.
327 These include i) the response time of the system (18-24 hours), ii) auxin can have subtle effects on adult
328 behaviour (circadian rhythms) and iii) induced levels of expression are not always as high as the maximum
329 levels obtained without the system. For example, as male adults eat less than their female counterparts,
330 lower expression of the transgene is observed in males (Figure 2 – figure supplement 1). However, as GAL4
331 induced expression is inherently strong, reduced transgene expression may not be an issue. This is evident
332 in our circadian rhythm behavioural experiment, where there is a robust response in both males and females
333 when driving expression of *UAS*-Kir2.1 (Figure 6C and 6D). While a 24-hour induction time is not instant, it is
334 both practical and convenient for researchers performing experiments on adult flies in the lab.

335 There is scope for AGES to be optimised in the future. One approach would be to utilise a mutated version
336 of TIR1 that can accommodate bulky analogues of auxin. Such mutants, in combination with modified auxins
337 (5-Ph-IAA or 5-adamantyl-IAA) allow the degron system to work with much lower levels of auxin (>500-fold
338 less) (Yesbolatova et al., 2020; Zhang et al., 2022) and demonstrate a more rapid degradation. Efforts to test
339 these modifications with AGES are underway.

340 In summary, AGES offers the fly community a cheap, safe, and easy system for temporal transgene expression
341 using existing GAL4 lines. Furthermore, it does not require a shift in temperature changes for induction of
342 gene expression. It is a particularly promising tool for research fields where researchers want to avoid using
343 temperature shifts (e.g. aging, behavioural genetics and neuropathology), and will undoubtedly have wide-
344 ranging benefits for multiple fields of study.

345

346 ## Materials and Methods

347 ### Generation of *AGES* line

348 The *AtTIR1-T2A-AID-GAL80-AID* sequence was synthesized by Twist Biosciences (twistbioscience.com)
349 (Figure 1 – Supplement 1). The *GAL80* sequence (from *S. cerevisiae*) and the *AtTIR1* sequence (Zhang et al.,
350 2015) were codon optimized for *Drosophila* for more effective translation. The *αTub84B* promoter was
351 amplified from a *tubulin-eGFP* plasmid (gift from M. Dionne) using the following primers: tub_FWD:
352 GATATCAAGCTTGCACAGGTCC and tub-RV: GTACCTTCACGCTGTGGATGAGG. The *αTub84B* and *AtTIR1-T2A-*
353 *AID-GAL80-AID* sequences were cloned into *pattB* (Bischof et al., 2007) using Gibson assembly (Gibson et al.,

354 2009). Successful clones were sequenced verified. Annotated sequence is in Figure 1 – figure supplement 2.
355 Microinjection was performed by *Cambridge Fly Facility* using the VK00040 line, which has an *attB* site at
356 location att3B (87B10) on chromosomal arm 3R. Injected adult males were collected and mated to w^{1118} virgin
357 females to identify transgenics (orange eyes). *tub*-TIR1-T2A-AID-GAL80-AID is available at the Bloomington
358 Drosophila Stock Center (stock #92470) or the VDRC Stock Center (stock #311020).

359 **Fly stocks and food**

360 In this study *c564*-GAL4*, UAS*-nls-GFP */ CyO-actin*-GAL4-GFP flies were used to perform fluorescent reporter
361 experiments in the adult and larval fat body, and for quantification of *GFP* mRNA expression. Moreover, these
362 flies were used in adult survival assays. *CantonS* flies were used in the developmental survival and
363 developmental timeline experiments. For fluorescent reporter experiments in the adult brain we used *elav*-
364 GAL4*, UAS*-nls-GFP. For behavioural experiments, *Iso31* flies were used as a background strain (Ryder et al.,
365 2004), and we used *PDF-Gal4* (Park et al., 2000) and *PDF*-GeneSwitch-GAL4 (BDSC 81116) to drive expression
366 of *UAS*-Kir2.1 (BDSC 6597). Flies were kept at 25 °C on and standard *Drosophila* food (recipe in supplementary
367 material) was supplemented (just before being aliquoted into vials/bottles/plates) with auxin (K-NAA (1-
368 Naphthaleneacetic acid potassium salt) available from Phytotech (#N610) or Glentham Life Sciences
369 (GK2088)) at varying concentrations. For behavioral experiments, flies were raised and entrained on standard
370 food, and transferred to activity monitoring tubes containing 2% agar, 5% sucrose, supplemented with
371 indicated concentrations of RU-486 in ethanol (Mifepristone, Sigma-Aldrich) or NAA (Phytotech, #N600).

372 **Immunohistochemistry**

373 Adult and larval brains were dissected in 1x PBS and fixed for 25 minutes at room temperature in 4%
374 formaldehyde (methanol free) in 0.3% Triton X-100 PBS (PBST). They were washed four times for 1 hour with
375 0.3% PBST. Normal Goat Serum (2 % in PBST) was used for tissue blocking (RT, 15 min- 1 hour) and subsequent
376 overnight primary antibody incubation. All tissue washes were done in PBST.
377 Primary antibodies used were chicken anti-GFP (Abcam #13970, 1:2000), guinea pig anti-Dpn 1:10,000
378 (Caygill and Brand, 2017) and rat anti-Elav 1:500 (Developmental Studies Hybridoma Bank, DSHB). Secondary
379 antibodies used include Alexa Fluor® 488, 545 and 633 at a concentration of 1:200 (Life technologies) and
380 tissue was incubated for 1.5 hours at room temperature. Tissue was mounted on standard glass slides in
381 Vectashield Mounting medium (Vector Laboratories). Brains were imaged using a Zeiss LSM 510 microscope.
382 Analysis of acquired images was done using Fiji (Schindelin et al., 2012).

383 **Imaging and Image Analysis**

384 Live imaging of GFP in adult flies and larvae was performed using a Nikon SMZ 1500 microscope. In larvae, to
385 identify earliest auxin-induction effects, animals were then placed on 1 mM auxin- supplemented food and

386 imaged at 0, 1, 5, 10 and 25 hour intervals. Adults were placed on food containing varying auxin
387 concentrations for 24 hours prior to imaging.
388 Live GFP levels were quantified in 6 animals per condition (including negative and positive controls). Binary
389 images were created by using a threshold (Outsu's thresholding) and the Wand Tool in Fiji was used to trace
390 the GFP-positive area as region of interest (ROI). The ROI corresponded to abdomens in adults or whole
391 larvae. Mean pixel intensity per ROI was calculated using the Measure plugin in Fiji for a total of 6 biological
392 replicates. Statistical significance analysed using One-way ANOVA (normally distributed) or Kruskal-Wallis
393 test with Dunn pair-wise comparison (non-normally distributed). Parametric t-test or non-parametric
394 Wilcoxon tests were used to compare individual conditions with respective controls. Statistical tests were
395 completed in *R* (v3.6.3)and plots were performed using the software GraphPad Prism version 9 for Windows.

396 **Developmental and Survival Assays**

397 All animals were kept at 25 °C. To determine the effects of auxin on larval development, 10 mated Canton-S
398 flies were allowed to lay eggs on fly food containing 0, 1, 5, or 10 mM of auxin for a maximum of 24 hours,
399 or until ~50 eggs were counted. 5 replicates were obtained for each concentration, except for 0 mM where
400 4 replicates were used. Daily emergence of L3 larvae, and presence of pupae were recorded to determine
401 the time required for egg-to-L3, and egg-to-pupae development for each concentration.

402 Adult survival assays were performed on *c564*-GAL4*, UAS*-nls-GFP */ AGES* flies on three replicates, with 20
403 flies per replicate. Male and female animals were separated 5 days post-eclosion. Each replicate was
404 continuously exposed to 0 mM, 5mM and 10 mM Auxin food. Death was scored daily until all flies were
405 deceased. Statistical tests and plots were performed using the software GraphPad Prism version 9 for
406 Windows.

407 **RNA extraction and RT qPCR**

408 In order to quantify whether levels of *GFP* mRNA expression changed in the presence of Auxin, the *c564*-
409 GAL4*, UAS*-nls-GFP/ AGES flies were placed on 0 mM, 5 mM and 10 mM auxin-supplemented food for 24 hrs.
410 The *c564*-GAL4*, UAS*-nls-GFP*/ +* flies were used as a positive control whilst the *AGES* flies were used as a
411 negative control (lacking both the GAL4 driver and the *UAS*-nls-GFP). The 0 mM was included to investigate
412 whether there was any leaky GAL4 activity in absence of auxin.

413 Total RNA was extracted from whole adult flies, 3 per replicate per condition, using a standard TRIzol ®
414 extraction protocol (Scientific, 2016). DNA was degraded using RNAse-free DNAse (Thermo Scientific) and
415 cDNA synthesis was performed using the iScript™ cDNA synthesis kit (Bio-Rad), following manufacturer's
416 instructions. Real time quantitative PCR was performed with iTaq™ Universal SYBR® Green Supermix (Bio-
417 Rad) using the StepOnePlus™ Real-Time PCR System (Applied Biosystems).

418 The gene *RpL4* (*Ribosomal Protein L4*) was used as reference gene. The *GFP* mRNA expression levels were
419 calculated using the ΔΔCt method (ΔCt = Ct [mean Ct of reference gene] – Ct [target]; ΔΔCt = ΔCt [target] –
420 mean ΔCt [control]) (Livak and Schmittgen, 2001). Target refers to the auxin concentrations (0mM, 5mM,
421 10mM) and control refers to the *AGES* flies. Primers used: RpL4_FW - 5'-TCCACCTTGAAGAAGGGCTA-3',
422 RpL4_RV – 5'-TTGCGGATCTCCTCAGACTT-3', GFP_FW – 5'-GAGCTGTACAAGAGCAGGCA-3', GFP_RV – 5'-
423 GTTGACGGCGTTTTCGTTCA-3'. Statistical significance analysed Kruskal-Wallis test with Dunn pair-wise
424 comparison in *R* (v3.6.3).

425

426 ***Drosophila* activity monitoring assay**

427 We evaluated circadian locomotor rhythms using the Drosophila activity monitoring (DAM) system
428 (Trikinetics). Male and female flies of the indicated genotype were entrained in a 12 h light: 12 h dark cycle
429 prior to loading individual 5-7 day old flies into glass DAM tubes containing control or experimental diets.
430 For AGES flies, the control diet consisted of standard DAM food. For GeneSwitch flies, the control diet
431 consisted of 1% ethanol vehicle in standard DAM food. Flies were monitored via DAM assay for three days
432 in 12:12 LD at 25° C, followed by eight days in constant darkness (DD). Circadian locomotor parameters
433 were analyzed using ClockLab software (Actimetrics) for data from days 2-8 of DD. Period length was
434 determined by χ^2 periodogram analysis, and relative rhythm power at 24 h was determined using fast
435 Fourier transform (FFT). FFT power and average daily activity counts for each sex were compared by two-
436 way ANOVA by genotype and food substrate, with Tukey's *post hoc* comparisons. Because period length
437 could not be estimated for arrhythmic flies, period length was compared by Student's *t*-test for flies of the
438 same genotype on different food substrates.

439

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450

451 ## Figure Legends

452 **Figure 1. Design and action of AGES**. A tubulin promoter ubiquitously expresses the bicistronic TIR1 and
453 AGES cassette. AID-tagged GAL80 is cleaved from the TIR1 due to the presence of the T2A sequence. In the
454 absence of auxin, GAL80 can inhibit GAL4 activity, whilst the presence of auxin induces degradation of
455 GAL80, allowing GAL4 to drive the expression of *UAS*-transgene(s).

456 **Figure 2. AGES effectively induces GAL4 activity in *Drosophila* adults**. A) Ventral images of live females
457 that express GAL4 in fat body tissue. Ingestion of food containing auxin (>= 5 mM for 24 hours) induces
458 GAL4 activity and the expression of GFP. B) Quantification of GFP levels. Pixel intensity thresholding was
459 performed to isolate abdomens as regions of interest. The average pixel intensities from 6 replicates were
460 quantified and analysed using Kruskal-Wallis test with Dunn pair-wise comparison (***, $p < 0.001$ and **, p
461 <0.01). C) qPCR data for *GFP* mRNA levels using different concentrations of auxin (3 biological replicates).
462 Values were normalised to housekeeping gene *RpL4* (*Ribosomal Protein L4*) and relative expression levels
463 (compared to the negative control) were calculated using the ΔΔCt method. Y-axis displaying ΔΔCt values
464 and statistics done using Kruskal-Wallis test with Dunn pair-wise comparison (*, $p < 0.05$). See Figure 2 -
465 source data 1 for raw data.

466 **Figure 3. AGES allows induction of GAL4 activity in *Drosophila* larvae**. A) GFP fluorescence quantification
467 in fat body tissue after induction (for 24 hours) on food with different concentrations of auxin. The average
468 pixel intensities from 6 larvae were quantified and analysed using Kruskal-Wallis test with Dunn pair-
469 wise comparison (***, $p < 0.001$ and **, p <0.01). B) Time course of GFP expression in fat body tissue when
470 using 5 mM auxin. The average pixel intensities from 6 larvae were quantified and analysed using
471 Kruskal-Wallis test with Dunn pair-wise comparison (**, p <0.01) C) Representative images of larvae fed on
472 different concentrations of auxin. D) Representative images of larvae imaged at each time interval since
473 induction. See Figure 3 - source data 1 for raw data.

474 **Figure 4. AGES allows induction of GAL4 activity in the *Drosophila* adult and larval brain.** A) GFP
475 fluorescence driven by *Or85a*-GAL4 in the antennal lobe, with and without auxin. B) GFP fluorescence
476 quantification in the antennal lobe 24 hours after induction (4-6 replicates). Statistics performed using
477 Ordinary one-way ANOVA (*, $p < 0.05$). C) GFP fluorescence driven by *grh*-GAL4 in the larval ventral nerve
478 cord, with and without auxin. D) GFP fluorescence quantification in the ventral nerve cord 24 hours after
479 induction (6 replicates). Statistics performed using Ordinary one-way ANOVA (*, $p < 0.05$). See Figure 4 -
480 source data 1 for raw data.

481 **Figure 5. Impact of auxin on developmental timing and survival.** A) Time taken from egg-laying to
482 wandering L3 larvae. B) Time taken from egg-laying to pupation. C) Survival across developmental stages
483 (larval to pupal and pupal to adult) with 10 mM auxin. D) Survival of adult females during continuous
484 exposure to auxin. E) Survival of adult males during continuous exposure to auxin. Logrank test and

485 weighted Gehan-Breslow-Wilcoxon model (ns) were used for the adult survival assays. See Figure 5 - source
486 data 1 for raw data.

487 **Figure 6. AGES induced expression of Kir2.1 in PDF+ clock neurons ablates circadian locomotor rhythms.**
488 A, B) Representative double-plotted actograms for 5-10 day old male flies maintained on standard food (A)
489 or food supplemented with 2 mm NAA (B) for three days in 12:12 LD and 7 days in DD. Left: parental
490 control *UAS*-Kir2.1, centre: parental control *PDF*-GAL4; AGES, right: experimental *PDF*-GAL4/*UAS*-Kir2.1;
491 AGES/+. Bars indicate LD cycle, grey shaded days indicate constant darkness. C) Amplitude of circadian
492 rest:activity rhythms on DD days 2-8 represented by FFT power at 24 h for male *PDF*-GAL4;AGES> *UAS*-
493 Kir2.1 flies and their parental controls on standard food (orange) and food supplemented with 2 mM NAA
494 (blue). Points represent individual flies, box shows 25-75% confidence interval, median line and outliers. D)
495 24 h FFT power as in (C) for female *PDF*-GAL4;AGES> *UAS*-Kir2.1 flies and their parental controls. E) 24 h FFT
496 power for as in (C) male *PDF*-GAL4-Geneswitch>*UAS*-Kir2.1 flies and their parental controls maintained on
497 vehicle control food (orange) and food supplemented with 466 mM RU-486 (red). F) 24 h FFT power as in
498 (E) for female *PDF*-GAL4-Geneswitch>*UAS*-Kir2.1 flies and their parental controls. For all panels, means
499 were compared by two-way ANOVA by genotype and food substrate, see Figure 6 – figure supplement 3 for
500 *p*-values. Means sharing the same letter are not significantly different from one another by Tukey's *post*
501 *hoc* test ($p > 0.05$). See Figure 6 - source data 1 for raw data.

502

503 **Figure 1 – figure supplement 1.** Plasmid map of the AGES plasmid. Map of the *pattB-tubP-AtTIR1-P2A-*
504 *miniAID-Gal80-miniAID-SV40* plasmid (generated using Snapgene). The full sequence is in Figure 1 – figure
505 supplement 2.

506 **Figure 1 – figure supplement 2.**

507 Genbank (.gb) sequence file for *pattB-tubP-AtTIR1-P2A-miniAID-Gal80-miniAID-SV40*

508 **Figure 2 – figure supplement 1.**

509 **Figure 2 – figure supplement 1. AGES effectively induces GAL4 activity in *Drosophila* adult males**. A)
510 Ventral images of live males that express GAL4 in fat body tissue. Ingestion of food containing auxin (24
511 hours) induces GAL4 activity and the expression of GFP. B) Quantification of GFP levels (from 6 male
512 abdomens). Pixel intensity thresholding was performed to isolate abdomens as regions of interest. The
513 average pixel intensity intensities were quantified and analysed using Ordinary one-way ANOVA (**, $p =$
514 0.002, ****, $p < 0.0001$). C) qPCR data for *GFP* mRNA levels using different concentrations of auxin (3
515 biological replicates). Values were normalised to housekeeping gene *RpL4* (*Ribosomal Protein L4*) and
516 relative expression levels were calculated using the ΔΔCt method. Y-axis displaying ΔΔCt values and
517 statistics done using Ordinary one-way ANOVA (**, $p < 0.005$).

Figure 2- figure supplement 2. On-off dynamics of AGES in adult flies and stability of auxin fly food. A) Quantification of GFP levels in female abdomens (expressed in the fat body). Pixel intensity thresholding was performed to isolate abdomens as regions of interest. The average pixel intensity intensities were quantified and analysed using Ordinary one-way ANOVA (*, $p < 0.05$, ****, $p < 0.0001$). B) Quantification of GFP levels in female abdomens with freshly made auxin food and 15-week old food.

Figure 4 – figure supplement 1. AGES allows induction of pan-neuronal GAL4 activity in the adult brain. Confocal images of adult brains stained with anti-GFP and anti-Elav. Adult were fed food containing 5 mM auxin for 24 hours. Scale bars represent 20 µm.

Figure 5– figure supplement 1. Working concentrations of auxin do not impact locomotor function of wild type *Drosophila*. A) Larval crawling speed on 0 mM food (10 larvae) and 5 mM auxin food (10 larvae). B) Distance climbed in climbing assay for males and females on 0 mM food (5 separate vials and a total of 144 flies for males and 130 flies for females) and 10 mM auxin food (5 separate vials and a total of 141 flies for males and 141 flies for females). Analysed using a paired t-test.

Figure 6 – Supplement 1. Dose-dependent NAA effects on behaviour of AGES parental controls. Behavioural data for *PDF*-GAL4; AGES> *UAS*-Kir2.1 flies and their parental controls on standard food (orange), 2 mM NAA (blue) or 10 mM NAA (purple). A, B) 24-hour FFT power on DD days 2-8 for male (A) and female (B) *PDF*-GAL4; AGES> *UAS*-Kir2.1 flies. 2 mM NAA data is replotted from Fig. 6. Means were compared by two-way ANOVA by genotype and food substrate. Means sharing the same letter are not significantly different from one another by Tukey's *post hoc* test ($p > 0.05$). C, D) Period length for male (C) and female (D) flies. Means were compared by Student's *t*-test for flies of the same genotype on different food substrates. E, F) Average 24-hour activity counts for male (E) and female (F) flies. Means were compared by two-way ANOVA by genotype and food substrate. Means sharing the same letter are not significantly different from one another by Tukey's *post hoc* test ($p > 0.05$).

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