1 **Original Research Article**
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3 # Independent origins of the Golgi phosphoprotein 3 oncoprotein gene

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Abstract

Golgi phosphoprotein 3 (GOLPH3) is considered the first oncoprotein of the Golgi apparatus. It was identified as an evolutionarily conserved protein upon its discovery about 20 years ago, but its function remains puzzling in normal and cancer cells. The GOLPH3 gene is part of a group of genes that also includes the GOLPH3L gene. Because cancer has deep roots in multicellular evolution, studying the evolution of the GOLPH3 gene family in non-model species represents an opportunity to identify new model systems that could help better understand the biology behind this group of genes. The main goal of this study is to explore the evolution of the GOLPH3 gene family in birds as a starting point to understand the evolutionary history of this oncoprotein. We identified a repertoire of three GOLPH3 genes in birds. We found duplicated copies of the GOLPH3 gene in all main groups of birds other than paleognaths, and a single copy of the GOLPH3L gene. We suggest there were at least three independent origins for GOLPH3 duplicates. Amino acid divergence estimates show that most of the variation is located in the N-terminal region of the protein. Our transcript abundance estimations show that one paralog is highly and ubiquitously expressed, and the others were variable. Our results are an example of the significance of understanding the evolution of the GOLPH3 gene family, especially for unraveling its structural and functional attributes.

1   Introduction

Golgi phosphoprotein 3 (GOLPH3) is a highly conserved protein of the Golgi apparatus (Wu et al., 2000; Bell et al., 2001) considered the first oncoprotein of this subcellular compartment (Sechi et al., 2015). The GOLPH3 gene family comprises the conserved GOLPH3 gene and the GOLPH3L gene found only in vertebrates (Wu et al., 2000; Bell et al., 2001). Despite the vast amount of empirical evidence demonstrating the contribution of GOLPH3 to tumorigenesis and cancer, a full understanding of its molecular role has not yet emerged. This is mainly due to multiple functions attributed to GOLPH3 (Sechi et al., 2015), including, the sorting of Golgi glycosyltransferases (Ali et al., 2012; Isaji et al., 2014; Pereira et al., 2014), the modulation of focal adhesion dynamics (Arriagada et al., 2019), induction of membrane curvature (Rahajeng et al., 2019), and an intriguing function for a Golgi protein, regulating mitochondrial function (Nakashima-Kamimura et al., 2005). Because cancer has deep evolutionary roots that arise as a consequence of the multicellularity (Boddy et al., 2020), and is widespread across animals (Albuquerque et al., 2018), studying the evolution of the GOLPH3 gene family in non-model species can provide significant information for a comparative oncology approach, which is emerging as an integrative field to tackle cancer (Boddy et al., 2020).

The availability of whole-genome sequences opens an opportunity to understand the evolution of gene families. The annotation of gene repertoires in different species has revealed that copy number variation is an important source of variability that should be considered when making functional comparisons (Abegglen et al., 2015; Sulak et al., 2016). Phylogenetic reconstructions show that the evolution of gene families follows complex pathways, including gene gain and losses and independent origins, making it challenging to perform direct interspecies comparisons. Thus, understanding the variability of gene repertoires and their duplicative history represents an essential piece of information to understand the biological functions associated with a group of genes and make biologically meaningful comparisons. Today, the GOLPH3 gene family is viewed as a group of genes containing two paralogs (GOLPH3 and GOLPH3L) with 1:1 orthologs among most vertebrate species (Wu et al., 2000; Bell et al., 2001). Among amniotes, it is suggested that birds are less susceptible to cancer in contrast to mammals (Ratcliffe, 1933; Lombard and Witte, 1959; Effron et al., 1977; Pang et al., 2012; Møller et al., 2017; Boddy et al., 2020); although this information should be taken with caution given sampling bias (Hochberg and Noble, 2017). Thus, the study of genes associated with cancer in birds could provide clues about the genetic bases associated with this difference and suggest additional model systems that could help to understand the biology of the GOLPH3 gene family.

The main goal of this study was to understand the evolutionary history of the GOLPH3 gene family in birds. We took advantage of whole genome sequences in representative species of all main lineages of birds to understand the evolutionary pathways that gave rise to GOLPH3 paralogs. According to our assessment, we identified a repertoire of three GOLPH3 genes in birds. We found duplicated copies of the GOLPH3 gene in all main groups of birds other than paleognaths, and a single copy of the GOLPH3L gene that would be derived from the common ancestor of all birds. In the case of the GOLPH3 gene, our gene tree suggests at least three independent origins for the duplicated copies, in the ancestor of Galliformes and Anseriformes, in the ancestor of Anseriformes, and the ancestor of Neoaves. Divergence estimates between duplicated genes showed that most of the variation is located in the N-terminal region of the protein. Our transcript abundance estimations showed that one paralog was highly and ubiquitously expressed, while the others were variable. Our evolutionary analyses suggest a more

96   complex than anticipated evolutionary history of the GOLPH3 gene family, a scenario that could
97   have implications for cancer.
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2   Material and Methods
2.1   DNA sequences and phylogenetic analyses
We performed searches for GOLPH3 sequences in avian genomes in the National Center for
Biotechnology Information (NCBI) (Sharma et al., 2018) and the Ensembl v.102 databases (Yates
et al., 2020). We retrieved orthologs and paralogs from the NCBI (Sharma et al., 2018) using the
chicken (*Gallus gallus*), zebra finch (*Taeniopygia guttata*), and mallard (*Anas platyrhynchos*)
sequences using the program blast (blastn) (Altschul et al., 1990) against the non-redundant
database (nr) with default parameters. Additionally, we also retrieved sequences from the Ensembl
v.102 database (Yates et al., 2020). In cases where sequences are not complete we manually
annotated them. To do so, we first identified the genomic fragment containing the GOLPH3 gene
in Ensembl v.102 (Yates et al., 2020) or NCBI databases (Geer et al., 2010). Once identified,
genomic fragments were extracted, including flanking genes. After extraction, we manually
annotated GOLPH3 genes by comparing known exon sequences from a species that share a
common ancestor most recently in time to the species of which the genomic piece is being
annotated using the program Blast2seq v2.5 (Tatusova and Madden, 1999) with default
parameters. Accession numbers and details about the taxonomic sampling are available in
Supplementary Table S1.
We performed separate phylogenetic analyses for GOLPH3 and GOLPH3L paralogs.
Amino acid sequences were aligned using MAFFT v.7 (Katoh and Standley, 2013), allowing the
program to choose the alignment strategy (L-INS-i in both cases). Nucleotide alignments were
generated using the amino acid alignments as templates using the software PAL2NAL (Suyama et
al., 2006). We used the proposed model tool of IQ-Tree v.1.6.12 (Kalyaanamoorthy et al., 2017)
to select the best-fitting model of codon substitution, which selected MGK+F1X4+R3 for
GOLPH3 and MGK+F3X4+G4 for GOLPH3L. This approach uses a more realistic description of
the evolutionary process at the protein-coding sequence level by incorporating the genetic code
structure in the model. We used the maximum likelihood method to obtain the best trees using the
program IQ-Tree v1.6.12 (Trifinopoulos et al., 2016). We assessed support for the nodes using
three strategies: a Bayesian-like transformation of aLRT (aBayes test) (Anisimova et al., 2011),
SH-like approximate likelihood ratio test (SH-aLRT) (Guindon et al., 2010) and the ultrafast
bootstrap approximation (Hoang et al., 2018). In each case (GOLPH3 and GOLPH3L), we carried
out 25 independent runs to explore the tree space, and the tree with the highest likelihood score
was chosen. In both cases, GOLPH3 and GOLPH3L sequences from crocodiles and turtles were
used as outgroups (Supplementary Table S1).
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2.2   Molecular Evolution analysis
To measure variation in functional constraint among the GOLPH3L genes and to test for evidence
of positive selection, we estimated the omega parameter (d_N/d_S), using a maximum-likelihood
approach (Goldman and Yang, 1994) implemented in the CODEML module of the program
PAML v.4.8a (Yang, 2007). We implemented branch-site models, which explore changes in the
omega parameter for a set of sites in a specific branch of the tree to assess changes in their selective
regime (Yang and dos Reis, 2011). In this case, we conducted two separate analyses. In the first,
the ancestral branch of Galliformes was labeled as the foreground branch, while in the second, the
branch leading to manakins was labeled as a foreground branch. We compared the modified model

A (Yang et al., 2005; Zhang et al., 2005; Yang and dos Reis, 2011), in which some sites are allowed
to change to an omega value > 1 in the foreground branch, with the corresponding null hypothesis
of neutral evolution using a Likelihood Ratio Test (LRT). Three starting omega values (0.5, 1, and
2) were used to check the existence of multiple local optima. The Bayes Empirical Bayes (BEB)
method was used to identify sites under positive selection (Nielsen and Yang, 1998; Yang et al.,
2000).

2.3   Secondary structure and disordered region prediction and protein structure
homology modeling
Secondary structure and disordered region prediction was performed using the PredictProtein
server (https://predictprotein.org/)(Yachdav et al., 2014). Multiple sequence alignment was
performed using MAFFT v.7 server (https://mafft.cbrc.jp/alignment/server/) with default
parameters (Katoh et al., 2019). Multiple sequence alignment editing was performed using Jalview
software v.2.11.1.3 (Waterhouse et al., 2009). Protein structure homology modeling was
performed using the SWISS-MODEL server (https://swissmodel.expasy.org/)(Waterhouse et al.,
2018). Structural figures were prepared with PyMOL Molecular Graphics System, Version 2.0.6
Schrödinger, LLC.

2.4   Transcript abundance analyses
GOLPH3 transcript abundance was measured in chicken (*Gallus gallus*) and mallard (*Anas*
platyrhynchos). We collected three RNASeq libraries from brain, heart, intestine, kidney, liver,
lung, spleen, ovary, and testis from each species gathered from the NCBI Short Read Archive
(SRA) (Leinonen et al., 2011). Accession numbers can be found in Supplemental Table S2.
Reference transcript sequences were collected from Ensembl v.102 (Yates et al., 2020), and we
only included the longest transcript for each gene. For each library, adapters were removed using
Trimmomatic 0.38 (Bolger et al., 2014), and reads were filtered for quality using the parameters
HEADCROP:5, SLIDINGWINDOW:5:30, and MINLEN:50. We mapped quality filtered paired-
end RNAseq reads back to reference sequences using Bowtie 1.2.2 (Langmead et al., 2009) and
default parameters of RSEM (Li and Dewey, 2011). Normalization of raw read counts for each
species was performed using the estimateSizeFactors and estimateDispersions functions in
DESeq2 v1.26 (Love et al., 2014).

3   Results and Discussion
Understanding the evolution of gene families is an active research area in evolutionary biology.
Reconstructing the sister group relationship among gene family members represents a fundamental
piece of information to understand homologous relationships, duplicative history, and their modes
of evolution (Nei and Rooney, 2005; Glover et al., 2019). Further, the availability of whole-
genome sequences opens an opportunity to discover new genes (Himmel et al., 2020; Opazo et al.,
2020). Today, the GOLPH3 gene family is understood as a group of genes containing two paralogs
(GOLPH3 and GOLPH3L) with 1:1 orthologs among most vertebrate species (Wu et al., 2000;
Bell et al., 2001). However, our comparative analyses suggest that its evolutionary history is more
complicated than previously thought. The evolution of the GOLPH3 paralog in birds is
characterized by independent duplication events, whereas the evolutionary history of GOLPH3L
seems to be less dynamic, being possible to identify 1:1 orthologs in all bird species.

3.1   Independent duplication events characterize the evolution of the GOLPH3 gene family member in birds

Our gene tree did not significantly deviate from the most updated phylogenetic hypotheses for the main group of birds (Prum et al., 2015; Kimball et al., 2019; Feng et al., 2020; Kuhl et al., 2021)(Fig. 1), suggesting that GOLPH3 was present in the ancestor of the group as a single copy gene. We recovered a clade containing GOLPH3 sequences from paleognaths (ostriches, tinamous, and allies) sister to GOLPH3 sequences from all other birds (Fig. 1). Further, we recovered the sister group relationship of the GOLPH3 sequences from Galliformes (chickens, pheasants, and allies) and Anseriformes (ducks, swans, and allies), in turn, this clade was recovered sister to GOLPH3 sequences from Neoaves (Fig. 1).

We found a single copy gene, located on chromosome Z, in most paleognaths species, except in the white-throated tinamou (*Tinamus guttatus*), where two copies were identified (Fig. 1), suggesting that this species independently gave rise to a second GOLPH3 copy located on chromosome W. The location of these genes on sexual chromosomes, and given the sex-determination system of birds (Chue and Smith, 2011), indicates that only females (ZW) can express both paralogs. In the case of Neoaves, our tree topology is not well resolved, being difficult to anticipate details regarding the duplicative history of the GOLPH3 paralog in this group (Fig. 1). However, for a diversity of species (e.g., zebra finch, common canary, kakapo), we found duplicated copies on different chromosomes, suggesting that the duplication event that gave rise to them occurred in the ancestor of the group. Similar to the case of the white-throated tinamou (*Tinamus guttatus*), we found duplicated copies in the killdeer (*Charadrius vociferus*) that were recovered sister to each other (Fig. 1), suggesting that they arouse as a product of a species-specific gene duplication event.

The evolutionary history of the GOLPH3 gene in the clade that includes Galliformes and Anseriformes followed a more complicated evolutionary pathway (Fig. 1). According to our assessment, we found a repertoire of two copies in species belonging to both groups (Fig. 1); however, our gene tree suggests that the events that gave rise to them followed a pattern of gene birth-and-death (Hughes and Nei, 1989)(Fig. 2). The reconciliation of the gene tree with the species tree suggests that the last common ancestor of Anseriformes and Galliformes, which lived 80 million years ago approximately (Hedges et al., 2015), had a single copy gene that underwent a duplication event (Fig. 2), giving rise to a repertoire of two GOLPH3 copies (Fig. 2). One of the copies was retained in Galliformes (Fig. 2; GOLPH3.1$_{GA}$; red lineage), but lost in Anseriformes (Fig. 2; red lineage). This GOLPH3 gene copy is located on chromosome W. The other copy, also originated in the ancestor of Galliformes and Anseriformes, was also retained in Galliformes (Fig. 3; GOLPH3.2$_{GA}$; pink lineage) and is located on chromosome Z. In the last common ancestor of Anseriformes, this copy underwent a duplication event giving rise to two copies (Fig. 3; GOLPH3.2.1$_{A}$, purple lineage and GOLPH3.2.2$_{A}$, light purple lineage). Like other cases, in Anseriformes, one of the copies is located on chromosome Z, while the other on chromosome W; the exception is the mallard (*Anas platyrhynchos*) in which is found on chromosome 22.

Thus, the main groups of birds possess gene repertoires with different evolutionary origins (Fig. 2). Most paleognaths retained the ancestral condition of a single gene copy, whereas Galliformes, Anseriformes, and Neoaves possess duplicated copies that were originated independently (Fig. 2). Anseriformes and Neoaves gave rise to their repertoire in the ancestor of each group (Fig. 2), while Galliformes retained copies originated in the ancestor of Galliformes and Anseriformes (Fig. 2). The independent origin of gene families in different groups is not an unusual event during the evolutionary process (Goodman et al., 1987; Kriener et al., 2001;

Hoffmann et al., 2010; Opazo and Zavala, 2018; Gramzow et al., 2020); however, it should be
taken into account when making comparisons because non-orthologous genes - i.e., genes with
different evolutionary origin - are being compared. Our results also highlight the importance of
manual curation in defining the composition of gene families. The description of new genes is also
not uncommon (Wichmann et al., 2016; Himmel et al., 2020; Opazo et al., 2020), and their
discovery could be attributed to their presence in non-model species and/or the absence of
appropriate evolutionary analyses. The presence of species with different gene repertoires
represents an opportunity to understand the evolutionary fate of duplicated genes (Lynch and
Conery, 2000) and the biological functions associated with a group of genes. This phenomenon,
variation in gene copy number, has been associated with differences in susceptibility to diseases
in different taxonomic groups. For example, in the African elephant (*Loxodonta africana*), it has
been claimed that an expansion in the number of TP53 "the guardian of the genome" gene copies
could help to explain the lower risk of developing cancer in this large and long-lived animal
(Abegglen et al., 2015; Sulak et al., 2016). Similarly, in whales there are also an expansion of gene
families related to cancer, and an accelerated rate of evolution in genomic regions enriched with
pathways involved in cancer (Tollis et al., 2019; Tejada-Martinez et al., 2021). Further evidence
comes from bats, a group in which the lifespan exceeds the expectation based on their body size
(de Magalhães and Costa, 2009). In this group, it has been documented the expansion of several
genes, for example, FBXO31, which is related to cell cycle arrest and response to DNA damage
diminishing the probability of developing cancer (Santra et al., 2009; Seim et al., 2013; Zhang et
al., 2013). Thus, the expanded repertoire of GOLPH3 genes in birds could be part of a set of
genomic traits that account for their lower susceptibility to cancer than mammals.
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3.2   Molecular divergence between duplicated copies of GOLPH3 genes
According to our analyses, the divergence values between duplicated GOLPH3 copies are low. In
Galliformes, the divergence values ranged from 1.79% to 4.76%. However, by checking the amino
acid alignment, we realized that most of the observed differences are in the first ~50 amino acids
of the N-terminal region of the protein (Fig. 3). By estimating amino acid sequence divergence for
the N- and C- terminal regions separately, we observed that the divergence values for the C-
terminal region ranged from 1.79% to 2.47%, while for the N-terminal region, which represents
only ~1/6 of the amino acid sequence, ranged from 15.69% to 17.65%. In this group of birds we
found eleven amino acid positions in the alignment that unequivocally distinguish between both
paralogs (Fig. 3). Six of them are in the N-terminal region, while the others in the C-terminal
portion of the molecule (Fig. 3). In Anseriformes (Fig. 4), the divergence values range from 1.69%
to 4.49%, similar to those estimated for Galliformes. Also, most of the observed differences are in
the N-terminal region of the molecule with divergence values ranging from 5.66% to 9.62%. In
the case of the C-terminal region, the values varied from 0.8% to 2.9%. In this group of birds there
are three amino acid positions that unequivocally distinguish both paralogs (Fig. 4), one of them
is located on the N-terminal region of the molecule, whereas the other two in the C-terminal region
(Fig. 4). In Neoaves we found the same evolutionary pattern as described for Anseriformes and
Galliformes, i.e. most of the amino acid replacements are found in the N-terminal portion of the
protein. The divergence values for the whole protein ranged from 1.36% to 3.72%, for the N-
terminal region varies from 3.92% to 15.38%, whereas for the C-terminal region goes from 0.41%
to 2.06%. Unlike the previous cases we did not identify amino acid sites in the alignment that
distinguish both paralogs (Fig. 5).

These differences in amino acid sequence divergence for the N- and C- terminal regions of
the GOLPH3 paralogs could have arisen as a consequence of different structural and functional
constraints during its evolution. Secondary structure prediction indicates that the region
comprising the first ~40-60 amino acids of GOLPH3 is disordered in a variety of organisms (e.g.,
yeast, fruit fly, spotted gar, human) that share a common ancestor more than a billion of years ago
(Fig. 6A and Supplementary Figure 1). Accordingly, only the crystal structures of N-terminal
truncation variants of GOLPH3 and Vps74 (GOLPH3 in yeasts) have been solved (Schmitz et al.,
2008; Wood et al., 2009). Both structures are remarkably similar (backbone atom root mean square
deviation of ~1.0 Å), consisting of a single globular domain that is predominantly α-helical, with
a central four-helix bundle surrounded by solvent-exposed loops, and eight amphipathic helices
(Schmitz et al., 2008; Wood et al., 2009). The overall structure of the N-terminal truncated
GOLPH3 protein is unique, with no strong structural homology to known protein folds, resulting
so far challenging to predict its function based on its structure. Protein structure homology
modeling of GOLPH3.1$_{GA}$ and GOLPH3.2$_{GA}$ of chicken (Supplementary Figure 2) showed that of
the divergent amino acids in the C-terminal region only L255 in GOLPH3.1$_{GA}$ and Q256 in
GOLPH3.2$_{GA}$ are non-conservative (Fig. 3) (Barnes and Gray, 2003). The position of Q256 is
predicted to be exposed at the surface of GOLPH3.2$_{GA}$, like it is for Q260 in human GOLPH3.
However, it is intriguing the variant L255 in GOLPH3.1$_{GA}$ because the preferred position of
leucine residues is buried in protein hydrophobic cores. None of the divergent amino acids of the
C-terminal region in Anseriformes and Neoaves are structurally disfavored. The C-terminal region
of GOLPH3 is sufficient for GOLPH3 physical interaction with the membrane of the Golgi
apparatus (Wu et al., 2000). This interaction is mediated by a series of highly conserved residues
that are postulated to interact with phosphates groups and the inositol ring of phosphatidylinositol
4-phosphate located in the cytosolic leaflet of the Golgi membrane (Wood et al., 2009), set of
residues that are also conserved in both copies of GOLPH3 in birds (Figs. 3-5 and Supplementary
Figures 1 and 3). In contrast, the N-terminal disordered region has no known function. Intriguingly,
some proteins containing disordered regions have the capacity to undergo liquid-liquid phase
separation that could result in their partitioning in functional biomolecular condensates also known
as membrane-less compartments (Wu et al., 2000; Borcherds et al., 2020). However, it is unknown
whether GOLPH3 has this capacity. In any case, the distinct amino acid sequence divergence
values for the N-terminal disordered region of GOLPH3 suggest a much flexible functional role.
Thus, it will be important to determine whether this domain contributes in the functions of
GOLPH3 as oncoprotein.
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3.3   Evolution of GOLPH3L paralog
In contrast to GOLPH3, GOLPH3L is largely uncharacterized. Although the amino acid sequences
of human GOLPH3 and GOLPH3L are 78% similar (65% identical), it has been suggested that
GOLPH3L antagonizes the functions of GOLPH3 (Ng et al., 2013). Despite this, other reports
suggest a similar function to GOLPH3 for GOLPH3L in some types of cancer (Kunigou et al.,
2011; Sotgia et al., 2012; Feng et al., 2015, 2017; He et al., 2017). The evolutionary history of
GOLPH3L followed a different trajectory in comparison to the GOLPH3 gene (Fig. 7). In this
case, our gene tree recovered the main groups of birds according to the most updated organismal
phylogenies (Prum et al., 2015; Kimball et al., 2019; Feng et al., 2020; Kuhl et al., 2021);
nevertheless, it was not possible to define the relationships among them (Fig. 7). We will assume
that the lack of variability does not allow us to resolve their sister-group relationship, instead of
more complex evolutionary scenarios invoking gene duplications and reciprocal loss in the

ancestor of the main groups of birds. Thus, according to our results the GOLPH3L gene was
present in the ancestor of birds as a single copy gene (Fig. 2), and this gene was inherited by all
descendant lineages (Fig. 2). Thus, GOLPH3L genes in different bird species are 1:1 orthologs.
Amino acid divergence values show a similar trend as we described for GOLPH3, i.e., the N-
terminal portion of the protein is more divergent than the C-terminal region (Fig. 8). In the case of
Galliformes, the divergence values for the N-terminal part of the molecule ranged from 5.71% to
17.14%, while for the C-terminal region varies from 3.28% to 6.97%. In Anseriformes, the values
for the N-terminal region ranged from 7.89% to 26.32%, whereas for the C-terminal portion varies
from 2.05% to 4.92%. In the case of Neoves, the evolutionary trend is the same, although the
values for the N-terminal region are higher. Thus, the values for the N-terminal part of the molecule
ranged from 20% to 43.34%, while for the C-terminal region varied from 4.10% to 17.55%. The
secondary structure prediction indicates that this region in GOLPH3L, although shorter than in
GOLPH3, is also disordered (Fig. 6B).
One thing that seems interesting is the number of changes accumulated in the branch
leading to Galliformes and to manakins (Fig. 7). This phenomenon could be indicative of an
acceleration of the rate of fixation of amino acid changing mutations in the ancestors of both
groups. To test this hypothesis, we estimated the omega value (d_N/d_S), i.e., the ratio of non-
synonymous (d_N) to synonymous substitutions (d_S), in the branches leading to both groups. In
brief, if non-synonymous substitutions are neutral, then the rate of fixation of d_N and d_S will be
very similar, and $d_N/d_S \approx 1$. Under negative selection, most non-synonymous substitutions are
deleterious, and $d_N/d_S < 1$. Finally, under positive selection non-synonymous (d_N) replacements
are advantageous and will be fixed at a greater rate than synonymous substitutions (d_S) and in
consequence $d_N/d_S > 1$ (Yang and Bielawski, 2000). According to our analyses, in the ancestor of
Galliformes the model in which the omega value was estimated from the data was not significantly
different from the model in which the omega value was fixed to 1 (neutral evolution)(LRT=0.142,
P > 0.05). On the other hand, in the case of manakins the model in which the omega value was
estimated from the data (d_N/d_S=6.2) was significantly different from the null hypothesis of neutral
evolution (LRT=7.19, P < 0.01), indicating that the rate of fixation of non-synonymous
substitutions (d_N) is higher in comparison to the neutral expectation (d_S) and suggesting an event
of positive selection in the ancestor of manakins. According to the Bayes Empirical Bayes (BEB)
approach five sites (152C/G, 191R, 230A, 233R and 263G) were inferred under positive selection
with a posterior probability higher than 0.95. All of them are located in the C-terminal region of
the protein. Given the limited understanding of the biological functions associated with the
GOLPH3 gene family, in particular of the GOLPH3L gene (Sechi et al., 2015), it is challenging to
explain the consequences of positive selection in GOLPH3L in this group of birds. However, it
could be interesting to carry out functional assays in which the performance of manakins
GOLPH3L protein is compared to the one in other birds.
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3.4   Expression pattern of GOLPH3 gene family members
Our next step was to investigate the expression pattern of the GOLPH3 gene family members,
especially for the duplicated copies derived independently in different groups of birds. To do this,
we mapped RNASeq reads to reference gene sequences in the chicken (*Gallus gallus*) and mallard
(*Anas platyrhynchos*) and examined transcript abundance in a panel of nine tissues (Fig. 9). It is
important to say that in chicken, the duplicated GOLPH3 copies are located on chromosome Z
(GOLPH3.2$_{GA}$) and W (GOLPH3.1$_{GA}$), while GOLPH3L is on chromosome 25. Therefore,
females (ZW) can express all gene family members, whereas males (ZZ) can only express

GOLPH3.2$_{GA}$ and GOLPH3L. This situation is somewhat similar to the allelic trichromacy
observed in New World monkeys, where some females (XX) possess trichromatic color vision due
to a polymorphism of an opsin gene located on chromosome X, while males (XY) are all
dichromatic (Jacobs, 1996). The case is different in mallard, as one copy is located on chromosome
Z (GOLPH3.2.1$_A$) but GOLPH3.2.2$_A$ and GOLPH3L are autosomal genes, so both sexes can
potentially express all paralogs.
In both species, the paralog located on the Z chromosome (GOLPH3.2$_{GA}$ in chicken and
GOLPH3.2.1$_A$ in mallard) was highly and ubiquitously expressed across all tissues (Fig. 9). We
collected chicken libraries from both male and female tissues, and unfortunately the sex of the
individuals for some tissues was not declared (Supplemental Table S2). As such GOLPH3.1$_{GA}$
exhibited variable expression from mixed sex sampling (Fig 9A). Thus, GOLPH3.1$_{GA}$ was highly
expressed in the brain and ovary where all libraries were constructed from female individuals (Fig
9A). Although we do not know the sex of the individuals for the intestine libraries of the chicken,
based on the consistent expression of the gene located on chromosome W, we can presume that
they were all from female individuals (Fig. 9A). As a validation of what we mentioned above,
GOLPH3.1$_{GA}$ was not expressed in all known male tissue libraries, which was most noticeable in
the male specific testes (Fig. 9A). All autosomal paralogs in both species were variably expressed
among and within tissues (Fig 9). In the case of the chicken GOLPH3L, we recovered expression
in all tissues but at variable levels, from low values in the brain and heart to higher values in the
intestine and kidney (Fig. 9A). By contrast GOLPH3L was universally expressed in all mallard
tissues (Fig. 9B). GOLPH3.2.2$_A$ in mallard was not expressed in the brain, liver and testes, but
highly expressed in the ovary, spleen and intestine. In both species, all GOLPH3 paralogs were
consistently expressed in the intestine (Fig. 9).
In humans, both paralogs are ubiquitously expressed across all tissues (Uhlén et al., 2015),
suggesting that they are required for the maintenance of basic cellular functions (Eisenberg and
Levanon, 2013); however, GOLPH3L seems to be expressed at lower levels. Similarly, the relative
expression levels of GOLPH3 and GOLPH3L in several mammalian cell lines with epithelial,
fibroblast, myeloid and neuronal characteristics, and in a variety of tissues from mice also indicates
that GOLPH3 is also ubiquitously expressed at higher levels than GOLPH3L (Ng et al., 2013).
Further, GOLPH3L is expressed more in cells with secretory epithelial characteristics (Ng et al.,
2013), suggesting a distinct function for this gene family member. The expression pattern observed
in the mallard is similar to what is observed in model species (Fig. 9B). One of the GOLPH3
duplicates (GOLPH3.2.1$_A$) is expressed in all examined tissues at high levels, while GOLPH3L is
also expressed in all tissues, but at lower levels. The expression of the other duplicate
(GOLPH3.2.2$_A$) is variable, including tissues in which it is not detected (Fig. 9). The case of the
chicken seems to be more dissimilar. In this species one of the GOLPH3 duplicates (GOLPH3.2$_{GA}$)
possesses an expression pattern similar to the human GOLPH3 (Fig. 9A), however the other two
copies seem to follow a specific expression pattern (Fig. 9A).
408
4   Conclusions
Our study shows that the evolution of the GOLPH3 gene family followed a more
complicated evolutionary pathway than previously thought. Although the history of the GOLPH3L
paralog is according to the current knowledge, the one of GOLPH3 is not. The most exciting thing
about the evolution of GOLPH3 in birds is that they possess extra GOLPH3 gene copies never
described before, and that all main groups independently originated their repertoire. In other
words, they do not have the same evolutionary origin, and in consequence, they are not 1:1

orthologs and are not directly comparable. Most paleognaths retained the ancestral condition of a
single gene copy, whereas Galliformes, Anseriformes, and Neoaves possess duplicated copies
originated independently. Thus, birds represent a natural experiment of gene copy number
variation (Albertson et al., 2009), that in addition to the differences in expression of individuals of
different sex, could help us improve our understanding of the biological functions associated with
the GOLPH3 gene family. Our results also highlight the power of manually curating genetic data
to define gene repertoires, and the reconciliation of gene trees with species trees (Goodman et al.,
1979) to understand the duplicative history of gene families to perform biologically meaningful
comparisons (Gabaldón, 2008; Glover et al., 2019). Finally, the conservation of the N-terminal
portion of GOLPH3 paralogs as a disordered region for more than a billion of years of evolution
and the fact that it displays a higher degree of divergence among species, compared to the C-
terminal portion, strongly suggests that it performs an essential, specialized and adapted cellular
function conserved in distantly related species like yeasts and humans that remains to be
elucidated.
430

431 **Conflict of Interest**
432 The authors declare that the research was conducted in the absence of any commercial or financial
433 relationships that could be construed as a potential conflict of interest.
434

435 **Author contributions**
436 JCO and GAM designed the study. MWV, JG, KZ, JCO, GAM collected and/or analyzed data.
437 JCO and GAM wrote the manuscript. MWV, LV-C, FJM reviewed and edited the manuscript. All
438 authors contributed to the article and approved the submitted version.
439

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Supplementary material
The supplementary material for this article can be found online at:
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Figure legends
Figure 1. Maximum likelihood tree showing sister group relationships among GOLPH3 genes of birds. Numbers above the nodes correspond to support values from the ultrafast bootstrap routine. GOLPH3 sequences from crocodiles and turtles were used as outgroups (not shown). The scale denotes substitutions per site and colors represent gene lineages.

Figure 2. An evolutionary hypothesis regarding the evolution of the Golgi phosphoprotein 3 (GOLPH3) gene family in birds. According to our results, the last common ancestor of birds had a repertoire of two GOLPH3 genes (GOLPH3 and GOLPH3L). The GOLPH3L gene was inherited by all species of birds and maintained as a single copy gene (brown lineage). The evolution of the GOLPH3 gene is more complicated. The gene present in the bird ancestor was inherited by paleognaths (ostriches, tinamous and allies) and maintained as a single copy gene in most species. In the case of Neoaves (zebra finches, kakapos and allies), the ancestral GOLPH3 gene underwent a duplication event in the ancestor of the group giving rise to a repertoire of two genes (blue and light blue gene lineages). In the last common ancestor of Anseriformes (ducks, swans and allies) and Galliformes (chickens, pheasants and allies) the ancestral GOLPH3 gene underwent a duplication event giving rise to a repertoire of two genes, one of the copies was retained in Galliformes (red lineage), but lost in Anseriformes (red lineage). The other copy originated in the ancestor of Galliformes and Anseriformes, was also retained in Galliformes (pink lineage), however, in the last common ancestor of Anseriformes underwent a duplication event giving rise to the repertoire of two copies (purple and light purple lineages). We use the subscripts to indicate the ancestor in which the gene repertoires were originated, N = the ancestor of Neoaves, A = the ancestor of Anseriformes and GA = the galliform/anseriform ancestor. Silhouette images were obtained from PhyloPic (http://phylopic.org/).

Figure 3. Alignment of Golgi phosphoprotein 3 (GOLPH3) amino acid sequences from chicken (*Gallus gallus*), ring-necked pheasant (*Phasianus colchicus)*, turkey (*Meleagris gallopavo*) and japanese quail (*Coturnix japonica*). Amino acids positions that differentiate between paralogs are in pink and red. The N-terminal region of the protein is shaded.

Figure 4. Alignment of Golgi phosphoprotein 3 (GOLPH3) amino acid sequences from mallard (*Anas platyrhynchos*), Tufted duck (*Aythya fuligula)*, swan goose (*Anser cygnoides*) and ruddy duck (*Oxyura jamaicensis*). Amino acids positions that differentiate between paralogs are in purple and light purple. The N-terminal region of the protein is shaded.

Figure 5. Alignment of Golgi phosphoprotein 3 (GOLPH3) amino acid sequences from lance tailed manakin (*Chiroxiphia lanceolata*), kakapo (*Strigops habroptilus*), New caledonian crow (*Corvus moneduloides*) and zebra finch (*Taeniopygia guttata*). The N-terminal region of the protein is shaded.

Figure 6. Secondary structure and disordered region predictions of GOLPH3 (A) and GOLPH3L (B) in human (*Homo sapiens*), mouse (*Mus musculus*), chicken (*Gallus gallus*), Western clawed frog (*Xenopus tropicalis*), coelacanth (*Latimeria chalumnae*), spotted gar (*Lepisosteus oculatus*), thorny skate (*Amblyraja radiata*), fruit fly (*Drosophila melanogaster*), roundworm (*Caenorhabditis elegans*) and brewer yeast (*Saccharomyces cerevisiae*).

500 **Figure 7.** Maximum likelihood tree showing sister group relationships among GOLPH3L genes
501 of birds. Numbers above the nodes correspond to support values from the ultrafast bootstrap
502 routine. GOLPH3L sequences from crocodiles and turtles were used as outgroups (not shown).
503 The scale denotes substitutions per site.
504
Figure 8. Alignment of Golgi phosphoprotein 3L (GOLPH3L) amino acid sequences from mallard
(*Anas platyrhynchos*), Tufted duck (*Aythya fuligula)*, black swan (*Cygnus atratus*), ruddy duck
(*Oxyura jamaicensis*), chicken (*Gallus gallus*), ring-necked pheasant (*Phasianus colchicus)*,
turkey (*Meleagris gallopavo*), japanese quail (*Coturnix japonica*), lance tailed manakin
(*Chiroxiphia lanceolata*), kakapo (*Strigops habroptilus*), New caledonian crow (*Corvus
moneduloides*) and zebra finch (*Taeniopygia guttata*). The N-terminal region of the protein is
shaded.
512
513 **Figure 9**. Transcript abundance measurements of GOLPH3 paralogs across a panel of nine tissues
514 in A) chicken (*Gallus gallus*) and B) mallard (*Anas platyrhynchos*). The mean and standard
515 deviation are plotted for three replicates per tissue. In many cases the standard deviation was
516 smaller than the size of the circle representing the mean and appears hidden.
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741
742
743

N-terminal

| | 1 | 10 | 20 | 30 | 40 | 50 | 60 | 70 |

Mallard GOLPH3.2.1_A MTSLTQRSSGLVQRRTEASRS–AAAADKERGAGGG**P**EDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG
Tufted duck GOLPH3.2.1_A MTSLTQRSSGLVQRRTEASRS–AAAADKERGAGGG**P**EDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG
Swan goose GOLPH3.2.1_A MTSLTQRSSGLVQRRTEASRS–AAAADKERGAGGG**P**EDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG
Ruddy duck GOLPH3.2.1_A MTSLTQRSSGLVQRRTEASRS–AAAADKERGAGGG**P**EDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG

Mallard GOLPH3.2.2_A MTSLTQRSSGLVQRRTEASRS–AAAADKEWGAGGG**L**EDEGHRDELGDDEKGDSKETRLTLMEEVLLLGLKDREG
Tufted duck GOLPH3.2.2_A MTSLTQRSSGLVQRRTEASRS–AAAADKEWGAGGG**L**EDEGHRDELGDDVKGDSKETRLTLMEEVLLLGLKDREG
Swan goose GOLPH3.2.2_A MTSLTQRSSGLVQRRTEASRSAAAAAKERGTGGG**L**EDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDRXG
Ruddy duck GOLPH3.2.2_A MTSLTQRNSGLVQRRTEASRSAAAAADKERGAGGG**L**EDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG

| | 80 | 90 | 100 | 110 | 120 | 130 | 140 |

Mallard GOLPH3.2.1_A YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSD**AP**TGDVLLDEALKHIKETQPPETVQ
Tufted duck GOLPH3.2.1_A YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSD**AP**TGDVLLDEALKHIKETQPPETVQ
Swan goose GOLPH3.2.1_A YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSD**AP**TGDVLLDEALKHIKETQPPETVQ
Ruddy duck GOLPH3.2.1_A YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSD**AP**TGDVLLDEALKHIKETQPPETVQ

Mallard GOLPH3.2.2_A YTSFWNDCISSGLRGCMLIELALRGRLQLEACGIRRKSLLTRKVICKSD**TS**TGDVLLDEALKHIKETQPPETVQ
Tufted duck GOLPH3.2.2_A YTSFWNDCISSGLRGCMLIELALRGRLQLEACGIRRKSLLTRKVICKSD**TS**TGDVLLDEALKHIKETQPPETVQ
Swan goose GOLPH3.2.2_A YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSD**TS**TGDVLLDEALKHIKETQPPETVQ
Ruddy duck GOLPH3.2.2_A YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSD**TS**TGDVLLDEALKHIKETQPPETVQ

| | 150 | 160 | 170 | 180 | 190 | 200 | 210 | 220 |

Mallard GOLPH3.2.1_A NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Tufted duck GOLPH3.2.1_A NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Swan goose GOLPH3.2.1_A NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Ruddy duck GOLPH3.2.1_A NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK

Mallard GOLPH3.2.2_A NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Tufted duck GOLPH3.2.2_A NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Swan goose GOLPH3.2.2_A NWIELLS--
Ruddy duck GOLPH3.2.2_A NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK

| | 230 | 240 | 250 | 260 | 270 | 280 | 290 | 296 |

Mallard GOLPH3.2.1_A WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKANTNEVLWAVVAAFTK
Tufted duck GOLPH3.2.1_A WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKANTNEVLWAVVAAFTK
Swan goose GOLPH3.2.1_A WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKANTNEVLWAVVAAFTK
Ruddy duck GOLPH3.2.1_A WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKANTNEVLWAVVAAFTK

Mallard GOLPH3.2.2_A WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRMRQLLDLDPEVECMKANTNEVLWAVVAAFTK
Tufted duck GOLPH3.2.2_A WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRMRQLLDLDPEVECMKANTNEVLWAVVAAFTK
Swan goose GOLPH3.2.2_A ---
Ruddy duck GOLPH3.2.2_A WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKANTNEVLWAVVAAFTK

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                          1         10        20        30        40        50        60        70
Lance tailed manakin GOLPH3.1ₙ   MTSLTQRSSGLVQRRTEASRS--AAADKERGAGGGSDDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG
Kakapo GOLPH3.1ₙ                 MTSLTQRSSGLVQRRTEASRS--AAADKERGAGGGSEDEGRRDESGDDEKGDSKETRLTLMEEVLLLGLKDREG
New Caledonian crow GOLPH3.1ₙ   MTSLTQRSSGLVQRRTEASRSAAAAADKERGAGGGPEDEGRRDESGDDEKGDSKETRLTLMEEVLLLGLKDREG
Zebra finch GOLPH3.1ₙ           MTSLTQRSSGLVQRRTEASRS-AAAADKERGAGGGPEDEGRRDESGDDEKGDSKETRLTLMEEVLLLGLKDREG

Lance tailed manakin GOLPH3.2ₙ   MTSLTQRSSGLVQRRTEASRS--AAADKDREAGGGSEDGGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG
Kakapo GOLPH3.2ₙ                 MTSLTQRSSGLVQRRTEASRS--TAAGKERGAGGGPEDEGRRDDPGDDEKGDSKEIRLTLMEEVLLLGLKDREG
New Caledonian crow GOLPH3.2ₙ   MTSLTQRSSGLVQRRTEASRS--AAADKEREAGGGSEDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG
Zebra finch GOLPH3.2ₙ           MTSLTQRSSGLVQRRTEAFRN--AAAAKEREAGGSSEDEGRRDEPGDDEKGDSKETRLTLMEEVLLLGLKDREG
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                                 80        90        100       110       120       130       140
Lance tailed manakin GOLPH3.1ₙ   YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSDAPTGDVLLDEALKHIKETQPPETVQ
Kakapo GOLPH3.1ₙ                 YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSDAPTGDVLLDEALKHIKETQPPETVQ
New Caledonian crow GOLPH3.1ₙ   YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSDAPTGDVLLDEALKHIKETQPPETVQ
Zebra finch GOLPH3.1ₙ           YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSDAPTGDVLLDEALKHIKETQPPETVQ

Lance tailed manakin GOLPH3.2ₙ   YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSDAPTGDVLLDEALKHIKETQPPETVQ
Kakapo GOLPH3.2ₙ                 YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLSRKVICKSDAPTGDVLLDEALKHIKETQPPETVQ
New Caledonian crow GOLPH3.2ₙ   YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSDAPTGDVLLDEALKHIKETQPPETVQ
Zebra finch GOLPH3.2ₙ           YTSFWNDCISSGLRGCMLIELALRGRLQLEACGMRRKSLLTRKVICKSDAPTGDVLLDEALKHIKETQPPETVQ
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                                 150       160       170       180       190       200       210       220
Lance tailed manakin GOLPH3.1ₙ   NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Kakapo GOLPH3.1ₙ                 NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
New Caledonian crow GOLPH3.1ₙ   NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Zebra finch GOLPH3.1ₙ           NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK

Lance tailed manakin GOLPH3.2ₙ   NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Kakapo GOLPH3.2ₙ                 NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVVDK
New Caledonian crow GOLPH3.2ₙ   NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
Zebra finch GOLPH3.2ₙ           NWIELLSGETWNPLKLHYQLRNVRERLAKNLVEKGVLTTEKQNFLLFDMTTHPLTNNNIKQRLIKKVQEAVLDK
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                                 230       240       250       260       270       280       290    296
Lance tailed manakin GOLPH3.1ₙ   WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKTNTNEVLWAVVAAFTK
Kakapo GOLPH3.1ₙ                 WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKANTNEVLWAVVAAFTK
New Caledonian crow GOLPH3.1ₙ   WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKTNTNEVLWAVVAAFTK
Zebra finch GOLPH3.1ₙ           WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKTNTNEVLWAVVAAFTK

Lance tailed manakin GOLPH3.2ₙ   WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKANMNEVLWAVVAAFTK
Kakapo GOLPH3.2ₙ                 WVNDPHRMDKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEAECMKANTNEVLWAVVAAFTK
New Caledonian crow GOLPH3.2ₙ   WVNDPHRMEKRLLALVYLAHASDVLENAFAPLLDEQYDLATKRVRQLLDLDPEVECMKANMNEVLWAVVAAFTK
Zebra finch GOLPH3.2ₙ           WVNDPHRMEKRLLALVYLAHASDVLENAFSPLLDEQYDLATKRVRQLLDLDPEVECMKANMNEVLWAVVAAFTK
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(A) GOLPH3

(B) GOLPH3L

GOLPH3L

N-terminal

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        1        10        20        30        40        50        60        70        80        90
Mallard GOLPH3L              MTTLTHRGRRSEAGGSVGKRLDGEEE---ERDP--EEDDDSKGTRLTLMEEVLLLGLKDK-GYTSFWNDCISSGLRGGILIELALRGRIQLEPLTA
Tufted duck GOLPH3L          MTTLTHRGRRTEAGGSVGKRVDGEE----ERDL---EDDDSKGTRLTLMEEVLLLGLKDKQGYTSFWNDCISSGLRGGILIELALRGRIQLEPLTA
Black swan GOLPH3L           MTTLTHRGRRTEAGGSTGKKLDDEEEAAAERDP---EDDDSKGTRLTLMEEVLLLGLKDKQGYTSFWNDCISPGLRGGILVELALRGRIQLEPLTT
Ruddy duck GOLPH3L           MTTLTHRGRRGDAGGSVGKKLDDEEEAAAERDP---EDDDSKGTRLTLMEEVLLLGLKDKQGYTSFWNDCISPGLRGGILVELALRGRIQLEPLTA

Chicken GOLPH3L              MTTLTRRGWRGD------KRTEPEEDAAGDREQPDDDSDNSKATRLTLMEEVLLLGLQDKEGYPSFWNDCLSPGLRGGILIELALRGRIHLEPLTA
Ring-necked pheasant GOLPH3L MTTLTRRGWRGD------KRTEPEEDVAADRELPDDDSDNSKAPRLTLMEEVLLLGLQDKEGYPSFWNDCLSPGLRGGILIELALRGRIHLEPLTA
Turkey GOLPH3L              MTTLTRRGWRGD------KRTEPEEDAAADRELPDDDSDNSKAPRLTLMEEVLLLGLQDKEGYPSFWNDCLSPGLRGGILIELALRGRIHLEPLTA
Japanese quail GOLPH3L      MTTLTHRGWRGD------KRAEPEEDVAADKELPDDDSDNSKAPRLTLMEEVLLLGLQDKEGYPSFWNDCLSPGLRGGILIELALRGRIHLEPLTA

Lance tailed manakin GOLPH3L MTTLTHRARRGDPGGGSAKRPDGEEDGAADREPGEEEPDDSKDIRLTLMEEVLLLGLKDKEGYTSFWNDCISSGLRGGILIELALRGRIRLEPLAL
Kakapo GOLPH3L              MTTLTHRSRRAE-GRSTDRKPTGEEDAAADRDLNDEDPDDSKDIRLTLMEEVLLLGLKDKEGYTSFWNDCISSGLRGGILIELAFRGRIRLEPPSM
New Caledonian crow GOLPH3L  MTTLTHRGRRVE-GRNSDKKPDGEEDAAADRDLNEDDPDDSKDIRLTLMEEVLLLGLKDKEGYTSFWNDCISSGLRGGILIELACLRGRIRLEPLSL
Zebra finch GOLPH3L          MTTLTHRGRRAE-GRSADRRTEGEEEAAADGELSQEHPEDSKDIRLTLMEEVLLLGLKDKEGYTSFWNDCISSGLRGGILIELALRGRIRLEPPAL
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        100       110       120       130       140       150       160       170       180       190
Mallard GOLPH3L              RKKRLLDRKVHLKSDAPTGDILLDETLRHIKATEPAETVQTWIELLTGETWNPFKLQYQLRNVRERIAKNLVEKGILTTEKQNFLLFDMTTHPVSN
Tufted duck GOLPH3L          RKKRLLDRKVHLKSDAPTGDILLDETLRHIKATEPAETVQTWIELLTGETWNPFKLQYQLRNVRERIAKSLVEKGILTTEKQNFLLFDMTTHPVSN
Black swan GOLPH3L           RKKRLLDRKVHLKSDTPTGDILLDETLRHIKATEPAETVQTWIELLTGETWNPFKLQYQLRNVRERIAKNLVEKGILTTEKQNFLLFDMTTHPVSN
Ruddy duck GOLPH3L           RKKRLLDRKVHLKSDTPTGDILLDETLRHIKATEPAETVQTWIELLTGETWNPFKLQYQLRNVRERIAKSLVEKGILTTEKQNFLLFDMTTHPVSN

Chicken GOLPH3L              RKKRLLDRKVLLKSAAPTGDVLLDETLRHIKASESTETVQTWIELLTGETWNPFKLQYQLRNVRERVAKGLVEKGILTTGKQSFLLFDMTTHPICD
Ring-necked pheasant GOLPH3L RKKRLLDRKVLLKSAAPTGDVLLDETLRHIKASESMETVQTWIELLTGETWNPFKLQYQLRNVRERVAKGLVEKGILTTGKQSFLLFDMTTHPICD
Turkey GOLPH3L              RKKRLLDRKVLLKSAAPTGDVLLDETLRHIKASESTETVQTWIELLTGETWNPFKLQYQLRNVRERVAKGLVEKGILTTGKQSFLLFDMTTHPVCD
Japanese quail GOLPH3L      RKKRLLDRKVLLKSAAPTGDVLLDETLRHIKAAESIETVQTWIELLTGETWNPFKLQYQLRNVREHVAKGLVEKGILTTGKQSFLLFDMTTHPVCD

Lance tailed manakin GOLPH3L RRKRLLDRKVLLKSDTPTGDVLLDETLRHIKGSECPETVPTWIELLTGETWNPFKLQFQLRNVRERLAKALVEKGILGTEKQNFLLFDMTTHPLSN
Kakapo GOLPH3L              RKKRLLERKVLLKSDAPTGDVLLDETLRHIKATEAAESVQTWIELLTGETWNPFKLQYQLRNVRERVAKALVEKGILTTEKQNFLLFDMTTHPVSN
New Caledonian crow GOLPH3L  RKKRLLERKVLLKSDAPTGDVLLDETLRHIKATESAETVQTWIELLTGETWNPFKLQYQLRNVRERLAKALVEKGILTTEKQNFLLEDMTTHPVSN
Zebra finch GOLPH3L          RRKRLLERKVLLKSDAPTGDVLLDETLRHIKATEPAETVQSWIELLTGETWNPFKLQYQLRNVRERLAKALVEKGILTTEKQNFLLFDMTTHPVSN
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        200       210       220       230       240       250       260       270       280   286
Mallard GOLPH3L              AAEKQRLLKRLQEGVLERWAGDPHRMDRRTLALLVLAHSSDVLENAFSSLTDERYEAAMNRTKEVLDADPEVEAAKNR-GTEMIWAILAAFNKA
Tufted duck GOLPH3L          AAEKQRLLKRLQEGVLERWAGDPHRMDRRTLALLVLAHSSDVLENAFSSLTDDRYEVAMNRTKEVLDADPEVEAAKNR-DTEMIWAILAAFNKA
Black swan GOLPH3L           AAEKQRLLKRLQEGVLERWAGDPHRMDRRTLALLVLAHSSDVLENAFSSLADDRYEVAMNRTKEVLDADPEVEAAKGR-GTEMIWAILAAFNKA
Ruddy duck GOLPH3L           AAEKQRLLKRLQEGVLERWVGDPHRMDRRTLALLVLAHSSDVLENAFSSLEDDRYKVAMNRTKEVLDADPEVEAAKGR-GTEMIWAILAVFNKA

Chicken GOLPH3L              TTEKQRLLKKLQGSVLERWTGDLRRMDHRLLALLVLAHASDVLEKALGSLEDTQYEMAMSRAKDVLEANAELEAAQGR-GTEVIWAVLAAFSRA
Ring-necked pheasant GOLPH3L TTEKQRLLRKLQDSLLERWTGDPRRMDRRLLALLVLAHASDVLEKALGGLEDTQYEMAMSRVRDVLEADAELEAAQGR-GTEVIWAVLAAFNRA
Turkey GOLPH3L              TTEKQRLLRKLQDSVLERWMGDPRRMDRRLLALLVLAHASDVLEKALGGLEDTQYEMAMSRAKDVLEADAELEAAQGR-GTEVIWAVLAAFSRA
Japanese quail GOLPH3L      TTEKQRLLRKLQSSLLERWTGDVRRMERRLLALLVLAHTSDVLEKALGGLEDTQYELAMSRAKDVLEADTELEAAQGR-GTEVIWAVLAALSRA

Lance tailed manakin GOLPH3L GAEKLRLQRRLQESVLERWARDPRRLEPRTLALLVLAHSSDVLEGVFGALADDRYELALARARDLLDLDPELEAAKARGGCEMIWAVLAAFNKG
Kakapo GOLPH3L              GTEKQRLVRRLQESVLERWVNDPHRMDRRTLALLVLAHSSDVLENVFASLADDKYDVAMNRTKDLLDMDPEVEAAKAK-GTEMVWAVLAAFNKS
New Caledonian crow GOLPH3L  ASEKQRLVKKLQESVLERWVNDPQRMERRTLALLVLAHASDVLENVFASLADDKYDVAMNRTKDLLDMDPEVEAAKAK-GTEMIWAVLAAFNKS
Zebra finch GOLPH3L          ASEKQRLLKKLQESVLERWVNEPQRMERRTLALLVLAHASDVLENVFGSLADDKYDVAMNRTKDLLDMDPEVEAAKGK-GTEMIWAVLAAFNKS
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