- 1 **Title**: Ectomycorrhizas accelerate decomposition to a greater extent than arbuscular mycorrhizas
- 2 in a northern deciduous forest
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## 10 Abstract

11 It has been proposed that ectomycorrhizal (EcM) fungi slow down decomposition by competing with free-living saprotrophs for organic nutrients and other soil resources (known as the "Gadgil 12 effect"), thereby increasing soil carbon sequestration. As such, this Gadgil effect should depend 13 14 on soil organic matter age and quality, but this remains unstudied. In addition, the Gadgil effect is 15 not expected to occur in arbuscular mycorrhizal (AM) forests since AM fungi cannot access 16 directly nutrients from soil organic matter, yet few direct comparisons between EcM and AM 17 forests have been made. We performed a two-year reciprocal decomposition experiment of soil 18 organic horizons (litter - L, fragmented - F, humic - H) in adjacent temperate deciduous forests 19 dominated by EcM or AM trees. Litterbags were made of different mesh sizes allowing or 20 excluding ingrowth of external fungal hyphae, which are primarily mycorrhizal in these forests 21 other than for the most-recent superficial litter horizon. As expected, organic matter originating 22 from deeper horizons and from EcM forests was of lower quality (e.g. higher lignin to nitrogen 23 ratios) and decomposed more slowly. However, contrary to the Gadgil effect, organic matter 24 exposed to external fungal hyphae (i.e. primarily mycorrhizal) actually decomposed faster in both 25 forest types, and this effect was strongest in EcM forests, particularly in the F horizon. 26 Unexpectedly, organic matter decomposition was faster in EcM than in AM forests, regardless of 27 organic matter origin. Overall, our study reinforces the view that temperate EcM forests store 28 greater amounts of soil organic carbon than AM forests, but suggests that this is due to factors other than the Gadgil effect. 29

30

#### 31 Keywords

Organic matter decomposition; carbon cycle, nitrogen cycle, temperate forest, vertical
segregation; Gadgil effect; mycorrhizal fungi; *Acer saccharum*; *Fagus grandifolia*

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## 35 Introduction

Forests cover much of the land surface, and represent the largest terrestrial carbon (C) pool 36 37 globally (Dixon and others 1994; Baldrian 2017). A majority of that C is stored in forest soils, 38 especially in northern forests (Lal 2005; Crowther and others 2019). Soil C storage is controlled 39 by many abiotic and biotic factors such as climate, vegetation, topography and nutrient 40 availability that interact together (Averill and others 2014; Carvalhais and others 2014; Wiesmeier and others 2019). However, belowground biotic factors, such as microorganisms, also 41 42 play an important role, directly influencing soil C inputs (i.e. litter quantity and quality) and 43 outputs (i.e. decomposition) (Schimel and Schaeffer 2012). For example, soil microorganisms 44 such as fungi can produce recalcitrant organic matter that decomposes slowly or they can produce extracellular enzymes that break down organic matter (Frey 2019). As a result, soil fungi play a 45 major role in forest C cycling (Kubartová and others 2008; Bardgett and Wardle 2010; Orwin and 46 others 2011). 47

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49 A long-standing hypothesis about the effects of fungi on the soil C cycle is the "Gagdil effect" 50 (Gadgil and Gadgil 1971; Fernandez and Kennedy 2016). This hypothesis suggests 51 ectomycorrhizal (EcM) fungi slow down litter decomposition, potentially due to competition 52 between EcM fungi and free-living saprotrophs for organic nutrients. Because EcM fungi acquire 53 their C in highly labile form via plant hosts (Smith and Read 2008) in exchange for nutrients such 54 as nitrogen and phosphorus, they would leave behind C-rich but nutrient-poor organic matter, 55 potentially favoring soil C accumulation (Read and others 2004; Averill and others 2014). On the 56 other hand, some EcM fungi have the capacity to oxidize organic matter, directly influencing 57 decomposition and indirectly influencing saprotrophic organisms (Lindahl and Tunlid 2015;

58 Verbruggen and others 2017). Saprotrophic fungi could also be impacted by EcM fungi through 59 mycoparatism, antibiosis and alteration of abiotic conditions (Fernandez and Kennedy 2016; Zak and others 2019). The Gagdil effect has only been supported by a few studies but seems to be 60 61 largely context dependent, for example to litter quality (Smith and Wan 2019) and moisture level 62 (Koide and Wu 2003). Compared to EcM fungi, it is considered that arbuscular mycorrhizal 63 (AM) fungi lack the capacity to produce enzymes that break down organic matter (Tisserant and 64 others 2013; Tedersoo and Bahram 2019). AM fungi would not compete directly with 65 saprotrophic fungi, therefore it is expected that decomposition would be quicker in AM forests 66 compared to EcM forests, but this still remains an open question (Fernandez and Kennedy 2016; 67 Frey 2019). In fact, AM fungi may even enhance directly organic matter decomposition in some cases via a "priming effect", promoting the activity of free-living saprotrophs (Hodge 2017; Frey 68 69 2019). A better understanding of the roles that different mycorrhizal types play in organic matter 70 decomposition is thus needed.

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72 Because fungal types and taxa differ strongly in their vertical distribution, especially in well-73 stratified soil such as podzols (Dickie and others 2002; Rosling and others 2003; Bahram and 74 others 2015), the strength and direction of the Gadgil effect could vary across soil organic 75 horizons, yet most previous studies have only considered the uppermost litter layer. Strong vertical segregation of fungal guilds occurs across podzol profile: saprotrophic fungi dominate 76 77 the litter horizon, and can still be abundant in upper organic horizons where mycorrhizal fungi 78 increasingly dominate (Lindahl and others 2007; Clemmensen and others 2015; Carteron and others 2020). It is recognized that overlapping niches between different groups of fungi can 79 80 generate competition for soil resources (Bödeker and others 2016; Mujic and others 2016). 81 Therefore, the greatest potential for mycorrhizal fungi to inhibit saprotrophs, and thus slow down

82 organic matter decomposition, should lie just below the layer of fresh litter. It has been suggested 83 that these interactions might help to explain differences in the amount and vertical distributions 84 of soil C in EcM systems (Clemmensen and others 2013; Kyaschenko and others 2017) and between EcM- and AM-dominated forests at different depth or horizons (Phillips and others 85 86 2013; Soudzilovskaia and others 2015; Craig and others 2018). By competing with saprotrophs 87 for organic nutrients, EcM fungi may promote C accumulation more than AM fungi that cannot 88 directly access these resources. These vertically segregated interactions among fungal guilds need 89 to be better understood because they play an important role in regulating organic matter 90 accumulation (Frey 2019). 91 92 Local adaptation to microbial guilds based on soil properties could also be an important factor 93 influencing decomposition via what has been termed the "home-field advantage" (HFA; van der

94 Wal and others 2013) hypothesis. This HFA predicts that litter decomposition is faster in "home 95 soils" due to adaptation of the decomposer community to the chemical composition of the "home 96 litter" (Gholz and others 2000; Austin and others 2014). Using published data on mass loss from 97 125 reciprocal litter transplants, Veen and others (2015) have shown that this HFA increases 98 decomposition rates by 7.5% on average. However, the strength of the HFA might depend on the 99 context such as plant identity, litter quality and moisture level (Veen and others 2015; Wang and 100 others 2020). Some studies suggest that AM litter shows higher HFA than EcM litter (Midgley 101 and others 2015; Jacobs and others 2018). On the other hand, because EcM litter tends to be more 102 recalcitrant (Keller and Phillips 2019), it could be expected that EcM litter decays faster in EcM

103 forest with saprotrophs better adapted to decompose recalcitrant organic matter. In any case,

104 further investigation is needed to better understand the effect of microbial decomposers driving

the HFA depending on litter type and the stage of litter decomposition (Li and others 2020; Linand others 2020).

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108 The main objective of our study was to assess the impact of EcM and AM strategies on the 109 decomposition of soil organic matter in organic horizons in northern forests. First, we determined 110 stocks of C and nutrients in the upper 20 cm of soil in adjacent forest plots dominated by AM or 111 EcM trees. Then, we performed a litterbag experiment using a reciprocal transplant of organic 112 matter from AM and EcM forest enabling us to isolate site vs. organic matter quality effects on 113 decomposition. Litterbags were composed of different mesh size that allowed (44  $\mu$ m) or 114 excluded  $(1 \,\mu m)$  ingrowth of fungal hyphae. It is well established that fungal hyphae can move 115 freely across large pore-size (30-50 µm) mesh but prevents in-growth of roots, while small pore-116 size (0.5-1 µm) mesh further blocks fungal hyphae (Johnson and others 2001; He and others 117 2004; Teste and others 2009). Since mycorrhizal fungi development requires association with 118 living plant roots, only saprotrophs can develop inside the small-mesh litterbags and access 119 organic matter. Mycorrhizal fungi can colonize organic-rich soil (Lindahl and others 2007; Bunn 120 and others 2019) and, can even be abundant in organic horizons (this system; see Carteron and 121 others 2020). Therefore, large-mesh litterbags allow to follow the decomposition of organic 122 matter in the presence of external fungal hyphae, which are primarily mycorrhizal in these forests 123 other than for the most-recent superficial litter horizon where saprotrophs dominate (Carteron and 124 others 2020). Decomposition of the three upper organic horizons (litter - L, fragmented - F, 125 humic - H) was followed by measuring changes in soil mass, and changes in C and nitrogen (N) 126 over two years. In addition, the fate of C fractions was followed in decomposing L samples and 127 potential access of N by mycorrhiza in the F samples. We hypothesized that the impact of 128 mycorrhizas on organic matter decomposition would differ between AM and EcM forests. More

129 specifically, we expected based on the Gadgil effect hypothesis that EcM forests would store a 130 higher amount of C in the topsoil and show slower organic matter decomposition due to the 131 inhibition of saprotrophs by EcM fungi and lower litter quality, whereas these effects would be 132 smaller in AM forests. In addition, we hypothesized that the slowing down of C cycle by EcM 133 fungi would be strongest in the fragmented (F) horizon where litter-derived organic materials, 134 free-living saprotrophs, mycorrhizal fungi and roots coincide (Clemmensen and others 2013; 135 Cotrufo and others 2015; Carteron and others 2020). Due to microbial adaptations of the 136 decomposer community, we also hypothesized that litter would decompose fastest in their 137 "home" forests relative to "away" forests (Veen and others 2015). Specifically, mass loss of 138 organic matter from AM soil would be highest when incubated in AM forest and mass loss of 139 EcM organic matter highest in EcM forest.

140

#### 141 Material and methods

#### 142 Study area and site selection

143 Our study was conducted in a northern temperate forest at the Université de Montréal's field 144 station (Station de biologie des Laurentides, Saint-Hippolyte, Québec, Canada). The mean annual 145 temperature is 4.3 °C and total annual precipitation is 1195 mm, with ~25% falling as snow 146 (based on 1981–2010 data, meteorological station #7037310, Saint-Hippolyte). Soils consist of 147 podzols with moder humus formed from Precambrian anorthosite (Bélanger and others 2004; Courchesne and others 2005). We selected ten 20 m  $\times$  20 m plots from Carteron and others 148 149 (2020), either dominated by EcM or AM trees (Table S1), and grouped into five clusters or 150 "blocks" (n = 5 blocks, each containing one plot of each of the two mycorrhizal types, EcM and 151 AM). These pairs of EcM-AM sites were clustered together to minimize variation in 152 environmental conditions (e.g. slope, aspect, elevation) within each block. Previous root

153	colonization and molecular analyses on the same sites showed that forests dominated by EcM
154	trees had the highest EcM fungal abundances while forests dominated by AM trees had the
155	highest AM fungal abundances. Carteron and others (2020) also found strong shifts from
156	saprotrophic to mycorrhizal fungal dominance with increasing soil depth in both forest types,
157	especially across surface organic horizons.
158	
159	Soil carbon and nutrient stocks
160	Carbon and nutrient stocks were quantified by measuring C, N, phosphorus (P) concentrations
161	and thickness for all horizons in the upper 20 cm of soil, as reported in Carteron and others
162	(2020). Soil bulk density was measured simultaneously for the five horizons in three randomly-
163	positioned locations replicates per plot using an auger, and values from these locations were
164	averaged across sites. The horizons considered were litter (L), fragmented (F), humic (H), and
165	mineral horizons Ae and B.
166	
167	Organic matter collection
168	In each plot, organic matter samples were collected separately from the three organic horizons,
169	namely: L, F and young H (i.e. most recent layer) from two pits. Samples were homogenized by
170	horizon within each plot. Samples were collected in July 2016. A subsample from each horizon
171	by plot was preserved at 4 °C as inoculum (see below). Another subsample was oven-dried at 60
172	°C for 72 h and ground for chemical analyses. The rest of the organic matter was air-dried before
173	being used to fill the bags.
174	

175 Litterbag design

176 Litterbags were 15 cm  $\times$  15 cm in size and designed to have three compartments (L, F, H; in the 177 same order in which they occur through the soil profile) separated by 44 µm-pore polyethylene 178 mesh (PETEX® 07-40/12; Sefar Inc., Buffalo, NY, USA). Our use of 44 µm-pore mesh ensured 179 that hyphae could grow across compartments within each bag, an important process for 180 decomposition (i.e. to allow for translocation of nutrients and C across horizons), while still 181 keeping L, F, and H horizons separate for later retrieval. The outer mesh of the litterbags was 182 made with either the same 44  $\mu$ m-pore polyethylene mesh described above or 1  $\mu$ m-pore mesh 183 from the same material (PETEX® 07-1/2; Sefar Inc., Buffalo, NY, USA). Large pore size (30-50 184 µm) mesh has been widely used to assess the effect of mycorrhizal hyphal colonization since 185 more than a decade ago (Johnson and others 2001), by excluding fine roots but not fungal hyphae 186 (He and others 2004; Teste 2008). Thus, our litterbags made with 44-µm pore size mesh allow to 187 study decomposition in the presence of mycorrhizal hyphae (and other saprotrophic fungi located 188 outside of the bag). By contrast, the small 1-µm pore size mesh prevents most external fungal 189 hyphae to grow through the litterbag (Teste and others 2006). Because most mycorrhizal hyphae 190 cannot grow within the bag (as mycorrhizal fungi are obligate biotrophs), this bag design allows 191 us to study organic matter decomposition in the absence of mycorrhizal fungi. Litterbags of 50 192 µm-pore size mesh have been found to allow ingrowth of mycorrhizal fungi (i.e. Teste and others 193 2006; Sterkenburg and others 2018), which are abundant in our F and H horizons of our plots 194 (Carteron and others 2020). By contrast, free-living saprotrophic fungi should be present in all 195 bags (of 1-µm and 44-µm pore sizes) since all bags were inoculated with horizon-specific organic 196 matter from the same plot prior to being installed in the field. For this reason, while we recognize that the 1-µm mesh bags exclude all external hyphal ingrowth (mycorrhizal and free-living), for 197 198 simplicity we refer to this treatment as "mycorrhizal exclusion" hereafter since 1-µm mesh bags

exclude this particular fungal guild. Mycorrhizal fungal hyphae should be present in the 44-um 199 200 mesh bags, being a very important component of the fungal community in the soil other than for 201 the L horizon (Carteron and others 2020). Polyethylene mesh was selected over nylon mesh (e.g. 202 NITEX®, Sefar Inc. Buffalo, NY, USA) because it is much more resistant to degradation when buried in soil (Colin and others 1981). Microscope observations showed no evidence that the 203 204 bags were breached after two years. Previous studies using 0.5 µm-pore size mesh litter bags 205 found that environmental conditions, particularly moisture levels, were similar inside and outside 206 the bags (Allison and others 2013) and, that soil water moved freely across the mesh within 207 minutes (Teste and others 2009). Our own observations confirmed that water moved freely across 208 membranes of both mesh size via capillary action as long as there was contact between the litter 209 inside the bag and the membrane itself; such conditions were maintained throughout the field 210 experiment since the litterbags were buried under the litter layer and secured firmly on the ground. The 1 and 44  $\mu$ m-pore size mesh have air permeability values of > 95 ±15 l.(m<sup>2</sup>.s)<sup>-1</sup> at 211 212 200 Pa (provided by Sefar inc.). In total, 160 litterbags were used (Fig. S1), from which 90 213 prevented most hyphal ingrowth of all external fungi. Each bag was stored within a 1-mm mesh 214 nylon bag to provide additional physical protection for the less robust 44- or 1-µm PETEX® 215 mesh.

216

#### 217 *Litterbag preparation and collection*

Weighed air-dry organic matter was transferred to litterbags (2.85 g for L and 4.75 g for F and H horizons). Horizon specific fresh inoculum (~5 % of dry-weight equivalent) was added to each horizon from the receiving plot to ensure that plot-specific microbial biota, including free-living saprotrophic fungi, could colonize each litterbag. Water content was determined from oven-dried

222	sub-samples at 60 °C for dry-mass inoculum conversion. Filled litterbags were put back in situ
223	October 2016, directly on top of the H horizon (with L horizon facing up) and covered by a thin
224	layer of fresh litter. Litterbags were secured on the ground with small stakes and tied together
225	with nylon fishing line to a central stake to facilitate retrieval of bags. Two spatial replicates
226	within each plot were installed. A total of 160 bags were collected after one and two years of
227	residence (i.e. field incubation) for 480 samples analyzed (Fig. S1).
228	
229	Soil analysis
230	Initial subsamples of ground horizons L, F and H were weighed (5.0, 6.0 and 7.0 mg $\pm$ 0.2
231	respectively) and analyzed to estimate C and N contents by dry combustion in a CN analyzer
232	(Vario Micro Cube; Elementar, New-Jersey, United States,
233	dx.doi.org/10.17504/protocols.io.udces2w). The concentrations of soluble cell contents (e.g. non-
234	structural carbohydrates), hemicellulose, cellulose and lignin (% dry weight) were also
235	determined on these initial samples by sequential digestion (Fiber Analyzer 200; ANKOM
236	technology, dx.doi.org/10.17504/protocols.io.yinfude). After one and two years, organic matter
237	samples were retrieved from litterbags, oven-dried at 60 °C for at least 72 h and then weighed to
238	estimate mass loss percentage. These samples were then ground with a cyclone mill (Cyclone
239	Sample Mills, UDY Corporation, Colorado, United States), using a 2-mm screen. Concentrations
240	of C and N were also determined using the method described above. Thirty subsamples of the
241	initial horizons, and all the F horizons after two years of residence were analyzed for $\delta^{15}N$ with a
242	Micromass model Isoprime 100 isotope ratio mass spectrometer coupled to an Elementar Vario
243	MicroCube elemental analyser in continuous flow mode.
244	

245 Statistical analyses

Differences in organic matter stocks among forest types were evaluated using a linear mixed-246 247 effects model with forest type (AM or EcM as soil provenance) as a fixed factor and block as a 248 random factor. Horizon was added as fixed factor for the modeling of initial soil chemistry. To 249 predict the changes in mass (within the litterbags), linear mixed-effects models were also used by 250 adding as fixed factors outside fungal hyphae (i.e. size of mesh pore) excluded (1 µm) or not (44 251 μm). Finally, forest of residence (AM or EcM forest) and time (one or two years) were added as 252 fixed factors to compare decomposition in the two forest types including relevant interactions 253 among fixed factors (see Table S2 for more details). Models were compared using the Akaike 254 information criterion corrected for small sample size (AIC $_{\rm c}$ ). Validation of the models was done by visual inspection of the residuals. Spatial replicates within one plot were averaged prior to 255 256 analyses. Eleven bags with damaged mesh were removed from the analysis. Statistical analyses were performed using the R software (R Core Team 2018) and the following packages *dplyr* 257 258 (Wickham and others 2017), emmeans (Lenth 2019), ggplot2 (Wickham 2016), ggpubr (Kassambara 2018), nlme (Pinheiro and others 2012). Data and R scripts can be found at 259 260 https://github.com/alexiscarter/decompo myco.

261

## 262 **Results**

263 Organic matter stocks

Stocks of C were higher in EcM forest stands compared to AM stand within the upper 20 cm of soil (one-way analysis of variance, P < 0.001; Fig. S2) as observed in the organic horizons (Table 1). Stands dominated by EcM trees stored 14% more C than AM stands in surface soils. The soil C:N ratio also differed among forest types, with higher values in EcM stands (P = 0.024; Fig. S3). By contrast, there were no differences in soil C:P ratio among forest types (Fig. S4).

269

#### 270 Initial soil chemistry

271	Soil C, cellulose and hemicellulose concentrations decreased from L to H horizons in both forest
272	types, while lignin was highest in the F horizon and in EcM stands overall (23% in AM forest
273	and 26 % in EcM forest; Table S3). By contrast, soil total [N] increased slightly with soil depth in
274	both forest types (Table S3). As a result, soil C:N and lignin:N ratios were higher in EcM forest
275	for the three organic horizons compared to AM forest (Table S3). $\delta^{15}N$ values showed similar
276	increases from L to H horizons in both forest types but the F horizon in EcM forest was slightly
277	enriched (but not significantly, $P = 0.224$ ; Table 1). Horizons tended to be thicker in EcM forest
278	(Table 1).

279

## 280 Effect of residence on decomposition: AM vs. EcM forests

281 In AM and EcM stands, older (i.e. deeper) horizons decomposed more slowly than younger ones 282 (Fig. 1). Organic matter loss was slower in the litterbags of 1 µm-pore size mesh in all horizons 283 of both types of forests. However, the slowing down of decomposition due to mycorrhizal fungal 284 exclusion was only statistically significant in the F horizons in stands dominated by AM (-3.7 %, 285 P = 0.02; Fig. 1a) and EcM (-4.4 %, P = 0.019; Fig. 1b). Differences in the effects of the 286 mycorrhizal exclusion treatments among forest types increased between one and two years of incubation (Fig. 2). Overall, decomposition was slower in AM compared to EcM stands, ranging 287 288 from -0.8% (P > 0.05) of mass loss after one year to -3% (P < 0.001) after two years of 289 incubation. After two years, decomposition of organic matter originating from EcM and AM soils 290 was higher in EcM stands (Fig. 2).

291

#### 292 Effect of provenance: AM vs. EcM forests

293	Litter originating from AM stands decomposed more quickly than EcM litter after one year but
294	this was no longer the case after two years ( $P > 0.05$ ; Fig. S5). While changes in the litter C:N
295	ratio remained similarly low for both soil origins (~1, Fig. S6), the EcM litter showed a clear
296	increase (~1.2) suggesting a lower loss in N compared to C. Similarly, changes in lignin:N ratio
297	were also stronger for EcM litter (Fig. S7).
298	

299 Effect of provenance and residence on C fractions and N

300 Mycorrhizal exclusion did not affect concentrations of soluble contents nor hemicellulose in litter

301 incubated in AM and EcM stands. Compared to EcM stands, decomposition was slower in AM

302 stands for litter cellulose (-4.12 %, P = 0.001; Fig. S8) and lignin (-6.56 but P > 0.05; Fig. S9).

303 Mycorrhizal exclusion slowed down decomposition of cellulose (-3.79 %, P = 0.003) and lignin

304 (-5.13% but P > 0.05). Overall, N loss was reduced by mycorrhizal exclusion (-2.83 %, P < 0.05).

305 0.001) and reduced in AM stands (+2.7 %, P < 0.001). After two years, <sup>15</sup>N enrichment was

higher in EcM stands (F horizons only, P = 0.046) but effect of mycorrhizal exclusion was rather

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307

#### 309 Discussion

low (Fig. S10).

310 No evidence of a Gadgil effect in either forest mycorrhizal type was observed. In fact, the

311 opposite effect was observed, in that decomposition was faster in the presence of EcM or AM

fungi than in their absence. Contrary to our hypothesis, decomposition was faster in EcM- than in

- 313 AM-dominated forests. However, as predicted, decomposition was higher in upper horizons (i.e.
- 314 "younger" soil), and the net effect of the external fungal network on decomposition was
- 315 significant in the fragmented (F) horizons. The F horizon is located just below the litter (L),
- 316 where most decomposition studies tend to focus. Our results suggest that the Gadgil effect is not

317	a universal pattern in EcM forests, and that mycorrhizal fungi may actually accelerate rather than
318	slow down decomposition (Frey 2019). In agreement with these results, we found that
319	decomposition was faster in EcM forests regardless of organic matter origin, suggesting an HFA
320	in EcM but not AM forests.
321	
322	Several abiotic and biotic factors can impact litter decomposition, such as climate and soil fauna
323	(Hättenschwiler and others 2005; Steidinger and others 2019). However, given the importance of
324	fungi in soil decomposition processes, there has been much interest in exploring the potential

325 effects of interguild fungal interactions over C and nutrient dynamics (Dighton and others 1987;

326 Verbruggen and others 2017). Mycorrhizal fungi can inhibit saprotrophs by competing for

nutrients, resulting in slower organic matter decomposition and promotion of C accumulation

328 (Frey 2019). We took advantage of a natural experiment of co-occurring patches of AM and EcM

329 trees under similar environmental conditions but distinct fungal communities and soil chemistry

330 (Carteron and others 2020) to test if contrasting mycorrhizal strategies exerted different control

on organic matter decomposition (Phillips and others 2013; Dickie and others 2014). However,

contrary to the Gadgil effect hypothesis, our results showed that both EcM and AM fungi

accelerate organic matter decomposition in this northern deciduous forest. This might occur if the

overall positive effect of mycorrhizal hyphae and other external fungi on decomposition was

335 greater than any potential negative impacts of competition with saprotrophs. In addition,

336 mycorrhizal fungi combined with their local microbial community in EcM forests tended to

degrade cellulose and lignin more quickly compared to AM forests. By isolating the effect of

338

decomposition tends to be higher in EcM than AM forests regardless of soil origin and incubation

mycorrhizas, microbial communities and local environmental conditions, our study shows that

time. Our results challenge the view that EcM fungi slow down litter and soil decomposition

compared to AM fungi (Tedersoo and Bahram 2019 and references therein). They also suggest
that more attention should be paid to priming *vs.* inhibitory effects of different mycorrhizal types
on the decomposition of organic matter (Kuzyakov 2010).

344

345 Ectomycorrhizal fungi have traditionally been suggested to slow down litter decomposition via 346 their negative competitive effects on free-living saprotrophs (Gadgil and Gadgil 1971; Fernandez 347 and Kennedy 2016). In our field experiment, we have found that EcM fungi in fact accelerated 348 the decomposition across the three upper organic horizons over two years, particularly in the 349 fragmented (F) horizon. Fernandez & Kennedy (2016) suggested a number of important 350 environmental factors that could modulate the inhibiting impact of EcM fungi on free-living 351 saprotrophs, which might help to explain our results. First, organic matter recalcitrance was 352 relatively low in this broadleaf forest, with lignin:N ratios below 20. Similarly, the C:N ratio was 353 below 30, making N less limiting for saprotrophs compared to other studies where a Gadgil effect 354 was observed (Smith and Wan 2019). Secondly, the studied podzols were well-stratified and 355 exhibited a strong vertical segregation with distinct fungal communities with strong shifts from 356 saprotrophic to mycorrhizal fungal dominance with increasing soil depth (Carteron and others 357 2020), thereby reducing opportunities for interguild competition. Finally, decreased soil moisture 358 due to EcM fungi can impede decomposition processes (Koide and Wu 2003), but our system is 359 located in a northern temperate forest characterized by a humid continental climate with 360 precipitations throughout the year, where water is not thought to be limiting. At least three other 361 experimental studies have found a positive combined effect of roots and EcM fungi on decomposition (Zhu and Ehrenfeld 1996; Subke and others 2011; Malik 2019). In our case, it is 362 363 worth noting that the strongest positive net effect was observed in the fragmented horizon where 364 there are: (i) High root colonization by EcM and AM fungi, (ii) high abundance of saprotrophic

and mycorrhizal fungi and (iii) high fine root density (Carteron and others 2020). Most studies
that have studied the impact of mycorrhizas on decomposition have focus on the most recent litter
(L) layer, whereas here we show that important processes occur in deeper (organic) horizons. Our
results suggest that vertical stratification should be taken into account to better understand the
effect of mycorrhizas on the decomposition process.

370

371 Arbuscular mycorrhizal fungi are known to produce compounds that can, for example, alter 372 microbial community or promote soil aggregation thus modulating decomposition rate (e.g. 373 Hodge and others 2001; Gui and others 2017; Xu and others 2018). Decomposition can even be 374 reduced by AM fungi, potentially through antagonistic interactions with free-living saprotrophs 375 (Leifheit and others 2015; Carrillo and others 2016). In our field experiment, we found no 376 evidence of a Gadgil effect exerted by AM fungi that would counterbalance their positive impacts 377 on decomposition. As expected, decomposition in the upper three organic horizons in AM forest 378 was not reduced with the 1-µm mesh bags (in which mycorrhizal hyphae were excluded), but in 379 fact tended to increase. Given that AM fungi lack a strong degradation machinery (Tedersoo and 380 Bahram 2019), our results support the view that priming of organic matter decomposition might 381 be an important nutrient acquisition strategy for them (Wurzburger and Brookshire 2017). 382 Greater priming in AM systems may result from AM fungal necromass and the lack of genetic 383 capacity from AM fungi to directly access organic nutrients (Frey 2019). It is worth noting that 384 with the 1-µm mesh bags, decomposition in AM forests tended to be slower than in EcM forests, 385 suggesting that their free-living saprotrophic communities have different capacities to degrade organic matter (see results after two years in Fig. 2). Microbial communities in AM forest might 386 387 be less efficient at degrading organic matter due to their more easily-decomposed litter contrary 388 to what has been observed for other AM systems in microcosm experiments (Taylor and others

2016). Evaluating the response of the saprotrophic community using molecular tools over longterm experiment would be an interesting way to better understand decomposition processes *in situ*, in order to complement studies that focus on laboratory manipulations of mycorrhizal abundance (Verbruggen and others 2017). It would also allow us to experimentally assess if the abundance of saprotrophs shifts in deeper horizons when AM and EcM fungi are excluded (e.g. Lindahl and others 2010; Sietiö and others 2019).

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396 Leaf litter decomposition rates are known to be positively linked with initial N concentration and 397 inversely with lignin (Prescott 2005; Berg and McClaugherty 2014). Overall net effect of 398 mycorrhizas over decomposition is known to be controlled by substrate quality and local 399 microbial community composition (Fernandez and others 2019; Smith and Wan 2019). As 400 expected, we found a strong effect of soil depth with deeper (i.e. "older") horizons decomposing 401 more slowly. In general, the litter of EcM-associated trees tend to have a lower quality than AM 402 trees such as lower C:N and lignin:N ratios (Lin and others 2017), which may drive soil C 403 accumulation in the short-term. In our field sites, litter in EcM stands had lower quality compared 404 to AM stands. The EcM stands were mostly composed of American beech. However, American 405 beech litter is less recalcitrant than many conifers (Moore and others 1999), which may explain 406 discrepancies with other studies from coniferous EcM forests in which Gadgil effects have been 407 observed (Fernandez and Kennedy 2016; Smith and Wan 2019). In temperate forests, AM plants tend to produce leaf litter that decomposes more rapidly in situ than that of EcM plants (Keller 408 409 and Phillips 2019). Similarly, litter originating from AM patches decomposed more quickly than 410 EcM litter after one year but interestingly, this was not the case after two years in our experiment. 411 This is consistent with previous studies showing that sugar maple leaf litter tend to decomposes 412 more quickly during the first years after senescence (McHale and others 1998; Lovett and others

413 2016), but tends to be more similar to American beech after several years (within standard error 414 range, see Lovett and others (2016). Community of decomposers in EcM forests may be efficient 415 at decomposing recalcitrant organic matter (Fernandez and others 2019). Contrary to the results of Midgley and others (2015) obtained from another study system, we observed HFA in EcM 416 417 forests but not in AM forests. The efficiency of the microbial decomposers present in EcM soil 418 for decomposing organic matter may be high regardless of litter type and quality. Furthermore, 419 we found no evidence that fragmented (F) and humic (H) horizons in AM stands decomposed 420 faster than the same horizons in EcM stands (but see Jacobs and others 2018). Taken together, 421 these results suggest that the significant impact of initial litter chemistry on decomposition 422 diminishes after the first year of decomposition and that microbial decomposer community may 423 adapt to "home" substrate quality.

424

425 Reducing decomposer diversity reduces litter decomposition rate (Handa and others 2014; Li and others 2020), but this effect is context-dependent and the effect of soil fauna is variable across 426 427 focal species (Makkonen and others 2012). Smaller mesh size are known to reduce the potential 428 diversity of soil fauna that are important for decomposition processes (Hättenschwiler and others 429 2005). In our study, patches were mainly composed of American beech or sugar maple, and 430 previous studies indicate that maple litter is generally preferred over beech litter by the soil fauna (Hättenschwiler and Bretscher 2001; Jacob and others 2010). However, the difference in 431 decomposition between American beech and sugar maple seems to decrease over time (Lovett 432 433 and others 2016) and to be dependent on stand type of incubation (Côté and Fyles 1994). Unlike most studies, we used litterbags that were designed to follow decomposition of the upper three 434 435 organic horizons while avoiding soil trenching and tree girdling which confound the effects of 436 roots and mycorrhizal fungi (Fernandez and Kennedy 2016). Trenching is the historical and most

437 widely used method to test the Gadgil effect, but it is known to directly affect soil drainage, 438 increase soil moisture by impeding root water uptake and strongly disturb the system (Gadgil and 439 Gadgil 1971; Fisher and Gosz 1986; Fernandez and Kennedy 2016). Tree girdling is the most 440 extreme alternative as it kills trees, also preventing further research on the same site. In our 441 experiment, the initial disturbance may have increased labile C but the persistence of this effect 442 after two years was assumed to be rather limited. Furthermore, the observed effects of our 443 exclusion treatment on mass loss increased between the first and second years of incubation 444 suggesting persistent biological effects. Decreasing mesh size might have decreased soil moisture 445 but we observed no impact on litter soluble content losses suggesting a rather low effect caused 446 by mesh size, at least on the most labile fractions of C. It is possible that the use of litterbags with 447 small mesh size limited the exposure to biophysical perturbations, which might hamper mass loss 448 (Prescott 2005; Berg and McClaugherty 2014), but this was common to all treatments. To better 449 predict soil C processes and stocks, more research may be needed to understand how interaction 450 between mycorrhizas, soil fauna, plant inputs and variables such as soil moisture, or bulk density 451 impact decomposition (Lin and others 2017).

452

453 Our sampling design allowed us to spatially distinguish decomposition processes in the upper 454 three horizons and assess the fate of young to older organic matter overcoming some limits of 455 short-term experiments. The overall net effect of mycorrhizas on decomposition was positive regardless of mycorrhizal type, but varied throughout the soil profile. Further analyses would 456 457 allow us to better understand if the greater decomposition could be due to higher microbial biomass inside the 44 µm-pore mesh bags, leading to higher enzymatic activities. As expected 458 459 from previous studies (e.g., Averill and others 2014; Soudzilovskaia and others 2019), C stocks 460 were greater in EcM stands compared to neighboring AM stands in this northern temperate forest

461 even though decomposition was greater in EcM soils, and positively influenced by the broader 462 fungal network. This indicates the potential importance of others factors such as litter quantity, soil fauna and moisture level in regulating C dynamics. The quality and composition of litter is 463 important for short-term C release, but the microbial community, including root-associated fungi 464 465 and mycorrhizal-associated organisms (Netherway and others 2020), potentially have a strong 466 impact on a longer-term which is important for C sequestration (Cotrufo and others 2015). 467 Overall, our study shows that forests dominated by different mycorrhizal strategies have distinct 468 impacts in soil organic matter dynamics. The EcM forests store higher soil organic C but support 469 microbial decomposer communities that are more efficient at degrading organic matter than those 470 of adjacent AM forests, rejecting the Gadgil effect as a driver of C accumulation in the northern 471 temperate forests.

472

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483

## 484 Authors' contributions

- 485 EL and AC conceived the ideas and designed methodology; AC and FC collected and analyzed
- the data; AC and EL interpreted the results; AC led the writing of the manuscript. All authors
- 487 contributed critically to the drafts and gave final approval for publication.

488

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#### 745 **Conflict of Interest**

The authors declare that they have no conflict of interest.

## 747 Table legends

- **Table 1.** Initial soil chemistry characteristics: C:N ratio, lignin:N ratio,  $\delta^{15}$ N, thickness and
- carbon stocks of the upper three horizons (litter L, fragmented F, humic H) of arbuscular
- 751 mycorrhizal (AM) and ectomycorrhizal (EcM) forest. Means and standard deviation are shown (*n*
- 752 = 5).

	Horizon	AM-dominated	Standard	EcM-dominated	Standard	
		forest	deviation	forest	deviation	
C:N ratio	L	22.71	2.53	27.36	1.81	
	F	20.50	1.01	22.32	1.36	
	Н	18.41	0.85	20.09	1.48	
lignin:N ratio	L	9.68	1.40	13.05	1.74	
	F	10.70	1.69	12.57	0.99	
	Н	8.62	1.30	10.50	1.50	
δ <sup>15</sup> N (‰)	L -3.26		0.36	-3.16	0.63	
	F	-2.48	0.40	-1.96	0.48	
	Н	-0.72	0.59	-0.74	0.36	
Thickness (cm)	L	0.71	0.22	0.81	0.30	
	F	2.36	0.68	3.44	0.65	
	Н	4.21	0.95	7.19	2.73	
Carbon stock	L	0.89	0.28	1.05	0.43	
$(kg.m^2)$	F	2.68	0.76	4.24	0.86	
	Н	2.97	0.67	6.99	4.00	

## 754 Figure legends

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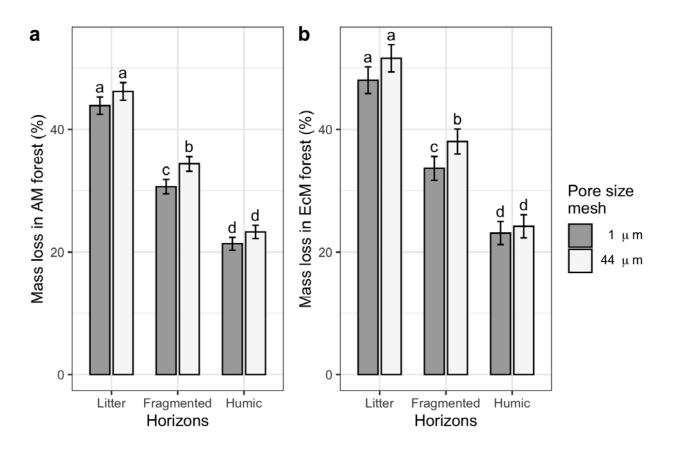
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- dominated by (a) arbuscular mycorrhiza (AM) or (b) ectomycorrhiza (EcM) in litterbags with
- pore mesh size of 1  $\mu$ m (grey bars) and 44  $\mu$ m (white bars). Means  $\pm$  1 SE are shown (n = 20).
- 759 Multiple comparison using Tukey's honestly significant difference post-hoc test, different letters
- 760 within each panel indicates significant differences (*P*-value < 0.05).

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**Figure 2.** Percentage of mass loss after one and two years of incubation in forests dominated by arbuscular mycorrhiza (AM) or ectomycorrhiza (EcM) in litterbags with pore size mesh of 1  $\mu$ m (top panels) and 44  $\mu$ m (bottom panels) and organic matter provenance from AM and EcM. Means  $\pm$  1 SE are shown (n = 30). Multiple comparison using Tukey's honestly significant difference post-hoc test, different letters indicates significant differences (*P*-value < 0.05).

# 767 **Figure 1**



# 769 **Figure 2**

