

Relation of inflammatory marker trajectories with frailty and aging in a 20-year longitudinal study

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1 **Abstract**

2 Little is known about the development of low-grade inflammation with age and its relationship with the
3 onset of frailty. In this exploratory study, we investigated 18 inflammatory markers measured in blood of
4 144 individuals aged 65-75 years at study endpoint, collected over 20 years at five-year intervals. IFN γ -
5 induced markers and platelet activation markers changed in synchrony over time. Chronically elevated levels
6 of IL-6-related markers, such as CRP and sIL-6R, were associated with frailty and becoming frail over time,
7 poorer lung function, or less physical strength. Overweight was a possible driver of these associations. More
8 and stronger associations were detected in women, such as between increasing sCD14 levels and frailty,
9 indicating possible monocyte overactivation. Multivariate prediction of frailty showed low accuracy but
10 confirmed the main results. In summary, we documented 20-year temporal changes of inflammatory markers
11 in an aging population, and related these to clinically relevant health outcomes.

12 **1 Introduction**

13 Understanding the aging process is important for finding ways to prevent or delay age-related morbidity and
14 thus to ensure good quality of life for an elderly population that is rapidly expanding worldwide (United
15 Nations, 2017). One factor that may relate to ‘successful’ aging is an adequately functioning immune system,
16 since an efficient immune response is essential to combat pathogens. Inflammatory processes can be out of
17 balance in the elderly, leading to persistent low-grade inflammation (Baylis et al., 2013; Franceschi et al.,
18 2017). It is thought that those with long-lasting low-grade inflammation have reduced responses to pathogens
19 and carcinogenesis, and that they have more auto-inflammatory responses, and therefore being more prone
20 to developing age-related diseases and to becoming frail (Baylis et al., 2013; Franceschi et al., 2017). One
21 potential driver of chronic low-grade inflammation could be the amount of body fat, since adipocytes are
22 thought to activate the immune system directly (Ghigliotti et al., 2014).

23 It is still largely unknown when and how low-grade inflammation develops during aging and how it is related
24 to frailty. The few longitudinal studies performed on this subject showed that frail people often had low-
25 grade inflammation for a long period of time (Gale et al., 2013; Puzianowska-Kuźnicka et al., 2016; Samson
26 et al., 2019; Walker et al., 2018). In most studies, including our own (Samson et al., 2019), the presence of
27 chronic low-grade inflammation was defined by the plasma concentrations of only one or two inflammatory
28 markers, notably CRP and IL-6. While these are the markers most commonly used to investigate low-grade
29 inflammation (Puzianowska-Kuźnicka et al., 2016), inflammation is a complex process in which many proteins
30 are involved. Some studies already suggested that looking at a larger panel of inflammatory biomarkers,
31 including a broader range of (chemotactic) cytokines, would improve the understanding of the relationship
32 between low-grade inflammation and age-related diseases (Morrisette-Thomas et al., 2014). In addition,
33 taking sex differences into account while studying the processes seems relevant because women generally
34 have a higher frailty index score than men but do reach a higher age (Gordon et al., 2017).

35 In order to gain more insight into long-lasting low grade inflammation in relation to frailty and differences
36 therein between men and women, we performed an exploratory study using data and blood samples from
37 a selection of participants (n=144) of the longitudinal Doetinchem Cohort study. Blood samples and data
38 were collected at 5-year intervals covering a period of approximately 20 years. We had several aims in our
39 study. First, we wanted to explore the development of low-grade inflammation in an aging population, by
40 investigating if and how concentrations of multiple inflammatory markers change with age. Secondly, we
41 wanted to know if exposure to low-grade inflammation during a prolonged period of time relates to frailty
42 or becoming frail, and to more specific aging-related clinical outcomes, such as decreased physical fitness

43 (handgrip strength) and decline of lung function (spirometry). Lastly, we investigated if overweight is an
44 important factor in these relationships.

45 2 Results

46 2.1 Study population characteristics

47 The study group consisted of 144 participants, (73 men and 71 women; Figure 1), with an average age of
48 68.3 years (min: 59.7, max: 73.5 years) at the endpoint of follow-up (Table 1). Average follow-up time was
49 19.3 years (min: 14.4, max: 20.9 years). Since blood samples were taken every five years, a maximum of
50 five samples per individual was available for analysis, which was the case for most participants (n=107).
51 Of the other study participants, either four samples (n=34) or three samples (n=3) were available. Highest
52 concentrations were in the order of 10^6 pg mL⁻¹, for example of soluble CD14 (sCD14)(average endpoint level:
53 $2.20 * 10^6$ pg mL⁻¹) and C-reactive protein (CRP) ($1.22 * 10^6$ pg mL⁻¹) (Table S1), and lowest concentrations
54 were around 1 pg mL⁻¹, for example of interleukins such as IL-10 and IL-6 (average endpoint level: 0.65 and
55 2.31 pg mL⁻¹, respectively). Concentrations of sIL-2R and IL-1 β , were below detection limit in > 40% of
56 the cases, and therefore these were excluded from analysis, leaving a total of 18 inflammatory markers for
57 investigation (Table S1).

Table 1: Baseline characteristics of the study population

	n	144
Women, %(n)		49.3 (71)
Age at baseline (yrs.)	49 (SD: 2.6, range: 42.4 - 53.5)	
Age at endpoint (yrs.)	68.3 (SD: 2.8, range: 59.7 - 73.5)	
Mean follow-up timespan (yrs.)	19.3 (SD: 1.8, range: 14.4 - 20.9)	
BMI	26.5 (SD: 4, range: 20.5 - 46.8)	

Note:

Numbers are mean (SD) unless otherwise stated

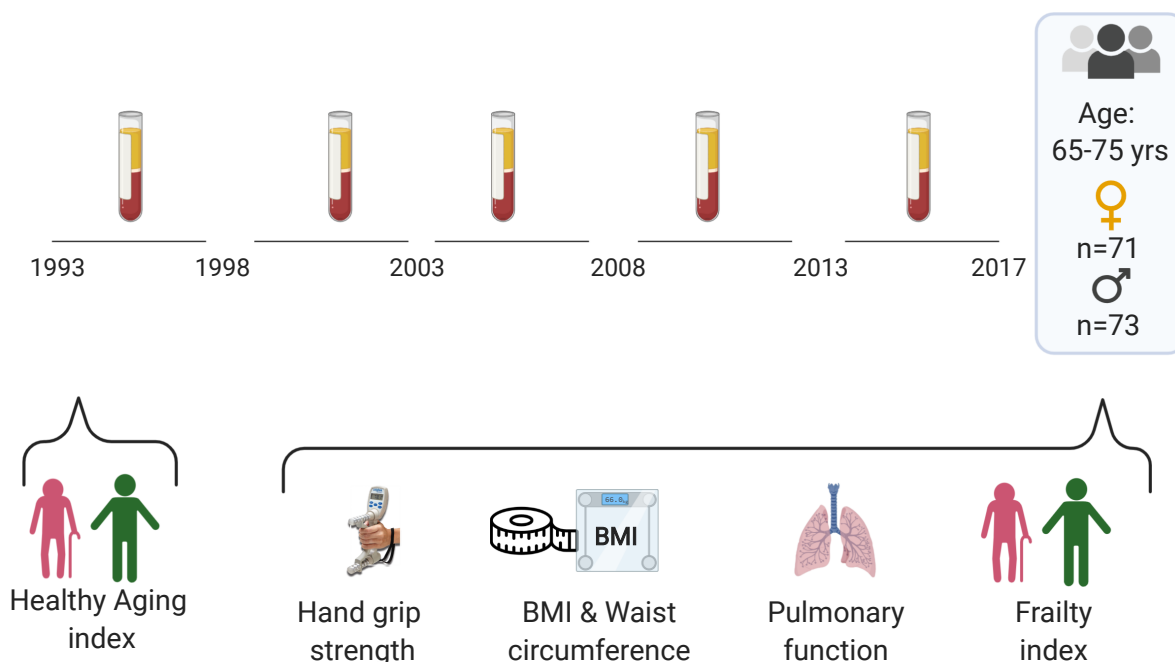


Figure 1: Timeline of the study. Participants of 65-75 years of age have been followed for more than 20 years, plasma samples were stored at 5-years intervals. At start, healthy aging index scores were determined. At last measurement, a frailty index on basis of 35 parameters including lung function and handgrip strength were determined which are used for separate analysis.

58 2.2 Sex-specific changes in concentrations of inflammatory markers during ag- 59 ing

60 We calculated the average inflammatory marker levels over 20-year follow-up by means of the area under the
61 concentration versus time curve (AUC) for each individual and each inflammatory marker separately. The
62 AUC values differed between men and women for CCL5/RANTES, showing continuously higher levels in
63 women (Figure S1). Several other dissimilarities between men and women became apparent when investigat-
64 ing the influence of age on the inflammatory marker levels. In men and women, an increase during aging was
65 observed in the levels of CXCL10/IP-10, CXCL11/I-TAC and CCL27/C-TACK (Figure 2). In women, this
66 increase was also seen for CRP, sIL-6R, CCL2/MCP-1, CCL11/Eotaxin, and sCD14, until age 60 approx-
67 imately, after which age the trajectories of inflammatory markers in men and women appeared to become
68 more similar (Figure 2). Indeed, women showed higher concentrations of CCL5/RANTES and BDNF, and
69 lower concentrations of CCL11/Eotaxin and CCL2/MCP-1 at study baseline, while no differences were found
70 between men and women in inflammatory marker levels at study endpoint (data not shown). The increase
71 in inflammatory marker levels in women could be influenced by the major hormonal changes associated with
72 menopause, since most women have their menopause before 60 years of age. Therefore, we compared the
73 levels of inflammatory markers just before the menopause with those shortly after menopause in all women

74 of whom these data were available at both timepoints (40 out of 70 women; Figure S2). Average self-reported
75 age of menopause was 50.3 years (95% CI 39.7-60.9), and average time between measurement before and
76 after menopause was 5.3 years (95% CI 2.6-8.1). Concentrations were higher after menopause for the inflam-
77 matory markers CCL2/MCP-1, CCL11/Eotaxin, sGP130, CCL27/C-TACK, and CXCL10/IP-10. Thus, our
78 data suggest that inflammatory marker trajectories differ between men and women but become more similar
79 with older age, which is possibly in part explained by hormonal changes in women during menopause.

80 **2.3 Correlations between inflammatory markers at study endpoint**

81 Testing of the correlations between inflammatory markers at endpoint revealed multiple associations (Figure
82 3A). All detected associations were positive, except for CCL27/C-TACK with P selectin, BDNF, sCD40L,
83 and IL-10, and P selectin with sCD14. The strongest positive associations ($\rho > 0.50$) were observed for
84 IL-6 with IL-10 ($\rho = 0.72$), IL-6 with sCD40L ($\rho = 0.58$), IL-6 with CXCL9 ($\rho = 0.53$), sCD40L with CXCL9
85 ($\rho = 0.50$), and for BDNF with CCL5/RANTES ($\rho = 0.68$). These associations were found both in men and in
86 women but were stronger in women (Figure S3A,B). Other positive but weaker associations were seen between
87 the structurally related IFN γ -inducible chemokines CXCL9, CXCL10/IP-10 and CXCL11/I-TAC (Figure
88 3A); again, these associations were stronger in women than in men (Figure S3A,B). Positive associations
89 were found between BDNF and most of the CXCL- and CCL- chemokines (CCL1/I-309, CCL2/MCP-1,
90 CCL5/RANTES, CCL11/Eotaxin, CXCL9, CXCL11/I-TAC) when men and women were analyzed together
91 and stratified by sex (Figure 3A). When analyzing men and women separately, these were detected only in
92 women (Figure S3A,B). Furthermore, weak correlations were found between several inflammatory markers
93 involved in the IL-6 pathway, such as between sIL-6R and sGP130 ($\rho = 0.20$), and between sGP130 and IL-6
94 itself ($\rho = 0.12$) (Figure 3A). A weak association was found between higher levels of CRP and higher levels
95 of sIL-6R ($\rho = 0.18$) as well as with higher levels of CCL11/Eotaxin ($\rho = 0.20$) (Figure 3A). When analyzed
96 separately by sex, these relationships were detected only in women (Figure S3A,B).

97 **2.4 Similarities between inflammatory markers in terms of temporal changes** 98 **over 20 years**

99 Since there is high variability between individuals in concentrations of inflammatory markers, a cross-sectional
100 analysis is somewhat restricted. Therefore, we looked for synchronous changes of biomarkers, meaning that
101 when biomarkers change together this is probably caused by an underlying biological process (in response to a
102 stimulus, or under homeostasis) inducing these changes. To do so, we determined in which of the four possible

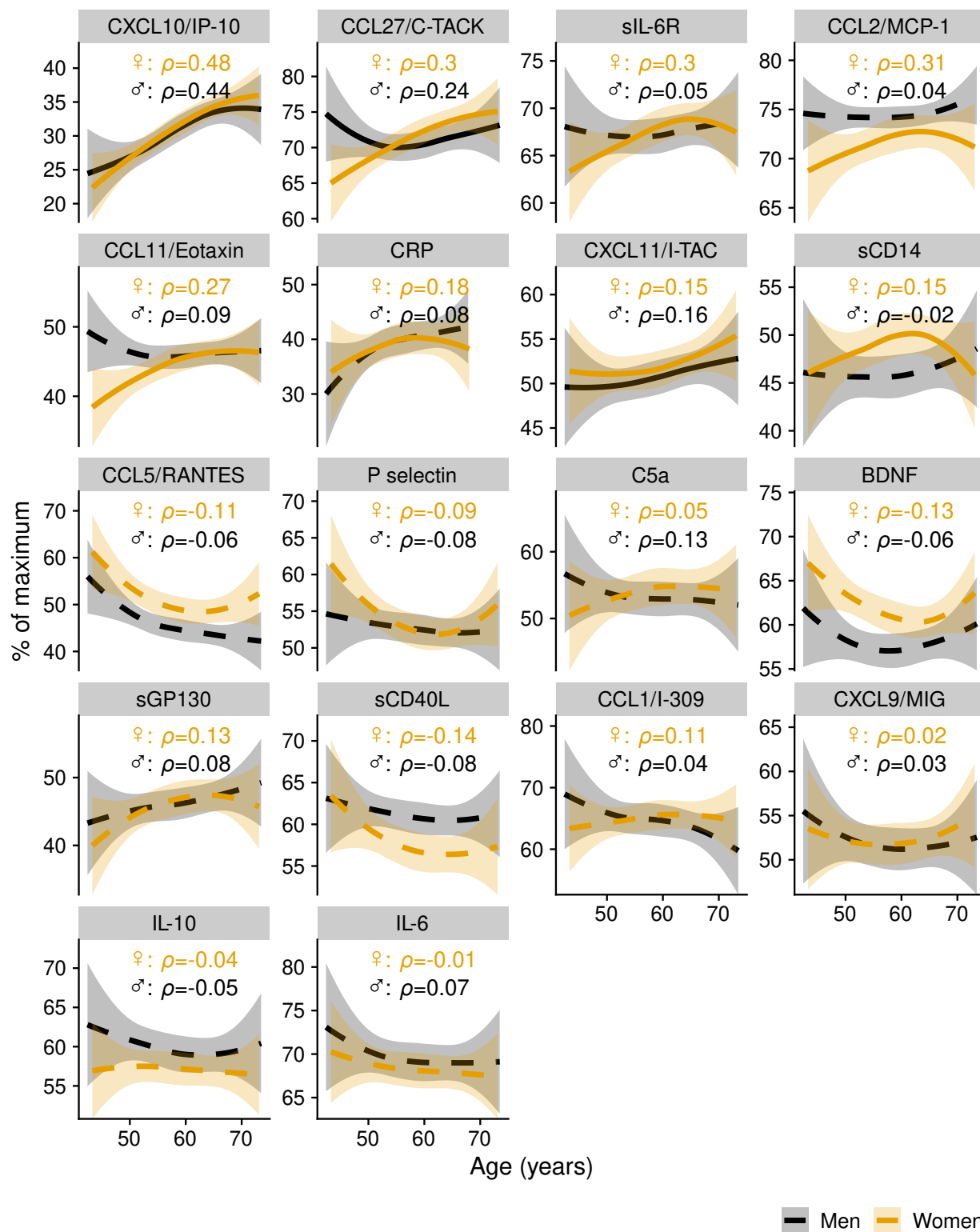


Figure 2: Trajectories of inflammatory markers related to age. The average trajectories are shown with 95% confidence interval estimated by local polynomial regression, separately for men (n=73) and women (n=71). Dashed line: no association found. Continuous line: association found between inflammatory marker trajectory and age. Y axis shows the % of maximum concentration per biomarker. Average concentrations per biomarker (pg mL^{-1}) are indicated in Table S1.

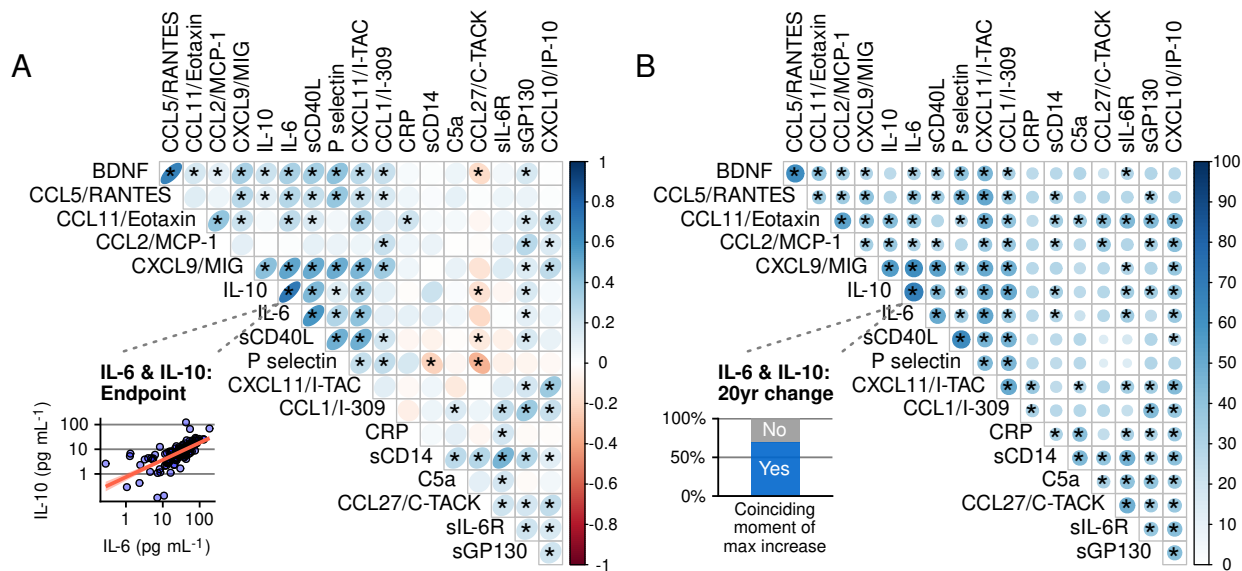


Figure 3: Relationships between inflammatory markers shown as A) correlation between pairs of inflammatory markers at study endpoint and B) similarity between pairs of inflammatory marker trajectories during about 20years of follow-up. In (A) the direction and strength of the association in is visualized with an oval shape and a color gradient. The inset shows the correlation between one pair of biomarkers (IL-6 and IL-10) as an example of the analysis. In (B) the blue gradient color and the size of the circles shows the percentage of participants of which a pair of biomarkers had the highest increase in concentration at the same moment in 20 years of follow-up. This was seen in 69% of the participants for the pair of biomarkers IL-6 and IL-10 (see inset). *= an association between two inflammatory markers, with false discovery rate being set at a maximum of 15%. n=144

103 time intervals in the 20 years of follow-up time a marker had its greatest increase in concentration. Then, we
 104 tested if this moment was the same for every pair of biomarkers in the majority of the participants. In both
 105 men and women the most distinct synchronous changes were again found with IL-6 and IL-10 (Figure 3B),
 106 and with markers of the innate immune system, such as those of platelet activation (sCD40L and P-selectin)
 107 and those related to chemotaxis and granulocyte activation (CCL11/Eotaxin and CCL5/RANTES) (Figure
 108 3B). The concentrations of structurally related chemokines CXCL9, CXCL11/I-TAC and, to a lesser extent,
 109 CXCL10/IP-10 also tended to change in unison over time in both men and women. In addition, CXCL9 and
 110 11 were related to multiple other inflammatory markers, such as IL-10 and IL-6, but also RANTES/CCL5, P
 111 selectin, and sCD40L, indicating the sensitivity of inflammatory marker levels for IFN γ -pathway activation
 112 in aged men and women.

113 In this longitudinal analysis inflammatory marker profiles of men and women appear somewhat more similar
 114 than in the cross-sectional analysis. Thus, while RANTES/CCL5 correlated with more markers in women
 115 than in men looking at the marker levels at study endpoint (Figure S3A,B), this appeared to be less so when
 116 looking at synchronous changes in markers over time (Figure S3C,D). Furthermore, we did not find differences
 117 between men and women in the maximum levels of biomarkers over the entire 20-year time period, nor in the
 118 magnitude of the maximum increase in inflammatory marker levels (data not shown). In summary, multiple

119 inflammatory marker levels were correlated with each other, with IL-6 showing the strongest correlations
120 with other markers, followed by markers related to platelet activation and the IFN γ -pathway activation.
121 Correlations between inflammatory markers at endpoint were stronger and more abundant in women, but
122 this tendency was less pronounced when comparing the temporal change in biomarker levels, with no strong
123 evidence found for higher immune marker reactivity in women.

124 **2.5 Relationships of inflammatory markers with BMI and waist circumference**

125 We investigated the relationship between the inflammatory marker profile and two measures of body fat,
126 namely body mass index (BMI) and waist circumference. The average BMI increased with age in both
127 sexes over the 20 years of follow-up (men: $\rho=0.24$, women: $\rho=0.37$, Figure S4A). In women, the average
128 BMI values and waist circumference during the follow-up were positively associated with the AUC of CRP
129 ($\rho=0.55$) and sIL-6R ($\rho=0.28$) (Figure S4B and C), both related to the IL-6 pathway. In men, a larger waist
130 circumference was positively associated with CRP levels, but associations between average BMI levels and
131 inflammatory markers were not observed.

132 **2.6 Levels of inflammatory markers related with frailty**

133 Continuously elevated CRP trajectories were related to frailty at study endpoint (frailty index based on 35
134 deficits, Table S2) as was shown previously (Samson et al., 2019), and this association turned out to be one of
135 the strongest when analyzed together in our inflammatory marker panel (correlation of study endpoint frailty
136 with the area under the curve (AUC) of the CRP trajectory in men: $\rho=0.44$, in women: $\rho=0.50$; see Figure
137 4A&B). Associations of frailty with other inflammatory markers were only found in women, showing a higher
138 AUC of sIL-6R ($\rho=0.34$) and sCD14 ($\rho=0.33$), but lower AUC of sCD40L ($\rho=-0.29$) in frailer women. BMI
139 values were also higher on average at each time of measurement in frail men and women (Figure 4C). When
140 associations of frailty with the inflammatory markers were adjusted for BMI, two correlations were found,
141 namely a positive association of CRP values with frailty and an association that was not found without
142 adjusting for BMI, namely a positive one of BDNF with frailty in men ($\rho=0.35$ and $\rho=0.21$, respectively).
143 We further investigated whether age-related increases or decreases of inflammatory markers over the past 20
144 years were specifically related to frailty at study endpoint. The age-related increases in sCD14 and sIL-6R
145 levels seen in women (Figure 3) turned out to be related to frailty at study endpoint (Figure 4D, $\rho=0.42$
146 and $\rho=0.35$, respectively). In addition, while no age-related increase in C5a levels were seen, C5a levels were
147 seen to increase with age with higher frailty index scores in women (Figure 4D, $\rho=0.30$). These results were

148 also found after adjusting for both BMI and for baseline marker concentrations.

149 **2.7 Prediction of frailty with multiple inflammatory protein trajectories**

150 Since multiple associations were found between inflammatory protein trajectories and frailty, we further
151 investigated this relationship by studying how well the AUCs of all the biomarkers together could predict
152 the frailty index score, using a random forest algorithm with BMI and age at last measurement also included
153 in the model. The predictive accuracy was 12% in men and 10% in women. With this low predictive accuracy,
154 only the top results of the algorithm could be interpreted (Figure 5). These top results confirmed findings of
155 the association study, with BMI and CRP being the most important predictors with comparable variable
156 importance in men and women, and sCD14 closely following albeit only in women.

157 **2.8 Trajectories of inflammatory markers related to an increase in frailty**

158 Next, the relation was studied between the development of low-grade inflammation and the onset of frailty.
159 This was done in two different ways. First, we investigated whether chronic low-grade inflammation was
160 related to an increase in frailty over a period of five years, i.e. between measurement rounds 5 and 6. While
161 the AUCs of several inflammatory markers were found to be related to frailty, no associations were observed
162 between them and the increase in frailty over the five-year period (Supplementary correlation tables, tab
163 “Frailty_change”). Secondly, to investigate the risk of becoming frail prospectively over a longer period of
164 time, we selected a subgroup of individuals who were ‘healthy’ and thus were likely to have a low, favorable
165 frailty index score at the start of our study; healthy at study baseline was defined according to an alternative
166 health index that could be assessed at study baseline (Healthy Ageing Index score, score of 9 or 10 out of
167 10, higher score being more favorable; 31 women and 27 men; Figure 6A) (Dieteren et al., 2020). Then we
168 investigated within these selected ‘healthy’ participants whether those with continuously higher (or lower)
169 levels of inflammatory markers had a higher frailty index score at study endpoint, and thus became frailer
170 over time. In this subgroup, we observed that in women frailty development was associated with higher AUCs
171 of CRP and sIL6R ($\rho=0.44$ and 0.56 respectively, Figure 6B&C). In men, no associations with frailty were
172 found. In the relationship between inflammatory markers and becoming frail body fat could be important,
173 since BMI was continuously higher in participants who became frail (Figure 6D). Indeed, after adjusting for
174 BMI, the associations of the inflammatory markers with the chance of becoming frail in women were weaker
175 and no longer passed the criteria for qualifying as a “detected association” (correlation coefficient of frailty
176 and sIL-6R: $\rho=0.49$; of frailty and CRP: $\rho=0.22$). Thus, it is likely that BMI plays an important role in the

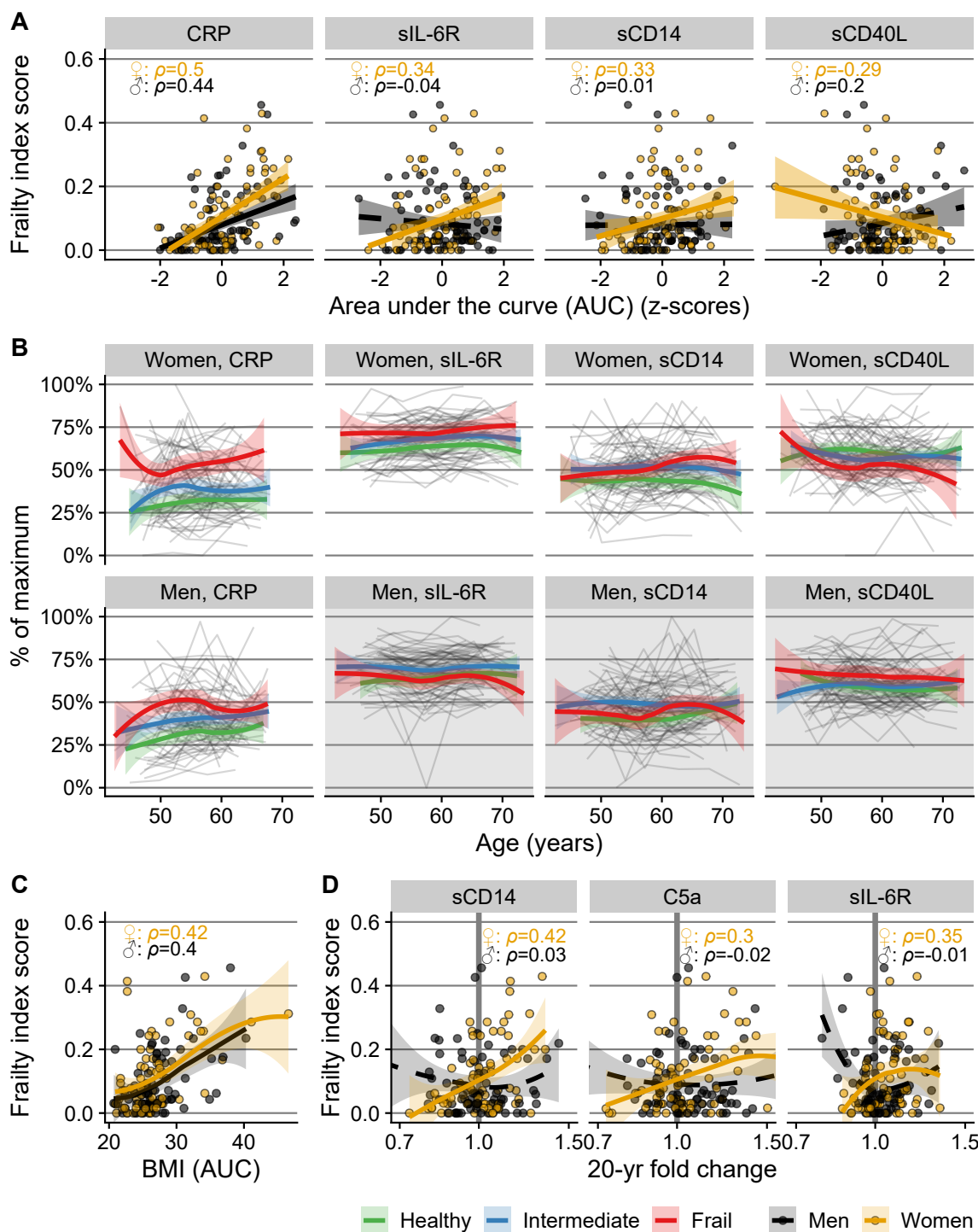


Figure 4: Relation between frailty at endpoint and inflammatory marker levels over time in men (n=73) and women (n=71). To capture the cumulative ‘exposure,’ the inflammatory markers in (A) are expressed as area under the inflammatory marker concentration curve versus time (AUC), standardized to take into account different follow-up periods, and transformed into z-scores for visualization and better comparison between inflammatory markers. (B) shows the trajectories of individuals (grey lines) and the local polynomial regression lines with 95% confidence intervals per frailty category (bold colored lines) based on frailty index score at study endpoint (Concentrations are scaled to % of maximum concentration per marker). This complements the analysis in (A) and visualizes in more detail whether marker levels are elevated chronically. The frailty categories in (B) were used for illustration purposes; for the statistical analyses, the continuous frailty index score was used. Inflammatory markers were displayed when their AUC showed an association with the frailty index score at endpoint in at least one of the sexes. Plot area backgrounds in (B) are grey if no association was found. (C) Body mass index (BMI) over time, expressed as AUC values. (D) 20-year fold change in inflammatory markers, all over 20 years. A vertical reference line of no increase (fold change of 1.0) is shown in bold grey.

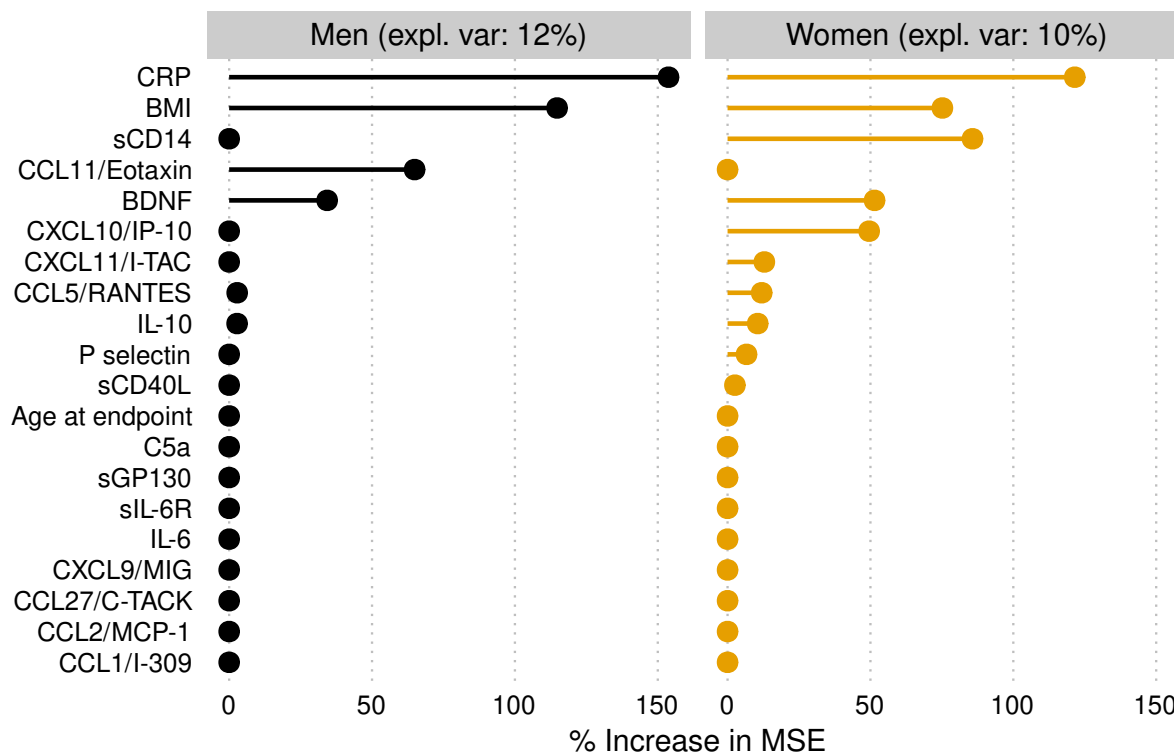


Figure 5: Variable importance of the inflammatory marker concentrations over the past 20 years (area under the curve) per individual in predicting frailty at study endpoint using a random forest algorithm. Age at study endpoint was included in the model. Expl. Var: explained variance. % increase in MSE: percentage increase in mean squared error of the prediction of frailty after the variable is replaced by random noise. A higher value thus means that the variable is more important in predicting frailty.

177 association of chronic inflammation over time and frailty.

178 2.9 Trajectories of inflammatory markers related to lung function

179 As lung function is an important determinant of health known to decline with age, and for which clinically
 180 accepted quantitative measures have been defined, we investigated the relation between inflammatory marker
 181 trajectories over the past 20 years and spirometric measurements at endpoint. In women, pulmonary function,
 182 in terms of Forced Expiratory Volume in one second (FEV1), tended to be lower when trajectories of CRP
 183 were higher (correlation of the AUC of CRP with FEV1, $\rho=-0.48$), but higher with elevated trajectories of
 184 CXCL11/I-TAC ($\rho=0.33$), and sCD40L ($\rho=0.33$) (Figure 7A). In men, no associations were found between
 185 the inflammatory marker trajectories and the FEV1, although the patterns of CRP were more or less similar
 186 to those in women. A different pulmonary function measure, Forced Vital Capacity (FVC), was found
 187 to be negatively related to CRP concentrations in women ($\rho=-0.48$) adding to the notion that pulmonary
 188 function is affected by chronic inflammation. No relationships were found between the inflammatory marker

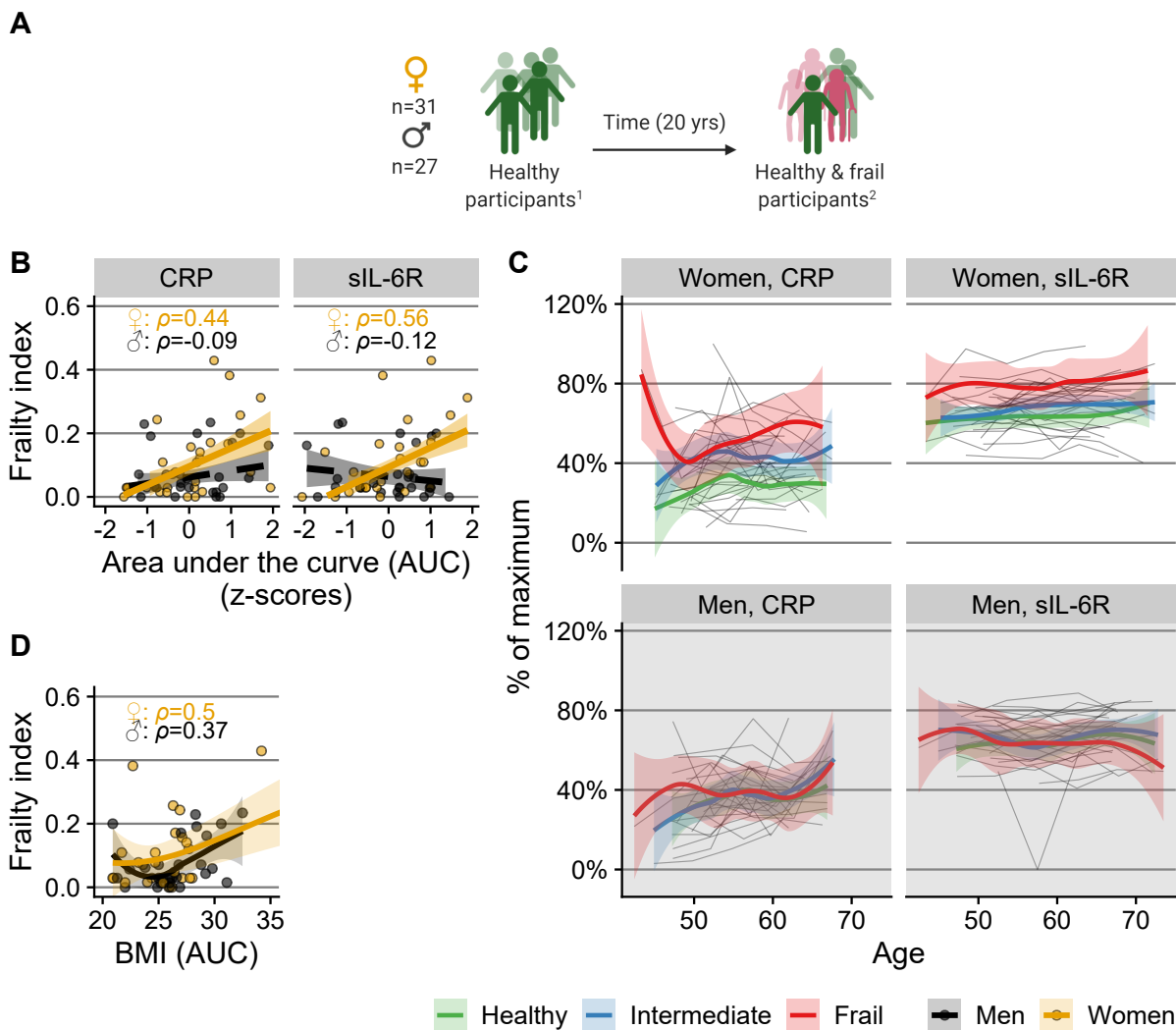


Figure 6: (A) Subgroup of participants that were ‘healthy’ at baseline. ¹Health status at baseline is defined by a healthy aging index (healthy aging index score = 9 or 10 out of 10). ²Health status at endpoint is defined by a frailty index score. Relation of frailty at study endpoint with trajectories of CRP and sIL-6R (B,C) and with BMI (D) in the subgroup of people that were ‘healthy’ at study baseline. To capture the cumulative ‘exposure,’ trajectories of CRP and sIL-6R in (B) and of BMI in (D) are expressed as area under the concentration/level versus time curve per individual. Grey lines in (C) are individual trajectories. Bold colored lines in (B) are (robust) linear regression lines, and in (C) and (D) local polynomial regression lines with 95% confidence intervals. In (C), the color denotes the ‘frailty category’ at study endpoint. Frailty categories are used for visualization of the longitudinal trajectory; for the statistical analyses the continuous frailty index score was used. Concentrations on y-axis in (C) are scaled to % of maximum concentration per marker.

189 trajectories and the FEV1/FVC ratio.

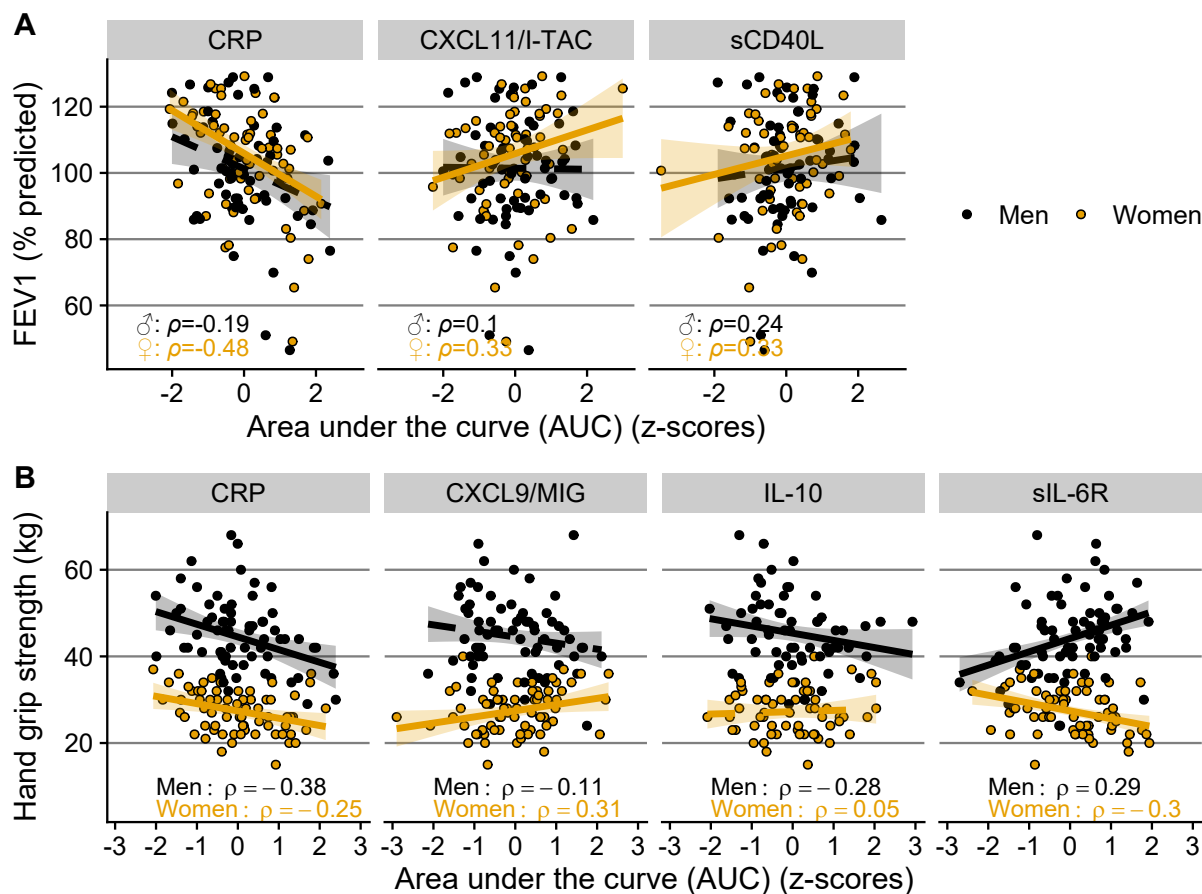


Figure 7: Inflammatory marker trajectories in men and women of those that showed an association with physiological clinical parameters of aging at study endpoint; (A) lung function assessed by forced expiratory volume in one second (FEV1), and (B) handgrip strength measured as an approximation of physical strength. As a measure of cumulative ‘exposure,’ inflammatory marker levels were assessed as area under the inflammatory marker concentration versus time curve per individual (AUC) during the 20-year follow-up. AUC values were transformed to z-scores for visualization. % predicted: FEV1 value compared to expected reference values from the Global Lung Initiative (Quanjer et al., 2012) specific for age, length, and sex. An association is indicated by a continuous trendline, otherwise a dashed trendline is shown.

190 2.10 Trajectories of inflammatory markers related to handgrip strength

191 Another important determinant of health that declines with advancing age is muscle strength, which can
 192 be represented approximately by handgrip strength. Handgrip strength, as measured with a dynamometer,
 193 was related to several inflammatory marker trajectories in men and in women (Figure 7B). In both sexes
 194 we found a negative association between the AUCs of CRP trajectories and handgrip strength, but only in
 195 men did this association remain after adjusting for BMI. In men, a negative association was found between
 196 IL-10 trajectories and handgrip strength ($\rho=-0.28$), which remained after adjusting for BMI. Contrasting
 197 correlations were found between sIL-6R and handgrip strength in men and women, with men showing a

198 positive correlation ($\rho=0.29$) and women a negative one ($\rho=-0.30$). After adjusting for BMI, however, these
199 associations with sIL-6R were lost. Women also showed a positive correlation of handgrip strength with
200 CXCL9 ($\rho=0.30$) which was lost after adjusting for BMI. Taken together, chronically elevated levels of CRP
201 and IL-10 are associated with lower hand grip strength in men.

202 **3 Discussion**

203 In this unique longitudinal study with 20 years of follow-up data, we quantified chronic low-grade inflam-
204 mation in an aging population using multiple inflammatory markers and showed that changes of markers
205 of the IL-6 pathway, of platelet activation and of monocyte activation are associated to clinically relevant
206 health outcomes at older age, such as frailty. BMI and waist circumference turned out to be important
207 factors in this process, since they are related to both frailty and markers of the IL-6 pathway. Associations
208 were stronger and more abundant in women, although inflammatory marker trajectories in men and women
209 became more similar with age. Although the power to predict frailty using the inflammatory marker tra-
210 jectories was weak, probably due to the heterogeneity in the study population and the small numbers of
211 individuals, the results were in agreement with those of the association studies.

212 **3.1 Age-related increase of inflammatory marker levels**

213 We detected an increase in multiple biomarker levels with advancing age, which is in line with the reported
214 chronic low-grade inflammation at older age (Baylis et al., 2013; Franceschi et al., 2006). Clear increases in
215 IFN γ -related chemokines with age were seen in both men and women. Higher concentrations of inflammatory
216 markers with advancing age were seen in both men and women in the IFN γ -induced and structurally related
217 chemokines CXCL10/IP-10 and CXCL11/I-TAC. Elevated concentrations of these chemokines could be a
218 sign of both innate and adaptive immune activation and could be clinically important, since they have been
219 proposed as possible biomarkers for heart failure (Altara et al., 2015). CXCL10/IP-10 has previously been
220 described as increasing with age in studies with shorter follow-up time (Hearps et al., 2012; Hsu et al.,
221 2019). Also CCL27/C-TACK levels increased with age in men and women, possibly related to accumulating
222 skin damage in elderly, since CCL27/C-TACK is a cytokine involved in T-cell-mediated homing to the skin
223 (Homey et al., 2002; Richmond et al., 2019).

224 In women, other inflammatory markers were also found to increase with advancing age, including the IL-6
225 pathway related markers CRP and sIL-6R and two chemokines related to innate immune cell activation,

226 namely CCL2/MCP-1 and CCL11/Eotaxin. These sex-specific increases all seemed to reach a plateau at
227 around the age of 60 years. Differences in the immune profile between men and women could be due to
228 hormonal differences, as stated previously (Bupp, 2015; Furman et al., 2014). This is in line with our
229 data suggesting that hormonal shifts due to menopause can partially explain why concentrations of multiple
230 inflammatory markers increase over time in women before the age of 60. Of note is that, while sIL-6R and
231 CRP were found to increase with age in women, IL-6 levels were not. This may be explained by the molar
232 excess of sIL-6R in the circulation, which naturally binds IL-6. Still, we expected to find an association
233 between IL-6 levels and age, since IL-6 levels were found to be increased with advancing age in several
234 previous studies (Ferrucci et al., 2005; Giuliani et al., 2001; Puzianowska-Kuźnicka et al., 2016), which
235 is why it is thought to be a key component in chronic low-grade inflammation in the elderly. There are,
236 however, other studies that also did not find a relationship between IL-6 and age (Beharka et al., 2001; Hsu
237 et al., 2019; Van Epps et al., 2016), one of which claimed this was due to the limited age range of their
238 participants (Van Epps et al., 2016).

239 **3.2 Chronic low-grade inflammation related to frailty**

240 In our study we investigated how chronic low-grade inflammation is related to frailty and to an increase in
241 frailty. As expected, the association of higher CRP levels in frailer men and women that we found in our
242 previous study (Samson et al., 2019) was detected in this subcohort again, even in a smaller selection of
243 participants (n=144 instead of 289), and with a different analysis approach, i.e. by investigating the area
244 under the curve of the inflammatory markers. Moreover, CRP turned out to be one of the inflammatory
245 markers with the strongest association with frailty and with the chance of becoming frail over time, followed
246 by sIL6R in women, indicating a role for the IL-6 pathway. Previous literature about immune marker
247 alterations preceding the risk of becoming frail is limited, with one study showing associations of higher
248 CRP and the risk of becoming frail (Gale et al., 2013). In another study, associations were reported between
249 IL-6 and ‘incident frailty’ over 5 years. sIL-6R was not measured in that study (Hsu et al., 2019). Probably
250 a major driver in this process is (over)weight, since the associations with frailty in our study were not
251 detected anymore after adjustments for BMI. A plausible reason why BMI influences these results is that
252 a higher BMI both increases the risk of becoming frail as well as raises the inflammatory biomarker levels
253 (Ghigliotti et al., 2014). Excessive adipose tissue, especially visceral adipose tissue, is known to contribute
254 to chronic inflammation by increased secretion of adipokines and inflammatory cytokines and by promoting
255 accumulation of macrophages (Ellulu et al., 2017; Ghigliotti et al., 2014; Ouchi et al., 2011). It has been
256 inferred that waist circumference is a better estimate of visceral pro-inflammatory body fat than BMI in

257 the elderly (Crow et al., 2019). Indeed, we saw stronger associations of IL-6 pathway markers with waist
258 circumference than with BMI in men, possibly because waist circumference is thought to give a better
259 estimate of visceral pro-inflammatory body fat than BMI in the elderly (Crow et al., 2019). Of note is
260 that even after adjusting for BMI, frailer women but not men also showed higher levels of sCD14, mostly
261 because of an increase in this marker's level in the preceding 20 years. sCD14 is a marker of monocyte
262 activation since it is released from monocytes after stimulation with TLR ligands (Shive et al., 2015), in
263 particular lipopolysaccharide (LPS). sCD14 also binds to LPS and it has been suggested that sCD14 is
264 necessary for an adequate response to LPS in platelets (Damien et al., 2015) and can enhance sensitivity
265 for LPS in endothelial cells (Lloyd-Jones et al., 2008), suggesting that sCD14 is important in the defense
266 against bacterial infections. Its increase in concentration over time in frailer women is in line with previous
267 reports showing higher monocyte numbers in frailer women (Leng et al., 2009; Samson et al., 2020).

268 While most associations between inflammatory marker levels and frailty were positive, as expected when low-
269 grade inflammation occurs more frequently in frail participants, in women we also found a negative association
270 of sCD40L levels with frailty, although only when not adjusted for BMI. sCD40L, which is released mainly by
271 activated platelets (Danese, 2003; Nagasawa et al., 2005), is a soluble membrane glycoprotein involved in B
272 cell responses (Jenabian et al., 2014) and in macrophage and monocyte signaling (Suttles & Stout, 2009). It
273 thus plays an important role in the communication between innate and adaptive immune responses. sCD40L
274 is commonly seen as a pro-inflammatory molecule (Aloui et al., 2016), but it might be immunosuppressive as
275 was suggested in HIV infection and in cancer by inducing expansions of regulatory T cell populations (Huang
276 et al., 2012; Jenabian et al., 2014), and suppressing T cell proliferation (Huang et al., 2012), suggesting that
277 lower levels of sCD40L might indicate failing regulation with higher low-grade inflammation as a result.

278 **3.3 Lung function and physical strength**

279 Higher overall CRP levels are often associated with worse outcomes in specific health-related parameters,
280 such as lung function (Ahmadi-Abhari et al., 2014) and handgrip strength (Cesari et al., 2014; Smith et al.,
281 2019; Tuttle et al., 2020). We largely confirmed these results in our study, showing that inflammatory marker
282 levels are persistently higher during 20 years in people with reduced lung function and handgrip strength,
283 which also could indicate that low-grade inflammation in the elderly is indeed 'chronic.' In addition, only in
284 women, higher levels of sCD40L and CXCL11/I-TAC were found to be associated with better lung function
285 (by means of FEV1 and FVC), which we believe is the first time these markers were related to lung function
286 in a community-dwelling population. While sCD40L was shown in previous studies to possibly have anti-
287 inflammatory roles as discussed above, CXCL11/I-TAC was not, and we would have expected a negative

288 correlation of this marker with lung function because of its pro-inflammatory effect in the IFN γ pathway.
289 Such a negative association was seen in cross-sectionally performed studies in sarcoidosis (Arger et al., 2019)
290 and other lung diseases (Kameda et al., 2020), showing higher levels of CXCL11/I-TAC with lower lung
291 function, which is in contrast with our findings. Further studies are needed to confirm our findings in a
292 community-dwelling population and to explain why in our study this relationship was found only in women.
293 Interestingly, in men we also found a negative correlation of handgrip strength with concentrations of IL-10,
294 a cytokine commonly seen as anti-inflammatory. A similar relationship of higher muscle strength (knee
295 extension strength) and lower IL-10 values was also described in a previous study (Calvani et al., 2017),
296 although in most studies no associations between IL-10 and grip strength were detected (Cesari et al., 2004;
297 Goldeck et al., 2016). Our results also show that IL-10 correlates strongly with several pro-inflammatory
298 markers such as IL-6, which is in line with previous studies (Calvani et al., 2017; Hsu et al., 2019). Further
299 research is needed to confirm the results and to investigate whether these elevated IL-10 levels could be a
300 compensatory response to low-grade inflammation.

301 **3.4 Strengths and limitations**

302 Our longitudinal analysis of a well described cohort is a major strength of this study. To the best of our
303 knowledge, our study is unique as an extensive inflammatory panel was repeatedly measured at 5-year
304 intervals in the same individuals over a period of 20 years. Since variation within-individuals was much
305 smaller than the variation between individuals, we had more power to detect changes of protein levels with
306 advancing age than in cross-sectional studies, even when these have larger sample sizes. Furthermore, while
307 sample size for cross-sectional analyses of a single analyte was limited compared to large cohort studies,
308 our study made use of a large panel of inflammatory markers, thus giving a more comprehensive insight in
309 inflammation than studies that measured only a few inflammatory markers. Also, since inflammatory marker
310 levels can vary considerably between studies due to methodological differences, an advantage of our study
311 was that all inflammatory markers were measured with the same assay and that all samples of the same
312 individual were measured in the same plate. Of note is that we did not have a frailty index measurement at
313 baseline, which is a limitation in our study of the relationship between levels of inflammatory markers and
314 becoming frail. Thus, while we assumed that the participants who we selected to be healthy at baseline had
315 a low frailty index score, this selection was based on a relatively simple health score rather than the frailty
316 index score that we used at study endpoint. Other limitations are that the study population could be less
317 representative for the general population due to, for instance, selective dropout of participants commonly
318 seen in longitudinal cohorts, although the response rate in the Doetinchem cohort study is good, generally

319 above 70% (Picavet et al., 2017). Furthermore, the prolonged storage of the plasma samples could affect the
320 sample quality. However, the samples were consistently stored in low temperature and the trajectories were
321 remarkably stable within individuals, indicating that protein degradation was probably of minor influence.

322 **3.5 Concluding remarks**

323 In conclusion, our exploratory study gives unique insight into longitudinal changes in the immune profile of
324 aging men and women over a period of about 20 years, and revealed multiple associations of immune markers
325 with clinically relevant outcomes at study endpoint, such as frailty, handgrip strength and lung function.
326 More specifically, the findings implicate an important role for the IL-6 pathway due to elevated markers
327 such as CRP and, only in women, monocyte activation due to the involvement of sCD14. As BMI and waist
328 circumference are related to elevations of immune markers in the IL-6 pathway, chronic inflammation might
329 be an important mediator of the relationship between BMI and frailty.

330 **4 Methods**

331 The Doetinchem cohort study (DCS) is a population-based longitudinal study of 7769 participants that have
332 been followed since 1987 (Picavet et al., 2017; Verschuren et al., 2008). Every five years plasma samples were
333 taken and data regarding the participants' life style and health were collected (1). For the present study,
334 a subgroup of 144 people aged 65-75 years was selected from the DCS, stratified by sex and with equal
335 numbers of healthiest, intermediate and frailest participants according to the classification used in (Samson
336 et al., 2019). In brief, the healthiest/frailest were defined as those with the 15% lowest/highest frailty index
337 score (see below) among their age- and sex-matched peers. The individuals selected were participating in
338 the study at least until 2016, and had participated in at least four out of the five investigated measurement
339 rounds. The measurements of the present study were performed as an extension of the ones in the entire
340 DCS, with the sample size being restricted by budgetary and logistic constraints.

341 **4.1 Frailty index**

342 Details on the frailty index score used in this study can be found elsewhere (Samson et al., 2019). Its definition
343 was based on previous studies (Collerton et al., 2012; Mitnitski et al., 2001) and in our implementation it
344 consists of 35 potential age- and health-related deficits, covering a broad range of 'domains,' including
345 cognitive, physical, and psychological functioning. Difference of the score used in this study compared to

346 the one previously used and validated in the DCS (Samson et al., 2019) is that the latest available data
347 was included, and that the deficit regarding overweight was left out of the score, so that we could better
348 investigate the influence of overweight on our results. Details and cutoff values for all of the 35 deficits
349 are shown in Table S2. All deficits are bound between zero (best possible score) and one (worst possible),
350 and the frailty index is the average score per individual. The frailty index was calculated in the two latest
351 available DCS assessment rounds (rounds 5 and 6). Frailty categories (see DCS subgroup selection above)
352 were only used for visualization in some figures; for statistical analyses, the continuous frailty index score
353 was used.

354 4.2 Healthy aging index

355 The healthy aging index was measured at study baseline and was used to select a subgroup of participants
356 that was 'healthy' at the start of follow-up. The score is based on health deficits defined by systolic blood
357 pressure, plasma glucose levels, creatinine levels, lung function (forced vital capacity), and general cognitive
358 function (Nooyens et al., 2011), as described elsewhere (Dieteren et al., 2020). Except for plasma glucose
359 levels, the deficits were comparable to some of those included in the frailty index score. Every item has
360 similar weight in the score, and the score ranges from 0 to 10, with 10 being the best and 0 the worst
361 possible outcome (with all deficits present).

362 4.3 Measurements of inflammatory markers

363 Plasma samples were collected repeatedly within the same individuals over approximately 20 years between
364 1991-2017 (1). The samples had been stored at -80°C and thawed shortly before measuring a large panel of
365 inflammatory markers, consisting of the following cytokines, chemokines and soluble receptors: C-C Motif
366 Chemokine Ligand (CCL) 1/I-309, CCL2/MCP-1, CCL5/RANTES, CCL11/Eotaxin, CCL27/C-TACK, C-
367 X-C Motif Chemokine Ligand (CXCL) 9 /MIG, CXCL10/IP-10, CXCL11/I-TAC, Interleukin (IL)-1 β , IL-10,
368 IL-6, soluble CD40 ligand (sCD40L), soluble CD14 (sCD14), soluble IL-2 receptor (sIL-2R), soluble IL-6
369 receptor (sIL-6R), soluble glycoprotein 130 (sGP130), Complement 5a (C5a), Brain-derived neurotrophic
370 factor (BDNF), and P selectin. Concentrations of the markers were measured using a validated bead-based
371 multiplex immunoassay (Flexmap 3D®, Luminex) at the Multiplex Core Facility lab, University Medical
372 Center Utrecht, The Netherlands. All detection antibodies were coupled to magnetic beads and were tested
373 for specificity. Samples were thawed at room temperature just before the measurement. To minimize assay
374 variation within individuals, all samples from the same participant (n=6) were measured on the same plate.

375 Measurement of samples took place in 2018, in three independent batches. Levels of C-reactive protein (CRP)
376 in plasma had been measured previously in DCS round 2,3,4, and 5 (Hulsegge et al., 2016). Apart from
377 sIL-2R and IL-1 β , the proportion of samples in which concentrations were below the limit of quantification
378 (LOQ) was less than 20% for all inflammatory markers. Concentrations below the LOQ were imputed with
379 a random value below the LOQ based on maximum likelihood estimation (Lubin et al., 2004). sIL-2R and
380 IL-1 β were excluded from analysis since more than 65% of the samples were below the LOQ. For IL-6 and
381 IL-10, 85 samples (often in the same participants) were excluded due to possible non-specific binding to
382 beads in the IL-10 or IL-6 region, which reduced the sample size for these cytokines to n=129 out of the
383 total n=144 individuals (n=65 men, n=64 women). The sample size of sGP130 was reduced (n=67 men,
384 n=66 women) since sGP130 was not measured in the first batch of cytokine measurements.

385 **4.4 Anthropometric measurements**

386 Body mass index was calculated as body mass in kilograms divided by the square of the participants' height
387 in meters. A tape measure was used to measure waist circumference, which was done with the participant
388 in an upright standing position and holding the tape measure horizontal around the waist, at the midpoint
389 between the lower rib and the iliac crest.

390 **4.5 Handgrip strength**

391 Participants applied as much force as possible with their dominant hand to a hydraulic dynamometer (Jamar),
392 with their elbow in a 90 degree angle. The highest applied force out of three separate attempts was used.

393 **4.6 Lung function**

394 At study endpoint, a heated pneumotachometer (E Jaeger, Wurzburg, Germany) was used to measure forced
395 expiratory volume in one second (FEV1) and forced vital capacity (FVC) as previously explained in detail
396 (van Oostrom et al., 2018). Spirometric measurements were done and evaluated according to the American
397 Thoracic Society and European Respiratory Society guidelines (Enright et al., 1991). FEV1 and FVC
398 values were transformed to the percentage of predicted values, with predictions based on the Global Lung
399 Initiative's (2012) reference values (Quanjer et al., 2012) specific for age, length, sex, and ethnicity, using the
400 R macro provided on [https://www.ers-education.org/guidelines/global-lung-function-initiative/spirometry-](https://www.ers-education.org/guidelines/global-lung-function-initiative/spirometry-tools.aspx)
401 [tools.aspx](https://www.ers-education.org/guidelines/global-lung-function-initiative/spirometry-tools.aspx).

402 **4.7 Statistical analysis**

403 **4.7.1 Longitudinal protein trajectory estimators**

404 To estimate the “inflammatory burden over time” for each individual and each inflammatory marker the area
405 under the concentration versus time curve (AUC) was calculated and was divided by the total time of follow-
406 up to adjust for possible differences in follow-up time. The AUC values of every inflammatory marker were
407 subsequently log transformed and mean-centered per measurement batch to adjust for possible confounding
408 batch effects. To estimate the changes in inflammation over time, we also calculated the within-individual
409 proportional increase or decrease per year. To do this, we used a slope of log-transformed concentration over
410 time per individual, calculated in a median based linear model (Komsta, 2019), as an estimator for rate of
411 change over time. A median based linear model is much less sensitive to outliers or to deviations from a
412 normal distribution than ordinary linear regression models (Theil, 1950; Wilcox, 1998).

413 **4.7.2 Association studies**

414 All the results were adjusted for multiple testing by controlling the false discovery rate (Benjamini &
415 Hochberg, 1995) separately for each analysis. A nominal false discovery rate of at most 15% (meaning,
416 roughly speaking, an average of 1.5 false discoveries per 10 discoveries presented as such) was tolerated
417 in view of the number of tests carried out. For testing associations, we used the permutation versions of
418 the Spearman and Wilcoxon-Mann-Whitney tests blocked for certain background or confounding variables
419 (e.g. age) as implemented in the R package coin (Hothorn et al., 2008) with p-values estimated by simulation.
420 For an explanation of the concept of blocking in experimental designs, see (Krzywinski & Altman, 2014).
421 Details of all association studies are shown in Supplementary tables.

422 To investigate whether changes in plasma concentrations of inflammatory markers change with age, we
423 carried out Spearman tests between each marker’s concentrations and age blocking by participant. Thus,
424 we focused on within-individual increases in the marker’s concentration during the 20 year of follow-up.
425 Should a marker tend to increase or decrease with age, the Spearman correlation between the marker’s
426 concentration and an participant’s age should tend to exhibit consistently large or consistently small values
427 across the cohort.

428 Single associations between an inflammatory marker’s trajectory (the AUC) and sex were tested with the
429 Wilcoxon-Mann-Whitney test, blocking for age at last measurement (in two categories, 65-70 and 70-75
430 years) and BMI (three categories) to adjust for these variables.

431 Associations between inflammatory markers at the study's endpoint were adjusted for age at last mea-
432 surement and the immunoassay batch number (three batches). Longitudinal relationships between pairs of
433 biomarkers were tested by investigating synchronous changes in biomarker levels. For every pair of markers,
434 we investigated whether the moment of maximum increase in one marker coincided with that in another
435 marker more often than what might be expected by chance, by carrying out one-tailed binomial tests pos-
436 tulating the 'probability of success' of $\frac{1}{4}$ under the null hypothesis of no association.

437 To address our research question whether chronic low-grade inflammation is related to frailty, we carried out
438 several association studies, all separately for men and women. First, we tested associations, with Spearman's
439 tests, between the AUC of each marker's inflammatory trajectory and the frailty index score, adjusted for
440 age at last measurement. Analogous associations were tested using BMI as additional blocking variable,
441 to get an idea about the importance of BMI in these relationships. Then, we investigated the association
442 between the individual's slope of the inflammatory protein trajectories and frailty and at endpoint, using
443 Spearman's tests blocked by the baseline level of the marker (in tertiles) and BMI. Next, the relation was
444 investigated between every immune marker trajectory and change in frailty index score from round 5 to
445 round 6, adjusting the associations for age at last measurement and for the initial frailty index score at
446 round 5 (using tertiles for blocking). The latter was done to adjust for a possible regression to the mean
447 effect and to give more weight to smaller increases in frailty index of people that already had a high index
448 in the beginning.

449 The tested relationships of the inflammatory markers with lung function parameters were also adjusted for
450 smoking with two blocking categories: non-smokers and former smokers. The relationships with handgrip
451 strength were adjusted for age and were repeated with adjustments for BMI.

452 **4.7.3 Prediction analysis**

453 We used a random forest prediction algorithm (Liaw & Wiener, 2002) to investigate whether frailty could
454 be predicted using several dependent variables, namely the AUCs of all the inflammatory markers, age
455 at endpoint, and BMI. The overall performance of the prediction analysis was assessed in terms of the
456 percentage explained variance. The importance of the independent variables to predict frailty was assessed
457 by ranking their 'importance.' This variable importance was quantified in terms of the percentage increase
458 in mean-squared error (MSE) when the effect of that variable was removed.

459 All statistical analyses were performed with R version 3.6.2 (R Core Team, 2019), with several general
460 packages for data processing (Wickham et al., 2020; Wickham & Henry, 2020) and for visualization (Pedersen,

461 2019; Wickham, 2016; Wilke, 2019). Correlations between inflammatory markers were visualized using the
462 corrplot package (Wei & Simko, 2017).

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468 6 Competing interests

469 None to declare.

470 7 References

- 471 Ahmadi-Abhari, S., Kaptoge, S., Luben, R. N., Wareham, N. J., & Khaw, K.-T. (2014). Longitudinal
472 Association of C-Reactive Protein and Lung Function Over 13 Years: The EPIC-Norfolk Study. *American*
473 *Journal of Epidemiology*, *179*(1), 48–56. <https://doi.org/10.1093/aje/kwt208>
- 474 Aloui, C., Prigent, A., Tariket, S., Sut, C., Fagan, J., Cognasse, F., Chakroun, T., Garraud, O., & Laradi, S.
475 (2016). Levels of human platelet-derived soluble CD40 ligand depend on haplotypes of CD40LG-CD40 -
476 ITGA2. *Scientific Reports*, *6*(1), 24715. <https://doi.org/10.1038/srep24715>
- 477 Altara, R., Gu, Y.-M., Struijker-Boudier, H. A. J., Thijs, L., Staessen, J. A., & Blankesteyn, W. M. (2015).
478 Left Ventricular Dysfunction and CXCR3 Ligands in Hypertension: From Animal Experiments to a
479 Population-Based Pilot Study. *PLOS ONE*, *10*(10), e0141394. [https://doi.org/10.1371/journal.pone.](https://doi.org/10.1371/journal.pone.0141394)
480 [0141394](https://doi.org/10.1371/journal.pone.0141394)
- 481 Arger, N. K., Ho, M., Woodruff, P. G., & Koth, L. L. (2019). Serum CXCL11 correlates with pulmonary
482 outcomes and disease burden in sarcoidosis. *Respiratory Medicine*, *152*, 89–96. [https://doi.org/10.1016/](https://doi.org/10.1016/j.rmed.2019.04.005)
483 [j.rmed.2019.04.005](https://doi.org/10.1016/j.rmed.2019.04.005)
- 484 Baylis, D., Bartlett, D. B., Patel, H. P., & Roberts, H. C. (2013). Understanding how we age: Insights into
485 inflammaging. *Longevity & Healthspan*, *2*, 8–8. <https://doi.org/10.1186/2046-2395-2-8>

- 486 Beharka, A. A., Meydani, M., Wu, D., Leka, L. S., Meydani, A., & Meydani, S. N. (2001). Interleukin-
487 6 Production Does Not Increase With Age. *The Journals of Gerontology: Series A*, *56*(2), B81–B88.
488 <https://doi.org/10.1093/gerona/56.2.B81>
- 489 Benjamini, Y., & Hochberg, Y. (1995). Controlling the false discovery rate: A practical and powerful
490 approach to multiple testing. *Journal of the Royal Statistical Society. Series B (Methodological)*, *57*(1),
491 289–300. <https://doi.org/10.2307/2346101>
- 492 Bupp, M. R. G. (2015). Sex, the aging immune system, and chronic disease. *Cellular Immunology*, *294*(2),
493 102–110. <https://doi.org/10.1016/j.cellimm.2015.02.002>
- 494 Calvani, R., Marini, F., Cesari, M., Buford, T. W., Manini, T. M., Pahor, M., Leeuwenburgh, C., Bern-
495 abei, R., Landi, F., & Marzetti, E. (2017). Systemic inflammation, body composition, and physical
496 performance in old community-dwellers. *Journal of Cachexia, Sarcopenia and Muscle*, *8*(1), 69–77.
497 <https://doi.org/10.1002/jcsm.12134>
- 498 Cesari, M., Gambassi, G., van Kan, G. A., & Vellas, B. (2014). The frailty phenotype and the frailty index:
499 Different instruments for different purposes. *Age Ageing*, *43*(1), 10–12. [https://doi.org/10.1093/ageing/
500 aft160](https://doi.org/10.1093/ageing/aft160)
- 501 Cesari, M., Penninx, B. W. J. H., Pahor, M., Lauretani, F., Corsi, A. M., Williams, G. R., Guralnik, J.
502 M., & Ferrucci, L. (2004). Inflammatory Markers and Physical Performance in Older Persons: The
503 InCHIANTI Study. *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*,
504 *59*(3), M242–M248. <https://doi.org/10.1093/gerona/59.3.M242>
- 505 Collerton, J., Martin-Ruiz, C., Davies, K., Hilkens, C. M., Isaacs, J., Kolenda, C., Parker, C., Dunn, M.,
506 Catt, M., Jagger, C., von Zglinicki, T., & Kirkwood, T. B. (2012). Frailty and the role of inflammation,
507 immunosenescence and cellular ageing in the very old: Cross-sectional findings from the Newcastle 85+
508 Study. *Mech Ageing Dev*, *133*(6), 456–466. <https://doi.org/10.1016/j.mad.2012.05.005>
- 509 Crow, R. S., Lohman, M. C., Titus, A. J., Cook, S. B., Bruce, M. L., Mackenzie, T. A., Bartels, S. J., &
510 Batsis, J. A. (2019). Association of Obesity and Frailty in Older Adults: NHANES 1999. *The Journal
511 of Nutrition, Health & Aging*, *23*(2), 138–144. <https://doi.org/10.1007/s12603-018-1138-x>
- 512 Damien, P., Cognasse, F., Eyraud, M.-A., Arthaud, C.-A., Pozzetto, B., Garraud, O., & Hamzeh-Cognasse,
513 H. (2015). LPS stimulation of purified human platelets is partly dependent on plasma soluble CD14 to
514 secrete their main secreted product, soluble-CD40-Ligand. *BMC Immunology*, *16*(1). [https://doi.org/
515 10.1186/s12865-015-0067-2](https://doi.org/10.1186/s12865-015-0067-2)

- 516 Danese, S. (2003). Activated platelets are the source of elevated levels of soluble CD40 ligand in the circula-
517 tion of inflammatory bowel disease patients. *Gut*, *52*(10), 1435–1441. [https://doi.org/10.1136/gut.52.](https://doi.org/10.1136/gut.52.10.1435)
518 10.1435
- 519 Dieteren, C. M., Samson, L. D., Schipper, M., van Exel, J., Brouwer, W. B. F., Verschuren, W. M. M., &
520 Picavet, H. S. J. (2020). The Healthy Aging Index analyzed over 15 years in the general population: The
521 Doetinchem Cohort Study. *Preventive Medicine*, 106193. <https://doi.org/10.1016/j.ypmed.2020.106193>
- 522 Ellulu, M. S., Patimah, I., Khaza'ai, H., Rahmat, A., & Abed, Y. (2017). Obesity and inflammation: The
523 linking mechanism and the complications. *Archives of Medical Science*, *4*, 851–863. [https://doi.org/10.](https://doi.org/10.5114/aoms.2016.58928)
524 5114/aoms.2016.58928
- 525 Enright, P. L., Johnson, L. R., Connett, J. E., Voelker, H., & Buist, A. S. (1991). Spirometry in the
526 Lung Health Study: 1. Methods and Quality Control. *American Review of Respiratory Disease*, *143*(6),
527 1215–1223. <https://doi.org/10.1164/ajrccm/143.6.1215>
- 528 Ferrucci, L., Corsi, A., Lauretani, F., Bandinelli, S., Bartali, B., Taub, D. D., Guralnik, J. M., & Longo,
529 D. L. (2005). The origins of age-related proinflammatory state. *Blood*, *105*(6), 2294–2299. <https://doi.org/10.1182/blood-2004-07-2599>
- 531 Franceschi, C., Bonafè, M., Valensin, S., Olivieri, F., De Luca, M., Ottaviani, E., & De Benedictis, G. (2006).
532 Inflamm-aging: An Evolutionary Perspective on Immunosenesence. *Annals of the New York Academy*
533 *of Sciences*, *908*(1), 244–254. <https://doi.org/10.1111/j.1749-6632.2000.tb06651.x>
- 534 Franceschi, C., Garagnani, P., Vitale, G., Capri, M., & Salvioli, S. (2017). Inflammaging and ‘Garb-aging.’
535 *Trends in Endocrinology & Metabolism*, *28*(3), 199–212. <https://doi.org/10.1016/j.tem.2016.09.005>
- 536 Furman, D., Hejblum, B. P., Simon, N., Jovic, V., Dekker, C. L., Thiebaut, R., Tibshirani, R. J., & Davis,
537 M. M. (2014). Systems analysis of sex differences reveals an immunosuppressive role for testosterone in
538 the response to influenza vaccination. *Proceedings of the National Academy of Sciences*, *111*(2), 869–874.
539 <https://doi.org/10.1073/pnas.1321060111>
- 540 Gale, C. R., Baylis, D., Cooper, C., & Sayer, A. A. (2013). Inflammatory markers and incident frailty in
541 men and women: The English Longitudinal Study of Ageing. *Age (Dordr)*, *35*(6), 2493–2501. <https://doi.org/10.1007/s11357-013-9528-9>
- 542
- 543 Ghigliotti, G., Barisione, C., Garibaldi, S., Fabbi, P., Brunelli, C., Spallarossa, P., Altieri, P., Rosa, G.,
544 Spinella, G., Palombo, D., Arsenescu, R., & Arsenescu, V. (2014). Adipose Tissue Immune Response:
545 Novel Triggers and Consequences for Chronic Inflammatory Conditions. *Inflammation*, *37*(4), 1337–1353.

546 <https://doi.org/10.1007/s10753-014-9914-1>

547 Giuliani, N., Sansoni, P., Girasole, G., Vescovini, R., Passeri, G., Passeri, M., & Pedrazzoni, M. (2001).
548 Serum interleukin-6, soluble interleukin-6 receptor and soluble Gp130 exhibit different patterns of age- and
549 menopause-related changes. *Experimental Gerontology*, *36*(3), 547–557. [https://doi.org/10.1016/S0531-](https://doi.org/10.1016/S0531-5565(00)00220-5)
550 [5565\(00\)00220-5](https://doi.org/10.1016/S0531-5565(00)00220-5)

551 Goldeck, D., Pawelec, G., Norman, K., Steinhagen-Thiessen, E., Oettinger, L., Haehnel, K., & Demuth, I.
552 (2016). No strong correlations between serum cytokine levels, CMV serostatus and hand-grip strength in
553 older subjects in the Berlin BASE-II cohort. *Biogerontology*, *17*(1), 189–198. [https://doi.org/10.1007/](https://doi.org/10.1007/s10522-015-9577-9)
554 [s10522-015-9577-9](https://doi.org/10.1007/s10522-015-9577-9)

555 Gordon, E. H., Peel, N. M., Samanta, M., Theou, O., Howlett, S. E., & Hubbard, R. E. (2017). Sex
556 differences in frailty: A systematic review and meta-analysis. *Experimental Gerontology*, *89*, 30–40.
557 <https://doi.org/10.1016/j.exger.2016.12.021>

558 Hearps, A. C., Martin, G. E., Angelovich, T. A., Cheng, W.-J., Maisa, A., Landay, A. L., Jaworowski, A.,
559 & Crowe, S. M. (2012). Aging is associated with chronic innate immune activation and dysregulation
560 of monocyte phenotype and function. *Aging Cell*, *11*(5), 867–875. [https://doi.org/10.1111/j.1474-9726.](https://doi.org/10.1111/j.1474-9726.2012.00851.x)
561 [2012.00851.x](https://doi.org/10.1111/j.1474-9726.2012.00851.x)

562 Homey, B., Alenius, H., Müller, A., Soto, H., Bowman, E. P., Yuan, W., McEvoy, L., Lauerma, A. I.,
563 Assmann, T., Bünemann, E., Lehto, M., Wolff, H., Yen, D., Marxhausen, H., To, W., Sedgwick, J.,
564 Ruzicka, T., Lehmann, P., & Zlotnik, A. (2002). CCL27CCR10 interactions regulate T cellmediated skin
565 inflammation. *Nature Medicine*, *8*(2), 157–165. <https://doi.org/10.1038/nm0202-157>

566 Hothorn, T., Hornik, K., Wiel, M. A. van de, & Zeileis, A. (2008). Implementing a Class of Permutation
567 Tests: The **Coin** Package. *Journal of Statistical Software*, *28*(8). <https://doi.org/10.18637/jss.v028.i08>

568 Hsu, B., Hirani, V., Cumming, R. G., Naganathan, V., Blyth, F. M., Wright, F. C., Waite, L. M., Seibel,
569 M. J., Handelsman, D. J., & Le Couteur, D. G. (2019). Cross-Sectional and Longitudinal Relationships
570 Between Inflammatory Biomarkers and Frailty in Community-dwelling Older Men: The Concord Health
571 and Ageing in Men Project. *The Journals of Gerontology: Series A*, *74*(6), 835–841. [https://doi.org/10.](https://doi.org/10.1093/gerona/glx142)
572 [1093/gerona/glx142](https://doi.org/10.1093/gerona/glx142)

573 Huang, J., Jochems, C., Talaie, T., Anderson, A., Jales, A., Tsang, K. Y., Madan, R. A., Gulley, J. L., &
574 Schlom, J. (2012). Elevated serum soluble CD40 ligand in cancer patients may play an immunosuppressive
575 role. *Blood*, *120*(15), 3030–3038. <https://doi.org/10.1182/blood-2012-05-427799>

- 576 Hulsegge, G., Herber-Gast, G. C., Spijkerman, A. M., Susan, H., Picavet, J., van der Schouw, Y. T., Bakker,
577 S. J., Gansevoort, R. T., Dolle, M. E., Smit, H. A., & Monique Verschuren, W. M. (2016). Obesity and
578 Age-Related Changes in Markers of Oxidative Stress and Inflammation Across Four Generations. *Obesity*
579 *(Silver Spring)*, 24(6), 1389–1396. <https://doi.org/10.1002/oby.21515>
- 580 Jenabian, M. A., Patel, M., Kema, I., Vyboh, K., Kanagaratham, C., Radzioch, D., Thébault, P., Lapointe,
581 R., Gilmore, N., Ancuta, P., Tremblay, C., & Routy, J. P. (2014). Soluble CD40-ligand (sCD40L,
582 sCD154) plays an immunosuppressive role via regulatory T cell expansion in HIV infection. *Clinical &*
583 *Experimental Immunology*, 178(1), 102–111. <https://doi.org/10.1111/cei.12396>
- 584 Kameda, M., Otsuka, M., Chiba, H., Kuronuma, K., Hasegawa, T., Takahashi, H., & Takahashi, H. (2020).
585 CXCL9, CXCL10, and CXCL11; biomarkers of pulmonary inflammation associated with autoimmunity in
586 patients with collagen vascular diseases associated interstitial lung disease and interstitial pneumonia with
587 autoimmune features. *PLOS ONE*, 15(11), e0241719. <https://doi.org/10.1371/journal.pone.0241719>
- 588 Komsta, L. (2019). *Mblm: Median-based Linear Models*.
- 589 Krzywinski, M., & Altman, N. (2014). Analysis of variance and blocking. *Nature Methods*, 11(7), 699–700.
590 <https://doi.org/10.1038/nmeth.3005>
- 591 Leng, S. X., Xue, Q. L., Tian, J., Huang, Y., Yeh, S. H., & Fried, L. P. (2009). Associations of neutrophil and
592 monocyte counts with frailty in community-dwelling disabled older women: Results from the Women’s
593 Health and Aging Studies I. *Exp Gerontol*, 44(8), 511–516. <https://doi.org/10.1016/j.exger.2009.05.005>
- 594 Liaw, A., & Wiener, M. (2002). Classification and regression by randomForest. *R News*, 2(3), 18–22.
- 595 Lloyd-Jones, K. L., Kelly, M. M., & Kubes, P. (2008). Varying Importance of Soluble and Membrane
596 CD14 in Endothelial Detection of Lipopolysaccharide. *The Journal of Immunology*, 181(2), 1446–1453.
597 <https://doi.org/10.4049/jimmunol.181.2.1446>
- 598 Lubin, J. H., Colt, J. S., Camann, D., Davis, S., Cerhan, J. R., Severson, R. K., Bernstein, L., & Hartge, P.
599 (2004). Epidemiologic Evaluation of Measurement Data in the Presence of Detection Limits. *Environ-*
600 *mental Health Perspectives*, 112(17), 1691–1696. <https://doi.org/10.1289/ehp.7199>
- 601 Mitnitski, A. B., Mogilner, A. J., & Rockwood, K. (2001). Accumulation of deficits as a proxy measure of
602 aging. *Scientific World Journal*, 1, 323–336. <https://doi.org/10.1100/tsw.2001.58>
- 603 Morrisette-Thomas, V., Cohen, A. A., Fülöp, T., Riesco, É., Legault, V., Li, Q., Milot, E., Dusseault-
604 Bélanger, F., & Ferrucci, L. (2014). Inflamm-aging does not simply reflect increases in pro-inflammatory

- 605 markers. *Mechanisms of Ageing and Development*, 139, 49–57. [https://doi.org/10.1016/j.mad.2014.06.](https://doi.org/10.1016/j.mad.2014.06.005)
606 005
- 607 Nagasawa, M., Zhu, Y., Isoda, T., Tomizawa, D., Itoh, S., Kajiwara, M., Morio, T., Nonoyama, S., Shimizu,
608 N., & Mizutani, S. (2005). Analysis of serum soluble CD40 ligand (sCD40L) in the patients undergoing
609 allogeneic stem cell transplantation: Platelet is a major source of serum sCD40L. *European Journal of*
610 *Haematology*, 74(1), 54–60. <https://doi.org/10.1111/j.1600-0609.2004.00342.x>
- 611 Nooyens, A. C. J., Bueno-de-Mesquita, H. B., van Boxtel, M. P. J., van Gelder, B. M., Verhagen, H., &
612 Verschuren, W. M. M. (2011). Fruit and vegetable intake and cognitive decline in middle-aged men
613 and women: The Doetinchem Cohort Study. *British Journal of Nutrition*, 106(5), 752–761. <https://doi.org/10.1017/S0007114511001024>
- 614
- 615 Ouchi, N., Parker, J. L., Lugus, J. J., & Walsh, K. (2011). Adipokines in inflammation and metabolic
616 disease. *Nature Reviews Immunology*, 11(2), 85–97. <https://doi.org/10.1038/nri2921>
- 617 Pedersen, T. L. (2019). *Patchwork: The composer of plots* [Manual].
- 618 Picavet, H. S. J., Blokstra, A., Spijkerman, A. M. W., & Verschuren, W. M. M. (2017). Cohort Profile
619 Update: The Doetinchem Cohort Study 1987-2017: Lifestyle, health and chronic diseases in a life course
620 and ageing perspective. *Int J Epidemiol*, 46(6), 1751–1751g. <https://doi.org/10.1093/ije/dyx103>
- 621 Puzianowska-Kuźnicka, M., Owczarz, M., Wieczorowska-Tobis, K., Nadrowski, P., Chudek, J., Slusarczyk,
622 P., Skalska, A., Jonas, M., Franek, E., & Mossakowska, M. (2016). Interleukin-6 and C-reactive protein,
623 successful aging, and mortality: The PolSenior study. *Immunity & Ageing*, 13(1), 21. [https://doi.org/](https://doi.org/10.1186/s12979-016-0076-x)
624 10.1186/s12979-016-0076-x
- 625 Quanjer, P. H., Stanojevic, S., Cole, T. J., Baur, X., Hall, G. L., Culver, B. H., Enright, P. L., Hankinson, J.
626 L., Ip, M. S. M., Zheng, J., Stocks, J., & the ERS Global Lung Function Initiative. (2012). Multi-ethnic
627 reference values for spirometry for the 3-yr age range: The global lung function 2012 equations. *European*
628 *Respiratory Journal*, 40(6), 1324–1343. <https://doi.org/10.1183/09031936.00080312>
- 629 R Core Team. (2019). *R: A language and environment for statistical computing* [Manual]. R Foundation for
630 Statistical Computing.
- 631 Richmond, J. M., Strassner, J. P., Essien, K. I., & Harris, J. E. (2019). T-cell positioning by chemokines in
632 autoimmune skin diseases. *Immunological Reviews*, 289(1), 186–204. <https://doi.org/10.1111/imr.12762>
- 633 Samson, L. D., Boots, A. M. H., Ferreira, J. A., Picavet, H. S. J., de Rond, L. G. H., de Zeeuw-Brouwer,
634 M., Verschuren, W. M. M., Buisman, A.-M., & Engelfriet, P. (2020). In-depth immune cellular profiling

- 635 reveals sex-specific associations with frailty. *Immunity & Ageing*, 17(1), 20. <https://doi.org/10.1186/s12979-020-00191-z>
- 636
- 637 Samson, L. D., Boots, A. M. H., Verschuren, W. M. M., Picavet, H. S. J., Engelfriet, P., & Buisman, A.-
638 M. (2019). Frailty is associated with elevated CRP trajectories and higher numbers of neutrophils and
639 monocytes. *Experimental Gerontology*, 125, 110674. <https://doi.org/10.1016/j.exger.2019.110674>
- 640 Shive, C. L., Jiang, W., Anthony, D. D., & Lederman, M. M. (2015). Soluble CD14 is a nonspecific marker
641 of monocyte activation. *AIDS (London, England)*, 29(10), 1263–1265. <https://doi.org/10.1097/QAD.0000000000000735>
- 642
- 643 Smith, L., Yang, L., & Hamer, M. (2019). Handgrip strength, inflammatory markers, and mortality. *Scandinavian Journal of Medicine & Science in Sports*, sms.13433. <https://doi.org/10.1111/sms.13433>
- 644
- 645 Suttles, J., & Stout, R. D. (2009). Macrophage CD40 signaling: A pivotal regulator of disease protection and
646 pathogenesis. *Seminars in Immunology*, 21(5), 257–264. <https://doi.org/10.1016/j.smim.2009.05.011>
- 647 Theil, H. (1950). A rank-invariant method of linear and polynomial regression analysis. *Indagationes Mathematicae*, 12, 85–91.
- 648
- 649 Tuttle, C. S. L., Thang, L. A. N., & Maier, A. B. (2020). Markers of inflammation and their association
650 with muscle strength and mass: A systematic review and meta-analysis. *Ageing Research Reviews*, 64,
651 101185. <https://doi.org/10.1016/j.arr.2020.101185>
- 652 United Nations. (2017). *World Population Ageing 2015*. UN. <https://doi.org/10.18356/88fa44e7-en>
- 653 Van Epps, P., Oswald, D., Higgins, P. A., Hornick, T. R., Aung, H., Banks, R. E., Wilson, B. M., Burant,
654 C., Gravenstein, S., & Canaday, D. H. (2016). Frailty has a stronger association with inflammation than
655 age in older veterans. *Immunity & Ageing*, 13(1), 27. <https://doi.org/10.1186/s12979-016-0082-z>
- 656 van Oostrom, S. H., Engelfriet, P. M., Verschuren, W. M. M., Schipper, M., Wouters, I. M., Boezen,
657 M., Smit, H. A., Kerstjens, H. A. M., & Picavet, H. S. J. (2018). Aging-related trajectories of lung
658 function in the general populationThe Doetinchem Cohort Study. *PLOS ONE*, 13(5), e0197250. <https://doi.org/10.1371/journal.pone.0197250>
- 659
- 660 Verschuren, W. M., Blokstra, A., Picavet, H. S., & Smit, H. A. (2008). Cohort profile: The Doetinchem
661 Cohort Study. *Int J Epidemiol*, 37(6), 1236–1241. <https://doi.org/10.1093/ije/dym292>
- 662 Walker, K. A., Walston, J., Gottesman, R. F., Kucharska-Newton, A., Palta, P., & Windham, B. G. (2018).
663 Midlife Systemic Inflammation is Associated with Frailty in Later Life: The ARIC Study. *J Gerontol A Biol Sci Med Sci*, gly045–gly045. <https://doi.org/10.1093/gerona/gly045>
- 664

- 665 Wei, T., & Simko, V. (2017). *R package "corrplot": Visualization of a correlation matrix* [Manual].
- 666 Wickham, H. (2016). *Ggplot2: Elegant graphics for data analysis*. Springer-Verlag New York.
- 667 Wickham, H., François, R., Henry, L., & Müller, K. (2020). *Dplyr: A grammar of data manipulation*
668 [Manual].
- 669 Wickham, H., & Henry, L. (2020). *Tidyr: Tidy messy data* [Manual].
- 670 Wilcox, R. (1998). A Note on the Theil-Sen Regression Estimator When the Regressor Is Random and the
671 Error Term Is Heteroscedastic. *Biometrical Journal*, 40(3), 261–268. [https://doi.org/10.1002/\(sici\)1521-](https://doi.org/10.1002/(sici)1521-4036(199807)40:3%3C261::Aid-bimj261%3E3.0.Co;2-v)
672 [4036\(199807\)40:3%3C261::Aid-bimj261%3E3.0.Co;2-v](https://doi.org/10.1002/(sici)1521-4036(199807)40:3%3C261::Aid-bimj261%3E3.0.Co;2-v)
- 673 Wilke, C. O. (2019). *Cowplot: Streamlined plot theme and plot annotations for 'Ggplot2'* [Manual].

674 8 Supplementary information

Table S1: Summary characteristics of the cytokines and chemokines in the cohort

Protein	Baseline concentration	Endpoint concentration	AUC
C-reactive protein			
CRP	1.03(0.93 – 1.14) * 10 ⁶	1.22(1.11 – 1.34) * 10 ⁶	1.56(1.43 – 1.70) * 10 ⁶
CC chemokines			
CCL1/I-309	3.30(3.17 – 3.43)	3.34(3.22 – 3.46)	3.38(3.27 – 3.50)
CCL2/MCP-1	1.03(1.00 – 1.06) * 10 ²	1.07(1.04 – 1.10) * 10 ²	1.08(1.06 – 1.11) * 10 ²
CCL5/RANTES	8.83(8.42 – 9.26) * 10 ⁴	7.60(7.27 – 7.95) * 10 ⁴	8.00(7.73 – 8.27) * 10 ⁴
CCL11/Eotaxin	4.76(4.64 – 4.89)	4.97(4.84 – 5.11)	5.08(4.97 – 5.19)
CCL27/C-TACK	9.34(8.97 – 9.74) * 10 ²	1.03(0.99 – 1.08) * 10 ³	1.08(1.05 – 1.12) * 10 ³
CXC chemokines			
CXCL9/MIG	3.08(2.95 – 3.23)	2.98(2.85 – 3.11)	3.15(3.05 – 3.26)
CXCL10/IP-10	2.37(2.29 – 2.46) * 10 ²	2.83(2.73 – 2.93) * 10 ²	3.01(2.92 – 3.10) * 10 ²
CXCL11/I-TAC	6.07(5.64 – 6.53)	6.19(5.76 – 6.65)	6.98(6.59 – 7.39)
Interleukins			
IL-6	2.43(2.23 – 2.65)	2.31(2.13 – 2.51)	2.61(2.45 – 2.78)
IL-10	0.71(0.66 – 0.77)	0.66(0.62 – 0.72)	0.75(0.71 – 0.80)
Soluble receptors			
sCD14	2.15(2.10 – 2.21) * 10 ⁶	2.21(2.15 – 2.27) * 10 ⁶	2.27(2.22 – 2.32) * 10 ⁶
sCD40L	4.14(3.90 – 4.40) * 10 ²	3.69(3.48 – 3.91) * 10 ²	4.05(3.87 – 4.24) * 10 ²
sIL-6R	2.27(2.22 – 2.32) * 10 ⁴	2.35(2.30 – 2.41) * 10 ⁴	2.37(2.33 – 2.42) * 10 ⁴
Other			
P selectin	7.95(7.43 – 8.50) * 10 ⁴	6.80(6.40 – 7.22) * 10 ⁴	8.04(7.73 – 8.36) * 10 ⁴
sGP130	2.19(2.16 – 2.22) * 10 ⁴	2.23(2.20 – 2.26) * 10 ⁴	2.25(2.23 – 2.27) * 10 ⁴
C5a	4.65(4.38 – 4.93) * 10 ³	4.90(4.62 – 5.18) * 10 ³	5.06(4.79 – 5.34) * 10 ³
BDNF	1.85(1.79 – 1.91) * 10 ⁴	1.69(1.63 – 1.77) * 10 ⁴	1.75(1.69 – 1.81) * 10 ⁴

Note:

Concentrations ($pg * mL^{-1}$) and AUC values ($years * pg * mL^{-1}$) are geometric mean values with 95% confidence intervals.

Table S2: Frailty index components

No	Frailty index component	Description	Value: 0	Value: 0.5	Value: 1
1	RR	High (systolic) blood pressure	$RR < 160$		$RR \geq 160$

Table S2: Frailty index components (*continued*)

No	Frailty index component	Description	Value: 0	Value: 0.5	Value: 1
2	Cardiologic Disease	One or more of the following conditions: myocardial infarction, bypass-surgery, balloon dilatation, cardiac catheterization, pacemaker implantation, large blood vessel surgery, hospitalization due to cardiac failure	No		One or more prevalent
3	Diabetes	Prevalence of Diabetes	No		Yes
4	Hearing	Inability to maintain a conversation in a group of 3 or more people due to hearing impairment (with hearing aid if needed)	Yes or with some effort		No or with great effort
5	Malignancy	(History of) any form of malignancy	No		Yes
6	Joint Inflammation	Chronic joint inflammation in the past year	No		Yes
7	Osteoporosis	Osteoporosis diagnosed by a medical doctor, in the past year,	No		Yes
8	Lower Back Pain	Severe lower back complaints in the past year (including lumbar herniated nucleus pulposus)	No		Yes
9	CVA	(History of) stroke	No		Yes
10	Migraine	Migraine prevalence in the past year	No		Yes

Table S2: Frailty index components (*continued*)

No	Frailty index component	Description	Value: 0	Value: 0.5	Value: 1
11	Neurologic Disease	One or more of the following neurological diseases, diagnosed by a medical doctor: m. Parkinson, Multiple Sclerosis, epilepsy	No		Yes
12	Asthma	Asthma diagnosed by a doctor and one or more asthma attacks in the past year	No		Yes
13	Spirometry Ratio	Poor lung function quantified by spirometry measurements: first second of forced expiration divided by the forced vital capacity (FEV1/FVC)	$FEV1/FVC > 0.70$		$FEV1/FVC \leq 0.70$
14	Digestive Tract	Severe bowel disorders in the past year, diagnosed by a medical doctor	No		Yes
15	Vertigo	Vertigo with falling the past 12 months	No vertigo	Some ver-tigo	Severe vertigo
16	Pain	Limited in daily activities due to pain	No limitation	Some limi-tation	Severe limitation
17	Incontinence	Unintentional urine incontinence past 12 months	No		Yes
18	Ankle Brachial Index	Poor ankle brachial index (ABI, the ratio of the systolic blood pressure in the ankle to the blood pressure in the arm)	$ABI > 0.9$		$ABI \leq 0.9$

Table S2: Frailty index components (*continued*)

No	Frailty index component	Description	Value: 0	Value: 0.5	Value: 1
19	Health Perception	(Subjective) perception of poor health	No or some impairment		Severe impairment
20	Eyesight	Bad eyesight perception, facial recognition within 4 meters (with glasses/ contact lenses if needed)	No		Yes
21	Renal Function	Poor renal function (estimated Glomular Filtration Rate (eGFR), calculated with plasma creatinine concentrations) (Inker et al., 2012)	$eGFR \geq 60$		$eGFR < 60$
22	Cognitive Speed	Poor cognitive speed. Z-scores corrected for measurements per person and for education level. Scores derived from the Stroop Color-Word Test and the Letter-Digit Substitution Test, as described previously (Nooyens et al., 2011)	Not belonging to 10% participants with lowest z-score within the Doetinchem cohort		Belonging to 10% participants with lowest z-score within the Doetinchem cohort
23	Cognitive Memory	Poor cognitive memory. Z-scores corrected for measurements per person and for education level. Scores derived from the Verbal Learning Test, as described previously (Nooyens et al., 2011)	Not belonging to 10% participants with lowest z-score within the Doetinchem cohort		Belonging to 10% participants with lowest z-score within the Doetinchem cohort

Table S2: Frailty index components (*continued*)

No	Frailty index component	Description	Value: 0	Value: 0.5	Value: 1
24	Cognitive Flexibility	Poor cognitive flexibility. Z-scores corrected for measurements per person and education level. Scores derived from the Stroop Color-Word Test, as described previously (Nooyens et al., 2011)	Not belonging to 10% participants with lowest z-score within the Doetinchem cohort		Belonging to 10% participants with lowest z-score within the Doetinchem cohort
25	Physical Inactive	Not meeting the Dutch healthy exercise norm.(Kemper, 2000). In addition: belonging to the 25th lowest percentile of walking activity and the 10th percentile lowest low/medium/high intensive activities in the Doetinchem cohort	No		Yes
26	ADL	Limited in washing and dressing due to health	No limitation	Some limitation	Severe limitation
27	Household	Limited in daily activities (cooking, cleaning) due to health	No limitatlion	Some limitation	Severe limitation
28	Walking	Limited in in walking 100 meters	No limitatlion	Some limitation	Severe limitation
29	Lifting	Limited in lifting or carrying groceries due to health	No limitatlion	Some limitation	Severe limitation

Table S2: Frailty index components (*continued*)

No	Frailty index component	Description	Value: 0	Value: 0.5	Value: 1
30	Walking Stairs	Limited in climbing stairs	No limitatlion	Some limi- tation	Severe limitation
31	Grip Strength	Poor grip strength (cutoff points as described previously) (Fried et al., 2001)			
		<i>Men, BMI</i> ≤ 24	> 29		≤ 29
		<i>Men, 24 < BMI</i> ≤ 26	> 30		≤ 30
		<i>Men, 26 < BMI</i> ≤ 28	> 30		≤ 30
		<i>Men, BMI</i> > 28	> 32		≤ 32
		<i>Women, BMI</i> ≤ 24	> 17		≤ 17
		<i>Women, 23 < BMI</i> ≤ 26	> 17.3		≤ 17.3
		<i>Women, 26 < BMI</i> ≤ 29	> 18		≤ 18
		<i>Women, BMI</i> > 29	> 21		≤ 21
32	Depressed	Feeling depressed the past week	No limitatlion	Some limi- tation	Severe limitation
33	Happiness	Feeling unhappy the past 4 weeks	No limitatlion	Some limi- tation	Severe limitation
34	Mental Effort	Feeling as if every activity costs effort during the past week	No limitatlion	Some limi- tation	Severe limitation
35	Getting Going	Not being able to get going the past week	No limitatlion	Some limi- tation	Severe limitation

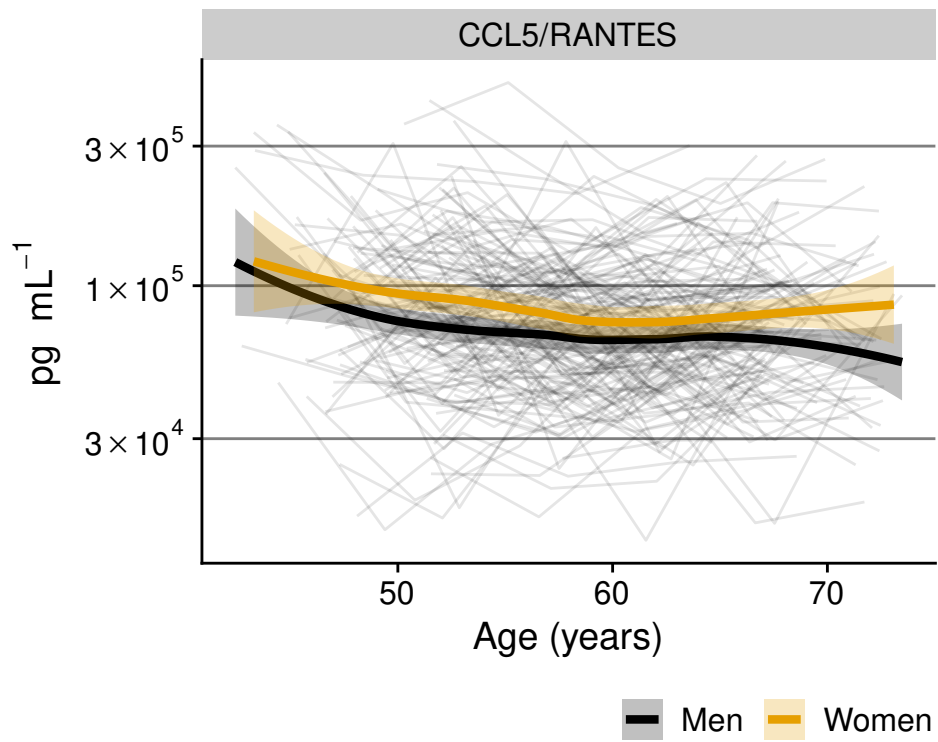


Figure S1: The concentration of CCL5/RANTES over 20 years in men and women (showing that this is continuously higher in women).

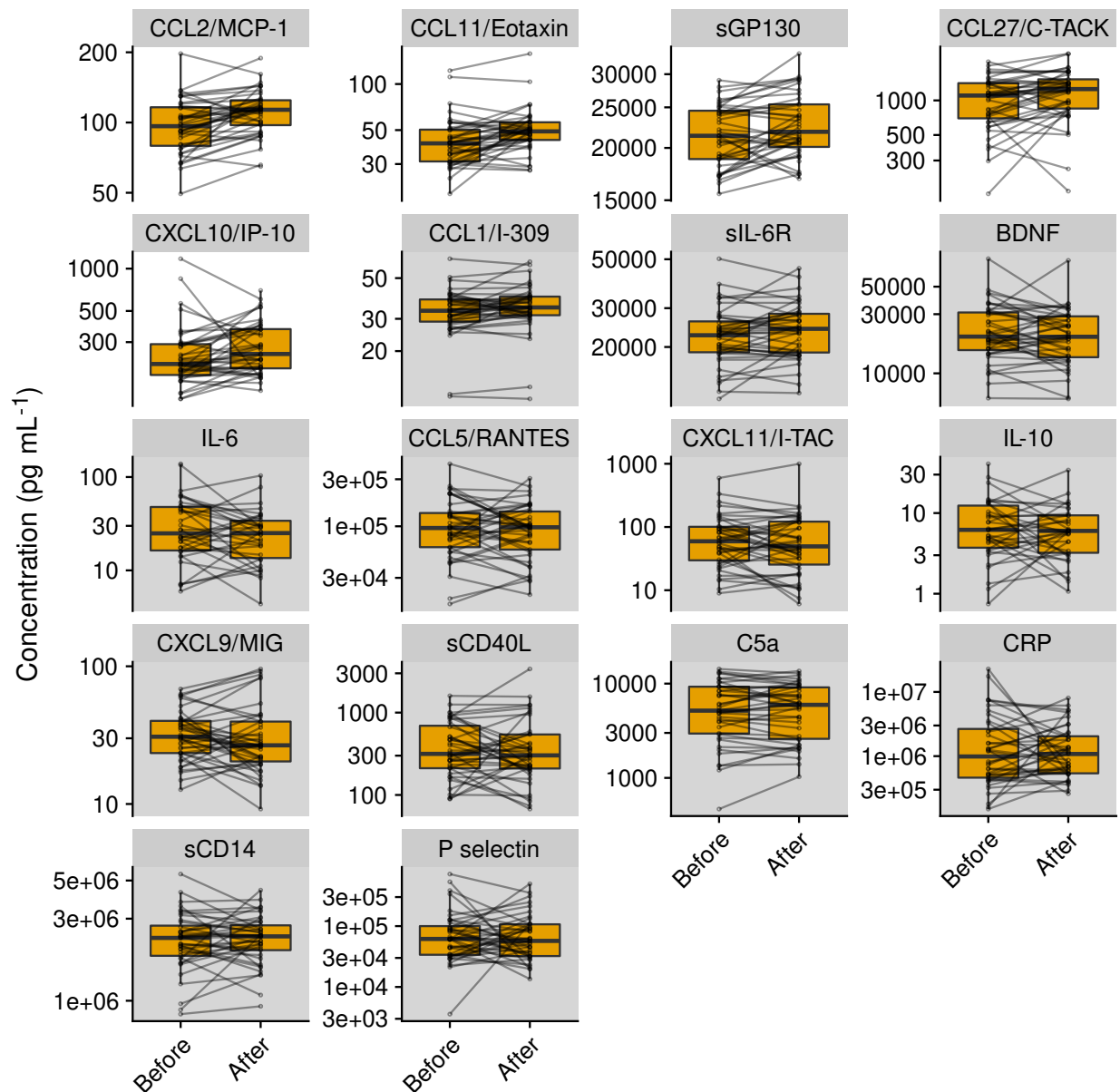


Figure S2: Menopause is related to changes in inflammatory marker profile. Inflammatory marker concentrations are shown shortly before and shortly after menopause (average difference: 5.3 years) in women of whom data at both timepoints are available (n=40/70). Tiles of biomarkers in which an association was found with menopause are shown in a white background, others in a grey background.

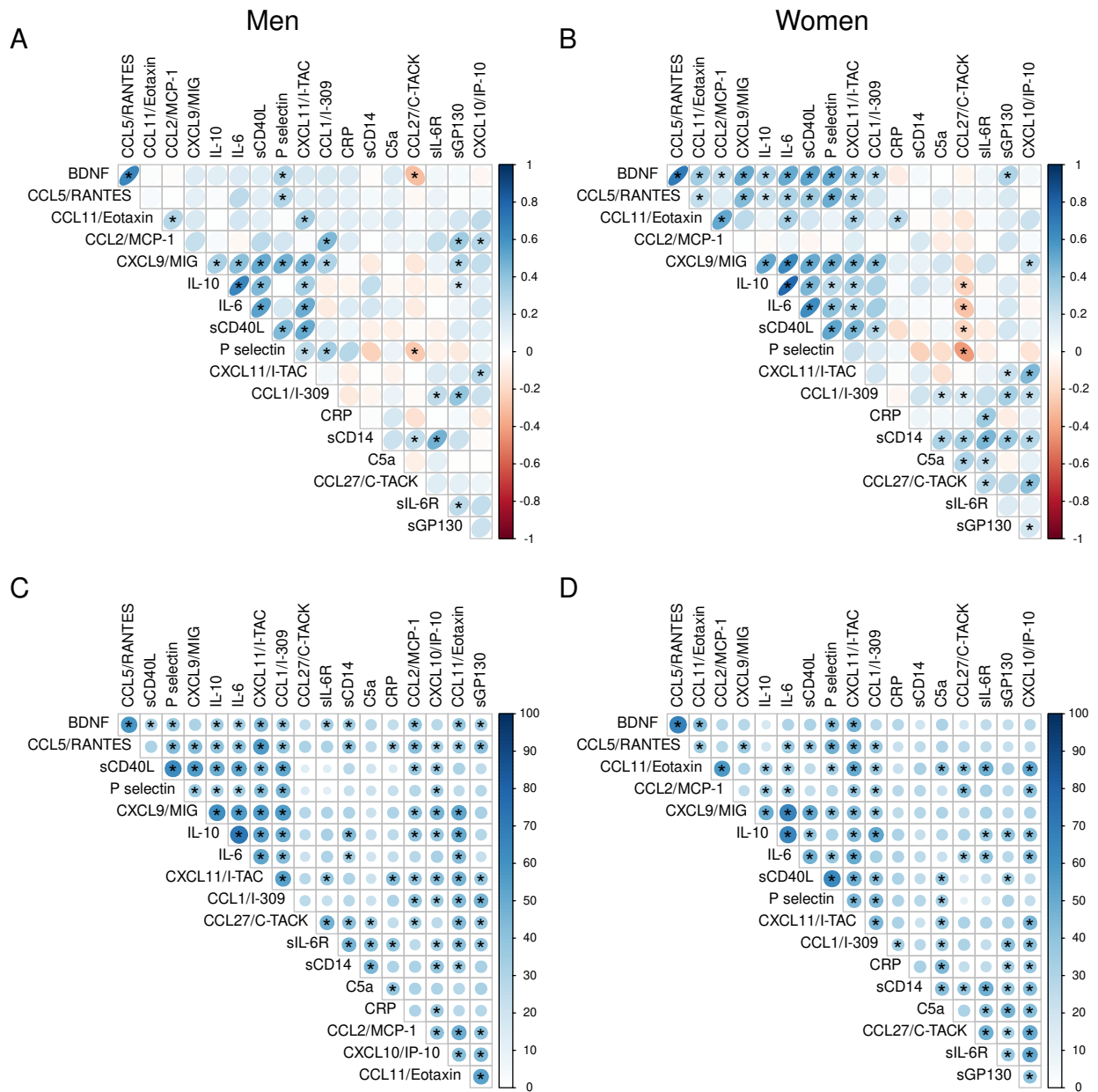


Figure S3: Relationships between inflammatory markers shown as (A,B) correlation between pairs of inflammatory markers at study endpoint and (C,D) similarity between pairs of inflammatory marker trajectories during about 20years of follow-up. In (A,B) the direction and strength of the association is visualized with an oval shape and a color gradient. In (C,D) the blue gradient color and the size of the circles shows the percentage of participants of which a pair of biomarkers had the highest increase in concentration at the same moment in 20 years of follow-up. * = an association between two inflammatory markers, with false discovery rate being set at a maximum of 15%. n=71 women, n=73 men.

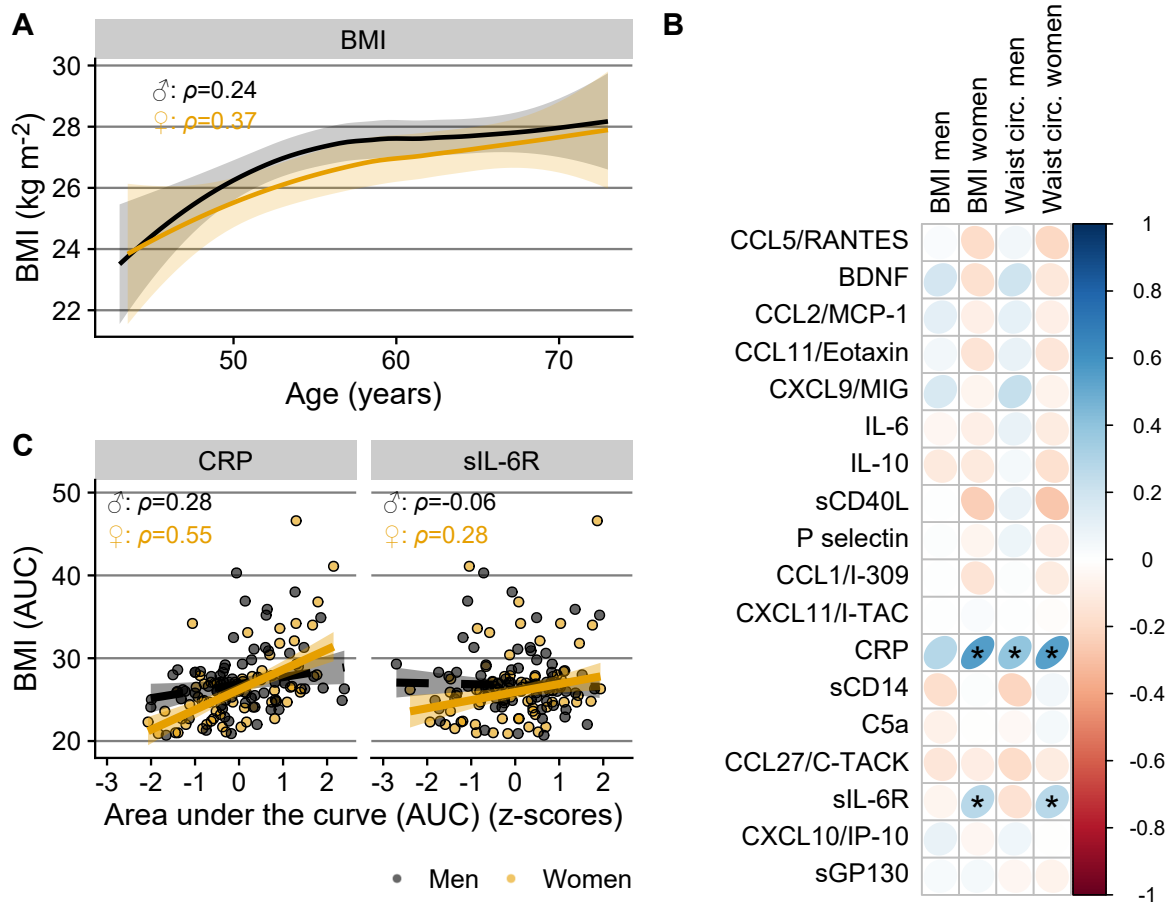


Figure S4: Relationship between body mass and inflammatory markers. (A) Local polynomial regression lines showing change in BMI over time in men and women. (B) Relationship of inflammatory marker levels with BMI and waist circumference of men (n=73) and women (n=71). Direction and strength of the associations in (B) are visualized with an oval shape and a color gradient. To capture the cumulative “exposure,” both BMI and inflammatory markers in (B) and (C) are expressed as area under the curve of levels/concentrations versus time, standardized to take into account different follow-up periods, and transformed into z-values. (C) BMI values (AUC) related to the AUC of CRP and sIL-6R. Trendlines in (C) are (robust) linear regression lines with 95% confidence interval.