PDBrenum: a webserver and program providing Protein Data Bank files renumbered according to their UniProt sequences

Bulat Faezov[1,2]
Roland L. Dunbrack, Jr[2,*]

[1] Institute of Fundamental Medicine and Biology
Kazan Federal University
Kazan, 420008, Russian Federation

[2] Institute for Cancer Research
Fox Chase Cancer Center
333 Cottman Avenue
Philadelphia PA 19111
USA

* Correspondence: roland.dunbrack@fccc.edu

May 5, 2021

Abstract

The Protein Data Bank (PDB) was established at Brookhaven National Laboratories in 1971 as an archive for biological macromolecular crystal structures. In early 2021, the database has more than 175,000 structures solved by X-ray crystallography, nuclear magnetic resonance, cryo-electron microscopy, and other methods. Many proteins have been studied under different conditions, including binding partners such as ligands, nucleic acids, or other proteins; mutations, and post-translational modifications, thus enabling extensive comparative structure-function studies. However, these studies are made more difficult because authors are allowed by the PDB to number the amino acids in each protein sequence in any manner they wish. This results in the same protein being numbered differently in the available PDB entries. For instance, some authors may include N-terminal signal peptides or the N-terminal methionine in the sequence numbering and others may not. In addition to the coordinates, there are many fields that contain information regarding specific residues in the sequence of each protein in the entry. Here we provide a webserver and Python3 application that fixes the PDB sequence numbering problem by replacing the author numbering with numbering derived from the corresponding UniProt sequences. We obtain this correspondence from the SIFTS database from PDBe. The server and program can take a list of PDB entries or a list of UniProt identifiers (e.g., "P04637" or "P53_HUMAN") and provide renumbered files in mmCIF format and the legacy PDB format for both asymmetric unit files and biological assembly files provided by PDBe.
Availability: Source code is freely available at https://github.com/Faezov/PDBrenum. The webserver is located at: http://dunbrack3.fccc.edu/PDBrenum.
Contact: bulat.faezov@fccc.edu or roland.dunbrack@fccc.edu.

Introduction

The Protein Data Bank (PDB) is a database for the three-dimensional structural data of biological macromolecules, including proteins and nucleic acids [1]. The data, typically obtained by X-ray crystallography, NMR spectroscopy, or cryo-electron microscopy, and submitted by scientists from around the world, are freely accessible through the World Wide PDB (wwPDB) http://www.wwpdb.org/ and three wwPDB partner sites, https://www.rcsb.org [2], https://www.ebi.ac.uk/pdbe [3], and https://pdbj.org [4]. The PDB provides useful and fundamental information about tens of thousands of proteins. For many proteins, there are 10s or even 100s of available structures performed under varying conditions, including the presence of different binding partners such as inhibitors, nucleic acids, or other proteins, or with mutations and post-translational modifications. However, in each structure in the PDB, authors are allowed to number protein sequences in any way they wish. This includes the coordinates and any functional or structural annotations contained within the PDB files. Authors commonly number according to sequences deposited in gene databanks such as GenBank [5] or UniProt [6]. So, for instance, a domain from a protein that is not at the N-terminus of the natural sequence may start with its position in the full-length sequence. However, different authors may choose different conventions for this numbering. Authors may or may not include the N-terminal methionine or N-terminal signal sequences in the numbering, both of which may be cleaved off to form the mature protein. For example, in PDB entry 3lvp [7], which is a structure of the kinase domain of human IGF1R, the DFG motif amino acids are numbered 1153-1155. But in PDB entry 3d94 [8], these residues are labeled as residues 1123-1125, because the numbering is that of the mature protein, which does not include the 30 amino acid signal sequence cleaved from the N-terminus of the preprotein. Proteins in the PDB often include N-terminal sequence tags, and the numbering of these residues can be just about anything including negative numbers, 0, or numbers that seem to indicate the residues are from the same gene as the protein under study. These inconsistent numbering schemes compromise structural bioinformatics studies that seek to compare multiple structures

of a single protein or structures within protein families across the PDB. They also affect mapping of sequence annotation data (such as mutation data) to structural information in the PDB, since any structure downloaded from the PDB may or may not have the same numbering scheme as the sequence database.

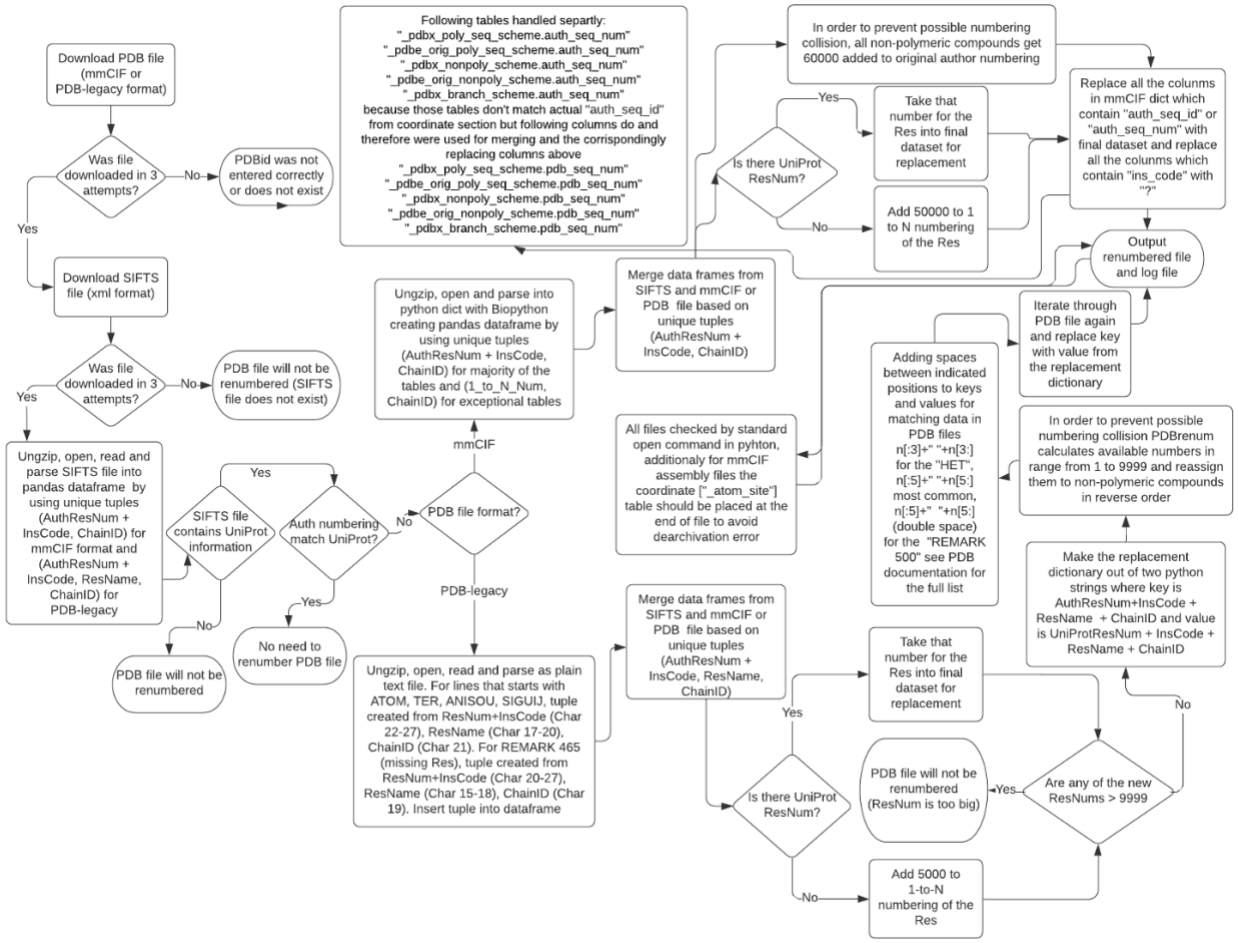

Figure 1. Flow-chart describing basic procedure of PDBrenum

The problem of inconsistent numbering, insertion codes, negative residue numbers, and other problems have been discussed previously but not addressed in any rigorous way (e.g. https://proteopedia.org/wiki/index.php/Unusual_sequence_numbering and https://www3.cmbi.umcn.nl/wiki/index.php/Residue_number). Mapping PDB structures to UniProt has been attempted a number of times, including SSMAP [9], Seq2Struct [10], and PDBSWS [11], although these servers did not renumber actual coordinate files, instead only providing the mapping of residue numbers from the PDB to UniProt. Only the PDBSWS server is still functioning.

In this paper, we present downloadable computer code in Python3 and a webserver that provide PDB files where the amino acids in all fields are renumbered according to their UniProt sequences. We obtain the correspondence between protein sequences in PDB chains with UniProt entries from the SIFTS database available from PDBe (https://www.ebi.ac.uk/pdbe/docs/sifts/) [12]. Our program and webserver provide renumbered

files in mmCIF [13] and legacy PDB format [14] for both asymmetric unit files (the coordinates deposited by authors) obtained from the RCSB server and biological assembly files (provided by the authors or calculated with the program PISA [15]) in mmCIF format obtained from PDBe, which distributes them for all PDB entries. The webserver is easy to use—the user enters a list of PDB codes ("1abc") or UniProt identifiers in the accession code ("P04637") or SwissProt ID ("P53_HUMAN") format, selects which kinds of files to download (mmCIF and/or legacy-PDB format; asymmetric units and/or biological assemblies), and with one click enables the download of a zip file containing the requested files.

(A)

PDBe	PDB	UniProt	AccessionID	UniProt_50k
(1, ARG, A)	(null, ARG, A)	50000	NaN	50001
(2, GLY, A)	(null, GLY, A)	50000	NaN	50002
(3, SER, A)	(null, SER, A)	50000	NaN	50003
(4, HIS, A)	(null, HIS, A)	50000	NaN	50004
(5, HIS, A)	(null, HIS, A)	50000	NaN	50005
(6, HIS, A)	(null, HIS, A)	50000	NaN	50006
(7, HIS, A)	(null, HIS, A)	50000	NaN	50007
(8, HIS, A)	(null, HIS, A)	50000	NaN	50008
(9, HIS, A)	(null, HIS, A)	50000	NaN	50009
(10, GLY, A)	(null, GLY, A)	50000	NaN	50010
(11, SER, A)	(0, SER, A)	50000	NaN	50011
(12, ALA, A)	(1, ALA, A)	(54, A, A)	P30044	54
(13, PRO, A)	(2, PRO, A)	(55, P, A)	P30044	55
(14, ILE, A)	(3, ILE, A)	(56, I, A)	P30044	56
(15, LYS, A)	(4, LYS, A)	(57, K, A)	P30044	57
(16, VAL, A)	(5, VAL, A)	(58, V, A)	P30044	58
(17, GLY, A)	(6, GLY, A)	(59, G, A)	P30044	59
(18, ASP, A)	(7, ASP, A)	(60, D, A)	P30044	60
(19, ALA, A)	(8, ALA, A)	(61, A, A)	P30044	61
(20, ILE, A)	(9, ILE, A)	(62, I, A)	P30044	62
(21, PRO, A)	(10, PRO, A)	(63, P, A)	P30044	63
(22, ALA, A)	(11, ALA, A)	(64, A, A)	P30044	64

(B)

PDBe	PDB	UniProt	AccessionID	UniProt_50k
(81, PHE, D)	(100, PHE, D)	(82, F, D)	Q4PRK9	82
(82, THR, D)	(101, THR, D)	(83, T, D)	Q4PRK9	83
(83, LYS, D)	(102, LYS, D)	(84, K, D)	Q4PRK9	84
(84, ALA, D)	(103A, ALA, D)	(85, A, D)	Q4PRK9	85
(85, PRO, D)	(103B, PRO, D)	(86, P, D)	Q4PRK9	86
(86, GLY, D)	(103C, GLY, D)	(87, G, D)	Q4PRK9	87
(87, LYS, D)	(103D, LYS, D)	(88, K, D)	Q4PRK9	88
(88, SER, D)	(103E, SER, D)	(89, S, D)	Q4PRK9	89
(89, ASP, D)	(105A, ASP, D)	(90, D, D)	Q4PRK9	90
(90, LYS, D)	(105B, LYS, D)	(91, K, D)	Q4PRK9	91
(91, GLU, D)	(106, GLU, D)	(92, E, D)	Q4PRK9	92
(92, TRP, D)	(107, TRP, D)	(93, W, D)	Q4PRK9	93

(C)

PDBe	PDB	UniProt	AccessionID	UniProt_50k
(1, HIS, A)	(-2, HIS, A)	50000	NaN	50001
(2, HIS, A)	(-1, HIS, A)	50000	NaN	50002
(3, MET, A)	(1, MET, A)	(1, M, A)	P21856	1
(4, ASP, A)	(2, ASP, A)	(2, D, A)	P21856	2
(5, GLU, A)	(3, GLU, A)	(3, E, A)	P21856	3
(6, GLU, A)	(4, GLU, A)	(4, E, A)	P21856	4
(7, TYR, A)	(5, TYR, A)	(5, Y, A)	P21856	5
(8, ASP, A)	(6, ASP, A)	(6, D, A)	P21856	6
(9, VAL, A)	(7, VAL, A)	(7, V, A)	P21856	7
(10, ILE, A)	(8, ILE, A)	(8, I, A)	P21856	8
(11, VAL, A)	(9, VAL, A)	(9, V, A)	P21856	9

Figure 2. Small fragments of the Pandas dataframes assembled from SIFTS files: (A) 2vl3, (B) 2aa3, and (C) 1d5t. Entry 2vl3 contains a His tag that is not observed in the coordinates. Chain D of 2aa3 contains insertion codes (column 2) for some residues. Entry 1d5t contains a His tag with negative author residue numbers (column 2). The PDBe column in each image contains data for each amino acid in tuples (SeqResNum, ResName, and EntityId), where SeqResNum is the position of the amino acid in the sequence numbered from 1 to N (the length of the sequence). This field acts as the Pandas dataframe index for the whole table, since it is unique for each amino acid. The PDB column contains tuples (AuthResNum + InsCode, ResName, and ChainID). The UniProt column contain tuples of (UniProtResNum, UniProtResName, ChainID) and if there is no UniProt residue number, it contains the number "50,000". The next column contains the UniProt AccessionID. The column UniProt_50k provides the final numbering of residues in the PDBrenum output file: it is the UniProt number when it is available, and 50,000+SeqResNum when there is no UniProt for a chain that has a UniProt. Chains with no UniProt in SIFTS are not renumbered.

Methods

PDBrenum was written in Python with use of Python 3.6, BioPython 1.76 [16], Pandas 0.25.1 [17], and Numpy 1.17 modules within Jupyter-Notebook 6.0.1 [18] and PyCharm 2020.2 (https://www.jetbrains.com/pycharm/) as an integrated development environment on a Ubuntu 20.04 operating system.

Figure 1 represents a basic scheme of the PDBrenum workflow. First, PDBrenum downloads the structure files (in PDB or/and mmCIF format) and corresponding SIFTS files (in .xml format). The program downloads files in three attempts; if there is no success in three attempts, the assumption is that there is no such file (sometimes servers might not respond or respond with errors, but it is very unlikely to get three bad responses from the server in a row). PDBrenum then parses the SIFTS file to obtain numbering data for each amino acid in each protein chain in the file and places the results in a Pandas dataframe:

- ***PDBChainID:*** the `label_asym_id` in mmCIF coordinates (from `entityId` in SIFTS). Note: this does not correspond to `entity_id` in the mmCIF files, which instead is an integer that indicates the molecule identity (i.e., each protein sequence and each ligand type gets an entity_id).
- ***AuthChainID:*** the `auth_asym_id` in mmCIF coordinates (from `PDB:dbChainID` in SIFTS)
- ***SeqResNum***: the `label_seq_id` in mmCIF coordinates, which is the number of each residue in each protein construct when numbered from 1 to N, the number of residues in the protein chain (from `PDBe:dbResNum` in the SIFTS file)
- ***AuthResNum***: the `auth_seq_id` in mmCIF coordinates, which is the author residue number (from `PDB:dbResNum` in SIFTS)
- ***InsCode***: the `pdbx_PDB_ins_code` labels in mmCIF coordinates, which are insertion codes, if any attached to residue numbers in legacy PDB files to distinguish residues inserted in the sequence (from upper-case letters in `PDB:dbResNum` in SIFTS)
- ***ResName*:** the `label_comp_id` and `auth_comp_id` in mmCIF coordinates, which is the residue name in three letter code (from `PDB:dbResName` in SIFTS)
- ***AccessionID:*** for UniProt entry (if any) (from `UniProt:dbAccessionId` in SIFTS)
- ***UniProtResNum***: residue number in UniProt reference sequence (if any) (from `UniProt:dbResNum` in SIFTS)
- ***UniProtResName***: amino acid type in UniProt sequence (if any) in one letter code (from `UniProt:dbResName` in SIFTS)

The typical Pandas dataframe will look like the ones shown in Figure 2. The dataframe correlates different numbering systems for the amino acids in each protein chain. The unique residue numbering is the 1 to N numbering (SeqResNum) for each chain, where N is the chain length. This is referred to as `label_seq_i`d in the mmCIF file coordinate (`_atom_site`) records. The tuple representing (SeqResNum, ResName, and PDBChainID), denoted `label_seq_id`, `label_comp_id`, and `label_asym_id` in the mmCIF coordinates, is shown in the first column where the combination of the residue number and chain id act as a Pandas index or key for the table. The second numbering system is that used by the authors in the coordinates of the mmCIF file, which is represented in the “PDB” column. It consists of tuples (AuthResNum + InsCode, ResName, and AuthChainID). These values are denoted `auth_seq_id`, `pdbx_PDB_ins_code`, `auth_comp_id`, and `auth_asym_id` in the coordinate section of mmCIF files respectively. Insertion codes are letters attached to some residue numbers by authors to create new residue identifiers for inserted residues in a sequence. They are common in antibody numbering systems [19].

For most amino acids in the PDB, the SIFTS database has a UniProt reference and residue number, which is given in the 3rd column in each dataframe. When there is no UniProt number given in SIFTS for some residues in a chain (usually for sequence tags), we place the number 50,000 in this column. The resulting numbering system (given in the column labeled

“UniProt_50k”) that PDBrenum will use as a replacement for the author numbering system is the UniProt number where it is available and 50,000+SeqResNum when there is no UniProt number. This guarantees that there will be no collision between a UniProt residue number, and the numbers assigned to sequence tags and other insertions that are not part of the UniProt numbering system.

After reading and processing the SIFTS file for an entry, PDBrenum uncompresses and reads the gzipped mmCIF file as a Python dictionary using BioPython. The dictionary created by the BioPython function MMCIF2DICT forms keys from each mmCIF table and item name as a single string (e.g., `_atom_site.Cartn_x`). The corresponding value for each key is a Python list (e.g., the x-coordinates for all atoms in an entry).

In order to renumber PDB files according to UniProt from SIFTS, we need to identify corresponding values in all tables in the mmCIF files and all records in the PDB format files. SIFTS contains the PDBChainIDs, the author ChainIDs, the author residue numbers (with appended insert codes, if they exist), and the 1-to-N numbering of each chain. For each table (e.g. `_atom_site` or `_pdbx_validate_torsion`), we detect whether the table has residue numbers and chain identifiers that may be compared to the SIFTS data described above.

For the ChainIDs, tables may contain both the author ChainIDs and the PDB’s ChainIDs, but in some tables only the author ChainIDs exist (e.g., table `_struct_ref_seq`) and are labeled either as `auth_asym_id`, `pdb_strand_id` or `pdbx_strand_id`. These ChainIDs agree with the `auth_asym_id` in the coordinates. According to the mmCIF dictionary, all variants (with suffixes and prefixes (e.g., `struct_sheet_range.beg_auth_asym_id`) of `auth_asym_id`, `pdb_strand_id`, and `pdbx_strand_id`) correspond to the author ChainIDs in the coordinates and SIFTS, and thus can be used by PDBrenum to translate protein residue numbering into UniProt.

Many tables do *not* contain the 1-to-N numbering of each chain (SeqResNum in Figure 2), and so we need to interpret the values in the author numbering (with insert codes, if any) in each table in order to renumber according to UniProt from SIFTS. Author sequence numbering may be designated as `auth_seq_id`, and may be prefixed or suffixed with other identifiers, e.g., `auth_seq_id_1` or `beg_auth_seq_id`. We used the mmCIF dictionary to determine that all identifiers that contain `auth_seq_id` or its variants are children of `_atom_site.auth_seq_id` in the coordinate records, i.e. the author residue numbering in the coordinates that corresponds to the author numbering in SIFTS.

mmCIF files contain three tables that provide information on the sequence numbering of molecules in the structure: `_pdbx_poly_seq_scheme`, `_pdbx_nonpoly_seq_scheme`, and `_pdbx_branch_scheme`. These tables include four residue numbering schemes: `seq_id`, `pdb_seq_num`, `ndb_seq_num`, and `auth_seq_num`. For proteins and other polymers, `seq_id` in the `_pdbx_poly_seq_scheme` is the 1 to N numbering of the chain. These numbers are also found in the `ndb_seq_num` column. The `pdb_seq_num` column corresponds to the residue numbering in the coordinates, according to the mmCIF dictionary. The dictionary indicates that the `auth_seq_num` in the three tables may or may not correspond to the numbering in the coordinates. We found about 2000 files where these numbers differ from `pdb_seq_num`. These residue numbers appear to be those originally deposited by the authors and in these 2000 files, the “author” residue numbering has been altered by the PDB in the coordinates and other tables, and in the legacy PDB format files. This information is apparently kept in the file for reference. It is not used in any other table in any current PDB entry or in the legacy PDB format files. As it

turns out, there is only one instance of a similar prefixed identifier, pdbx_auth_seq_num (_struct_ref_seq_dif.pdbx_auth_seq_num), and it corresponds to pdb_seq_num, not auth_seq_num in the pdbx_poly_seq_scheme table, so it is renumbered by PDBrenum according to SIFTS.

Insert codes may be designated "ins_code", "PDB_ins_code", or "pdb_ins_code", and these names may contain prefixes and suffixes. According to the dictionary, all of these are children of _atom_site.pdbx_PDB_ins_code, and thus will agree with any insert codes present in SIFTS.

PDBrenum forms a Pandas dataframe for each mmCIF table (e.g. _atom_site) with index names equal to the Python tuple consisting of (AuthResNum + InsCode, ChainID), and merges it with the same combination (labeled "PDB" in Figure 2) from the SIFTS dataframe. This merged table is then used to replace the AuthResNum values with the UniProtResNum or 50,000+SeqResNum values. Non-polymeric molecule types such as small ligands are also renumbered as 60,000+their residue number (_pdbx_nonpoly_seq_scheme.pdb_seq_num) in the mmCIF file. With a tuple consisting of the author chainID the author residue number, and the insert code (if any), the following items in mmCIF files are renumbered:

```
_atom_site_anisotrop.pdbx_auth_seq_id
_atom_site.auth_seq_id
_pdbx_distant_solvent_atoms.auth_seq_id
_pdbx_refine_tls_group.beg_auth_seq_id
_pdbx_refine_tls_group.end_auth_seq_id
_pdbx_struct_chem_comp_diagnostics.auth_seq_id
_pdbx_struct_conn_angle.ptnr1_auth_seq_id
_pdbx_struct_conn_angle.ptnr2_auth_seq_id
_pdbx_struct_conn_angle.ptnr3_auth_seq_id
_pdbx_struct_mod_residue.auth_seq_id
_pdbx_struct_sheet_hbond.range_1_auth_seq_id
_pdbx_struct_sheet_hbond.range_2_auth_seq_id
_pdbx_struct_special_symmetry.auth_seq_id
_pdbx_unobs_or_zero_occ_atoms.auth_seq_id
_pdbx_unobs_or_zero_occ_residues.auth_seq_id
_pdbx_validate_chiral.auth_seq_id
_pdbx_validate_close_contact.auth_seq_id_1
_pdbx_validate_close_contact.auth_seq_id_2
_pdbx_validate_main_chain_plane.auth_seq_id
_pdbx_validate_peptide_omega.auth_seq_id_1
_pdbx_validate_peptide_omega.auth_seq_id_2
_pdbx_validate_planes.auth_seq_id
_pdbx_validate_polymer_linkage.auth_seq_id_1
_pdbx_validate_polymer_linkage.auth_seq_id_2
_pdbx_validate_rmsd_angle.auth_seq_id_1
_pdbx_validate_rmsd_angle.auth_seq_id_2
_pdbx_validate_rmsd_angle.auth_seq_id_3
_pdbx_validate_rmsd_bond.auth_seq_id_1
_pdbx_validate_rmsd_bond.auth_seq_id_2
_pdbx_validate_symm_contact.auth_seq_id_1
_pdbx_validate_symm_contact.auth_seq_id_2
_pdbx_validate_torsion.auth_seq_id
_struct_conf.beg_auth_seq_id
_struct_conf.end_auth_seq_id
_struct_conn.ptnr1_auth_seq_id
_struct_conn.ptnr2_auth_seq_id
_struct_mon_prot_cis.auth_seq_id
_struct_mon_prot_cis.pdbx_auth_seq_id_2
_struct_ncs_dom_lim.beg_auth_seq_id
_struct_ncs_dom_lim.end_auth_seq_id
_struct_sheet_range.beg_auth_seq_id
_struct_sheet_range.end_auth_seq_id
_struct_site_gen.auth_seq_id
_struct_site.pdbx_auth_seq_id

_pdbx_nonpoly_scheme.pdb_seq_num
_pdbx_poly_seq_scheme.pdb_seq_num
_pdbx_branch_scheme.pdb_seq_num
_struct_ref_seq_dif.pdbx_auth_seq_num

_struct_ref_seq.pdbx_auth_seq_align_beg
_struct_ref_seq.pdbx_auth_seq_align_end
```

As noted above, the pdbx_poly_seq_scheme, pdbx_nonpoly_seq_scheme, and the pdbx_branch_scheme (for branched sugars) contain the 1-to-N numbering (called seq_id), the author residue numbering corresponding to the coordinates (pdb_seq_num), and an extra residue number (auth_seq_num) that may differ from pdb_seq_num, containing historical data from the original file deposition. When it differs from pdb_seq_num, it is not used elsewhere in the mmCIF files. We renumber pdb_seq_num according to UniProt, and replace auth_seq_num in this table with the values of pdb_seq_num in the PDB-issued mmCIF file. That way, our files have a table that provides a correspondence between the 1-to-N numbering, the UniProt numbering, and the residue numbering of the original mmCIF file obtained from the PDB (Figure 3).

```
#
loop_
_pdbx_poly_seq_scheme.asym_id
_pdbx_poly_seq_scheme.entity_id
_pdbx_poly_seq_scheme.seq_id
_pdbx_poly_seq_scheme.mon_id
_pdbx_poly_seq_scheme.ndb_seq_num
_pdbx_poly_seq_scheme.pdb_seq_num
_pdbx_poly_seq_scheme.auth_seq_num
_pdbx_poly_seq_scheme.pdb_mon_id
_pdbx_poly_seq_scheme.auth_mon_id
_pdbx_poly_seq_scheme.pdb_strand_id
_pdbx_poly_seq_scheme.pdb_ins_code
_pdbx_poly_seq_scheme.hetero
A 1 1   THR 1   18  18  THR THR A . n
A 1 2   PRO 2   19  19  PRO PRO A . n
A 1 3   LYS 3   20  20  LYS LYS A . n
A 1 4   PRO 4   21  21  PRO PRO A . n
A 1 5   LYS 5   22  22  LYS LYS A . n
A 1 6   ILE 6   23  23  ILE ILE A . n
A 1 7   VAL 7   24  24  VAL VAL A . n
A 1 8   LEU 8   25  25  LEU LEU A . n
A 1 9   VAL 9   26  26  VAL VAL A . n
A 1 10  GLY 10  27  27  GLY GLY A . n
A 1 11  SER 11  28  28  SER SER A . n
A 1 12  GLY 12  29  29  GLY GLY A . n
A 1 13  MET 13  30  30  MET MET A . n
A 1 14  ILE 14  31  31  ILE ILE A . n
A 1 15  GLY 15  32  32  GLY GLY A . n
A 1 16  GLY 16  33  33  GLY GLY A . n
A 1 17  VAL 17  35  35  VAL VAL A . n
```

```
#
loop_
_pdbx_poly_seq_scheme.asym_id
_pdbx_poly_seq_scheme.entity_id
_pdbx_poly_seq_scheme.seq_id
_pdbx_poly_seq_scheme.mon_id
_pdbx_poly_seq_scheme.ndb_seq_num
_pdbx_poly_seq_scheme.pdb_seq_num
_pdbx_poly_seq_scheme.auth_seq_num
_pdbx_poly_seq_scheme.pdb_mon_id
_pdbx_poly_seq_scheme.auth_mon_id
_pdbx_poly_seq_scheme.pdb_strand_id
_pdbx_poly_seq_scheme.pdb_ins_code
_pdbx_poly_seq_scheme.hetero
A 1 1   THR 1   2     18  THR THR A . n
A 1 2   PRO 2   3     19  PRO PRO A . n
A 1 3   LYS 3   4     20  LYS LYS A . n
A 1 4   PRO 4   5     21  PRO PRO A . n
A 1 5   LYS 5   6     22  LYS LYS A . n
A 1 6   ILE 6   7     23  ILE ILE A . n
A 1 7   VAL 7   8     24  VAL VAL A . n
A 1 8   LEU 8   9     25  LEU LEU A . n
A 1 9   VAL 9   10    26  VAL VAL A . n
A 1 10  GLY 10  11    27  GLY GLY A . n
A 1 11  SER 11  12    28  SER SER A . n
A 1 12  GLY 12  13    29  GLY GLY A . n
A 1 13  MET 13  14    30  MET MET A . n
A 1 14  ILE 14  15    31  ILE ILE A . n
A 1 15  GLY 15  16    32  GLY GLY A . n
A 1 16  GLY 16  17    33  GLY GLY A . n
A 1 17  VAL 17  18    35  VAL VAL A . n
```

Figure 3. Renumbering of the pdbx_poly_seq_scheme table from 2aa3 processed by PDBrenum. Left: the original file from the PDB. Right: the renumbered file from PDBrenum. The original author numbering is given in column 6 of the table on the left (18, 19, 20, etc.), which is replaced with the UniProt numbering (entry Q9PRK9) for this chain (2,3,4, etc) in column 6 of the table on the right. The original author numbering has been placed in column 7 of the table on the right (i.e., in the auth_seq_num position).

For the space-delimited legacy PDB format files, if the line starts with ("ATOM"), ("TER"), ("ANISOU") or ("SIGUIJ") the program gets columns 22:26, 27, 17:20, 21 as residue number (AuthResNum), insertion code (InsCode), residue name (AuthResName), and ChainID respectively. For special lines “REMARK 465” records which list missing residues, columns 20:26, 27, 15:18, 19 are obtained as the residue number, insertion code, residue name, and ChainID correspondingly. The two data frames are merged and if the residue does not have a UniProt residue number then it gets default_PDB_num (default_PDB_num = 5000 + SeqResNum). In order to prevent possible numbering collisions, PDBrenum calculates available numbers in the range from 1 to 9999 and reassigns them to non-polymeric compounds in reverse order. After PDBrenum makes the replacement dictionary out of two python strings where the keys are AuthResNum (4 Char) + InsCode (1 Char) + ResName (3 Char) + ChainID (1 Char) and the values are UniProtResNum + InsCode + ResName + ChainID, where the values have the same number of characters. Finally, PDBrenum processes each line of the PDB file, replacing keys with values.

The value offset for non-UniProt residues, default_PDB_num (5,000), can be reset (with “--set_default_mmCIF_num” flag) and default_mmCIF_num (50,000) (with “--set_default_PDB_num” flag) as you wish but we recommend it to be big enough so it will not be the same as any other numbers but not bigger then 9000 for PDB format because it might go over the 4-character limit of 9999).

There are some chains in the PDB that are chimeras containing sequence from two or more UniProt entries. In cases where there is no collision of residue numbering, then the sequences are numbered according to the UniProt sequences in the SIFTS file. In cases when there is a

collision, the longest sequence is taken as the default. The only exception to this is for a small number of UniProt sequences that are commonly used as crystallization chaperones, and are therefore not the target of main interest. These include UniProt codes GFP_AEQVI, GCN4_YEAST, C562_ECOLX, ENLYS_BPT4, MALE_ECOLI.

All the files that were processed by PDBrenum get the name tag "_renum" (e.g. 2aa3_renum.cif.gz) and we insert REMARKs in them.

REMARK for mmCIF files:

```
loop_
_database_PDB_remark.id       1
_database_PDB_remark.text
;File processed by PDBrenum: http://dunbrack3.fccc.edu/PDBrenum
Author sequence numbering is replaced with UniProt numbering according to
alignment by SIFTS (https://www.ebi.ac.uk/pdbe/docs/sifts/).
Only chains with UniProt sequences in SIFTS are renumbered.
Residues in UniProt chains without UniProt residue numbers in SIFTS
(e.g., sequence tags) are given residue numbers 50000+label_seq_id
(where label_seq_id is the 1-to-N residue numbering of each chain.
Ligands are numbered 50000+their residue number in the original file.
The _poly_seq_scheme table contains a correspondence between the
1-to-N sequence (seq_id), the new numbering based on UniProt (pdb_seq_num =
auth_seq_id in the _atom_site records), and the author numbering
in the original mmCIF file from the PDB (auth_seq_num).
;
#
```

REMARK for legacy PDB-format files:

```
REMARK   0  File processed by PDBrenum: http://dunbrack3.fccc.edu/PDBrenum
REMARK   0  Author sequence numbering is replaced with UniProt numbering
REMARK   0  according to alignment by SIFTS
REMARK   0  (https://www.ebi.ac.uk/pdbe/docs/sifts/).
REMARK   0  Only chains with UniProt sequences in SIFTS are renumbered.
REMARK   0  Residues in UniProt chains without UniProt residue numbers in SIFTS
REMARK   0  (e.g., sequence tags) are given residue numbers 5000+label_seq_id
REMARK   0  (where label_seq_id is the 1-to-N residue numbering of each chain.
REMARK   0  Ligands are numbered 5000+their residue number in the original
REMARK   0  file. The _poly_seq_scheme table contains a correspondence between
REMARK   0  the 1-to-N sequence (seq_id), the new numbering based on UniProt
REMARK   0  (pdb_seq_num = auth_seq_id in the _atom_site records), and
REMARK   0  the author numbering in the original mmCIF file from the PDB
REMARK   0  (auth_seq_num).
```

Setting up PDBrenum

As a prerequisites, anaconda should be installed https://docs.anaconda.com/anaconda/install/. The following will set up a conda environment for running PDBrenum locally:

```
(base) $ git clone https://github.com/Faezov/PDBrenum.git
(base) $ cd PDBrenum
(base) $ conda create -n PDBrenum python=3.6 numpy=1.17 pandas=0.25.1 biopython=1.76
tqdm=4.36.1 ipython=7.8.0 requests=2.25.1 lxml=4.6.2
(base) $ conda activate PDBrenum
```

***Running PDBrenum**:*

Help can be obtained with this command:

```
(PDBrenum) $ python3 PDBrenum.py -h
```

The user can provide PDBids directly as a list of arguments (-rfla --renumber_from_list_of_arguments):

```
(PDBrenum) $ python3 PDBrenum.py -rfla 1d5t 1bxw 2vl3 5e6h -mmCIF
(PDBrenum) $ python3 PDBrenum.py -rfla 1d5t 1bxw 2vl3 5e6h -PDB
(PDBrenum) $ python3 PDBrenum.py -rfla 1d5t 1bxw 2vl3 5e6h -mmCIF_assembly
(PDBrenum) $ python3 PDBrenum.py -rfla 1d5t 1bxw 2vl3 5e6h -PDB_assembly
```

or put PDBids in a text file (comma, space, tab, or newline delimited) (-rftf --renumber_from_text_file):

```
(PDBrenum) $ python3 PDBrenum.py -rftf input.txt -mmCIF
(PDBrenum) $ python3 PDBrenum.py -rftf input.txt -PDB
(PDBrenum) $ python3 PDBrenum.py -rftf input.txt -mmCIF_assembly
(PDBrenum) $ python3 PDBrenum.py -rftf input.txt -PDB_assembly
```

The user can renumber the entire PDB in a given format (by default in mmCIF if no format is provided):

```
(PDBrenum) $ python3 PDBrenum.py -redb -mmCIF
(PDBrenum) $ python3 PDBrenum.py -redb -PDB
(PDBrenum) $ python3 PDBrenum.py -redb -mmCIF_assembly
(PDBrenum) $ python3 PDBrenum.py -redb -PDB_assembly
```

Note that sometimes on Windows installations, the biopython module might be installed incorrectly by conda and it will cause a module error in python. To resolve this problem simply run:

```
(PDBrenum) $ pip install biopython==1.76
```

PDBrenum was heavily tested on all PDB structure files in both formats and on all popular operating systems (Linux, Mac and Windows).

PDBrenum uses multiprocessing by default using all available cores, but the user can set a limit to the number of processors by providing number to `-nproc` flag.

The user can also change where input output files will go by using these self-explanatory flags (with absolute paths):

```
"-sipm", "--set_default_input_path_to_mmCIF"
"-sipma", "--set_default_input_path_to_mmCIF_assembly"
"-sipp", "--set_default_input_path_to_PDB"
"-sippa", "--set_default_input_path_to_PDB_assembly"
"-sips", "--set_default_input_path_to_SIFTS"
"-sopm", "--set_default_output_path_to_mmCIF"
"-sopma", "--set_default_output_path_to_mmCIF_assembly"
"-sopp", "--set_default_output_path_to_PDB"
"-soppa", "--set_default_output_path_to_PDB_assembly"
```

By default, files go here:

```
default_input_path_to_mmCIF = current_directory + "/mmCIF"
default_input_path_to_mmCIF_assembly = current_directory + "/mmCIF_assembly"
default_input_path_to_PDB = current_directory + "/PDB"
default_input_path_to_PDB_assembly = current_directory + "/PDB_assembly"
default_input_path_to_SIFTS = current_directory + "/SIFTS"
default_output_path_to_mmCIF = current_directory + "/output_mmCIF"
default_output_path_to_mmCIF_assembly = current_directory + "/output_mmCIF_assembly"
default_output_path_to_PDB = current_directory + "/output_PDB"
default_output_path_to_PDB_assembly = current_directory + "/output_PDB_assembly"
```

Also, by default all files gzipped if you want have them unzipped please use `"-offz"` or `"--set_to_off_mode_gzip"`

1	SP	PDB_id	chain_PDB	chain_auth	UniProt	SwissProt	uni_len	chain_len	renum	5k_or_50k
2	+	2aa3	A	A	Q4PRK9	Q4PRK9_PLAVI	315	316	315	1
3	+	2aa3	B	B	Q4PRK9	Q4PRK9_PLAVI	315	315	315	0
4	+	2aa3	C	C	Q4PRK9	Q4PRK9_PLAVI	315	318	315	3
5	+	2aa3	D	D	Q4PRK9	Q4PRK9_PLAVI	315	315	315	0

Figure 4. Log file of PDBrenum on PDB entry 2aa3. SP is special case it is either "+" for cases when there is no clash in UniProt numbers or "*" for the case when biggest UniProt was taken unless it is in the exception list [GFP_AEQVI, GCN4_YEAST, C562_ECOLX, ENLYS_BPT4, MALE_ECOLI]. PDB_id is 4-character long PDB identifier, chain_PDB is the chain identifier given by the PDB in mmCIF files (label_asym_id), chain_auth is a chain identifier given by author of the structure (auth_asym_id), comp_uni is 6-character long UniProt identifier usually starts with "P", human_uni is the human readable UniProt (SwissProt) identifier, prot_len, uni_len, and chain_len represent total quantity of residues in a given protein, UniProt, chain, renum represents total quantity of residues that were renumbered, 5k_or_50k represents quantity of residues that were renumbered by adding 5000 to 1-to-N numbering to for PDB-legacy format or 50000 to 1-to-N numbering to for mmCIF format.

Result and Discussion

PDBrenum returns a log file and renumbered structure files, shown in Figures 4 and 5 respectively for PDB entry 2aa3. In Fig 5, green arrows point to the columns which were changed.

We have created a webserver (http://dunbrack3.fccc.edu/PDBrenum) that will take a list of PDB entry codes or a list of UniProt identifiers (as Accession IDs or SwissProt IDs) and with one click of the mouse, will return a zip file with the requested mmCIF or legacy-PDB format files of the asymmetric units and/or biological assemblies (Figure 6). The output files can also be accessed programmatically via direct http links:

http://dunbrack3.fccc.edu/PDBrenum/output_PDB/2aa3_renum.pdb.gz
http://dunbrack3.fccc.edu/PDBrenum/output_mmCIF/2aa3_renum.cif.gz
http://dunbrack3.fccc.edu/PDBrenum/output_PDB_assembly/2aa3_renum.pdb1.gz
http://dunbrack3.fccc.edu/PDBrenum/output_mmCIF_assembly/2aa3-assembly-1_renum.cif.gz

On April 26, 2020, the PDB had 176,507 structures. When processed with PDBrenum:

1) 3,659 structures (2.1%) do not have SIFTS (mostly DNA or RNA-only files)
2) 5,919 structures (3.4%) have SIFTS files but do not have any UniProt data in SIFTS (mostly

antibodies)

3) 75,359 structures (42.7%) were not changed because all of the proteins have UniProt numbering
4) 23,448 structures (13.3%) were changed due only to the presence of sequence tags or other residues with no UniProt numbers
5) 49,422 structures (28.0%) were changed due to differences between UniProt and author residue numbering only
6) 18,700 structures (10.6%) were changed due to both UniProt/author numbering differences and sequence tags or other non-UniProt residues

The large percentage of files (38.6%) that contain proteins that are not numbered according to a standard for each protein (UniProt) demonstrates the need for the approach enabled by PDBrenum.

(A)

```
loop_
_atom_site.group_PDB
_atom_site.id
_atom_site.type_symbol
_atom_site.label_atom_id
_atom_site.label_alt_id
_atom_site.label_comp_id
_atom_site.label_asym_id
_atom_site.label_entity_id
_atom_site.label_seq_id
_atom_site.pdbx_PDB_ins_code
_atom_site.Cartn_x
_atom_site.Cartn_y
_atom_site.Cartn_z
_atom_site.occupancy
_atom_site.B_iso_or_equiv
_atom_site.pdbx_formal_charge
_atom_site.auth_seq_id
_atom_site.auth_comp_id
_atom_site.auth_asym_id
_atom_site.auth_atom_id
_atom_site.pdbx_PDB_model_num
ATOM   1    N N   . THR A 1 1   ? 21.071  -3.780  0.959   0.50 33.08 ? 18  THR A N   1
ATOM   2    C CA  . THR A 1 1   ? 20.920  -4.893  -0.028  0.50 33.12 ? 18  THR A CA  1
ATOM   3    C C   . THR A 1 1   ? 20.260  -4.505  -1.390  0.50 32.86 ? 18  THR A C   1
ATOM   4    O O   . THR A 1 1   ? 19.219  -5.071  -1.781  0.50 33.29 ? 18  THR A O   1
ATOM   5    C CB  . THR A 1 1   ? 20.217  -6.140  0.640   0.50 33.37 ? 18  THR A CB  1
ATOM   6    O OG1 . THR A 1 1   ? 20.264  -7.284  -0.235  0.50 34.48 ? 18  THR A OG1 1
ATOM   7    C CG2 . THR A 1 1   ? 18.758  -5.827  1.056   0.50 32.86 ? 18  THR A CG2 1
ATOM   8    N N   . PRO A 1 2   ? 20.845  -3.532  -2.116  0.50 31.64 ? 19  PRO A N   1
ATOM   9    C CA  . PRO A 1 2   ? 21.882  -2.598  -1.716  0.50 30.98 ? 19  PRO A CA  1
ATOM   10   C C   . PRO A 1 2   ? 21.307  -1.477  -0.840  0.50 29.29 ? 19  PRO A C   1
ATOM   11   O O   . PRO A 1 2   ? 20.113  -1.468  -0.510  0.50 29.80 ? 19  PRO A O   1
ATOM   12   C CB  . PRO A 1 2   ? 22.357  -2.013  -3.052  0.50 30.71 ? 19  PRO A CB  1
ATOM   13   C CG  . PRO A 1 2   ? 21.162  -2.030  -3.898  0.50 31.48 ? 19  PRO A CG  1
ATOM   14   C CD  . PRO A 1 2   ? 20.450  -3.321  -3.522  0.50 32.18 ? 19  PRO A CD  1
ATOM   15   N N   . LYS A 1 3   ? 22.165  -0.537  -0.480  1.00 27.74 ? 20  LYS A N   1
ATOM   16   C CA  . LYS A 1 3   ? 21.851  0.470   0.566   1.00 24.76 ? 20  LYS A CA  1
ATOM   17   C C   . LYS A 1 3   ? 20.859  1.474   -0.070  1.00 21.20 ? 20  LYS A C   1
ATOM   18   O O   . LYS A 1 3   ? 21.118  1.943   -1.164  1.00 20.53 ? 20  LYS A O   1
ATOM   19   C CB  . LYS A 1 3   ? 23.173  1.165   0.968   1.00 25.30 ? 20  LYS A CB  1
ATOM   20   C CG  . LYS A 1 3   ? 23.163  2.034   2.252   1.00 25.01 ? 20  LYS A CG  1
ATOM   21   C CD  . LYS A 1 3   ? 23.196  1.149   3.512   1.00 29.51 ? 20  LYS A CD  1
ATOM   22   C CE  . LYS A 1 3   ? 22.595  1.857   4.796   1.00 29.55 ? 20  LYS A CE  1
ATOM   23   N NZ  . LYS A 1 3   ? 22.037  0.855   5.799   1.00 28.73 ? 20  LYS A NZ  1
```

```
loop_
_atom_site.group_PDB
_atom_site.id
_atom_site.type_symbol
_atom_site.label_atom_id
_atom_site.label_alt_id
_atom_site.label_comp_id
_atom_site.label_asym_id
_atom_site.label_entity_id
_atom_site.label_seq_id
_atom_site.pdbx_PDB_ins_code
_atom_site.Cartn_x
_atom_site.Cartn_y
_atom_site.Cartn_z
_atom_site.occupancy
_atom_site.B_iso_or_equiv
_atom_site.pdbx_formal_charge
_atom_site.auth_seq_id
_atom_site.auth_comp_id
_atom_site.auth_asym_id
_atom_site.auth_atom_id
_atom_site.pdbx_PDB_model_num
ATOM   1    N N   . THR A 1 1   ? 21.071  -3.780  0.959   0.50 33.08 ? 2    THR A N   1
ATOM   2    C CA  . THR A 1 1   ? 20.920  -4.893  -0.028  0.50 33.12 ? 2    THR A CA  1
ATOM   3    C C   . THR A 1 1   ? 20.260  -4.505  -1.390  0.50 32.86 ? 2    THR A C   1
ATOM   4    O O   . THR A 1 1   ? 19.219  -5.071  -1.781  0.50 33.29 ? 2    THR A O   1
ATOM   5    C CB  . THR A 1 1   ? 20.217  -6.140  0.640   0.50 33.37 ? 2    THR A CB  1
ATOM   6    O OG1 . THR A 1 1   ? 20.264  -7.284  -0.235  0.50 34.48 ? 2    THR A OG1 1
ATOM   7    C CG2 . THR A 1 1   ? 18.758  -5.827  1.056   0.50 32.86 ? 2    THR A CG2 1
ATOM   8    N N   . PRO A 1 2   ? 20.845  -3.532  -2.116  0.50 31.64 ? 3    PRO A N   1
ATOM   9    C CA  . PRO A 1 2   ? 21.882  -2.598  -1.716  0.50 30.98 ? 3    PRO A CA  1
ATOM   10   C C   . PRO A 1 2   ? 21.307  -1.477  -0.840  0.50 29.29 ? 3    PRO A C   1
ATOM   11   O O   . PRO A 1 2   ? 20.113  -1.468  -0.510  0.50 29.80 ? 3    PRO A O   1
ATOM   12   C CB  . PRO A 1 2   ? 22.357  -2.013  -3.052  0.50 30.71 ? 3    PRO A CB  1
ATOM   13   C CG  . PRO A 1 2   ? 21.162  -2.030  -3.898  0.50 31.48 ? 3    PRO A CG  1
ATOM   14   C CD  . PRO A 1 2   ? 20.450  -3.321  -3.522  0.50 32.18 ? 3    PRO A CD  1
ATOM   15   N N   . LYS A 1 3   ? 22.165  -0.537  -0.480  1.00 27.74 ? 4    LYS A N   1
ATOM   16   C CA  . LYS A 1 3   ? 21.851  0.470   0.566   1.00 24.76 ? 4    LYS A CA  1
ATOM   17   C C   . LYS A 1 3   ? 20.859  1.474   -0.070  1.00 21.28 ? 4    LYS A C   1
ATOM   18   O O   . LYS A 1 3   ? 21.118  1.943   -1.164  1.00 20.53 ? 4    LYS A O   1
ATOM   19   C CB  . LYS A 1 3   ? 23.173  1.165   0.968   1.00 25.30 ? 4    LYS A CB  1
ATOM   20   C CG  . LYS A 1 3   ? 23.163  2.034   2.252   1.00 25.01 ? 4    LYS A CG  1
ATOM   21   C CD  . LYS A 1 3   ? 23.196  1.149   3.512   1.00 29.51 ? 4    LYS A CD  1
ATOM   22   C CE  . LYS A 1 3   ? 22.595  1.857   4.796   1.00 29.55 ? 4    LYS A CE  1
ATOM   23   N NZ  . LYS A 1 3   ? 22.037  0.855   5.799   1.00 28.73 ? 4    LYS A NZ  1
```

(B)

```
loop_
_struct_conf.conf_type_id
_struct_conf.id
_struct_conf.pdbx_PDB_helix_id
_struct_conf.beg_label_comp_id
_struct_conf.beg_label_asym_id
_struct_conf.beg_label_seq_id
_struct_conf.pdbx_beg_PDB_ins_code
_struct_conf.end_label_comp_id
_struct_conf.end_label_asym_id
_struct_conf.end_label_seq_id
_struct_conf.pdbx_end_PDB_ins_code
_struct_conf.beg_auth_comp_id
_struct_conf.beg_auth_asym_id
_struct_conf.beg_auth_seq_id
_struct_conf.end_auth_comp_id
_struct_conf.end_auth_asym_id
_struct_conf.end_auth_seq_id
_struct_conf.pdbx_PDB_helix_class
_struct_conf.details
_struct_conf.pdbx_PDB_helix_length
HELX_P HELX_P1  1  GLY A 12  ? LYS A 25  ? GLY A 29  LYS A 43  1 ? 14
HELX_P HELX_P2  2  ASN A 38  ? TYR A 55  B ASN A 57  TYR A 73  1 ? 18
HELX_P HELX_P3  3  SER A 65  ? LYS A 70  ? SER A 84  LYS A 89  5 ? 6
HELX_P HELX_P4  4  ASP A 96  ? CYS A 116 ? ASP A 111 CYS A 131 1 ? 21
HELX_P HELX_P5  5  PRO A 127 ? GLY A 140 ? PRO A 141 GLY A 154 1 ? 14
HELX_P HELX_P6  6  PRO A 142 ? ASN A 144 ? PRO A 156 ASN A 158 5 ? 3
HELX_P HELX_P7  7  GLY A 150 ? ASN A 167 ? GLY A 164 ASN A 181 1 ? 18
HELX_P HELX_P8  8  CYS A 169 ? ARG A 171 ? CYS A 183 ARG A 185 5 ? 3
HELX_P HELX_P9  9  LYS A 189 ? TYR A 191 ? LYS A 203 TYR A 205 5 ? 3
HELX_P HELX_P10 10 LEU A 199 A ASN A 205 ? LEU A 210 ASN A 215 1 ? 7
HELX_P HELX_P11 11 THR A 209 ? ASN A 222 ? THR A 220 ASN A 234 1 ? 14
HELX_P HELX_P12 12 ASN A 222 ? LEU A 231 ? ASN A 234 LEU A 243 1 ? 10
HELX_P HELX_P13 13 TYR A 235 ? LYS A 251 ? TYR A 247 LYS A 263 1 ? 17
HELX_P HELX_P14 14 GLN A 266 ? GLY A 268 ? GLN A 278 GLY A 280 5 ? 3
HELX_P HELX_P15 15 ASN A 294 ? ALA A 313 ? ASN A 300 ALA A 327 1 ? 20
HELX_P HELX_P16 16 GLY B 12  ? LYS B 25  ? GLY B 29  LYS B 43  1 ? 14
HELX_P HELX_P17 17 ASN B 38  ? TYR B 55  B ASN B 57  TYR B 73  1 ? 18
HELX_P HELX_P18 18 SER B 65  ? LYS B 70  ? SER B 84  LYS B 89  5 ? 6
HELX_P HELX_P19 19 ASN B 93  ? ASP B 95  ? ASN B 100 ASP B 110 5 ? 3
HELX_P HELX_P20 20 ASP B 96  ? CYS B 116 ? ASP B 111 CYS B 131 1 ? 21
```

```
loop_
_struct_conf.conf_type_id
_struct_conf.id
_struct_conf.pdbx_PDB_helix_id
_struct_conf.beg_label_comp_id
_struct_conf.beg_label_asym_id
_struct_conf.beg_label_seq_id
_struct_conf.pdbx_beg_PDB_ins_code
_struct_conf.end_label_comp_id
_struct_conf.end_label_asym_id
_struct_conf.end_label_seq_id
_struct_conf.pdbx_end_PDB_ins_code
_struct_conf.beg_auth_comp_id
_struct_conf.beg_auth_asym_id
_struct_conf.beg_auth_seq_id
_struct_conf.end_auth_comp_id
_struct_conf.end_auth_asym_id
_struct_conf.end_auth_seq_id
_struct_conf.pdbx_PDB_helix_class
_struct_conf.details
_struct_conf.pdbx_PDB_helix_length
HELX_P HELX_P1  1  GLY A 12  ? LYS A 25  ? GLY A 13  LYS A 26  1 ? 14
HELX_P HELX_P2  2  ASN A 38  ? TYR A 55  B ASN A 39  TYR A 56  1 ? 18
HELX_P HELX_P3  3  SER A 65  ? LYS A 70  ? SER A 66  LYS A 71  5 ? 6
HELX_P HELX_P4  4  ASP A 96  ? CYS A 116 ? ASP A 97  CYS A 117 1 ? 21
HELX_P HELX_P5  5  PRO A 127 ? GLY A 140 ? PRO A 128 GLY A 141 1 ? 14
HELX_P HELX_P6  6  PRO A 142 ? ASN A 144 ? PRO A 143 ASN A 145 5 ? 3
HELX_P HELX_P7  7  GLY A 150 ? ASN A 167 ? GLY A 151 ASN A 168 1 ? 18
HELX_P HELX_P8  8  CYS A 169 ? ARG A 171 ? CYS A 170 ARG A 172 5 ? 3
HELX_P HELX_P9  9  LYS A 189 ? TYR A 191 ? LYS A 190 TYR A 192 5 ? 3
HELX_P HELX_P10 10 LEU A 199 A ASN A 205 ? LEU A 200 ASN A 206 1 ? 7
HELX_P HELX_P11 11 THR A 209 ? ASN A 222 ? THR A 210 ASN A 223 1 ? 14
HELX_P HELX_P12 12 ASN A 222 ? LEU A 231 ? ASN A 223 LEU A 232 1 ? 10
HELX_P HELX_P13 13 TYR A 235 ? LYS A 251 ? TYR A 236 LYS A 252 1 ? 17
HELX_P HELX_P14 14 GLN A 266 ? GLY A 268 ? GLN A 267 GLY A 269 5 ? 3
HELX_P HELX_P15 15 ASN A 294 ? ALA A 313 ? ASN A 295 ALA A 314 1 ? 20
HELX_P HELX_P16 16 GLY B 12  ? LYS B 25  ? GLY B 13  LYS B 26  1 ? 14
HELX_P HELX_P17 17 ASN B 38  ? TYR B 55  B ASN B 39  TYR B 56  1 ? 18
HELX_P HELX_P18 18 SER B 65  ? LYS B 70  ? SER B 66  LYS B 71  5 ? 6
HELX_P HELX_P19 19 ASN B 93  ? ASP B 95  ? ASN B 94  ASP B 96  5 ? 3
HELX_P HELX_P20 20 ASP B 96  ? CYS B 116 ? ASP B 97  CYS B 117 1 ? 21
```

Figure 5. Renumbering PDB entry 2aa3. Screenshot of files 2aa3 before (left) and after (right) PDBrenum run "_atom_site" (coordinate section) (A) and "struct_conf" (B), green arrows pointed to the columns which were changed.

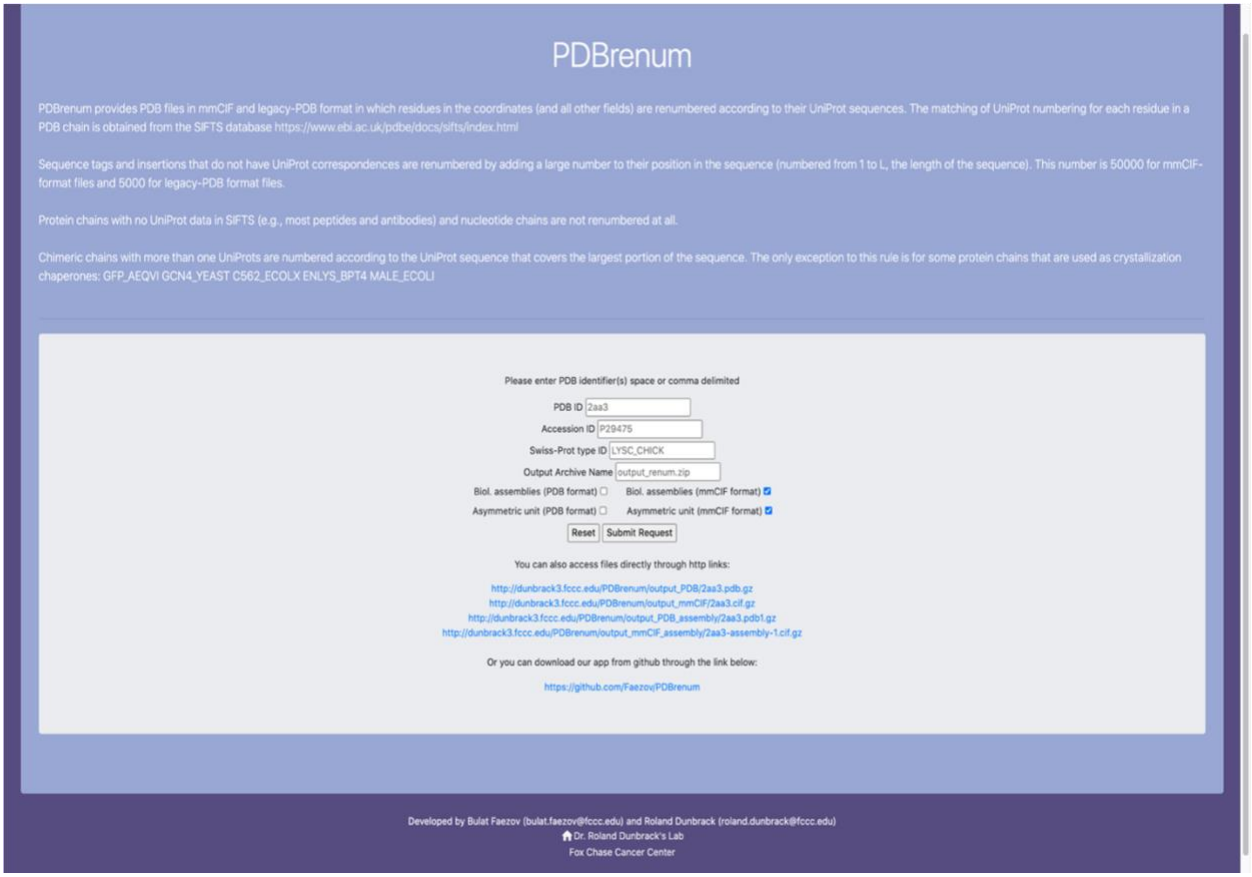

Figure 6. Screenshot of the PDBrenum server. The server takes in a list of PDB entry codes (comma, space, tab, or newline separated) or a list of UniProt (e.g. P38398) or SwissProt (e.g., BRCA1_HUMAN) accession codes. The user can choose whether to obtain mmCIF and/or PDB-format files, and whether to obtain asymmetric units and/or biological assemblies with check boxes. If more than one file is requested, a zip file is returned, and the name of this file can be specified.

We feel it is very important that PDBrenum provides renumbered files for both mmCIF and legacy-PDB format. mmCIF is the current standard for PDB files, and provides significantly more information than the original PDB format developed in the 1970s. However, many programs still exist that take only the legacy PDB format, and so it is still worthwhile to provide these files. Finally, we also feel it is important to provide the biological assembly files. More than half of crystal structures in the PDB have annotated assemblies that are different from the asymmetric unit [20]. While RCSB does not provide these files in mmCIF format at this time, they are available from PDBe. Even when the assembly consists of multiple copies of the asymmetric unit, every chain in the assembly files from PDBe has its own unique chainID in the auth_asym_id fields, and so can be processed like any PDB file for further analysis. As an example of the utility of this function in PDBrenum, in Figure 7 we show the result of downloading the renumbered biological assembly structures of UniProt BMP2_HUMAN.

Acknowledgments
This work was supported by NIH grant R35 GM122517. We thank Vivek Modi and Mitchell Parker for testing the PDBrenum program.

Figure 7. Biological assemblies of human bone morphogenetic protein 2 (BMP2_HUMAN downloaded with PDBrenum. BMP2 is a homodimer (orange and blue) that binds Type I (magenta) and Type II receptors (cyan), RGM domain family members (yellow), and von Willebrand factor C-terminal domains (green).

References

1. Berman H, Henrick K, Nakamura H, Markley JL. The worldwide Protein Data Bank (wwPDB): ensuring a single, uniform archive of PDB data. Nucleic Acids Res. 2007;35(suppl_1):D301-D3.
2. Burley SK, Berman HM, Bhikadiya C, Bi C, Chen L, Di Costanzo L, et al. RCSB Protein Data Bank: biological macromolecular structures enabling research and education in fundamental biology, biomedicine, biotechnology and energy. Nucleic Acids Res. 2019;47(D1):D464-D74.
3. Armstrong DR, Berrisford JM, Conroy MJ, Gutmanas A, Anyango S, Choudhary P, et al. PDBe: improved findability of macromolecular structure data in the PDB. Nucleic Acids Res. 2020;48(D1):D335-D43.
4. Kinjo AR, Bekker GJ, Wako H, Endo S, Tsuchiya Y, Sato H, et al. New tools and functions in data-out activities at Protein Data Bank Japan (PDBj). Protein Sci. 2018;27(1):95-102.
5. Sayers EW, Cavanaugh M, Clark K, Ostell J, Pruitt KD, Karsch-Mizrachi I. GenBank. Nucleic Acids Res. 2019;47(D1):D94-D9.
6. UniProt Consortium. UniProt: a worldwide hub of protein knowledge. Nucleic Acids Res. 2019;47(D1):D506-D15.
7. Nemecek C, Metz WA, Wentzler S, Ding FX, Venot C, Souaille C, et al. Design of potent IGF1-R inhibitors related to bis-azaindoles. Chem Biol Drug Des. 2010;76(2):100-6.
8. Wu J, Li W, Craddock BP, Foreman KW, Mulvihill MJ, Ji Qs, et al. Small-molecule inhibition and activation-loop trans-phosphorylation of the IGF1 receptor. The EMBO journal. 2008;27(14):1985-94.
9. David FP, Yip YL. SSMap: a new UniProt-PDB mapping resource for the curation of structural-related information in the UniProt/Swiss-Prot Knowledgebase. BMC Bioinformatics. 2008;9(1):1-12.
10. Via A, Zanzoni A, Helmer-Citterich M. Seq2Struct: a resource for establishing sequence-structure links. Bioinformatics. 2005;21(4):551-3.
11. Martin AC. Mapping PDB chains to UniProtKB entries. Bioinformatics. 2005;21(23):4297-301.
12. Dana JM, Gutmanas A, Tyagi N, Qi G, O'Donovan C, Martin M, et al. SIFTS: updated Structure Integration with Function, Taxonomy and Sequences resource allows 40-fold increase in coverage of structure-based annotations for proteins. Nucleic Acids Res. 2019;47(D1):D482-D9.
13. Young J, Westbrook J, Feng Z, Sala R, Peisach E, Oldfield T, et al. PDBx/mmCIF: the foundation for the wwPDB onedep system. Foundations of Crystallography. 2017;70:C1361.
14. Abola EE, Sussman JL, Prilusky J, Manning NO. [29] Protein data bank archives of three-dimensional macromolecular structures. Methods Enzymol. 1997;277:556-71.
15. Krissinel E, Henrick K. Inference of macromolecular assemblies from crystalline state. J Mol Biol. 2007;372:774-97.
16. Hamelryck T, Manderick B. PDB file parser and structure class implemented in Python. Bioinformatics. 2003;19(17):2308-10.
17. McKinney W, editor Data structures for statistical computing in python. Proceedings of the 9th Python in Science Conference; 2010: Austin, TX.
18. Kluyver T, Ragan-Kelley B, Pérez F, Granger BE, Bussonnier M, Frederic J, et al. Jupyter

Notebooks-a publishing format for reproducible computational workflows2016.
19. Abhinandan K, Martin AC. Analysis and improvements to Kabat and structurally correct numbering of antibody variable domains. Mol Immunol. 2008;45(14):3832-9.
20. Xu Q, Dunbrack RL. ProtCID: a data resource for structural information on protein interactions. Nature communications. 2020;11(1):1-16.