# Apparent diet digestibility of captive colobines in relation to stomach types with special reference to fibre digestion

Satoru Hoshino<sup>1,2</sup>, Satoru Seino<sup>3</sup>, Takashi Funahashi<sup>4</sup>, Tomonori Hoshino<sup>4</sup>, Marcus Clauss<sup>5</sup>, Ikki Mastuda<sup>4,6,7,8</sup>, Masato Yayota<sup>2,9\*</sup>

<sup>1</sup> The United Graduate School of Agricultural Science, Gifu University, Gifu 501-1193, Japan

<sup>2</sup> Zoo Biology Research Center, Gifu University, Gifu 501-1193, Japan

<sup>3</sup> Preservation and Research Center, City of Yokohama, Yokohama 241-0804, Japan

<sup>4</sup> Japan Monkey Centre, Inuyama, Aichi, 484-0081 Japan

<sup>5</sup> Clinic for Zoo Animals, Exotic Pets and Wildlife, Vetsuisse Faculty, University of Zürich, Winterthurerstrasse 260, 8057 Zürich, Switzerland

<sup>6</sup> Chubu University Academy of Emerging Sciences, 1200, Matsumoto-cho, Kasugai-shi, Aichi 487-8501, Japan

<sup>7</sup> Wildlife Research Center of Kyoto University, 2-24 Tanaka-Sekiden-cho, Sakyo, Kyoto,
606-8203 Japan

<sup>8</sup> Institute for Tropical Biology and Conservation, Universiti Malaysia Sabah, Jalan UMS,
88400 Kota Kinabalu, Sabah, Malaysia

<sup>9</sup> Faculty of Applied Biological Sciences, Gifu University, Gifu 501-1193, Japan

\* Corresponding author

E-mail: <u>yayo@gifu-u.ac.jp (MY)</u>

## Abstract

Colobine monkeys are known for the anatomical complexity of their stomachs, making them distinct within the primate order. Amongst foregut fermenters, they appear peculiar because of the occurrence of two different stomach types, having either three ('tripartite') or four ('quadripartite', adding the praesaccus) chambers. The functional differences between tri and quadripartite stomachs largely remain to be explained. In this study, we aim to compare the apparent digestibility (aD) in tripartite and quadripartite colobines. Hence, we measured the aD in two colobine species, Nasalis larvatus (quadripartite) and Trachypithecus cristatus (tripartite), in two zoos. We also included existing colobine literature data on the aD and analysed whether the aD of fibre components is different between the stomach types to test the hypothesis of whether quadripartite colobines show higher aD of fibre components than tripartite colobines did. Our captive N. larvatus specimen had a more distinctively varying nutrient intake across seasons with a larger seasonal variation in aD than that of a pair of T. cristatus, which mostly consumed commercial foods with a lower proportion of browse and less seasonal variation. We observed higher aD of dry matter (DM), neutral detergent fibre (NDF) and acid detergent fibre (ADF) in the N. larvatus specimen, suggesting a higher gut capacity of N. larvatus provided by the additional praesaccus forestomach chamber. Based on the analysis of literature data for aD, we also found that quadripartite species achieved higher fibre digestibility at similar dietary fibre levels compared with tripartite species, supporting the hypothesis that the additional gut capacity offered by the praesaccus facilitates a longer retention and hence more thorough microbial fermentation of plant fibre.

## Introduction

Primates display a diverse array of digestive adaptations while covering various trophic niches, from folivory to frugivory, gummivory, insectivory, omnivory and nearly exclusive carnivory in some human populations [1, 2]. In particular, Old World monkeys of the subfamily Colobinae are known for the anatomical complexity of their stomachs, making them distinct within the primate order as the only 'foregut fermenters'. Their stomachs are complex and multi-chambered, harbouring a symbiotic microbiome that digest plant fibre and detoxify defensive plant chemicals, possibly allowing them to exploit a diet of leaves in greater quantities than other simple-stomached primates [3, 4]. Because of such anatomical complexity with their folivory, colobines have historically often been considered difficult to maintain healthy under zoo feeding regimens, especially when compared with frugivorous and/or omnivorous primates with simple stomachs [5, 6].

In previous studies, two different types of colobine forestomach have been distinguished. The so-called 'tripartite' type comprises a saccus, tubiform and glandular stomach part; these can be found in all colobines. The 'quadripartite' type has an additional blind sac, or pouch, named 'praesaccus', which is thought to represent an additional chamber. It is found in the genera *Procolobus, Piliocolobus, Rhinopithecus, Pygathrix* and *Nasalis* [7-9]. Notably, genera with a quadripartite stomach are notoriously difficult to maintain and breed in captivity, especially in temperate regions [10-13] where constant supply of fresh browse is especially difficult in autumn and winter, and chemical composition of fresh browse differs across plant species and seasons [14]. Therefore, to reduce gastrointestinal disorders and enhance health and survival in captive colobines, identifying an appropriate diet considering nutritional differences across plant species and seasons, in relation to their digestive physiology, is one of the goals for *ex situ* animal

management.

The functional differences between tri and quadripartite stomachs, however, largely remain unexplained. Matsuda, Chapman and Clauss [15] compiled literature data on the natural diet of colobine species to investigate the role of the praesaccus, suggesting that a larger gut capacity provided by an additional praesaccus is an important characteristic by which colobines survive on diets with a particularly high proportion of leaves. Thus, the higher intake capacity for species with quadripartite stomach would be assumed to be detrimental in the case of more digestible (commercial) diets in captivity than those in the wild, thereby leading to malfermentation by highly digestible components such as sugars or starch [5, 6]. Conversely, it may be assumed that species with tripartite stomachs are less susceptible to extreme bouts of malfermentation when fed highly digestible diets, simply because of their relatively reduced intake capacity. Evidently, these speculations remain to be tested.

One approach is to compare the apparent digestibility (aD), i.e. the ratio of the difference of the ingested and faecal nutrients to the ingested nutrients, in tripartite and quadripartite colobines to obtain information relevant for evaluating the digestive capacity for fibre. The aD evaluates the ability to break down and absorb nutrients, like fibre contained in browse/leaves, and thus measuring aD facilitates insights into the digestive adaptations and capacities of a species, and – maybe more importantly – also comparisons between species or species groups. It has previously been quantified in some colobines in comparison to simple-stomached primates, indicating that colobines show higher aD of fibre components, e.g. neutral detergent fibre (NDF) and acid detergent fibre (ADF), than simple-stomached primates, such as *Macaca fuscata* [16], *Alouatta* spp. [17] and *Nomascus siki* [18]. However, to our knowledge, a study focusing on comparing aD between tripartite and quadripartite colobines has not yet been undertaken. In our

comparison of aD between the colobines with different stomach types, we expected that quadripartite species with a putatively larger gut capacity would display higher aD of fibre than tripartite ones.

As a first preliminary approach to this question, we examined the aD of two captive colobine species, *Nasalis larvatus* (quadripartite) and *Trachypithecus cristatus* (tripartite), in two temperate region zoos. We focused on the seasonal difference of their aD to evaluate the effects of seasonal variation in nutrient composition throughout the year on their digestive efficiency. Additionally, to test the hypothesis of whether quadripartite colobines show higher aD of fibre components than tripartite colobines, we included existing colobine literature data on the aD and analysed whether the aD of fibre components is different between the stomach types.

## Materials and methods Ethics statement

We conducted the feeding experiments of proboscis monkey (N. larvatus) and silvered langur (*T. cristatus*) in the Yokohama Zoological Gardens, Zoorasia (approval ID: #256) and Japan Monkey Centre in Japan (approval ID: #2018-016), respectively. Invasive or stressful approaches such as capture, manual restraint or anaesthesia were not performed in this study. The materials were collected from animals non-invasively. This study was approved by the Welfare of Gifu University (approval ID: #17092). All animal experiment procedures were conducted following the Guidelines for Proper Conduct of Animal (Science Experiments Council of 2006: Japan, http://www.scj.go.jp/ja/info/kohyo/pdf/kohyo-20-k16-2e.pdf) and the Guidelines of Animal Research and Welfare of Gifu University (2008; https://www.gifuu.ac.jp/20150821-12a-experi.pdf).

## **Digestion trials**

In the Yokohama Zoological Gardens, the experiments were performed with one male *N. larvatus* (14 years old) housed individually. Three digestive trials were conducted in different seasons: autumn (3rd–16th September 2017), winter (2nd–15th January 2018) and summer (8th–15th June 2018). Each trial was composed of two different continuous periods: the acclimatisation (seven days) and sampling (seven days) periods. We were compelled to shorten one acclimatisation (8–12 June 2018) and sampling (13–15 June 2018) period due to heavy rain that soaked animal faeces and leftover leaves in the cage. Note that since the mean retention time for different markers in the whole digestive tract of *N. larvatus* has been reported as approximately 40 h [19], we believe that the shorter sampling period (72 h), though not ideal, is still suitable for assessing digestibility in this species. The animal was fed a mixed diet of seasonally available fresh leaves with branches and twigs harvested nearby the zoo and supplied by several commercial company or private farms (Ogawana farm, Dairi-en and Shiramori-en, Yokohama and Ohnishi Agricultural Corp. Yagazishima farm, Nago), vegetables and commercial pellets (S1 Table) two times daily. Water was freely available at all times.

In the Japan Monkey Centre, the trials were performed with two adult male *T. cristatus* (16 and 17 years old) housed together. Two experiments were conducted in different seasons: summer (30 July to 12 August 2018) and winter (14–27 February 2019) applying the same design as in *N. larvatus*, i.e. two continuous periods. Note that as their diets commonly fed did not differ in composition as much throughout the year compared to those in *N. larvatus*, we assessed these the *T. cristatus* and their diets only in two different seasons, selected for a maximum contrast of climatic conditions. The *T. cristatus* were fed a mixed diet of fresh leaves, vegetables and commercial pellets (S2 Table) three

times daily. The amounts of each feed were the same between trials across the seasons. Tree leaves harvested by the zoo staffs inside the zoological garden, were fed with branch and twigs once a day at noon, and the other feeds provided by commercial suppliers, were fed in the morning and evening.

We measured the body mass of all animals before and after the sampling periods in each experiment (*N. larvatus*: Digital Platform Scale, DP-8100, Yamato, Japan; *T. cristatus*: SD75LJP, OHAUS Corporation, USA), by offering weighing platforms on which the animals stepped voluntarily.

### **Sampling procedures**

Feed intake was quantitatively recorded over seven sampling days. Each food item was weighed before it was offered to the animals and left in their enclosures until the next feeding session. S1-S2 Table shows the mean ( $\pm$ standard error) daily amount of offered food per animal. All leftover food was removed, and the enclosure was cleaned before fresh food items were offered. All food items and leftovers were weighed with accuracy of 2 g (browse, UDS-500N, Yamato, Japan) or 1 g (others, UH-3201, A&D Company, Japan). Leftover weights were adjusted by deriving a desiccation factor from the measured moisture lost from similar sets of food placed in a desiccation pan in an area adjacent to the primate enclosures. For the two *T. cristatus* at Japan Monkey Centre housed together, individual feed intake and faeces output were calculated as the average of total measures divided by two.

We collected equal amounts of each feedstuff at every feeding time during the sampling period and stored them in a refrigerator (10°C: trials for *N. larvatus*; 4°C: trials for *T. cristatus*) and a freezer (-18 to 20°C: only leaves and banana peel for *T. cristatus*) for nutritional analysis. We also collected all faeces shortly before every feeding time and

immediately preserved them in the freezer. We mixed each feed sample and then collected 100 g of leaves of each browse species and all amounts of other foods as representative samples. We mixed all faeces with 1 ml/100 g of 10% formalin solution and preserved 500 g faeces as a representative sample [20].

After measuring the fresh weight of representative samples, in the case of *N*. *larvatus*, we lyophilised leaf and faeces samples using a freeze-dryer (DC400/800, Yamato, Japan). For other feedstuff samples (vegetables and commercial pellets), we used an air-forced dry oven (DKM812, Yamato, Japan) for 48 h at 60°C. Note that we mashed soybean and peanuts in a mortar due to their high fat content and then soaked and washed the mixture with ether to extract fat. After fat extraction, the soybean and peanuts were dried using an air-forced dry oven (DKM600, Yamato, Japan) for 3 h at 60°C. In the case of *T. cristatus*, we dried all feedstuff and faecal samples using an air-forced dry oven (DKM812, Yamato, Japan) for 48 h at 60°C.

We ground leaf and faecal samples using a Wiley mill through a 2-mm screen and ground other feed samples using a coffee mill to avoid heat denaturation of sugars in the feeds.

## **Estimation of apparent digestibility (aD)**

We used the average daily feed intake and faecal output during the sampling term to estimate the aD in each trial. We analysed the dry matter (DM), crude ash (CA), crude protein (CP) and acid detergent fibre (ADFom) in accordance with AOAC 930.15, 942.05, 990.03 and 973.18, respectively [21]. We then analysed neutral detergent fibre (aNDFom) according to the method of van Soest, Robertson and Lewis [22]. Detergent fibre data are presented without residual ash. We calculated aD of each nutrient (N) according to the following equation:  $aD_N$  (%) = (N<sub>feed intake</sub>-N<sub>feces</sub>) / N<sub>food intake</sub> × 100.

## Literature data and analysis

Together with our new experimental data, as shown in Table 1, we used published aD data of seven colobine species, including four tripartite species (13 datasets) [16, 17, 23-26] and two quadripartite species (five datasets) [17, 18, 27] to compare digestive capacity between the stomach groups. These published data included total feed and fibre intake and aD of DM, NDF and ADF, and for some but not all studies the body mass of the animals used.

Due to the limited data, we only focussed on two-factorial models and did not perform models with more variables/factors. First, we tested two-factorial linear models that linked the aD to either combination of body mass, stomach type and NDF content of the ingested diet. Second, in the larger dataset (including the studies did not provide body mass information), we examined the relationship between aD in DM, NDF and ADF and the dietary NDF content in each colobine group with different stomach type to estimate the slope and intercept using linear regression analysis. Analysis of covariance served to compare the slopes and intercepts of regression lines between the stomach types. Note that we could not conduct the same statistical analysis for the aD of CP because of the lack of published data. Additionally, because Nijboer et al. [28] analysed only crude fibre as fibre contents and Coudrat and Cabana [18] did not specify DM contents in diets and faeces with the total intake (see Table 1), we could not include these data in our analysis. All statistical analyses were performed in Spyder (Python 3.7).

## Results

## **Digestion trials**

The body mass of Nasalis larvatus in autumn, winter and summer was 17.3, 18.9 and

17.8 kg, respectively, shortly before beginning of the sampling periods. The composition and amount of the diet were different amongst trials (S1 Table). The ratio of leaves/other feed intake were 7.4:1, 3.3:1 and 3.5:1, in autumn, winter and summer. Browse species generally contained more fibre than other feeds (S3–S5 Table), possibly leading to different proportion of ADFom in the diet in autumn, i.e. 33.3%, 29.4% and 30.1% of DM in autumn, winter and summer, respectively. There was notable seasonal variation in the nutritional composition of the browse species (S3–S5 Table). The DM and ash contents of laurel (*Machilus thunbergii*) in winter were 5%–10% higher than those in summer and winter. The aNDFom and ADFom of each browse species in winter were 5%–10% lower than those in summer and autumn. On the other hand, nutrient contents of fruits, vegetables, beans, starchy foods and commercial products in *N. larvatus* did not differ between the seasons (S3–S5 Table). The aD of DM was 69.9%, 79.6% and 73.7% in autumn, winter and summer, respectively (Table 2). Likewise, the aD of CP, aNDFom and ADFom varied amongst the three seasons (Table 2).

The body mass of the two *T. cristatus* was 7.6 and 7.6 kg in summer and 6.8 and 7.7 kg in winter shortly before beginning of the sampling period. There was strong seasonality in the nutritional composition of their browse (S6–S7 Table). The DM and ash contents in the browse species in *T. cristatus* were higher, and CP, aNDFom and ADFom were lower in winter than in summer. The other diet items, i.e. fruits,vegetables, starchy foods and commercial product, did not differ between the two seasons (S6–S7 Table). The aD of DM, ash and ADFom did not differ between the two seasons; only the aD of CP and aNDFom changed between the two seasons (Table 2).

#### Literature data

#### **Apparent digestibility**

Body mass differed significantly between species with a tripartite (mean = 7.39 kg; sd = 0.71) or a quadripartite (13.7 kg; sd = 4.23) stomach (Z = 3.40, p < 0.001).

In a model linking aD DM with body mass and stomach type ( $R^2 = 0.016$ , p = 0.862), neither body mass (F=0.015, p = 0.903) nor stomach type (F= 0.072, p= 0.792) were significant. However, for aD NDF and aD ADF, particularly stomach type approached significance in both models, and body mass in the model for aD ADF (whole model:  $R^2 = 0.170$ , p = 0.187 and  $R^2 = 0.252$ , p = 0.074; body mass F = 2.252, p = 0.150 and F = 3.025, p = 0.074; stomach type: F = 3.866, p = 0.064 and F = 3.025, p = 0.074; respectively).

On the other hand, in a model linking aD DM with body mass and NDF contents of the ingested diet ( $R^2 = 0.571$ , p = 0.0005), only the NDF content (F= 22.04, p < 0.001) was significant but not body mass (F = 0.015, p = 0.903). For aD NDF and aD ADF, the neither body mass nor NDF content were significant (whole model:  $R^2 = 0.287$ , p = 0.047and  $R^2 = 0.364$ , p = 0.017; body mass: F = 2.252, p = 0.150 and F = 2.998, p = 0.100; NDF contents: F = 2.138, p = 0.160 and F = 2.744, p = 0.114, respectively). Note that in comparison to the models presented further below, sample sizes were limited as not all literature provided data on body mass.

The final model for the limited dataset evaluated the effects of aD DM on the stomach type and NDF content ( $R^2 = 0.550$ , p = 0.001), where only the NDF content (F= 22.04, p = 0.0002) was significant but not stomach type (F = 0.072, p = 0.792). For aD NDF and aD ADF, only stomach type was significant or approached significance (whole model:  $R^2 = 0.291$ , p = 0.045 and  $R^2 = 0.406$ , p = 0.009; stomach type: F = 3.866, p = 0.064 and F = 6.382, p = 0.021; NDF contents: F = 2.138, p = 0.160 and F = 2.744, p = 0.114, respectively).

In the larger dataset, there was a significant negative relationship between the

aD of DM and the NDF content of the ingested diet of six colobine species (r = -0.770, p < 0.001). The difference in the regression slopes and intercepts for aD of DM vs. NDF content of the ingested diet between the stomach types was not significant (Fig 1; slope, t = 1.079; p = 0.293; intercept, t = 1.092, p = 0.287). There was also a significant negative relationship between the aD of NDF or the aD of ADF and the NDF content of the ingested diet of six colobine species (NDF, r = -0.459, p = 0.021; ADF, r = -0.439, p = 0.028). The differences in the regression intercepts for both aD of NDF and aD of ADF vs. NDF content were significant (NDF, t = -2.559, p = 0.018; ADF, t = -2.487, p = 0.021) between the stomach types (Fig 1), while the slopes were not different (NDF, t = 0.267, p = 0.792; ADF, t = 0.152, p = 0.881).

## Discussion

As expected, we confirmed the variation in the aD because of the nutrient differences in the diets across the seasons. Each browse species' nutrient composition changed seasonally, as reported for deciduous tree species in North American zoos [14], whereas those of commercial food items were stable (S3–S7 Table). The aD of DM, aNDFom and ADFom in *N. larvatus* clearly changed across seasons: aD DM by 9.1%, aD aNDFom by 11.2 % and aD ADFom by 5.1%. On the other hand, aD was relatively stable between the two different seasons in *T. cristatus*, although the total DM intake in winter was slightly higher than that in summer. As in the present study, Edwards and Ullrey [17] noted that the aD of DM in three colobine species (*Colobus guereza, Pygathrix nemaeus* and *Trachypithecus francoisi*) decreased with an increased fibre level in the diet, supporting that aD is affected by a variation in food composition and seasonally varying nutrient contents in feeds. Consequently, the aD of *N. larvatus* had a larger seasonal variation, with more distinctively varying nutrient intake across seasons, than that of *T. cristatus*,

which mostly consumed commercial foods with a lower proportion of browse. Evaluating the nutrient digestibility of captive colobines on different diets throughout the year may contribute to their health management and predict intake requirements across different diets and seasons.

Comparison of the nutrient composition (especially fibre) of faeces in freeranging and captive individuals has been proposed to obtain information relevant for the improvement of diets of colobines [10]. *Nasalis larvatus* in our study had faecal NDF contents (35%–41% in DM) that were higher than those reported in other captive conspecifics, i.e. 17% [27] (mean of two different values), but lower than those of freeranging ones, i.e. 53%–70% [10]. Although faecal NDF contents of free-ranging *T. cristatus* are not available, those in our study (30%–31%) were comparable to other closely related species in captivity, i.e. 37% (mean of six different values in *T. auratus*) [24] and 31% (mean of three different values in *T. francoisi*) [23, 29], although still far lower than those reported for free-ranging *N. larvatus*. Altering the diets of captive colobines to include more fibre, comparable to those of free-ranging ones, may be recommendable.

In the present study, the NDF level of the total DM intake was much higher in the quadripartite species *N. larvatus* (35.6%–37.9%) than in the tripartite species *T. cristatus* (12.9%–15.0%), because of a much higher proportion of browse fed to the former. This zoo practice may stem from the impression that quadripartite species are generally more difficult to maintain in captivity; therefore, more effort is undertaken to provide them with feed items considered natural for them, mainly browse. Correspondingly, we observed a higher DM intake (% BW, shown in S3–S5 Table) in *N. larvatus* compared with those in *T. cristatus* that could be interpreted as compensation for the higher fibre levels. However, regardless of the higher fibre and intake levels, we

observed higher aD in DM, aNDFom and ADFom (Table 2) in the *N. larvatus* specimen. This observation is most parsimoniously explained by a higher gut capacity in the proboscis monkey, provided by the additional praesaccus forestomach chamber. Typically, a higher relative food intake leads to shorter digesta retention times and can also compromise digestibility [30, 31, cf. also Fig 2]. However, a higher gut capacity can mitigate this effect, and this may be the main adaptive value of the praesaccus in quadripartite species [15].

Unfortunately, the currently available information for tri- and quadripartite species is biased in terms of the body mass of the investigated specimens. Although the overall distribution of tri- and quadripartite stomach types across colobine genera [15] does not suggest a body size effect on stomach anatomy, quadripartite specimens of the literature data collection were significantly heavier than tripartite specimens. This is due to the inclusion of *N. larvatus* – the largest colobine, with a quadripartite stomach. Given an older but prevailing assumption in the ecophysiological literature that body mass is positively related to digesta retention time and digestibility [32], this could lead to the suspicion that differences between the stomach types are body size effects. However, this is refuted on the one hand by our own analyses that showed no effect of body mass or a trend subordinate to the trend of stomach type, and on the other hand by more recent literature. An effect of body size on digesta retention and digestion has been contested by various large empirical data collections as well as by theoretical considerations [31, 33, 34], including exclusive analyses of primate data.

The results of our analysis for aD using the larger literature data set further suggested functional differences between the stomach types. There was no difference in how dietary fibre content influenced the overall DM digestibility, which is, in most of the diets used in these studies, most likely dominated by the digestion of non-fibrous carbohydrates and protein. However, stomach type had a significant effect on how dietary fibre levels affected the digestibility of fibre itself. Here, quadripartite species achieved higher fibre digestibility at similar dietary fibre levels than tripartite species, suggesting that the additional gut capacity offered by the praesaccus facilitates longer retention and hence more thorough microbial fermentation of plant fibre. One notable tendency was that *N. larvatus* achieved a particularly high digestibility, driving the difference between the two stomach types. In this species, regurgitation and remastication have been observed in the wild [35], and a smaller faecal particle size of *N. larvatus* compared with other colobines has been reported [36]. Because particle size reduction via chewing is one of several key factors affecting digestibility [37], we cannot determine whether the difference in digestibility observed here is related to stomach type or chewing efficiency. Ideally, in future studies, similar diets should be employed as this will allow meaningful comparison of faecal particle size; chewing behaviour should also be observed.

Unfortunately, the current available data on digesta retention times in colobine species does not allow testing for a general difference between tripartite and quadripartite species (Fig 2). Between species, retention times need to be compared in relation to the food intake level [30, 38]. However, the ranges of intake level in published studies hardly overlap between the stomach groups, making a reasonable comparison impossible. To test whether quadripartite species achieve longer digesta retention because of a higher gut fill, comparative studies with different species on a similar (possibly browse-dominated) diet would be required where intake, digestibility and digesta retention are assessed in the same experiment, additionally facilitating the calculation of gut fill [39, 40]. Ideally, such a study would also address the problem of the very limited sample size of the present

experiments. Until such a study is performed, our results must be considered preliminary, delivering plausible hypotheses.

It should be noted that we cannot exclude the possibility of the effects of specific fibre-digesting bacteria in the presaccus in quadripartite species. So, far, only a few analyses of the forestomach microbiome are available for colobines. Although the recent developments in sequencing technology describe the foregut microbiome in some colobines, e.g. *N. larvatus* [41, 42] and *Rhinopithecus roxellana* [43], the function of these microbe species has not been evaluated. However, there is currently also no reason to assume that the praesaccus should harbour a fundamentally different microbiome from the saccus. Detailed studies about differences in the microbiome at different forestomach locations, as available in ruminants [e.g. 44], do not exist for colobines so far. In contrast to the detailed knowledge about the differential function of individual forestomach sections in ruminants and camelids [e.g. 45], there is, to date, no indication of the differential function of the provision of sheer fermentation chamber capacity, and this may well also apply to colobines.

## Acknowledgements

Our appreciation goes to the animal caretakers and veterinarians from Zoorasia and Japan Monkey Centre, namely, Ryuta Kawasaki and Kei Watanabe, without whom this project would not have been possible. General supervision and administrative supports were provided by Dr. Takashi Hayakawa and Dr. Koshiro Watanuki as the counterparts of Japan Monkey Centre. In particular, S.H. thanks to Dr. T. Hayakawa for his kind arrangement of the freezer to preserve the collected samples at his laboratory of the Primate Research Institute, Kyoto University and for his technical editing of this manuscript.

## **Author contributions**

SH, IM, MC and MY conceptualised the idea and drafted the manuscript; SH performed the feeding trials; SH, IM, MC and MY performed and interpreted the statistical analysis; SS, TF and TH arranged the sampling in the zoos and MY organised the projects. All authors contributed to the final version of the manuscript.

## References

Campbell CJ, Fuentes A, MacKinnon KC, Bearder SK, Stumpf RM. Primates in perspective.
 edition S, editor. New York: Oxford University Press; 2011.

Lambert JE. Primate digestion: Interactions among anatomy, physiology, and feeding ecology.
 Evolutionary Anthropology: Issues, News, and Reviews. 1998;7(1): 8-20. doi: 10.1002/(sici)1520-6505(1998)7:1<8::aid-evan3>3.0.co;2-c.

Chivers D. Functional anatomy of the gastrointestinal tract. In: Davies A, Oates J, editors.
 Colobine monkeys: their ecology, behaviour and evolution. Cambridge: Cambridge University Press;
 1994. pp. 205–227.

Bauchop T, Martucci RW. Ruminant-like digestion of the langur monkey. Science.
 1968;161(3842): 698-700. doi: 10.1126/science.161.3842.698.

5. Clauss M, Dierenfeld ES. The nutrition of browsers. In: Fowler ME, Miller RE, editors. Zoo and Wild Animal Medicine: Current Therapy. 6 ed. St. Louis: Saunders Elsevier; 2008. pp. 444-454.

Matsuda I, Clauss M. Morphology and physiology of colobine digestive tracts. In: Matsuda I,
 Grueter CC, Teichroeb JA, editors. The Colobines: Natural History, Behaviour and Ecological Diversity.
 Cambridge: Cambridge University Press; in press.

Langer P. The mammalian herbivore stomach. Stuttgart/New York: Gustav Fischer Verlag;
 1988.

Langer P. Comparative anatomy of the gastrointestinal tract in Eutheria: Taxonomy,
 biogeography and food. Vol I: Afrotheria, Xenarthra and Euarchontoglires. Vol II: Laurasiatheria, general
 discussion. Berlin: De Gruyter; 2017.

9. Caton JM. The morphology of the gastrointestinal tract of *Pygathrix nemaeus*. In: Jablonski NG, editor. Natural history of the doucs and snub-nosed monkeys. Singapore: World Scientific; 1998. pp. 129-149.

 Matsuda I, Bernard H, Tuuga A, Nathan S, Sha JCM, Osman I, et al. Fecal nutrients suggest diets of higher fiber levels in free-ranging than in captive proboscis monkeys (*Nasalis larvatus*). Front Vet Sci. 2017;4:246. doi: 10.3389/fvets.2017.00246. PubMed PMID: 29404345; PubMed Central PMCID: PMCPMC5780573.

11. Struhsaker TT. The Red Colobus Monkeys: Variation in Demography, Behavior, and Ecology of Endangered Species. New York: Oxford University Press; 2010.

 Hollihn UWE. Remarks on the breeding and maintenance of Colobus monkeys *Colobus* guereza, Proboscis monkeys *Nasalis larvatus* and Douc langurs *Pygathrix nemaeus* in zoos. International Zoo Yearbook. 1973;13(1): 185-188. doi: 10.1111/j.1748-1090.1973.tb02146.x.

Lhota S, Sha JCM, Bernard H, Matsuda I. Proboscis monkey conservation: beyond the science.
 In: Behie AM, Teichroeb JA, Malone NM, editors. Primate Research and Conservation in the
 Anthropocene. Cambridge Cambridge University Press; 2019. pp. 182-196.

Nijboer J, Dierenfeld ES. Comparison of diets fed to southeast Asian colobines in North American and European zoos, with emphasis on temperate browse composition. Zoo Biology.
1996;15(5): 499–507. doi: 10.1002/(sici)1098-2361(1996)15:5<499::aid-zoo6>3.0.co;2-6.

Matsuda I, Chapman CA, Clauss M. Colobine forestomach anatomy and diet. J Morphol.
 2019;280(11): 1608-1616. doi: 10.1002/jmor.21052. PubMed PMID: 31424606.

16. Sakaguchi E, Suzuki K, Kotera S, Ehara A, editors. Fibre digestion and digesta retention time in Macaque and Colobus monkeys. XIIIth congress of the Int Primatolgy Society; 1991: Elsevier

18

Edwards MS, Ullrey DE. Effect of dietary fiber concentration on apparent digestibility and digesta passage in non-human primates. II. Hindgut- and foregut-fermenting folivores. Zoo Biology.
1999;18(6): 537-549. doi: 10.1002/(sici)1098-2361(1999)18:6<537::aid-zoo8>3.0.co;2-f.

18. Coudrat C, Cabana F. Preliminary results on the food intake and nutrient digestibility of southern white-cheeked gibbons (*Nomascus siki*) and red-shanked douc langurs (*Pygathrix nemaeus*) at the Endangered Primate Rescue Center, Vietnam. Vietnamese Journal of Primatology. 2019;3(1): 71-76.

19. Matsuda I, Sha JC, Ortmann S, Schwarm A, Grandl F, Caton J, et al. Excretion patterns of solute and different-sized particle passage markers in foregut-fermenting proboscis monkey (*Nasalis larvatus*) do not indicate an adaptation for rumination. Physiol Behav. 2015;149: 45-52. doi:

10.1016/j.physbeh.2015.05.020. PubMed PMID: 26004169.

20. Ishida M. Digestive trial for ruminants. In: Ishibashi T, editor. Experimental Methods for Animal Nutrition. Tokyo: Yokendo; 2001. pp. 190-197.

21. AOAC. Official Methods of Analysis of AOAC International. Gaithersburg, MD: Association of Official Analytical Chemists International; 2012.

van Soest PJ, Robertson JB, Lewis BA. Methods for Dietary Fiber, Neutral Detergent Fiber,
 and Nonstarch Polysaccharides in Relation to Animal Nutrition. Journal of Dairy Science. 1991;74(10):
 3583-3597. doi: 10.3168/jds.S0022-0302(91)78551-2.

Nijboer J, Becher F, van der Kuilen J, Beynen AC. Chemical analysis and consistency of faeces produced by captive monkeys (*Francois langurs, Trachypithecus francoisi*) fed supplemental fibre.
Veterinary Quarterly. 2001;23(2): 76-80. doi: 10.1080/01652176.2001.9695086. PubMed PMID: 11361103.

24. Nijboer J, Clauss M, Olsthoorn M, Noordermeer W, Huisman TR, Verheyen C, et al. Effect of diet on the feces quality in javan langur (*Trachypithecus auratus auratus*). Journal of Zoo and Wildlife Medicine. 2006;37(3): 366–372. PubMed PMID: 17319137.

25. Oftedal OT, Jakubasz M, Whetter P. Food intake and diet digestibility by captive black and

white colobus (*Colobus guereza*) at the National Zoological Park. Ann Proc Amer Assoc Zoo Vet. 1982;33: (Abstract).

26. Watkins BE, Ullrey DE, Whetter PA. Digestibility of a high-fiber biscuit-based diet by black and white colobus (*Colobus guereza*). American Journal of Primatology. 1985;9(2): 137-144. doi: 10.1002/ajp.1350090207.

27. Dierenfeld ES, Koontz FW, Goldstein RS. Feed intake, digestion and passage of the proboscis monkey (*Nasalis larvatus*) in captivity. Primates. 1992;33(3): 399-405. doi: 10.1007/bf02381201.

28. Nijboer J, Clauss M, van de Put K, van der Kuilen J, Woutersee H, Beynen AC. Influence of two different diets on fluid and particle retention time Javan langur (*Trachypithecus auratus auratus*). Der Zoologische Garten. 2007;77(1): 36-46. doi: 10.1016/j.zoolgart.2007.06.004.

29. Nijboer J. Fibre intake and faeces quality in leaf-eating primates: Utrecht University; 2006.

30. Clauss M, Streich WJ, Nunn CL, Ortmann S, Hohmann G, Schwarm A, et al. The influence of natural diet composition, food intake level, and body size on ingesta passage in primates. Comp Biochem Physiol A Mol Integr Physiol. 2008;150(3): 274–281. doi: 10.1016/j.cbpa.2008.03.012. PubMed PMID: 18450489.

31. Müller DW, Codron D, Meloro C, Munn A, Schwarm A, Hummel J, et al. Assessing the Jarman-Bell Principle: Scaling of intake, digestibility, retention time and gut fill with body mass in mammalian herbivores. Comp Biochem Physiol A Mol Integr Physiol. 2013;164(1): 129–140. doi: 10.1016/j.cbpa.2012.09.018. PubMed PMID: 23047052.

32. Demment MW, Van Soest PJ. A nutritional explanation for body-size patterns of ruminant and nonruminant herbivores. The American Naturalist. 1985;125(5): 641-672. doi: 10.1086/284369.

33. Clauss M, Steuer P, Muller DW, Codron D, Hummel J. Herbivory and body size: allometries of diet quality and gastrointestinal physiology, and implications for herbivore ecology and dinosaur gigantism. PLoS One. 2013;8(10): e68714. doi: 10.1371/journal.pone.0068714. PubMed PMID: 24204552; PubMed Central PMCID: PMCPMC3812987. 34. Steuer P, Südekum K-H, Tütken T, Müller DWH, Kaandorp J, Bucher M, et al. Does body
mass convey a digestive advantage for large herbivores? Functional Ecology. 2014;28(5): 1127-1134. doi:
10.1111/1365-2435.12275.

Matsuda I, Murai T, Clauss M, Yamada T, Tuuga A, Bernard H, et al. Regurgitation and remastication in the foregut-fermenting proboscis monkey (*Nasalis larvatus*). Biology letters. 2011;7(5):
786-789. doi: 10.1098/rsbl.2011.0197. PubMed PMID: 21450728; PubMed Central PMCID: PMC3169055.

Matsuda I, Tuuga A, Hashimoto C, Bernard H, Yamagiwa J, Fritz J, et al. Faecal particle size in free-ranging primates supports a 'rumination' strategy in the proboscis monkey (*Nasalis larvatus*).
 Oecologia. 2014;174: 1127-1137. doi: 10.1007/s00442-013-2863-9. PubMed PMID: 24380969.

37. Hummel J, Clauss M, Südekum K-H. Aspects of food comminution in ungulates and their consequences for energy budget. In: Martin T, von Koenigswald W, editors. Mammalian teeth - form and function. Munich: Dr. Friedrich Pfeil; 2020.

38. Levey DJ, Martinez del Rio C. Test, rejection, and reformulation of a chemical reactor-based model of gut function in a fruit-eating bird. Physiol Biochem Zool. 1999;72(3): 369-383. doi: 10.1086/316663. PubMed PMID: 10222331.

39. Munn AJ, Tomlinson S, Savage T, Clauss M. Retention of different-sized particles and derived gut fill estimate in tammar wallabies (*Macropus eugenii*): physiological and methodological considerations. Comp Biochem Physiol A Mol Integr Physiol. 2012;161(2): 243-249. doi:

10.1016/j.cbpa.2011.11.003. PubMed PMID: 22094100.

40. Munn A, Stewart M, Price E, Peilon A, Savage T, Van Ekris I, et al. Comparison of gut fill in sheep (*Ovis aries*) measured by intake, digestibility, and digesta retention compared with measurements at harvest. Canadian Journal of Zoology. 2015;93(10): 747-753. doi: 10.1139/cjz-2014-0314.

41. Hayakawa T, Nathan S, Stark DJ, Saldivar DAR, Sipangkui R, Goossens B, et al. First report of foregut microbial community in proboscis monkeys: are diverse forests a reservoir for diverse

microbiomes? Environ Microbiol Rep. 2018;10(6): 655-662. doi: 10.1111/1758-2229.12677. PubMed PMID: 29992728.

42. Suzuki-Hashido N, Tsuchida S, Hayakawa T, Sakamoto M, Azumano A, Seino S, et al. *Lactobacillus nasalidis* sp. nov., isolated from the forestomach of a captive proboscis monkey (*Nasalis larvatus*). International Journal of Systematic and Evolutionary Microbiology. 2021;71(4). doi: 10.1099/ijsem.0.004787.

43. Zhou X, Wang B, Pan Q, Zhang J, Kumar S, Sun X, et al. Whole-genome sequencing of the snub-nosed monkey provides insights into folivory and evolutionary history. Nat Genet. 2014;46(12): 1303-1310. doi: 10.1038/ng.3137. PubMed PMID: 25362486.

44. Zeitz JO, Ineichen S, Soliva CR, Leiber F, Tschuor A, Braun U, et al. Variability in microbial population and fermentation traits at various sites within the forestomach and along the digestive tract as assessed in goats fed either grass or browse. Small Ruminant Research. 2016;136: 7-17. doi:

10.1016 / j. small rum res. 2015. 12.029.

Idalan N, Martin LF, Clauss M. Physical characteristics of gastrointestinal content of llama (*Lama glama*). J Anim Physiol Anim Nutr (Berl). 2019;103(4): 1015-1022. doi: 10.1111/jpn.13116.
PubMed PMID: 31050031.

46. Schwarm A, Ortmann S, Fritz J, Flach E, Rietschel W, Clauss M. No distinct stratification of ingesta particles and no distinct moisture gradient in the fore-stomach of non-ruminants: The wallaby, peccary, hippopotamus, and sloth. Mammalian Biology - Zeitschrift für Säugetierkunde. 2013;78(6): 412–421. doi: 10.1016/j.mambio.2013.04.001.

47. Schwarm A, Ortmann S, Wolf C, Streich WJ, Clauss M. Passage marker excretion in red kangaroo (*Macropus rufus*), collared peccary (*Pecari tajacu*) and colobine monkeys (*Colobus angolensis*, *C. polykomos*, *Trachypithecus johnii*). Journal of experimental zoology Part A, Ecological genetics and physiology. 2009;311(9): 647-661. doi: 10.1002/jez.552. PubMed PMID: 19551808.

48. Matsuda I, Espinosa-Gomez FC, Ortmann S, Sha JCM, Osman I, Nijboer J, et al. Retention

bioRxiv preprint doi: https://doi.org/10.1101/2021.03.02.433677; this version posted May 30, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

marker excretion suggests incomplete digesta mixing across the order primates. Physiol Behav. 2019;208:

112558. doi: 10.1016/j.physbeh.2019.112558. PubMed PMID: 31125579.

## **Supporting information**

S1 Table. Feed composition, and offered and leftover amounts (g as fed/day) in each digestion trial of an adult male of *Nasalis larvatus*. \*<sup>1</sup> Weight unit is  $X \pm SE$  g, as fed/day/animal. \*<sup>2</sup> We measured the weight of the whole branch, including leaves and twigs.

S2 Table. Feed composition, and offered and leftover amounts (g as fed/day/two animals) in each digestion trial of two adults of *Trachypithecus cristatus*. \*1 Weight unit is  $X \pm SE$  g FM/day. \*2 We measured the weight of the whole branch, including leaves and twigs.

**S3 Table.** Nutrient contents in the feeds and DM intake of an adult male of *Nasalis larvatus* in September 2017 (autumn). DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash; ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg). \*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse. \*2 DM intake of each feed/total DM intake. \*3 BW of the proboscis monkey was 17.3 kg at the beginning of the second week of the experiment.

**S4 Table.** Nutrient contents in the diet and DM intake of an adult male of *Nasalis larvatus* in January 2018 (winter). DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash; ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg). \*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse. \*2 DM intake of

each feed/total DM intake. \*3 BW of the proboscis monkey was 18.9 kg.at the beginning of the second week of the experiment. \*4 The pelleted feed was soaked in water before feeding

**S5 Table.** Nutrient contents in the diet and DM intake of an adult male of *Nasalis larvatus* in June 2018 (summer). DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash; ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg). \*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse. \*2 DM intake of each feed/total DM intake. \*3 BW of the proboscis monkey was 17.8 kg at the beginning of the second week of the experiment.

**S6 Table.** Nutrient contents in the diet and DM intake of two adults *Trachypithecus cristatus* in August 2018 (summer). DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash; ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg). \*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse. \*2 DM intake of each feed/total DM intake. \*3 BW of two silver lutungs were 7.6 and 7.6 kg at the beginning of the second week of the experiment.

**S7 Table. Nutrient contents in the diet and DM intake of two adults of** *Trachypithecus cristatus* **in February 2019 (winter).** DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash; ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body

mass (kg). \*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse. \*2 DM intake of each feed/total DM intake. \*3 BW of two silver lutungs were 6.8 and 7.7 kg at the beginning of the second week of the experiment

	olobus monkey	Body mass			stuff		Ap	parent digestibi	Reference	
Stomach compartments	species	(kg)	Browse	Fruits Vegetable	Artificial Food	Others*1	DM	NDF	ADF	
		10.9			✓ (15ADF)		80.3	77.0	80.1	Educada and Hilana [17]
		10.7			✓ (30ADF)		78.7	74.3	56.2	Edwards and Ullrey [17]
	Colobus guereza	7.0 *2	1	1	1		65	56	56	Nijboer et al. [24] and
	Colobus guerezu	7.0 *	1	1	1		68	63	56	Nijboer [29]
		8.9		1	1		83 (82-84)	68	68	Oftedal, Jakubasz and Whetter [25]
		8.0		1	1	1	$87.1\pm0.1$	$81.3\pm1.0$	$69.3\pm0.5$	Watkins, Ullrey and Whetter [26]
	3 Trachypithecus francoisi	6.0			✓ (15ADF)		82.4±3.0	$79.3\pm5.3$	$82.3\pm4.8$	Edwards and Ullrey [17]
3		6.0			✓ (30ADF)		$76.7\pm3.5$	$75.7\pm7.7$	$76.9 \pm 12.8$	Edwards and Onrey [17]
			1	1	1		58.9	46.6	38.5	
		-	1	1	1		64.7	62.2	56.1	Nijboer et al. [23]
			1	1	1		74.0	68.0	65.3	
	Trachypithecus auratus	6.0	1	1	1	1	68	-	-	Nijboer et al. [28]
	~ *	0.0	1			1	59	-	-	5 2 5
	Trachypithecus cristatus	-					$75.7 \pm 5.5$	$68.9 \pm 5.8$	$61.6\pm7.5$	Sakaguchi et al. [16]
							91	70	53	
	Trachypithecus obscurus	6.0 *2					84	77	54 65	Nijboer et al. [24]
							78 78	74 64	65 60	•
			V	V	······	V		86.4	85.0	
	Nasalis larvatus	9.0 *2		v 1		· ·	88.3 88.8	86.2	85.0 86.0	Dierenfeld, Koontz and Goldstein [27]
		12.1 (15ADF) Pygathrix nemaeus 11.75 (30ADF)		v		v	00.0	80.2	80.0	
4				(15ADF)			76.2	66.5 66.6		Edwards and Ullrey [17]
	Pygathrix nemaeus				73.9	69.8	67.6			
		-	1	1	. ,		-	89.2	83.9	Coudrat and Cabana [18]

Table 1. Summary of colobine apparent digestibility and body mass in the previous studies

\*1 Others containing non-fibrous carbohydrate (rice, bread, sweet potato, cereal and Saint John's bread), animal protein (boiled egg, meat and mealworm) and vegetable

protein (tofu).

\*2 Median of specimens used for digestive trials.

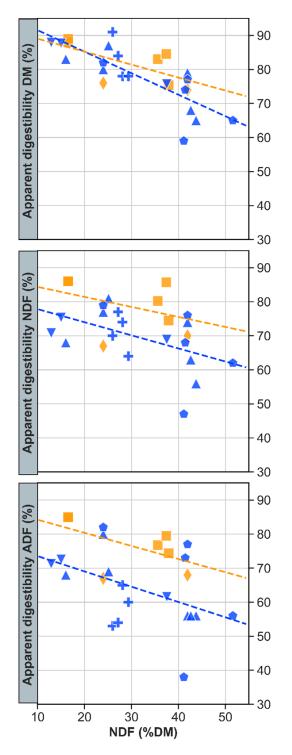
	N	asalis larvat	us	Trachyp crist	
	2017	2018	2018	2018	2019
	$\mathbf{Sep}$	Jan	Jun	Aug	Feb
	(Autumn)	(Winter)	(Summer)	(Summer)	(Winter)
Body mass (kg)	17.3	18.9	17.8	7.6 / 7.6	6.8 / 7.7
DM intake (g DM/day/animal)	702.0	654.9	664.3	146.0	133.3
Fecal output (g DM/day/animal)	172.3	109.1	102.8	17.6	16.4
Ingested diet					
OM (%DM)	91.3	90.6	92.6	93.8	93.4
CP (%DM)	11.7	11.7	11.9	7.3	7.9
aNDFom %DM)	37.9	35.3	37.4	12.9	15.0
ADFom (%DM)	33.3	29.1	30.1	10.3	12.7
Feces					
OM (%DM)	89.3	88.5	87.4	88.4	88.3
CP (%DM)	17.3	14.9	19.0	26.3	25.0
aNDFom %DM)	39.4	41.4	34.5	31.1	30.0
ADFom (%DM)	34.7	40.2	39.8	23.3	28.2
aD DM (%)	75.5	83.0	84.5	87.9	87.7
aD OM (%)	76.0	83.7	85.4	88.6	88.3
aD CP (%)	63.8	78.8	75.3	56.4	61.1
aD αNDFom (%)	74.5	80.4	85.7	70.8	75.4
aD ADFom (%)	74.4	77.0	79.5	72.7	72.6

#### Table 2. DM intake, faecal output and apparent digestibility (aD) in Nasalis

#### larvatus and Trachypithecus cristatus

DM, dry matter; OM, organic matter; CP, crude protein; aNDFom, neutral detergent fibre;

ADFom, acid detergent fibre



Stomach type

- Tripartite
- Quadripartite

#### Species

- ▲ Colobus guereza
- Nasalis larvatus
- ♦ Pygathrix nemaeus
- ▼ Trachypithecus cristatus
- Trachypithecus francoisi
- + Trachypithecus obscurus

Fig 1. Relationship between apparent digestibility of dry matter (DM, %), neutral detergent fibre (NDF, %) or acid detergent fibre (ADF, %) and neutral detergent fibre intake (%) in six colobine species: *Colobus guereza* [17], *Nasalis larvatus* [27],

### Pygathrix nemaeus [17], Trachypithecus cristatus [16], Trachypithecus francoisi [17,

**23] and** *Trachypithecus obscurus* **[24].** Species and stomach type, i.e. tri- or quadripartite, are indicated in different shape and colour symbols, respectively.

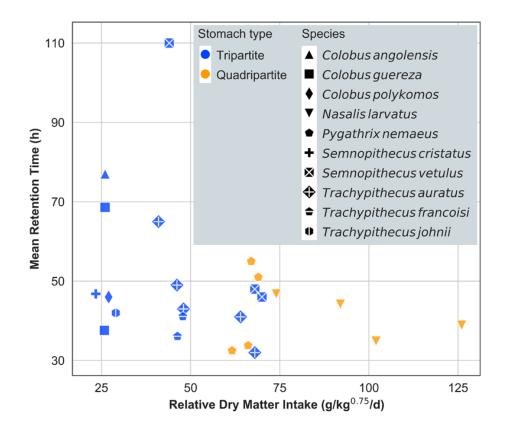


Fig 2. Relationship between mean retention time (h) and relative dry matter intake (g/kg<sup>0.75</sup>/d) in 10 colobine species: *Colobus angolensis* [47], *Colobus guereza* [17], *Nasalis larvatus* [19], *Pygathrix nemaeus* [17], *Semnopithecus cristatus* [16], *Semnopithecus vetulus* [48], *Trachypithecus auratus* [28], *Trachypithecus francoisi* [17] and *Trachypithecus johnii* [47]. Species and stomach type, i.e. tri- or quadripartite, are indicated in different shape and colour symbols, respectively.

		$2017~{ m Sep}$	(Autumn)	2018 Jar	n (Winter)	2018 Jun	(Summer)
		Offered*1	Leftover <sup>*1</sup>	$Offered^{*1}$	Leftover <sup>*1</sup>	Offered <sup>*1</sup>	Leftover <sup>*1</sup>
	Yoshino cherry (Prunus yedoensis Matumura)	$1551 \pm 120$	$996 \pm 128$	-	-	$1537 \pm 420$	$1114 \pm 287$
	Bamboo-leaf oak (Quercus myrsinifolia)	$1708 \pm 221$	$1441 \pm 190$	$2111 \pm 279$	$1930 \pm 276$	-	-
	Chinquapin ( <i>Castanopsis sieboldii</i> )	$1611 \pm 146$	$1405 \pm 124$	$969 \pm 110$	$358\pm264$	-	-
$Browse^{*2}$	Laurel ( <i>Machilus thunbergii</i> )	$1077 \pm 126$	$846 \pm 99$	$2583 \pm 426$	$2113\pm373$	$3317 \pm 931$	$2682 \pm 619$
Drowse -	Glossy privet ( <i>Ligustrum lucidum</i> )	$1339 \pm 130$	$1177 \pm 108$	$1040 \pm 73$	$894 \pm 77$	$1799 \pm 626$	$1637 \pm 595$
	Willow (Salix spp.)	$761 \pm 139$	$452 \pm 96$	-	-	-	-
	Hibiscus ( <i>Hibiscus spp.</i> )	-	-	$187 \pm 22$	$58\pm8$	-	-
	Japanese spindletree ( <i>Euonymus japonicus</i> )	-	-	$768 \pm 35$	$571 \pm 39$	$758\pm88$	382±131
Fruit	Apple	$129\pm6$	0	163±1	3±2	$167 \pm 5$	0
	Carrot	$123\pm\!2$	1±1	$125 \pm 1$	$93\pm2$	$122 \pm 13$	$14 \pm 7$
	Green bean	$124 \pm 2$	$2\pm1$	$173 \pm 3$	$101 \pm 5$	$158 \pm 2$	$9\pm2$
Vegetable	Broccoli	$108 \pm 2$	0	$148 \pm 3$	$3\pm 2$	$122 \pm 12$	0
	Asparagus	$104 \pm 1$	0	$101 \pm 1$	$4\pm1$	$77\pm8$	$2\pm2$
	Cucumber	$199 \pm 4$	$5\pm5$	121±6	3±3	$97 \pm 5$	0
Beans	Soy bean	31±1	0	41±1	0	$37\pm2$	0
Deans	Peanuts	$25\pm1$	$6\pm1$	$73\pm2$	$26 \pm 1$	$61 \pm 4$	$29 \pm 0$
Commercial product	Primate L/S biscuit banana (Mazuri)	16±1	2±0	141±2	2±1	58±0	0

\*1 Weight unit is  $X \pm SE$  g, as fed/day/animal.

 $*^{2}$  We measured the weight of the whole branch, including leaves and twigs.

#### S2 Table. Feed composition, and offered and leftover amounts (g as fed/day/two animals) in each digestion trial of two adults of

#### Trachypithecus cristatus

		2018 (Sum	0		9 Feb inter)	
		Offered <sup>*1</sup>	Leftover <sup>*1</sup>	Offered <sup>*1</sup>	Leftover <sup>*1</sup>	
$Browse^{*2}$	Bamboo-leaf oak (Quercus myrsinifolia)	300±0	$269\pm5$	300±1	$280 \pm 4$	
Fruit	Apple	$600 \pm 1$	$81 \pm 16$	$600 \pm 0$	1±1	
	Banana peel	$80 \pm 0$	$3\pm1$	$80 \pm 0$	0	
Vegetable	Carrot	$60 \pm 1$	1±1	$60 \pm 0$	0	
	Cabbage	900±0	$69 \pm 17$	$901 \pm 0$	$9\pm2$	
Starchy food	Sweet potato	$320 \pm 0$	0	$321 \pm 0$	0	
Commercial product	Leaf-eater primate diet - Mini-biscuit (Mazuri)	20±0	0	20±0	0	

\*1 Weight unit is  $X \pm SE$  g FM/day.

\*2 We measured the weight of the whole branch, including leaves and twigs.

	DM (%)	Ash (% DM)	CP (% DM)	aNDFom (% DM)	ADFom (%DM)	DMI (g DM/day)	DMI*1 (%)	DMI*2 (%)	DMI* <sup>3</sup> (g DM/kg <sup>0.75</sup> /day)
Yoshino cherry (Prunus yedoensis Matumura)	34.15	8.06	9.24	29.47	28.28	189.6	-	27.0	22.4
Bamboo-leaf oak (Quercus myrsinifolia)	41.95	8.38	10.90	49.96	36.26	111.8	-	15.9	13.2
Chinquapin (Castanopsis sieboldii)	39.39	6.65	9.96	49.47	39.36	81.0	-	11.5	9.6
Laurel (Machilus thunbergii)	36.28	5.67	8.45	53.06	46.09	83.9	-	12.0	9.9
Glossy privet (Ligustrum lucidum)	34.86	9.82	8.96	31.02	33.80	56.2	-	8.0	6.6
Willow (Salix spp.)	31.16	12.86	14.11	34.09	34.05	96.5	-	13.7	11.4
All browses	35.79	8.52	10.26	39.85	34.99	619.0	-	88.2	73.0
Apple	12.71	9.16	2.51	9.48	8.42	16.4	100.0	2.3	1.9
Carrot	5.54	18.82	10.38	19.74	21.51	6.8	99.3	1.0	0.8
Green bean	4.83	14.43	22.47	25.24	23.88	5.9	98.7	0.8	0.7
Broccoli	6.11	12.33	32.77	18.10	18.80	6.6	100.0	0.9	0.8
Asparagus	3.73	14.68	30.51	24.99	20.11	3.9	100.0	0.6	0.5
Cucumber	1.86	28.35	26.45	24.43	23.30	3.6	97.6	0.5	0.4
All fruits/vegetables	5.54	13.98	15.60	17.20	16.47	43.2	99.5	6.2	5.1
Soy bean	37.67	6.49	50.23	14.46	13.24	11.7	100.0	1.7	1.4
Peanuts	89.16	3.66	28.75	39.94	37.04	15.4	75.2	2.2	1.8
Primate L/S biscuit banana (Mazuri)	92.30	8.48	18.46	32.16	23.97	12.8	89.0	1.8	1.5
Others	64.17	6.03	29.87	29.98	25.88	39.9	85.7	5.7	4.7
Ingested diet	27.30	8.71	11.70	37.89	33.33	702.0	_	100.0	82.8

S3 Table. Nutrient contents in the feeds and DM intake of an adult male of *Nasalis larvatus* in September 2017 (autumn)

DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash; ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg)

\*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse.

\*2 DM intake of each feed/total DM intake.

\*3 BW of the proboscis monkey was 17.3 kg at the beginning of the second week of the experiment.

	DM (%)	Ash (% DM)	CP (% DM)	aNDFom (% DM)	ADFom (%DM)	DMI (g DM/day)	DMI*1 (%)	DMI*2 (%)	DMI* <sup>3</sup> (g DM/kg <sup>0.75</sup> /day)
Bamboo-leaf oak (Quercus myrsinifolia)	45.13	10.21	6.63	44.19	(70DM) 32.59	(g Divi/day) 81.7	(70) -	12.47	<u>(g Divi/kg /udy)</u> 9.0
Chinquapin ( <i>Castanopsis sieboldii</i> )	42.09	6.11	8.67	43.38	36.06	56.7	-	8.66	6.3
Laurel ( <i>Machilus thunbergii</i> )	46.24	10.19	6.63	42.12	36.08	217.4	-	33.20	24.0
Glossy privet ( <i>Ligustrum lucidum</i> )	29.32	10.15	12.98	24.50	18.40	42.8	_	6.54	4.7
Hibiscus ( <i>Hibiscus spp.</i> )	22.83	13.74	12.64	12.97	14.67	29.4	_	4.48	3.2
Japanese spindletree (Euonymus japonicus)	32.36	12.59	7.84	28.07	24.61	63.7	_	9.73	7.0
All browses	39.10	10.28	7.93	37.51	31.19	491.7	_	75.1	54.2
Apple	14.62	3.87	2.03	9.92	8.28	23.5	98.3	3.58	2.6
Carrot	7.69	8.76	9.11	13.87	15.40	2.4	25.1	0.37	0.3
Green bean	5.44	12.95	23.88	24.65	24.28	3.9	41.4	0.59	0.4
Broccoli	10.81	9.16	32.11	14.27	13.61	15.7	98.3	2.40	1.7
Asparagus	4.52	12.40	29.97	26.26	19.75	4.4	95.9	0.67	0.5
Cucumber	2.54	16.92	21.39	18.75	19.49	3.0	97.3	0.46	0.3
All fruits/vegetables	8.48	7.78	16.31	14.33	12.95	52.8	79.5	8.1	5.8
Soy bean	39.95	5.91	45.44	17.73	17.25	16.3	99.7	2.48	0.3
Peanuts	95.65	3.69	26.01	44.24	38.20	44.5	63.9	6.80	4.9
Primate L/S biscuit banana (Mazuri)*4	35.55	8.22	20.31	33.18	21.91	49.5	98.8	7.56	5.5
Others	48.69	6.05	26.32	35.37	27.80	110.3	79.0	16.8	12.2
Ingested diet	31.08	9.36	11.71	35.28	29.15	654.8	-	100.0	72.2

S4 Table. Nutrient contents in the diet and DM intake of an adult male of *Nasalis larvatus* in January 2018 (winter)

DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash; ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg)

\*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse.

\*2 DM intake of each feed/total DM intake.

- \*3 BW of the proboscis monkey was 18.9 kg.at the beginning of the second week of the experiment.
- \*4 The pelleted feed was soaked in water before feeding

	DM (%)	Ash (% DM)	CP (% DM)	aNDFom (% DM)	ADFom (%DM)	DMI (g DM/day)	DMI*1 (%)	DMI*2 (%)	DMI* <sup>3</sup> (g DM/kg <sup>0.75</sup> /day)
Yoshino cherry (Prunus yedoensis Matumura)	28.74	9.14	10.60	22.09	19.68	121.6	-	18.3	14.0
Laurel (Machilus thunbergii)	37.38	4.96	7.46	55.23	44.02	237.4	-	35.7	27.4
Glossy privet (Ligustrum lucidum)	28.66	11.90	7.65	31.00	30.49	46.4	-	7.0	5.4
Japanese spindletree (Euonymus japonicus)	29.53	8.93	9.61	33.96	23.84	110.9	-	16.7	12.8
All browses	77.73	7.42	8.68	40.68	32.73	516.4		77.7	59.6
Apple	12.39	5.54	1.50	8.29	9.42	20.7	100.0	3.1	2.4
Carrot	5.28	8.58	6.09	14.81	15.68	5.7	88.5	0.9	0.7
Green bean	6.08	8.75	18.07	24.20	24.80	9.0	94.1	1.4	1.0
Broccoli	7.13	12.58	36.77	21.18	21.01	8.7	100.0	1.3	1.0
Asparagus	5.25	10.95	39.55	24.42	18.50	3.9	97.8	0.6	0.5
Cucumber	3.00	17.55	27.53	18.35	19.10	2.9	100.0	0.4	0.3
All fruits/vegetables	7.67	8.76	15.41	15.86	16.08	50.9	97.3	7.7	5.9
Soy bean	40.97	5.86	48.24	17.64	16.13	15.3	100.0	2.3	1.8
Peanuts	88.94	3.84	27.69	33.87	29.95	28.5	52.7	4.3	3.3
Primate L/S biscuit banana (Mazuri)	92.30	8.48	18.46	32.16	23.97	53.3	100.0	8.0	6.1
Others	14.60	6.56	26.88	30.93	23.36	97.0	79.2	14.6	11.2
Ingested diet	27.23	7.40	11.85	37.35	30.09	664.3	-	100.0	76.7

S5 Table. Nutrient contents in the diet and DM intake of an adult male of *Nasalis larvatus* in June 2018 (summer)

DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash;

ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg)

\*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse.

\*2 DM intake of each feed/total DM intake.

\*3 BW of the proboscis monkey was 17.8 kg at the beginning of the second week of the experiment.

	DM (%)	Ash (% DM)	CP (% DM)	aNDFom (% DM)	ADFom (%DM)	DMI (g DM/day)	DMI*1 (%)	DMI*2 (%)	DMI* <sup>3</sup> (g DM/kg <sup>0.75</sup> /day)
Bamboo-leaf oak (Quercus myrsinifolia)	51.48	7.60	9.19	56.62	34.56	16.0	-	2.0	1.7
Apple	14.17	6.12	1.53	5.24	6.24	73.5	95.9	28.1	8.0
Banana peel	8.81	17.68	8.48	30.10	30.67	6.8	94.6	3.2	0.7
Cabbage	8.74	8.62	15.24	14.23	14.50	69.8	98.5	21.7	7.6
All fruits/vegetables	10.76	7.80	8.22	10.55	11.19	150.1	96.9	52.9	16.4
Carrot	10.69	7.59	7.23	10.34	11.27	6.3	97.0	2.0	0.7
Sweet potato	31.66	3.33	2.74	6.88	3.55	101.3	99.2	34.2	11.1
Leaf-Eater Primate Diet – Mini-Biscuit (Mazuri)	91.40	7.90	22.92	27.40	18.90	18.3	100.0	6.9	2.0
Others	31.54	4.21	5.90	10.03	6.16	126.0	99.3	43.1	13.8
Ingested diet	16.00	6.24	7.27	12.85	10.30	292.1	-	100.0	31.9

S6 Table. Nutrient contents in the diet and DM intake of two adults *Trachypithecus cristatus* in August 2018 (summer)

DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash; ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg)

\*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse.

\*2 DM intake of each feed/total DM intake.

\*3 BW of two silver lutungs were 7.6 and 7.6 kg at the beginning of the second week of the experiment.

	DM (%)	Ash (% DM)	CP (% DM)	aNDFom (% DM)	ADFom (%DM)	DMI (g DM/day)	DMI*1 (%)	DMI*2 (%)	DMI* <sup>3</sup> (g DM/kg <sup>0.75</sup> /day)
Bamboo-leaf oak (Quercus myrsinifolia)	50.65	7.64	8.67	56.74	41.46	(g Divi/day) 10.5	-	3.9	<u>(g Divi/kg / (uly)</u> 1.2
Apple	13.01	4.64	1.94	8.96	7.65	74.9	95.9	28.1	8.5
Banana peel	11.27	15.89	6.77	28.23	28.82	8.5	94.6	3.2	1.0
Cabbage	6.51	10.90	15.32	19.78	22.87	57.7	98.5	21.7	6.6
All fruits/vegetables	4.59	15.76	29.10	30.31	15.41	141.2	96.9	52.9	16.1
Carrot	9.30	9.09	5.74	14.15	15.79	5.4	97.0	2.0	0.6
Sweet potato	28.64	4.09	5.31	8.44	4.19	91.2	99.2	34.2	10.4
Leaf-Eater Primate Diet – Mini-Biscuit (Mazuri)	91.40	7.86	22.92	27.43	18.87	18.3	100.0	6.9	2.1
Others	14.48	9.85	23.46	14.16	16.27	114.9	99.3	43.1	13.1
Ingested diet	13.63	6.60	15.00	12.71	7.93	266.6	-	100.0	30.3

S7 Table. Nutrient contents in the diet and DM intake of two adults of *Trachypithecus cristatus* in February 2019 (winter)

DM, dry matter; ash, crude ash; CP, crude protein; aNDFom, neutral detergent fibre assayed with a heat stable amylase exclude residual ash;

ADFom, acid detergent fibre excluded residual ash; DMI, dry matter intake; BW, body mass (kg)

\*1 Intake/offered of each feed (DM-basis). Browses were weighed as the whole branch, including leaves and twigs, but nutrient contents were analysed only for leaves. Thus, we could not calculate offered DM amounts of browse.

\*2 DM intake of each feed/total DM intake.

\*3 BW of two silver lutungs were 6.8 and 7.7 kg at the beginning of the second week of the experiment.