Fungal taste for minerals: the ectomycorrhizal fungus *Paxillus involutus* triggers specific genes when extracting potassium from different silicates

4 Pinzari F.^{1,2,*}, Jungblut A.D.¹, Cuadros J.³

⁶ ¹Life Sciences Department, Natural History Museum, Cromwell Road, SW7 5BD London, UK

² Council of National Research of Italy (CNR), Institute for Biological Systems (IBS), Monterotondo
 (Rome), Italy.

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³ Earth Sciences Department, Natural History Museum, Cromwell Road, SW7 5BD London, UK

Corresponding Author: Flavia Pinzari, Life Sciences Department, Natural History Museum,
 Cromwell Road, SW7 5BD, United Kingdom, email: f.pinzari@nhm.ac.uk

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16 **Running title:** Transcriptomic response of ectomycorrhiza to potassium mining

18 Abstract

19 Silicates make up about 90% of the Earth's crust and constitute the main source of mineral nutrients for microorganisms and plants. Fungi can actively weather silicates to extract nutrients. However, it 20 21 is unclear whether they are able to obtain the same amounts of nutrients and use the same mechanisms 22 when tapping into different mineral sources. We performed a microcosm experiment using the ectomycorrhizal basidiomycetes Paxillus involutus and the silicates K-vermiculite, muscovite and 23 phlogopite as only potassium sources, as they show a different resistance for the removal of K cations 24 25 from the mineral structure. A combination of transcriptomic, elemental and SEM analyses showed that different minerals stimulated specific weathering mechanisms and led to a change in fungal genes 26 expression. The differential expression of the fungal genes generated alternative chemical attacks on 27 the minerals, resulting in a tailored dissolution and selective uptake of chemical elements according 28 29 to the leachability of K from the silicate mineral. The K uptake capacity of the fungus was highest with vermiculite in comparison to growth on phlogopite and muscovite. The findings provide new 30 insights into fungal-mineral interactions that will help to interpret key processes for the homeostasis 31 32 of soil environments.

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34 Keywords

35 Fungi, mineral weathering, *Paxillus involutus*, potassium, RNAseq, silicates.

37 **1. Introduction**

38 Weathering of silicates by fungi is an important process in soil formation and chemical evolution of rocks [1-3]. The ability to dissolve minerals applies to many fungi and in particular to mycorrhizal 39 fungi, which form mutual symbiotic associations with the plant root system and are able to increase 40 41 the dissolution and transformation of silicate minerals while extracting phosphorus (P), potassium (K), calcium (Ca), magnesium (Mg) and iron (Fe), especially under nutrient limitations [2]. The study 42 of the genomes of ectomycorrhizal fungi and their transcripts have shown how fungi evolved complex 43 44 mechanisms for the assimilation and exchange of carbon (C), P and nitrogen-based compounds [4). However, metal nutrients have received much less attention, and their biogeochemical routes are 45 almost unknown [2]. Free-living and mycorrhizal fungi can use a range of mineral dissolution 46 mechanisms, including acidification of the microenvironment via excretion of protons, phosphoric 47 acid, organic acids, and carbon dioxide (CO₂) [5-7]. Fungi also use the production of extracellular 48 polymeric substances to adsorb and accumulate cations and decrease the saturation for those elements 49 [8]. Exudation of organic complex-forming molecules [8-9] is as well an essential mechanism for the 50 modification of water chemistry (e.g., concentrating salts) and viscosity within biofilms that increases 51

52 water reactivity with the mineral surface [10]. In addition to multiple mechanisms, there also seem to be different intensities of mineral weathering by fungi [11]. The production of oxalic acid by a fungus, 53 54 for example, is modulated according to the type of mineral it comes into contact [12]. It is also known that there are genes that can be associated with specific weathering mechanisms and that these can be 55 regulated by the depletion of certain nutrients from the culturing media [13]. What is unclear is 56 57 whether, in light of the different structural ways in which nutrients may be part of the mineralogy of silicates, the fungi not only are able to modulate the intensity of weathering but also to implement 58 different functional strategies. The ability of fungi to change the metabolic strategy depending on the 59 type of mineral from which they derive nutrients would have implications for both the ecology and 60 evolution of fungi and fungi-plant systems [6, 14]. Fungal-derived chemical equilibria in the 61 biosphere would vary based on gene expression mechanisms and regulatory processes whose control 62 would disclose new frontiers in agriculture and soil management. In fact, a better understanding of 63 the regulation of weathering processes by fungi could improve predictions of ecosystem functions 64 [15-16]. An active fungal regulation in silicates weathering would also impact Earth system models 65 and provide new variables to the global C cycle [17-19]. 66

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This study tested the hypothesis that fungi would show distinct mechanisms of attack and differential 68 gene expression profiles when grown on specific minerals to obtain limiting nutrients such as K. The 69 70 fungus Paxillus involutus was selected for the experiments as it is a Basidiomycetes of global significance [20]. It can form mycorrhizae with many species of trees [21] but is capable also of 71 72 growing in axenic culture [22] and can weather minerals [12, 23-25]. P. involutus was grown in 73 microcosm experiments under three different conditions where the only sources of K were the minerals phlogopite, K-vermiculite and muscovite, and compared with a positive control with K 74 available in the media and a negative control without any K. These silicates have similar metal 75 76 nutrient contents but have different degrees of weatherability, which could trigger a specific fungal strategy to obtain nutrients and reveal a differential genetic expression. The growth and response of 77 the fungi were evaluated using RNAseq-based differential gene expression. The mycelium was 78 analysed to evaluate the uptake of nutrients using inductively coupled plasma mass spectroscopy 79 (ICP-MS), and the minerals were analysed to detect traces of weathering and patterns of interaction 80 with the fungal hyphae by means of scanning electron microscopy (SEM). 81 82

83 **2. Materials and Methods**

85 2.1 Fungal material and experimental set-up

86 Cultures of P. involutus (Batsch) Fr. (strain ATCC 200175, www.atcc.org) (Basidiomycota, Boletales) were reactivated and maintained aseptically on Modified Melin–Norkrans (MMN) agar 87 (Supplementary Materials S1). The purity and identity of P.involutus ATCC 200175 strain was 88 verified by Sanger sequencing (Supplementary Materials S2). The microcosms were prepared 89 according to Wei et al. [26] with some modifications (Supplementary Materials S3). Three different 90 minerals with decreasing level of K leachability were used: muscovite, phlogopite, K-vermiculite. 91 92 Their structural formulas are specified in Supplementary Materials S4. All cultures were kept in the dark at 25°C throughout the experiment. The microcosms were incubated for 21 days. Ten biological 93 replicates from each experiment were used for RNA extraction, five from each experiment were 94 95 dismounted and used for ICP-MS analysis, and the corresponding minerals were used in SEM analysis. After the experiments, fungal biomass was determined by weight using five microcosms 96 from each experiment (dry weight, 50°C, 48 h). The pH of the agar was measured at the beginning 97 and the end of the experiments (Electrode n.662-1164, VWR International Ltd, Leicestershire, U.K.). 98

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100 2.2. Microscopic and chemical analysis of fungal biomass and minerals

101 Mineral samples were analysed before and after the experiment using SEM as described in 102 Supplementary Materials S5. Groups of five replicates from each experiment were used to analyse sodium (Na), Mg, aluminium (Al), Fe, P, K, and Ca in the fungal mycelium by digestion and analysis
by ICP-MS (Agilent Technologies 7700), as described in Supplementary Materials S6.

105106 2.3 RNA extraction and libraries preparation

At the end of the experiment, fungal biomass was removed from the cellophane membrane and ground into a powder in liquid nitrogen. mRNA was extracted using the RNAeasy® PlantMini Kit (Qiagen, Manchester, U.K.) and libraries prepared using TruSeq Stranded mRNA Kit (Illumina, Cambridge, UK) (Supplementary Materials S7 and 8). Libraries (5 biological replicates for each experiment, for a total of 25 samples) were sequenced with Illumina NovaSeq 6000 (2 x 100 bp readlength) by the CeGAT Company (Germany).

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114 2.4 Bioinformatic analysis

Full details on the sequence processing are provided in Supplementary Materials S9. Trimmed reads 115 were aligned to the *Paxillus involutus* reference genome (GCA 000827475.1) using STAR (v2.5.2b) 116 [27]. A count table was created using the HTSeq package [28] implemented in OmicBox (version 117 1.2.4) [29-30]. EdgeR version 2.12 [ref. 31] was used to perform pairwise differential expression 118 analysis (DEA) between pairs of experimental conditions (5 replicates per experimental condition). 119 Functional annotation of differentially expressed genes (DEGs) was based on *P.involutus* annotation 120 from the EnsemblFungi server (http://fungi.ensembl.org) with assembly accession number 121 GCA_000827475.1 [ref. 21], and by using the BLASTx algorithm within OmicsBox version 1.2.4 122 123 (BioBam) [29-30]. Functional annotation of sequences was further achieved using InterProScan [29-30, 31] and EggNOG-mapper [32]. Gene Set Enrichment Analysis (GSEA) was used to determine 124 whether an a priori defined set of genes showed statistically significant, concordant differences 125 between two biological states [34]. The normalised enrichment score (NES) was used to compare 126 analysis results across gene sets. The false discovery rate (FDR), was used as an estimate of the 127 probability that a gene set with a given NES represented a false positive finding. FDR cut-off was 128 129 fixed at 5%.

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131 **3. Results**

132 3.1 Fungal growth

P. involutus mycelium grew in all experiments showing good replicability of the in-vitro system 133 134 utilised (Fig. S1). The pH of the agar decreased in all the experiments from its initial value of 4.7. Table S1 shows the pH values in the five experiments at the end of the incubation, with a higher pH 135 in the positive control (Cp) than in the microcosms with the minerals. The pH values in the treatments 136 137 with minerals were significantly different from the positive control, whereas the pH values in the negative control (Cm) were not significantly different from the positive control or the experiments 138 with the minerals. The comparison of the fungal biomass content obtained in the five experiments is 139 shown in Table S2. The wet fungal biomass was significantly higher in the positive control, followed 140 by the experimental treatment with phlogopite, while significantly lower values were measured in the 141 K-vermiculite, muscovite and negative control experiments. 142

144 3.2 Fungal weathering of minerals: ICP-MS results

The fungus absorbed more K from K-vermiculite than from the other two minerals (Table 1). There 145 146 was a trend indicating a slightly higher concentration of K in experiments run with phlogopite than muscovite (Table 1), but the K content of mycelium grown in the presence of muscovite and 147 phlogopite was not significantly different from that recorded in the negative control. Table 2 shows 148 the Pearson correlation coefficients (r) calculated between the mycelial concentrations of the 149 elements. The coefficient values showed that the absorption of K was positively correlated with P, 150 Ca and Mg, while it was negatively correlated with the absorption of Na. Moreover, Fe and Al showed 151 a strong positive correlation (r=0.99). 152

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154 3.3 Scanning Electron Microscopy of mineral surfaces

The SEM imaging showed that the three minerals were altered by the growth of the fungus during 155 the microcosm experiments. Comparing SEM images of minerals before and after the experiments 156 suggested that *P. involutus* when forced to grow under limited availability of K, produced different 157 effects in each silicate mineral. In muscovite, the surface of the crystal appeared eroded and uneven 158 (Fig. 1A). At higher magnification, alterations were observed at the contours of the fungal hyphae 159 consisting in the localised removal of surface layers of the mineral (Fig. 1B). In phlogopite, large 160 portions of mineral layers were removed (Fig. 1C). In the high-resolution images, the surface of the 161 phlogopite crystals was fragmented and furrowed, and characteristic depositions arranged in small 162 regular clusters were observed between the cracks (Fig. 1D). Fungal mycelium appeared singularly 163 adherent to the crystal when grown on the K-vermiculite. Here, forms of surface erosion were also 164 visible at low magnification suggesting that the fungus exerted a mechanical action (Fig. 1E). 165 However, the waxy consistency of the superficial layer of K-vermiculite, at times excavated and 166 raised to form small accumulations of material, suggests that a chemical action also took place to 167 make an initially intact crystalline structure malleable. High magnification images of K-vermiculite 168 169 surface show some hyphae flattened and others filled with precipitated material (Fig. 1F).

171 3.4 Differential expression of P. involutus during K weathering on K-vermiculite, muscovite and 172 phologpite minerals.

The transcriptome sequencing yielded 966.96 million reads, with a total number of bases (in Gb) of 173 194.89, an average GC content of trimmed FASTQ reads (average of all samples) of 53.0% to 54.0% 174 and a Q30 quality score threshold value between 91.3% and 93.07% (Table S3). Multidimensional 175 scaling analysis (MDS) (Fig. 2) [35] was performed to evaluate the dissimilarity between 176 transcriptomes of *P.involutus* obtained in response to its growth on K-vermiculite, phlogopite and 177 muscovite and in comparison to the negative and positive controls. In the two-dimensional scatterplot 178 (Fig. 2) the distances represent the log2 fold change (log2 FC) between samples. The transcriptomes 179 obtained with minerals were different from the positive control. The transcriptome from the K-180 vermiculite microcosm showed a high similarity between replicates and grouped separately from all 181 the other experiments. The transcriptomes from phlogopite, muscovite and the negative control 182 overlapped and showed a general higher variability compared to positive control and K-vermiculite 183 treatment. 184

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From the heatmap (Fig. 3) emerges that there are two significant clusters of genes with distinct 186 regulation for the positive and negative controls. The fungus in the presence of K-vermiculite showed 187 a gene expression partially similar to the positive control. With muscovite and phlogopite the gene 188 expression was similar to the negative control. The number of genes differentially expressed by the 189 fungus in each comparison are shown in Table 3. Overall, the number of genes differentially 190 191 expressed between the experiments with the minerals and the negative control is lower than the number of genes obtained in the contrasts against the positive control. The similarity between the 192 condition of K deprivation and that of bioavailability conditioned by the mineral source is also shown 193 by the Volcano plots that display both the genes differentially expressed (FDR < 0.05) and those that 194 did not show a change in their expression level between contrasting conditions (Figure S2). The 195 number of significantly up and down-regulated genes (FDR < 0.05) between the experiments and 196 197 controls is summarised in Table 3. In the pairwise comparison between transcriptomes of P. involutus grown on the three minerals, and the negative control, the number of differentially expressed genes 198 decreases from K-vermiculite (439) to phlogopite (47) and muscovite (29). There were very few 199 DEGs between muscovite and negative control, which indicates that the physiological state of the 200 fungus in the presence of muscovite was similar to being under the condition of K deprivation. 201 202

Fig. 4 and Supplementary Spreadsheet 1 show the genes that the fungus either triggered or repressed at the presence of the three minerals and the positive control, compared to the negative control based on FDR values and log2FC scores. The differential expression between positive and negative control
 identified marked differences between the condition of total bioavailability of K and its unavailability,
 with genes showing up to 6 log2FC values. However, the top genes were assigned as hypothetical
 proteins without associated specific function.

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210 3.5 Gene set enrichment analysis (GSEA)

The genes that were identified as differentially expressed between pairs of treatments were grouped 211 by their involvement in biological processes, molecular functions and cellular compartments. In Fig. 212 5 and Supplementary Spreadsheet 2 are shown the normalised enrichment score values (NES) 213 obtained for comparisons between the experiments in which the fungus grew in the presence of the 214 three minerals (V, P and M) compared to the negative control condition (Cm). Among the molecular 215 functions stimulated in the experiments with phlogopite there were carboxylic-acid metabolic process 216 (GO:0019752), organic-acid metabolic process (GO:0006082), oxoacid metabolic process 217 (GO:0043436) and lipid metabolic process (GO:0006629). The molecular functions stimulated in the 218 case of muscovite were different and significantly fewer, compared to the other two minerals. The 219 220 only molecular functions overexpressed in P.involutus when incubated with muscovite were monooxygenase activity (GO:0004497) and oxidoreductase activity (GO:0016705-MF). 221 Interestingly, the biological processes that were under-expressed (compared to the negative control 222 223 condition) were response to heat (GO:0009408), cellular response to heat (GO:0034605) and ribonucleoprotein complex subunit organisation (GO:0071826). There were also differences 224 225 observed in the gene assignment for the fungus grown with K-vermiculite as the only source of K. In 226 this condition, both molecular functions and biological processes were associated with the cytoskeleton, and significant overexpression of gene sets related to cytoskeletal protein binding 227 (GO:0008092) and cytoskeleton organisation (GO:0007010). Moreover, with K-vermiculite, 228 catabolic process (GO:0009056), vesicle-mediated transport (GO:0016192), and cellular response to 229 stimulus (GO:0051716 i.e. processes resulting in a change in state or activity of a cell) were also 230 231 overexpressed.

233 **4. Discussion**

The present study confirmed that *Paxillus involutus* is able to weather K from silicates such as Kvermiculite, muscovite, and phlogopite, but we also showed for the first time that the gene expression patterns and metabolic response differed according to K leachability from different types of silicate minerals. Differences in chemical and physical attack by the fungus on the three minerals were also documented by chemical and imaging analyses.

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240 *4.1 Effects on pH and uptake of K and other mineral nutrients*

The pH of the agar under the mycelium decreased significantly in all K-depletion treatments, which 241 is likely due to secretion of acids by the fungus in response to either the absence of K or the presence 242 of K containing minerals in the agar. This stimulation of acid secretion appeared stronger in the 243 presence of minerals than in the negative control, which reinforces the idea that it is not the absence 244 of K, but the presence of minerals containing it, that stimulated the secretion of acids into the culture 245 medium. K concentration in the mycelium was negatively correlated with Na concentration, but 246 positively correlated with P. There was no clear correlation observed between Mg and Ca. The 247 248 negative correlation of K with Na was likely due to the use of Na as a substitute for K by the fungus. This can be explained by K being the primary inorganic cation in the fungal cell cytoplasm and an 249 essential macro-element that is accumulated against a transmembrane concentration gradient. If the 250 K deficiency occurs in the presence of Na, fungal cells can take Na+ instead of H+ as a substitute for 251 the missing K [36]. While fungi can utilise Na to grow in the absence of K, Na is also a toxic element 252 in excessive quantities [37]. Thus, the recorded changes in gene expression that depended on K 253 limitation could have been linked to functional modifications needed at membrane level to assimilate 254 255 Na instead of K, and mechanisms to react to Na toxicity [38].

257 We also observed a positive correlation of K and P, which could be due either to differential P uptake, or to differential P excretion. The findings agree with previous studies, where a strong correlation 258 between P and K cellular levels was observed for the ectomycorrhizal fungus Hebeloma 259 cylindrosporum [39]. Jentschke et al. [40] found that the fungus P. involutus translocated K and Mg 260 to Norway spruce plants, with the translocation of these elements depending on their coupling with P 261 uptake during the process. Our findings also suggest that K was one of the major counter-ions of 262 polyphosphate (polyP) granules, especially of soluble polyP short-chains, mainly located in fungal 263 vacuoles [41]. K and P have been shown to be located in the same vacuolar fungal compartments of P. 264 involutus [42-43]. P uptake and polyphosphates synthesis and accumulation could result in the 265 accumulation of a high negative charge in the cell, which results in the activation of cation import 266 mechanism for maintaining overall cellular charge neutrality. Therefore, K is both an element directly 267 involved in fungal nutrition, but also a component necessary for cell homeostasis and efficient uptake and 268 storage of inorganic phosphate. Although the fungus grew in K-depleted experiments and P was not 269 limiting, less P was assimilated than in the positive control. This indicates that the limitation in the 270 271 bioavailability of K could have affected P assimilation.

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The SEM imaging allowed to document visible effects of metabolic action by the fungus on minerals 273 274 at microscale level. The K extraction led to the formation of furrows, cracks, swelling, erosions and bio-precipitation of material on crystals. Some of the effects are due to the production of organic 275 acids, the adhesion of hyphae and leaching of elements other than K. For example, the mycelium 276 277 grown in the presence of phlogopite showed significantly higher assimilation of Fe and Al than the other experiments. Ectomycorrhizal fungi can assimilate Fe [44] and are also capable of absorbing 278 Al [45]. Similarly, Bonneville et al. [44] showed that Paxillus involutus could oxidise Fe(II) to Fe(III) 279 in biotite, increasing with this process the mineral weathering. The authors suggested that Fe(II) 280 oxidation induces the loss of the newly formed Fe(III) from the octahedral sheets and formation of 281 amorphous Fe oxy-hydroxides within the silicate layers, which induces volume increase, strain in the 282 biotite lattice and the formation of fractures. 283

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4.2 Differential gene expression of P. involutus as a response to growth on silicates with variable K leachability

287 The gene expression profiles by P. involutus were distinct for mycelium grown on K-vermiculite in comparison to muscovite and phlogopite as well as to controls. The muscovite and phlogopite 288 experiments had similar profiles and overlapped with the negative control. The differential expression 289 290 of genes matched to the concentration of K found in the mycelium after the experiments, and there was a positive relationship between the accessibility of the element and its absorption by the fungus. 291 This suggested the fungus modulated the gene expression according to both the amount of K that it 292 293 was able to assimilate from the silicates but also reflected the strength with which the three minerals retained the cation. As the three minerals showed some variation in the weathering of other elements 294 such as Fe, Mg, Na and Al, the gene expression profile might also be affected by other factors than 295 296 leachability of K from the respective mineral, including toxicity of released metals, and solubility of the other cations in the growth environment [46-47]. 297

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299 In the presence of K-vermiculite, the fungus was able to derive the K by its displacement from the interlayer sites of vermiculite by other cations in the medium, which is an exchenge mechanism 300 similar to how fungi obtain minerals in soil environments [48]. This suggests that the fungus was not 301 subjected to the same level of K deprivation as with the other two minerals. However, it still had to 302 activate different mechanisms for the assimilation of K than under condition of its unlimited 303 bioavailability (in positive control). In the case of muscovite, which is the mineral most strongly 304 retaining K, the gene expression profiles suggested a response similar to growth in the absence of K. 305 There were only 21 genes up- and 8 down-regulated by the fungus in the presence muscovite in 306

comparison to the negative control, moreover, only a few of these were found to be differentially 307 expressed for more than 2 logFC. In terms of K effectively being assimilated by the fungus, the 308 difference corresponds to about one microgram per dry gram of mycelium. These few genes are 309 therefore those that separate the condition of total K depletion from that of a minimum bioavailability. 310 The most differentially expressed genes (downregulated more than 2 logFC in the presence of 311 muscovite, compared to negative control) were a glycoside hydrolase family 79 protein from the 312 membrane (PAXINDRAFT 153975, GO:0016020), a hypothetical protein associated to the nucleus, 313 involved in regulation of transcription by RNA polymerase II and with DNA-binding transcription 314 factor activity (PAXINDRAFT_176752, P:GO:0006357; F:GO:0000981; F:GO:0008270) and an 315 RnaseH-domain-containing protein, with RNA phosphodiester bond hydrolytic function, binding 316 nucleic acids (PAXINDRAFT 91567, P:GO:0090502; F:GO:0003676; F:GO:0004523). The 317 glycoside hydrolase family 79 protein includes endo-beta-N-glucuronidase (EC 3.2.1.31) and 318 enzymes with β -glucanase activity, that in fungi are thought to be necessary in morphogenetic events 319 that require controlled hydrolysis of the cell wall [49]. 320

Comparison of positive and negative control conditions showed that more genes were up regulated 321 322 under unlimited K bioavailability than under stress conditions of K depletion. Which corresponds to a large down-regulation of genes in the negative control. This can be read as a response to stress with 323 324 the deactivation of functions in order to overcome the nutrients limitation. Still, a significant down regulation of genes in the positive control microcosms experiments was also observed, indicating the 325 existence of a number of mechanisms that have evolved to respond to the absence of K in the 326 327 environment. However, the fungus still grew under K-depleted conditions as the dry weight of the mycelium in the negative control was not significantly different to biomass in the positive control. 328 This could be due to the increased uptake of Na to replace K as cation, as Na content of the mycelium 329 in the negative control was at least seven times that of the positive control. The change in gene 330 expression triggered by K limitation could have therefore been linked to functional modifications 331 needed at membrane level to assimilate Na instead of K, and to limit any toxic effects by the increased 332 uptake of Na in to the cells. Similar mechanisms of reaction of P. involutus to limiting nutritional 333 conditions have been observed by Paparokidou et al. [50] who showed that inorganic P starvation 334 induces in the fungus expression of newly identified putative high-affinity Pi transporter genes, while 335 reducing the expression of putative organic acid transporters. 336 337

4.3 Differences in functional genes and processes for K leaching from silicates

Phlogopite, K-vermiculite and muscovite led to distinct transcriptional responses in the fungus, 339 suggesting some mechanism by the fungus to sense and respond to its mineral surroundings. Gene 340 expression of the fungus in the presence of K-vermiculite compared to the condition of total K 341 depletion highlighted the overexpression of genes that interact with cytoskeleton components which 342 have a role in the assembly or disassembly of cytoskeletal structures such as actin. They are also 343 involved in vesicle-mediated transport, and suggests that adhesion mechanisms could have had a role 344 in K uptake. Indeed, fungi, in addition to dissolving the minerals with the production of acidic 345 substances, have shown in some studies to carry out physical attacks on minerals through hyphal 346 adhesion. It has been shown for example that the release of K was enhanced by a factor of 3-4 by 347 direct contact between K-feldspar and illite surfaces and the fungus [51]. Attachment of fungi to the 348 mineral surfaces has also been shown to cause a more efficient release of elements from biotite [52-349 350 53]. Physical mechanisms of action may be therefore particularly efficient for some cations such K.

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Phlogopite triggered the overexpression of genes associated with the chemical reactions and pathways involving carboxylic acids, organic acids and also oxoacid metabolic process. An oxoacid is a compound which contains oxygen and which produces a conjugate base by loss of positive hydrogen ion(s). Many organic acids, like carboxylic acids, are oxoacids. Van Schöll et al. [25] showed how ectomycorrhizal fungi lacking mineral nutrients like P, Mg, and K might modify the composition of organic acids they exude. In other fungi, oxalate exudation was found to be higher when the mycelium

was in contact with rock grains containing K [54]. Furthermore, the biological processes that were 358 stimulated by phlogopite involved the hydrolytic activity linked to P-containing anhydrides for the 359 catalytic reaction that produces nucleoside diphosphate and a phosphate from nucleoside 360 triphosphate. There are the adenosine triphosphatases (ATPases) that generate electrical and chemical 361 ion gradients across membranes and transporters for a wide range of solutes across membranes. 362 Another gene up-regulated by the fungus in the presence of phlogopite, compared to the negative 363 control, is a phosphoglycerate mutase-like protein, which was annotated as an integral component of 364 the plasma membrane. The top up-regulated gene in K-vermiculite was also a hypothetical protein 365 that is located in the membrane and is involved in ATP-coupled electron transport and electron 366 transfer activity. This highlights the involvement in transport mechanisms during leaching and uptake 367 of K from silicates. 368

Monooxygenase and oxidoreductase activities were the only molecular functions significantly 370 overexpressed as gene sets in the presence of muscovite. Fungi produce heme-containing peroxidases 371 and peroxygenases, flavin-containing oxidases, and different copper-containing oxidoreductases 372 373 [55]. Moreover, in some basidiomycetes unspecific peroxygenases can catalyse a variety of monooxygenation reactions with H₂O₂ as the source of oxygen and final electron acceptor. Oxidation 374 processes are more energy-demanding than acid production, and therefore they might have been used 375 376 by the fungus after failure to leach K under acidic conditions, as suggested by the low uptake of K in the mycelium and the limited uptake of Al, compared to phlogopite, and by the evidence that fungal 377 weathering of Al from muscovite occur by releasing organic acids [56]. Xiao et al. [57] and Wang et 378 379 al. [58] suggested that genes encoding multicopper oxidase have a significant role in the weathering of K-bearing silicates in Aspergillus spp. Our study identified a gene encoding for di-copper centre-380 containing protein with oxidoreductase and metal ion binding function as up-regulated by the fungus 381 grown with muscovite. Wang et al. [59] also showed with an electrochemical experiment that 382 multicopper oxidases could enhance electrons transfer during fungal weathering of K-bearing 383 silicates, promoting electron transport at the edge of the minerals. 384

385 There are four families of putative K transport systems identified in ectomycorrhizal fungi [60]. HAK 386 and Trk transporters, TOK and SKC channels, the latter representing voltage-dependent K-selective 387 channels. In particular, Trk transporters are membrane proteins involved in K uptake that are 388 389 widespread among fungi [61]. HcTrk1, one member of these transporters that are present in some ectomycorrhizal fungi [62] was described as a Na+ /K+ transporter. However, in P. involutus none 390 of the genes belonging to K transporters (HAK and Trk transporters, or to TOK and SKC channels) 391 392 were significantly up-regulated in the microcosms with the minerals compared to the negative control. In contrast, several genes associated with transport systems were down-regulated in the mycelia 393 grown with K-vermiculite compared to negative control, including a P-loop containing nucleoside 394 triphosphate hydrolase protein associated to ATPase-coupled transmembrane transporter. There was 395 also a gene down-regulated in the muscovite experiment that is associated with a general substrate 396 transmembrane transporter (major facilitator superfamily MFS), and an integral component of the 397 398 plasma membrane. The MFS are membrane proteins that import and export target substrates such as metabolites, oligosaccharides, amino acids and oxyanions. They utilise the electrochemical gradient 399 of the target substrate (uniporter), or act as a co-transporter where transport is coupled to the 400 401 movement of a second substrate.

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The presented study is based on laboratory experiments, however the fungus *P. involutus* is abundant in the environment and forms mycorrhizae with many tree species. Many soil environments can be cation-poor and therefore the ability of *P. involutus* to access K from minerals would give an ecological advantage. Therefore, the fungi might have even evolved specific cellular and metabolic mechanisms [48, 63]. An adaptation of the fungi to low K conditions could help to explain the observed down regulation of many genes in the positive control microcosms experiments. The down regulated genes under unlimited K conditions were predicted to be linked to fundamental fungal
biological processes, such as "translation", "carbohydrate metabolic processes" or "ribosome
biogenesis". A large set of genes (over 200) associated to the "mitochondrion" cellular components
were also significantly down-regulated. This suggest that the fungi might had a lower energy
expenditure and cellular activity in the presence readily bioavailable K.

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415 **5. Conclusions**

The geo-biological cycles are governed by complex interactions and feedback mechanisms between 416 living organisms and rocks. Filamentous fungi play a key role as they collaborate symbiotically with 417 plants, decompose and recycle organic matter and extract nutrients from minerals. One of the 418 fundamental nutrients for growth is K, and the global transcriptome analysis showed that Paxillus 419 involutus could switch on or off different genes and metabolic pathways depending on the minerals 420 from which it is forced to obtain K. The differential expression of the fungal genes generated 421 alternative chemical attacks on the minerals, resulting in a tailored dissolution and selective uptake 422 of chemical elements. The fungus also showed an excellent exchange capacity with K-vermiculite, 423 424 which appeared to create a different condition from that represented by phlogopite and muscovite. Understanding the strategy deployed by microorganisms to obtain mineral nutrients under different 425 426 conditions provides new insights for interpreting the processes on which biodiversity and the 427 homeostasis of soil environments depend.

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429 **6. Supplementary material**

Supplementary Methods: S1, MMN complete medium composition; S2, Confirmation of purity and
identity of *P. involutus* strain; S3, Microcosm experiment; S4. Mineral substrates used for fungal
weathering experiments; S5, SEM of mineral surfaces; S6, ICP-MS of mycelium; S7, RNA
extraction; S8, Library preparation using TruSeq Stranded mRNA (Illumina) kit; S9, Bioinformatic
analysis; S10, Statistical tests.

- 436 Supplementary Tables: Supplementary Table S1, pH of agar under the mycelium; Supplementary
 437 Table S2, Average fungal biomass; Supplementary Table S3, Results of the sequencing of fungal
 438 mRNA.
- *Supplementary Figures:* Supplementary Figure S1, Diagram showing the setup of the experiment;
 Supplementary Figure S2, Volcano plots
- 441 Supplementary Spreadsheets: Supplementary Spreadsheet 1, Excel file containing the list of top up-
- regulated and down-regulated genes; Supplementary Spreadsheet 2, Excel file containing the list of
 GSEA results.
- 444

445 Availability of data and material (data transparency)

The datasets generated during and analysed during the current study are available in the Geo Submission Omnibus (GEO) public database (geo@ncbi.nlm.nih.gov) with the accession number GSE158973.

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460 **Conflicts of interest/Competing interests**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

463464 Authors' contributions

- 465 F.P. conceived the design of the study, conducted molecular analyses, analysed and interpreted
- transcriptomic data and wrote the manuscript. A.J. conceived the design of the study, supervised themolecular and genetic part of project, and wrote the manuscript. J.C. conceived the design of the
- study, supervised the geochemical aspects of the project and wrote the manuscript.
- 469

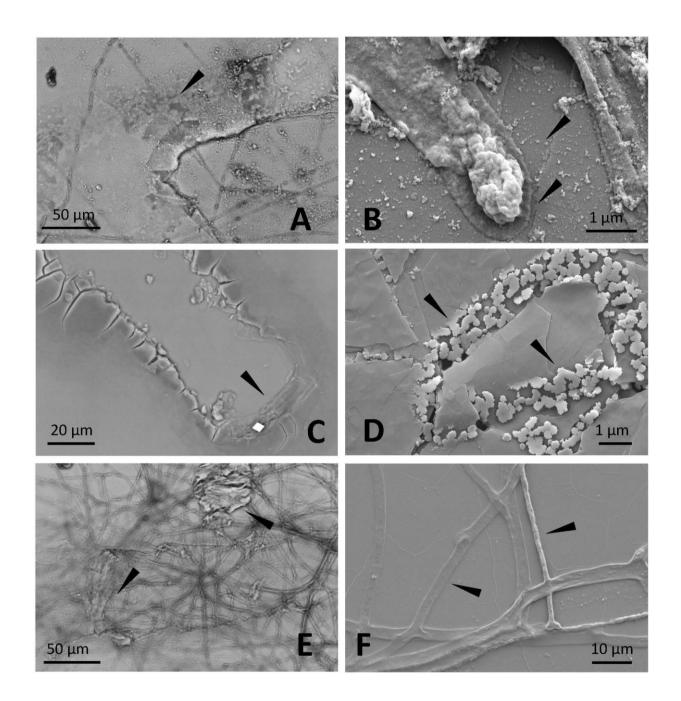
470 **References**

- [1] Hoffland E, Kuyper TW, Wallander H, Plassard C, Gorbushina AA, Haselwandter K, *et al.* The
 role of fungi in weathering. Front Ecol Environ. 2004; 2: 258–264.
- [2] Cuadros J. Clay minerals interaction with microorganisms: A review. Clay Miner. 2017; 52(2):
 235-261.
- [3] Fomina M, Skorochod I. Microbial Interaction with Clay Minerals and Its Environmental and
 Biotechnological Implications. *Minerals*. 2020; *10(10)*: 861.
- 477 [4] van der Heijden MGA, Martin FM, Selosse M-A, Sanders IR. Mycorrhizal ecology and evolution:
 478 the past, the present, and the future. New Phytol. 2015; 205: 1406–1423.
- [5] Rosling A, Lindahl BD, Taylor AFS, Finlay RD. Mycelial growth and substrate acidification of
 ectomycorrhizal fungi in response to different minerals. FEMS Microbiol Ecol. 2004; 47(1): 31–
 37. https://doi.org/10.1016/S0168-6496(03)00222-8.
- [6] Fomina M, Burford EP, Gadd G. Fungal dissolution and transformation of minerals: significance
 for nutrient and metal mobility. In: Gadd GM (ed.). Fungi in Biogeochemical Cycles. (Cambridge
 University Press, Cambridge) (ISBN: 978-0-521-84579-3). 2006. pp 236–266.
- [7] Balogh-Brunstad Z, Kent Keller C, Thomas Dickinson J, Stevens F, Li C, Bormann BT. Biotite
 weathering and nutrient uptake by ectomycorrhizal fungus, *Suillus tomentosus*, in liquid-culture
 experiments. Geochim Cosmochim Acta. 2008; 72: 2601–2618.
- [8] Bray AW, Oelkers EH, Bonneville S, Wolff-Boenisch D, Potts NJ, Fones GR, Benning LG. The
 effect of pH, grain size, and organic ligands on biotite weathering rates. Geochim Cosmochim
 Acta. 2015; 164: 127–145.
- [9] Adeyemi AO, Gadd GM. Fungal degradation of calcium-, lead- and silicon-bearing minerals.
 Biometals. 2005; 18: 269–281.
- [10] Cuadros J, Afsin B, Jadubansa P, Ardakani M, Ascaso C, Wierzchos J. Pathways of volcanic
 glass alteration in laboratory experiments through inorganic and microbially-mediated processes.
 Clay Miner. 2013; 48: 423–445.
- [11] Pinzari F, Cuadros J, Napoli R, Canfora L, Baussà Bardají D. Routes of phlogopite weathering
 by three fungal strains. Fungal Biol. 2016; 120(12): 1582–1599.
- [12] Schmalenberger A, Duran AL, Bray AW, Bridge J, Bonneville S, Benning LG, *et al.* Oxalate
 secretion by ectomycorrhizal *Paxillus involutus* is mineral-specific and controls calcium
 weathering from minerals. Sci Rep; 2015: 5: 1–14.
- [13] Xiao B, Lian B, Sun L, Shao W. Gene transcription response to weathering of K-bearing minerals
 by *Aspergillus fumigatus*. Chem Geol. 2012; 306:1–9.
- [14] Naranjo-Ortiz MA, Gabaldón T. Fungal evolution: major ecological adaptations and
 evolutionary transitions. Biol Rev Camb Philos Soc. 2019: 94(4): 1443–1476.
- [15] Finlay RD, Mahmood S, Rosenstock N, Bolou-Bi EB, Köhler SJ, Fahad Z, *et al.* Reviews and
 syntheses: Biological weathering and its consequences at different spatial levels from nanoscale
 to global scale. Biogeosciences. 2020; 17: 1507–1533.
- [16] Hu A, Wang J, Sun H, Niu B, Si G, Wang J. *et al.* Mountain biodiversity and ecosystem
 functions: interplay between geology and contemporary environments. ISME J. 2020; 14: 931–
 944. <u>https://doi.org/10.1038/s41396-019-0574-x</u>

- [17] Penman DE, Caves Rugenstein JK, Ibarra DE, Winnick MJ. Silicate weathering as a feedback
 and forcing in Earth's climate and carbon cycle. Earth-Science Reviews. 2020; 209: 103298,
- 513 [18] Treseder KK, Lennon JT. Fungal traits that drive ecosystem dynamics on land. Microbiol Mol
 514 Biol Rev. 2015; 79(2): 243–262.
- [19] Zanne A, Abarenkov K, Afkhami M, Aguilar-Trigueros CA, Bates S, Bhatnagar JM, *et al.*Fungal functional ecology: bringing a trait-based approach to plant-associated fungi. Biol Rev.
 2020; 95(2): 409–433.
- [20] Jargeat P, Chaumeton JP, Navaud O, Vizzini A, Gryta H. The *Paxillus involutus* (Boletales,
 Paxillaceae) complex in Europe: genetic diversity and morphological description of the new
 species *Paxillus cuprinus*, typification of *P. involutus* s.s., and synthesis of species boundaries.
 Fungal Biol. 2014; 118(1): 12–31. doi: 10.1016/j.funbio.2013.10.008.
- [21] Kohler A, Kuo A, Nagy LG, Morin E, Barry KW, Buscot F, *et al.* Convergent losses of decay
 mechanisms and rapid turnover of symbiosis genes in mycorrhizal mutualists. Nat Genet. 2015;
 47(4): 410–415. doi: 10.1038/ng.3223.
- [22] Lapeyrie F, Chilvers GA, Bhem CA. Oxalic acid synthesis by the mycorrhizal fungus *Paxillus involutus* (Batsch EX FR) F.R. New Phytol. 1987; 106:139–146.
- 527 [23] Smits MM, Bonneville S, Benning LG, Banwart SA, Leake JR. Plant-driven weathering of
 528 apatite the role of an ectomycorrhizal fungus. Geobiology. 2012; 10: 445–456.
- [24] Saccone L, Gazze S, Duran A, Leake J, Banwart S, Ragnarsdottir K, *et al.*, High resolution
 characterisation of ectomycorrhizal fungal-mineral interactions in axenic microcosm
 experiments. Biogeochemistry. 2012; 111: 411–425. doi:10.1007/s10533-011-9667-y.
- [25] van Schöll L, Smits MM, Hoffland E. Ectomycorrhizal weathering of the soil minerals muscovite
 and hornblende. New Phytol. 2006; 171(4): 805–813.
- [26] Wei Z, Kierans M, Gadd G. A Model Sheet Mineral System to Study Fungal Bioweathering of
 Mica. Geomicrobiol J. 2012; 29: 323–331.
- [27] Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S *et al.* STAR: ultrafast universal
 RNA-seq aligner. Bioinformatics. 2013; 29 (1): 15–21.
- [28] Anders S, Pyl PT, Huber W, HTSeq—a Python framework to work with high-throughput
 sequencing data. Bioinformatics. 2015; 31(2): 166–169.
- [29] Conesa A, Götz S, Garcia-Gomez JM, Terol J, Talon M, Robles M. Blast2GO: a universal tool
 for annotation, visualisation and analysis in functional genomics research. Bioinformatics. 2005;
 21: 3674–3676.
- [30] Conesa A, Götz S. Blast2GO: A Comprehensive Suite for Functional Analysis in Plant
 Genomics. Int J Plant Genomics. 2008; 1–13. 2008: 619832. doi: 10.1155/2008/619832.
- [31] Robinson MD., McCarthy DJ. and Smyth GK. (2010). edgeR: a Bioconductor package for
 differential expression analysis of digital gene expression data. Bioinformatics (Oxford, England).
 2010; 26(1): 139–140.
- [32] Labarga A, Valentin F, Anderson M, Lopez R. Web Services at the European Bioinformatics
 Institute, Nucleic Acids Res. 2007; 35 (Issue suppl-2): W6–W11.
 https://doi.org/10.1093/nar/gkm291.
- [33] Huerta-Cepas J, Forslund S, Coelho LP, Szklarczyk D, Jensen L, von Mering C, Bork P. Fast
 Genome-Wide Functional Annotation through Orthology Assignment by eggNOG-Mapper. Mol
 Biol Evol. 2017; 34. 10.1093/molbev/msx148.
- [34] Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, *et al.* Gene set
 enrichment analysis: A knowledge-based approach for interpreting genome-wide expression
 profiles. PNAS. 2005; 102(43): 15545–15550.
- [35] Chen Y, Meltzer PS. Gene Expression Analysis via Multidimensional Scaling. Current Protocols
 in Bioinformatics. 2005; Chapter 7: Unit 7.11. doi: 10.1002/0471250953.bi0711s10.
- [36] Rodriguez-Navarro, A. Potassium transport in fungi and plants. Biochim Biophys Acta. 1999;
 1469 (2000): 1–30.

- [37] Camacho M, Ramos J, Rodríguez-Navarro A. Potassium requirements of *Saccharomyces cerevisiae*. Curr Microbiol. 1981; 6: 295–299.
- [38] Gostinčar C, Lenassi M, Gunde-Cimerman N, Plemenitaš A. Fungal adaptation to extremely
 high salt concentrations. Adv Appl Microbiol. 2011; 77: 71–96.
- [39] Garcia K, Delteil A, Conejero G, Becquer A, Plassard C, Sentenac H, Zimmermann S. Potassium nutrition of ectomycorrhizal *Pinus pinaster*: overexpression of the *Hebeloma cylindrosporum*HcTrk1 transporter affects the translocation of both K and P in the host plant. New Phytol. 2014; 201: 951–960.
- [40] Jentschke G, Brandes B, Kuhn AJ, Schröder WH, Godbold DL. Interdependence of phosphorus,
 nitrogen, potassium and magnesium translocation by the ectomycorrhizal fungus *Paxillus involutus*. New Phytol. 2001; 149: 327–337.
- 572 [41] Bücking H, Heyser W. Elemental composition and function of polyphosphates in ectomycorrhizal fungi an X-ray microanalytical study. Mycol Res. 1999; 103(1): 31–39.
- 574 [42] Orlovich DA, Ashford AE. Polyphosphate granules are an artefact of specimen preparation in
 575 the ectomycorrhizal fungus *Pisolithus tinctorius*. Protoplasma. 1993; 173: 91–105.
- [43] Ashford AE, Vesk PA, Orlovich DA, Markovina A-L, Allaway WG. Dispersed polyphosphate
 in fungal vacuoles of *Eucalyptus pilularis/Pisolithus tinctorius* ectomycorrhizas. Fungal Genet
 Biol. 1999; 28: 21–33.
- [44] Bonneville SC, Bray A, Benning L. Structural Fe(II) Oxidation in Biotite by an Ectomycorrhizal
 Fungi Drives Mechanical Forcing. Environ sci & technol. 2016; 50(11): 5589–5596.
- [45] Väre H. Aluminium polyphosphate in the ectomycorrhizal fungus *Suillus variegatus* (Fr.) O.
 Kunze as revealed by energy dispersive spectrometry. New Phytol. 1990; 116: 663–668.
- [46] Gadd GM. Metals, minerals and microbes: geomicrobiology and bioremediation. Microbiology.
 2010; 156(3): 609–643.
- [47] Illmer P, Buttinger R. Interactions between iron availability, aluminium toxicity and fungal siderophores. Biometals. 2006; 19(4): 367–377.
- [48] Haro R, Benito B. The Role of Soil Fungi in K+ Plant Nutrition. Int J Mol Sci. 2019; 20(13):
 3169. doi:10.3390/ijms20133169.
- [49] Dueñas-Santero E, Martín-Cuadrado AB, Fontaine T, Latgé J P, del Rey F, Vázquez de Aldana
 C. Characterization of glycoside hydrolase family 5 proteins in *Schizosaccharomyces pombe*.
 Eukaryotic cell. 2010; 9(11): 1650–1660.
- [50] Paparokidou C, Leake JR, Beerling DJ *et al.* Phosphate availability and ectomycorrhizal
 symbiosis with *Pinus sylvestris* have independent effects on the *Paxillus involutus* transcriptome.
 Mycorrhiza. 2012; 31: 69–83.
- [51] Lian B, Wang B, Pan M, Liu C, Teng HH. Microbial release of potassium from K-bearing
 minerals by thermophilic fungus *Aspergillus fumigatus*. Geochim Cosmochim Acta. 2008; 72:87–
 98.
- [52] Bonneville S, Smits MM, Brown A, Harrington J, Leake JR, Brydson R, Benning LG. Plant driven fungal weathering: Early stages of mineral alteration at the nanometer scale. Geology.
 2009; 37(7): 615–618.
- [53] Ahmed E, Holmström SJM. Microbe-mineral interactions: The impact of surface attachment on
 mineral weathering and element selectivity by microorganisms. Chem Geol. 2015; 403: 13–23.
- [54] van Hees PA, Rosling A, Essén S, Godbold DL, Jones DL, Finlay RD. Oxalate and ferricrocin
 exudation by the extramatrical mycelium of an ectomycorrhizal fungus in symbiosis with *Pinus sylvestris*. New Phytol. 2006; 169(2): 367–77. doi: 10.1111/j.1469-8137.2005.01600.x.
- [55] Martínez AT, Ruiz-Dueñas FJ, Camarero S, Serrano A, Linde D, Lund H *et al.* Oxidoreductases
 on their way to industrial biotransformations. Biotechnol Adv. 2017; 35(6); 815–831.
- [56] Song M, Pedruzzi I, Peng Y, Li P, Liu J-F, Yu J. K-Extraction from Muscovite by the Isolated
 Fungi. Geomicrobiol J. 2015; 32(9): 771–779.
- [57] Xiao L, Sun Q, Lian B. A Global View of Gene Expression of *Aspergillus nidulans* on
 Responding to the Deficiency in Soluble Potassium. Curr Microbiol. 2016; 72(4): 410–419.

- [58] Wang W, Lian B, Pan L. An RNA-sequencing study of the genes and metabolic pathways
 involved in *Aspergillus niger* weathering of potassium feldspar. Geomicrobiol J. 2015; 32(8):
 689–700.
- [59] Wang W, Sun Q, Lian B. Redox of Fungal Multicopper Oxidase: A Potential Driving Factor for
 the Silicate Mineral Weathering, Geomicrobiol J. 2018; 35(10): 879–886.
- [60] Garcia K and Zimmermann SD. The role of mycorrhizal associations in plant potassium
 nutrition. Front. Plant Sci. 2014; 5: 337. doi: 10.3389/fpls.2014.00337
- [61] Benito B, Garciadeblás B, Fraile-Escanciano A *et al.* Potassium and sodium uptake systems in
 fungi. The transporter diversity of *Magnaporthe oryzae*. Fungal Genet Biol. 2011; 48: 812–822.
- [62] Corratgé C, Zimmermann S, Lambilliotte R, Plassard C, Marmeisse R, Thibaud JB, *et al.* Molecular and functional characterization of a Na+-K+ transporter from the Trk family in the
 ectomycorrhizal fungus *Hebeloma cylindrosporum*. J. Biol. Chem. 2007; 282: 26057–26066.
- [63] Heaton LLM, Jones NS, Fricker MD. A mechanistic explanation of the transition to simple
 multicellularity in fungi. Nat Commun. 2020; 11: 2594. <u>https://doi.org/10.1038/s41467-020-</u>
 16072-4.
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630 Figure 1. Scanning Electron Microscopy imaging of the minerals after fungal growth. On the left (A, C, E), images obtained in variable pressure mode with backscattering detector, on non-coated 631 samples. On the right (B, D, F), images captured in a high vacuum on gold-palladium coated samples. 632 A) muscovite: the arrow indicates superficial erosion of the mineral that occurred after the exposure 633 to fungal activity; B) muscovite: the arrows point to the tip of a fungal hypha attached to the surface 634 of the mineral, and the superficial trenching; C) phlogopite: the arrow points to the edge of a furrow 635 636 caused on phlogopite surface by the fungus; D) phlogopite: the arrows indicate regular clusters of depositions between the crack caused by the fungus onto the surface of the crystal; E) K-vermiculite: 637 the arrows indicate surface erosion, with a localised scratching and slipping of the surface layer of 638 the crystal; F) K-vermiculite: the arrows indicate fungal hyphae adhering to the mineral's surface, 639 with some flattened, also due to the high vacuum, and others filled with some precipitated material. 640 641

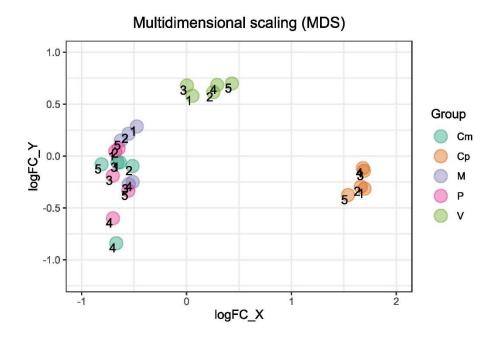


Figure 2. Multidimensional Scaling analysis of transcriptomes obtained from the *P. involutus* grown
 K-vermiculite, muscovite and phologpite as well as positive and negative controls. The distances
 represent the log2 fold change between samples. The symbols differentiate between experiments, the
 numbers represent the biological replicates within each experiment.

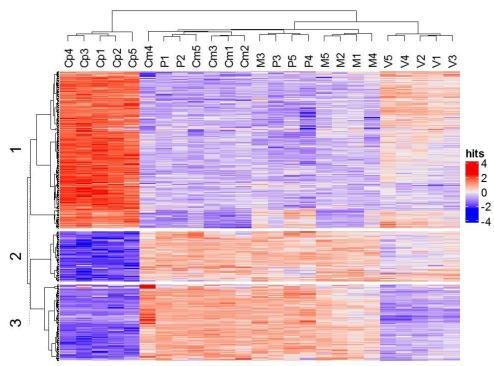


Figure 3. Heatmap of the top 1000 differentially expressed genes (ranked by FDR). It is a twodimensional visual representation of data in which a range of colours represents numerical values of points. The dendrograms added to the left, and top are produced by a hierarchical clustering method based on Euclidean distance computed between the differentially expressed genes (left) and samples (top).

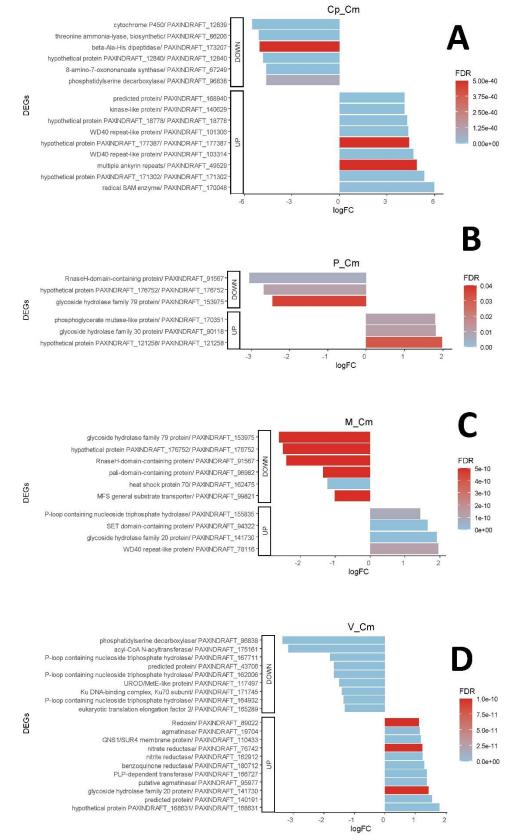


Figure 4. Top Up- and Down-regulated genes between the experiments in which the fungus grew in the presence of the three minerals (V, P and M) compared to the negative control situation (Cm).

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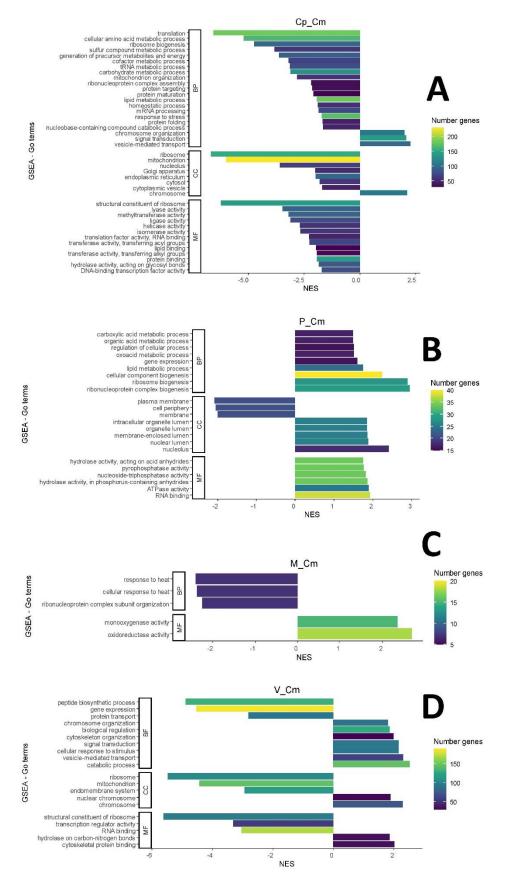


Figure 5. Gene Set Enrichment Analysis (GSEA). Treatments with the three minerals (V, P and M)
and the positive (Cp) control, compared to the negative (Cm) control condition.

Table 1. Uptake of elements by the fungal mycelium. Values are the means $(\pm SD)$ from n = 5replicates. Means in a column without a common letter (a/b/c) differ for P < 0.05, as analysed by oneway ANOVA and Tukey's post hoc test.

Ρ Κ Mg AI Na Са Fe ppm ($\mu q q^{-1}$ of dry mycelium) 0.06±0.001 b Ср 304.11±11.05 a 7.16±0.35 a 110.54± 5.70 a 6.55±0.38 c 3.20±0.17 a 0.33± 0.04 b 5.82±0.75 a 0.07±0.01 b 70.35± 10.49 b 2.99± 0.52 ab Cm 5.52±1.69 c 47.76±11.47 b 0.33± 0.04 b М 6.80±0.96 c 6.21±0.85 a 0.28±0.09 b 73.24± 11.07 b 51.35±8.31 b 2.56± 0.41 b 0.48± 0.09 b Ρ 8.64±2.91 c 6.67±1.15 a 1.12±0.78 a 68.57±7.90 b 47.64±8.00 b 2.73± 0.23 ab 0.93± 0.50 a V 0.16±0.04 b 79.52± 4.52 b 2.63± 0.23 ab 25.11±4.31 b 6.75±0.47 a 64.57±4.81 a 0.41± 0.06 b Pr > F (Model) < 0.0001 0.10 0.0001 < 0.0001 < 0.0001 0.04 0.0001 Significant Yes No Yes Yes Yes Yes Yes

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Table 2. Correlation coefficient (Pearson) between elements concentration in the fungal mycelium
across all the experiments. Values range from -1 (complete negative linear correlation between
variables) to 1 (complete positive linear correlation).

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Variables	Na	Mg	Al	Р	K	Ca	Fe
Na	1	-0.08	0.20	-0.59	-0.87	-0.33	0.22
Mg	-0.08	1	0.39	0.67	0.42	0.54	0.43
Al	0.20	0.39	1	-0.28	-0.28	-0.10	0.99
Р	-0.59	0.67	-0.28	1	0.89	0.63	-0.26
К	-0.87	0.42	-0.28	0.89	1	0.49	-0.28
Са	-0.33	0.54	-0.10	0.63	0.49	1	-0.10
Fe	0.22	0.43	0.99	-0.26	-0.28	-0.10	1

Values in bold are statistically significant (p < 0.05)

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679**Table 3.** Number of up- and down-regulated genes between pairs of treatments (FDR < 0.05), positive</th>680control (C plus K= Cp), K-vermiculite (V), phlogopite (P), muscovite (M) and negative control (C681minus K= Cm). In parenthesis those genes that were up- and down-regulated for more than 2 logFC,682without parenthesis those genes that were up- and down-regulated for more than 1 logFC. Total DEGs683is the sum of up and down regulated genes for (FDR < 0.05).</td>

	Up-regulated	Down-regulated	Total significant DEGs	Total genes shared between pairs of conditions	
	Log FC>1	Log FC>-1			
	(Log FC>2)	(Log FC>-2)			
M/Cm	21 (0)	8 (5)	29	968	
P/Cm	39 (1)	8 (5)	47	1335	
V/Cm	191 (11)	248 (35)	439	5008	
Cp/Cm	832 (111)	676 (155)	1508	7536	
M/Cp	583 (103)	744 (120)	1327	7355	
P/Cp	730 (103)	891 (120)	1621	7732	
V/Cp	198 (39)	428 (62)	626	5786	