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A meta-analysis of effects of feeding seaweed on beef and dairy cattle performance and methane yield

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## 22 Abstract

23           There has been considerable interest in the use of red seaweed, and in particular  
24 *Asparagopsis taxiformis*, to increase production of cattle and to reduce greenhouse gas  
25 emissions. We hypothesized that feeding seaweed or seaweed derived products would increase  
26 beef or dairy cattle performance as indicated by average daily gain (ADG), feed efficiency  
27 measures, milk production, and milk constituents, and reduce methane emissions. We used  
28 meta-analytical methods to evaluate these hypotheses. A comprehensive search of Google  
29 Scholar, Pubmed and ISI Web of Science produced 14 experiments from which 23 comparisons  
30 of treatment effects could be evaluated. Red seaweed (*A. taxiformis*) and brown seaweed  
31 (*Ascophyllum nodosum*) were the dominant seaweeds used. There were no effects of treatment  
32 on ADG or dry matter intake (DMI). There was an increase in efficiency for feed to gain by  
33  $0.41 \pm 0.22$  kg per kg [standardized mean difference (SMD) =  $0.70 \pm 0.35$ ;  $P = 0.001$ ], but not  
34 for gain to feed ( $P = 0.215$ ), although the direction of the change was for improved efficiency.  
35 The type of seaweed used was not a significant covariable for ADG and DMI. Milk production  
36 was increased with treatment on weighted mean difference (WMD;  $1.35 \pm 0.44$  kg/d;  $P$   
37  $<0.001$ ); however, the SMD of 0.45 was not significant ( $P = 0.111$ ). Extremely limited data  
38 suggest the possibility of increased percentages of milk fat ( $P = 0.040$ ) and milk protein ( $P =$   
39  $0.001$ ) on DerSimonian and Laird (D&L) WMD evaluation. The limited data available indicate  
40 dietary supplementation with seaweed produced a significant and substantial reduction in  
41 methane yield by  $5.28 \pm 3.5$  g/kg DMI ( $P = 0.003$ ) on D&L WMD evaluation and a D&L SMD  
42 of  $-1.70$  ( $P = 0.001$ ); however, there was marked heterogeneity in the results ( $I^2 > 80\%$ ). In  
43 one comparison, methane yield was reduced by 97%. We conclude that while there was  
44 evidence of potential for benefit from seaweed use to improve production and reduce methane  
45 yield more *in vivo* experiments are required to strengthen the evidence of effect and identify

- 46 sources of heterogeneity in methane response, while practical applications and potential risks
- 47 are evaluated for seaweed use.

## 48 **Introduction**

49           There has been considerable interest in the use of red seaweed, and in particular  
50 *Asparagopsis taxiformis* to increase production of cattle and to reduce greenhouse gas  
51 emissions [1, 2]. However, several different seaweeds have been fed to cattle and include  
52 brown seaweed (*Ascophyllum nodosum*), and *Saragssum wightii*. A commercial product  
53 ‘*Tasco*’ has been developed based on *A. nodosum* [3].

54           To date, there have been several reviews that have provided qualitative overviews of  
55 the production responses and the extent of inhibition of methane emissions when seaweed was  
56 included in the diets of beef and dairy cows [4, 5]. However, there has been no comprehensive  
57 quantitative review of this subject. Given that studies have evaluated the effects of seaweeds  
58 on beef cattle production, on dairy cattle production, and on methane emissions, there is clear  
59 potential to evaluate the use of seaweeds in cattle production and methane emissions using  
60 meta-analytical methods. We hypothesized that feeding seaweed or seaweed derived products  
61 would increase beef or dairy cattle performance as indicated by average daily gain (ADG), feed  
62 efficiency measures, milk production, and milk constituents, and reduce methane emissions.

## 63 **Materials and methods**

### 64 **Literature search**

65           A comprehensive search of the English language literature used the US National  
66 Library of Medicine National Institutes of Health through PubMed  
67 (<http://www.ncbi.nlm.nih.gov/pubmed>), Google Scholar (<http://scholar.google.com/>), and the  
68 ISI Web of Science (<http://apps.webofknowledge.com>). The search was conducted on 21  
69 January 2021 and searches were based on the following key words with no limits included:

70 seaweed and cattle. We searched the reference lists of papers obtained to identify other studies.  
71 One additional paper was identified from a personal communication.

72 For Google Scholar, 28,400 citation results occurred, and the screening of papers  
73 stopped when 50 sequential citations were not relevant. Whereas only 58 and 55 results  
74 occurred from Pubmed and ISI Web of Science, respectively. In one case, the authors of an  
75 article were contacted to clarify results and to provide additional information.

## 76 **Inclusion criteria**

77 Papers were primarily screened on their citation title by 2 reviewers and secondarily  
78 screened based on the full text. Experiments were included in the analysis if they met the  
79 following inclusion criteria developed by *Scibus* (Camden, NSW, Australia): were full  
80 manuscripts from peer-reviewed journals; experiments were *in vivo* and the animals studied  
81 were cattle; the experiments evaluated use of seaweed or seaweed derived products for dietary  
82 supplementation of cattle; they were randomized; they had a description of the randomization  
83 processes employed; they had appropriate analysis of data; they contained sufficient data to  
84 determine the effect size for production outcomes (e.g., the number of cattle or pens in each  
85 treatment and control group); they had a measure of effect so that the data were amenable to  
86 effect size (ES) analysis for continuous data (e.g., standardized mean difference, SMD); and  
87 they had a measure of variance (SE or SD) for each effect estimate or treatment and control  
88 comparisons. Studies that could not be adequately interpreted, used purposive and non-  
89 representative sampling methods or where authors did not respond to clarify their approach,  
90 were excluded. Note, one article was included from the pre-print server for Biology, bioRxiv  
91 (<https://www.biorxiv.org/>).

92 Fig. 1 depicts a PRISMA diagram [6] of the flow of data collection for the meta-  
93 analysis. The PRISMA checklist is provided in S1 File. After the initial search and screening

94 61 different articles (experiments) were identified and papers without a full text (5) were  
95 excluded providing 56 papers that were assessed for eligibility. A total of 42 were excluded for  
96 the following reasons: the abstract was in English but the full article was in another language  
97 (3 experiments), the experiment was *in vitro* (8 experiments), the article was a review or book  
98 chapter (7 articles), the experiment had group feeding resulting in pseudo-replication (2  
99 experiments), the experiment was off topic or had irrelevant outcomes (20 experiments), or the  
100 experiment lacked measures of variance (2 experiments). A list of articles excluded with the  
101 reason is provided in S1 Table. A total of 14 experiments with 23 treatment comparisons were  
102 included in the meta-analysis. A list of the experiments and comparisons included in the meta-  
103 analysis is provided in Table 1.

104 **Fig. 1. PRISMA flow diagram (adapted from [6]) of the systematic review from initial**  
105 **search and screening to final selection of publications to be included in the meta-analysis**  
106 **on seaweed in cattle.**

## 107 **Data extraction**

108 All data extracted from each of the experiments that met the inclusion criteria were  
109 audited by up to three reviewers. The descriptive data extracted included experiment design,  
110 and details about the experiment and the animals used. Design details included the number of:  
111 animals or pens, animals/group, and pens/group; experimental and analytical unit (animal or  
112 pen). Experimental details included: the number of days on feed, the number of days treatment  
113 products were fed, the dose of treatment administered, and diet and delivery methods of  
114 product. Animal details included: class of cattle (steers or heifers, or dairy cows), production  
115 system (dairy or beef), initial body weight of control and treatment groups, and type of housing  
116 and feeding systems. Key descriptive data are provided in Table 1.

117            Output variables extracted for meta-analysis included: final body weight (FBW, kg),  
118    ADG (kg/head/d), dry matter intake (DMI; kg/head/d), gross feed efficiency [ratio of gain to  
119    feed (G:F) and ratio of feed to gain (F:G)], milk yield (kg/d), milk fat percentage, milk protein  
120    percentage, and methane yield (g/kg DMI) (Tables 2 and 3).

**Table 1. Descriptive information for the comparisons included in the dataset**

Author and Reference	Year	Design	Breed	Production system	Unit of interest	Number of replicates		Study length (d)	Seaweed category	Dose of seaweed	Initial body weight (kg)	
						Control	Treatment				Control	Treatment
Allen et al. [7]	2001	RBD	ANG, ANG×HF, BRAH	Beef	Pen	12	12	146.5	<i>A. nodosum</i>	3.4 kg/ha	330.0 ± 17.32	325.0 ± 17.32
Allen et al. [7]	2001	RBD	ANG, ANG×HF, BRAH	Beef	Pen	12	12	146.5	<i>A. nodosum</i>	3.4 kg/ha	360.0 ± 17.32	355.0 ± 17.32
Anderson et al. [8]	2006	RBD	English crossbred	Beef	Pen	4	4	24	<i>A. nodosum</i>	2.0 %	381.5 ± 9.86	384.5 ± 9.86
Anderson et al. [8]	2006	RBD	English crossbred	Beef	Pen	4	4	14	<i>A. nodosum</i>	2.0 %	381.5 ± 9.86	388.2 ± 9.86
Anderson et al. [8]	2006	RBD	English crossbred	Beef	Pen	4	4	38	<i>A. nodosum</i>	2.0 %	381.5 ± 9.86	385.9 ± 9.86
Antaya et al. [9]	2019	RBD	Jersey	Dairy	ANI	10	10	28	<i>A. nodosum</i>	113 g/hd/d	420.0 ± 44.00	400.0 ± 36.00
Carter et al. [10]	2000	RCT	Predominately British crosses	Beef	Pen	4	4	56	<i>A. nodosum</i>	273 g/hd/d		
Cvetkovic et al. [11]	2004	RCT	Holstein	Dairy	ANI	12	12	21	<i>A. nodosum</i>	114 g/hd/d		
Gravett [12]	2000	RBD	ANG & ANG cross	Beef	Pen	10	10	14	<i>A. nodosum</i>	1.0 %		
Kidane et al. [13]	2015	4x4 LS	Norwegian Red	Dairy	ANI	6	6	28	<i>A. nodosum</i>	160 g/hd/d		
Williams et al. [3]	2009	2x2 fact	ANG crossbred	Beef	ANI	6	6	13	<i>A. nodosum</i>	1.0 %	367.9 ± 20.58	367.0 ± 20.58
Williams et al. [3]	2009	2x2 fact	ANG crossbred	Beef	ANI	6	6	13	<i>A. nodosum</i>	1.0 %	343.4 ± 20.58	329.8 ± 20.58
Kinley et al. [2]	2020	RBD	BRAH×ANG	Beef	ANI	5	5	90	<i>A. taxiformis</i>	0.05 %		
Kinley et al. [2]	2020	RBD	BRAH×ANG	Beef	ANI	5	5	90	<i>A. taxiformis</i>	0.1 %		
Kinley et al. [2]	2020	RBD	BRAH×ANG	Beef	ANI	5	5	90	<i>A. taxiformis</i>	0.2 %		
Roque et al. [1]	2020	RCT	ANG×HF	Beef	ANI	7	7	147	<i>A. taxiformis</i>	0.25 %	357.0 ± 24.37	348.0 ± 24.37
Roque et al. [1]	2020	RCT	ANG×HF	Beef	ANI	7	6	147	<i>A. taxiformis</i>	0.5 %	357.0 ± 24.37	350.0 ± 22.56
Stefenoni et al. [14]	2021	4x4 LS	Holstein	Dairy	ANI	20	20	7	<i>A. taxiformis</i>	0.25 %		
Stefenoni et al. [14]	2021	4x4 LS	Holstein	Dairy	ANI	20	20	7	<i>A. taxiformis</i>	0.5 %		
Bendary et al. [15]	2013	RCT	Friesian	Dairy	ANI	6	6	150	Other <sup>a</sup>	50 g/hd/d	534.0 ± 13.04	534.0 ± 13.04
Sharma & Datt [16]	2020	RCT	Karan fries	Beef	ANI	6	6	150	Other <sup>b</sup>	1.5 %	415.9 ± 13.10	403.4 ± 14.10
Sharma & Datt [16]	2020	RCT	Karan fries	Beef	ANI	6	6	150	Other <sup>b</sup>	3.0 %	415.9 ± 13.10	406.6 ± 10.15
Singh et al. [17]	2015	RCT	Sahiwal	Dairy	ANI	4	4	126	Other <sup>c</sup>	20 %		
									<b>Mean ± SD</b>		<b>388.1 ± 18.26</b>	<b>382.9 ± 17.35</b>



RBD = randomized block design; RCT = randomized controlled trial; LS = latin square; ANG = Angus; HF = Hereford; BRAH = Brahman; ANI = animal; *A. nodosum* = *Ascophyllum nodosum*; *A. taxiformis* = *Asparagopsis taxiformis*; Other = seaweed that is not *A. nodosum* or *A. taxiformis*

<sup>a</sup> Seaweed meal (Crossgates Bioenergetics-Seaweeds Company, Gargrave, North Yorkshire, United Kingdom)

<sup>b</sup> *Kappaphycus alvarezii* & *Gracilaria Salicornia*

<sup>c</sup> *Sargassum wightii*

**Table 2. Mean  $\pm$  SD of control and treatment group production outcomes for each comparison included in analysis**

Author and reference	Year	Seaweed category	Final body weight (kg)		Average daily gain (kg/d)		Dry matter intake (kg/d)		Feed to gain (kg/kg)		Gain to feed (kg/kg)	
			Control	Treatment	Control	Treatment	Control	Treatment	Control	Treatment	Control	Treatment
Allen et al. [7]	2001	<i>A. nodosum</i>	555.0 $\pm$ 24.25	552.0 $\pm$ 24.25	1.61 $\pm$ 0.10	1.63 $\pm$ 0.10			6.70 $\pm$ 0.55	6.30 $\pm$ 0.55		
Allen et al. [7]	2001	<i>A. nodosum</i>	578.0 $\pm$ 24.25	570.0 $\pm$ 24.25	1.57 $\pm$ 0.10	1.55 $\pm$ 0.10			7.20 $\pm$ 0.55	6.90 $\pm$ 0.55		
Anderson et al. [8]	2006	<i>A. nodosum</i>	553.8 $\pm$ 17.58	552.5 $\pm$ 17.58	1.45 $\pm$ 0.12	1.41 $\pm$ 0.12					9.14 $\pm$ 0.72	9.08 $\pm$ 0.72
Anderson et al. [8]	2006	<i>A. nodosum</i>	553.8 $\pm$ 17.58	544.4 $\pm$ 17.58	1.45 $\pm$ 0.12	1.31 $\pm$ 0.12					9.14 $\pm$ 0.72	9.69 $\pm$ 0.72
Anderson et al. [8]	2006	<i>A. nodosum</i>	553.8 $\pm$ 17.58	567.1 $\pm$ 17.58	1.45 $\pm$ 0.12	1.52 $\pm$ 0.12					9.14 $\pm$ 0.72	9.02 $\pm$ 0.72
Antaya et al. [9]	2019	<i>A. nodosum</i>	408.0 $\pm$ 35.73	392.0 $\pm$ 35.73			18.1 $\pm$ 1.26	19.3 $\pm$ 1.26				
Carter et al. [10]	2000	<i>A. nodosum</i>			0.86 $\pm$ 0.18	0.73 $\pm$ 0.18			7.52 $\pm$ 3.40	5.78 $\pm$ 3.40		
Cvetkovic et al. [11]	2004	<i>A. nodosum</i>					22.7 $\pm$ 1.87	22.5 $\pm$ 1.87				
Gravett [12]	2000	<i>A. nodosum</i>			1.31 $\pm$ 0.16	1.36 $\pm$ 0.16			6.23 $\pm$ 0.41	6.00 $\pm$ 0.41		
Kidane et al. [13]	2015	<i>A. nodosum</i>					18.1 $\pm$ 1.15	18.1 $\pm$ 1.15				
Williams et al. [3]	2009	<i>A. nodosum</i>	368.5 $\pm$ 19.96	365.4 $\pm$ 19.96	0.05 $\pm$ 0.66	-0.13 $\pm$ 0.66						
Williams et al. [3]	2009	<i>A. nodosum</i>	378.3 $\pm$ 19.96	364.5 $\pm$ 19.96	2.68 $\pm$ 0.66	2.66 $\pm$ 0.66						
Kinley et al. [2]	2020	<i>A. taxiformis</i>			1.21 $\pm$ 0.38	1.24 $\pm$ 0.36	9.0 $\pm$ 1.77	8.0 $\pm$ 1.14	7.45 $\pm$ 1.39	6.95 $\pm$ 0.58		
Kinley et al. [2]	2020	<i>A. taxiformis</i>			1.21 $\pm$ 0.38	1.52 $\pm$ 0.29	9.0 $\pm$ 1.77	10.5 $\pm$ 1.43	7.45 $\pm$ 1.30	6.60 $\pm$ 0.58		
Kinley et al. [2]	2020	<i>A. taxiformis</i>			1.21 $\pm$ 0.38	1.47 $\pm$ 0.16	9.0 $\pm$ 1.77	9.4 $\pm$ 0.49	7.45 $\pm$ 0.38	6.42 $\pm$ 0.58		
Roque et al. [1]	2020	<i>A. taxiformis</i>	589.0 $\pm$ 29.37	572.0 $\pm$ 29.37	1.60 $\pm$ 0.16	1.52 $\pm$ 0.16	11.3 $\pm$ 0.77	10.4 $\pm$ 0.77			0.14 $\pm$ 0.03	0.15 $\pm$ 0.03
Roque et al. [1]	2020	<i>A. taxiformis</i>	589.0 $\pm$ 29.37	587.0 $\pm$ 27.19	1.60 $\pm$ 0.16	1.56 $\pm$ 0.15	11.3 $\pm$ 0.77	9.7 $\pm$ 0.71			0.14 $\pm$ 0.03	0.16 $\pm$ 0.02
Stefenoni et al. [14]	2021	<i>A. taxiformis</i>	642.0 $\pm$ 77.37	645.0 $\pm$ 77.37			25.3 $\pm$ 5.81	24.5 $\pm$ 5.81				
Stefenoni et al. [14]	2021	<i>A. taxiformis</i>	642.0 $\pm$ 77.37	635.0 $\pm$ 77.37			25.3 $\pm$ 5.81	23.5 $\pm$ 5.81				
Bendary et al. [15]	2013	Other					17.1 $\pm$ 0.42	17.2 $\pm$ 0.42				
Sharma & Datt [16]	2020	Other	426.9 $\pm$ 12.02	417.2 $\pm$ 13.71			12.2 $\pm$ 0.24	11.7 $\pm$ 0.19				
Sharma & Datt [16]	2020	Other	426.9 $\pm$ 12.02	418.4 $\pm$ 9.76			12.2 $\pm$ 0.24	12.0 $\pm$ 0.18				
Singh et al. [17]	2015	Other	338.9 $\pm$ 18.70	334.2 $\pm$ 16.50			9.3 $\pm$ 1.40	9.6 $\pm$ 0.70				
		<b>Mean <math>\pm</math> SD</b>	<b>506.9 <math>\pm</math> 28.87</b>	<b>501.1 <math>\pm</math> 28.54</b>	<b>1.38 <math>\pm</math> 0.26</b>	<b>1.38 <math>\pm</math> 0.24</b>	<b>15.0 <math>\pm</math> 1.79</b>	<b>14.7 <math>\pm</math> 1.57</b>	<b>7.14 <math>\pm</math> 1.14</b>	<b>6.42 <math>\pm</math> 0.95</b>	<b>5.54 <math>\pm</math> 0.44</b>	<b>5.62 <math>\pm</math> 0.44</b>

*A. nodosum* = *Ascophyllum nodosum*; *A. taxiformis* = *Asparagopsis taxiformis*; Other = seaweed that is not *A. nodosum* or *A. taxiformis*.

**Table 3. Mean  $\pm$  SD of control and treatment group milk production and methane outcomes for each comparison included in analysis**

Author	Year	Seaweed category	Milk volume (kg/d)		Milk fat (%)		Methane (g/kg DMI)	
			Control	Treatment	Control	Treatment	Control	Treatment
Antaya et al. [9]	2019	<i>A. nodosum</i>	14.4 $\pm$ 1.90	15.2 $\pm$ 1.90	4.4 $\pm$ 0.60	4.5 $\pm$ 0.60	22.6 $\pm$ 2.78	20.6 $\pm$ 2.78
Cvetkovic et al. [11]	2004	<i>A. nodosum</i>	33.5 $\pm$ 2.08	35.3 $\pm$ 2.08	3.9 $\pm$ 0.45	3.6 $\pm$ 0.45		
Kidane et al. [13]	2015	<i>A. nodosum</i>	15.7 $\pm$ 1.57	16.0 $\pm$ 1.57	4.3 $\pm$ 0.30	4.1 $\pm$ 0.30		
Kinley et al. [2]	2020	<i>A. taxiformis</i>					11.0 $\pm$ 1.92	10.0 $\pm$ 3.85
Kinley et al. [2]	2020	<i>A. taxiformis</i>					11.0 $\pm$ 1.92	6.8 $\pm$ 4.02
Kinley et al. [2]	2020	<i>A. taxiformis</i>					11.0 $\pm$ 1.92	0.3 $\pm$ 0.31
Roque et al. [1]	2020	<i>A. taxiformis</i>					17.5 $\pm$ 2.65	9.5 $\pm$ 2.65
Roque et al. [1]	2020	<i>A. taxiformis</i>					17.5 $\pm$ 2.65	5.0 $\pm$ 2.45
Stefenoni et al. [14]	2021	<i>A. taxiformis</i>	40.2 $\pm$ 8.59	40.0 $\pm$ 8.59	3.6 $\pm$ 0.51	3.6 $\pm$ 0.51	13.9 $\pm$ 3.00	14.4 $\pm$ 3.00
Stefenoni et al. [14]	2021	<i>A. taxiformis</i>	40.2 $\pm$ 8.59	37.6 $\pm$ 8.59	3.6 $\pm$ 0.51	3.6 $\pm$ 0.51	13.9 $\pm$ 3.00	9.8 $\pm$ 3.00
Bendary et al. [15]	2013	Other	12.6 $\pm$ 0.44	14.1 $\pm$ 0.44	3.2 $\pm$ 0.02	3.2 $\pm$ 0.02	11.0 $\pm$ 1.92	10.0 $\pm$ 3.85
Singh et al. [17]	2015	Other	7.3 $\pm$ 2.30	8.8 $\pm$ 1.50	5.3 $\pm$ 0.16	5.4 $\pm$ 0.17		
		<b>Mean <math>\pm</math> SD</b>	<b>23.4 <math>\pm</math> 3.64</b>	<b>23.9 <math>\pm</math> 3.52</b>	<b>4.0 <math>\pm</math> 0.36</b>	<b>4.0 <math>\pm</math> 0.36</b>	<b>14.8 <math>\pm</math> 2.48</b>	<b>9.5 <math>\pm</math> 2.76</b>

*A. nodosum* = *Ascophyllum nodosum*; *A. taxiformis* = *Asparagopsis taxiformis*; Other = seaweed that is not *A. nodosum* or *A. taxiformis*; DMI = dry matter intake

## 121 **Statistical analysis**

122 Data were structured to allow a classical meta-analytical evaluation of differences in  
123 responses of the experimental groups. Many of the experiments in this analysis used multiple  
124 treatment comparisons (nesting), and therefore the data had a hierarchical structure. For this  
125 reason, meta-regression using multi-level models was used to evaluate the effects of  
126 experiment and treatment by taking into account this hierarchical structure [18-20].

127 Initial data exploration included production of basic statistics using Stata (Version 16,  
128 StataCorp. LP, College Station, TX) to examine the data for errors and to estimate the means  
129 and measures of dispersion. Normality of the data was examined for continuous variables, by  
130 visual and statistical appraisal.

131 Stata was also used to analyze differences in responses by SMD analysis which is also  
132 called ES analysis. These methods have been published in detail in [21] and [22]. The  
133 difference between treatment and reference groups means, which is termed ‘treatment’ in the  
134 following description, was standardized using the SD of reference and treatment groups. The  
135 SMD estimates were pooled using the DerSimonian and Laird random effects models (D&L)  
136 [23] and, in the case of methane yield, with the more conservative Knapp-Hartung method (K-  
137 H) [24]. Only random effects models were used, as previous work concluded that when there  
138 was uncertainty in the evaluative units caused by clustering of observations, the random effects  
139 model was appropriate [25].

140 Robust regressions models (RR) were produced that account for the nested effect of  
141 comparisons within experiment [18] and analysed using “*robumeta*” (Stata) as applied by [26].  
142 The RR were developed to account for the two-stage cluster sampling inherent when the ES  
143 estimates are derived from a total of  $n = k_1 + k_2 + \dots + k_m$  estimates from comparisons that  
144 were collected by sampling  $m$  clusters of experiments, that is several comparison estimates are  
145 derived from the same experiment [18]. Hence, sampling  $k_j \geq 1$  estimates within the  $j^{\text{th}}$  cluster

146 for  $j = 1, \dots, m$ . Briefly, in this test the mean ES from a series of experiments is described as  
147 follows: In this case, the regression model has only an intercept  $b_1$  and the weighted mean has  
148 the form:

$$149 \quad b_1 = \frac{\sum_{j=1}^m \sum_{i=1}^{k_1} w_{ij} T_{ij}}{\sum_{j=1}^m \sum_{i=1}^{k_1} w_{ij}}$$

150 where  $m$  is the total number of experiments,  $k$  the total number of comparisons in the extracted  
151 database and  $w_{ij}$  is the weighting for comparisons within experiments and  $T_{ij}$  is the vector of  
152 the ES estimates of comparisons within experiments. If all the estimates in the same experiment  
153 are given identical weights, the robust variance estimate ( $v^R$ ) reduces to:

$$154 \quad v^R = \frac{\sum_{j=1}^m w_j^2 (\check{T}_j - b_1)^2}{(\sum_{j=1}^m w_j)^2}$$

155 where  $\check{T}_j$  is the unweighted mean of the estimates in the  $j^{\text{th}}$  cluster,  $b_1$  is the estimate of the  
156 weighted mean, and  $w_j$  is the total weight given to estimates in the  $j^{\text{th}}$  cluster. This is a kind of  
157 weighted variance which reduces to  $(m-1)/m^2$  times the variance, when the weights within  
158 experiment are identical, and (since the correlation coefficient = 1 in this case) the robust  
159 regression standard error equals  $1/m$  times the variance of  $\check{T}_j$  estimated when the weights are  
160 equal. Several important aspects of the robust model are highlighted by [18] and the underlying  
161 assumptions that; the correlation structure of the  $T_j$  does not need be known to compute the  
162 pooled ES or  $V^R$ , only that the vectors of estimates from different experiments are independent  
163 and that regularity conditions are satisfied; the experiment or comparison level regressors do  
164 not need to be fixed; the theorem is asymptotic based on the number of experiments, rather  
165 than the number of comparisons; and the theorem is relatively robust to regularity assumptions.

166 A random effects weighted mean difference (WMD) between treatment and reference  
167 was estimated, with the weighting reflecting the inverse of the variance of the treatments

168 included according to the *nostandard* method in the *metan* model of Stata to allow an  
169 interpretation of treatment effects in familiar units (e.g. kg of FBW), rather than ES.

170 Forest plots were produced for both WMD and SMD results for each outcome variable  
171 that incorporated the D&L and RR estimates. The forest plots provided further allow a  
172 comparison of *A. taxiformis*, *A. nodosum*, and ‘*other*’ sources of seaweed evaluated with the  
173 D&L and RR methods. Additionally, plots were produced for initial body weight.

174 Variations among the comparison level SMD were assessed using a chi-squared (Q)  
175 test of heterogeneity. Heterogeneity in comparisons reflects underlying differences in clinical  
176 diversity of the research site and interventions, differences in experimental design and  
177 analytical methods, and statistical variation around responses. The clinical diversity of the site  
178 includes all the non-study design aspects of variation, such as facility design, environment, and  
179 cattle management that may be measured and controlled for in meta-analysis but are often not  
180 reported or measured. Identifying the presence and sources of the heterogeneity improves  
181 understanding of the responses to the interventions used. An  $\alpha$  level of 0.10 was used because  
182 of the relatively poor power of the  $\chi^2$  test to detect heterogeneity among small numbers of trials  
183 [27]. Heterogeneity of results among the comparisons was quantified using the  $I^2$  statistic [28].  
184 The  $I^2$  provides an estimate of the proportion of the true variance of effects of the treatment,  
185 that is the true variance,  $\tau^2$  ( $\tau^2$ ) divided by the total variance observed in the comparison [29]  
186 that reflect measurement error. Negative values of  $I^2$  are assigned a value of 0, consequently  
187 the value  $I^2$  lies between 0 and 100%. An  $I^2$  value between 0 and 40% might not be important,  
188 30 to 60% may represent moderate heterogeneity, 50 to 90% might represent substantial  
189 heterogeneity, and 75 to 100% might represent considerable heterogeneity [30]. Both  $I^2$  and  $\tau^2$   
190 are provided to allow readers the opportunity to evaluate both metrics.

191 A key focus of meta-analysis is to identify and understand the sources of heterogeneity  
192 or variation of response among comparisons. However, given the limited number of

193 experiments available the only meta-regression analyses suitable were for category of seaweed  
194 intervention for ADG and DMI and production system for DMI.

195 Presence of publication bias was investigated using funnel plots which are a simple  
196 scatter plot of the intervention effect estimates from individual comparisons plotted against  
197 comparison precision. The name '*funnel plot*' arises because precision of the intervention effect  
198 increases as the size and precision of a comparison increases. Effect estimates from  
199 comparisons with a small number of animal units will scatter more widely at the bottom of the  
200 graph and the spread narrows for those with higher numbers of units. In the absence of bias,  
201 the plot should approximately resemble a symmetrical (inverted) funnel. Funnel plots are  
202 available upon request.

## 203 **Results and discussion**

204 The literature that was amenable to quantitative review on seaweed use in cattle was  
205 reasonably limited with only 14 full texts suitable (Fig 1; Table 1). The experiments used were  
206 all published after the year 2000, indicating that they are relatively current. Although these  
207 were current some production data indicated only modest production performance (Tables 2  
208 and 3). Funnel plots produced indicated that publication bias was not likely (data not shown).  
209 The limited number of comparisons and even fewer experiments limited the type of meta-  
210 regressions that could be performed and the use of RR. Only 2 experiments, one on a dairy and  
211 one on a beef production system, used Latin Square designs and this precluded evaluation of  
212 the effect of study design. As the SD of these were similar to the randomized controlled designs  
213 adjustments to the error terms for these were not made.

214 Differences in FBW were significant for treatment for both RR SMD and RR WMD  
215 suggesting that the FBW was lower for treated cattle (Table 4). These findings were not  
216 supported by differences in ADG with all models showing little difference in ADG (Table 4;  
217 Fig 2). The numerically lower initial body weight for treated cattle supports the contention that

218 FBW differences were substantially influenced by initial BW differences (WMD D&L = -3.08  
219 kg; 95% CI = -7.62 to 1.46; P = 0.183; SMD D&L = -0.28; 95% CI = -0.57 to 0.02; P = -0.57  
220 to 0.02). The comparisons contributing to the observations on FBW and ADG differ but had  
221 considerable overlap as 9 comparisons were shared. There was no evidence of difference  
222 between *A. taxiformis* and *A. nodosum* interventions on FBW (data not shown) or ADG (Table  
223 4).

224

225 **Fig 2. Forest plot of the weighted mean difference (WMD) and 95% CI of the effect of**  
226 ***Ascophyllum nodosum* and *Asparagopsis taxiformis* seaweed intervention on the average**  
227 **daily gain (ADG; kg/d) of cattle.** The solid vertical line represents a mean difference of zero  
228 or no effect. Points to the left of the line represent a reduction in ADG, while points to the right  
229 of the line indicate an increase. Each square around the point effect represents the mean effect  
230 size for that comparison and reflects the relative weighting of the comparison to the overall  
231 effect size estimate. The larger the box, the greater the comparison contribution to the overall  
232 WMD estimate. The weights that each comparison contributed are in the right-hand column.  
233 The upper and lower limit of the line connected to the square represents the upper and lower  
234 95% CI for the WMD. The overall pooled WMD and 95% CI pooled using the DerSimonian  
235 and Laird (D+L) [23] and robust meta-analytical random effects models [18, 26] are indicated  
236 by the respective diamonds at the bottom. The heterogeneity measure,  $I^2$  is a measure of  
237 residual variation among comparisons included in the meta-analysis. The ADG was not  
238 heterogeneous as indicated by the overall  $I^2$  of 0%.



239 **Table 4. Summary of the meta-analysis using classical meta-analysis methods for the effects of seaweed on production measures. The**  
 240 **Table provides the number (N) of experiments and comparisons for each evaluation, the weighted mean difference (WMD) and**  
 241 **standardized mean difference (SMD) using both the DerSimonian and Laird (D&L) and robust regression (RR) methods, and the P-value,**  
 242 **estimated heterogeneity ( $I^2$ ) and comparison and experiment variance ( $\tau^2$ ) of these estimates when available.**

Measure	N comparisons (N experiments)	Effect (95% CI)	P-value	Heterogeneity ( $I^2$ , %)	Variance ( $\tau^2$ )	Meta-regressions (coefficient $\pm$ SE; P-value; $\tau^2$ )
<b>Final body weight</b>						
WMD (D&L; kg)	15 (8)	-6.57 (-12.23 to -0.90)	0.023	0	0	
WMD (RR; kg)	15 (8)	-5.71 (-11.84 to -0.37)	0.039		0	
SMD (D&L)	15 (8)	-0.23 (-0.48 to 0.02)	0.067	0	0	
SMD (RR)	15 (8)	-0.27 (-0.52 to -0.02)	0.041		0	
<b>Average daily gain</b>						
WMD (D&L; kg/d)	14 (7)	-0.01 (-0.05 to 0.03)	0.730	0	0	
WMD (RR; kg/d)	14 (7)	0.01 (-0.09 to 0.07)	0.711		0	<i>A. taxiformis</i> compared to <i>A. nodosum</i> as reference 0.05 $\pm$ 0.13; P = 0.726; $\tau^2$ = 0
SMD (D&L)	14 (7)	-0.01 (-0.31 to 0.29)	0.947	0	0	
SMD (RR)	14 (7)	-0.03 (-0.49 to 0.42)	0.863		0	<i>A. taxiformis</i> compared to <i>A. nodosum</i> as reference 0.36 $\pm$ 0.50; P = 0.538; $\tau^2$ = 0
<b>Dry matter intake</b>						
WMD (D&L; kg/d)	14 (9)	-0.28 (-0.63 to 0.07)	0.119	60.95	0.35	
WMD (RR; kg/d)	14 (9)	-0.33 (-0.99 to 0.48)	0.47		0	Dairy compared to beef as reference 0.76 $\pm$ 0.38; P = 0.106; $\tau^2$ = 0 <i>A. nodosum</i> compared to 'Other' as reference 0.54 $\pm$ 0.51; P = 0.364; $\tau^2$ = 0 <i>A. taxiformis</i> compared to 'Other' as reference

						-0.43 ± 0.77; P = 0.622; $\tau^2 = 0$
SMD (D&L)	14 (9)	-0.31 (-0.75 to 0.14)	0.177	59.4	0.39	
SMD (RR)	14 (9)	-0.25 (-0.91 to 0.41)	0.393		0	Dairy compared to beef as reference 0.83 ± 0.75; P = 0.324; $\tau^2 = 0$ <i>A. nodosum</i> compared to 'Other' as reference 0.75 ± 0.78; P = 0.389; $\tau^2 = 0$ <i>A. taxiformis</i> compared to 'Other' as reference 0.14 ± 0.85; P = 0.874; $\tau^2 = 0$
<b>Feed to gain</b>						
WMD (D&L)	7 (4)	-0.41 (-0.63 to -0.20)	0.001	0.1	0	
SMD (D&L)	7 (4)	-0.70 (-1.01 to -0.31)	0.001	0	0	
<b>Gain to feed</b>						
WMD (D&L)	5 (2)	0.02 (-0.01 to 0.04)	0.133	0	0	
SMD (D&L)	5 (2)	0.35 (-0.21 to 0.92)	0.215	0	0	
<b>Milk yield</b>						
WMD (D&L; kg/d)	7 (6)	1.35 (0.91 to 1.78)	<0.001	0	0	
SMD (D&L)	7 (6)	0.45 (-0.11 to 1.09)	0.111	65.1	0.39	
<b>Milk fat</b>						
WMD (D&L; %)	7 (6)	0.06 (0.00 to 0.12)	0.040	7.0	0	
SMD (D&L)	7 (6)	0.12 (-0.49 to 0.78)	0.703	66.2	0.41	
<b>Milk protein</b>						
WMD (D&L; %)	6 (5)	0.06 (0.03 to 0.08)	0.001	20.9	0	
SMD (D&L)	6 (5)	0.59 (-0.14 to 1.33)	0.113	73.8	0.56	
<b>Methane</b>						
WMD (D&L; g/kg DMI)	8 (5)	-5.28 (-8.78 to -1.78)	0.003	94.2	23.6	
SMD (D&L)	8	-1.70	0.001	84.0	1.61	

	(5)	(-2.73 to -0.67)				
SMD (K-H) <sup>a</sup>	8 (5)	-1.94 (-3.89 to -0.01)	0.051	84.0	3.57	

243 *A. nodosum* = *Ascophyllum nodosum*; *A. taxiformis* = *Asparagopsis taxiformis*; Other = seaweed that is not *A. nodosum* or *A. taxiformis*; DMI =

244 dry matter intake

245 <sup>a</sup> Knapp-Hartung method [24]

246           There was no effect of treatment on DMI (Table 4; Fig 3) and neither the effects of  
247   dairy or beef production system nor type of seaweed significantly influenced results (Table 4).  
248   Interestingly, these results were heterogenous among comparisons indicating substantial  
249   variations in experimental measurement ( $I^2 > 60\%$ ; Table 4). The F:G was evaluated in 7- and  
250   the G:F in 5- experiments. The F:G was reduced by a significant 0.41 kg per kg with an ES of  
251   0.70 (Table 4); however, these are the less conservative D&L measures as there were  
252   insufficient data to evaluate the RR or the effects of differences in seaweed type on F:G. The  
253   more limited number of experiments on G:F were not significant ( $P = 0.215$ ); however, the  
254   point direction for the SMD (D&L = 0.35) was consistent with improved feed efficiency from  
255   feeding seaweed (Table 4).

256

257   **Fig 3. Forest plot of the weighted mean difference (WMD) and 95% CI of the effect of**  
258   **seaweed intervention on the dry matter intake (DMI; kg/d) of cattle.** Effects for  
259   *Ascophyllum nodosum* and *Asparagopsis taxiformis* and 'Other' seaweed interventions are  
260   provided as well as an overall effect. The solid vertical line represents a mean difference of  
261   zero or no effect. Points to the left of the line represent a reduction in DMI, while points to the  
262   right of the line indicate an increase. Each square around the point effect represents the mean  
263   effect size for that comparison and reflects the relative weighting of the comparison to the  
264   overall WMD estimate. The larger the box, the greater the comparison contribution to the  
265   overall estimate. The weights that each comparison contributed are in the right-hand column.  
266   The upper and lower limit of the line connected to the square represents the upper and lower  
267   95% CI for the WMD. The overall pooled WMD and 95% CI pooled using the DerSimonian  
268   and Laird (D+L) [23] and robust meta-analytical random effects models [18, 26] are indicated  
269   by the respective diamonds at the bottom. The heterogeneity measure,  $I^2$  is a measure of

270 residual variation among comparisons included in the meta-analysis. The DMI was  
271 substantially heterogeneous as indicated by the overall  $I^2$  of 60.9%.

272

273 Milk production was evaluated in only 6 experiments; however, the results were a  
274 significant D&L WMD of 1.35 kg/d increase with treatment. However, the D&L SMD of 0.45  
275 was not significant and was heterogeneous ( $I^2 = 65.1%$ ; Table 4). There were no significant  
276 effects on percentages of milk fat or milk protein on SMD, which were both heterogeneous ( $I^2$   
277 = 66.2% and 73.8%, respectively). However, the WMD for both milk fat and protein  
278 percentages were significantly increased by 0.06% (Table 4). The milk production results  
279 contrast with the lack of effect on ADG of treatment, but may be consistent with the efficiency  
280 improvement in F:G. The differences in SMD and WMD results reflect sparse data and  
281 differences in the weighting between these measures.

282 There is considerable interest in the potential for *Asparagopsis* to reduce methane  
283 emissions and methane yield [1, 2, 14, 31]. The very limited data available for the meta-analysis  
284 provide support for the effect to reduce methane yields *in vivo* with a D&L WMD of  $-5.28 \pm$   
285 3.5 g/kg of DMI, D&L SMD of  $-1.70$  or K-H SMD of  $-1.94$  indicating a substantial reduction  
286 in methane yields. There was marked heterogeneity in the results ( $I^2 > 80%$ ; Table 4; Fig 4). In  
287 one comparison the reduction in methane yield with treatment was 97% [2]. These results are  
288 consistent with the observations made in *in vitro* studies on the effects of *A. taxiformis* on  
289 methane emissions [4] providing further evidence methane emissions is markedly reduced. The  
290 mechanism for the reduction in methane emissions and methane yields has been attributed to  
291 the bromoform and di-bromochloromethane content of the seaweeds [32, 33] that inhibit  
292 methane emissions. However, there are concerns that halogenated gases associated with the  
293 bromoforms could cause damage to the ozone layer [4, 34]. At the higher dose of 0.5%  
294 inclusion of *A. taxiformis*, [14] found that DMI and milk production and energy corrected milk

295 production were significantly lower than controls and that the milk contained markedly  
296 increased concentrations of iodine (> 5 times the control) and bromide (approximately 8 times  
297 the control). In the experiment of [14], the concentration of iodine in milk of cows given 0.5%  
298 *A. taxiformis* was approximately 3 mg/L, and assuming that a child <3 yr old can drink milk at  
299 1 L/d this is approximately 15 times the upper tolerable intake level [35]. Iodine concentrations  
300 in *A. taxiformis* have been reported to range from 8.1 to 11.6 g/kg DM of seaweed [36]. Further,  
301 [37] reported that approximately 31% of ingested iodine is transferred to milk indicating there  
302 is potential that when cows are fed dietary supplements of *A. taxiformis*, iodine concentrations  
303 in milk could be substantially greater than those reported by [14].

304 Although the present analysis indicates that the supplementary feeding of *A. taxiformis*  
305 to beef and dairy cattle has some positive effects on animal production and desirable inhibitory  
306 effects on methane yields, questions are raised, albeit in a single study, that relate to iodine  
307 concentration in *A. taxiformis* and the potential challenges this may bring regarding resultant  
308 iodine concentration in milk when feeding *A. taxiformis* to lactating dairy cows.

309 More *in vivo* experiments are required to strengthen the evidence of production and  
310 methane effects in both beef and dairy cows fed under partial mixed ration and pasture-based  
311 systems. These studies should use a range of *Asparagopsis* preparations/sources, examine  
312 effects on feed intake, and identify sources of heterogeneity in methane response, while  
313 practical applications and potential risks are evaluated for seaweed use.

314

315 **Fig 4. Forest plot of the effect size or standardized mean difference (SMD; standardized**  
316 **using the z-statistic) and 95% CI of the effect of seaweed intervention on methane yield**  
317 **from cattle.** The solid vertical line represents a mean difference of zero or no effect. Points to  
318 the left of the line represent a reduction in methane yield, while points to the right of the line  
319 indicate an increase. Each square around the point effect represents the mean effect size for

320 that comparison and reflects the relative weighting of the comparison to the overall effect size  
321 estimate. The larger the box, the greater the comparison contribution to the overall estimate.  
322 The weights that each comparison contributed are in the right-hand column. The upper and  
323 lower limit of the line connected to the square represents the upper and lower 95% CI for the  
324 effect size. The overall pooled effects size or SMD and 95% CI pooled using the DerSimonian  
325 and Laird (D+L) [23] and robust meta-analytical random effects models [18, 26] are indicated  
326 by the respective diamonds at the bottom. The heterogeneity measure,  $I^2$  is a measure of  
327 residual variation among comparisons included in the meta-analysis. Methane yield was  
328 considerably heterogeneous as indicated by the overall  $I^2$  of 84.0%.

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## 332 **References**

- 333 1. Roque BM, Venegas ME, Kinley R, deNys R, Neoh TL, Duarte TL, et al. Red seaweed  
334 (*Asparagopsis taxiformis*) supplementation reduces enteric methane by over 80 percent in beef  
335 steers. bioRxiv. 2020. <https://doi.org/10.1101/2020.07.15.204958>
- 336 2. Kinley RD, Martinez-Fernandez G, Matthews MK, de Nys R, Magnusson M, Tomkins  
337 NW. Mitigating the carbon footprint and improving productivity of ruminant livestock  
338 agriculture using a red seaweed. Journal of Cleaner Production. 2020:120836.  
339 <https://doi.org/10.1016/j.jclepro.2020.120836>
- 340 3. Williams J, Spiers D, Thompson-Golden L, Hackman T, Ellersieck M, Wax L, et al.  
341 Effects of Tasco in alleviation of heat stress in beef cattle. Prof Ani Sci. 2009;25:109-17.  
342 [https://doi.org/10.15232/S1080-7446\(15\)30693-8](https://doi.org/10.15232/S1080-7446(15)30693-8)

- 343 4. Abbott DW, Aasen IM, Beauchemin KA, Grondahl F, Gruninger R, Hayes M, et al.  
344 Seaweed and seaweed bioactives for mitigation of enteric methane: Challenges and  
345 opportunities. *Animals*. 2020;10:2432.
- 346 5. Vijn S, Compart DP, Dutta N, Foukis A, Hess M, Hristov AN, et al. Key considerations  
347 for the use of seaweed to reduce enteric methane emissions from cattle. *Frontiers in Veterinary*  
348 *Science*. 2020;7:1135. <https://doi.org/10.3389/fvets.2020.597430> PMID: 33426018
- 349 6. Moher D, Liberati A, Tetzlaff J, Altman DG, The PG. Preferred reporting items for  
350 systematic reviews and meta-analyses: The PRISMA statement. *PLOS Medicine*.  
351 2009;6:e1000097. <https://doi.org/10.1371/journal.pmed.1000097> PMID: 19621072
- 352 7. Allen VG, Pond KR, Saker KE, Fontenot JP, Bagley CP, Ivy RL, et al. Tasco-forage:  
353 III. Influence of a seaweed extract on performance, monocyte immune cell response, and  
354 carcass characteristics in feedlot-finished steers. *J Anim Sci*. 2001;79:1032-40.  
355 <https://doi.org/10.2527/2001.7941032x> PMID: 11325177
- 356 8. Anderson M, Blanton Jr J, Gleghorn J, Kim S, Johnson J. *Ascophyllum nodosum*  
357 supplementation strategies that improve overall carcass merit of implanted English crossbred  
358 cattle. *Asian Austral J Anim*. 2006;19:1514-8. <https://doi.org/10.5713/ajas.2006.1514>
- 359 9. Antaya NT, Ghelichkhan M, Pereira ABD, Soder KJ, Brito AF. Production, milk  
360 iodine, and nutrient utilization in Jersey cows supplemented with the brown seaweed  
361 *Ascophyllum nodosum* (kelp meal) during the grazing season. *J Dairy Sci*. 2019;102:8040-58.  
362 <https://doi.org/10.3168/jds.2019-16478> PMID: 31279546
- 363 10. Carter J, Stovall T, Gill D, Confer A, Smith R, Ball R. Nutritional benefits of feeding a  
364 pelleted supplement manufactured from north atlantic seaweed to transit-stressed feedlot cattle:  
365 Animal performance and medical costs. *Anim Sci Res Report P-980*. 2000:65-9.
- 366 11. Cvetkovic B, Shirley JE, Brouk MJ. Impact of dried seaweed meal on heat-stressed  
367 lactating dairy cattle. *Dairy Day*. 2004:59-61.



- 368 12. Gravett RB. The effects of *Ascophyllum nodosum* on immune function, performance,  
369 and carcass characteristics of sheep and cattle: Masters Thesis Texas Tech University; 2000.
- 370 13. Kidane A, Nesheim IL, Larsen HJ, Thuen E, Jensen SK, Steinshamn H. Effects of  
371 supplementing mid-lactation dairy cows with seaweed and vitamin E on plasma and milk  $\alpha$ -  
372 tocopherol and antibody response to immunization. J Agric Sci. 2015;153:929-42.  
373 <https://doi.org/10.1017/S0021859615000052>
- 374 14. Stefenoni H, Räisänen S, Cueva S, Wasson D, Lage C, Melgar A, et al. Effects of the  
375 macroalga *Asparagopsis taxiformis* and oregano leaves on methane emission, rumen  
376 fermentation, and lactational performance of dairy cows. J Dairy Sci. 2021;104:Article in  
377 Press. <https://doi.org/10.3168/jds.2020-19686> PMID: 33516546
- 378 15. Bendary M, Bassiouni M, Ali M, Gaafar H, Shamas AS. Effect of premix and seaweed  
379 additives on productive performance of lactating friesian cows. Int Res J Agric Sci Soil Sci.  
380 2013;3:174-81.
- 381 16. Sharma A, Datt C. Supplementation effect of red seaweed powder on dry matter intake,  
382 body weight and feed conversion efficiency in crossbred cows. Journal of Entomology and  
383 Zoology Studies. 2020;8:1056-9.
- 384 17. Singh BK, Chopra RC, Rai SN, Verma MP, Mohanta RK. Nutritional evaluation of  
385 seaweed on nutrient digestibility, nitrogen balance, milk production and composition in  
386 sahiwal cows. Proc Natl Acad Sci, India, Sect B Biol Sci. 2015;87:437-43.  
387 <https://doi.org/10.1007/s40011-015-0616-8>
- 388 18. Hedges LV, Tipton E, Johnson MC. Robust variance estimation in meta-regression with  
389 dependent effect size estimates. Res Synth Methods. 2010;1:39-65.  
390 <https://doi.org/10.1002/jrsm.5> PMID: 26056092

- 391 19. Van den Noortgate W, López-López JA, Marín-Martínez F, Sánchez-Meca J. Three-  
392 level meta-analysis of dependent effect sizes. *Behav Res Methods*. 2013;45:576-94.  
393 <https://doi.org/10.3758/s13428-012-0261-6> PMID: 23055166
- 394 20. St-Pierre N. Invited review: Integrating quantitative findings from multiple studies  
395 using mixed model methodology. *J Dairy Sci*. 2001;84:741-55.  
396 [https://doi.org/10.3168/jds.S0022-0302\(01\)74530-4](https://doi.org/10.3168/jds.S0022-0302(01)74530-4) PMID: 11352149
- 397 21. Lean IJ, Rabiee AR, Duffield TF, Dohoo IR. Invited review: Use of meta-analysis in  
398 animal health and reproduction: Methods and applications. *J Dairy Sci*. 2009;92:3545-65.  
399 <https://doi.org/10.3168/jds.2009-2140> PMID: 19620636
- 400 22. Golder HM, Lean IJ. A meta-analysis of lasalocid effects on rumen measures, beef and  
401 dairy performance, and carcass traits in cattle. *J Anim Sci*. 2016;94:306-26.  
402 <https://doi.org/10.2527/jas.2015-9694> PMID: 26812337
- 403 23. DerSimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin Trials*.  
404 1986;7:177-88. [https://doi.org/10.1016/0197-2456\(86\)90046-2](https://doi.org/10.1016/0197-2456(86)90046-2) PMID: 26343745
- 405 24. Knapp G, Hartung J. Improved tests for a random effects meta-regression with a single  
406 covariate. *Stat Med*. 2003;22:2693-710. <https://doi.org/10.1002/sim.1482> PMID: 12939780
- 407 25. White IR, Thomas J. Standardized mean differences in individually-randomized and  
408 cluster-randomized trials, with applications to meta-analysis. *Clinical Trials*. 2005;2:141-51.  
409 <https://doi.org/10.1191/1740774505cn081oa> PMID: 16279136
- 410 26. Tanner-Smith EE, Tipton E. Robust variance estimation with dependent effect sizes:  
411 Practical considerations including a software tutorial in Stata and SPSS. *Res Synth Methods*.  
412 2014;5:13-30. <https://doi.org/10.1002/jrsm.1091> PMID: 26054023
- 413 27. Egger M, Smith GD. Principles of and procedures for systematic reviews. In: Egger M,  
414 Smith GD, G. AD, editors. *Systematic reviews in health care meta-analysis in context*. 23-42.  
415 London: British medical journal books; 2001.

- 416 28. Higgins JPT, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Stat Med*.  
417 2002;21:1539-58. <https://doi.org/10.1002/sim.1186> PMID: 12111919
- 418 29. Borenstein M, Higgins JPT, Hedges LV, Rothstein HR. Basics of meta-analysis:  $I^2$  is  
419 not an absolute measure of heterogeneity. *Res Synth Methods*. 2017;8:5-18.  
420 <https://doi.org/10.1002/jrsm.1230>
- 421 30. Higgins JPT, Green S. *Cochrane handbook for systematic reviews of interventions*  
422 version 5.1.0 [updated march 2011]. 2011 [cited 11/30/16]. The Cochrane Collaboration, [cited  
423 11/30/16]. Available from: [www.cochrane-handbook.org](http://www.cochrane-handbook.org).
- 424 31. Chagas JC, Ramin M, Krizsan SJ. *In vitro* evaluation of different dietary methane  
425 mitigation strategies. *Animals*. 2019;9:1120. <https://doi.org/10.3390/ani9121120> PMID:  
426 31835803
- 427 32. Machado L, Magnusson M, Paul NA, Kinley R, de Nys R, Tomkins N. Identification  
428 of bioactives from the red seaweed *Asparagopsis taxiformis* that promote antimethanogenic  
429 activity *in vitro*. *J Appl Phycol*. 2016;28:3117-26. <https://doi.org/10.1007/s10811-016-0830-7>
- 430 33. Van Nevel C, Demeyer D. Control of rumen methanogenesis. *Environ Monit Assess*.  
431 1996;42:73-97. <https://doi.org/10.1007/BF00394043> PMID: 24193494
- 432 34. Eckard R, Clark H. Potential solutions to the major greenhouse-gas issues facing  
433 Australasian dairy farming. *Anim Prod Sci*. 2020;60:10-6. <https://doi.org/10.1071/AN18574>
- 434 35. National Institutes of Health: Office of Dietary Supplements (NIH). Iodine: Fact sheet  
435 for health professionals 2020 [Available from: [https://ods.od.nih.gov/factsheets/Iodine-  
436 HealthProfessional/](https://ods.od.nih.gov/factsheets/Iodine-HealthProfessional/)].
- 437 36. Nunes N, Valente S, Ferraz S, Barreto MC, de Carvalho MP. Validation of a  
438 spectrophotometric methodology for a rapid iodine analysis in algae and seaweed casts. *Algal*  
439 *Res*. 2019;42:101613. <https://doi.org/10.1016/j.algal.2019.101613>

440 37. Flachowsky G, Franke K, Meyer U, Leiterer M, Schöne F. Influencing factors on iodine  
441 content of cow milk. Eur J Nutr. 2014;53:351-65. <https://doi.org/10.1007/s00394-013-0597-4>  
442 PMID: 24185833

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## 444 **Supporting information**

445 **SI Table. List of references that were rejected at secondary screening and the reasons**

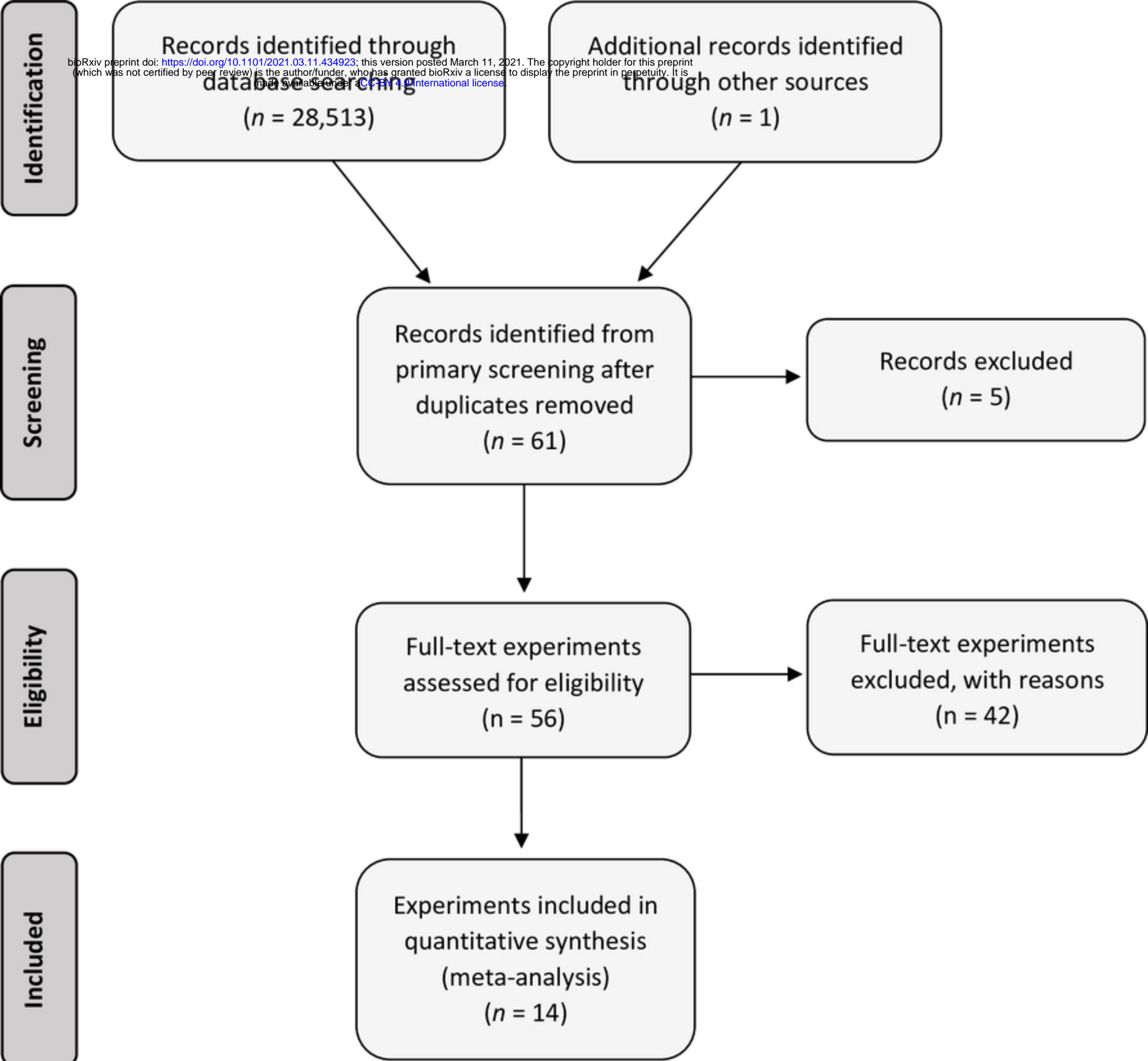


Fig 1

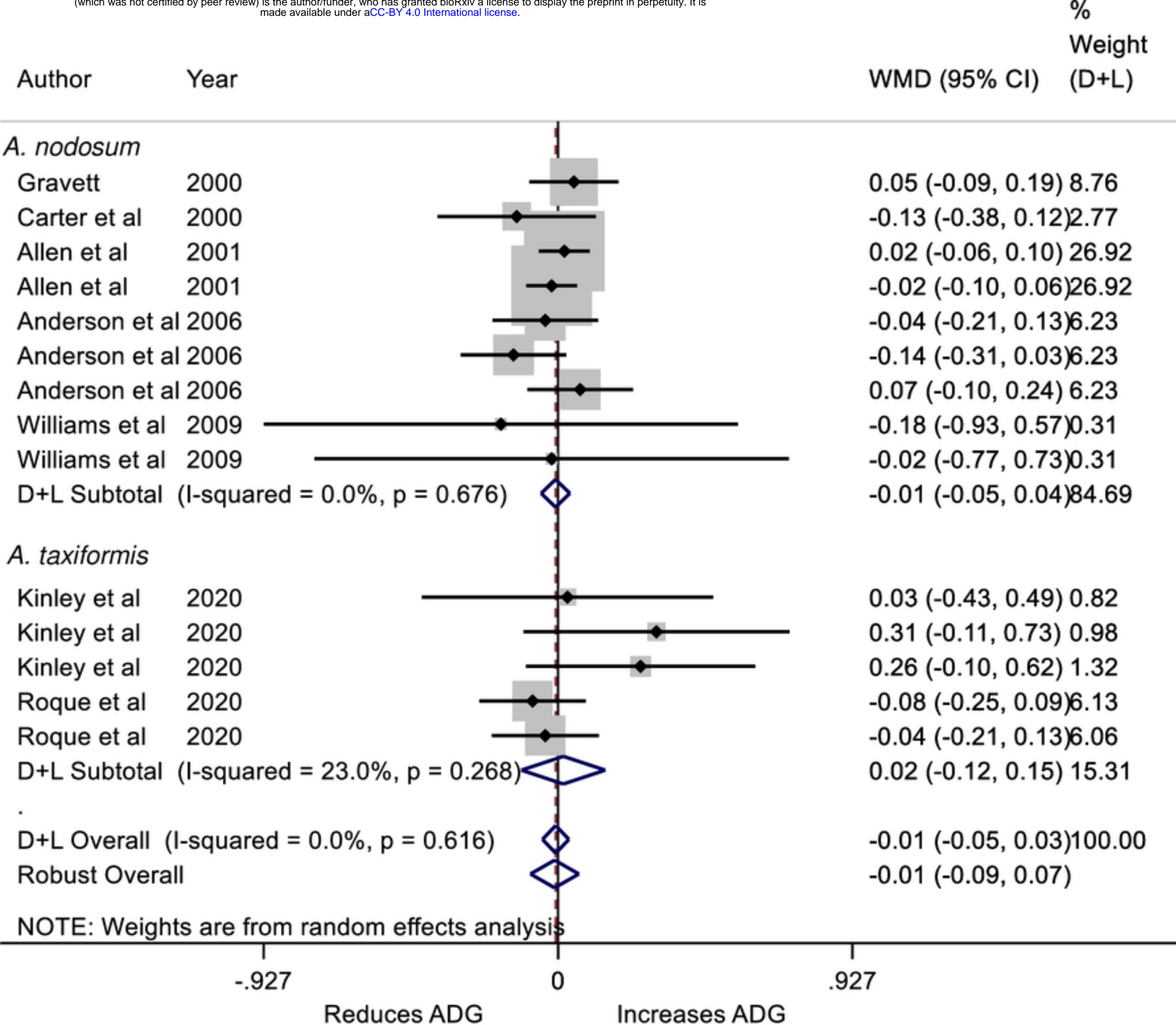


Fig 2

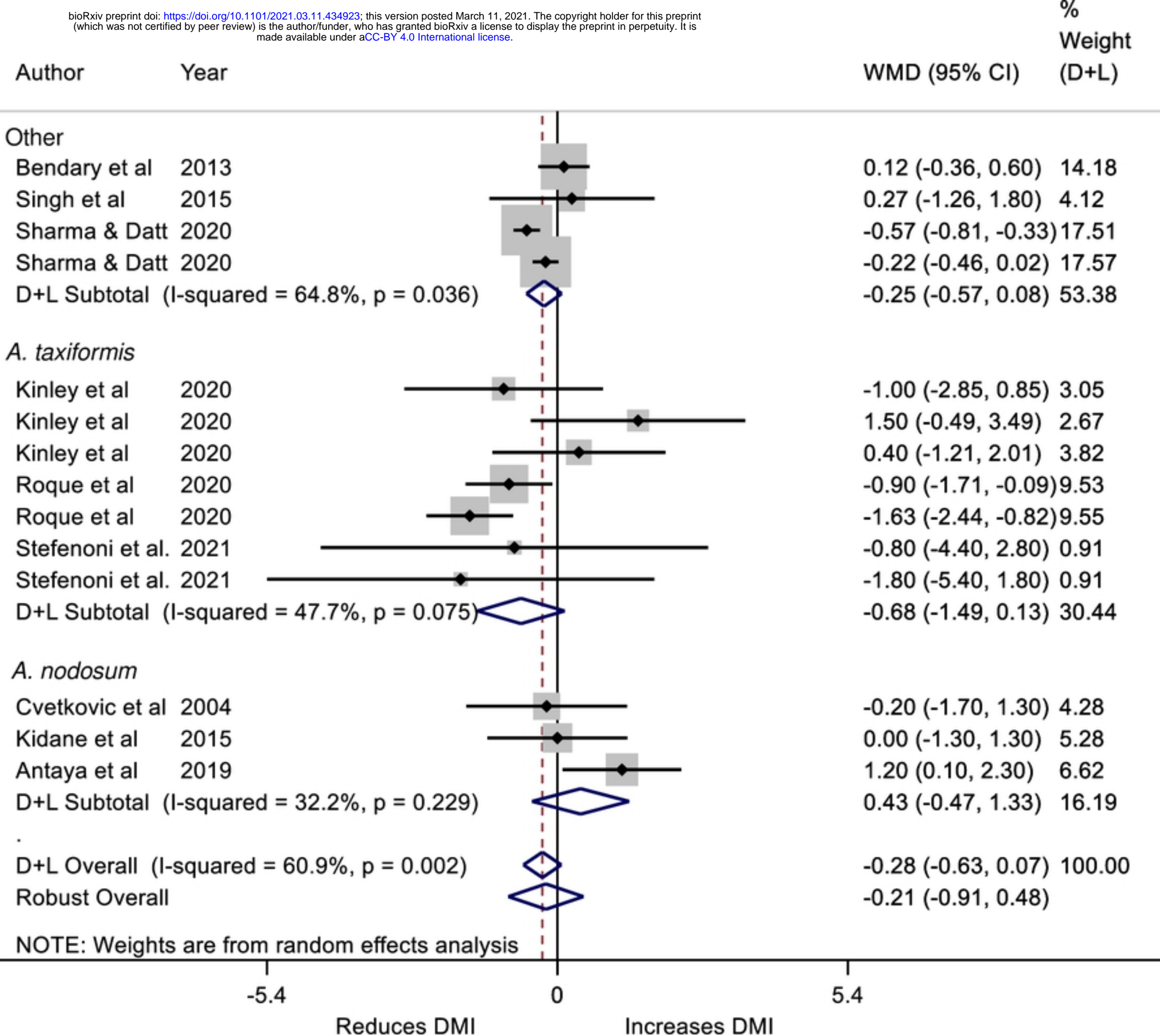


Fig 3

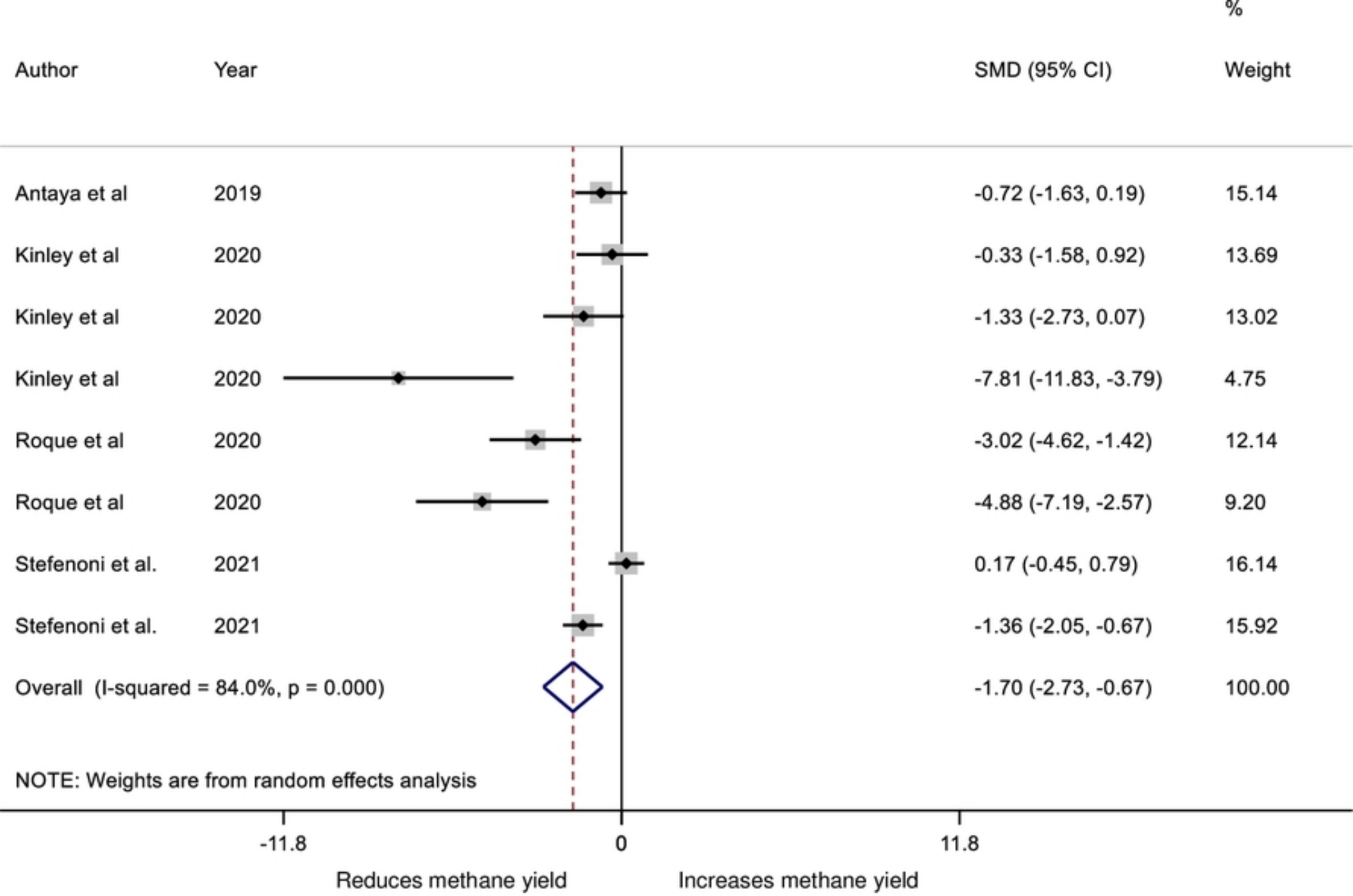


Fig 4