

1 **Asc-Seurat – Analytical single-cell Seurat-based web**
2 **application**

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15 **Abstract**

16 **Summary:** Single-cell RNA sequencing (scRNA-seq) has become a popular approach
17 for studying the transcriptome, providing a powerful tool for discovering and
18 characterizing cell types and their developmental trajectories. However, scRNA-seq
19 analysis is complex, requiring a continuous, iterative process to refine the data
20 processing and uncover relevant biological information. We present Asc-Seurat, a
21 feature rich workbench, providing a user-friendly and easy-to-install web application
22 encapsulating the necessary tools for an all-encompassing and fluid scRNA-seq data
23 analysis.

24 **Availability and implementation:** Asc-Seurat is available at
25 https://github.com/KirstLab/asc_seurat/ and released under GNU 3 license.

26 **Contact:** mkirst@ufl.edu

27 **Supplementary information:** Supplementary data are available at *Bioinformatics*
28 online.

29 **1. Introduction**

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31 Single-cell technologies dramatically enhanced our capacity to characterize
32 tissues and their cell types. By quantifying individual cells' gene expression, single-
33 cell RNA sequencing (scRNA-seq) substantially increases the resolution of
34 transcriptome profiles from whole tissues or organs by disentangling gene
35 expression signals from bulk cell populations down to cell specific contribution (Stark
36 et al. 2019).

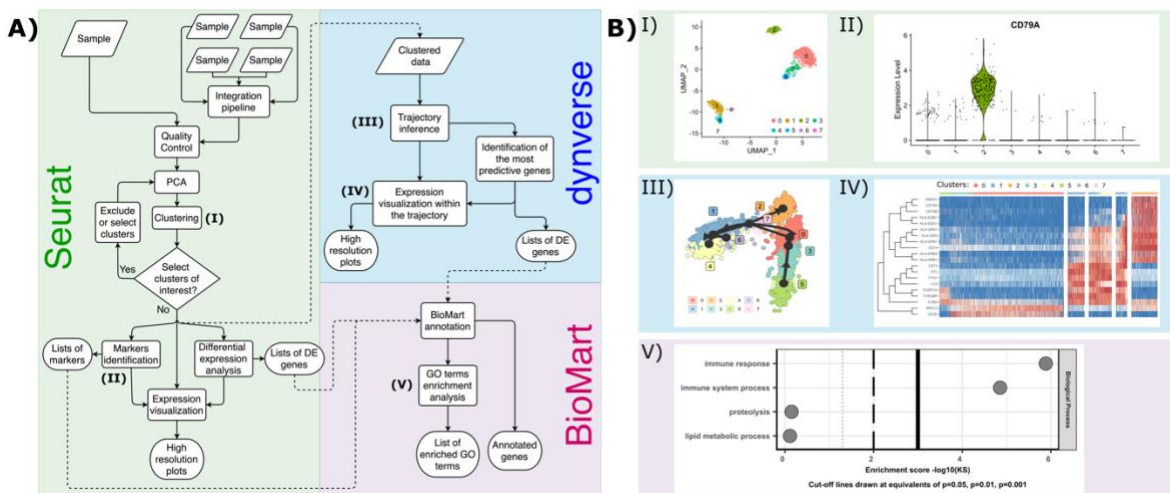
37 Several software has been developed to address the different aspects of
38 scRNA-seq data analysis. Seurat (Stuart *et al.*, 2019) is an R package widely used
39 for scRNA-seq data processing, cell clustering and analysis of differentially

40 expressed genes (DEGs) of single or multiple samples. Further analyses, like single-
41 cell lineage development trajectory inference tools are provided by software such as
42 dynverse (Saelens et al., 2019), in which multiple models can be evaluated and
43 DEGs identified in these trajectories.

44 The Seurat's and dynverse's functions are simple to use, and the results
45 reported are intuitive to interpret. Nevertheless, the dependency on a command-line
46 interface and the need for knowledge of the R programming language poses a major
47 barrier for researchers with limited computational expertise. Moreover, each
48 analytical step outcome is strongly influenced by data quality and execution
49 parameters, requiring the continuous manipulation of these parameters and
50 reevaluation of subsets of data. Tools that simplify this iterative process and
51 integration across analysis platforms are essential for biologists.

52 Here we present Asc-Seurat (Analytical single-cell Seurat-based web
53 application), an easy to install interactive web application implemented using Shiny
54 (<https://shiny.rstudio.com/>). Asc-Seurat provides a comprehensive scRNA-seq
55 analysis workbench (Figure 1-A), with the integration of many algorithmic capabilities
56 from Seurat and dynverse, combined with gene functional annotation using BioMart
57 (Smedley *et al.*, 2009) via the biomaRt package (Durinck *et al.*, 2009).

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60 **Figure 1. Asc-Seurat workflow overview (A) and demonstrative results using the 10x's**

61 **PBMC dataset (B).** Asc-Seurat is built on three analytical cores (A). Using Seurat, users

62 explore scRNA-seq data to identify cell types, markers, and DEGs. Dynverse allows the

63 evaluation and visualization of developmental trajectories and identifies DEGs on these

64 trajectories. Asc-Seurat also implements BioMart for functional annotation and GO terms

65 enrichment analysis. B-I) UMAP plot demonstrating the cluster of cells detected in the PBMC

66 sample. B-II) Visualization of the expression of a gene (CD79A) identified as a marker for

67 cluster 2. B-III) Visualization of the developmental trajectory inferred using slingshot (Street

68 *et al.*, 2018), a model in dynverse. B-IV) Heatmap showing the gene expression of 20 DEGs,

69 ranked by their importance in predicting the trajectory. B-V) Enriched GO terms, in the

70 category biological process, in the set of 50 most important DEGs within the trajectory

71 shown in B-III.

72 **2. Asc-Seurat functionalities**

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74 By employing Asc-Seurat, users have access to 1) a rich and responsive

75 graphical user interface, enabling iterative analysis parametrization; 2) a container-

76 based (Docker, <https://www.docker.com/>) distribution that handles all software

77 dependencies and simplifies installation; 3) integration of multiple samples; 4)

78 selection of cell clusters of interest for reanalysis; 5) search of gene markers and
79 DEG; 6) incorporation of dozens of models for cell trajectory inference; 7)
80 identification of DEGs within trajectories; 8) functional annotation of genes; 9) search
81 for enriched GO terms; and 10) a broad set of publication-ready graphs, among
82 other features.

83 **3. Asc-Seurat use case**

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85 To demonstrate Asc-Seurat's functionalities, we analyzed the publicly available
86 10x's Peripheral Blood Mononuclear Cells (PBMC) dataset (**Figure 1-B**). A stepwise
87 demonstration can be found in the **Supplementary material**.

88 **3.1. Quality control, clustering, and markers identification**

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90 Asc-Seurat workflow starts with raw scRNA-seq data input and provides several
91 options to exclude poor quality cells. Violin plots are generated to show the
92 distribution of cells before and after filtering (**Figure S2**). Expression levels can be
93 normalized and scaled (Figure S3), followed by clustering and cluster visualization
94 using UMAP and t-SNE (**Figure 1.B-I and Figure S4**). After clustering, Asc-Seurat
95 simplifies the selection and reanalysis of subsets of cells from clusters of interest.
96 Moreover, the expression of genes can be visualized at the individual cell (**Figure**
97 **S7**) or cluster level (**Figure 1.B-II and Figure S8**). Asc-Seurat also implements
98 methods to identify cell-type markers and genes differentially expressed among
99 clusters (**Figure S5**).

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101 **3.2. Trajectory inference and differential expression**

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103 Several algorithms are available on Asc-Seurat to model and infer developmental
104 trajectories among cell clusters (**Figure 1.B-III and Figure S11**), allowing the
105 visualization of gene expression in individual cells within the trajectory (**Figure S13**)
106 and the identification of DEGs along the trajectory (**Figure 1.B-IV, Figure S12**).

107 **3.3. Functional annotation and enrichment analysis**

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109 Annotation of genes of interest (e.g., markers, DEGs) can be retrieved by the
110 Asc-Seurat BioMart module. Moreover, it is possible to search for enriched Gene
111 Ontology (GO) terms (**Figure 1.B-V and Figure S15**) using topGO (Alexa and
112 Rahnenfuhrer, 2020; Bioconductor). These capabilities allow the biological
113 interpretation of scRNA-seq results and real-time hypotheses generation.

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120 **Conflict of Interest:** none declared.

121 **Data availability**

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123 The 10x's PBMC dataset is publicly available at

124 https://cf.10xgenomics.com/samples/cell/pbmc3k/pbmc3k_filtered_gene_bc_matrice

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