Journal: Molecular Ecology Resources 2 Subject area: Computer Programs SPART, a versatile and standardized data exchange format 3 for species partition information 4 5 Aurélien Miralles^{1*}, Jacques Ducasse², Sophie Brouillet¹, Tomas Flouri³, Tomochika Fujisawa⁴, Paschalia Kapli³, L. Lacey Knowles⁵, Sangeeta Kumari⁶, Alexandros 7 Stamatakis^{7,8}, Jeet Sukumaran⁹, Sarah Lutteropp⁷, Miguel Vences⁶*, Nicolas Puillandre¹* 9 ¹Institut de Systématique, Évolution, Biodiversité (ISYEB), Muséum national d'Histoire naturelle, CNRS, 10 Sorbonne Université, EPHE, Université des Antilles 57 rue Cuvier, CP 50, 75005 Paris, France 11 12 ²49 rue Eugène Carrière, 75018 Paris, France ³Centre for Life's Origins and Evolution, Department of Genetics, Evolution and Environment, University 13 14 College London, London WC1E 6BT, UK ⁴Center for Data Science and Education and Research, Shiga University, 1-1-1 Banba, Hikone, 522-8522, Shiga, 15 16 Japan 17 ⁵Department of Ecology and Evolution, University of Michigan, Ann Arbor, MI 48109, USA 18 ⁶Braunschweig University of Technology, Zoological Institute, Mendelssohnstraße 4, 38106 Braunschweig, 19 Germany 20 ⁷Computational Molecular Evolution Group, Heidelberg Institute for Theoretical Studies, Schloss-Wolfsbrunnenweg 35, 69118 Heidelberg Germany 21 22 ⁸Institute for Theoretical Informatics, Karlsruhe Institute of Technology, Am Fasanengarten 5, 76131 Karlsruhe, 23 Germany 24 ⁹Biology Department, LS 262, San Diego State University, 5500 Campanile Drive, San Diego, CA 92182-4614, 25 USA 26 *Corresponding authors 27 28 Contact: miralles.skink@gmail.com, m.vences@tu-braunschweig.de, nicolaspuillandre@gmail.com 29 30 **Short running title:** SPART, a standardized format for species partition

Abstract

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A wide range of data types can be used to delimit species and various computer-based tools dedicated to this task are now available. Although these formalized approaches have significantly contributed to increase the objectivity of SD under different assumptions, they are not routinely used by alpha-taxonomists. One obvious shortcoming is the lack of interoperability among the various independently developed SD programs. Given the frequent incongruences between species partitions inferred by different SD approaches, researchers applying these methods often seek to compare these alternative species partitions to evaluate the robustness of the species boundaries. This procedure is excessively time consuming at present, and the lack of a standard format for species partitions is a major obstacle. Here we propose a standardized format, SPART, to enable compatibility between different SD tools exporting or importing partitions. This format reports the partitions and describes, for each of them, the assignment of individuals to the "inferred species". The syntax also allows to optionally report support values, as well as original trees and the full command lines used in the respective SD analyses. Two variants of this format are proposed, overall using the same terminology but presenting the data either optimized for human readability (matricial SPART) or in a format in which each partition forms a separate block (SPART.XML). ABGD, DELINEATE, GMYC, PTP and TR2 have already been adapted to output SPART files and a new version of LIMES has been developed to import, export, merge and split them. **Key words.** Species delimitation programs; SPART; Species partition format; Integrative taxonomy; LIMES v2.0

Introduction

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Species delimitation (SD) is a burgeoning, fully fledged research field in systematic biology (Sites & Marshall 2003; Camargo & Sites 2013; Flot 2015, Ducasse et al. 2020). SD benefits from the interpretation of species as independent evolutionary lineages (De Queiroz 1998, 2007) that can be distinguished from each other using a variety of operational SD criteria (Samadi & Barberousse 2006). In integrative taxonomy (Dayrat 2005; Padial et al. 2010), various lines of evidence and a wide range of data types can be used in formalised analytical workflows to propose species hypotheses, from DNA barcodes to phylogenomic data, discrete morphological characters, morphometric measurements, ecological traits, geographic occurrence, bioacoustic signals, metabolomic profiles, and others (Miralles et al. 2020). If many, and among them the earliest, formalised SD procedures are mostly carried out manually, e.g. by comparing trees with the geographic occurrence of individuals, calculating correlations between geographic and genetic distances, assessing steepness of hybrid zones, or seeking for correlation between genetic distance and morphological characters (Good & Wake 1992, Wiens & Penkrot 2002, Vieites et al. 2009, Flot et al. 2010, Weisrock et al. 2010, Puillandre et al. 2012a, Miralles & Vences 2013, Derkarabetian & Hedin 2014, Dufresnes et al. 2015), a substantial number of computer-based tools has been developed to delimit species, often based on statistical criteria. These programs can analyse large datasets, with a strong focus on the use of sequence data (Table 1). These methods have significantly contributed to increase the objectivity, repeatability, and speed of species delimitation inferences under different mathematical models and assumptions (e.g. Multispecies coalescent model, DNA barcode gap, haplotype fields of recombination, cf. de Queiroz 1998,

79 2007, Knowles & Carstens 2007, Yang & Rannala 2010, Carstens et al. 2013, Leavitt et al. 80 2015, Rannala 2015). 81 82 Although the number and importance of SD tools is likely to sharply increase in the immediate future, they are not yet routinely used in the majority of alpha-taxonomic studies 83 84 that result in the naming of over 15,000 new species of organisms every year (Miralles et al. 2020). One obvious shortcoming is the lack of interoperability among the various 85 86 independently developed SD programs, and the lack of comprehensive software suites that 87 offer various user-friendly features, such as those for data visualization and comparison of 88 results across methods. For instance, incongruent species partitions resulting from different 89 SD approaches applied to a given dataset are common. They can even be significant, if not 90 striking in some cases (such as excessive splitting or lumping leading to highly different 91 number of species delimited; Carstens et al. 2013, Miralles & Vences 2013, Dellicour & Flot 92 2015, Kapli et al. 2016, Postaire et al. 2016, Renner et al. 2017 for empirical cases; and Sukumaran & Knowles 2017, Chan et al 2020, Luo et al. 2018, Mason et al. 2020 and Zhang 93 94 et al. 2011 for more methodological studies on SD limitations). Integrative taxonomists will 95 seek to compare these alternative species partitions across SD approaches (but see Rannala 96 2015), and eventually estimate their robustness by integrating other data sources 97 (morphological variation, geographic distribution, etc), in order to make an informed choice-98 a procedure that is excessively time consuming at present, given the lack of a standard format 99 for species partitions.

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The main output of species delimitation, and therefore of any SD program, is a species partition. The term "partition" here follows the set theory concept: the organization of a set of elements into mutually-exclusive and jointly-comprehensive subsets, not including the empty subset (Hrbacek & Jech 1999). In an SD application, the elements are individuals (i.e. samples or specimens), and a specific species delimitation hypothesis is a particular assignment (i.e. a partition) of these individuals to subsets, where each subset corresponds to a distinct inferred species. Categories resulting from an SD analysis have been referred to by various terms, such as primary species hypothesis, operational taxonomic unit (OTUs), barcode index number (BINs; Ratnasingham & Hebert 2013), or even cluster (without any particular status (Fig. 1)), but all of them match the aforementioned definition of a subset. Furthermore, while some tools produce de novo species partitions (i.e. directly aggregating individuals into species hypotheses; exploratory methods), others statistically compare competing species hypotheses that have been defined a priori (hypothesis-testing methods), and these programs require a species partition as input. SD methods may also assign scores, either to the entire inferred partition (e.g., ASAP-score in the program ASAP; Puillandre et al. 2021), to the distinctiveness of each subset from the others (e.g., posterior probabilities in the programs BPP and bPTP; Yang & Rannala 2010; Zhang et al. 2013), or to the presence of each individual in a given subset (e.g., probability of placement in calculation of BINs, Ratnasingham & Hebert 2013).

A standardized Species PARTition format (SPART)

Typically, each SD program exports the resulting species partitions in its own idiosyncratic format. Some, for instance, provide a table of assignments of individual specimens to the

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subsets (e.g. GMYC) while others, conversely, list the different subsets with the included individuals (e.g. ABGD, PTP), whereas again others graphically report subsets on a tree topology (e.g. GMYC). These different formats may or may not include complementary data (e.g., scores, topologies, metadata, number of species delimited, etc.), and are not designed to be parsed by other tools for downstream analyses. Their manual conversion into a versatile and easily reusable plain text species partition (e.g., CSV) is not always straightforward. It can be particularly error prone and time consuming with large datasets, as species delimitations on several hundreds, or even thousands, of specimens are becoming common practice in molecular taxonomy (e.g., Ahrens et al. 2016, Renner et al 2017, Garcià-Melo et al. 2019, Hoffmann et al. 2019, Solihah et al. 2020, Christodoulou et al. 2020). We here propose a standardized species partition format, SPART, to enable compatibility between different tools producing (export) or using (import) species partitions. Our format facilitates: (1) statistical comparison of different alternative species partitions such as their overall congruence, similarity or resolving power, identification of the subsets that are congruently delimited (currently implemented in the program LIMES v2.0; Ducasse et al. 2020); (2) assessment of multiple competing SD hypotheses, including those used as input in e.g. DELINEATE and BPP to evaluate them (Sukumaran et al. 2020, Yang & Rannala 2010); (3) visualization and comparison of species partitions (e.g., DNA-based species partitions compared with manually-edited species partitions obtained from alternative methods and data

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such as Principal Component Analysis of morphometry, haplotype networks, geographic distribution, habitat type, external phenetic similarity, or simply, current taxonomy); (4) extraction, from original data files, of specific data for each subset under different species partition assumptions (e.g. lists of molecular and morphological diagnostic character states, descriptive statistics characterizing each of the inferred species, or ecological or distributional traits); and (5) potential taxonomic reassignment of specimens in databases. More generally, the SPART format is designed to be versatile and fully integrative in the sense that it can include any species partition descriptors, independently of the method or data-type used to generate the species partition (Fig. 2). SPART does not convey any interpretation on the quality of the species partition, nor on the pros and cons of the methods used to define them, but is simply a common format that seeks at homogenising the way species partitions are recorded. It can therefore be implemented in any method used to generate one or several species partitions as output. Likewise, any method using (analysing, comparing, automatically reassigning or graphically representing) multiple subsets of specimens might benefit from being able to import SPART files as input data. Matricial and serial implementation of the SPART format

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SPART files include information on one or multiple species partitions for a given set of elements (i.e. individuals) and use standardized terminology to denote the number of species partitions included in the file ("N_spartitions") and for each partition, the number of individuals ("N_individuals"), number of subsets ("N_subsets"), and the assignment of individuals to subsets ("Assignment") (Supporting information 1). The syntax also allows to optionally include support values for species partitions, subsets, and the assignment of individuals to subsets, as well as original trees and the full command line used in the respective SD analyses, the program version number as well as comments and species partition comparison indices as calculated with LIMES 2.0, a new version of LIMES (Ducasse et al. 2020) recently published. To account for the diversity of possible future applications, we propose two variants of the SPART format (for details see Supporting information 1). Both of these use largely the same terminology but represent the data differently: The first SPART variant is optimized for human readability and its syntax has been designed to be compatible with Nexus (a widely used data format in phylogenetic inference software: Maddison et al. 1997). This allows to include SPART specifications as blocks in Nexus files if required by future applications. If information from multiple partitions is included, then it is combined into a single block, presenting the respective assignments and assignment scores per individual from different species partitions concatenated on a single line, separated by separator symbols. This enables easy manual transformation into a spreadsheet format if required. Due to the presentation of information from multiple partitions in one block as a concatenated matrix, we denote this variant as matricial SPART format, or simply SPART.

The second SPART variant is optimized for machine readability, and relies on XML (eXtensible Markup Language), a lightweight data-interchange format that can be easily parsed and written by software tools, while it can still be read and written by humans as well. When information from multiple partitions is included, each partition forms a separate block containing information on the number of subsets, individual assignments and assignment scores. We therefore denote this variant as *SPART.XML* format.

Tools already implementing SPART and future perspectives

The proposed format is already implemented in several widely-used SD programs. Both the matricial SPART and SPART.XML output files are already generated by GUI-driven standalone versions (https://github.com/iTaxoTools) of ABGD, ASAP, GMYC, PTP, mPTP, TR2 and DELINEATE (Vences et al. submitted), by the native Python version of TR2, and in the web versions of ABGD and ASAP; and in progress for the Python versions of GMYC and PTP. Furthermore, the species partition comparison tool LIMES v2.0 has been expanded to import, export and convert SPART files, in particular to (1) compare, by calculating indices (e.g., Ctax, Ratx, Match Ratio, cf. Ducasse et al. 2020) for species partitions from SPART files (including each one or several species partitions); (2) merge species partitions included in different SPART files into one SPART file, (3) import species partition(s) table(s) from spreadsheet editors such as Microsoft EXCEL and save it (them) into a single SPART file. A new software tool named SPARTMAPPER has also been developed; it takes SPART files as input along with a tab-delimited series of geographical coordinates linked to specimen names, plots the distribution of alternative delimited species on a map, and exports a .kml file to visualize this information in Google Earth.

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note to reviewers: all of these new software versions will be released before or upon publication of this manuscript, and the web based ABGD and ASAP are already freely available since February 2021 (https://bioinfo.mnhn.fr/abi/public/abgd/abgdweb.html, https://bioinfo.mnhn.fr/abi/public/asap/); for review, various preliminary Windows executables are available under this link: https://hidrive.ionos.com/share/ohaymwcgjd#\$/Win%20executables] In the context of future work, we envisage the development of visualization tools to automatically illustrate information from species partitions along with support values and phylogenetic hypotheses (Fig. 1). There is still a long way to go before programs will be able to infer species based on combining evidence using different data sources such as genetics, morphology, ecology, behaviour, geographic distribution, etc. However, eventually, reliable computer-based, species delimitation procedures that mirror the procedures of integrative taxonomy will be at the core of next generation taxonomy (Vences 2020). Our SPART data exchange format would thus contribute to this next generation taxonomy, by simplifying computational approaches to completing the inventory of life on Earth. Acknowledgments We are grateful to Susanne Renner who stimulated this work by leading the priority program SPP 1991 "Taxon-Omics" of the Deutsche Forschungsgemeinschaft (DFG), specifically in the context of a grant on taxonomic data integration and management (RE 603/29-1), and to many members of the Taxon-Omics consortium and Guillaume Achaz (MNHN) for fruitful discussion. MV and SK were supported by DFG grant VE247/20-1, NP by the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation programme (grant agreement No. 865101), and AS and SL by the Klaus-Tschira foundation.

242 243 **REFERENCES** 244 245 246 247 Ahrens, D., Fujisawa, T., Krammer, H. J., Eberle, J., Fabrizi, S., & Vogler, A. P. (2016). 248 Rarity and incomplete sampling in DNA-based species delimitation. Systematic Biology, 65, 478–494, doi:10.1093/sysbio/syw002 249 Camargo, A., & Sites, J. Jr. (2013). Species delimitation: a decade after the renaissance. In: 250 The Species Problem - Ongoing Issues (ed. I. Y. Pavlinov). IntechOpen. 251 Carstens, B. C., Pelletier, T. A., Reid, N. M., & Satler, J. D. (2013). How to fail at species 252 delimitation. Molecular Ecolology, 22, 4369–4383. doi:10.1111/mec.12413 253 254 Chan, K. O., Hutter, C. R., Wood, P. L. Jr., Grismer, L. L., Das, I., & Brown, R. M. (2020). 255 Gene flow creates a mirage of cryptic species in a Southeast Asian spotted stream frog complex. Molecular Ecology, 29(20), 3970-3987. doi:10.1111/mec.15603 256 Christodoulou, M., O'Hara, T., Hugall, A. F., Khodami, S., Rodrigues, C. F., Hilario, A., 257 258 Vink, A., & Martinez Arbizu, P. (2020) Unexpected high abyssal ophiuroid diversity in polymetallic nodule fields of the northeast Pacific Ocean and implications for 259 conservation, Biogeosciences, 17, 1845–1876. doi:10.5194/bg-17-1845-2020 260 Dayrat, B. (2005). Toward integrative taxonomy. Biological Journal of the Linnean Society, 261 85, 407–415. doi:10.1111/j.1095-8312.2005.00503.x 262 Dellicour, S., & Flot J.-F. (2015). Delimiting species-poor data sets using single molecular 263 markers: a study of barcode gaps, haplowebs and GMYC. Systematic Biology, 64, 900-264 908. doi:10.1093/sysbio/syu130 265 de Queiroz, K. (1998). The general lineage concept of species, species criteria, and the 266 process of speciation. In: D.J. Howard & S.H. Berlocher, S.H. (Eds.), Endless Forms: 267 Species and Speciation. (pp. 57-75). New York: Oxford University Press. 268 de Queiroz, K. (2007). Species concepts and species delimitation, Systematic Biology, 56, 269 879-886. doi:10.1080/10635150701701083 270 271 Derkarabetian, S., & Hedin, M. (2014). Integrative taxonomy and species delimitation in harvestmen: a revision of the western North American genus Sclerobunus (Opiliones: 272 Laniatores: Travunioidea). PLoS One, 9, e104982. doi:10.1371/journal.pone.0104982 273 Ducasse, J., Ung, V., Lecointre, G., Miralles, A. (2020). LIMES: a tool for comparing 274 species partition. Bioinformatics, 2282–2283. doi:10.1093/bioinformatics/btz911 275 Dufresnes, C., Brelsford, A., Crnobrnja-Isailović, J., Tzankov, N., Lymberakis, P., & Perrin, 276 N. (2015). Timeframe of speciation inferred from secondary contact zones in the European 277 278 tree frog radiation (Hyla arborea group). BMC Evolutionary Biology 15, 1-8. doi: 279 10.1186/s12862-015-0385-2 Ence, D.D., & Carstens, B.C. (2011). SpedeSTEM: A rapid and accurate method for species 280 281 delimitation. Molecular Ecolology Resources, 11, 473–480. doi:10.1111/j.1755-282 0998.2010.02947.x

- Flot, J.-F., Couloux, A., & Tillier, S. (2010). Haplowebs as a graphical tool for delimiting species: a revival of Doyle's "field for recombination" approach and its application to the
- coral genus *Pocillopora* in Clipperton. *BMC Evolutionary Biology*, 10, 372.
- doi:10.1186/1471-2148-10-372

- Flot, J.-F. (2015). Species delimitation's coming of age, *Systematic Biology*, 64, 897–899.
- Fontaneto, D., Herniou, E., Boschetti, C., Caprioli, M., Melone, G., Ricci, C., & Barraclough,
- T.G. (2007). Independently evolving species in asexual bdelloid rotifers. *PLoS Biology*, *5*, e87. doi:10.1371/journal.pbio.0050087
- 291 Fujisawa, T., Aswad, A., Barraclough, T. G. (2016). A rapid and scalable method for
- multilocus species delimitation using Bayesian model comparison and rooted triplets.
- 293 *Systematic Biology*, 65(5), 759–771. doi:10.1093/sysbio/syw028
- García-Melo, J. E., Oliveira, C., Da Costa Silva, G. J., Ochoa-Orrego, L. E., Garcia Pereira, L.
- 295 H., Maldonado-Ocampo, J. A. (2019). Species delimitation of Neotropical characins
- 296 (Stevardiinae): Implications for taxonomy of complex groups. *PLoS ONE 14*(6),
- 297 e0216786. doi:10.1371/journal.pone.0216786
- Good, D. A., & Wake, D. B. (1992). Geographic variation and speciation in the torrent
- salamanders of the genus *Rhyacotriton* (Caudata: Rhyacotritonidae). *University of California Publications in Zoology*, *126*, 1–91.
- 301 Hofmann, E. P., Nicholson, K. E., Luque-Montes, I. R., Köhler, G., Cerrato-Mendoza, C. A.,
- Medina-Flores, M., Wilson, L. D., & Townsend, J. H. (2019). Cryptic diversity, but to
- 303 what extent? Discordance between single-locus species delimitation methods within
- mainland anoles (Squamata: Dactyloidae) of Northern Central America. *Frontiers in*
- 305 Genetics. 10,11. doi:10.3389/fgene.2019.00011
- 306 Hrbacek, K. & Jech, T. (1999). Introduction to set theory, third edition, revised and expanded.
- Monographs and Textbooks in pure and applied mathematics, vol. *220*, Marcel Dekker Inc. Ney York, Basel.
- 309 Jones, G. (2017). Algorithmic improvements to species delimitation and phylogeny estimation
- under the multispecies coalescent. Journal of Mathematical Biology, 74, 447.
- 311 doi:10.1007/s00285-016-1034-0
- 312 Jones, G., Aydin, Z., & Oxelman, B. (2014). DISSECT: An assignment free Bayesian
- discovery method for species delimitation under the multispecies coalescent.
- 314 *Bioinformatics*, 31, 991–998. doi:10.1093/bioinformatics/btu770
- Kapli, P., Lutteropp, S., Zhang, J., Kobert, K., Pavlidis, P., Stamatakis, A., & Flouri, T.
- 316 (2016). Multi-rate Poisson Tree Processes for single-locus species delimitation under
- Maximum Likelihood and Markov Chain Monte Carlo. *Bioinformatics*, 33, 1630–1638.
- doi:10.1093/bioinformatics/btx025
- Knowles, L. L., Carstens, B. C. (2007) Delimiting species without monophyletic gene trees.
- 320 *Systematic Biology*, 56 (6), 887–895. doi:10.1080/10635150701701091
- 321 Leavitt, S. D., Moreau, C. S., & Lumbsch, H. T. (2015). The dynamic discipline of species
- delimitation: Progress toward effectively recognizing species boundaries in natural
- populations. In *Recent Advances in Lichenology* (pp. 11–44). New Delhi: Springer.
- 324 doi:10.1007/978-81-322-2235-4_2
- Luo, A., Ling, C., Ho, S. Y. W., Zhu, C.-D. (2018). Comparison of methods for molecular
- species delimitation across a range of speciation scenarios. Systematic Biology, 67(5), 830–
- 327 846. <u>doi:10.1093/sysbio/syy011</u>
- 328 Maddison, D. R., Swofford, D. L., Maddison, W. P. (1997). NEXUS: An extensible file
- format for systematic information. Systematic Biology, 46, 590–621.
- 330 <u>doi:10.1093/sysbio/46.4.590</u>
- 331 Mason, N. A., Fletcher, N. K., Gill, B. A., Funk, C., Zamudio, K. R. (2020). Coalescent-based
- species delimitation is sensitive to geographic sampling and isolation by distance.
- 333 Systematics and Biodiversity, 18(3), 269–280. doi:10.1080/14772000.2020.1730475

- Masters, B. C., Fan, V., & Ross, H. A. (2011). Species Delimitation a Geneious plugin for
- the exploration of species boundaries. *Molecular Ecology Resources*, 11, 154–157.
- 336 doi:10.1111/j.1755-0998.2010.02896.x
- 337 Miralles, A. & Vences, M. (2013). New metrics for comparison of taxonomies reveal striking
- discrepancies among species delimitation methods in *Madascincus* lizards. *PlosONE*, 8,
- e68242. doi:10.1371/journal.pone.0068242
- 340 Miralles, A., Bruy, T., Wolcott, K., Scherz, M. D., Begerow, D., Beszteri, B., Bonkowski, M.,
- Felden, J., Gemeinholzer, B., Glaw, F., Glöckner, F. O., Hawlitschek, O., Kostadinov, I.,
- Nattkemper, T. W., Printzen, C., Renz, J., Rybalka, N., Stadler, M., Weibulat, T., Wilke,
- T., Renner, S., Vences, M. (2020). Repositories for taxonomic data: where we are and what
- is missing. *Systematic Biology*, 69, 1231–1253. doi:10.1093/sysbio/syaa026
- Monaghan, M. T., Wild, R., Elliot, M., Fujisawa, T., Balke, M., Inward, D. J., Lees, D. C.,
- Ranaivosolo R., Eggleton, P., Barraclough, T.G., & Vogler, A.P. (2009). Accelerated
- species inventory on Madagascar using coalescent-based models of species delineation.
- 348 *Systematic Biology*, 58, 298–311. doi:10.1093/sysbio/syp027
- Padial, J.M., Miralles, A., De la Riva, I., & Vences, M. (2010). The integrative future of taxonomy. *Frontiers in Zoology*, *7*, 16. doi:10.1186/1742-9994-7-16
- Pons, J., Barraclough, T. G., Gomez-Zurita, J., Cardoso, A., Duran, D. P., Hazell, S.,
- Kamoun, S., Sumlin, W. D., & Vogler, A. P. (2006). Sequence-based species delimitation
- for the DNA taxonomy of undescribed insects. *Systematic Biology*, 55, 595–609.
- 354 <u>doi:10.1080/10635150600852011</u>
- Postaire B., Magalon H., Bourmaud C. A., & Bruggemann J. H. (2016). Molecular species
- delimitation methods and population genetics data reveal extensive lineage diversity and
- 357 cryptic species in Aglaopheniidae (Hydrozoa). Molecular Phylogenetics and Evolution,
- 358 *105*, 36–49. doi:10.1016/j.ympev.2016.08.013
- Puillandre, N., Lambert, A., Brouillet, S., & Achaz, G. (2012b). ABGD, Automatic Barcode Gap Discovery for primary species delimitation, *Molecular Ecology*, *21*, 1864–1877.
- 361 doi:10.1111/j.1365-294X.2011.05239.x
- Puillandre, N., Modica, M. C., Zhang, Y., Sirovich, L., Boisselier, M. C., Cruaud, C.,
- Holford, M., Samadi, S. (2012a). Large-scale species delimitation method for hyperdiverse
- groups. *Molecular Ecology*, *11*, 2671–3691. doi:10.1111/j.1365-294X.2012.05559.x
 Puillandre, N., Brouillet, S., Achaz, G. (2021). ASAP: Assemble Species by Automatic
- 366 Partitioning. *Molecular Ecology Resources*, 21(2), 609–620. doi:10.1111/1755-0998.13281
- Rabiee, M., Mirarab, S. (2019). SODA: Multi-locus species delimitation using quartet frequencies. *bioRxiv*, 869396. doi:10.1101/869396
- Rannala, B. (2015). The art and science of species delimitation. *Current Zoology*, 61, 846–853. doi:10.1093/czoolo/61.5.846
- Ratnasingham, S., & Hebert P.D.N. (2013). A DNA-based registry for all animal species: the
- Barcode Index Number (BIN) system. *PLoS ONE*, 8: e66213.
- 373 <u>doi:10.1371/journal.pone.0066213</u>
- Renner, M.A., Heslewood, M.M., Patzak, S.D., Schäfer-Verwimp, A., & Heinrichs J. (2017).
- By how much do we underestimate species diversity of liverworts using morphological
- evidence? An example from Australasian *Plagiochila* (Plagiochilaceae:
- Jungermanniopsida). Molecular Phylogenetics and Evolution, 107, 576–593.
- 378 doi:10.1016/j.ympev.2016.12.018
- 379 Samadi, S., & Barberousse, A. (2006). The tree, the network, and the species. *Biological*
- *Journal of the Linnean Society*, 89(3), 509-521. doi:10.1111/j.1095-8312.2006.00689.x

- 381 Sites, J. W., & Marshall J. C. (2003). Delimiting species: a Renaissance issue in systematic
- biology. Trends in Ecology and Evolution, 18, 462–470. doi:10.1016/S0169-382
- 383 5347(03)00184-8
- 384 Sholihah, A., Delrieu-Trottin, E., Sukmono, T., Dahruddin, H., Risdawati, R., Elvira, R.,
- 385 Wibowo, A., Kusno, K., Busson, F., Sauri, S., Nurhaman, U., Zein, M. S. A., Fitriana, Y.,
- 386 Utama, I., Muchlisin, Z. A., Agnèse, J. F., Hanner, R., Wowor, D., Steinke, D., Keith, P.,
- Rüber, L., Hubert, N., (2020). Disentangling the taxonomy of the subfamily Rasborinae 387
- (Cypriniformes, Danionidae) in Sundaland through DNA barcodes. Scientific Reports, 10, 388 389 2818. doi:10.1038/s41598-020-59544-9
- Solís-Lemus, C., Knowles, L.L., & Ané, C. (2015). Bayesian species delimitation combining 390 multiple genes and traits in a unified framework. Evolution, 69, 492–507. 391 doi:10.1111/evo.12582 392
- Spöri, Y., & Flot, J. □ F. (2020). HaplowebMaker and CoMa: two web tools to delimit species 393 394 using haplowebs and conspecificity matrices. Methods in Ecology and Evolution, 11(11), 395 1434-1438. doi:10.1111/2041-210X.13454
- Sukumaran, J., & Knowles, L. (2017). Multispecies coalescent delimits structure, not species. 396 397 *Proceedings of the National Academy of Sciences USA*, 114(7), 1607–1612. 398 doi:10.1073/pnas.1607921114
- 399 Sukumaran, J., Holder, T. M., Knowles, L. L. (2020). Incorporating the speciation process into species delimitation. https://github.com/jeetsukumaran/delineate. 400
- 401 Vences, M. (2020). The promise of next-generation taxonomy. *Megataxa*, 1, 35–38. 402 doi:10.11646/megataxa.1.1.6
- Vences, M., Miralles, A., Brouillet, S., Ducasse, J., Fedosov, A., Kharchev, V., Kumari, S. 403 Patmanidis, S., Puillandre, N., Scherz, M. D., Kostadinov, I., Renner, S. S. (submitted). 404
- iTaxoTools 0.1: Kickstarting a specimen-based software toolkit for taxonomists. Submitted 405 manuscript will become publicly available on BioRxiv on 26 March 2021. 406
- Vieites, D. R., Wollenberg, K. C., Andreone, F., Köhler, J., Glaw, F., & Vences M. (2009). 407
- 408 Vast underestimation of Madagascar's biodiversity evidenced by an integrative amphibian 409 inventory. Proceedings of the National Academy of Sciences U. S. A., 106, 8267–8272.
- 410 doi:10.1073/pnas.0810821106
- Weisrock, D. W., Rasoloarison R. M., Fiorentino, I., Ralison, J. M., Goodman, S. M., 411
- 412 Kappeler, P. M., & Yoder, A.D. (2010). Delimiting species without nuclear monophyly in Madagascar's mouse lemurs. PLoS ONE, 5, e9883. doi:10.1371/journal.pone.0009883 413
- Wiens, J. J., & Penkrot, T. A. (2002). Delimiting species using DNA and morphological 414 variation and discordant species limits in spiny lizards (Sceloporus). Systematic Biology, 415
- 51, 69-91. doi:10.1080/106351502753475880 416
- 417
- Yang, Z., & Rannala, B. (2010). Bayesian species delimitation using multilocus sequence data. Proceedings of the National Academy of Sciences U. S. A., 107, 9264–9269. 418 doi:10.1073/pnas.0913022107 419
- 420 Yang, Z., & Rannala, B. (2014). Unguided species delimitation using DNA sequence data 421 from multiple loci. *Molecular Biology and Evolution*, 31, 3125–3135. 422 doi:10.1093/molbev/msu279
- Zhang, J., Kapli, P., Pavlidis, P., & Stamatakis, A. (2013). A general species delimitation 423 424 method with applications to phylogenetic placements. *Bioinformatics*, 29, 2869–2876. doi:10.1093/bioinformatics/btt499 425
- Zhang, C., Zhang, D. X., Zhu, T., Yang, Z. (2011). Evaluation of Bayesian coalescent method 426 of species delimitation. Systematic Biology, 60(6), 747–761. doi:10.1093/sysbio/syr071 427 428

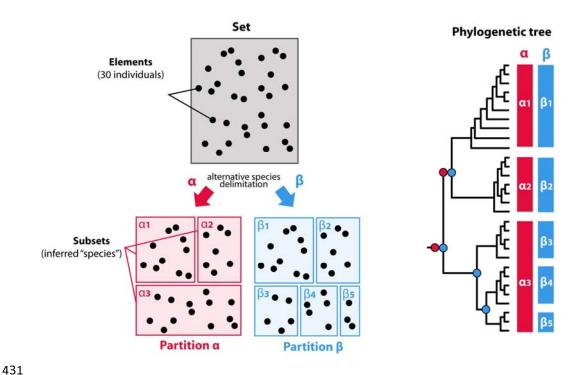


Figure 1. In mathematics, a partition of a set is a grouping of its elements into non-empty subsets, in such a way that every element is included in exactly one such subset. The main output of a species delimitation inference therefore corresponds to a partition, independently of the theoretical context, the biological input data, or the algorithms/models used. In our example, a set of 30 specimens is split by two different methods into two alternative partitions α and β , corresponding to 3 and 5 putative species (subsets), respectively. For the sake of clarity, these two alternative species partitions are represented as boxes reported next to each "species clade" in a phylogenetic tree, with hypothetical speciation events highlighted by circles via a corresponding color. Note that not all SD methods rely on a tree topology, and may therefore delimit non-monophyletic units (e.g., methods based on morphological or molecular divergence).

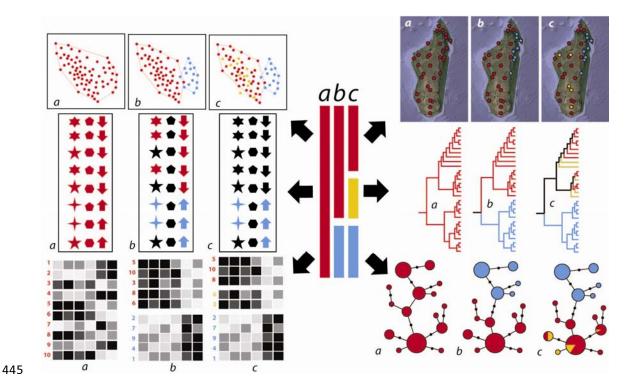


Figure 2. Illustration of exemplary potential applications of a species partition (SPART) file. If it can be parsed by other programs, SPART might facilitate the exploration of taxonomic datasets under various delimitation assumptions (such as morphometric Principal Component Analysis, automated extraction of diagnostic traits, heatmap of meristic morphological traits, distribution map, mitochondrial DNA-based phylogenetic tree, or haplotype network from nuclear DNA). In the present example, the partition b represents the optimal delimitation from a taxonomic perspective.

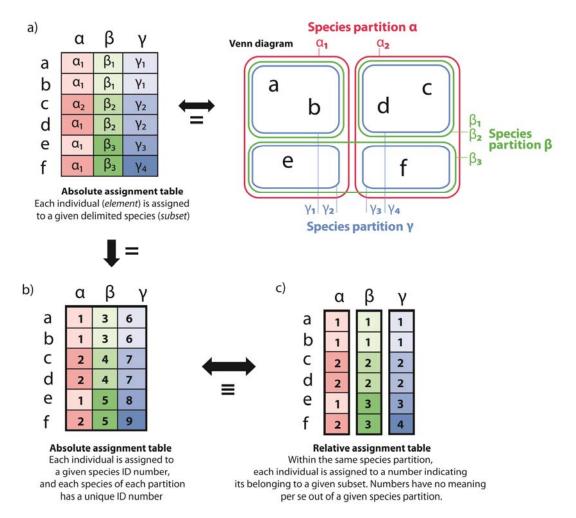


Figure 3. The SPART format can combine alternative species partitions of a same set of individuals (elements) into a unique multiple species partition file. (a) Example of set comprising six individuals split by three distinct SD analyses, resulting in three distinct species partitions (α , β and γ). All these species partitions are hierarchically compatible (i.e. they conform to the mathematical definition of nested sets), with the exception of the pair α - β (Venn diagram representing the alternative species partitions on the right, and corresponding assignment table on the left). These alternative species partitions can be coded in SPART either (b) by using a unique numbering for all the three species partitions (so that each species partition has its own set of species (subset) numbers) or (c) by using one numbering system per species partition. The latter representation allows combining different species partitions into a multiple species partition file without having to adjust each species or cluster number (subset). Both (b) and (c) are fully equivalent in SPART format, because the coding of each partition is independent from the others (subset assignment numbers have no meaning *per se*, they only indicate, within each partition, the common assignment to a specific subset).

Tools	General principle	Hypothetical partition needed as an input (a priori species assignement)	Optimal datasets and format	SPART impletementation	References
GMYC (mGMYC and bGMYC)	General mixed Yule- coalescent model	No	mtDNA – ultrametric gene tree	Yes *	Pons et al. (2006), Fontaneto et al. (2007), Monaghan et al. (2009)
BPP, iBPP	Multispecies coalescent model	Both options are possible	nDNA – multilocus alignments + (optionally in iBPP) matrix of morphological characters	In preparation	Yang & Rannala (2010, 2014), Solís- Lemus et al. (2015)
SPEDESTEM	Maximum likelihood and information theory	Yes	nDNA – ultrametric gene trees from multiple loci (nwk)	No	Ence and Carstens (2011)
ABGD	DNA barcode gap detection	No	mtDNA – sequence alignment or distance matrix	Yes	Puillandre et al. (2012)
SPECIES DELIMITATION	Coalescence / tree based approach	Yes	Topology (ultrametric tree)	No	Masters et al. (2011)
BINs	DNA barcode distance threshold + Markov clustering.	No	mtDNA – sequence alignment	No	Ratnasingham & Hebert (2013)
PTP (mPTP and bPTP)	Multi-rate Poisson ree processes model	No	Non ultrametric tree (nwk or NEXUS tree)	Yes (mPTP and bPTP) *	Zhang et al. (2013), Kapli et al. (2016)
DISSECT	Multispecies coalescent model	No	nDNA – multilocus alignments	No	Jones et al. (2014)
TR2	Multispecies coalescent model	No	nDNA – rooted gene trees from multiple loci (nwk)	Yes	Fujisawa et al. (2016)
STACEY	Multispecies coalescent model	No	nDNA – multilocus alignments	No	Jones (2017)
SODA	Quartet frequencies, based on coalescent model	No	Multiple gene tree topologies	Yes *	Rabiee & Mirarab (2019)
HaplowebMaker / CoMa	Mutual allelic exclusivity	No	nDNA – multilocus alignments	No	Spöri & Flot (2020)
ASAP	Distance-based partitions + coalescent-based scoring	No	mtDNA – sequence alignment or distance matrix	Yes	Puillandre et al. (2021)
DELINEATE	Multispecies coalescent model	Yes	Rooted ultrametric tree (nwk or NEXUS)	Yes *	Sukumaran et al. (2020)

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