

Predictors of zoonotic potential in helminths

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1 **Abstract**

2 Helminths are parasites that cause disease at considerable cost to public health and present a
3 risk for emergence as novel human infections. Although recent research has elucidated
4 characteristics conferring a propensity to emergence in other parasite groups (e.g. viruses), the
5 understanding of factors associated with zoonotic potential in helminths remains poor. We
6 applied an investigator-directed learning algorithm to a global dataset of mammal helminth traits
7 to identify factors contributing to spillover of helminths from wild animal hosts into humans. We
8 characterized parasite traits that distinguish between zoonotic and non-zoonotic species with
9 greater than 88% accuracy. Results suggest that helminth traits relating to transmission (e.g.
10 definitive and intermediate hosts) and geography (e.g. distribution) are more important to
11 predicting zoonotic species than morphological or epidemiological traits. Whether or not a
12 helminth causes infection in companion animals (cats and dogs) is the most important predictor
13 of propensity to cause human infection. Finally, we identified helminth species with high
14 modeled propensity to cause zoonosis (over 70%) that have not previously been deemed to be
15 of risk. This work highlights the importance of prioritizing studies on the transmission of
16 helminths that infect pets and points to the risks incurred by close associations with these
17 animals.

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19

20 **Keywords:** macroparasite, parasite trait, zoonosis, pet, intermediate host

21 **Introduction**

22 Understanding the factors that contribute to the emergence of novel infectious diseases
23 is a central concern to global public health [1]. Since most outbreaks of novel pathogens among
24 humans are due to spillover from animal hosts [2-4], identifying factors associated with the
25 propensity for transmission to humans is of high priority. Research in this area is particularly
26 urgent because the rate of human-wildlife contacts is increasing with changes to natural
27 landscapes and global climate [5], providing ample opportunities for human exposure to novel
28 hosts and pathogens [6, 7]. Identifying species that are potentially parasitic or pathogenic in
29 humans (i.e., those with high *zoonotic potential*) would enhance our understanding of the factors
30 underpinning spillover transmission from animal reservoirs, and enable preemptive approaches
31 to disease control.

32 One approach to evaluating zoonotic potential is to analyze pathogen and host traits
33 [e.g. 8]. Particularly, features distinguishing zoonotic from non-zoonotic parasites and their
34 reservoir host species can be used to predict which species are most likely to present high risk
35 of zoonotic exposure to people [9]. For example, work by Han, Schmidt [10] identified ‘fast’ life
36 history strategy (short-lived, short generation time) as a key predictor of the rodent species most
37 likely to be reservoirs of novel zoonotic pathogens. Trait analysis of zoonotic viruses revealed
38 that viruses which can replicate in cytoplasm are more likely to infect humans [11] and viruses
39 which infect nonhuman primates predict the transmissibility of a virus between humans [12].
40 Patterns in genome sequences of viruses have also yielded predictions on which hosts are
41 likely to be reservoirs of zoonosis and which arthropods are likely to be their vectors [13]. These
42 findings are of scientific interest concerning the current theoretical debate about why some
43 parasite species are more prone to spillover [14-16].

44 Parasitic helminths are a group of parasites that remains poorly studied in comparison to
45 viruses and bacteria, but may pose considerable future risk of human transmissibility. Helminths

46 are macroparasites, primarily known for chronic infections of the gastrointestinal tract, typically
47 caused by tapeworms (cestodes), roundworms (nematodes), or flatworms (trematodes),
48 although helminths can infect nearly all human tissues [17]. Helminths are also known to be
49 vectors for other zoonoses, such as the fever-causing bacteria *Neorickettsia sennestu*
50 transmitted by a trematode ingested via raw fish consumption [18]; although helminth vectoring
51 remains understudied [19]. Human-helminth associations have ancient origins [reviewed in 20],
52 but the relatively recent domestication of animals for food and companionship significantly
53 increased the number of parasites shared between humans and (domesticated) animals [21].
54 The agricultural revolution and associated practices, such as storage of crops in granaries, likely
55 created new links between humans and wildlife, providing additional opportunities for helminth
56 species to infect human hosts [22]. To this day, zoonotic helminths continue to emerge within
57 human populations, a process that may be further accelerated with the global trade of livestock,
58 climate change and growth in the demand for animal protein for human consumption [23].

59 Helminths are distinct from other human parasites, such as viruses and bacteria, in that
60 they commonly have complex life cycles that rely on one or more intermediate hosts [24, 25].
61 These intermediate hosts are necessary for the development of juvenile life stages (eggs and
62 larvae) and transmission to the definitive host, where the animal matures, reproduces and
63 produces propagules [26]. Intermediate hosts include a wide range of aquatic, terrestrial, wild
64 and domesticated animals [26], yet it is unknown how intermediate host identities are linked to
65 risk of helminthiasis in humans. In addition, transmission may occur directly (i.e., trophically,
66 vertically) and/or indirectly (i.e., via environment or arthropod vector). From a public health
67 perspective, most chronic infections are caused by soil-transmitted helminths [27], however, the
68 transmission modes of most zoonotic helminths have not previously been summarized. Thus,
69 identifying helminth biological and ecological traits that are linked to zoonosis can help to

70 improve our understanding of the factors that drive zoonotic potential in helminths and to better
71 manage risk of transmission to humans.

72 In addition to intrinsic biological and ecological traits such as identity of definitive and
73 intermediate hosts, transmission to humans also may be influenced by socio-economic factors
74 specific to regions where the parasites are found. Currently, most helminth infections in humans
75 are found in low and middle income countries of the tropics [27, 28], where disease prevention
76 and healthcare infrastructure vary greatly. Numerous parasitic worms such as hookworms
77 (genera *Ancylostoma* and *Necator*) are considered neglected tropical diseases which could be
78 eliminated with sufficient drug administration and effective interventions [28]. Further, given the
79 generally high animal biodiversity of tropical regions, it also may be that there are more host
80 species of potential zoonoses in this part of the world [29], although previous work indicates that
81 temperate regions contain more zoonotic helminths than tropical regions [9]. Thus, we
82 conjectured that geographic traits of helminths might be important factors for predicting the
83 probability that a species might infect humans. Despite the high variation in medical,
84 educational, and economic burden of human helminth infections worldwide [28], how the
85 different epidemiological and geographic factors relate to helminth zoonotic potential has been
86 unclear.

87 We investigated which traits of helminths are predictors of disease in humans. We
88 compiled a global dataset from existing databases and the published literature on more than
89 700 mammal helminth parasite species to examine the frequency of biological (transmission,
90 morphology), epidemiological, and geographical traits. We used boosted regression trees, an
91 ensemble learning technique, to navigate the high dimensionality of these data. These and
92 similar machine learning methods are rapidly developing approaches that can be applied to
93 heterogeneous covariates and are often robust to nonlinear interactions hidden in the data [30,
94 31]. Among over 70 variables, our machine learning approach identified key trait patterns

95 predicting helminth zoonosis. Specifically, whether a helminth species is zoonotic was best
96 predicted by three characteristics: (1) whether one of the hosts is a companion animal (i.e. dog,
97 cat), (2) whether an intermediate host is a fish (member of Chordata phylum), and (3) the
98 number of unique locations in which the helminth species has been detected. More generally,
99 this study adds to the growing body of literature used to inform strategies for preventing
100 helminth infection and mitigating risk of novel zoonoses.

101

102 **Methods**

103 *Data compilation*

104 We used the Global Mammal Parasite Database (GMPD) [32], which consists of over 700
105 species of helminths, representing three main phyla (Acanthocephala, Nematoda, and
106 Platyhelminthes) of parasitic helminths that infect wild mammals. Most emerging zoonotic
107 diseases originate from mammals [33] and therefore a mammal-focused analysis is well-suited
108 to identifying zoonotic risk factors. For each helminth species, we searched primary literature for
109 evidence of human infection originating from animal hosts to assign a binary response indicating
110 whether or not the helminth species is zoonotic. We acquired morphological information of
111 adults and eggs from Benesh, Lafferty [34] and Dallas, Gehman [35] databases, both of which
112 gathered information from the literature. To fill in gaps, we followed Dallas, Gehman [35] and
113 searched for missing morphological information from veterinary and parasitology references
114 (e.g. Taylor, Coop [36]), taxonomy references [26, 37], and primary literature. We extracted
115 minimum, mean, and maximum body length and width (in millimeters) of adult helminths from
116 the descriptions of each parasite species. We also extracted minimum, mean, and maximum
117 egg length and width (in millimeters). We compiled records of male and female body sizes when
118 that information was available. We recorded site of infection in the definitive host body when it
119 was provided.

120 We supplemented transmission information within the above references by extracting the
121 following: common name(s) of definitive and intermediate hosts, whether the species has a free-
122 living propagule stage (a binary variable), and if so, the stage of the free-living propagule as
123 egg, larva, or both (as can occur in species that pass through more than one intermediate host),
124 and the medium in which free-living stage(s) persist (soil, water, or both). We used the common
125 names of intermediate hosts to note the class or phyla to which the intermediate animal host
126 belongs, whether any of the host (definitive or intermediate) are domesticated animals (livestock
127 and pets), or companion pet animals (predominantly cats and dogs). For each species we noted
128 the transmission mode(s) to the definitive host as vertical (from parent to offspring),
129 environmental (propagules acquired from the soil, water, or both), vector (via biting arthropod),
130 or trophic (via consumption of intermediate host).

131 The GMPD provides geographical coordinates for each helminth species, which we
132 augmented with host-helminth occurrence data from London Natural History Museum (LNHM)
133 [38] available via R package *helminthR* [39]. Coordinates in the GMPD are from reported study
134 site coordinates, or centroids of the reported study area [32]. Helminth occurrences in LNHM
135 are georeferenced as centroids to the country or state (for the USA) level. In several instances
136 coordinates were not provided by the databases, which we then georeferenced based on the
137 location name using the *geocode* function [package *ggmap*; 40]. Some location names were
138 obscure, such as the portion of a continent (e.g. southern South America) or body of water (e.g.
139 southwest Atlantic), which we did not georeference. Next, based on the occurrence points of
140 each species, we calculated the number of unique locations and latitudinal range (minimum and
141 maximum), assigned a binary variable to indicate whether the species occurrences fall within
142 the tropical latitudes (between 23° 27' N and 23° 27' S), and quantified the number of
143 occurrences within tropical latitudes. We note that the number of unique locations reflects
144 geographic distribution and sampling effort. From occurrence data we also calculated the
145 number of countries, terrestrial ecoregions of the world (as defined by Olson, Dinerstein [41]),

146 and terrestrial zoogeographic realms (as defined by Holt, Lessard [42]) from which each
147 helminth species has been reported. Further, following Byers, Schmidt [43] we calculated range
148 size for each helminth species as the total area of the ecoregions in which the species has been
149 found. Finally, we calculated the mean gross domestic product (GDP) and human population
150 size of the countries (provided by package `rworldmap` in R [44]) in which the species has been
151 documented. Our final dataset consisted of 737 globally distributed helminth species
152 (supplemental materials Fig. S1) and 73 trait variables describing helminth species that we
153 included in our analyses. We classified the traits into one of four categories: transmission,
154 epidemiological, morphological, or geographical traits (see Table 1). For full descriptions of each
155 variable see supplementary materials.

156

157 *Predictive model*

158 We used boosted regression trees (BRT), a regression approach that permits missing data,
159 variable interactions, collinearity, and non-linear relationships between the response and
160 explanatory variables, which can be of mixed types [30, 45]. We fit a logistic-like predictive
161 model with the zoonotic status of the helminths (0: not zoonotic, 1: zoonotic) as the response
162 variable and the 73 traits as explanatory variables. Prior to analysis, we log transformed body
163 size variables, which were right skewed. We randomly selected 80% of the data as the training
164 set and reserved 20% for testing. Boosted regression trees were trained using the `gbm` package
165 in R [46] with Bernoulli distributed error. We ran permutations of the model with different
166 learning rates (1×10^{-5} to 1×10^{-2}) and tree depths (1 to 3) using the training set to identify
167 optimal learning parameters yielding the highest predictive performance (see supplementary
168 materials Fig. S2). The learning conditions that were identified as yielding highest accuracy as
169 assessed by the model AUC score (area under the receiver operating characteristic curve)
170 included setting the maximum number of trees to 50,000, a learning rate of 0.001, and an
171 interaction depth of 3. We used permutation procedures to compute relative importance scores

172 for each predictor variable using Friedman's algorithm [45]. We also build partial dependence
173 plots, showing the marginal effect of each variable on the predicted outcome of the primary
174 model [30, 45] (Fig. 1). Based on the results of the primary model, we ranked helminth species
175 by their mean predicted probability of being transmissible to humans (Fig. 2).

176 Finally, we repeated the above analysis using only the top 15 most important variables
177 predicted by the primary model trained on all 73 variables, and permuted the model 100 times.
178 To further evaluate the relative importance of trait category, we ran additional submodels, also
179 permuted 100 times, with one of the four trait categories (transmission, epidemiology,
180 morphology, geography) excluded (Fig. 3 and 4). We used R programming for all analyses [47].

181

182 **Results**

183 We examined 737 globally distributed helminth species of which 137 are known to infect
184 humans. Our boosted regression ensemble of models trained on 73 helminth traits distinguished
185 zoonotic versus nonzoonotic species in the test dataset with 88% accuracy ($AUC \pm SE = 0.88 \pm$
186 0.01) and identified several predictors of zoonotic helminths (Fig. 1). The most important
187 variable for accurately predicting zoonotic helminths was whether the helminth species is known
188 to infect a companion animal, followed by whether fish serve as intermediate hosts, and the
189 number of locations in which the helminth species has been documented. The fourth and fifth
190 most important traits predicting zoonotic status in helminths related to the size of terrestrial
191 zoogeographic regions observed for each helminth species (Fig. 1). Generally, the most
192 important traits were related to geography and transmission, while epidemiological and
193 morphological traits were least important (for the relative influence values of all 73 variables see
194 supplementary materials Table S1).

195 While not currently known to cause human infection, BRT models identified 3 mammal-
196 borne helminth species as likely to be zoonotic with >70% probability (Fig. 2) (in descending
197 order): *Paramphistomum cervi*, *Schistocephalus solidus*, and *Taenia pisiformis*.

198 Additional ensembles of BRT models restricted to the top 15 most important variables (as
199 identified by the primary models with 73 traits included, see Fig. 1) predicted the testing data
200 with higher accuracy (AUC = 0.91) compared to the primary models trained on all 73 traits (AUC
201 = 0.89). The restricted submodels trained on the 15 variables generally agreed on the ranking of
202 the importance of variables with the primary models (Fig. 3). Submodels trained on data without
203 one of the trait categories (i.e., leave-one-out) indicated that model trained on data without
204 morphological traits performed slightly worse (AUC = 0.90) compared to submodels with all trait
205 categories included (AUC = 0.91; Fig. 3), suggesting that including these features improved the
206 predictive accuracy of our models. Models trained on data with epidemiological traits left-out
207 performed best (AUC = 0.92; Fig. 3). Finally, models trained on data without geographical traits
208 or transmission traits performed worse than models with other categories left out (AUC = 0.87,
209 AUC = 0.89 respectively; Fig. 3). In submodels, companion animal host was the most important
210 variable, except for the submodel that excluded transmission traits (Fig. 4). For AUC scores and
211 the relative influence values of the variables in submodels see supplementary materials (Table
212 S2).

213

214 **Discussion**

215 Identifying pathogen traits associated with a propensity to spillover into humans is key
216 for understanding and predicting emergence of novel human diseases originating from wildlife.
217 We applied a machine learning algorithm to a large dataset of mammal helminths to identify
218 characteristics distinguishing zoonotic and non-zoonotic species, and to predict which species
219 currently classified as non-zoonotic have a high risk of 'spilling over' to humans in the future.
220 Our results indicate that helminths that infect companion animals (dogs and cats) and utilize fish
221 as intermediate hosts are more likely to cause human infection compared to other mammal-
222 borne helminths. The third strongest predictor of the ability to cause human infection was the
223 number of occurrences of helminth species, which indicates that widespread geographic

224 distribution might provide important transmission exposure to human hosts; however, we note
225 that this variable might also reflect sampling effort (see below). Overall, these results suggest
226 that the zoonotic potential of helminth species is related to the identity of both definitive and
227 intermediate hosts that come in direct and indirect contact with people, thereby providing
228 abundant opportunities for parasite transmission. Further, our findings highlight the importance
229 of transmission strategies in the ability of mammalian helminths to infect humans.

230 Particularly interesting is the predicted association between helminth zoonosis and
231 companion animals (predominantly cats and dogs in this study). Domestic cats and dogs are
232 hosts to numerous parasitic helminth species [36, 48] and represent an important link between
233 humans and wildlife for zoonosis [49]. Indeed, the role of cats and dogs in helminthiasis have
234 been well-documented for several parasites including the zoonotic tapeworm *Echinococcus*
235 *multilocularis* (see Richards et al. in this issue) and roundworm *Toxocara cati* [49]. While many
236 domesticated cats and dogs are “free-range” (i.e., not owned and cared for by humans), these
237 animals are ubiquitous and tend to live near humans for provisioned food and shelter. Further,
238 they hunt wild animals, consume animal parts (e.g. entrails) discarded by humans, and can
239 overlap with wildlife habitat and territories [50], even in urban areas where numerous wild
240 animals such as racoons, foxes, and coyotes thrive [51, 52]. The direct trophic interactions and
241 indirect contacts dog and cats have with wildlife provide numerous opportunities for
242 transmission of helminth parasites from wild to domestic animals, and eventually to humans.
243 Additionally, the human-pet-wildlife interface has been around for centuries as it surfaced
244 thousands of years ago with the domestication of cats 10,000 years ago and dogs 16,000 years
245 ago [53, 54]. Therefore, there has been ample opportunity for host-jumping and host-switching
246 events from wildlife to pets and humans, a process which is expected to accelerate with the
247 increasing size of the human population, associated companion animals, and activities that
248 impose close contact with wildlife.

249 Fish (freshwater or marine) as an intermediate host was identified as the third most
250 important trait for predicting zoonosis. This finding is not surprising as fish are well-documented
251 intermediate hosts to non-zoonotic parasitic worms that inflict humans [55]. One of the best-
252 known examples of zoonotic parasites transmitted by fish is nematode *Anisakis simplex*, which
253 have a complex life cycle with marine mammals as definitive host and high incidence among
254 human populations that eat raw fish [56]. Fish-borne helminths are transmitted via consumption
255 of raw, undercooked, or improperly preserved fish [57] and therefore fish represent an important
256 direct trophic link between humans and wildlife. While *wild* fish are a source of parasitic
257 helminths [55, 58], recent work indicates that farmed fish are also linked to zoonosis [59, 60].
258 Parasitic worm infections stemming from fish ingestion are increasing, likely due to the
259 significant increase in demand for fish meat associated with changes in dietary habits and
260 population growth [61]. Our finding elucidates fish as a key group of intermediate hosts linked to
261 helminthiasis and the importance of monitoring fish intended for human consumption for
262 parasitic worms to prevent and control zoonosis.

263 We also identified several geographical traits as important to predicting zoonotic
264 helminths. Specifically, the number of unique locations around the world, the number of
265 zoological realms in which helminths have been found, and the number of locations within the
266 tropics were relatively important predictors. Overall, these findings suggest that mammalian
267 parasitic helminths that are geographically widespread and able to persist in a range of habitat
268 types are also more likely to be zoonotic than their more ecological specialized counterparts,
269 possibly due to their ability to persist in different environmental conditions and exposure to
270 humans in varying environments.

271 It is important to note that study effort (and attendant bias) is likely interwoven through
272 several traits we included in this study. Particularly, the number of unique record locations might
273 not only capture distribution but also number of samples and therefore sampling effort. Indeed,
274 previous work shows that variation in sampling effort among parasitic species can predict the

275 number of localities in which the species are documented [62]. Companion animal (pet host)
276 trait might also reflect disproportionate study effort given the high access and relative ease of
277 sampling. Furthermore, veterinary diagnostics (e.g. fecal floats, snap tests) more frequently
278 performed on companion animals in high income countries might lead to higher discovery rate
279 of helminth species in these places. We found that submodels which included or excluded the
280 number of occurrences resulted in companion animal (pet host) remaining the most important
281 predictor of zoonotic status among the helminths, lending some assurance of the strong
282 statistical association between zoonotic status and pet host despite the influence of sampling
283 effort in helminth data.

284 Our model predicted several helminth species that are currently not known to infect
285 humans to have high probability (70% or higher) of causing zoonosis. The helminth species with
286 highest probability of causing human infection was a flatworm, *Paramphistomum cervi*, followed
287 by *Schistocephalus solidus*, and *Taenia pisiformis*. *Paramphistomum cervi* is environmentally
288 transmitted and requires a snail intermediate host that is accidentally ingested by wild mammals
289 and livestock ruminants (e.g. sheep and cattle), the definitive hosts [63]. Given that livestock
290 can share species of gastrointestinal helminths with farmers [64], *Paramphistomum cervi* may
291 be a likely candidate for spillover to humans. On the other hand, the flatworm *Schistocephalus*
292 *solidus* infects a copepods, fish, and fish-eating water birds [65], all of which have the potential
293 to provide trophic transmission to human host. *Taenia pisiformis* also appears likely to have the
294 pathway to directly infect humans since it utilizes rabbit intermediate hosts and carnivores
295 including cats and dogs as definitive hosts [66]. Indeed, consumption of wild rabbits by humans
296 is popular in some European countries [e.g. Spain; 67] and might facilitate host-switching to
297 humans for *Taenia pisiformis*. Identifying the three species of helminths and their traits serves
298 as an initial step in focusing efforts on surveillance and empirical work investigating the zoonotic
299 potential of these species.

300 In conclusion, we focused our study on parasitic helminth traits and used boosted
301 regression trees to quantify how the different transmission, geographic, morphological and
302 epidemiological factors relate to helminths' zoonotic potential. Our work suggests that helminths
303 found in cats and dogs are more likely to infect humans, and that consumption of fish by
304 humans may pose a greater risk of spillover. While our study examined over 700 mammalian
305 helminth species, many more parasitic worms are found in wildlife, and most are poorly
306 described with little known about their life cycles [68]. Key life cycle details, such as intermediate
307 host(s), are often assumed based on relation to better-studied species in the same genus.
308 Large gaps in our understanding of life cycles and transmission dynamics exist for most
309 parasitic worms, including those known to infect humans. Experimental infection work is largely
310 lacking, while detailed studies of life cycles are no longer common [68] as molecular studies
311 have eclipsed traditional experimental biology. Despite these knowledge gaps, the machine
312 learning approach we took point to key insights about zoonotic helminths. In particular, our
313 results highlight the importance of the interface between wildlife, companion animals, and
314 humans in determining risk of parasitic worm infections, which continue to cause significant
315 disease burden in developing countries [69], where semi-feral dogs and cats are generally not
316 treated for parasites and will likely continue to serve as a source of novel helminthiases.

317

318

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324

325

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336 **References**

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- 338 [1] Morens, D. M. & Fauci, A. S. 2013 Emerging infectious diseases: threats to human health
339 and global stability. *PLoS Pathogens* **9**, e1003467.
- 340 [2] Jones, K. E., Patel, N. G., Levy, M. A., Storeygard, A., Balk, D., Gittleman, J. L. & Daszak, P.
341 2008 Global trends in emerging infectious diseases. *Nature* **451**, 990.
- 342 [3] Taylor, L. H., Latham, S. M. & Mark, E. 2001 Risk factors for human disease emergence.
343 *Philosophical Transactions of the Royal Society of London B: Biological Sciences* **356**, 983-989.
- 344 [4] Johnson, C. K., Hitchens, P. L., Evans, T. S., Goldstein, T., Thomas, K., Clements, A., Joly,
345 D. O., Wolfe, N. D., Daszak, P. & Karesh, W. B. 2015 Spillover and pandemic properties of
346 zoonotic viruses with high host plasticity. *Scientific reports* **5**, 14830.
- 347 [5] Dickman, A. J. 2010 Complexities of conflict: the importance of considering social factors for
348 effectively resolving human-wildlife conflict. *Animal conservation* **13**, 458-466.
- 349 [6] Karesh, W. B., Dobson, A., Lloyd-Smith, J. O., Lubroth, J., Dixon, M. A., Bennett, M., Aldrich,
350 S., Harrington, T., Formenty, P. & Loh, E. H. 2012 Ecology of zoonoses: natural and unnatural
351 histories. *The Lancet* **380**, 1936-1945.
- 352 [7] Blum, A. J. & Hotez, P. J. 2018 Global “worming”: Climate change and its projected general
353 impact on human helminth infections. *PLoS Neglected Tropical Diseases* **12**, e0006370.
- 354 [8] Olival, K. J., Hosseini, P. R., Zambrana-Torrel, C., Ross, N., Bogich, T. L. & Daszak, P.
355 2017 Host and viral traits predict zoonotic spillover from mammals. *Nature* **546**, 646-650.
- 356 [9] Han, B. A., Kramer, A. M. & Drake, J. M. 2016 Global patterns of zoonotic disease in
357 mammals. *Trends in parasitology* **32**, 565-577.
- 358 [10] Han, B. A., Schmidt, J. P., Bowden, S. E. & Drake, J. M. 2015 Rodent reservoirs of future
359 zoonotic diseases. *Proceedings of the National Academy of Sciences* **112**, 7039-7044.
- 360 [11] Pulliam, J. R. & Dushoff, J. 2009 Ability to replicate in the cytoplasm predicts zoonotic
361 transmission of livestock viruses. *The Journal of infectious diseases* **199**, 565-568.
- 362 [12] Walker, J. W., Han, B. A., Ott, I. M. & Drake, J. M. 2018 Transmissibility of emerging viral
363 zoonoses. *PloS One* **13**, e0206926.
- 364 [13] Babayan, S. A., Orton, R. J. & Streicker, D. G. 2018 Predicting reservoir hosts and
365 arthropod vectors from evolutionary signatures in RNA virus genomes. *Science* **362**, 577-580.
- 366 [14] Plowright, R. K., Parrish, C. R., McCallum, H., Hudson, P. J., Ko, A. I., Graham, A. L. &
367 Lloyd-Smith, J. O. 2017 Pathways to zoonotic spillover. *Nature Reviews Microbiology* **15**, 502-
368 510.
- 369 [15] Becker, D. J., Washburne, A. D., Faust, C. L., Pulliam, J. R., Mordecai, E. A., Lloyd-Smith,
370 J. O. & Plowright, R. K. 2019 Dynamic and integrative approaches to understanding pathogen
371 spillover. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 20190014.
- 372 [16] Holmes, E. C. 2013 What can we predict about viral evolution and emergence? *Current*
373 *opinion in virology* **3**, 180-184.
- 374 [17] Bogitsh, B. J., Carter, C. E. & Oeltmann, T. N. 2019 *Human parasitology*. 5th ed.
375 Cambridge, MA, USA, Academic Press.
- 376 [18] Dittrich, S., Phuklia, W., Turner, G. D., Rattanavong, S., Chansamouth, V., Dumler, S. J.,
377 Ferguson, D. J., Paris, D. H. & Newton, P. N. 2015 *Neorickettsia sennetsu* as a neglected cause
378 of fever in South-East Asia. *PLoS Neglected tropical diseases* **9**, e0003908.
- 379 [19] Perkins, S. E. & Fenton, A. 2006 Helminths as vectors of pathogens in vertebrate hosts: A
380 theoretical approach. *International journal for parasitology* **36**, 887-894.
- 381 [20] Ledger, M. L. & Mitchell, P. D. 2019 Tracing zoonotic parasite infections throughout human
382 evolution. *International Journal of Osteoarchaeology*, 1-12.

- 383 [21] Morand, S., McIntyre, K. M. & Baylis, M. 2014 Domesticated animals and human infectious
384 diseases of zoonotic origins: domestication time matters. *Infection, Genetics and Evolution* **24**,
385 76-81.
- 386 [22] Reinhard, K. J., Ferreira, L. F., Bouchet, F., Sianto, L., Dutra, J., Iñiguez, A., Leles, D., Le
387 Bailly, M., Fugassa, M. & Pucu, E. 2013 Food, parasites, and epidemiological transitions: a
388 broad perspective. *International Journal of Paleopathology* **3**, 150-157.
- 389 [23] Gordon, C. A., McManus, D. P., Jones, M. K., Gray, D. J. & Gobert, G. N. 2016 The
390 increase of exotic zoonotic helminth infections: The impact of urbanization, climate change and
391 globalization. *Advances in parasitology* **91**, 311-397.
- 392 [24] Freeland, W. & Boulton, W. 1992 Coevolution of food webs: parasites, predators and plant
393 secondary compounds. *Biotropica*, 309-327.
- 394 [25] Parker, G., Ball, M. & Chubb, J. 2015 Evolution of complex life cycles in trophically
395 transmitted helminths. I. Host incorporation and trophic ascent. *Journal of evolutionary biology*
396 **28**, 267-291.
- 397 [26] Olsen, O. W. 1962 *Animal parasites: their biology and life cycles*. Minneapolis, Minnesota,
398 Burgess Publishing Company.
- 399 [27] De Silva, N. R., Brooker, S., Hotez, P. J., Montresor, A., Engels, D. & Savioli, L. 2003 Soil-
400 transmitted helminth infections: updating the global picture. *Trends in parasitology* **19**, 547-551.
- 401 [28] Hotez, P. J., Brindley, P. J., Bethony, J. M., King, C. H., Pearce, E. J. & Jacobson, J. 2008
402 Helminth infections: the great neglected tropical diseases. *The Journal of clinical investigation*
403 **118**, 1311-1321.
- 404 [29] Guernier, V., Hochberg, M. E. & Guégan, J.-F. 2004 Ecology drives the worldwide
405 distribution of human diseases. *PLoS Biology* **2**, e141.
- 406 [30] Elith, J., Leathwick, J. R. & Hastie, T. 2008 A working guide to boosted regression trees.
407 *Journal of Animal Ecology* **77**, 802-813.
- 408 [31] Breiman, L. 2001 Statistical modeling: The two cultures. *Statistical science* **16**, 199-231.
- 409 [32] Stephens, P. R., Pappalardo, P., Huang, S., Byers, J. E., Farrell, M. J., Gehman, A., Ghai,
410 R. R., Haas, S. E., Han, B. & Park, A. W. 2017 Global mammal parasite database version 2.0.
411 *Ecology* **98**, 1476-1476.
- 412 [33] Woolhouse, M. E. & Gowtage-Sequeria, S. 2005 Host range and emerging and reemerging
413 pathogens. *Emerging infectious diseases* **11**, 1842.
- 414 [34] Benesh, D. P., Lafferty, K. D. & Kuris, A. 2017 A life cycle database for parasitic
415 acanthocephalans, cestodes, and nematodes. *Ecology* **98**, 882-882.
- 416 [35] Dallas, T., Gehman, A. L. M., Aguirre, A. A., Budischak, S. A., Drake, J. M., Farrell, M. J.,
417 Ghai, R., Huang, S. & Morales-Castilla, I. 2019 Contrasting latitudinal gradients of body size in
418 helminth parasites and their hosts. *Global Ecology and Biogeography* **28**, 804-813.
- 419 [36] Taylor, M., Coop, R. & Wall, R. 2016 *Veterinary parasitology*. 4th ed. New Delhi, India,
420 Wiley Blackwell.
- 421 [37] Skrjabin, K. 1965 *Trematodes of animals and man. Essentials of trematodology. Volume*
422 *XVIII*. Jerusalem, Israel, Israel Program for Scientific Translations.
- 423 [38] Gibson, D., Bray, R. & Harris, E. 2005 Host-parasite database of the Natural History
424 Museum, London. . (
- 425 [39] Dallas, T. 2016 helminthR: an R interface to the London Natural History Museum's host-
426 parasite database. *Ecography* **39**, 391-393.
- 427 [40] Kahle, D. & Wickham, H. 2013 R package ggmap: Spatial Visualization with ggplot2. v
428 3.0.0. (
- 429 [41] Olson, D. M., Dinerstein, E., Wikramanayake, E. D., Burgess, N. D., Powell, G. V.,
430 Underwood, E. C., D'amico, J. A., Itoua, I., Strand, H. E. & Morrison, J. C. 2001 Terrestrial
431 ecoregions of the world: A new map of life on Earth: A new global map of terrestrial ecoregions
432 provides an innovative tool for conserving biodiversity. *BioScience* **51**, 933-938.

- 433 [42] Holt, B. G., Lessard, J.-P., Borregaard, M. K., Fritz, S. A., Araújo, M. B., Dimitrov, D., Fabre,
434 P.-H., Graham, C. H., Graves, G. R. & Jønsson, K. A. 2013 An update of Wallace's
435 zoogeographic regions of the world. *Science* **339**, 74-78.
- 436 [43] Byers, J. E., Schmidt, J., Pappalardo, P., Haas, S. E. & Stephens, P. R. 2019 What factors
437 explain the geographical range of mammalian parasites? *Proceedings of the Royal Society B*
438 **286**, 20190673.
- 439 [44] South, A. 2016 R package rworldmap: Mapping Global Data. v.1.3-6. (
- 440 [45] Friedman, J. H. 2001 Greedy function approximation: a gradient boosting machine. *Annals*
441 *of statistics*, 1189-1232.
- 442 [46] Greenwell, B., Boehmke, B., Cunningham, J. & Developers, G. 2019 R package gbm:
443 Generalized Boosted Regression Models. v.2.1.5. (
- 444 [47] R Core Team. 2020 R: A Language and Environment for Statistical Computing. R
445 Foundation for Statistical Computing. (4.0.3 ed. Vienna, Austria.
- 446 [48] Foreyt, W. J. 2013 *Veterinary parasitology reference manual*. 5th ed. Ames, Iowa, USA,
447 Wiley-Blackwell.
- 448 [49] Deplazes, P., van Knapen, F., Schweiger, A. & Overgaauw, P. A. 2011 Role of pet dogs
449 and cats in the transmission of helminthic zoonoses in Europe, with a focus on echinococcosis
450 and toxocarosis. *Veterinary parasitology* **182**, 41-53.
- 451 [50] Otranto, D., Cantacessi, C., Dantas-Torres, F., Brianti, E., Pfeffer, M., Genchi, C., Guberti,
452 V., Capelli, G. & Deplazes, P. 2015 The role of wild canids and felids in spreading parasites to
453 dogs and cats in Europe. Part II: Helminths and arthropods. *Veterinary parasitology* **213**, 24-37.
- 454 [51] Scott, D. M., Berg, M. J., Tolhurst, B. A., Chauvenet, A. L., Smith, G. C., Neaves, K.,
455 Lochhead, J. & Baker, P. J. 2014 Changes in the distribution of red foxes (*Vulpes vulpes*) in
456 urban areas in Great Britain: findings and limitations of a media-driven nationwide survey. *PLoS*
457 *One* **9**.
- 458 [52] Greenspan, E., Nielsen, C. K. & Cassel, K. W. 2018 Potential distribution of coyotes (*Canis*
459 *latrans*), Virginia opossums (*Didelphis virginiana*), striped skunks (*Mephitis mephitis*), and
460 raccoons (*Procyon lotor*) in the Chicago Metropolitan Area. *Urban Ecosystems* **21**, 983-997.
- 461 [53] Perri, A., Widga, C., Lawler, D., Martin, T., Loebel, T., Farnsworth, K., Kohn, L. & Buenger,
462 B. 2019 New evidence of the earliest domestic dogs in the Americas. *American Antiquity* **84**, 68-
463 87.
- 464 [54] Driscoll, C. A., Macdonald, D. W. & O'Brien, S. J. 2009 From wild animals to domestic pets,
465 an evolutionary view of domestication. *Proceedings of the National Academy of Sciences* **106**,
466 9971-9978.
- 467 [55] Chai, J.-Y., Murrell, K. D. & Lymbery, A. J. 2005 Fish-borne parasitic zoonoses: status and
468 issues. *International journal for parasitology* **35**, 1233-1254.
- 469 [56] Audicana, M. a. T., Ansotegui, I. J., de Corres, L. F. & Kennedy, M. W. 2002 Anisakis
470 simplex: dangerous—dead and alive? *Trends in parasitology* **18**, 20-25.
- 471 [57] Cross, J. H. 2018 Fish-and invertebrate-borne helminths. In *Foodborne Disease Handbook*.
472 *Volume 2: Viruses, Parasites, Pathogens, and HACCP* (eds. Y. H. Hui, S. A. Sattar, K. D.
473 Murrell, W. Nip & P. S. Stanfield), p. 249, 2nd ed. Boca Raton, Florida, USA, CRC Press.
- 474 [58] Dearnorff, T. L. 1991 Epidemiology of marine fish-borne parasitic zoonoses. *The Southeast*
475 *Asian Journal of Tropical Medicine and Public Health* **22**, 146-149.
- 476 [59] dos Santos, C. A. L. & Howgate, P. 2011 Fishborne zoonotic parasites and aquaculture: a
477 review. *Aquaculture* **318**, 253-261.
- 478 [60] Hung, N., Madsen, H. & Fried, B. 2013 Global status of fish-borne zoonotic trematodiasis in
479 humans. *Acta Parasitologica* **58**, 231-258.
- 480 [61] Broglia, A. & Kapel, C. 2011 Changing dietary habits in a changing world: emerging drivers
481 for the transmission of foodborne parasitic zoonoses. *Veterinary parasitology* **182**, 2-13.
- 482 [62] Walther, B. A., Cotgreave, P., Price, R., Gregory, R. & Clayton, D. H. 1995 Sampling effort
483 and parasite species richness. *Parasitology today* **11**, 306-310.

- 484 [63] Chaoudhary, V., Hasnani, J., Khyalia, M. K., Pandey, S., Chauhan, V. D., Pandya, S. S. &
485 Patel, P. 2015 Morphological and histological identification of *Paramphistomum cervi*
486 (Trematoda: Paramphistoma) in the rumen of infected sheep. *Veterinary world* **8**, 125.
- 487 [64] Squire, S. A., Yang, R., Robertson, I., Ayi, I., Squire, D. S. & Ryan, U. 2018 Gastrointestinal
488 helminths in farmers and their ruminant livestock from the Coastal Savannah zone of Ghana.
489 *Parasitology research* **117**, 3183-3194.
- 490 [65] Barber, I. & Scharsack, J. 2010 The three-spined stickleback-Schistocephalus solidus
491 system: an experimental model for investigating host-parasite interactions in fish. *Parasitology*
492 **137**, 411-424.
- 493 [66] Beveridge, I. & Rickard, M. D. 1975 The development of *Taenia pisiformis* in various
494 definitive host species. *International journal for parasitology* **5**, 633-639.
- 495 [67] Sevillano Morales, J., Moreno-Ortega, A., Amaro Lopez, M. A., Arenas Casas, A., Cámara-
496 Martos, F. & Moreno-Rojas, R. 2018 Game meat consumption by hunters and their relatives: a
497 probabilistic approach. *Food Additives & Contaminants: Part A* **35**, 1739-1748.
- 498 [68] Blasco-Costa, I. & Poulin, R. 2017 Parasite life-cycle studies: a plea to resurrect an old
499 parasitological tradition. *Journal of Helminthology* **91**, 647-656.
- 500 [69] Pullan, R. L., Smith, J. L., Jasrasaria, R. & Brooker, S. J. 2014 Global numbers of infection
501 and disease burden of soil transmitted helminth infections in 2010. *Parasites & vectors* **7**, 37.
502

Table 1. Top 15 most important variables used to predict helminth zoonoses status.

Colors of the rows correspond to the four trait categories: geographical traits are in pink, transmission traits are in green, morphological traits are in blue and epidemiological traits are in orange. Color scheme also applies to Fig. 3 and 4.

Variable	Description
Transmission	
Pet host	Binary variable indicating whether the host (final or intermediate) is a companion animal (predominantly dog and cat)
Fish intermediate host	Binary variable indicating whether an intermediate host is a fish
Geography	
Number of locations	Number of distinct locations (based on coordinates) a helminth species was observed in
Number of zoogeographic realms	Number of terrestrial zoogeographic realms (as defined in Holt et al 2013) a helminth species was located in
Number of tropical sites	Number of tropical sites the parasites was observed in
Number of ecoregions	Number of terrestrial ecoregions (as defined by World Wildlife Fund)
Number of countries	Number of countries the helminth parasite was observed in
Morphology	
Male length (mean)	Mean male length in millimeters
Female length (max)	Maximum female length in millimeters
Egg width (max)	Maximum egg width in micrometers
Female length (min)	Minimum female length in millimeters
Male length (min)	Minimum male length in millimeters
Female width (max)	Maximum female width in millimeters
Epidemiology	
Human population (mean)	Mean human population of the countries in which the helminth species is found
Gross domestic product (mean)	Mean gross domestic product (GDP) of the countries in which the helminth species occurs

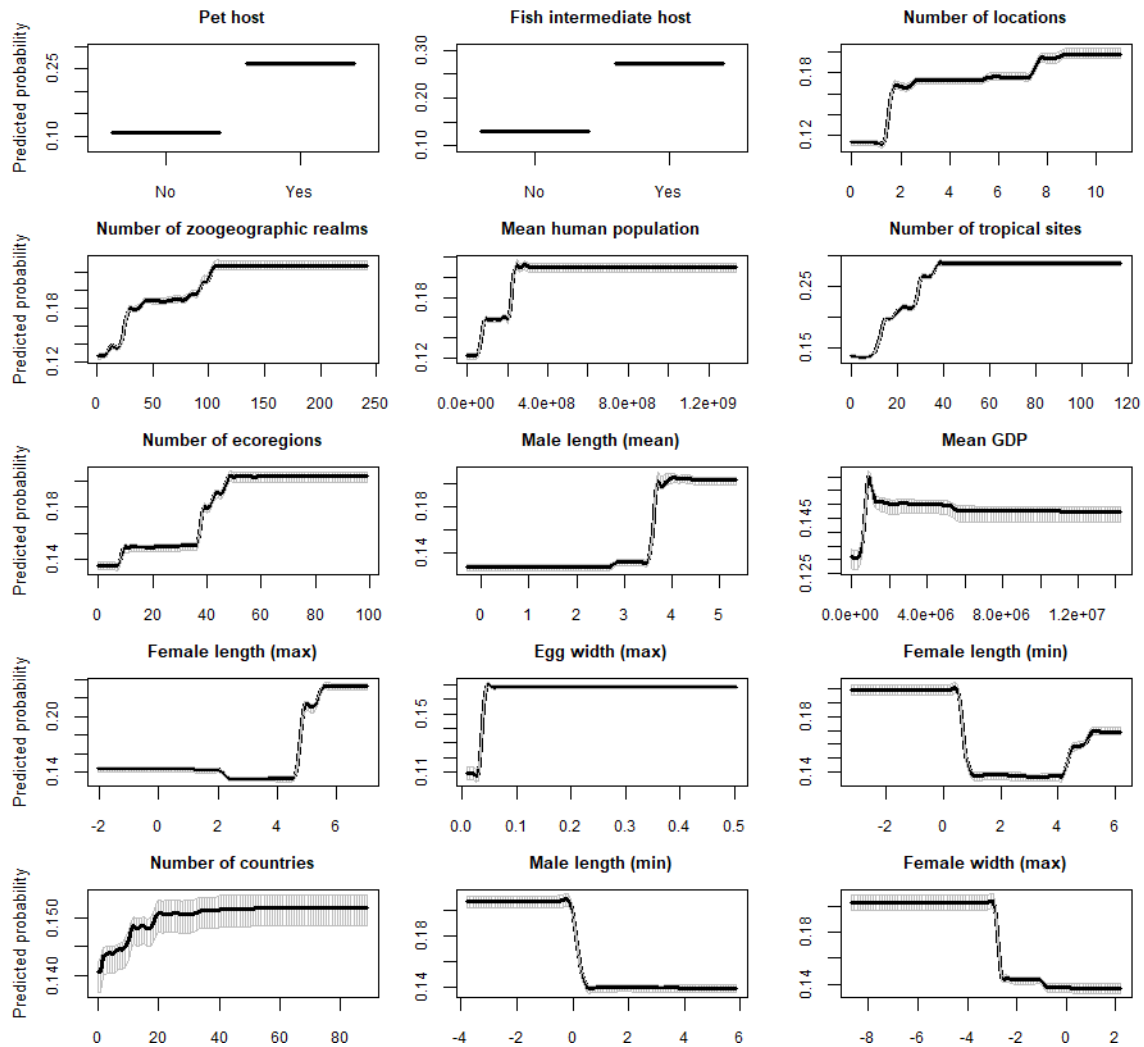


Figure 1. Partial dependence plots for top 15 most important variables. Plots are based on permutations of the primary boosted regression tree model that included 73 variables. Importance of the variables is ordered from left to right, then top to bottom. Black lines represent the median predicted probability, while shaded regions represent the corresponding 95% confidence interval across 100 permutations of the model.

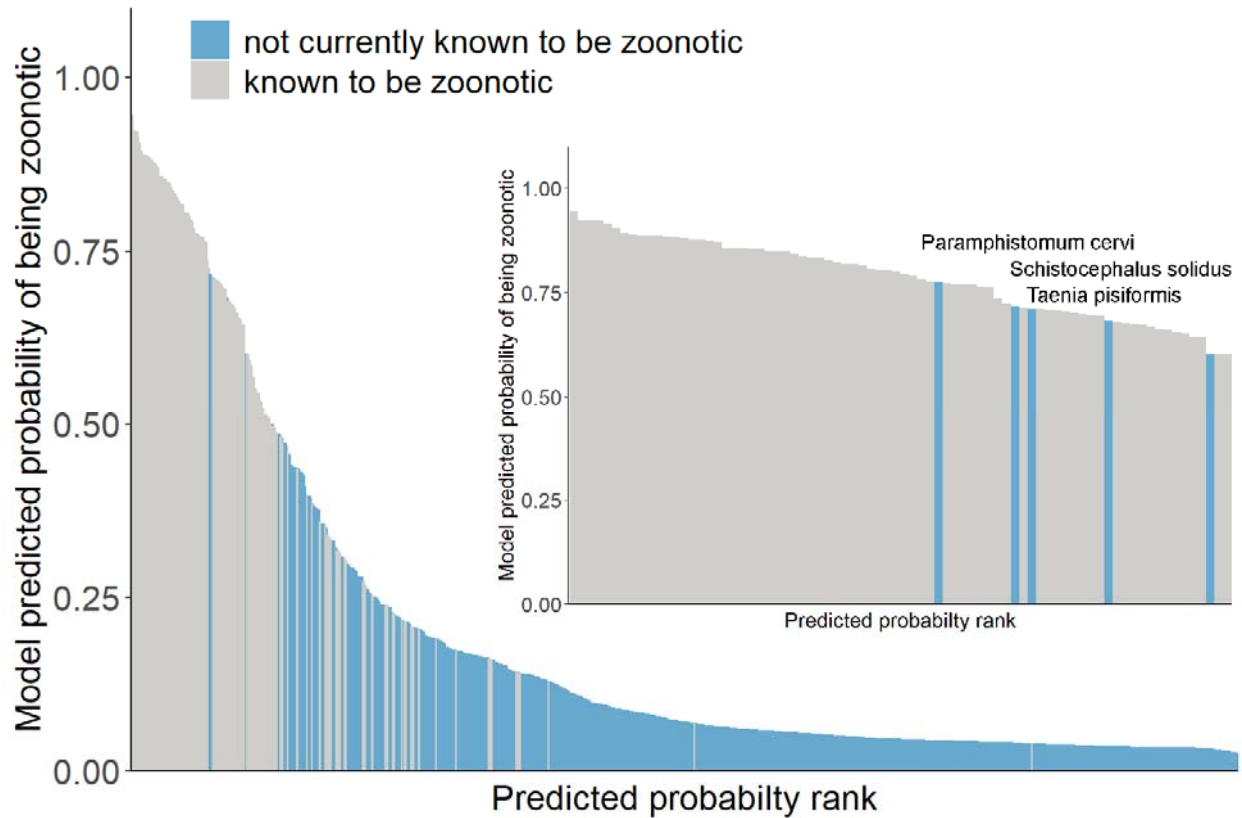


Figure 2. Predicted zoonotic helminth risk index. Average model-predicted probability of being zoonotic as ranked by the primary boosted regression tree model. Blue bars represent species not known to be transmissible to humans from wildlife and gray bars are species known to be transmissible to humans from wild hosts and are confirmed by the model to be zoonotic. Inset: zoonosis risk of helminth species with model-predicted probabilities greater than 70%. Names of top 3 species not currently known to be zoonotic appear above the bars and include *Paramphistomum cervi*, *Schistocephalus solidus*, and *Taenia pisiformis* (in descending order).

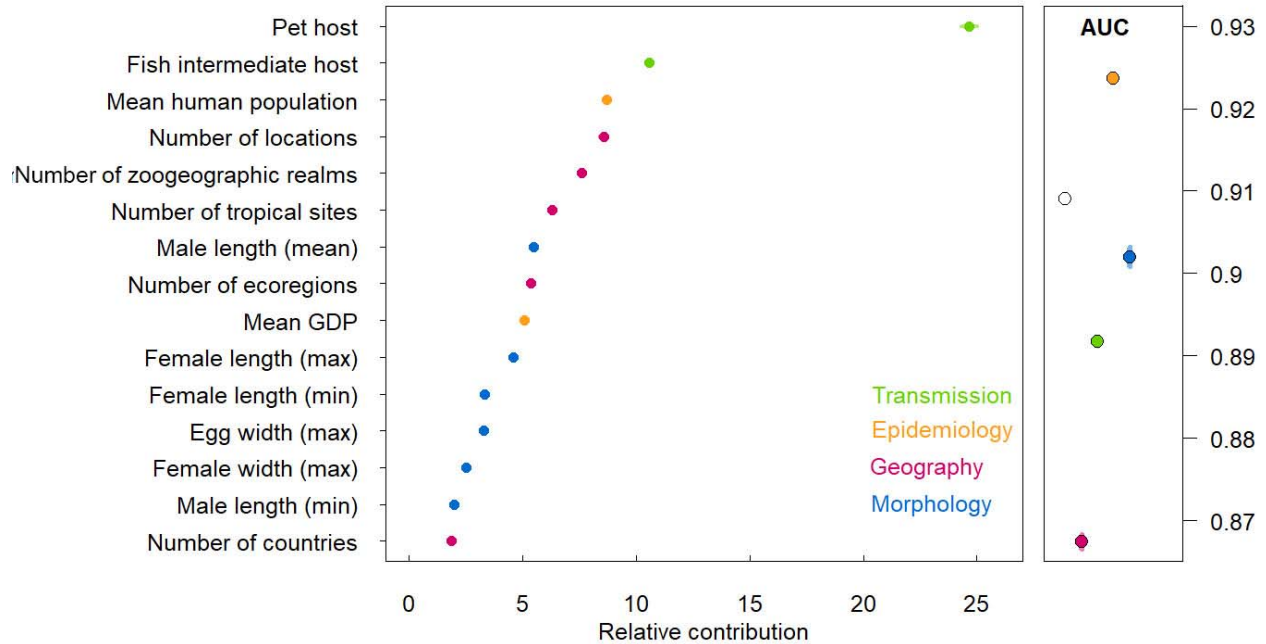


Figure 3. Variable importance values by permutation, averaged over 100 models trained on all four categories of traits (left panel), show relative importance of transmission traits (green), epidemiological traits (orange), geographical traits (maroon), and morphological traits (blue). Average model accuracy for each submodel trained on all four trait categories (white symbol), all trait categories except: morphological traits (blue), epidemiological traits (orange), transmission traits (green), or geographical traits (maroon). Error bars represent the standard deviation from 100 model permutations.

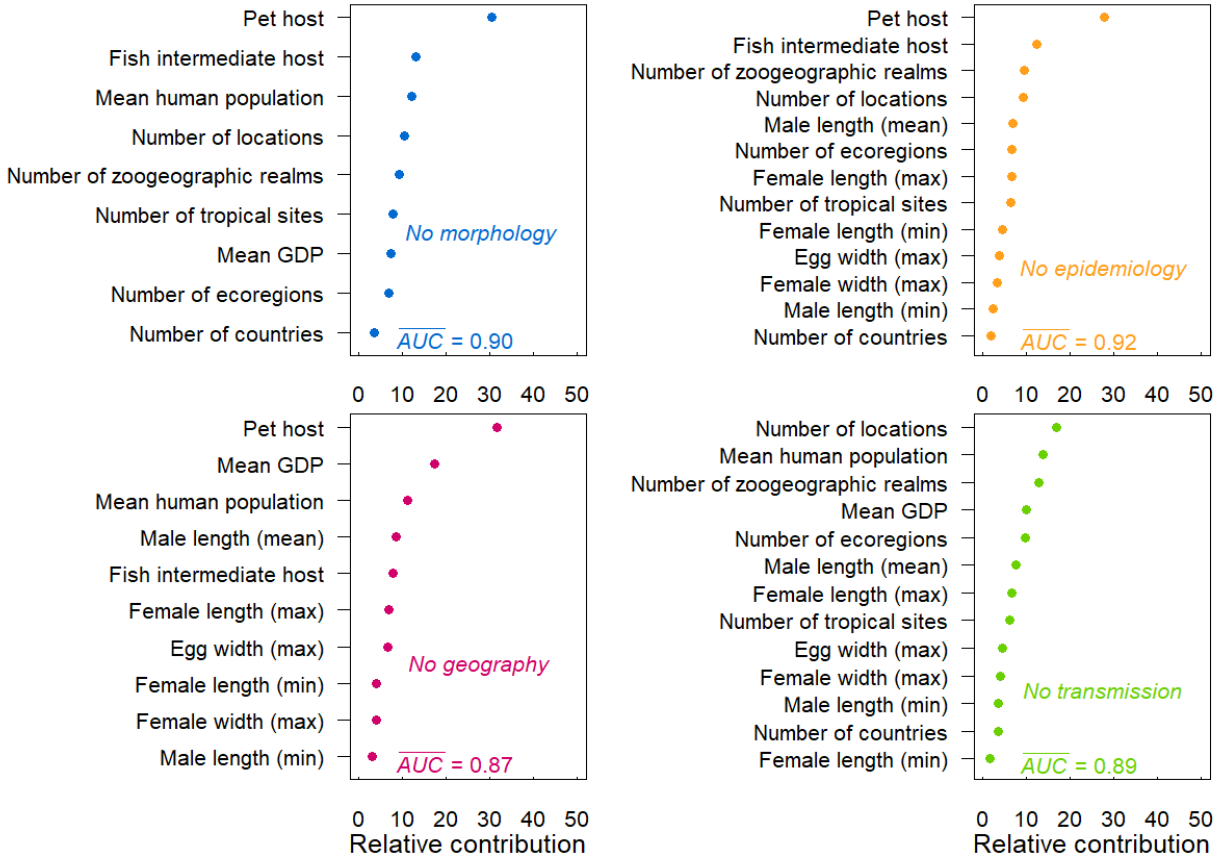


Figure 4. Variable importance values averaged over 100 model permutations trained on all categories of traits except: morphology (top left - blue), epidemiological traits (top right - orange), geography (bottom left - maroon), and transmission (bottom right- green).