Sterilizing Effects of Novel Regimens Containing TB47, Clofazimine and Linezolid in a Murine Model of Tuberculosis

Wei Yu, [a,b,c,*] Yusuf Buhari, [a,b,c,*] Shuai Wang, [a,b,c] Xirong Tian, [a,b,d] H. M. Adnan Hameed, [a,b] Zhili Lu, [a,b] Gift Chiwala, [a,b,c] Md Shah Alam, [a,b,c] Gregory M. Cook, [f,g] Dmitry A. Maslov, [h] Nanshan Zhong, [b,e] Tianyu Zhang [a,b,c,#]

[a] State Key Laboratory of Respiratory Disease, Guangzhou Institutes of Biomedicine and Health (GIBH), Chinese Academy of Sciences (CAS), Guangzhou 510530, China

[b] Guangdong-Hong Kong-Macao Joint Laboratory of Respiratory Infectious Diseases, Guangzhou Institutes of Biomedicine and Health (GIBH), Chinese Academy of Sciences (CAS), Guangzhou 510530, China

[c] University of Chinese Academy of Sciences (UCAS), Beijing 100049, China

[d] Institute of Physical Science and Information Technology, Anhui University, Hefei, 230601, China

[e] State Key Laboratory of Respiratory Disease, National Clinical Research Center for Respiratory Disease, The National Center for Respiratory Medicine, The First Affiliated Hospital of Guangzhou Medical University, Guangzhou, 510230, China

[f] Department of Microbiology and Immunology, School of Biomedical Sciences, University of Otago, Dunedin, 9054, New Zealand

[g] Maurice Wilkins Centre for Molecular Biodiscovery, The University of Auckland,

Private Bag, 92019, Auckland 1042, New Zealand

[h] Laboratory of Bacterial Genetics, Vavilov Institute of General Genetics, Russian

Academy of Sciences, Moscow, 119333, Russia

[*] These authors contributed equally to this work.

[#] Corresponding author: Tianyu Zhang

Tel: +86-20-32015270

E-mail: zhang_tianyu@gibh.ac.cn

Running title: Novel regimens for drug-resistant TB

Keywords: tuberculosis, cytochrome bc_1-aa_3 oxidase, TB47, murine model, drug

resistance

ABSTRACT

Toxicity and inconvenience associated with the use of injectable drug-containing regimens for tuberculosis (TB) have made all-oral regimens a preferred alternative. Widespread resistance to fluoroquinolones and pyrazinamide makes it essential to identify new drug candidates and study their effects on current regimens for TB. TB47 is a pyrazolo[1,5-a]pyridine-3-carboxamide with powerful synergistic *in vitro* and *in vivo* activities against mycobacteria, especially with clofazimine. Here, we investigated the bactericidal and sterilizing activities of novel oral regimens containing TB47 + clofazimine + linezolid, and the potential roles of levofloxacin and/or pyrazinamide in such drug combinations. Using a well-established mouse model, we assessed the effect of these regimens on bacterial burden in the lung during treatment and relapse (4 months after stopping treatment + immunosuppression). Our findings indicate that the TB47 + clofazimine + linezolid + pyrazinamide, with/without levofloxacin, regimens had fast-acting (4 months) sterilizing activity and no relapse was observed. When pyrazinamide was excluded from the regimen, treatment times were longer (5-6 months) to achieve sterilizing conditions. We propose that TB47 + clofazimine + linezolid can form a highly sterilizing block for use as an alternative pan-TB regimen.

INTRODUCTION

Although tuberculosis (TB) is a treatable infectious disease, in 2019, an estimated 10 million people developed active TB, with 1.2 million deaths as well as 208,000 deaths among HIV/TB co-infected individuals (1). For patients with drug-susceptible TB, standard treatment with the four powerful first-line anti-TB drugs results in acceptable cure rates. Patients who are infected with strains resistant to at least rifampicin (RIF) and isoniazid, called multidrug-resistant TB (MDR-TB), possess limited treatment alternatives and are practically incurable by the standard first-line treatment. Approximately 500,000 new cases of MDR-TB occur annually with a low treatment success rate at $\leq 57\%$ (1). It usually takes 9 - 12 months for regimens containing at least 4 - 6 drugs, including at least one injectable agent, to treat patients with MDR-TB (2). More seriously for patients, extensively drug-resistant TB (XDR-TB) is MDR-TB plus resistance to a fluoroquinolone (FQ) and an injectable agent, with an unacceptably low treatment success rate of only $\leq 39\%$ (3). Novel drugs and regimens to shorten and improve MDR/XDR-TB treatment success are urgently needed. If such regimens do not contain RIF and isoniazid, they may be applicable to both drug-susceptible and MDR-TB. Furthermore, if injectable drugs and FQs are also not included in the combination, such regimens may be useful for XDR-TB treatment. For instance, the recent FDA-approved combination, BPaL (bedaquiline + pretomanid + linezolid), is a fully oral regimen with relatively shorter course of treatment that has shown remarkably promising

clinical effect against both MDR- and XDR-TB with manageable side effects. More of

such regimens are desired to improve TB treatment.

TB47, a pyrazolo[1,5-a]pyridine-3-carboxamide, is a promising anti-mycobacterial

agent as its mechanism of action is not shared with other existing drugs (4). It can block

the cytochrome bc_1-aa_3 oxidase complex by targeting the QcrB subunit, thereby causing a

reduction in intracellular ATP and eventually inhibiting the growth of mycobacteria (5,6).

Although mono-therapy with TB47 is rather bacteriostatic than bactericidal in mouse TB

models due to rerouting of energy metabolism from QcrB to the cytochrome *bd* oxidase

(7,8), it has been proved to provide good synergistic effects in combination with either

pyrazinamide (PZA) or RIF, demonstrating a potential role in combination therapies (6).

With cytochrome *bd* inactivation, TB47 holds a significant bactericidal potential (6). We

revealed that TB47 exerted high bactericidal and sterilizing activity and corresponding

remarkable treatment-shortening effect in mice infected with *Mycobacterium ulcerans*

which lacks cytochrome *bd* activity naturally (5,7). It has also shown a very good

synergistic activity with clofazimine (CLF) against *Mycobacterium abscessus* and *M.*

tuberculosis both *in vitro* and *in vivo* (10,11), and even played a remarkable role in

shortening MDR-TB treatment duration from ≥ 9 to 5 months in a well-established

murine model of TB (11). CLF competes with menaquinone and kills bacteria by

generating lethal levels of reactive oxygen species (12), and regimens containing CLF

and cytochrome bc_1 inhibitors or other electron transport chain-targeting drugs provide

enhanced killing of both replicating and non-replicating *M. tuberculosis* (13–15)*,*

suggesting that dual or multiple targeting of the electron transport chain may be a novel

approach to enhanced killing of mycobacteria (7,10).

Considering the extensive resistance to FQs (16) and PZA (17–19) by DR-TB, the

serious side effects and inconvenience associated with injectable drugs, as well as the fact

that the patent-expiring linezolid (LZD) showed a very good activity against DR-TB in

clinical use (20), we evaluated the effect of partially or totally replacing them with LZD

in drug combinations containing TB47 + CLF as a block to study their benefits in new

regimens and to explore the possibility of developing a novel, fully oral pan-TB regimen.

RESULTS

Establishment of *M. tuberculosis* infection in female BALB/c mice. The scheme of the

experimental design is shown in Table 1. On the day after infection (Day -15), *M.*

tuberculosis H37Rv reached a cell density of 4.64 ± 0.19 mean $\log_{10}$ CFU (colony-

forming unit) in the lungs. At the time of treatment initiation (Day 0), the *M. tuberculosis*

burden increased to 8.18 ± 0.14 mean $\log_{10}$ CFU, demonstrating that a heavy infection

was established in the lungs. All nine untreated mice (Table 1) succumbed to TB

infection between day 22 and day 25 post-infection.

Bactericidal activities of the test regimens. Details of lung CFU counts obtained at

various time points are presented in Table 2. While two of five mice in the NCTZ group,

and only one of five mice in the LNCTZ group were culture negative (Table 2), all mice

treated with other regimens remained culture positive after 2.5 months of treatment. The

mean $\log_{10}$ CFU counts in the lungs declined significantly in all groups during the course

of treatment (Table 2, Figure 1). LZD was used only for 3 months due to side effects (11).

Addition of levofloxacin (LVX) to the AEZCT regimen resulted in stronger bactericidal

activity compared to the AEZCT regimen alone ($P = 0.0003 < 0.001$) (Figure 1a). In

contrast, contribution of LVX to the bactericidal activities of NCT and NCTZ was not

significant ($P > 0.05$) (Figure 1b). However, there were significant differences between

PZA-containing (NCTZ and LNCTZ) and corresponding PZA-lacking regimens (NCT

and LNCT, $P < 0.0001$ and $P < 0.001$, respectively) (Figure 1b). Compared to ALEZCT,

the stronger regimen containing amikacin (AMK), the injectable drug, NCTZ showed

better bactericidal activity ($P = 0.04 < 0.05$), NCT showed less bactericidal activity ($P =$

$0.02 < 0.05$), LNCT and LNCTZ showed no difference ($P = 0.097$, $P = 0.65$, respectively)

(Table 2).

Relapse rates after treatment cessation. *M. tuberculosis*-infected mice were subjected

to various duration of treatment using various combination regimens, after which a

number of mice were held for 3.5 months followed by two weekly immunosuppression

treatment with dexamethasone (DXM) prior to euthanasia and detection of *M.*

tuberculosis in lungs for relapse (Table 1). Treatment with 2ALEZCT/2LEZC for 4

months resulted in a 13.33 % (2/15) relapse rate, which later reduced to 0 % (0/15) with

another month of LEZC (Table 3). These results are almost the same with those from our

previous study (11). In contrast, treatment with 2AEZCT/3EZCT for 5 months and

2AEZCT/4EZCT for 6 months yielded a 33.33 % (5/15) and 0 % (0/15) relapse rates,

respectively (Table 3), which indicated that LVX played a role in shortening the treatment

duration in ALEZCT combination, though it did not show a sharp activity in lowering the

M. tuberculosis burden in lungs during the 2.5 months of treatment as mentioned above.

Similarly, 3LNCT/2LCT for 5 months achieved 0% relapse rate, a month earlier than

3NCT/3CT did, which also suggested that LVX has played a role in the LNCT

combination. Unexpectedly, after 4 and 5 months of treatment, both 3NCTZ/2CTZ- and

3LNCTZ/2LCTZ-treated mice showed 0 % (0/15) relapse rates (Table 3), which could

not provide evidence of benefit of LVX in LNCTZ combination. This was consistent with

the lack of difference in their bactericidal activities. We speculate that the actual cure

time could be even shorter than 4 months for LNCTZ and NCTZ groups. Adding PZA

into NCT significantly ($P < 0.0001$) shortened the treatment duration as 3NCTZ/2CTZ

achieved 0 % (0/15) relapse rate at least 2 months earlier than 3NCT/4CT, which was

accordant to the bactericidal activities of these two regimens. We could not provide

evidence of benefit of PZA in the combination 3LNCTZ/2LCTZ as we had no cohort of

mice relapsed in either 3LNCTZ/2LCTZ or 3LNCT/4LCT at designed time points. All

four oral regimens could achieve 0 % (0/15) relapse rate in 4 to 6 months, which were

shorter than or equal to the treatment duration of AMK-containing regimens (5 to 6

months). This indicated that LZD may be able to replace ALE or ALEZ in the control

regimen 2ALEZCT/3LEZC and still provide equal or even better outcome.

DISCUSSION

Potency of the standard short-course regimen consisting of first-line drugs (RIF +

isoniazid + PZA + ethambutol (EMB)) for drug-susceptible TB requires at least 6

months of treatment. However, for DR-TB, the duration of treatment is even longer and

treatment options are limited. Emergence of DR-TB, therefore, poses a significant threat

to individual patient management and global TB control efforts. For this reason, a

plethora of safer and accessible novel regimens that could shorten the duration of DR-TB

treatment are urgently needed (21), one of which is the recent FDA-approved BPaL.

Injectable drugs are not recommended for TB treatment due to their toxicity, poor

efficacy and causation of irreversible adverse effects, and may regularly need to be taken

at healthcare facilities. The WHO suggests the phasing out of injectable drug-containing

regimens and adoption of a shorter all-oral bedaquiline-containing regimen for treatment

of MDR/RR-TB (22). All-oral regimens, therefore, as opposed to injectable drug-

containing regimens, are a preferred alternative for TB treatment as shown by improved

clinical outcomes. It has become quite important to study the synergistic activities of

novel drugs/compounds with no cross-resistance with other anti-TB drugs as activities of

some compounds may be determined by efficacies of their companion drug(s) (23).

TB47 is an antimycobacterial candidate that has a good safety profile and ideal

pharmacokinetic parameters (5,6), as well as good synergistic effects with other TB drugs

(6,10). We used the Bangladesh regimen + TB47 (ALEZCT) from our previous study

(11) as a positive control here, in which we further confirmed the highly synergistic

activity of TB47 + CLF in shortening MDR-TB treatment duration to ≤ 5 months with

no relapse in the murine TB model. The Bangladesh regimen is not commonly used in

some places partly due to high prevalence of resistance to FQs and PZA in DR-TB

patients (17,24), especially in countries with high TB burden (25,26), for example, China,

and their roles in the regimen have not been fully elucidated in an animal model.

Therefore, considering the severity of resistance to FQs (16) and PZA (17–19) and the

serious side effects and inconvenience associated with injectable drugs, we evaluated the

effect of partially or totally replacing them in the drug combinations with the repurposed

LZD to study their benefits in the new regimens based on LZD + CLF + TB47 as a

backbone.

Rather than holding cohorts of mice for six months prior to relapse assessment as

described before (11), here, mice were held for 3.5 months followed by two weekly

immunosuppression treatment with DXM and the relapse results were comparable to our

previous study (11). We propose that four months, including immunosuppression after

treatment cessation, are long enough to monitor relapse, though the half-life of CLF is

long and may interfere with relapse evaluation.

Here, we incorporated TB47 + CLF into all test regimens to further study regimens

containing this suite of compounds. The regimen containing NCT showed very good

bactericidal and sterilizing activities (Table 2 and Table 3), which may cure TB in 6

months and could possibly form an all-oral pan-TB regimen for any form of DR-TB.

Inclusion of PZA in a combination regimen has been thought to even further shorten the

course of TB treatment provided the infecting strain remains PZA-susceptible (27). Based

on NCT, adding PZA could sharply shorten the duration further by two months or more

(Table 3). This points to the remarkable role of PZA in combination with NCT, and PZA

should be used when necessary.

As for LVX, it exhibited both bactericidal and sterilizing benefits in the regimen

2ALEZCT/3LEZCT over 2AEZCT/4EZCT (Figure 1, Table 2, Table 3). However, both

the bactericidal and sterilizing activities of NCT/3CT and 3LNCT/3LCT, which

contained NCT were indifferent ($P > 0.05$), even though they showed 6.67% and 0%

relapse rates, respectively after 5 months of treatment. Adding LVX into NCT-containing

regimens used here had only a limited or even no additive effect. Our data supports that

notion that LVX and other FQs may be removable in such regimens. However, if LVX

should be withdrawn from the Bangladesh regimen containing TB47 (ALEZCT),

administration of the regimen would be subject to an extension by one month.

Because CLF + LZD + TB47 were prepared and administered together in this

experiment, we speculate that in all LZD-containing regimens, had the administration of

LZD been separated from administration of its companion drugs by at least 4 hours (27),

their bactericidal effects might have been better than what was observed here as LZD

affects the absorption of companion agents (28).

It is noticeable that "BPaL" has shown remarkably promising effect against both

MDR- and XDR-TB in a phase III clinical trial (NCT02333799) by curing > 90% DR-TB

patients in 6 months, though most patients have had a reduction in dose or interruption of

LZD during treatment. However, side effects of the drug have been shown to be

dependent on dose and duration of treatment and were manageable by withdrawal of

LZD or interruption and resumption of treatment without sabotaging the efficacy of the

regimen (20). Those justify the withdrawal of LZD from LZD-containing regimens after

the first three months of treatment in this study. Besides the side effects, cost and

accessibility of the "BPaL" regimen is also a cause for concern by many TB patients.

In conclusion, we report potentially effective all-oral regimens based on TB47 + LZD

+ CLF, which could achieve complete sterilization after 6 months of treatment and even

4 months with addition of PZA in a mouse TB model. The benefit of adding LVX was not

quite obvious, especially in the NCT-based regimens, likely because its activity was

masked by other companion agents. Though it is too early to infer that the early

bactericidal and sterilizing potentials of the regimens in this study will necessarily

translate directly to human TB, the promising potentials of the regimens warrant further

studies to ascertain their clinical applicability in treatment of MDR- or even XDR-TB.

MATERIALS AND METHODS

Ethical approval. All animal procedures were approved by the Institutional Animal Care and Use Committee of the Guangzhou Institutes of Biomedicine and Health, Chinese Academy of Sciences (#2018053).

Mycobacterial strains and growth conditions. A mouse-passaged *M. tuberculosis* H37Rv strain (ATCC 27294) preserved at -80ºC was subcultured in Middlebrook 7H9 broth (Difco, Detroit, MI, USA) supplemented with 10% oleic acid-albumin-dextrose-catalase enrichment medium (BBL, Sparks, MD, USA), 0.2% glycerol, and 0.05% Tween 80 at 37ºC. Broth cultures of *M. tuberculosis* at the logarithmic phase with OD_{600} between 0.8 and 1.0 were used for the infection.

Antimicrobials. All drugs used in the study were purchased from Meilun bio (Dalian, China) except for TB47 and DXM. TB47 was synthesized and supplied by Shanghai Abotchem Co. Ltd (Shanghai, China) with a purity of $\geq$ 95%. DXM was purchased from Sigma-Aldrich (St. Louis, MO, USA). AMK and DXM were dissolved in sterile phosphate-buffered saline (PBS). LVX, EMB, and PZA were dissolved in distilled water. LZD, CLF, and TB47 were suspended in 0.4% sodium carboxymethyl cellulose (Solarbio, Beijing, China). All drug solutions/suspensions were prepared to deliver the drug(s) and dose(s) in a total volume of 0.2 mL per gavage or 0.1 mL per injection, based on the average mouse body weight of 20 g. Drug solutions and suspensions were

prepared weekly and stored at 4ºC prior to use.

Aerosol infection and determination of bacterial burden. Five to six weeks old female

BALB/c mice were purchased from Charles River Laboratories (Beijing, China).

Following several days of acclimatization, 362 mice were infected with *M. tuberculosis*

H37Rv at a high dose by an inhalation exposure system (Glas-Col, Terre Haute, IN,

USA). The mice were block randomized into treatment groups and timed sacrifice

cohorts before the start of treatment. Nine mice with 3 mice from each infection run were

humanely sacrificed 1 day after infection (Day -15) and on the day of treatment initiation

(Day 0) to determine the CFU counts of bacteria implanted in the lungs and at the start of

treatment, respectively. *In vivo* bactericidal activities of regimens were measured by lung

CFU counts during treatment for each time point.

Chemotherapy. Drugs were prepared as previously described (11,27). The daily doses

(mg/kg) are as follows: AMK 100, LVX 300, EMB 100, PZA 150, CLF 25, LZD 100 and

TB47 50. The scheme is shown in Table 1. Fifteen days after infection, mice were treated

once daily, 5 days per week (from Monday to Friday). AMK was administered

subcutaneously, and other drugs were administered orally by gavage. LVX + EMB or

CLF + LZD + TB47 were prepared and administered together in a single dose. PZA was

prepared and administered alone. The lung CFU count for each mouse was determined as

scheduled. Mice used for relapse assessment were subcutaneously injected with DXM

(10 mg/kg) 3.5 months post-treatment cessation for immunosuppression. DXM was

administered again a week later, after which the mice were kept for a week prior to

sacrifice.

Assessment of treatment efficacy. Efficacy was assessed on the basis of lung CFU

counts at selected time points during treatment (a measure of bactericidal activity) and

the proportion of mice with culture-positive relapse 4 months after treatment completion

(a measure of sterilizing activity). Lungs were dissected from the mice and homogenized

in 2.0 mL sterile PBS using glass tissue grinders. Undiluted as well as 10-fold serial

dilutions of the homogenates were prepared and cultured with a volume of 0.5 mL per

7H11 agar plate. Since CLF carryover can affect the growth of *M. tuberculosis*, lung

homogenates from CLF-containing groups were plated on 7H11 agar containing 0.4%

(weight/volume) activated charcoal to adsorb residual CLF (29). Plates were incubated

for 4 to 5 weeks at 37ºC before CFU counting. Relapses were assessed 4 months

(including two immunosuppressions) post-treatment cessation by equally plating the

homogenate of the whole lung on 4 plates. If ≥ 1 colony appeared, the mouse was

considered as relapse.

Statistical analysis. CFU counts (x) were log-transformed to $\log_{10} (x + 1)$ prior

statistical analyses, using 0.05 significance level and 95 % confidence interval.

Bactericidal activities of all regimens were compared by two-way analysis of variance

with Tukey's post-test to correct for multiple comparisons. Sterilizing activities of all the regimens were analyzed by Fisher's exact test, adjusting for multiple comparisons. GraphPad Prism version 8.0.2 (GrahPad, San Diego, CA) was used for all statistical analyses.

ACKNOWLEDGMENTS

This work was supported by the National Mega-project of China for Innovative Drugs (2019ZX09721001-003-003), by the National Natural Science Foundation of China (NSFC,81973372, 21920102003), by Joint Research of the Russian Science Foundation-NSFC Collaboration (21-45-00018 & 82061138019), by the Health Research Council of New Zealand-NSFC Biomedical Collaboration Fund (20/1211 & 8206112800),the Chinese Academy of Sciences Grants (154144KYSB20190005), the Grant (SKLRD-OP-201919, SKLRD-OP-202113) from the State Key Laboratory of Respiratory Disease and First Affiliated Hospital of Guangzhou Medical University. This work was sponsored by Science and Technology Innovation Leader of Guangdong Province (2016TX03R095 to TZ), the UCAS (to Y.B.) and "One Belt One Road" (to A.M.S.) Master Fellowship Programs for international students, CAS-TWAS President's PhD Fellowship Program (to G.C.) for international students, and the Postdoctoral Fellowship from the University of Chinese Academy of Sciences (to H.M.A.H). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

References

1. World Health Organization (WHO). 2020. Global Tuberculosis Report. Available at: https://apps.who.int/iris/bitstream/handle/10665/336069/9789240013131-eng.pdf

2. World Health Organization. 2019. WHO Consolidated guidelines on drug-resistant tuberculosis treatment. Available at: https://apps.who.int/iris/bitstream/handle/10665/311389/9789241550529-eng.pdf?ua=1

3. World Health Organization (WHO). 2019. Global Tuberculosis Report. Available at: https://apps.who.int/iris/bitstream/handle/10665/329368/9789241565714-eng.pdf

4. Tang J, Wang B, Wu T, Wan J, Tu Z, Njire M, Baojie W, Scott GF, Tianyu Z, Xiaoyun L, Ke D. 2015. Design, Synthesis, and Biological Evaluation of Pyrazolo[1,5-a]pyridine-3-carboxamides as Novel Antitubercular Agents. ACS Med Chem Lett. 6(7):814–8.

5. Liu Y, Gao Y, Liu J, Tan Y, Liu Z, Chhotaray C, Huofeng J, Zhili L, Gift C, Shuai W, Gaelle M, Islam MM, Hameed HMA, Xingshan C, Changwei W, Xinjie L, Shouyong T, Tianyu Z. 2019. The compound TB47 is highly bactericidal against *Mycobacterium ulcerans* in a Buruli ulcer mouse model. Nat Commun 10(1):1–9. https://dx.doi.org/10.1038/s41467-019-08464-y

6. Lu X, Williams Z, Hards K, Tang J, Cheung CY, Aung HL, Bangxing W, Zhiyong L,

Xianglong H, Anne L, Lisa W, Courtney H, Xiantao Z, Zhe W, Kyu R, Ke D,

Zhang T, Cook GM. 2019. Pyrazolo[1,5- a]pyridine Inhibitor of the Respiratory

Cytochrome bcc Complex for the Treatment of Drug-Resistant Tuberculosis. ACS

Infect Dis. 2019;5(2):239–49.

7. Lee BS, Hards K, Engelhart CA, Hasenoehrl EJ, Kalia NP, Mackenzie JS, Sviriaeva E,

Chong SMS, Manimekalai MSS, Koh VH, Chan J, Xu J, Alonso S, Miller MJ,

Steyn AJC, Gruber G, Schnappinger D, Berney M, Cook GM, Moraski GC, Pethe

K. 2021. Dual inhibition of the terminal oxidases eradicates antibiotic-tolerant

Mycobacterium tuberculosis. EMBO Mol Med 13: e13207.

8. Kalia NP, Hasenoehrl EJ, Ab Rahman NB, Koh VH, Ang ML, Sajorda DR, Hards K,

Grüber G, Alonso S, Cook GM, Berney M, Pethe K. 2017. Exploiting the synthetic

lethality between terminal respiratory oxidases to kill *Mycobacterium tuberculosis*

and clear host infection. Proc Natl Acad Sci U S A 114:7426-743.

9. Gao Y, Hameed HMA, Liu Y, Guo L, Fang C, Tian X, Liu Z, Wang S, Lu Z, Islam

MM, Zhang T. 2020. Ultra-short-course and intermittent TB47-containing oral

regimens produce stable cure against Buruli ulcer in a murine model and prevent

the emergence of resistance for *Mycobacterium ulcerans*, Acta Pharm. Sin. B.,

S2211-3835(20)30796-6. https://doi.org/10.1016/j.apsb.2020.11.007.

10. Liu Y, Tan Y, Islam MM, Cao Y, Lu X, Zeng S, Hameed HMA, Zhou P, Cai X, Wang

S, Mugweru JN, Zhang G, Yin H, Liu J, Nuermberger E, Zhang T. 2020.

Assessment of clofazimine and TB47 combination activity against *Mycobacterium*

abscessus using a bioluminescent approach. Antimicrob Agents Chemother.

2020;64(3):1–13.

11. Yu W, Chiwala G, Gao Y, Liu Z, Sapkota S, Lu Z, Guo L, Khan SA, Zhong N, Zhang

T. 2020. TB47 and clofazimine form a highly synergistic sterilizing block in a

second-line regimen for tuberculosis in mice. Biomed Pharmacother.

131(2020)110782. https://doi.org/10.1016/j.biopha.2020.110782

12. Yano T, Kassovska-Bratinova S, Shin Teh J, Winkler J, Sullivan K, Isaacs A,

Schechter NM, Rubin H. 2011. Reduction of clofazimine by mycobacterial type 2

NADH: Quinone Oxidoreductase: A pathway for the generation of bactericidal

levels of reactive oxygen species. J Biol Chem. 2011;286(12):10276–87.

13. Berube BJ, Parish T. 2018. Combinations of respiratory chain inhibitors have

enhanced bactericidal activity against *Mycobacterium tuberculosis*. Antimicrob

Agents Chemother. 2018;62(1):1–10.

14. Lamprecht DA, Finin PM, Rahman MA, Cumming BM, Russell SL, Jonnala SR,

Adamson JH, Steyn AJC. 2016. Turning the respiratory flexibility of

Mycobacterium tuberculosis against itself. Nat Commun 2016;7:1–14. Available

from: https://dx.doi.org/10.1038/ncomms12393

15. Berube BJ, Russell D, Castro L, Choi S-R, Narayanasamy P, Parish T. 2019. Novel MenA inhibitors are bactericidal against *Mycobacterium tuberculosis* and synergize with electron transport chain inhibitors. Antimicrob Agents Chemother 63:e02661-18. https://doi.org/10.1128/AAC.02661-18.

16. Ali S, Zeb MT, Masood N, Afzal MT. 2018. Prevalence of Pyrazinamide Resistance in Khyber Pakhtunkhwa, Pakistan. Microb Drug Resist. 2018;00(00):1–5. https://doi.org/10.1089/mdr.2017.0234

17. Hameed HMA, Tan Y, Islam M, Lu Z, Chhotaray C, Wang S, Zhiyong L, Cuiting F, Shouyong T, Wing WY, Nanshan Z, Jianxiong L, Tianyu Z. 2020. Detection of Novel Gene Mutations Associated with Pyrazinamide Resistance in Multidrug-Resistant *Mycobacterium tuberculosis* Clinical Isolates in Southern China. Infect Drug Resist 2020:13 217–227.

18. Shi J, Su R, Zheng D, Zhu Y, Ma X, Wang S, Li H, Sun D. 2020. Pyrazinamide Resistance and Mutation Patterns Among Multidrug-Resistant *Mycobacterium tuberculosis* from Henan Province. Infect Drug Resist 2020:13 2929–2941.

19. Kurbatova E V, Cavanaugh JS, Dalton T, Click ES, Cegielski JP. 2013. Epidemiology of Pyrazinamide-Resistant Tuberculosis in the United States , 1999 – 2009. Clin

Infect Dis 2013;57(8):1081–93. DOI: 10.1093/cid/cit452

20. Conradie F, Diacon AH, Ngubane N, Howell P, Everitt D, Crook AM, Mendel CM,

Egizi E, Moreira J, Timm J, McHugh TD, Wills GH, Bateson A, Hunt R, Van

Niekerk C, Li M, Olugbosi M, Spigelman M. 2020. Treatment of highly drug-

resistant pulmonary tuberculosis. N Engl J Med. 2020;382(10):893–902.

doi:10.1056/NEJMoa1901814.

21. O'Brien RJ, Nunn PP. 2001. The need for new drugs against tuberculosis: Obstacles,

opportunities, and next steps. Am J Respir Crit Care Med. 2001;163(5):1055–8.

22. World Health Organization. 2019. Rapid Communication : Key changes to the

treatment of drug-resistant tuberculosis. Available at:

https://www.who.int/tb/publications/2019/WHO_RapidCommunicationMDR_TB2

019.pdf?ua=1

23. Xu J, Li SY, Almeida DV., Tasneen R, Barnes-Boyle K, Converse PJ, Upton AM,

Mdluli K, Fotouhi N, Nuermberger EL. 2019. Contribution of pretomanid to novel

regimens containing bedaquiline with either linezolid or moxifloxacin and

pyrazinamide in murine models of tuberculosis. Antimicrob Agents Chemother.

2019;63(5):1–14.

24. Hameed HMA, Tan Y, Islam MM, Guo L, Chhotaray C, Wang S, Liu Z, Gao Y, Tan S,

Yew WW, Zhong N, Liu J, Zhang T. 2019. Phenotypic and genotypic characterization of levofloxacin- and moxifloxacin-resistant *Mycobacterium tuberculosis* clinical isolates in southern China. J Thorac Dis. 2019;11(11):4613–25. https://dx.doi.org/10.21037/jtd.2019.11.03.

25. Singh PK, Jain A. 2019. Limited Scope of Shorter Drug Regimen for MDR TB Caused by High Resistance to Fluoroquinolone. Emerg Infect Dis 2019;25(9):1760–2. https://doi.org/10.3201/eid2509.190105.

26. Singh N, Kumar P, Singh U, Garg R, Jain A. 2021. Fluroquinolone drug resistance among MDR-TB patients increases the risk of unfavourable interim microbiological treatment outcome : An observational study. J Glob Antimicrob Resist 24(2021)40–44. https://doi.org/10.1016/j.jgar.2020.11.011.

27. Tasneen R, Betoudji F, Tyagi S, Li S-Y, Williams K, Converse PJ, Dartois V, Yang T, Mendel CM, Mdluli KE, Nuermberger EL. 2016. Contribution of oxazolidinones to the efficacy of novel regimens containing bedaquiline and pretomanid in a mouse model of tuberculosis. Antimicrob Agents Chemother 60:270 –277. doi:10.1128/AAC.01691-15.

28. Williams KN, Brickner SJ, Stover CK, Zhu T, Ogden A, Tasneen R, Tyagi S, Grosset GH, Nuermberger EL. 2009. Addition of PNU-100480 to First-Line Drugs Shortens the Time Needed to Cure Murine Tuberculosis. Am J Respir Crit Care

419   Med 2009(180) 371–376. doi:10.1164/rccm.200904-0611OC.

420   29. Tyagi S, Ammerman NC, Lia SY, Adamson J, Converse PJ, Swanson RV, Almeida

421   DV, Grosset JH. 2015. Clofazimine shortens the duration of the first-line treatment

422   regimen for experimental chemotherapy of tuberculosis. Proc Natl Acad Sci U S A

423   112(3) 869–874. doi:10.1073/pnas.1416951112.

431   Table 1. Scheme of the experiment.

Regimen[a]	Number of mice used to determine lung CFU counts at the given time points										
	D-15	D0	M1	M1.5	M2	M2.5	M4	M5	M6	M7	Total
Untreated	9	9	9								27
2ALEZCT/3LEZCT				5	5	5	(15)[b]	(15)			45
2AEZCT/4EZCT				5	5	5	(15)	(15)	(15)		60
3NCT/4CT			5	5	5	5		(15)	(15)	(15)	65
3NCTZ/2CTZ			5	5	5	5	(15)	(15)			50
3LNCT/4LCT			5	5	5	5		(15)	(15)	(15)	65
3LNCTZ/2LCTZ			5	5	5	5	(15)	(15)			50
Total	9	9	29	30	30	30	60	90	45	30	362

[a] Numbers in the drug regimen descriptions indicate the number of months (4 weeks/month) for which the drug combination was administered. A,

amikacin; L, levofloxacin; E, ethambutol; Z, pyrazinamide; C, clofazimine; N, linezolid; T, TB47. All the drugs were administered once a day, 5 days per

week, at the following doses (mg/kg): A 100, L 300, E 100, Z 150, C 25, N 100 and T 50. A was administered subcutaneously and others were orally.

D-15, one day after infection with *M. tuberculosis* H37Rv.

D0, day of treatment initiation.

M1, one month after treatment initiation, and so on.

[b] Numbers in parentheses indicate the number of mice that were held for 4 months after treatment completion before being killed for relapse assessment.

Table 2. Lung CFU counts during the course of experiment.

Regimen[a]	D-15	D0	M1	M1.5	M2	M2.5
Untreated	4.64 ± 0.19	8.18 ± 0.14				
2ALEZCT/3LEZCT				2.10 ± 0.25	1.32 ± 0.61	0.76 ± 0.63
2AEZCT/4EZCT				2.57 ± 0.08	2.12 ± 0.16	1.34 ± 0.39
3NCT/4CT			4.53 ± 0.11	2.25 ± 0.19	1.95 ± 0.21	1.31 ± 0.54
3NCTZ/2CTZ			3.67 ± 0.34	1.42 ± 0.48	0.95 ± 0.21	0.59 ± 0.59[b]
3LNCT/4LCT			4.41 ± 0.39	2.40 ± 0.24	1.60 ± 0.46	1.27 ± 0.48
3LNCTZ/2LCTZ			3.56 ± 0.20	1.71 ± 0.40	0.91 ± 0.51	0.93 ± 0.67[c]

[a] Numbers in the drug regimen descriptions indicate the number of months (4 weeks/month) for which the drug combination was administered. A, amikacin; L, levofloxacin; E, ethambutol; Z, pyrazinamide; C, clofazimine; N, linezolid; T, TB47. All the drugs were administered once a day, 5 days per week, at the following doses (mg/kg): A 100, L 300, E 100, Z 150, C 25, N 100 and T 50. A was administered subcutaneously and others were orally.

[b] Two of five mice were culture negative.

[c] One of five mice was culture negative.

D-15, one day after infection with *M. tuberculosis* H37Rv.

D0, day of treatment initiation.

M1, one month after treatment initiation, and so on.

448 Results are presented as Log_{10} CFU per lung ± standard deviation.

450 Table 3. Proportion of mice relapsing after treatment completion.

Duration of treatment (months)	Regimen[a]	No. of culture-positive mice/total No. of mice (% of culture-positive)[b]
4	2ALEZCT/3LEZCT	2/15 (13.33%) [4, 51][c]
	2AEZCT/4EZCT	5/15 (33.33%) [76, 88, 180, 550, 410][c]
	3NCTZ/2CTZ	0/15 (0%)
	3LNCTZ/2LCTZ	0/15 (0%)
5	2ALEZCT/3LEZCT	0/15 (0%)
	2AEZCT/4EZCT	5/15 (33.33%) [291, 161, +++, +++, +++][c]
	3NCT/4CT	1/15 (6.67%) [46][c]

	3NCTZ/2CTZ	0/15 (0%)
	3LNCT/4LCT	0/15 (0%)
	3LNCTZ/2LCTZ	0/15 (0%)
6	2AEZCT/4EZCT	0/15 (0%)
	3NCT/4CT	0/15 (0%)
	3LNCT/4LCT	0/15 (0%)
7	3NCT/4CT	0/15 (0%)
	3LNCT/4LCT	0/15 (0%)

[a] Numbers in the treatment regimen descriptions represent the number of months for which the drug combination was administered. Relapse was defined

as ≥ one *M. tuberculosis* colony appeared on the plates spread with the entire lung homogenate 4 months after the indicated duration of treatment. Each

lung homogenate was equally divided and plated on 4 plates.

[b] The proportion of mice with culture-positive relapse was determined by holding cohorts of mice for an additional 4 months after treatment completion.

[c] Values in brackets indicate the number of colonies from lung homogenates of mice with relapse after treatment completion.

+++ indicates too many to count.

Figure 1. Bactericidal activities of the studied regimens as observed during the course of experiment. (a) Adding LVX to AEZCT

had a noticeable effect on its bactericidal activity. (b) Addition of LVX did not result in significant contribution to activities of any of

the NCT-based regimens. However, addition of PZA resulted in significant differences between PZA-containing and PZA-lacking

regimens. Incorporation of both LVX and PZA enhanced the activity of NCT, but the activity of NCT with PZA alone equals or even

surpasses the activity of NCT with both LVX and PZA added. ns, not significant, $P > 0.05$; ***, $P < 0.001$; ****, $P < 0.0001$. D -15,

Day -15; M1, month 1, and so on.