

1 **ULTRASTRUCTURAL STUDY OF THE ESOPHAGUS AND**
2 **STOMACH OF *Arapaima gigas* (Schinz 1822), JUVENILE**
3 **PAICHE, CREATED EXCAVATED TANK**

4

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23 **Abstract**

24 Paiche (*Arapaima gigas*) belongs to the Kingdom *Animalia*, Phylum *Chordata*,

25 Class *Actinopterygii*, Order *Osteoglossiformes*, Family *Arapaimidae*, Genus

26 *Arapaima*, and its origin may date to the Jurassic period. The species has natural

27 habitat in the Amazonian rivers, found mainly in marginal lakes, being considered

28 an important fishing resource, with high market value and high demand for meat

29 and leather in both Brazilian and international trade. This study aims to describe

30 the morphology of the esophagus and stomach by light microscopy and scanning

31 electronics microscopy. The esophagus was presented as muscular, short,

32 tubular and fan-shaped in the cranial portion, also presenting deep longitudinal
33 folds, and the entire mucosa is covered by mucus secretory cells with distinct
34 morphological characteristics. Pirarurcu's stomach has a J-shape divided into
35 three regions: cardiac with a lighter aspect, fundus portion with few folds in the
36 mucosa, and pyloric with deeper folds, also presenting gastroliths in fundus and
37 pyloric portions. Both microscopy studies highlighted three glandular regions,
38 composed by mucoid columnar epithelial cells, gastric crypts with different
39 shapes and sizes depending on each portion, in which the different shapes of
40 the mucosal folds in each region of the stomach were evident, and digitiform
41 microsaliences were found in the cardiac region, and micro-orifices and
42 desmosome in the fundus region. Also, fundus and pyloric portions produce more
43 mucus than the cardiac. Then morphology found was consistent with the eating
44 habits and management of distinct characteristics of the digestive tract.

45

46 **Keywords:** arapaimidae, digestive tract, digitiform microsaliencia, taste sensory
47 cells

48

49 **Introduction**

50 Paiche (*Arapaima gigas* Schinz, 1822) is the largest scale fish of the Amazonian
51 rivers, being found mainly in locations with low or no currents, such as marginal
52 lakes [1]. *A. gigas* is considered the most important and emblematic species of
53 Amazonian ichthyofauna, not only for its large size, but also for the historical and
54 economic context of artisanal fishing [2]. Among the fish of the Order
55 *Osteoglossiformes*, paiche is the best known and, such as others of this order, it
56 is characterized by the presence of parasphenoid bone [3].

57 However, due to the high exploitation of natural resources and lack of inspection,
58 paiche is in a worrying situation, mainly due to the reduction of natural resources
59 and near disappearance of its habitat [4]. On the other hand, it presents itself as
60 an important fishing economic activity, as it is a neotropical fish, of large size,
61 high rusticity, easy adaptation to high rates of density and high temperatures, as
62 well as good feed conversion and excellent weight gain in low water quality
63 environments [5]. However, knowledge of the fishing biology of these species is
64 necessary, especially, to understand eating peculiarities, aiming to offer rational
65 measures of exploitation [6]. The architecture and the structure of the
66 gastrointestinal tract have been described in several fish species, mainly due to
67 the interest of wide variations in both morphology and functions related to
68 nutritional dynamics, physiological changes in aquaculture target species and
69 digestive pathologies [7-9].

70 According to Rodrigues and Cargnin-Ferreira [10], the morphological aspects of
71 the digestive tract of the *A. gigas*, in the juvenile phase is similar to other
72 carnivorous teleosts, allowing the species to ingest, store and digest large foods.
73 In particular, such studies are relevant to understand morphophysiological
74 particularities related to seizure, digestion, and absorption of food in target
75 aquaculture species, and thus reduce adaptive flexibility in response to changes
76 in diet, due to the decrease in the variation in nutrient composition in natural
77 environment, administered in captivity production [11].

78 Based on the aforementioned, the propose of this investigation is to describe the
79 morphological and structural aspects of the esophagus and stomach of the young
80 *Arapaima gigas*, raised in commercial creations in excavated tanks.

81

82 **Material and Methods**

83 Thus, esophagus and stomach were collected from six specimens of Paiche
84 (*Arapaima gigas*), in the juvenile phase, raised in an intensive system of tanks
85 excavated and fed with a pelletized commercial diet, from an aquaculture
86 property in the municipality of Pimenta Bueno/Rondônia/Brazil. The fish were
87 caught for the consumption of the property, by artisanal fishing, and after removal
88 of the viscera, the esophagus and stomach were washed in running water,
89 removing the food content present inside and fixed in 10% buffered
90 formaldehyde. The sample was processed at Facilite Advanced Center for
91 Diagnostic Imaging - CADI- FMVZ-USP.

92 Organ dissection and mucosal exposure were performed on a plank to
93 macroscopically observe the different structures. Then these organs were later
94 documented with a photographic camera. For microscopic analysis, tissue
95 samples were dehydrated in a series of increasing concentrations of ethanol (70
96 to 100%) and diaphanized in xylol, with subsequent inclusion in histological liquid
97 paraffin and then microtomed at 5µm thick and subsequently stained with
98 Hematoxylin-Eosin (HE). The images were obtained with the Nikon Eclipse E-
99 800 light microscope.

100 Part of the material was processed for scanning electron microscopy (SEM).
101 Samples fixed in 10% formaldehyde were dehydrated in increasing sets of
102 alcohols at concentrations of 50%, 70%, 90% and 100%, dried in LEICA critical
103 point apparatus in CPD300 (FMVZ-USP), fixed with carbon glue in aluminum
104 metal bases (stub) and metallized (sputting) with gold in the EMITECH K550
105 metallizer (FMVZ-USP), being analyzed and photographed under a LEO 435VP
106 scanning electron microscope (FMVZ-USP).

107 The project was presented and approved by the Research Ethics Committee on
108 the use of animals of the Federal University of Rondônia, with protocol No.
109 043/2019.

110

111 **Results**

112 Anatomically, the esophagus and stomach of the *Arapaima gigas* are tubular
113 organs composed of modified muscle fibers that connect the pharyngeal cavity
114 to the proximal intestine (Fig. 1A). The esophagus is presented in form of a
115 muscular and short rectilinear tube (Fig. 1 A, C), but at the pharyngo-esophageal
116 junction (cranial portion), a marked dilation of the fan-shaped organ (a) is
117 observed, and in the caudal portion a visible reduction of the lumen (b). On the
118 other hand, the stomach is a J-shaped muscle sac (Fig. 1 A), located on the left
119 side of the cavity, extending from the esophagus to the proximal bowel portion.
120 The esophageal mucosa is characterized by numerous longitudinal folds, most
121 evident in the medial and caudal region (arrows on Fig. 1 C).

122 As the esophageal mucosa is replaced by gastric, such folds become thicker and
123 more visible. However, after stomach fixation in 10% formaldehyde it was
124 possible to differentiate it into three distinct portions, mainly by the color and
125 shape of the distribution of the gastric folds, in: cranial or cardiac portion, with a
126 lighter aspect and the central sulcus and the mucosa folds parallel to the central
127 axis of the organ (c). In the medial or fundus portion, gastric folds are more
128 evident, with the presence of transverse sulcus (d) and in the caudal or pyloric
129 portion, gastric folds are higher with deep sulcus (e). During the opening of the
130 paiche's stomach (Fig. 1 B) the presence of small gastroliths was observed in the
131 fungal and pyloric portions (d, e).

132 In the microscopic study of light, it was observed that the esophagus is wrapped
133 by a stratified epithelium (Fig. 2 A) (line) with three different cell types: the simple
134 epithelial cells, the mucous cells and the claviform cells. The number of layers
135 varied from 1 to 12 layers of cells. The mucous cells presented higher numbers
136 and they were distributed among the layers of the stratified epithelium. By
137 histochemical methods, with hematoxylin and eosin (H/E) two cell types could be
138 differentiated: acidophil and basophil (Fig. 2 A).

139 Morphologically, the acidophilic cells presented cubiform shape, enucleated,
140 plasma membrane well demarcated and acidophylic cytoplasm of granulous
141 aspect. The basophilic cells are larger, presenting round-shaped and basophilic
142 cytoplasm. The epithelial cells present varied morphology from columnar to
143 cuboidal (Fig. 2 B) and they are observed in greater number in places where the
144 epithelium is composed of up to two layers of cells. Goblet cells (Fig. 2 A) are
145 observed in smaller numbers, nucleated, presenting a light halo around the
146 nucleus, being usually located in the medial region of the epithelium. The lamina
147 propria of the mucosa and submucosa (Fig. 2 B) are composed of dense, not
148 presenting gland, well-developed connective tissue and with no separation
149 between them. Bundles of smooth muscles infiltrating the connective tissue of the
150 mucous tunica were also observed, being more evident in the cranial portion of
151 the esophagus. The muscle wrap is formed by bundles of muscles in a vertical
152 direction.

153 By scanning electron microscopy (SEM) it was observed that esophageal
154 mucosa is composed of numerous folds and sulcus in different directions (Fig. 2
155 D, E), and it is wrapped by epithelial cuboid-shaped cells, with the apical portion
156 of some cells discontinued, suggesting the release of cytoplasmic content to

157 lumen in the form of cellular explosion, and large amount of mucus on the surface
158 and inside the interdigitations of the mucosa. However, in the cranial portion of
159 the esophagus, mucosal folds are not observed, and the lining epithelium is
160 formed by cells rounded to polyhedral, with a large amount of mucus.
161 Furthermore, numerous projections in taste bud-shapes (Fig. 2 C) are observed
162 on the epithelial surface, randomly distributed by the epithelium line. Such
163 projections are intra-epithelial structures formed by grouping of support cells of
164 the pavement type and in the center, numerous sensory cells with microvilli
165 immersed in the central pore (white arrowhead). However, the surface is formed
166 by epithelial cells with digitiform micro-saliences in the caudal portion of the
167 esophagus.

168 The stomach mucosa of the *A. gigas* is covered with the simple columnar
169 epithelium with the presence of numerous crypts and gastric folds, of morphology,
170 height and in different numbers between the portions. In the cardiac portion (Fig.
171 3 A), the crypts have the end of a rounded-to-round aspect, with the presence of
172 smaller secondary crypts among them. The mucosal lamina propria is formed by
173 glandular loose connective tissue, which is observed in the base region of the
174 crypts.

175 In the fundus portion (Fig. 3 B), the epithelial cells of wrapping are columnar, with
176 slightly eosinophilic cytoplasm and the crypts are elevated and slender with the
177 pointed end, besides being more frequent compared to the cardiac portion. In the
178 mucosal lamia propria, a greater number of glands than connective tissue is
179 observed, which are larger than the cardiac portion.

180 In the pyloric portion the epithelial cells present a columnar to balloon form, with
181 areas of epithelium hyperplasia, with up to three layers of cells. Such cells are

182 also observed in glands located in the mucosal lamina propria. The gastric crypts
183 of this region are thicker and shorter.

184 There is no separation between the mucous wrap and mucosal lamina propria,
185 however the connective tissue of the mucous wrap is formed of thicker and
186 denser collagen than the mucosal lamina propria. The muscular wrap is formed
187 of bundles of smooth muscles arranged in two layers, horizontally and vertically.
188 The ultramicroscopic scanning study of the surface of the paiche's stomach
189 mucosa presented it is formed of numerous folds and interdigitations (D). These
190 are overlaid by cuboid epithelial cells (E) with micro-orifices (arrowhead) on the
191 surface of the plasma membrane of these cells.

192 At the ends of the mucosa folds, epithelial cells present dilation of the intercellular
193 spaces, which allows the visualization of desmosome (arrows). Inside the
194 interdigitations, cuboid epithelial cells are replaced by epithelial cells with the
195 plasma membrane (Fig. 3 D) with digitiform micro-salience. These cells reduce
196 in quantity in the fundus portion and they were not found in the pyloric part. Mucus
197 was observed in the three regions of the stomach; however the pyloric and
198 cardiac portion were the sites with the highest amount, also presenting mucus
199 and more flattened villi (Fig. 3 F).

200

201 **Discussion**

202 The anatomical arrangement of the digestive system of the paiche (*A. gigas*) is
203 similar to other teleosts. However, the stomach has varied shapes and regions;
204 structurally adapted to food types. Anatomically, the stomach of the *A. gigas* is
205 elongated in J-shape, similar to the species *Rhamdia quelen* [7], with a chemical
206 region, and another mechanical, highly muscularized, with crushing function of
207 the ingested food, also observed in piscivore stomachs [12].

208 The esophagus of the *A. Gigas* is a muscular organ, with several folds of mucosa
209 and a fan-shaped dilation in the cranial portion with marked reduction of lumen
210 in the medial portion, thus enabling distension during ingestion of large foods.
211 Generally, this anatomical aspect has been observed in other carnivorous fish
212 such as *Hoplias malabaricus* [13], *Salminus brasiliensis* [14], and *H. lacerdae*
213 [15], as well as in other teleosts with different eating habits [15-17]. According to
214 Rodrigues and Cargnin-Ferreira [10] and Scadeng et al. [18] the several amount
215 of folds of mucosa supports the integrity of the esophageal wall, especially during
216 the sudden distension motivated by quick ingestion.

217 By the scanning microscopy and light microscopy, a variation in the morphology
218 of epithelial cells of esophageal wrapping was observed, thus, in the cranial
219 portion, the mucosa was covered by rounded to polyhedral cells with the
220 presence of taste corpuscle and in the other portions, secretory epithelial cells.
221 The gustatory system of teleosts is activated by water-soluble substances, and
222 these are involved in detection, selection, and ingestion.

223 According to Santos et al. [19] there is a close relationship between the
224 distribution pattern of taste corpuscles and the way fish locate and select foods,
225 in carnivores usually present in the buccopharyngeal cavity and little observed in
226 the esophagus. The presence of such corpuscles in the cranial portion of the
227 paiche esophagus, allow them to eject the already ingested food and even the
228 eversion of the stomach when they feed on something strange or not palatable,
229 as well as observed in salmon [20], and other teleosts [19].

230 In addition to the taste corpuscles, three different cell types were also observed
231 in the stratified epithelium of the esophagus, mainly in the medial and caudal
232 portions, the basophilic, acidophilic and claviform mucous cells. The epithelium

233 of the teleosts is quite variable according to species, which may vary from
234 stratified squamous to stratified columnar with three cell types: secretory cells of
235 neutral, acidic and mixed mucopolysaccharides [21], such cells were also
236 observed in the paiche's esophagus of this study. According to Rodrigues and
237 Cargnin-Ferreira [10] the goblet cells produce mucus that protect the esophageal
238 wall against chemical and mechanical action during the passage of food.
239 Nevertheless, acid mucus produced by acidophilic cells, is associated with
240 binders capable of aggregating food particles, besides efficiently protecting the
241 epithelium [22].

242 Bertin [23] describes on stomach of the teleosts different anatomical
243 conformations and it can be grouped into three types: siphonal, cecum, and
244 linear. The siphonal stomach has two branches, one descending, corresponding
245 to the cardiac and the pyloric branch that ascends. The cecum stomach on the
246 other hand, has two branches that interpose in the union region, the blind and
247 rectilinear fundus and a muscular tube. However, it was possible to anatomically
248 and histologically differentiate the stomach of the *Arapaima gigas* in three distinct
249 portions: the cardiac, fundus, and pyloric, all glandular and presenting different
250 histological characteristics, mainly at the ends of the gastric crypts, as well as
251 the shape and number of glands of the mucosal lamina propria. Such
252 morphological conformations corroborate the classification suggested by
253 Cardoso et al. [24], that divide the stomach according to the anatomical
254 structures and the presence or not of gastric glands in three parts, as described
255 in the paiche.

256 The study of the digestive tract of several fish species, particularly teleosts, has
257 attracted the attention of many researchers, mainly due to the wide structural

258 variation with diversity of eating habits and behaviors, which are not found in other
259 vertebrates. Based on anatomical and histological characteristics of the digestive
260 tract, we can comprehend the feeding of fish [25]. The observation of gastrolithic
261 pebbles inside the paiches' stomach of this study may be associated with an
262 adaptation to the type of pelletized food. For Magalhães et al. [26] the presence
263 of pebbles in the stomach is related to the maceration process of pelletized foods,
264 aided by stomach peristaltic waves. However, considering the low number of
265 specimens in these studies, further research with native and captive fish are
266 necessary to verify whether the presence of these pebbles is a food adaptation.
267 On the ultramicroscopic scanning study of the surface of the paiche stomach
268 mucosa, it was observed that it is formed by folds and interdigitations, also it is
269 overlaid with different cell types such as cuboid epithelial cells, which secrete
270 mucus, and epithelial cells with digitiform micro-salience. Cells with digitiform
271 micro-salience were also observed in the caudal portion of the paiche's
272 esophagus. Morphological studies based on *Colossoma macropomum* also
273 observed the presence of such cells, overlaying the esophagus [27]. According
274 to Carrassón [28], these cells protect the epithelial surface and anchor the mucus
275 secreted by goblet cells, besides being associated with fluids and ions absorption.

276

277 **Conclusion**

278 Based on anatomical and morphological aspects observed in the esophagus of
279 this study, the anatomical fan-shaped of the cranial portion, as well as the
280 presence of taste corpuscles, enables the storage of food before passing to the
281 stomach and the neurostimulation of flavors by taste corpuscles stimulate the
282 bundles of muscle present in the submucosa wrap, carrying out the regurgitation

283 or deglutition. Then morphology found was consistent with the eating habits and
284 management of distinct characteristics of the digestive tract. However, these
285 studies were carried out in juvenile fish, raised in aquaculture systems, fed with
286 pelletized feed, which may not be observed in *A. gigas* in natural environment.

287

288 **Bioethics and biosecurity committee approval**

289 The project was presented and approved by the Research Ethics Committee on
290 the use of animals of the Federal University of Rondônia, with protocol No.
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292

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419

420 **Figure captions**

421

422 **Fig 1.** In A, external photomacrography of the *A. gigas* digestive tube, (a) esophageal cranial
423 portion (b) caudal esophageal portion straight, short and muscular tube; (c, d, e) J-shaped
424 stomach. In C mucosa of the esophagus and stomach, (a) fan-shaped esophageal cranial portion
425 (b), medial portion and caudal (arrows) with presence of longitudinal folds in the esophageal
426 mucosa, in (c) cardiac portion of the stomach, lighter aspect, in (d) mucosa in gastric folds of the
427 fundus portion and (e) pyloric portion of the stomach. In B stomach (d, e) fungal and pyloric
428 portions respectively with presence of gastroliths.

429

430 **Fig. 2.** Esophageal photomicrograph *A. gigas*. Cross-section in A, B, (HE)/ML. In A esophageal
431 mucosa lyxiform cells with halo around the nucleus (cc), cuboid cells, granulous aspect (arrows),
432 the major, rounded basophilic cells and basophilic cytoplasm (*). B epithelial cells of prismatic-
433 to cuboids-shape, epithelium is formed by up to two layers of cells (arrowhead), stratified
434 epithelium with the presence of secretory epithelial cells (trace), mucosal and submucosa lamina
435 propria with dense connective tissue, no presenting gland, without separation between them (a),
436 muscle wrap with bundles of muscles in vertical direction (b). In C, D, E, and F scanning electron
437 microscopy (SEM) of the surface of the esophageal mucosa. In C cranial esophageal region,
438 taste corpuscles to the center sensory cells with microvilli immersed in the central pore (white
439 arrow head). In D and E presents the medial esophageal region, folds (db) and interdigitations
440 (s) in different directions wrapped by cuboid-shaped epithelial cells (*), mucus release (arrow),
441 and in F caudal portion of the esophageal surface is wrapped of epithelial cells with digitiform
442 micro-saliences (md).

443

444 **Fig. 3.** Photomicrography of *A. gigas* stomach. Cross-section in A, B, C (HE) in (ML). In A, cardiac
445 portion, crypts with a rounded-to-round aspect (**), and secondary crypts (*), B in the fundus
446 portion, column epithelial cells (arrow), and slightly eosinophilic cytoplasm with elevated and
447 slender crypts and pointed extremity (***), in the mucosal lamina propria, more glands (gl) than
448 connective tissue, these being larger than the cardiac portion. In C pyloric portion, baloon-shaped

449 columnar epithelial cells (cl), epithelium hyperplasia (line), and thicker and shorter gastric crypts
450 (dotted double arrow). In D, E, F in the SEM of the glandular surface of the stomach mucosa, in
451 the cardiac portion (D) numerous folds (db) and interdigitations (s) and inside the interdigitations,
452 cuboid epithelial cells replaced by epithelial cells with the digitiform micro-salience plasma
453 membrane (*). E, cubiform epithelial cells with the presence of micro orifices (arrowheads), and
454 at the ends of the mucosa folds the epithelial cells present dilation of the intercellular spaces,
455 which allows the visualization of desmosome (arrows). In F, the pyloric portion presents flatter
456 and thicker folds with presence of mucus (**).

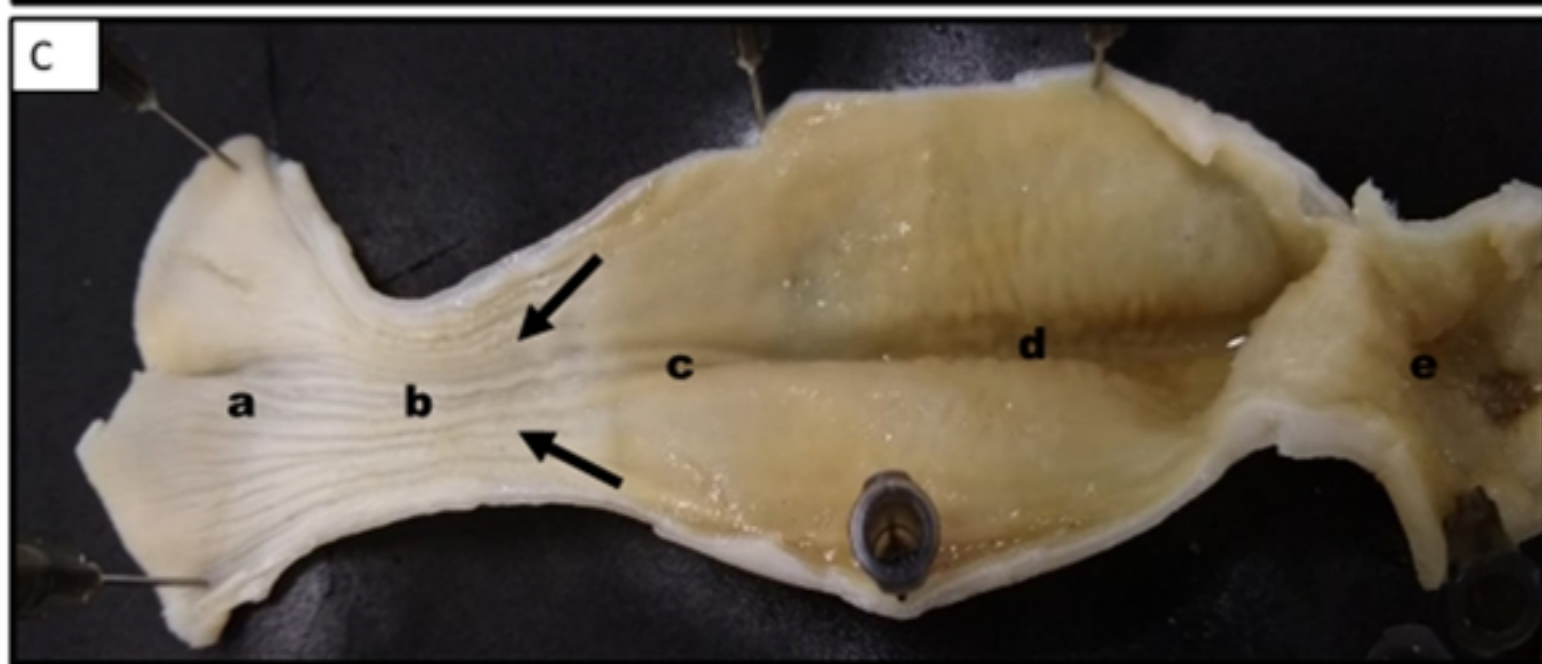


Figure 1

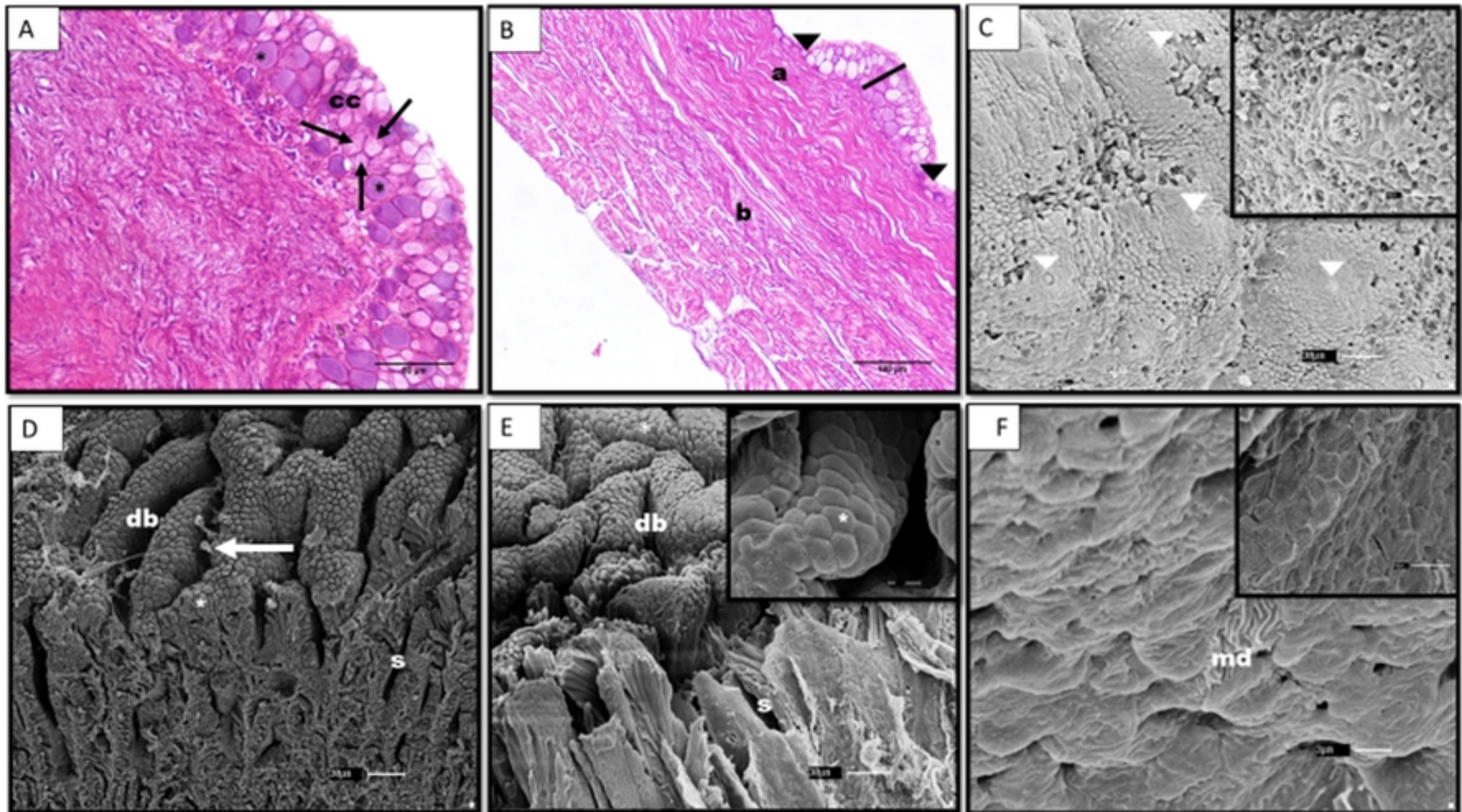


Figure 2

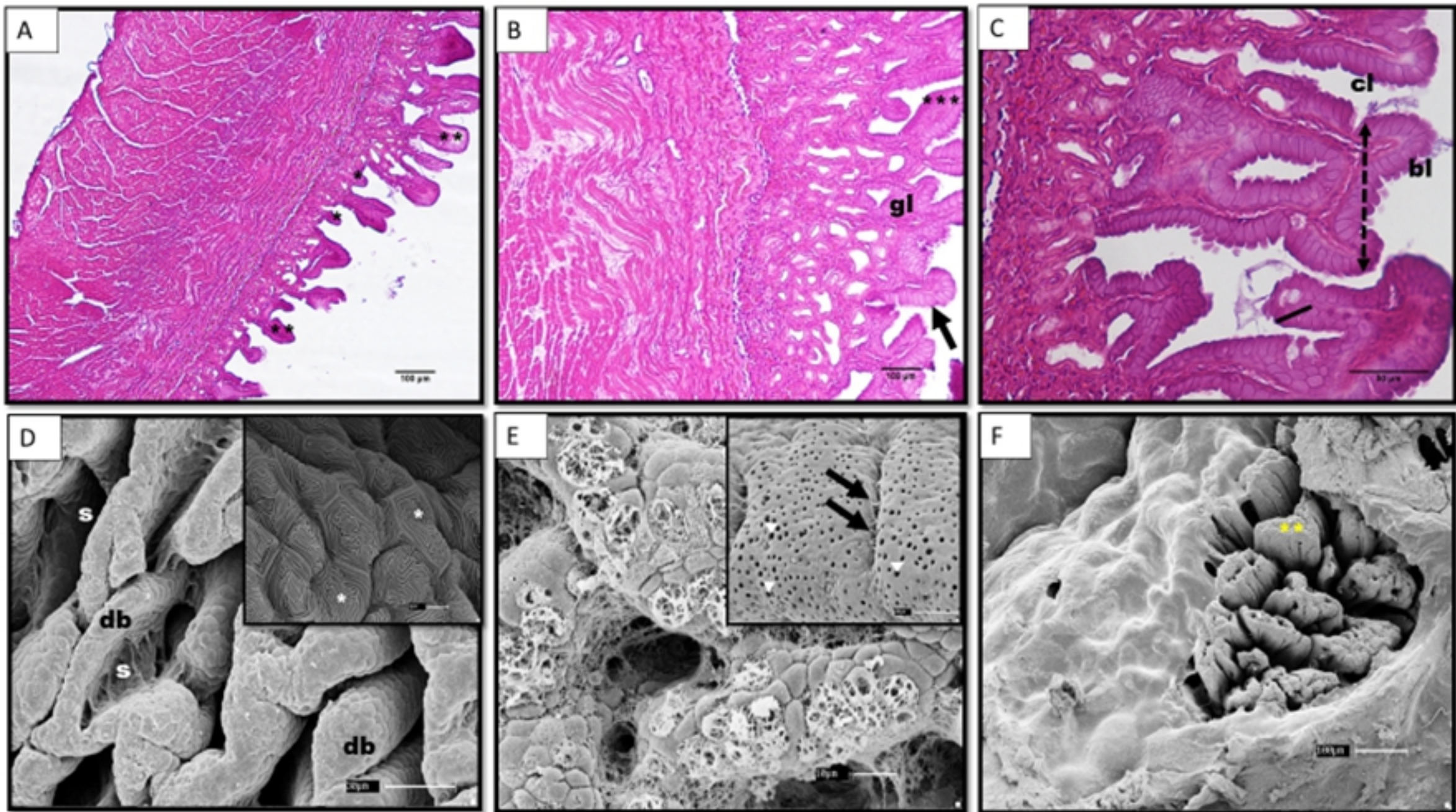


Figure 3