Shorter effective lifespan in laboratory populations of *D. melanogaster* can reduce

sexual selection

Avani Mital[1#], Manaswini Sarangi[2], Bodhisatta Nandy[3], Neha Pandey[4], Amitabh Joshi[5]

Evolutionary Biology Laboratory, Jawaharlal Nehru Centre for Advanced Scientific Research, Bengaluru

560064 India.

1: Avani Mital (# Corresponding author)

Present address: Biology Division, Department of Physics, Chemistry and Biology (IFM), Linköping University

Campus Valla, 58183, Sweden.

avani.mital@liu.se

2: Manaswini Sarangi

Present address: 4202 Biological Sciences Building, University of Michigan, MI 48109, USA

msarangi@umich.edu

3. Bodhisatta Nandy

Present address: Department of Biological Sciences, Indian Institute of Science Education and Research,

Berhampur 760010 India

nandy@iiserbpr.ac.in

4.nehapande2@jncasr.ac.in

5. ajoshi@jncasr.ac.in

Acknowledgment: We are grateful to Sajith V.S, Rajanna N., and Muniraju for help with the experiments, and two anonymous reviewers for helpful feedback. This work was funded by Jawaharlal Nehru Centre for Advanced Scientific Research (in-house funding to AJ), the Science and Engineering Research Board (SERB) JC Bose fellowship (recipient: AJ), and in part by personal funds of AJ. JNCASR provided doctoral fellowships to AM, MS, and NP, and a postdoctoral fellowship to BN.

Conflict of interest: The authors declare no conflict of interest.

Author contributions: AJ, AM and BN developed the overall theme of the study; AM and BN designed the specific experiments; AM, MS, BN and NP carried out the experiments; AM analysed the data and wrote the first draft of the paper; all authors contributed to the final draft and revision.

Data Availability statement: The raw data based on which we report results in this article is available as supplementary material (SM_2).

Abstract

The role of sexual selection in mediating levels of sexual conflict has been demonstrated in many experimental

evolution studies on *Drosophila* spp. where competition among males for mating was the target of selection.

Sexual selection has also been shown to affect the evolution of life-histories. However, the influence of

divergent life-histories on reproductive strategies and, therefore, sexual selection and possibly sexual conflict,

have been less well studied. We investigated *D. melanogaster* populations selected for a short development time

and early age at reproduction for changes in reproductive behaviour and traits that are proxies of sexual

selection. We report a large reduction in reproductive competition experienced by the males of these

populations, compared to ancestral populations that continue to not be consciously selected for rapid

development or early reproduction, likely leading to reduced sexual selection. We show that rapidly developing

and early reproducing populations have very low levels of mating in their lifetime (females are more or less

monandrous), low courtship levels, shorter copulation duration and longer time from eclosion to first mating,

compared to the controls. These results are discussed in the context of the previously demonstrated reduction of

inter-locus sexual conflict in these populations. We show that life-history strategies can have a large and

significant impact on sexual selection, with each influencing the other and contributing to the complexities of

adaptation.

Key Words: Sexual conflict, life-history evolution, male-male competition, courtship, reproductive behaviour,

experimental evolution.

Significance statement

Sexual conflict, often manifested as an arms-race between males and females trying to enhance their own

reproductive success at some cost to the other, is of great evolutionary interest because it can maintain genetic

variation in populations, prevent the independent optimization of male and female traits, and also promote

speciation. Sexual selection, or variation in mating success, is well known to affect levels of sexual conflict.

However, it is not so clear whether, and how, the regular evolution of life-histories also affects sexual selection.

Here, we show that life-history evolution in fruit fly populations selected for traits not directly related to sexual

conflict can, nevertheless, mediate the possible evolution of altered sexual conflict levels through effects on

sexual selection. Populations that evolved to develop to adulthood fast, and reproduce relatively early in life, are

shown to undergo less sexual selection, which can explain the low sexual conflict levels earlier observed in

them.

Introduction

Due in part to Darwin's independent treatment of natural selection (Darwin 1859, 1868) and sexual selection (Darwin 1871), the two sub-fields have largely developed separately, and the causes and consequences of viability/fecundity selection and sexual selection have often been studied by evolutionary biologists with somewhat differing backgrounds and interests. An unfortunate consequence of this outcome has been that studies of life-history evolution and sexual selection did not meaningfully intersect for many decades, even though they both focus on opportunities for, and timing and distribution of, reproductive output, albeit from somewhat different points of view. Parental investment per offspring differs between males and females of most sexually reproducing species (Bateman 1948; Trivers 1972), resulting in different reproductive strategies for male or female fitness maximization, often leading to differences in optimal mating rates for males and females (Bateman 1948; Andersson 1994). Such differences can give rise to inter-locus sexual conflict (Parker 1979; Chapman et al. 2003; Anrqvist and Rowe 2013), resulting in arms-race like dynamics with males evolving to manipulate female reproductive choices, and females, in turn, evolving to circumvent such manipulation (Chapman et al. 2003; Anrqvist and Rowe 2013). Sexual selection and sexual conflict are clearly intertwined and, indeed, have been studied together in some detail over the past few decades (e.g., Wigby and Chapman 2004; Linklater et al. 2007; Edwards et al. 2010; Nandy et al. 2013a, b). However, even though viability and fecundity selection can shape life-histories in ways that can either heighten or reduce sexual selection and, thereby, sexual conflict, life-history evolution has not yet been integrated with studies of sexual selection and sexual conflict in similar detail.

Several studies have provided evidence for sexual selection affecting life histories. For example, Hollis et al. (2017) report the evolution of faster development and maturation in monogamous compared to polygamous *D. melanogaster* populations. In males of the decorated cricket, *Gryllodes pirillas*, increasing reproductive effort with age was shown to correlate with slower ageing and longer lifespan (Archer et al. 2012). Similarly, Zajitschek et al. (2009) demonstrated sexual dimorphism in age dependent patterns of survival and reproduction in the field cricket *Teleogryllus commodus*, likely due to differences is age-dependent reproductive strategies between males and females. Recently, evidence for the evolution of longer development times and decreased desiccation and starvation resistance in polyandrous *D. pseudoobscura* lines compared to monogamous lines has also been reported (Garlovsky et al. 2021).

In contrast, the direct influence of life-history changes on sexual selection and sexual conflict has been

investigated relatively rarely, even though these influences can potentially have large effects on both sexual

selection and sexual conflict. For example, a life-history providing a relatively short duration of time for

reproduction can reduce sexual selection by constraining opportunities to re-mate, driving lower levels of

competition among males. The latter, in turn, could drive an evolutionary reduction in inter-locus sexual conflict

levels. Thus, effective adult lifespan (the duration of adult life in which reproduction can occur), in particular,

can alter the level of sexual selection, all else being equal. This can be particularly important in discrete

generation laboratory systems such as *Drosophila*, where continued adult lifespan beyond the point at which

eggs are collected to initiate the next generation does not contribute to Darwinian fitness. For example,

populations of *D. melanogaster* that experienced longer durations of effective adult life evolved increased male

offense and defense ability at late ages and an improved ability to induce typical female post mating responses,

compared to their controls (Service 1993; Service and Fales 1993; Service and Vossbrink 1996). Similarly,

selection for early reproduction in seed beetles *Callosobruchus maculatus* resulted in the evolution of more

frequent early-life mating compared to those selected for late reproduction (Maklakov et al. 2010).

One of the clearest demonstrations of life-history evolution affecting sexual conflict has come from a

study showing considerably reduced sexual conflict in populations of *D. melanogaster* subjected to long term

selection for rapid development and early reproduction in the laboratory relative to control populations from

which they were derived (Ghosh and Joshi 2012; Mital et al. 2021). This was shown to be largely driven by the

much smaller body size of flies from the selected populations (Mital et al. 2021), a consistent correlated

response to strong selection for rapid development in *D. melanogaster* across multiple studies (Zwaan et al.

1995; Nunney 1996; Chippindale et al. 1997a; Prasad et al. 2000). The observation that size reduction in these

selected populations likely drove the evolution of reduced levels of sexual conflict appears to be consistent with

studies showing smaller males to be less harmful to females than larger males (Partridge et al. 1987a, b; Pitnick

1991; Pitnick and Garcia-Gonzales 2002).

However, in addition to being selected for rapid development, which results in reduced body size, the

selected populations used by Ghosh and Joshi (2012) were also selected for relatively early reproduction

(around day 3 of adult life) compared to controls (around day 11 of adult life). The selected populations, thus,

have an effective adult lifespan that is less than one-third that of the controls. With a 3 day effective adult

lifespan, there is limited scope of multiple matings by females, especially if smaller, resource deprived females

fail to remate readily. This can result in a further reduction in sexual selection and sexual conflict in these

populations, beyond that due to the smaller body size of selected population males (Ghosh and Joshi 2012;

Mital et al. 2021). Here, we examine this hitherto unexplored aspect of the FEJ selection regime by asking

whether differences in the breeding ecology of these selected and control populations might also be affecting

sexual selection.

We looked at lifetime mating frequency, courtship frequency, female maturation time and copulation

duration in our selected and control populations. Mating frequency has been used as a proxy for potential sexual

selection previously (Kuijper and Morrow 2009). Courtship frequency is indicative of male mating effort since

D. melanogaster males perform extensive and energetically expensive courtship for which they likely bear a

fitness cost (Cordts and Partridge 1996; Anholt et al. 2020). Moreover, courtship by males potentially provides

an opportunity for females to exercise mate choice (Gavrilets et al. 2001; see Anholt et al. 2020 for review).

Copulation duration is another estimate of reproductive effort/investment by males as copulation in *D.*

melanogaster lasts substantially longer than required for sperm transfer alone, with the additional time spent in

the transfer of accessory gland proteins (Acps) (Gilchrist and Partridge 2000). Many of these Acps are known to

mediate sexually antagonistic effects in females (Chapman et al. 1995; Wolfner et al. 1999; Chapman 2001).

Methods

<u>Study Populations</u>

We used eight large, outbred *D. melanogaster* populations that have a common ancestry. Four of these

populations were selected for rapid pre-adult development and early reproduction and are referred to as the

FEJs (**F**aster developing, **E**arly reproducing, **J**B derived); the other four were ancestral controls, called the **JB**s

(**J**oshi **B**aseline). Details of the ancestry of these populations and their maintenance have been reported

previously (JB: Sheeba et al. 1998; FEJ: Prasad et al. 2000).

In summary, JBs are maintained on a 21day discrete generation cycle with eggs collected into 40

replicate vials (9.5 cm ht × 2.4 cm dia) per population (60-80 eggs/ 6mL of banana-jaggery medium). All adults

typically emerge by the 12th day from egg collection and are transferred to fresh food vials on days 12, 14 and

16. On day 18, all flies are collected into a Plexiglas cage ($25 \times 20 \times 15$ cm^3) provided with food supplemented

with additional yeast for three days, before starting the next generation from eggs laid during an 18 h time

window on day 21. The FEJ maintenance is similar except that only the first 20-25% of eclosing flies from each

vial are collected directly into a Plexiglas cage with food supplemented with additional yeast. After three days,

eggs are collected to initiate the next generation. All populations are maintained at a breeding adult number of

1500-1800 flies, under constant light, $25^{\circ}C \pm 1^{\circ}C$ and about 90% relative humidity. Thus, only the fastest

developing flies in the FEJs make it to the breeding pool and reproduce relatively early, i.e., on day three after

eclosion. Consequently, by the time of this study (~600 generations of FEJ selection), FEJs were being

maintained on a 10-day discrete generation cycle. Since each of the four FEJ populations has been derived from

one JB population, we could account for ancestry by treating FEJs and JBs with matched subscripts as random

blocks in the statistical analyses. Standardization (common control-type rearing conditions for a full generation

in order to equalize non-genetic parental effects) was done only prior to assaying reproductive maturity and

copulation duration in the FEJ and JB populations, as we wanted to assay courtship and mating frequencies

under conditions approximating the regular maintenance of the populations. It was not possible to record

behaviour data blind as the focal flies from the JB and FEJ populations are easily distinguishable based on their

body size.

<u>Mating and Courtship Frequencies</u>

For this assay, we used flies derived from eggs laid in the running cultures, without standardization, as

we wanted to assess breeding ecology differences that FEJ and JB flies experience during their maintenance,

including those potentially due to non-genetic parental effects. Since the maintenance regimes and development

times of the FEJ and JB populations are quite different, assaying them under conditions closely mimicking their

normal maintenance necessitates some differences in the assay protocols for the two sets of populations. For

JBs, flies from these assay populations were collected on day 12 from egg-lay, corresponding to the time all

flies would have eclosed in their regular cultures, pairing five males and five females per vial with fresh banana-

jaggery food medium (12 vials per population) to facilitate courtship observations. Observation vials were again

set up on days 14 and 16 as described for day 12. For observing flies in the last two days in cages prior to egg

lay, we set up 100 pairs of flies per Plexiglas cage ($22 \times 18 \times 18$ cm^3) with a plate of fresh food medium

covered with a paste of live yeast, as in their regular maintenance. We set up three replicate observation cages

per population; these were slightly smaller than the maintenance cages, to facilitate behavioural observations.

Flies were first collected into a regular cage (from holding vials in case of JBs and culture vials in case of FEJs)

and then 100 males and 100 females were lightly anesthetized (using carbon dioxide) and transferred into each

replicate observation cage. In case of the fast developing FEJs, only the first ~25% of the eclosing flies (~day

6.5 from eggs) were selected to become part of the observation set and directly shifted to observation cages with

a plate of fresh food medium covered with a paste of live yeast, as in their regular maintenance.

We took three 'instantaneous' observations every four hours (corresponding to one time point) starting

from day one of observation (i.e., day 12 from JB culture initiation (egg-lay), ~day 6.5 from FEJ culture

initiation) noting the number of males observed to be performing a courtship behaviour (Spieth 1974) towards a

female, and the number of pairs in copula. Observations were conducted over 144 hours (six days) in the vials

for only JBs, as the FEJs do not have a corresponding vial stage as adults in their regular maintenance, and for

72 hours (three days) in Plexiglas cages for both JBs and FEJs, for each replicate population. We calculated the

daily average of courtship and mating frequency (fraction of males that were courting or mating) for each day.

Daily average frequencies were then summed over all days of the effective lifetime, i.e., nine days for JB

populations and three days for FEJ populations, each for mating or courtship.

<u>Copulation Duration and Female Maturation Time</u>

To obtain adults of the same age at the beginning of the assay, FEJ cultures were started about 60 hours

after the JB cultures, to account for the difference in their egg to adult development times. We allowed an egg

laying time of only 30 minutes, in order to have a narrow eclosion distribution, and collected females that

emerged within a one-hour time window. Females were paired with one day old virgin males (single pair per

food vial), with 20-25 pairs per population, under each of two conditions (presence or absence of live-yeast

supplement on food) and kept undisturbed for continuous observation. In their regular maintenance, the first

matings occur in cages in the presence of food supplemented with additional yeast for FEJs, but in vials without

yeast for JBs. We recorded time from eclosion till first mating (maturation time) and copulation duration under

both yeasted and unyeasted conditions. The duration of observation was for 56 hours, or until 80% of pairs per

population, per treatment, had mated, whichever occurred first.

<u>Statistical Analyses</u>

We used mixed model analyses of variance (ANOVA) for data from all the assays. Average time till

maturation for a population, lifetime courtship and mating frequency per population, and proportion of wild-

type offspring averaged across females per population were used as the response variables. Tukey's HSD test

was used for comparing individual means and calculating all confidence intervals for post-hoc analyses. Since

some data were fractional, we also repeated each analysis for those response variables with arcsine square root

transformed values to check if there were any differences obtained in the significance of the fixed factors.

Selection regime and yeasted/non-yeasted conditions for time till maturation and copulation duration, and

selection regime for courtship and mating rate assays were treated as fixed factors in the analysis. All

experiments were performed on four replicate populations of a maintenance regime. Replicate populations with

matching numerical subscripts (FEJ$_i$ and JB$_i$; i = 1-4) were assayed together, and also share the same ancestry.

Those pairs were therefore treated as random blocks in the analyses. All analyses were carried out using

STATISTICA™ using Windows Release 5.0B (StatSoft Inc. 1995).

Results

<u>Mating and Courtship Frequencies</u>

The differences between FEJ and JB lifetime mating (F=1020.928, P< 0.0001) and courtship frequencies

(F=1372.439, P< 0.0001) were statistically significant, with JBs showing many fold higher frequencies of both

courtship and mating (Fig. 1a, b; Table 1). When we compared the mating and courtship frequencies only within

the cage stage (the last three days of their effective lifespan), courtship frequencies were still significantly

greater for JB than for FEJ (F=910.352, P<0.001), but their mating frequencies did not differ significantly

(F=4.163, P=0.1339; Fig. 1c, d; Table 1). Moreover, although overall courtship frequencies were very low for

FEJ, the results from the analysis of data from only within the cages indicated a very low courtship requirement

for arousal of FEJ females for copulation, given that mating frequencies in the cage were very similar for JB and

FEJ, despite courtship frequencies being much lower in FEJ (Fig. 1c, d). It is also worth noting that both

courtship and mating frequencies dropped by almost an order of magnitude for JB when only the cage stage was

considered, indicating that most mating in the regular cultures of JBs probably occurs during the vial transfer

stages, before the flies from all vials representing one replicate population are collected into a cage.

<u>Copulation Duration and Female Maturation Time</u>

Maturation time of the JB and FEJ differed significantly (F=149.885, P=0.0011, Table 2), as did

copulation duration (F=15.260, P=0.0297, Table 3), with JB females taking a shorter time from eclosion to first

mating than FEJ females, consistent with previous reports (Prasad 2004; Ghosh-Modak 2009), and mating for

longer than FEJ females (Fig. 2a, b). The drastic reduction in FEJ development time, especially pupal duration,

may have resulted in the postponement of many aspects of reproductive development to adulthood (Prasad

2004), causing FEJ females to take ~10 hours longer than JB females to mature (Fig. 2a). Although there is

considerable evidence for change in mating behaviour with adult diet and nutrition in flies (Fricke et al. 2008;

Schultzhaus et al. 2018; Duxbury and Chapman 2020), our results did not demonstrate any effect of additional

yeast on female maturation time or copulation duration; there was no significant main effect of environment or

of the selection-by-environment interaction (Table 2, 3). Female condition (adults being fed a protein rich diet in

this case) is also known to increase male courtship vigour through an increased attractiveness of the female

(Long et al. 2009), but such effects on the FEJ or JB males, or differences in these effects, did not manifest in

our assays.

Discussion

Sexual conflict is thought to be an incidental by-product of sexual selection (Trivers 1972, Parker 1979) and

sexual conflict can be affected by the intensity and scope of sexual selection. Several studies on insects have

investigated how reproductive strategies may evolve upon changing their breeding ecology. These studies

establish a strong relationship between sexual selection, altered by manipulating the degree of competition

among males for mating, and sexual conflict related traits. For example, populations of *Drosophila* sp. were

subjected to either monogamy (Holland and Rice 1999; Pitnick et al. 2001; Crudgington et al. 2005; Hollis et al.

2014; Wensing et al. 2017), or different operational sex ratios (Wigby and Chapman 2004; Linklater et al. 2007;

Edwards et al. 2010; Nandy et al. 2013a, b) in order to experimentally alter the degree of polygamy and,

therefore, sexual selection experienced by the flies. Traits that promote male-specific fitness reduced in the

monogamy adapted populations (Holland and Rice 1999; Pitnick et al. 2001; Crudgington et al. 2005; Wensing

et al. 2017), and females from these populations experienced greater fitness loss when paired with males

adapted to polygamy than to monogamy (Holland and Rice 1999; Crudgington et al. 2005). Similarly, females

from populations with male-biased sex ratio had higher resistance to mate harm compared to females from

female biased sex ratio populations (Wigby and Chapman 2004; Nandy et al. 2014). In contrast to these studies,

in our FEJ populations, sexual selection and sexual conflict have been altered as a correlated response to

selection on life-history traits like development time and adult age at effective reproduction.

Alterations in life history are also predicted to affect sexual selection and, consequently, the evolution

of sexually antagonistic traits (Bonduriansky et al. 2008; Adler and Bonduriansky 2014), and at least a subset of

these effects is expected to be through alterations in the breeding ecology of a population. Breeding ecology,

which includes not only the timing of reproductive activity, but also various other aspects of reproduction and

mating behaviour, such as male-female encounter rate, promiscuity, sexual dimorphism etc., is expected to be

defined by the life history strategy adopted by the population. For example, if males in a population have a

"live-fast, die-young" life history strategy, male-male competition is expected to be intense, all else being equal

(Bonduriansky et al. 2008, and references therein). A few investigations have shown the evolution of sexually

selected traits as a consequence of selection for life history traits such as lifespan (Service 1993; Service and

Fales 1993; Service and Vossbrink 1996; Makklakov et al. 2010). These reports generally support the idea that

evolution of life history traits can result in changes in male reproductive investment, which may further result in

reduction in sexual conflict in the population. Yun et al. (2017) showed that the sexual conflict in a population

living in a physically complex habitat can be less than that in a simple habitat. If this effect of habitat type is due

to females' access to a spatial refuge, then male-female encounter rate is directly implicated as an important

determinant of the level of conflict in that population. Therefore, factors such as, locomotor activity, opportunity

of interaction etc. are also expected to be important modulators of the conflict.

Our results complement and extend these studies, with our observations on breeding ecology of

laboratory populations of *Drosophila melanogaster* that have earlier been shown to have evolved reduced levels

of inter-locus sexual conflict as a correlated response to selection for rapid egg-to-adult development and early

reproduction (Ghosh and Joshi 2012; Mital et al. 2021), suggesting a reduced opportunity for sexual selection in

these populations. A reduction in the time available for mating in the FEJ populations appears to considerably

reduce reproductive competition by constraining re-mating opportunities. Thus, the lower levels of sexual

conflict in these populations, compared to their ancestral controls (Ghosh and Joshi 2012), earlier shown to be

partly due to their smaller body size (Mital et al. 2021), are also likely to be partly a consequence of reduced

sexual selection and male-male competition for mates. Moreover, we also find males from our selected

populations to have substantially lower courtship frequency and shorter copulation duration, indicating

potentially reduced investment in pre- and post-copulatory competition. Taken together, our present study and

those of Ghosh and Joshi (2012) and Mital et al. (2021), to our knowledge, constitute one among very few

examples directly connecting life-history evolution, sexual selection and sexual conflict.

In conditions closely approximating the regular maintenance of the FEJ and JB populations, we found

that the FEJ flies have a much lower mating rate than the JB controls. To better interpret the large difference in

life-time mating rates between the two sets of populations, we scaled the mating frequency results by the

maximum possible number of matings achievable in that time for that population. We multiplied the FEJ mating

rate by 24 (maximal possible mating events being once every hour) to obtain an expected number of life-time

mating events, which got rounded off to one mating during their effective adult life. In comparison, the

projected life-time mating events for a JB fly is six matings during their effective adult life (scaling factor of 12,

mating once every two hours). These scaling factors were chosen based on the mating duration and likely

recovery required by FEJ and JB males before attempting re-mating (A Mital, personal observation) and we

believe these are overestimates of the maximum number of matings possible within a day. Therefore, one and

six are conservative estimates of the expected lifetime mating events per fly for FEJs and JBs, respectively.

One limitation of our assay is that it only tells us that FEJ adults in their regular cultures are, on an

average, probably getting to mate just once, as compared to an average of six matings for JB adults in their

cultures. In the absence of data on the variance of male mating success in these populations, it could be

speculated that the FEJ populations are likely monandrous, since females are typically refractive after a mating

and tend not to re-mate for quite some time, whereas some males could be obtaining multiple matings and others

none at all. In that case, although post-copulatory sexual selection would be low, there might be substantial pre-

copulatory sexual selection on males in the FEJ populations. However, we believe this is unlikely to be the case

in the FEJ populations, for the following reason.

There has, to our knowledge, been only one study of the variance of male mating success in large,

outbred *Drosophila* populations under assay conditions approximating their running cultures (Joshi et al. 1999).

That study was carried out on the B populations of Rose (1984), which are the ultimate ancestors of our JB

control populations but differ in having been maintained on a 14 day discrete generation cycle with no cage

maintenance at any life-stage. Joshi et al. (1999) examined the mating success of individual marked males over

the relevant part of adult life. Except for early eclosing males that found themselves in vials with a female-

biased sex-ratio, there was no evidence for a non-Poisson distribution of male mating success (Joshi et al. 1999).

Even those early eclosing males showed a non-Poisson distribution of male mating success in only three out of

five replicate populations, and in each of those populations, the variance in male mating success was less than

half of the mean. Early eclosing males, thus, were obtaining a large number of matings, due to the availability of

more than one female per male, but mating success varied even less than expected through chance alone if all

males had equal probabilities of mating (Joshi et al. 1999).

If we examine the life-history in the FEJ, JB, B and UU (first described in Shiotsugu et al. 1997)

populations, we can predict that it is likely that the FEJ populations have both reduced pre- and post-copulatory

sexual selection, compared to the JB controls. The B populations (Rose 1984) are ancestral to the rest; the UU

were derived from the B in 1991 and shifted from a two week to a three week discrete generation cycle

(Shiotsugu et al. 1997), the JB were derived from the UU in 1997 and remained on a three week cycle (Sheeba

et al. 1998), and the FEJ were derived from the JB in 1998 and subjected to selection for rapid development and

early reproduction (Prasad et al. 2000). The B populations were not specifically selected for rapid development

by truncating the eclosion distribution. However, compared to the UU populations, the B populations showed

reduced development time (Shiotsugu et al. 1997); the development times of the UU and JB populations are

comparable, though they were never assayed together. The development time data coincide well with the

effective adult life span of these various sets of populations: FEJ are the fastest developing and have an effective

adult life of about 3.5 days, followed by the B with an effective adult life of about 5 days, and then the UU and

JB that are the slowest developing and have an effective adult life of about 11 days. In the B populations, the

number of matings obtained by a male in its effective lifetime ranged from 0 to 7, with a mean of 1.8 (data in

Table 1, Joshi et al. 1999). This mean mating success, based on direct observations of marked males, is

considerably lower than what we infer for the JB controls from our assay (6 lifetime matings), but higher than

what we infer for the FEJ (1 lifetime mating). Given these observations on the B populations, which have

somewhat rapid development and short effective lifespan, we expect that is likely that males in our FEJ

populations are undergoing considerably less pre- and post-copulatory sexual selection as compared to the JB

controls.

Another result indicating considerable change in reproductive behaviour of our selected populations is

the large reduction in their lifetime courtship frequency, compared to controls (Fig. 1b), which persisted even in

the cage stage (last three days of effective adult life, Fig. 1d) whereas similar differences in mating frequency

were not seen in the cage stage (Fig. 1c). There are multiple plausible causes for this result, as we discuss

below.

The period between eclosion and first reproduction (maturation time) is typically the only opportunity

for adult females to feed and thereby accumulate additional resources for egg production (Luckinbill et al 1985;

Chippindale et al. 1997b). Such compensatory feeding would be especially important for FEJ females given

their small size and low lipid content at eclosion (Prasad 2004). The already resource limited FEJ females are

expected to then reduce all energetically expensive activities, which may render increased receptivity selectively

advantageous. That FEJ males are significantly less harmful and females less resistant to mate harm has already

been reported earlier (Ghosh and Joshi 2012, Mital et al. 2021). Moreover, frugal courtship by males and a low

arousal threshold in females is likely to be selectively advantageous also because mating at least once is

necessary to have any fitness, and the time available for reproduction in the FEJ breeding ecology is short and

further reduced by a long maturation time, (Fig. 2a). In addition, since FEJs have also evolved a significantly

different developmental schedule and maturation time (Prasad 2004; Ghosh-Modak 2009; Fig. 2a),

developmental changes may further contribute to poor courtship by males, especially since males complete

reproductive activity at a very young adult age of about three days. For instance, in some studies on *D.*

melanogaster, males have been found to keep maturing reproductively, especially in terms of accessory gland

growth, for up to six days after eclosion, with younger males tending to have poor sperm competitive ability and

courtship effort as compared to more mature males (Ruhmann et al. 2016). Finally, males from the controls JB

populations may be selected for high courtship effort during the final three days of their adult life, as they have

about 80% paternity share of the offspring that form the next generation if they mate with females during the

'cage' stage (Supplementary Material, Fig. S1) (Fig. 2c, d). JB males would gain a high fitness reward for

mating effort in this time despite low chance of mating success. This may also indicate high variation in male

mating success in the control population, a long-standing argument for stronger selection in males for

exaggerated reproductive traits (Bateman 1948; Andersson 1994).

More generally, however, low courtship frequencies despite similar mating frequencies among males

that we see in the cage stage (Fig. 1c, d) can also be explained in the context of the chase-away selection model

for the evolution of exaggerated sexual traits in *D. melanogaster* (Holland and Rice 1998; Gavrilets et al. 2001).

The model suggests that if persistent male mating effort is harmful to females (sexual conflict) then high female

resistance towards such sensory manipulation will evolve. This may be interpreted as increased female

bias/preference, in turn selecting for higher courtship effort from males (Holland and Rice 1998, 1999). Since

persistent male courtship is known to reduce female survival in *D. melanogaster* (Partridge and Fowler 1990),

reduced sexual conflict driven by low male-male competition in FEJs may release the sexes from this co-

evolutionary cycle, and appear as lack of female choice and male courtship effort. Therefore, low courtship

frequency in FEJs, nevertheless resulting in amating frequency similar to that of the JBs in the last three days of

effective adult life may be driven by female ecological constraints, changes in development, reduced male-male

competition, or a combination of these. Put together, our results on the lifetime mating and courtship frequencies

in the FEJ populations r suggest the evolution of reduced sexual selection and conflict as a consequence of

strong selection for rapid development and an extremely short effective adult life.

The interplay between life-history and sexual selection is expected to ultimately determine the

optimum investment strategy for a population. In our study populations, the changes that have likely resulted in

reduced sexual selection are also expected to release the flies from certain reproductive investment demands.

We observed that FEJ flies mate for around 15 minutes on average, compared to about 25 minutes for JB flies

(Fig. 2b). Production of Acps is known to be costly to males (Chapman and Edwards 2011) and it has been

suggested that in *D. melanogaster*, most sperm transfer occurs during the first half of mating (Gilchrist and

Partridge 2000), the remaining time being spent in the transfer of Acps responsible for the female post-mating

response. We speculate that, reduced energy investments in courtship and in the non-sperm components of the

ejaculate in the FEJs would not only explain their shorter copulation duration (Fig. 2b) but potentially also

permit the evolution of even smaller flies, further pushing the boundaries of rapid development for which they

experience strong selection. Changes in resource use in FEJs have been demonstrated previously; FEJs have

evolved larger egg size (B M Prakash and A Joshi, unpublished data) and more eggs produced per unit dry

weight by females (Prasad 2004; Ghosh-Modak 2009) than JBs. It is therefore likely that the optimum

investment strategy for male FEJs has diverged from that of the ancestral controls. This is further supported by

the observations that greater starvation resistance per unit lipid has evolved in the FEJs than JBs (Prasad 2004),

and that Acp genes in young FEJ males are downregulated compared to the JBs (Satish 2010; K M Satish, P

Dey and A Joshi, unpublished data).

Conclusion

Our results, taken together with those from the studies of Joshi et al. (1999), Ghosh and Joshi (2012), and Mital

et al. (2021), strongly suggest that the changed breeding ecology of our fast developing and early reproducing

FEJ populations has likely reduced both pre- and post-copulatory sexual selection and, consequently, facilitated

the earlier observed evolution of lower inter-locus sexual conflict experienced by them, as compared to their

controls. The shorter effective adult life of FEJs, together with strong directional selection for rapid

development, has resulted in small flies that appear to exhibit substantially reduced competition among males

for mating. These observations complement other studies that have reported reduced sexual conflict upon

directly selecting for different levels of male-male competition (Introduction) and exemplify the potential

impact of selection on life-history related traits (development time and age at reproduction) on levels of sexual

selection and conflict via major changes in reproductive behaviour. This is especially relevant as differences in

sexual selection and sexual conflict are often implicated in the emergence of speciation phenotypes (Parker and

Partridge 1998). We note that the FEJs and JBs have diverged in their reproductive behaviour to a degree that

incipient reproductive isolation has occurred (Ghosh and Joshi 2012). Our work, therefore, also highlights that

selection on traits not directly associated with sexual conflict can, nonetheless, drive the evolution of

reproductively isolating mechanisms, either directly through rapid development (Mital et al. 2021) or indirectly

by affecting sexual selection, or both. Dissecting out such nuanced interactions of life-history and reproductive

strategy in affecting sexual selection and sexual conflict is important for understanding the myriad evolutionary

consequences that may accompany adaptation to specific ecological challenges.

438 **Tables**

439 **Table 1** Results summary of ANOVA for effect of selection on mating and courtship frequencies[1]

Response Variable	df	MS	*F*	*P*
Lifetime Mating Fr	1	0.5502	1020.928	< 0.0001
Lifetime Courtship Fr	1	120.2025	1372.439	< 0.0001
Mating Fr in cage	1	0.0001	4.163	0.1339
Courtship Fr in cage	1	0.0010	910.352	<0.001

446 **Table 2** Results summary of ANOVA for maturation time[2]

Effect	df	MS	*F*	*P*
Selection	1	535.6813	149.885	0.0011
Environment	1	1.6626	0.203	0.6823

---

[1] Summary of results of a one-way ANOVA each for lifetime and cage mating and courtship frequencies. Main effects of selection regime for the two traits are shown. In this design, random factors and interaction effects cannot be tested for significance and have been left out for brevity.

[2] Selection regime and environment (yeasted or non-yeasted) are fixed factors in the ANOVA. Random factors and interactions cannot be tested for significance in this design, and have been left out for brevity.

Selection × Environment	1	1.0137	0.003	0.9542

Table 3 Results summary of ANOVA for copulation duration[3]

Effect	df	MS	*F*	*P*
Selection	1	143.8742	15.260	0.0297
Environment	1	0.6389	0.6460	0.4802
Selection × Environment	1	0.0867	0.0326	0.8680

---

[3] Selection regime and environment (yeasted or non-yeasted) are fixed factors in the ANOVA. Random factors and interactions cannot be tested for significance in this design, and have been left out for brevity.

Figures

Fig. 1 a) Mean lifetime mating frequency averaged across the four replicate populations of JB and FEJ. b) Mean lifetime

courtship frequency averaged across four replicate populations of JB and FEJ. c) Mean mating frequency in the cage, during

the final three days of effective adult life, averaged across four replicate populations of JB and FEJ. d) Mean courtship

frequency in the cage, averaged across the four replicate populations of JB and FEJ. Error bars are 95% confidence intervals

around the means.

508 Fig. 2 a) Mean maturation time, or mating latency of virgins from eclosion, averaged across four replicate populations of JB

509 and FEJ, under yeasted (Y) and non-yeasted (NY) conditions. b) Mean copulation duration of the first mating averaged

510 across four replicate populations of JB and FEJ, under yeasted (Y) and non-yeasted (NY) conditions. Error bars are 95%

511 confidence intervals around the means.

References

Adler MI, Bonduriansky R (2014) Sexual conflict, life span, and aging. Cold Spring Harb Perspect Biol. 6:a017566. https://doi: 10.1101/cshperspect.a017566

Andersson M (1994) Sexual Selection. Princeton University Press, Princeton, USA

Anholt RHR, O'Grady P, Wolfner MF, Harbison ST (2020) Evolution of reproductive behaviour. Genetics 214:49–73 https://doi.org/10.1534/genetics.119.302263

Archer CR, Zajitschek F, Sakaluk SK, Royle NJ, Hunt J (2012) Sexual selection affects the evolution of lifespan and ageing in the decorated cricket *Gryllodes sigillatus*. Evolution 66:3088–100 https://doi.org/10.1111/j.1558-5646.2012.01673.x

Arnqvist G, Rowe L (2013) Sexual Conflict. Princeton University Press, Princeton, USA

Bateman AJ (1948) Intra-sexual selection in *Drosophila*. Heredity 2:349–368 https://doi.org/10.1038/hdy.1948.21

Bonduriansky R, Makklakov A, Zajitschek F, Brooks R (2008) Sexual selection, sexual conflict and the evolution of ageing and lifespan. Func Ecol 22:443–453 https://doi.org/10.1111/j.1365-2435.2008.01417.x

Chapman T, Liddle LF, Kalb JM, Wolfner MF, Partridge L (1995) Cost of mating in *Drosophila melanogaster* females is mediated by male accessory gland products. Nature 373:241–244 https://doi.org/10.1038/373241a0

Chapman T (2001) Seminal fluid-mediated fitness traits in *Drosophila*. Heredity 87:511 https://doi.org/10.1046/j.1365-2540.2001.00961.x

Chapman T, Arnqvist G, Bangham J, Rowe L (2003) Sexual conflict. Trends Ecol Evol 18:41–47

Chapman T and Edward D (2011) Mechanisms underlying reproductive trade-offs: cost of reproduction. Pp. 137– 152 *in* T. Flatt and A. Heyland, eds. Life-history evolution: understanding the proximate mechanism. Oxford Univ. Press, Oxford, U.K. https://doi.org/10.1093/acprof:oso/9780199568765.003.0011

Chippindale AK, Alipaz AJ, Chen HW, Rose MR (1997a) Experimental evolution of accelerated development in *Drosophila*. 1. Developmental speed and larval survival. Evolution 51:1536–1551 https://doi.org/10.1111/j.1558-5646.1997.tb01477.x

Chippindale AK, Leroi AM, Saing H, Borash DJ, Rose MR (1997b) Phenotypic plasticity and selection in *Drosophila* life history evolution. 2. Diet, mates and the cost of reproduction. J Evol Biol 10:269–293

Cordts R, Partridge L (1996) Courtship reduces longevity of male *Drosophila melanogaster*. Anim Behav 52:269–278

Crudgington HS, Beckerman AP, Brüstle L, Green K, Snook RR (2005) Experimental removal and elevation of

sexual selection: does sexual selection generate manipulative males and resistant females? Am Nat 165:72–87

https://doi.org/10.1086/429353

Darwin C (1859) On the origin of species by means of natural selection, or the preservation of favoured races in

the struggle for life. Murray, London, UK

Darwin C (1868) The variation of animals and plants under domestication (2 Vols). Murray, London, UK

Darwin C (1871) The descent of man and selection in relation to sex (2 Vols). Murray, London, UK (Facsimilie:

Princeton University Press, Princeton, NJ, USA, 1981)

Duxbury EML, Chapman T (2020) Sex-specific responses of life span and fitness variation in developmental vs

adult   diets   in   *Drosophila   melanogaster*.   J   Gerontol   Biol   Sci   75:1431–1438

https://doi.org/10.1093/gerona/glz175

Edward DA, Fricke C, Chapman T (2010) Adaptations to sexual selection and sexual conflict: insights from

experimental evolution and artificial selection. Philos Trans R Soc Lond., B, Biol Sci 365:2541–2548

https://dx.doi.org/10.1098%2Frstb.2010.0027

Fricke C, Bretman A, Chapman T (2008) Adult male nutrition and reproductive success in *Drosophila*

melanogaster. Evolution 62:3170–3177 https://doi.org/10.1111/j.1558-5646.2008.00515.x

Garlovsky MD, Holman L, Brooks AL, Snook RR (2021) Sexual selection affects the evolution of physiological

and life-history traits. BioRxiv https://doi.org/10.1101/2021.03.23.436586

Gavrilets S, Arnqvist G, Friberg U (2001) The evolution of female mate choice by sexual conflict. Proc Royal

Soc B: Biol Sci 268:531–539 https://dx.doi.org/10.1098%2Frspb.2000.1382

Ghosh-Modak, S (2009) Evolution of life-history traits, canalization and reproductive isolation in laboratory

populations of *Drosophila melanogaster* selected for faster pre-adult development and early reproduction

(Doctoral dissertation), Jawaharlal Nehru Centre for Advanced Scientific Research, Bengaluru, India

Ghosh SM, Joshi A (2012) Evolution of reproductive isolation as a by‐product of divergent life‐history

evolution   in   laboratory   populations   of   *Drosophila   melanogaster*.   Ecol   Evol   2:3214–3226

https://dx.doi.org/10.1002%2Fece3.413

Gilchrist AS, Partridge L (2000) Why it is difficult to model sperm displacement in *Drosophila melanogaster*:

the   relation   between   sperm   transfer   and   copulation   duration.   Evolution   54:534–542

https://doi.org/10.1111/j.0014-3820.2000.tb00056.x

Holland B, Rice WR (1998) Perspective: chase☐away sexual selection: antagonistic seduction versus resistance.

Evolution 52:1–7 https://doi.org/10.1111/j.1558-5646.1998.tb05132.x

Holland B, Rice WR (1999) Experimental removal of sexual selection reverses intersexual antagonistic co-

evolution and removes a reproductive load. PNAS 96:5083–5088 https://doi.org/10.1073/pnas.96.9.5083

Hollis B, Houle D, Yan Z, Kawecki TJ, Keller L (2014) Evolution under monogamy feminizes gene expression

in *Drosophila melanogaster*. Nat Commun 5:3482 https://doi.org/10.1038/ncomms4482

Hollis B, Keller L, Kawecki TJ (2017) Sexual selection shapes development and maturation rates in *Drosophila*.

Evolution 71:304–-314 https://doi.org/10.1111/evo.13115

Joshi A, Do MH, Mueller LD (1999) Poisson distribution of male mating success in laboratory populations of

Drosophila melanogaster. Genet. Res. 73:239–249

Kuijper B and Morrow EH (2009) Direct observation of female mating frequency using time lapse photography.

FLY 3:118–120 https://doi.org/10.4161/fly.8053

Linklater JR, Wertheim B, Wigby S, Chapman T (2007) Ejaculate depletion patterns evolve in response to

experimental manipulation of sex ratio in *Drosophila melanogaster*. Evolution 61:2027–2034

https://doi.org/10.1111/j.1558-5646.2007.00157.x

Long TAF, Pischedda A, Stewart AD, Rice WR (2009) A cost of sexual attractiveness to high-fitness females.

PLoS Biol 7:e1000254 https://doi.org/10.1371/journal.pbio.1000254

Luckinbill LS, Clare MJ (1985) Selection for life span in *Drosophila melanogaster*. Heredity 55:9–18

https://doi.org/10.1038/hdy.1985.66

Makklakov AA, Cayeteno L, Brooks RC, Bonduriansky R (2010) The roles of life-history selection and sexual

selection in the adaptive evolution of mating behaviour in a seed beetle. Evolution 64:1273–1282

https://doi.org/10.1111/j.1558-5646.2009.00904.x

Mital A, Sarangi M, Joshi A (2021) Evolution of lower levels of inter-locus sexual conflict in *D. melanogaster*

populations under strong selection for rapid development. BioRxiv https://doi.org/10.1101/2021.02.08.430125

Nandy B, Chakraborty P, Gupta V, Ali SZ, Prasad NG (2013)*a* Sperm competitive ability evolves in response to

experimental alteration of operational sex ratio. Evolution 67:2133–2141 https://doi.org/10.1111/evo.12076

Nandy B, Gupta V, Sen S, Udaykumar N, Samant MA, Ali SZ, Prasad NG (2013)*b* Evolution of mate-harm,

longevity and behaviour in male fruit flies subjected to different levels of inter-locus conflict. BMC Evol Biol

13:212 https://doi.org/10.1186/1471-2148-13-212

Nandy B, Gupta V, Udaykumar N, Samant MA, Sen S, Prasad NG (2014) Experimental evolution of female traits under different levels of intersexual conflict in *Drosophila melanogaster*. Evolution 68:412–425 https://doi.org/10.1111/evo.12271

Nunney L (1996) The response to selection for fast larval development in *Drosophila melanogaster* and its effect on adult weight: an example of a fitness trade☐off. Evolution 50:1193–1204 https://doi.org/10.1111/j.1558-5646.1996.tb02360.x

Parker GA (1979) Sexual selection and sexual conflict. Pages 123–166 in M. S. Blum and N. A. Blum, eds. Sexual Selection and Reproductive Competition in Insects, Academic Press, Athens, GA, USA

Partridge L, Fowler K, Trevitt S, Sharp W (1986) An examination of the effects of males on the survival and egg-production rates of female *Drosophila melanogaster*. J Insect Physiol 32:925–929 https://doi.org/10.1016/0022-1910(86)90140-X

Partridge L, Hoffmann A, Jones JS (1987)*a* Male size and mating success in *D. melanogaster* and *D. pseudoobscura* under field conditions. Anim Behav 35:468–476 https://doi.org/10.1016/S0003-3472(87)80272-5

Partridge L, Ewing A, Chandler A (1987)*b* Male size and mating success in *Drosophila melanogaster*: the roles of male and female behaviour. Anim Behav 35:555–562 https://doi.org/10.1016/S0003-3472(87)80281-6

Partridge L, Fowler K (1990) Non-mating costs of exposure to males in female *Drosophila melanogaster*. J Insect Physiol 36:419–425 https://doi.org/10.1016/0022-1910(90)90059-O

Pitnick S (1991) Male size influences mate fecundity and remating interval in *Drosophila melanogaster*. Anim Behav 41:735–745 https://psycnet.apa.org/doi/10.1016/S0003-3472(05)80340-9

Pitnick S, Miller GT, Reagan J, Holland B (2001) Males' evolutionary responses to experimental removal of sexual selection. Proc Royal Soc B: Biol Sci 268:1071–1080 https://doi.org/10.1098/rspb.2001.1621

Pitnick S, García–González F (2002) Harm to females increases with male body size in *Drosophila melanogaster*. Proc Royal Soc B: Biol Sci 269:1821–1828 https://doi.org/10.1098/rspb.2002.2090

Prasad NG, Shakarad M, Gohil VM, Sheeba V, Rajamani M, Joshi A (2000) Evolution of reduced pre-adult viability and larval growth rate in laboratory populations of *Drosophila melanogaster* selected for shorter development time. Genet Res 76:249–259 https://doi.org/10.1017/s0016672300004754

Prasad NG (2004) *Life-history evolution in laboratory populations of Drosophila melanogaster subjected to selection for faster development and early reproduction* (Doctoral dissertation). Jawaharlal Nehru Centre for Advanced Scientific Research, Bengaluru, India

Parker GA, Partridge L (1998) Sexual conflict and speciation. Philos Trans R Soc Lond B, Biol Sci 353:261–74

https://doi: 10.1098/rstb.1998.0208

Rose MR (1984) Laboratory evolution of postponed senescence in *Drosophila melanogaster*. Evolution

38:1004–1010

Ruhmann H, Wensing KU, Neuhalfen N, Specker JH, Fricke C (2016) Early reproductive success in *Drosophila*

males is dependent on maturity of the accessory gland. Behav Ecol 27:1859–1868

https://doi.org/10.1093/beheco/arw123

Satish KM (2010) Reverse evolution and gene expression studies on populations of *Drosophila melanogaster*

selected for rapid pre-adult development and early reproduction (Doctoral dissertation). Jawaharlal Nehru

Centre for Advanced Scientific Research, Bengaluru, India

Schultzhaus JN, Bennet CJ, Iftikar H, Yew JY, Mallet J, Carney GE (2018) High-fat diet alters *Drosophila*

melanogaster sexual behaviour and traits: decreased attractiveness and changes in pheromone profiles. Sci Rep

8:5387 https://doi.org/10.1038/s41598-018-23662-2

Service PM (1993) Laboratory evolution of longevity and reproductive fitness components in male fruit flies:

mating ability. Evolution 47:387–399 https://doi.org/10.2307/2410059

Service PM, Fales AJ (1993) Evolution of delayed reproductive senescence in male fruit flies: sperm

competition. Genetica 91:111–125 https://doi.org/10.1007/BF01435992

Service PM, Vossbrink RE (1996) Genetic variation in "first" male effects on egg laying and re-mating by

female *Drosophila melanogaster*. Behav Genet 26:39–48 https://doi.org/10.1007/BF02361157

Sheeba V, Madhyastha NA, Joshi A (1998) Oviposition preference for novel versus normal food resources in

laboratory populations of *Drosophila melanogaster*. J Biosci 23:93–100 https://doi.org/10.1007/BF02703000

Shiotsugu J, Leroi AM, Yashiro H, Rose MR, Mueller LD (1997) The symmetry of correlated selection

responses in adaptive evolution: an experimental study using *Drosophila*. Evolution 51:163–172

Spieth HT (1974) Courtship behaviour in *Drosophila*. Annu Rev Entomol 19:385–405

https://doi.org/10.1146/annurev.en.19.010174.002125

StatSoft Inc. (1995) STATISTICA^{TM} for Windows. Tulsa, OK, USA

Trivers R (1972) Parental Investment and Sexual Selection. Pages 136–179 *in* B. Cambell, ed. Sexual Selection

& the Descent of Man, Aldine de Gruyter, New York, USA

Wensing KU, Koppik M, Fricke C (2017) Precopulatory but not postcopulatory male reproductive traits diverge

in response to mating system manipulation in *Drosophila melanogaster*. Ecol and Evol 7:10361–10378

https://doi.org/10.1002/ece3.3542

Wigby S, Chapman T (2004) Female resistance to male harm evolves in response to manipulation of sexual

conflict. Evolution 58:1028–1037 https://doi.org/10.1111/j.0014-3820.2004.tb00436.x

Wolfner MF (1997) Tokens of love: functions and regulation of *Drosophila* male accessory gland products.

Insect Biochem Mol Biol 27:179–192 https://doi.org/10.1016/s0965-1748(96)00084-7

Yun L, Chen PJ, Singh A, Agrawal AF, Rundle HD (2017) The physical environment mediates male harm and

its   effects   on   selection   in   females.   Proc   Royal   Soc   B: Biol   Sci   284:20170424

https://doi.org/10.1098/rspb.2017.0424

Zajitschek F, Bonduriansky R, Zajitschek SR, Brooks RC (2009) Sexual dimorphism in life history: age,

survival, and reproduction in male and female field crickets *Teleogryllus commodus* under seminatural

conditions. Am Nat 173:792–802 https://doi.org/10.1086/598486

Zwaan B, Bijlsma R, Hoekstra RF (1995) Artificial selection for developmental time in *Drosophila*

melanogaster in relation to the evolution of aging: direct and correlated responses. Evolution 49:635–648

https://doi.org/10.1111/j.1558-5646.1995.tb02300.x