Genetic mapping of sex and self-incompatibility determinants in the androdioecious plant *Phillyrea angustifolia*

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Abstract

The diversity of mating and sexual systems in Angiosperms is spectacular, but the factors driving their evolution remain poorly understood. In plants of the Oleaceae family, an unusual self-incompatibility (SI) system has been discovered recently, whereby only two distinct homomorphic SI specificities segregate stably. To understand the role of this peculiar SI system in preventing or promoting the diversity of sexual phenotypes observed across the family, an essential first step is to characterize the genetic architecture of these two traits. Here, we developed a high-density genetic map of the androdioecious shrub *P. angustifolia* based on a F1 cross between a hermaphrodite and a male parent with distinct SI genotypes. Using a double restriction-site associated digestion (ddRAD) sequencing approach, we obtained reliable genotypes for 196 offspring and their two parents at 10,388 markers. The resulting map comprises 23 linkage groups totaling 1,855.13 cM on the sex-averaged map. We found strong signals of association for the sex and SI phenotypes, that were each associated with a unique set of markers on linkage group 12 and 18 respectively, demonstrating inheritance of these traits as single, independent, mendelian factors. The *P. angustifolia* linkage map shows robust synteny to the olive tree genome overall. Two of the six markers strictly associated with SI in *P. angustifolia* have strong similarity with a recently identified 741kb chromosomal region fully linked to the SI phenotype on chromosome 18 of the olive tree genome, providing strong cross-validation support. The SI locus stands out as being markedly rearranged, while the sex locus has remained relatively more collinear between the two species. This *P. angustifolia* linkage map will be a useful resource to investigate the various ways by which the sex and SI determination systems have co-evolved in the broader phylogenetic context of the Oleaceae family.

Introduction

Modes of sexual reproduction are strikingly diverse across Angiosperms, both in terms of the proportion of autogamous vs. allogamous matings and in terms of the distribution of male and female sexual functions within and among individuals (BARRETT 1998; SAKAI AND WELLER 1999; DIGGLE *et al.* 2011). The conditions under which this diversity could arise under apparently similar ecological conditions and have evolved rapidly -sometimes even within the same family- have been a topic of intense interest in evolutionary biology (BARRETT 1998). The control of self-fertilization and the delicate balance between its costs and benefits is considered to be a central force driving this diversity. Avoidance of self-fertilization is sometimes associated with observable phenotypic variations among reciprocally compatible partners. These variations can be morphological (e.g. distyly) or temporal (e.g. protandry, protogyny in the case of heterodichogamy), but in many cases the flowers show no obvious morphological or phenological variation, and self-fertilization avoidance relies on so-called "homomorphic" self-incompatibility (SI) systems. These systems are defined as the inability of fertile hermaphrodite plants to produce zygotes through self-fertilization (LUNDQVIST 1956; DE NETTANCOURT 1977), and typically rely on the segregation of a finite number of recognition "specificities" whereby matings between individuals expressing cognate specificities are not successful at producing zygotes. At the genetic level, the SI specificities most commonly segregate as a single multi-allelic mendelian locus, the S locus. This locus contains at least two genes, one encoding the male determinant expressed in pollen and the other encoding the female determinant expressed in pistils, with the male specificity sometimes determined by a series of tandemly arranged paralogs (KUBO *et al.* 2015). The male and female determinants are both highly polymorphic and tightly linked, being inherited as a single non-recombining genetic unit. In cases where the molecular mechanisms controlling SI could be studied in detail, they were found to be remarkably diverse, illustrating their independent evolutionary origins across the flowering plants (IWANO AND TAKAYAMA 2012). Beyond the diversity of the molecular functions employed, SI systems can also differ in their genetic architecture.

In the Poaceae family for example, two independent loci (named S and Z) control SI (Yang, et al., 2008). In other cases, the alternate allelic specificities can be determined by presence-absence variants rather than nucleotide sequence variants of a given gene, such as *e.g.* in *Primula vulgaris*, where one of the two reproductive phenotypes is hemizygous rather than heterozygous for the SI locus (LI *et al.* 2016).

In spite of this diversity of molecular mechanisms and genetic architectures, a common feature of SI phenotypes is that they are all expected to evolve under negative frequency-dependent selection, a form of natural selection favoring the long-term maintenance of high levels of allelic diversity (WRIGHT 1939). Accordingly, large numbers of distinct SI alleles are commonly observed to segregate within natural and cultivated SI species (reviewed in CASTRIC AND VEKEMANS 2004). There are notable exceptions to this general rule, however, and in some species only two SI specificities seem to segregate stably. Most often in such diallelic SI systems, the two SI specificities are in perfect association with morphologically distinguishable floral phenotypes. In distylous species, for instance, two floral morphs called “pin” (L-morph) and “thrum” (S-morph) coexist (BARRETT 1992; BARRETT 2019). In each morph, the anthers and stigma are spatially separated within the flowers, but located at corresponding, reciprocal positions between the two morphs. Additional morphological differences exist, with S-morph flowers producing fewer but larger pollen grains than L-morph flowers (DULBERGER 1992). These morphological differences are believed to enhance the selfing avoidance conferred by the SI system but also to increase both male and female fitnesses (BARRETT 1990; BARRETT 2002; KELLER *et al.* 2014), although it is not clear which of SI or floral morphs became established in the first place (CHARLESWORTH AND CHARLESWORTH 1979).

The Oleacea family is another intriguing exception, where a diallelic SI system was recently found to be shared across the entire family (VERNET *et al.* 2016). In this family of trees, the genera *Jasminum* (2*n* = 26), *Fontanesia* (2*n* = 26) and *Forsythia* (2*n*= 28) are all heterostylous and are therefore all expected to possess a heteromorphic diallelic SI system; in *Jasminum fruticans* self- and within-morph fertilization are unsuccessful (DOMMÉE *et al.* 1992). The ancestral heterostyly gave rise to species with hermaphrodite (e.g.

Ligustrum vulgare, *Olea europaea*), androdioecious (e.g. *P. angustifolia*, *Fraxinus ornus*), polygamous (e.g. *Fraxinus excelsior*) and even dioecious (e.g. *Fraxinus chinensis*) sexual systems, possibly in association with a doubling of the number of chromosomes (2*n*= 46 in the Oleeae tribe) (TAYLOR 1945; WALLANDER AND ALBERT 2000). Evaluation of pollen germination success in controlled *in vitro* crossing experiments (whereby fluorescence microscopy is used to score the growth of pollen tubes reaching the style through the stigma; referred to below as the "stigma test") revealed the existence of a previously unsuspected homomorphic diallelic SI in one of these species, *P. angustifolia* (SAUMITOU-LAPRADE *et al.* 2010). In this androdioecious species (i.e. in which male and hermaphrodite individuals coexist in the same populations), hermaphrodite individuals form two morphologically indistinguishable groups of SI specificities that are reciprocally compatible but incompatible within groups, whereas males show compatibility with hermaphrodites of both groups (SAUMITOU-LAPRADE *et al.* 2010). This "universal" compatibility of males offsets the reproductive disadvantage they suffer from lack of their female function, such that the existence of the diallelic SI system provides a powerful explanation to the long-standing evolutionary puzzle represented by the maintenance of high frequencies of males in this species (PANNELL AND KORBECKA 2010; SAUMITOU-LAPRADE *et al.* 2010; BILLIARD *et al.* 2015; PANNELL AND VOILLEMOT 2015). Extension of the stigma test developed in *P. angustifolia* to other species of the same tribe including *L. vulgaris* (DE CAUWER *et al.* 2020), *F. ornus* (VERNET *et al.* 2016) and *O. europaea* (SAUMITOU-LAPRADE *et al.* 2017; DUPIN *et al.* 2020), demonstrated that all species exhibited some form of the diallelic SI system, but with no consistent association with floral morphology. Cross-species pollination experiments even showed that pollen from *P. angustifolia* was able to trigger a robust SI response on *O. europaea* and the more distant *F. ornus* and *F. excelsior* stigmas (the reciprocal is also true). This opens the question of whether the homomorphic diallelic SI determinants are orthologs across the Oleeae tribe, even in the face of the variety of sexual polymorphisms present in the different species. More broadly, the link between determinant of the homomorphic diallelic SI in the Oleeae tribe and those of the heteromorphic diallelic SI in the ancestral

109 diploid, largely heterostylous species, remains to be established (BARRETT 2019). Understanding the causes
110 of the long-term maintenance of this SI system and exploring its consequences on the evolution of sexual
111 systems in hermaphrodite, androdioecious, polygamous or dioecious species of the family represents an
112 important goal. The case of *P. angustifolia* is particularly interesting because it is one of the rare instances
113 where separate sexes decoupled from mating types can be studied in a single species (CHARLESWORTH
114 1978).

115 A first step toward a better understanding of the role of the diallelic SI system in promoting the
116 sexual diversity in Oleaceae is to characterize and compare the genetic architecture of the SI and sexual
117 phenotypes. At this stage, however, the genomic resources for most of these non-model species remain
118 limited. In this context, the recent sequencing efforts (UNVER *et al.* 2017; JIMÉNEZ-RUIZ *et al.* 2020) and the
119 genetic mapping of the SI locus in a biparental population segregating for SI groups in *Olea europaea*
120 (MARIOTTI *et al.* 2020) represent major breakthroughs in the search for the SI locus in Oleaceae. They have
121 narrowed down the SI locus to an interval of 5.4cM corresponding to a region of approximately 300kb, but
122 it is currently unknown whether the same region is controlling SI in other species. In *P. angustifolia,* based
123 on a series of genetic analysis of progenies from controlled crosses, Billiard *et al.* (2015) proposed a fairly
124 simple genetic model, where sex and SI are controlled by two independently segregating diallelic loci.
125 Under this model, sex would be determined by the "M" locus at which a dominant *M* allele codes for the
126 male phenotype (*i.e. M is a* female-sterility mutation leading e.g. to arrested development of the stigma)
127 and a recessive *m* allele codes for the hermaphrodite phenotype. The S locus would encode the SI system
128 and comprise a dominant allele *S2* and a recessive allele *S1.* The model thus hypothesizes that
129 hermaphrodites are homozygous *mm* at the sex locus, and fall into two groups of SI specificities, named
130 H_a and H_b carrying the *S1S1* and *S1S2* genotypes at the S locus, respectively (their complete genotypes
131 would thus be *mmS1S1* and *mmS1S2* respectively). The model also hypothesizes three male genotypes
132 (M_a: *mMS1S1*, M_b: *mMS1S2*, and M_c*: mMS2S2*). In addition, Billiard *et al.* (2015) experimentally showed

133 that, while males are compatible with all hermaphrodites, the segregation of sexual phenotypes varies
134 according to which group of hermaphrodites they sire: the progeny of H_a hermaphrodites pollinated by
135 males systematically consists of both hermaphrodites and males with a consistent but slight departure
136 from 1:1 ratio, while that of H_b hermaphrodites pollinated by the very same males systematically consists
137 of male individuals only. These segregation patterns suggests a pleiotropic effect of the *M* allele, conferring
138 not only female sterility and universal pollen compatibility, but also a complete male-biased sex-ratio
139 distortion when crossed with one of the two groups of hermaphrodites and a more subtle departure from
140 1:1 ratio when crossed with the other group of hermaphrodites (BILLIARD *et al.* 2015). The latter departure,
141 however, was observed on small progeny arrays only, and its magnitude thus comes with considerable
142 uncertainty.

143 In this study, we developed a high-density genetic map for the non-model tree *P. angustifolia* using
144 a ddRAD sequencing approach and used it to address three main questions related to the evolution of its
145 peculiar reproductive system. First, are the SI and sex phenotypes in *P. angustifolia* encoded by just two
146 independent loci, as predicted by the most likely segregation model of Billiard *et al.* (2015)? Second, which
147 genomic regions are associated with the SI and sex loci, and what segregation model do the SI and sex-
148 associated loci follow (i.e. which of the males or hermaphrodites, and which of the two SI phenotypes are
149 homozygous *vs.* heterozygous at either loci, or are these phenotypes under the control of hemizygous
150 genomic regions?). Third, what is the level of synteny between our *P. angustifolia* genetic map and the
151 recently published Olive tree genome (UNVER *et al.* 2017; MARIOTTI *et al.* 2020), both globally and
152 specifically at the SI and sex-associated loci?

153

154 ## Material and Methods

155 ### Experimental cross and cartography population

156 In order to get both the SI group and the sexual phenotype (males vs hermaphrodites) to segregate
157 in a single progeny array, a single maternal and a single paternal plant were chosen among the progenies
158 of the controlled crosses produced by (BILLIARD *et al.* 2015). Briefly, a H_a maternal tree (named 01.N-25,
159 with putative genotype mmS1S1) was chosen in the progeny of a (H_a x M_a) cross. It was crossed in March
160 2012 to a M_b father (named 13.A-06, putative genotype mM S1S2) chosen in the progeny of a (H_a x M_c)
161 cross, following the protocol of Saumitou-Laprade *et al.* (2010). Both trees were maintained at the
162 experimental garden of the "Plateforme des Terrains d'Expérience du LabEx CeMEB," (CEFE, CNRS) in
163 Montpellier, France. F1 seeds were collected in September 2012 and germinated in the greenhouse of the
164 "Plateforme Serre, cultures et terrains expérimentaux," at the University of Lille (France). Seedling
165 paternity was verified with two highly polymorphic microsatellite markers (VASSILIADIS *et al.* 2002), and
166 1,064 plants with confirmed paternity were installed in May 2013 on the experimental garden of the
167 "Plateforme des Terrains d'Expérience du LabEx CeMEB," (CEFE, CNRS) in Montpellier. Sexual phenotypes
168 were visually determined based on the absence of stigma for 1,021 F1 individuals during their first
169 flowering season in 2016 and 2017 (absence of stigma indicates male individuals). Twenty-one progenies
170 did not flower and 22 died during the test period. The hermaphrodite individuals were assigned to an SI
171 group using the stigma test previously described in Saumitou-Laprade *et al.* (2010; SAUMITOU-LAPRADE *et al.*
172 2017).

173

174 ### DNA extraction, library preparation and sequencing

175 In 2015, *i.e.* the year before sexual phenotypes were determined and stigma tests were
176 performed, 204 offspring were randomly selected for genomic library preparation and genotyping. Briefly,
177 DNA from parents and progenies was extracted from 100 mg of frozen young leaves with the Chemagic

178 DNA Plant Kit (Perkin Elmer Chemagen, Baesweller, DE, Part # CMG-194), according to the manufacturer's
179 instructions. The protocol was adapted to the use of the KingFisher Flex™ (Thermo Fisher Scientific,
180 Waltham, MA, USA) automated DNA purification workstation. The extracted DNA was quantified using a
181 Qubit fluorometer (Thermo Fisher Scientific, Illkirch, France). Genome complexity was reduced by double
182 digestion restriction associated DNA sequencing (ddRAD seq) (PETERSON *et al.* 2012) using two restriction
183 enzymes: *PstI*, a rare-cutting restriction enzyme sensitive to methylation recognizing the motif CTGCA/G,
184 and *MseI*, a common-cutting restriction enzyme (recognizing the motif T/TAA). The libraries were
185 constructed at the INRAE - AGAP facilities (Montpellier, France). Next-generation sequencing was
186 performed in a 150-bp paired-ends-read mode using three lanes on a HiSeq3000 sequencer (Illumina, San
187 Diego, CA, USA) at the Get-Plage core facility (Genotoul platform, INRAE Toulouse, France).
188

189 **GBS data analysis and linkage mapping**

190 Illumina sequences were quality filtered with the *process_radtags* program of Stacks v2.3
191 (CATCHEN *et al.* 2011) to remove low quality base calls and adapter sequences. We followed the Rochette
192 & Catchen protocol (ROCHETTE AND CATCHEN 2017) to obtain a *de novo* catalog of reference loci. Briefly, the
193 reads were assembled and aligned with a minimum stack depth of 3 (–m=3) and at most two nucleotide
194 differences when merging stacks into loci (–M=2). We allowed at most two nucleotide differences between
195 loci when building the catalog (–n=2). Both parental and all offspring FASTQ files were aligned to the *de*
196 *novo* catalog using Bowtie2 v2.2.6 (LANGMEAD AND SALZBERG 2012), the option 'end-to-end' and 'sensitive'
197 were used for the alignment. At this step, one .bam file was obtained per individual to construct the linkage
198 map with Lep-MAP3 (RASTAS 2017). A custom python script was used to remove SPN markers with reads
199 coverage <5. After this step, the script calls Samtools v1.3.1 and the script *pileupParser2.awk* (limit1=5) to
200 convert *.bam* files to the format used by Lep-MAP3. We used the *ParentCall2* module of Lep-MAP3 to
201 select loci with reliable parental genotypes by considering genotype information on parents and offspring.

202 The *Filtering2* module was then used to remove non-informative and distorted markers (dataTolerance =
203 0.0000001). The module *SeparateChromosomes2* assigned markers to linkage groups (LGs), after test,
204 where the logarithm of odds score (LodLimit) varied from 10 to 50 in steps of 5 then from 20 to 30 in steps
205 of 1 and the minimum number of SNP markers (sizeLimit) per linkage group from 50 to 500 in steps of 50
206 for each of the LodLimit. The two parameters, lodLimit = 27 and sizeLimit = 250, were chosen as the best
207 parameters to obtain the 23 linkage groups (as expected in members of the Oleoideae subfamily;
208 (WALLANDER AND ALBERT 2000). A custom python script removed loci with SNPs mapped on two or more
209 different linkage groups. The last module *OrderMarkers2* ordered the markers within each LG. To consider
210 the slight stochastic variation in marker distances between executions, the module was run three times
211 on each linkage group, first separately for the meiosis that took place in each parent (sexAveraged = 0)
212 and then averaged between the two parents (sexAveraged = 1). To produce the most likely final father and
213 mother specific maps and a final sex-averaged maps (DE-KAYNE AND FEULNER 2018), we kept for each map
214 the order of markers that had the highest likelihoods for each linkage group. In the end of some linkage
215 groups, we removed from the final genetic map markers that were clearly outliers i.e. that had orders of
216 magnitude more recombination to any marker than the typical average (Table 1). The original map is
217 provided in Figure S1.
218

219 **Sex and SI locus identification**

220 To identify the sex-determination system in *P. angustifolia* we considered two possible genetic
221 models. First, a "XY" male heterogametic system, where males are heterozygous or hemizygous (XY) and
222 hermaphodites are homozygous (XX). Second, a "ZW" hermaphrodite heterogametic system, where
223 hermaphodites are heterozygous or hemizygous (ZW) and males are homozygous (ZZ). We applied the
224 same logic to the SI determination system, as segregation patterns (Billiard et al. 2015) suggested that SI
225 possibly also has a heterogametic determination system, with homozygous H_a and heterozygous H_b. In the

226 same way as for sex, it is therefore possible to test the different models (XY, ZW or hemizygous) to
227 determine which SNPs are linked to the two SI phenotypes.

228 Based on this approach, we identified sex-linked and SI-linked markers on the genetic map by
229 employing SEX-DETector, a maximum-likelihood inference model initially designed to distinguish
230 autosomal from sex-linked genes based on segregation patterns in a cross (MUYLE *et al.* 2016). Briefly, a
231 new alignment of reads from each individual on the loci used to construct the linkage map was done with
232 bwa (LI AND DURBIN 2009). This new alignment has the advantage of retrieving more SNPs than used by
233 LepMap3, as SNPs considered as non-informative by LepMap3 can still be informative to distinguish among
234 sex- or SI-determination systems by SEX-DETector. The alignment was analyzed using Reads2snp (default
235 tool for SEX-DETector) (TSAGKOGEORGA *et al.* 2012) with option -par 0. We ran Reads2snp without the -aeb
236 (account for allelic expression bias) option to accomodate for the use of genomic rather than RNA-seq
237 data. For each phenotype (H_a vs. H_b and males vs. hermaphrodites), SEX-DETector was run for both a XY
238 and a ZW model with the following parameters: -detail, -L, -SEM, -thr 0.8, -E 0.05. For each run, SEX-
239 DETector also calculates the probability for X (or Z)-hemizygous segregation in the heterozygous
240 haplotypes. To compensate for the heterogeneity between the number of males (83) and hermaphrodites
241 (113) in our progeny array, each model was tested three times with sub-samples of 83 hermaphrodites
242 obtained by randomly drawing from the 113 individuals. We retained SNPs with a ≥80% probability of
243 following an XY (or ZW) segregation pattern, with a minimum of 50% individuals genotyped and less than
244 5% of the individuals departing from this model (due to either genotyping error or crossing-over).

245

246 **Synteny analysis with the olive tree**

247 To study synteny, we used basic local alignment search tool (BLAST) to find regions of local
248 similarity between the *P. angustifolia* ddRADseq loci in the linkage map and the *Olea europea var. sylvestris*
249 genome assembly (UNVER *et al.* 2017). This assembly is composed of 23 main chromosomes and a series

of 41,233 unanchored scaffolds for a total of 1,142,316,613 bp. Only loci with a unique hit with at least 85% identity over a minimum of 110 bp were selected for synteny analysis. Synteny relationships were visualized with *circos-0.69-6* (KRZYWINSKI *et al.* 2009). Synteny between linkage groups of *P. angustifolia* and the main 23 *O. europea* chromosomes was established based on the number of markers with a significant BLAST hit. At a finer scale, we also examined synteny with the smaller unanchored scaffolds of the assembly, as the history of rearrangement and allo-tetraploidization is likely to have disrupted synteny.

Data availability

Fastq files for all 204 offspring and both parents are deposited in the NCBI BioProject (SRA accession PRJNA724813). All scripts used can be accessed at https://github.com/Amelie-Carre/Genetic-map-of-Phillyrea-angustifolia .

261 ## Results

262 ### Phenotyping progenies for sex and SI groups

263 As expected, our cartography population segregated for sex and SI phenotypes, providing a
264 powerful resource to genetically map these two traits. Among the 1,021 F1 individuals that flowered
265 during the two seasons of phenotyping, we scored 619 hermaphrodites and 402 males, revealing a biased
266 sex ratio in favor of hermaphrodites (khi^2= 46.12, p-value=$1.28x10^{-11}$). Stigma tests were successfully
267 performed on 613 hermaphrodites (6 individuals flowered too late to be included in a stigma test),
268 revealing 316 H_a and 297 H_b, i.e. an equilibrated segregation of the two SI phenotypes (khi^2=1.22, p-value=
269 0.27). The random subsample of 204 F1 progenies chosen before the first flowering season for ddRAD-seq
270 analysis (see below) followed similar phenotypic proportions. Only 196 of the 204 progenies ended up
271 flowering, revealing 83 males and 113 hermaphrodites, among which 60 belonged to the H_a group and 53
272 to the H_b group.

273

274 ### Linkage mapping

275 The two parents and the 196 offspring that had flowered were successfully genotyped using a
276 ddRAD-seq approach. Our stringent filtering procedure identified 11,070 loci composed of 17,096 SNP
277 markers as being informative for Lep-MAP3. By choosing a LOD score of 27, a total of 10,388 loci composed
278 of 15,814 SNPs were assigned to, and arranged within, 23 linkage groups in both sex-averaged and sex-
279 specific maps (Table 1).

280 The linkage groups of the mother map were on average larger (78.88 cM) than the linkage groups
281 of the father map (73.40 cM) and varied from 22.73 cM to 112.38 cM and from 35 cM to 121.94 cM
282 respectively (Table 1, Figure S1). The total map lengths were 1586.57 cM, 1688.16 cM and 1814.19 cM in
283 the sex-averaged, male and female maps, respectively. The length of the linkage groups varied from 23.90

284 cM to 110.69 cM in the sex-averaged map, with an average of 683 SNPs markers per linkage group (Table
285 1).
286
287 **Sex and SI locus identification**

288 We found evidence that a region on linkage group 18 (LG18) was associated with the SI
289 phenotypes, with Hb hermaphrodites having heterozygous genotype, akin to a XY system. Indeed, when
290 comparing H_a and H_b, among the 38,998 SNPs analyzed by SEX-DETector, 496 had a probability of following
291 an XY pattern ≥0.80. We then applied two stringent filters by retaining only SNPs that had been genotyped
292 for more than 50% of the offspring (n=211), and for which less than 5% of the offspring departed from the
293 expected genotype under a XY model (n=23). Six of these 23 SNPs, distributed in 4 loci, followed a
294 segregation pattern strictly consistent with a XY model. These four loci are tightly clustered on the linkage
295 map and define a region of 1.230 cM on LG18 (Figure 1) in the sex-averaged map. Relaxing the stringency
296 or our thresholds, this region also contains five loci that strictly follow an XY segregation but with fewer
297 than 50% of offsprings successfully genotyped, as well as six loci with autosomal inheritance, possibly
298 corresponding to polymorphisms accumulated within allelic lineages associated with either of the
299 alternate SI specificities. Using the same filtering scheme, none of the SNPs was found to follow a ZW
300 pattern.

301 For the comparison of male and hermaphrodites, an average of 44,565 SNPs were analyzed by
302 SEX-DETector across the three subsamples, among which an average of 438 had a probability of following
303 an XY pattern ≥0.80. We applied the same set of stringent filters and retained an average of 171 SNPs
304 having been genotyped for at least 50% of the offspring, among which 41 had less than 5% of the offspring
305 departing from the expected genotype under a XY model and were shared across the three subsets. Thirty-
306 two of these SNPs followed a segregation pattern strictly consistent with a XY model. These 32 markers,
307 corresponding to 8 loci, are distributed along a region of 2.216 cM on linkage group 12 (LG12, Figure 1) in

308 the sex-averaged map. Relaxing the stringency or our thresholds, this region also contains five loci that
309 strictly follow an XY segregation pattern but with fewer than 50% of offspring successfully genotyped, as
310 well as 17 loci consistent with autosomal inheritance, possibly corresponding to polymorphisms
311 accumulated within allelic lineages associated with either of the alternate sex phenotypes. Again, no SNP
312 was found to follow a ZW pattern. This provides evidence that this independent region on LG12 is
313 associated with sex, with a determination system akin to a XY system where males have the heterogametic
314 genotype.
315

316 **Synteny analysis with the olive tree**

317 About half (49%) of the 10,388 *P. angustifolia* loci used for the genetic map had a significant BLAST
318 hit on the olive tree genome. Overall, the relative position of these hits was highly concordant with the
319 structure of the linkage map. Indeed, the vast majority (79.7%) of loci belonging to a given linkage group
320 had non-ambiguous matches on the same olive tree chromosome. Loci that did not follow this general
321 pattern did not cluster on other chromosomes, suggesting either small rearrangements or
322 mapping/assembly errors at the scale of individual loci. The order of loci within the linkage groups was
323 also well conserved with only limited evidence for rearrangements (Figure 2, Figure 3), suggesting that the
324 two genomes have remained largely collinear.

325 We then specifically inspected synteny between the linkage groups carrying either the sex or the
326 SI locus and the olive tree genome (Figure 4). Synteny was good for LG12, the linkage group containing the
327 markers associated with the sex phenotype. Among the 645 loci of LG12, 365 have good sequence
328 similarity in the olive tree genome. Eighty eight percent had their best hits on the same chromosome of
329 the olive tree (chromosome 12 per our numbering of the linkage groups), and the order of markers was
330 largely conserved along this chromosome. Six loci contained in the region associated with sex on LG12 had
331 hits on a single 1,940,009bp region on chromosome 12. This chromosomal interval contains 82 annotated

genes in the olive tree genome (Table S1). In addition, eight loci in the sex region had their best hits on a series of five smaller scaffolds (Sca393, Sca1196, Sca1264, Sca32932, Sca969) that could not be reliably anchored in the main olive tree assembly but may nevertheless also contain candidates for sex determination. Collectively, these scaffolds represent 1.849.345bp of sequence in the olive tree genome and contain 57 annotated genes (Table S1).

Synteny was markedly poorer for markers on LG18, the linkage group containing the markers associated with the SI specificity phenotypes (Figure 5). Of the 440 loci on LG18, 203 had non-ambiguous BLAST hits on the olive tree genome. Although a large proportion (89%) had their best hits on chromosome 18, the order of hits along that chromosome suggested a large number of rearrangements. This more rearranged order was also observed for the six markers that were strictly associated with SI in *P. angustifolia*. Two of them had hits on a single region of 741,403bp on the olive tree genome. This region contains 32 annotated genes (Table S2) and contains two markers that were previously found to be genetically associated with SI directly in the olive tree by Mariotti *et al.* (2020). Three markers more loosely associated with SI in *P. angustifolia* had hits on a more distant region on chromosome 18 (19,284,909-19,758,630Mb). The three other strongly associated markers all had hits on scaffold 269, which contains 15 annotated genes and represents 545,128bp. Nine other loci strongly or loosely associated with SI had hits on a series of seven other unanchored scaffolds (Sca1199, Sca1200, Sca1287, Sca1579, Sca213, Sca327, Sca502) that collectively represent 96 annotated genes (Table S2) and 2,539,637bp.

Discussion

Until now, studies have mostly relied on theoretical or limited genetic segregation analyses to investigate the evolution of sexual and SI phenotypes in *P. angustifolia* (VASSILIADIS *et al.* 2002; SAUMITOU-LAPRADE *et al.* 2010; HUSSE *et al.* 2013; BILLIARD *et al.* 2015). In this study, we created the first genetic map of the androdioecious species *P. angustifolia* and identified the genomic regions associated with these two

important reproductive phenotypes. The linkage map we obtained shows strong overall synteny with the olive tree genome, and reveals that sex and SI phenotypes segregate independently from one another, and are each strongly associated with a different genomic region (in LG18 and LG12, respectively).

The SI linked markers on LG18 are orthologous with the genomic interval recently identified by Mariotti *et al.* (2020) as the region controlling SI in the domesticated olive tree, providing strong reciprocal support that the determinants of SI are indeed located in this region. Interestingly, we observed a series of shorter scaffolds that could not previously be anchored in the main assembly of the olive tree genome but match genetic markers that are strictly linked to SI in *P. angustifolia.* These unanchored scaffolds provide a more complete set of genomic sequences that will be important to consider in the perspective of identifying the (currently elusive) molecular determinants of SI in these two species. We note that poor assembly of the S-locus region (MARIOTTI *et al.* 2020) was expected given the considerable levels of structural rearrangements typically observed in SI- and more generally in the mating type-determining regions (GOUBET *et al.* 2012; BADOUIN *et al.* 2015), making *P. angustifolia* a useful resource to map the SI locus in the economically important species *O. europeae*.

Our observations also provide direct support to the hypothesis that the determinants of SI have remained at the same genomic position at least since the two lineages diverged, 30 to 40 Myrs ago (BESNARD *et al.* 2009; OLOFSSON *et al.* 2019). Stability of the genomic location of SI genes has been observed in some Brassicaceae species, where the *SRK-SCR* system maps at orthologous positions in the Arabidopsis and Capsella genuses (GUO *et al.* 2011). In other Brassicaceae species, however, the SI system is found at different genomic locations, such as in Brassica and Leavenworthia. In the former, the molecular determinants have remained the same (also a series of *SRK-SCR* pairs, (IWANO *et al.* 2014), but in the latter SI seems to have evolved d*e novo* from exaptation of a pair of paralogous genes (CHANTHA *et al.* 2013; CHANTHA *et al.* 2017). Together with the fact that *P. angustifolia* pollen is able to trigger a robust SI response on *O. europaeae* stigmas (SAUMITOU-LAPRADE *et al.* 2017), our results provide strong support to the

380 hypothesis that the *P. angustifolia* and *O. europaeae* SI systems are homologous. Whether mating type
381 determinants occupy orthologous genomic regions in different species and rely on the same molecular
382 players has also been discussed in oomycetes by Dussert *et al.* (2020).

383 Several approaches could now be used to refine the mapping of SI in *P. angustifolia,* and ultimately
384 zero in on its molecular determinants. One possibility would require fine-mapping using larger offspring
385 arrays, starting from our cross for which only a fraction of all phenotyped individuals were genotyped.
386 Beyond the analysis of this controlled cross, evaluating whether the association of the SI phenotype still
387 holds for markers within a larger set of accessions from diverse natural populations will constitute a
388 powerful fine-mapping approach. Since the SI phenotypes seem to be functionally homologous across the
389 Oleeae tribe (VERNET *et al.* 2016), the approach could, in principle, be extended to more distant SI species
390 of the family like *L. vulgare* or *F. ornus*. Identification of sequences that have remained linked over these
391 considerable time scales would represent excellent corroborative evidence to validate putative SI
392 candidates. In parallel, an RNA-sequencing approach could be used to identify transcripts specific to the
393 alternate SI phenotypes.

394 While comparison to the closely related *O. europeae* genome is a useful approach for the mapping
395 of SI in *P. angustifolia*, it is *a priori* of limited use for mapping the sex-determining region, since the olive
396 tree lineage has been entirely hermaphroditic for at least 32.22 Myrs (confidence interval: 28-36 Myrs)
397 (FigS1 in OLOFSSON *et al.* 2019). Detailed exploration of the genomic region in the olive tree that is
398 orthologous to the markers associated with sexual morphs in *P. angustifolia* is however interesting, as it
399 may either have anciently played a role in sex determination and subsequently lost it, or alternatively it
400 may contain quiescent sex-determining genes that have been activated specifically in *P. angustifolia*. At a
401 broader scale, mapping and eventually characterizing the sex locus in other androdioecious species such
402 as *F. ornus* could indicate whether the different instances of androdioecy in the family represent
403 homologous phenotypes or independent evolutionary emergences.

Identifying the molecular mechanisms of the genes controlling SI and sex and tracing their evolution in a phylogenetic context would prove extremely useful. First, it could help understand the strong functional pleiotropy between sex and SI phenotypes, whereby males express universal SI compatibility (SAUMITOU-LAPRADE *et al.* 2010). In other words, males are able to transmit the SI specificities they inherited from their parents, but they do not express them themselves even though their pollen is fully functional. This intriguing feature of the SI system was key to solve the puzzle of why *P. angustifolia* maintains unusually high frequencies of males in natural populations (HUSSE *et al.* 2013), but the question of how being a male prevents expression of the SI phenotype in pollen is still open. A possibility is that the *M* allele of the sex locus contains a gene interacting negatively either with the pollen SI determinant itself or with a gene of the downstream response cascade. Identifying the molecular basis of this epistasis will be an interesting next step. Second, another intriguing feature of the system is segregation distortion, which is observed at several levels. Billiard *et al.* (2015) observed complete segregation bias in favor of males among the offspring of H_b hermaphrodites sired by males. Here, by phenotyping >1,000 offspring of a H_a hermaphrodite sired by a M_b male, we confirmed that this cross also entails a departure from Mendelian segregation, this time in favor of hermaphrodites, albeit of a lesser magnitude. Although the generality of this observation still remains to be determined by careful examination of the other possible crosses (H_a hermaphrodites x M_a and M_c males), it is clear that segregation distortion is a general feature of this system, as was already observed in other sex determination systems causing departures from equal sex ratios (e.g. KOZIELSKA *et al.* 2010). Beyond identification of the mechanisms by which the distortions arise, pinpointing the evolutionary conditions leading to their emergence will be key to understanding the role they may have played in the evolution of this reproductive system.

More broadly, while sex and mating types are confounded in many species across the tree of life and cannot be distinguished, the question of when and how sex and mating types evolve separately raises several questions. The evolution of anisogamy (and hence, sexual differentiation) has been linked to that

428 of mating types (CHARLESWORTH 1978). In volvocine algae for instance, the mating-type locus in isogamous
429 species is orthologous to the pair of U/V sex chromosomes in anisogamous/oogamous species, suggesting
430 that the sex-determination system derives from the mating-type determination system (GENG *et al.* 2014).
431 From this perspective the Oleaceae family is an interesting model system, where a SI system is ancestral,
432 and in which some species have evolved sexual specialization that is aligned with the two SI phenotypes
433 (e.g. in the polygamous *F. excelsior* males belong to the H_a SI group and can only mate with hermaphrodites
434 or females of the H_b group, and the sexual system of *F. excelsior* can be viewed as subdioecy (SAUMITOU-
435 LAPRADE *et al.* 2018). In other species, sexual phenotypes are disjoint from SI specificities and led to the
436 differentiation of males and hermaphrodites. For instance, in the androdiecious *P. angustifolia* and
437 probably *F. ornus*, the male determinant is genetically independent from the SI locus but fully linked to a
438 genetic determinant causing the epistatic effect over SI (BILLIARD *et al.* 2015; VERNET *et al.* 2016). Yet other
439 species have remained perfect hermaphrodites and have no trace of sexual differentiation whatsoever (*O.*
440 *europeae*). Understanding why some species have followed one evolutionary trajectory while others have
441 followed another will be an exciting avenue for future research (BILLIARD *et al.* 2011).
442

443 ## Author Contributions

444 All authors contributed to the study presented in this paper. PS-L and PV developed, designed and oversaw
445 the study; they coordinated the cross and carried out the phenotyping and stigma tests. CG performed the
446 seedling paternity analysis. SS performed DNA extraction, library preparation and organized sequencing.
447 AC and SG constructed the data analysis pipeline and AC, SS, PS-L and VC interpreted the results and wrote
448 the manuscript.
449

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Table 1. Comparison of the sex-averaged, male and female linkage maps. The values in this table are computed without the outliers SNP markers at the extremity of the linkage groups.

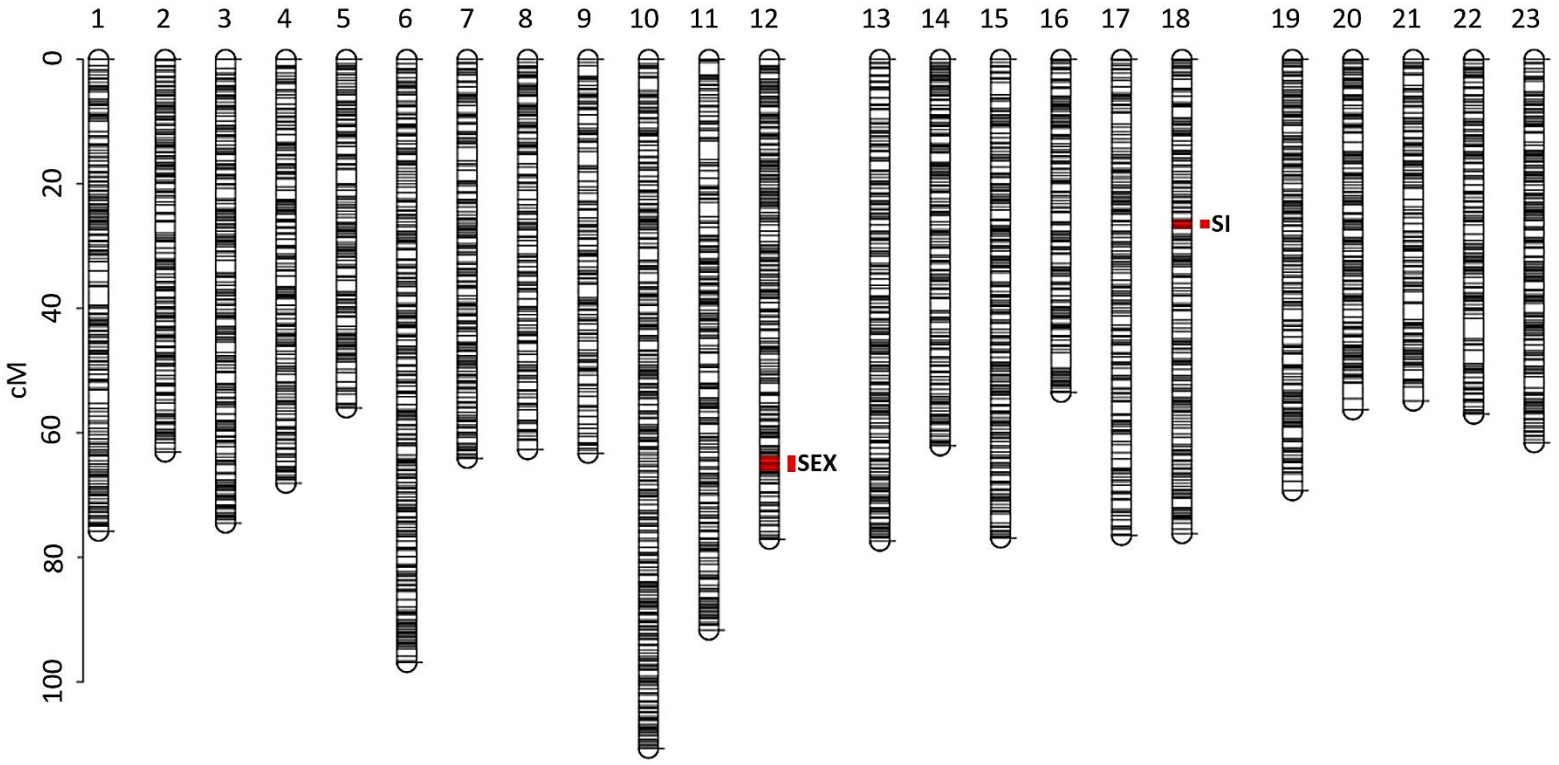
			Sex-averaged map			Paternal map			Maternal map		
Linkage group	**Number of SNPs**	**Number of SNPs (without outliers)**	**LG length (cM)**	**SNPs/cM**	**average intermarker distance**	**LG Length (cM)**	**SNPs/cM**	**average intermarker distance**	**LG Length (cM)**	**SNPs/cM**	**average intermarker distance**
1	854	839	75.78	11.07	0.09	79.30	10.58	0.09	92.11	9.11	0.11
2	633	621	23.90	25.98	0.10	35.00	17.74	0.12	22.73	27.32	0.11
3	676	676	74.50	9.07	0.11	61.94	10.91	0.09	85.42	7.91	0.13
4	535	535	68.07	7.86	0.13	71.63	7.47	0.13	69.82	7.66	0.13
5	502	494	56.02	8.82	0.11	50.58	9.77	0.10	67.84	7.28	0.14
6	877	877	96.89	9.05	0.11	90.81	9.66	0.10	103.63	8.46	0.12
7	609	601	64.14	9.37	0.11	68.93	8.72	0.11	64.99	9.25	0.11
8	486	479	62.71	7.64	0.13	91.89	5.21	0.19	119.25	4.02	0.25
9	408	406	63.28	6.42	0.16	56.06	7.24	0.14	71.04	5.72	0.18
10	1365	1361	110.69	12.30	0.08	121.95	11.16	0.09	112.38	12.11	0.08
11	793	783	91.66	8.54	0.12	80.40	9.74	0.10	108.84	7.19	0.14
12	973	969	77.12	12.56	0.08	91.52	10.59	0.09	88.09	11.00	0.09
13	849	848	77.40	10.96	0.09	77.29	10.97	0.09	80.77	10.50	0.10
14	566	565	62.12	9.10	0.11	72.25	7.82	0.13	71.39	7.91	0.13
15	750	747	76.92	9.71	0.10	82.98	9.00	0.11	96.01	7.78	0.13
16	591	589	53.53	11.00	0.09	56.93	10.35	0.10	69.56	8.47	0.12
17	613	613	76.52	8.01	0.13	70.58	8.69	0.12	83.94	7.30	0.14
18	660	659	76.22	8.65	0.12	77.26	8.53	0.12	81.99	8.04	0.12
19	806	806	69.29	11.63	0.09	91.10	8.85	0.11	79.49	10.14	0.10
20	547	531	56.26	9.44	0.11	67.56	7.86	0.13	62.91	8.44	0.12
21	479	476	54.86	8.68	0.12	69.49	6.85	0.15	54.14	8.79	0.11
22	550	544	57.05	9.54	0.11	55.64	9.78	0.10	61.44	8.85	0.11
23	690	684	61.64	11.10	0.09	67.10	10.19	0.10	66.42	10.30	0.10
average	687	682	68.98	10.28	0.11	73.40	9.46	0.11	78.88	9.29	0.12

621 **Figure 1.** *Phillyrea angustifolia* sex-averaged linkage map showing the grouping and position of 15703

622 SNPs. The length of each of the 23 linkage groups is indicated by the vertical scale in cM. The markers

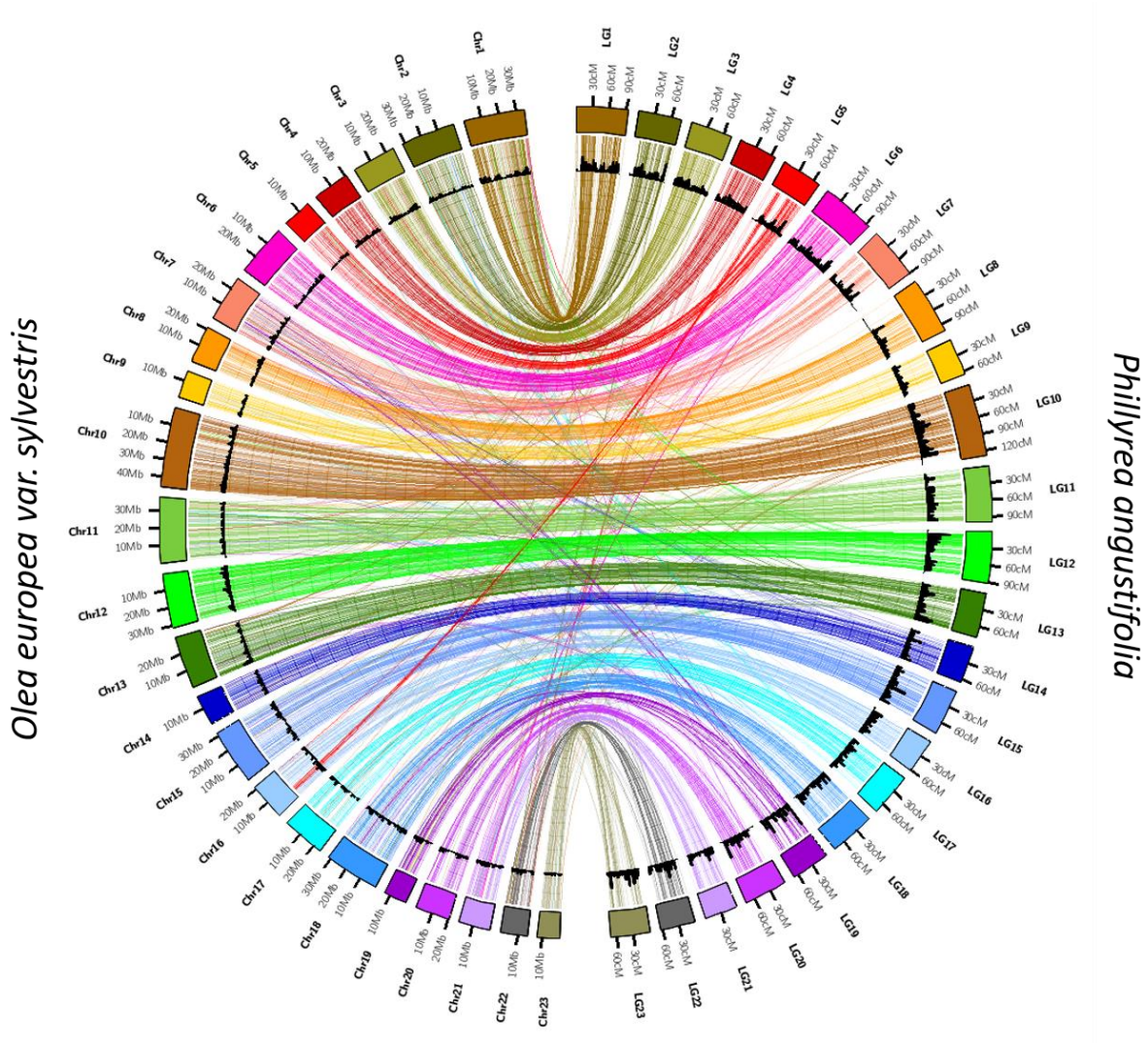
623 strictly linked to sex and self-incompatibility (SI) phenotypes are shown in red. Markers that were clearly

624 outliers at the end of some linkage groups were removed (see Table1, Figure S1).



625 **Figure 2.** Synteny plot identifying homologous *P. angustifolia* linkage groups (LG, scale in cM) with olive
626 tree chromosomes (Chr, scale in Mb). Lines connect markers in the *P. angustifolia* linkage map with their
627 best BLAST hit in the *O. europea* genome and are colored according to the linkage group. Variation of the
628 density of loci in bins of 3.125cM along linkage groups and 1 Mbp along chromosomes is shown in the
629 inner circle as a black histogram.

630



631 **Figure 3.** Visualization of chromosome-scale synteny by comparing the location of markers along the *P.*
632 *angustifolia* linkage groups (LG, scale in cM) with the location of their best BLAST hit along the homologous
633 olive tree chromosome (Chr, scale in Mbp). The vertical lines on LG12 and LG18 indicate the position of
634 markers strictly associated with sex and SI phenotypes in *P. angustifolia*, respectively. The horizontal line
635 on Chr18 indicates the chromosomal region containing the SI locus in *Olea europaea* according to Mariotti
636 *et al.* (2020).

637

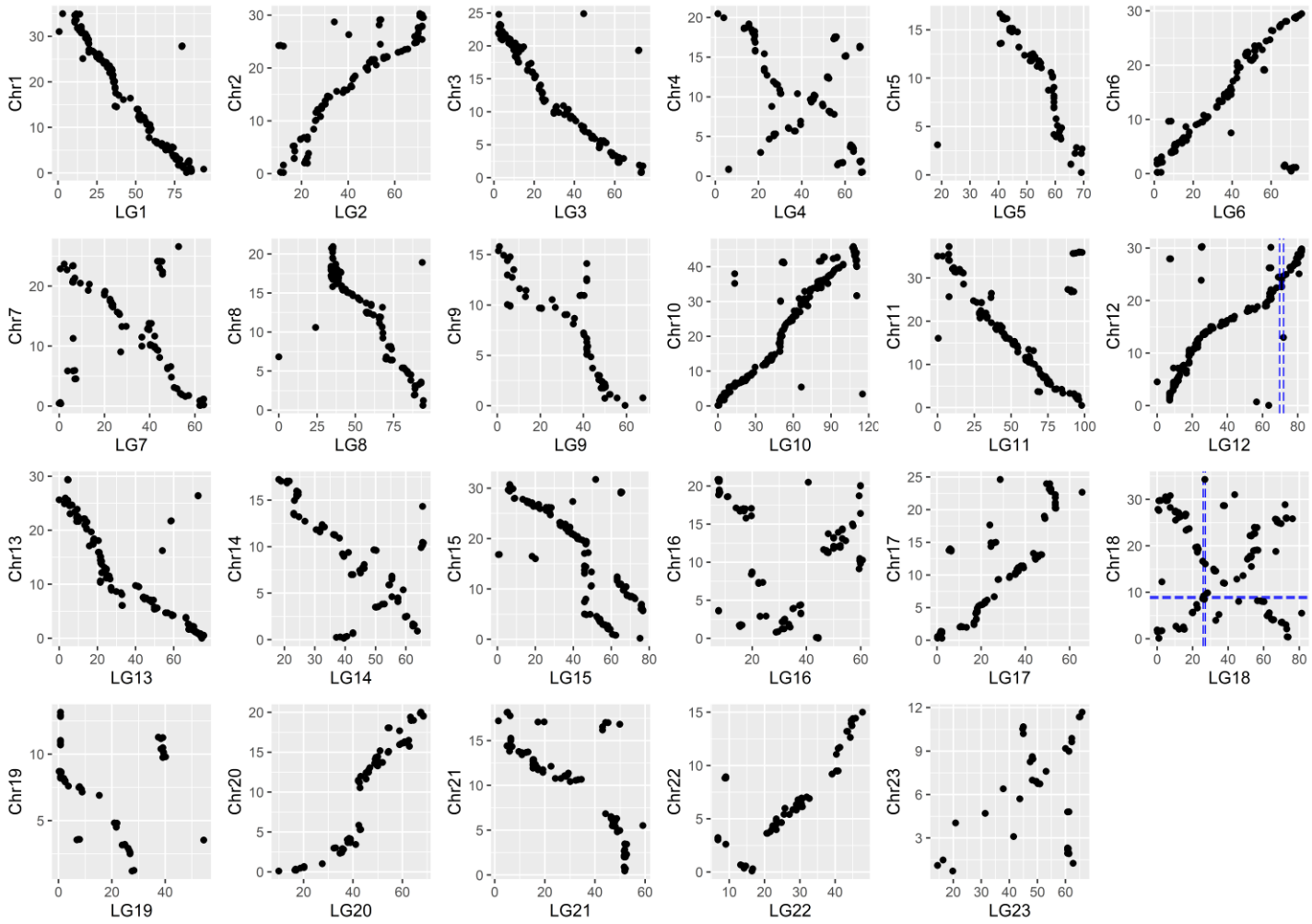


Figure 4. Synteny plot between the *P. angustifolia* linkage group 12 (scale in cM) and the olive tree chromosomes 12 and a series of unanchored scaffolds (scale in Mb). Lines connect markers in the *P. angustifolia* linkage map with their best BLAST hit in the *O. europea* genome. Green lines correspond to markers with autosomal inheritance. Black lines correspond to markers which strictly cosegregate with sex phenotypes (males *vs.* hermaphrodites). Red lines correspond to markers with strong but partial (95%) association with sex. Variation of the density of loci in bins of 3.125cM along linkage groups and 1 Mbp along chromosomes is shown in the inner circle as a black histogram.

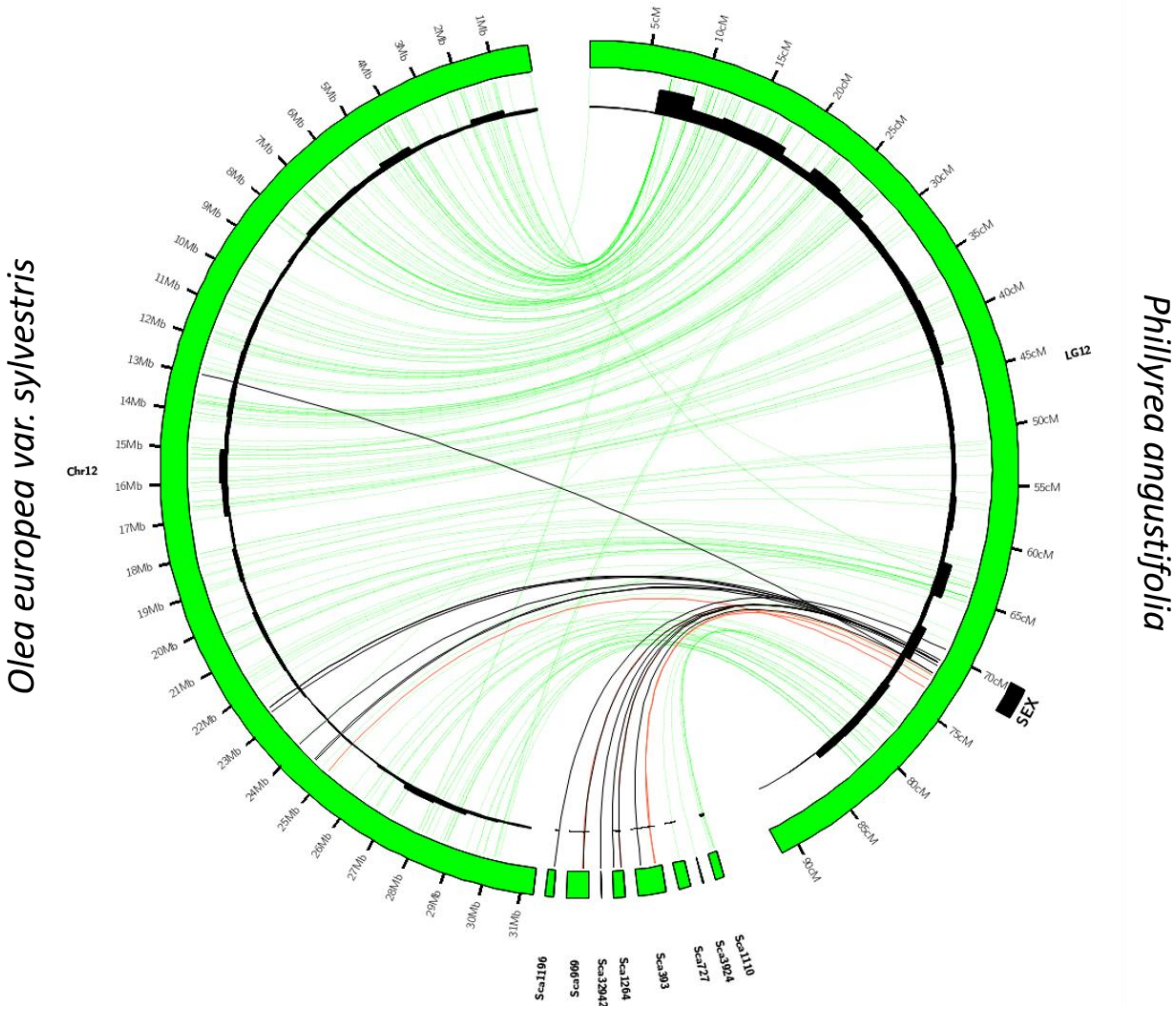
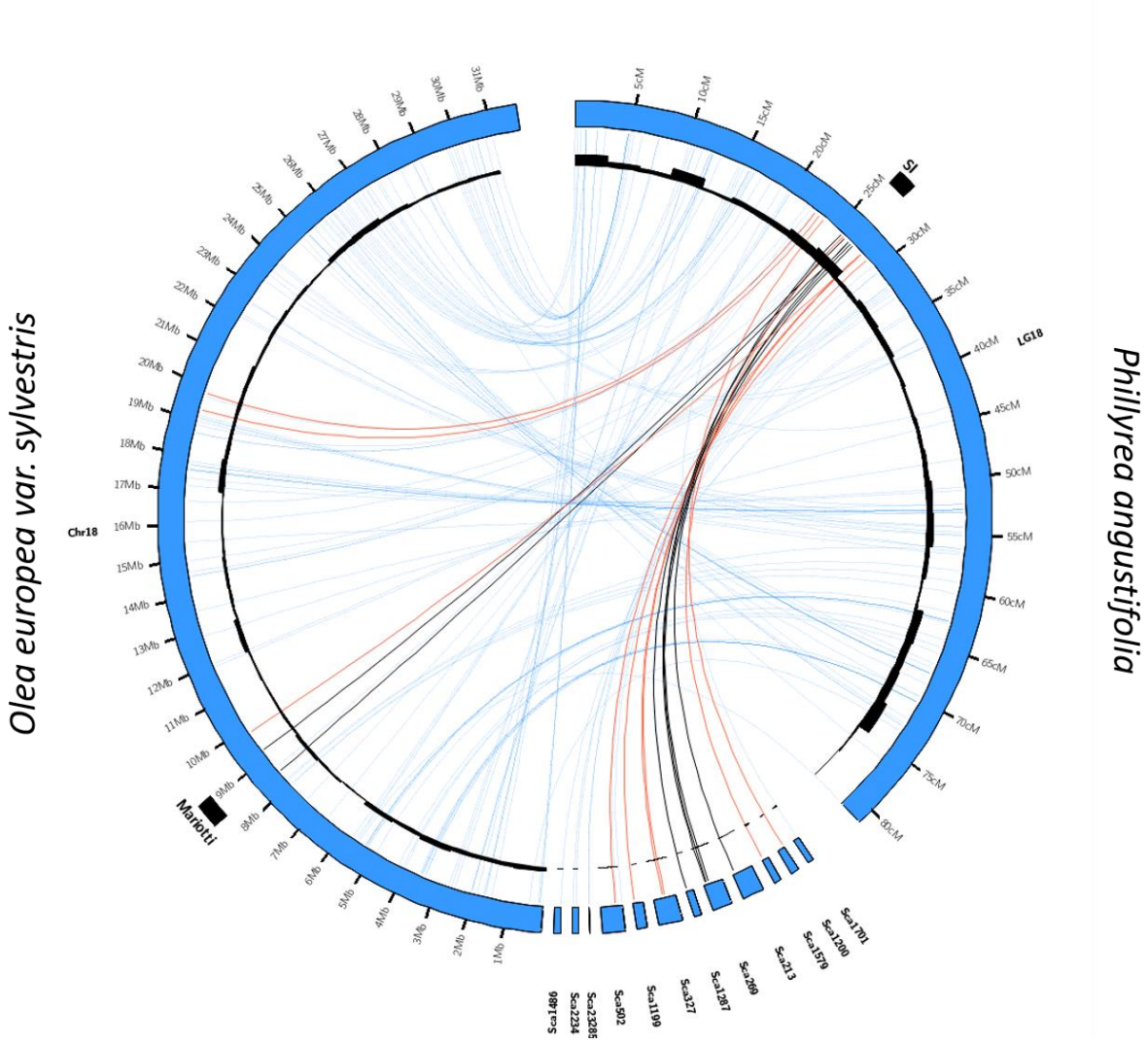



Figure 5. Synteny plot between the *P. angustifolia* linkage group 18 (scale in cM) and the olive tree chromosomes 18 and a series of unanchored scaffolds (scale in Mb). Lines connect markers in the *P. angustifolia* linkage map with their best BLAST hit in the *O. europea* genome. Blue lines correspond to markers with autosomal inheritance. Black lines correspond to markers which strictly cosegregate with SI phenotypes (H_a *vs.* Hb). Red lines correspond to markers with strong but partial (95%) association with SI. The region found to be genetically associated with SI in the olive tree by Mariotti *et al.* (2020) is shown by a black rectangle. Variation of the density of loci in bins of 3.125cM along linkage groups and 1 Mbp along chromosomes is shown in the inner circle as a black histogram.