<u>**Title**</u>

The dopamine circuit as a reward-taxis navigation system

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<u>**Abstract**</u>

Research on certain circuits in simple organisms, such as bacterial chemotaxis, has enabled the
formulation of mathematical design principles, leading to ever more precise experimental tests,
catalyzing quantitative understanding. It would be important to map these principles to the far
more complex case of a vertebrate behavioral circuit. Here, we provide such a mapping for the
midbrain dopamine system. Dopamine transmission plays a key role in learning, motivation,
and movement, but its systems-level function is not fully understood. We develop a minimal
mechanistic model of the dopamine circuit based on physiological and behavioral data, and
show that it can be mapped mathematically to the bacterial chemotaxis circuit. Just as
chemotaxis robustly climbs attractant gradients, the dopamine circuit performs 'reward-taxis'
where the attractant is the expected value of reward. The reward-taxis mechanism is based on
a circuit feature called fold-change detection, where the circuit outputs the temporal
logarithmic derivative of expected reward. The model can explain the general matching law,
in which the ratio of responses to concurrent rewards goes as the reward ratio to the power β.
It provides an accurate mechanistic value for β as the average gain/baseline ratio of the
dopaminergic neurons. Reward-taxis provides testable etiologies for specific dopamine-related
disorders.

Introduction

Foundational work in the field of systems biology showed how, once a biological circuit is well characterized in terms of components and behavior, general mathematical principles can be formed which not only shed light on the original circuit but can generalize to explain many other circuits. It is important to explore whether these principles apply also to a vertebrate behavioral circuit. To do so, it is ideal to begin with a well-characterized circuit in terms of neuronal dynamics, physiology, and behavior. A good candidate for such a circuit is the midbrain dopamine circuit.

Dopamine transmission in the midbrain participates in at least two major biological functions. The first role is the control of movement and motivation (Niv *et al*, 2007; Mazzoni *et al*, 2007; Dudman & Krakauer, 2016; da Silva *et al*, 2018; Shadmehr & Ahmed, 2020). Dopamine transmission regulates movement initiation and vigor on a sub-second level as shown in freely behaving mice (da Silva *et al*, 2018), and defects in dopamine transmission underlie the movement difficulties in Parkinson's disease (Meder *et al*, 2019).

The second role of dopamine is encoding *reward prediction errors* (Schultz *et al*, 1997). Reward prediction errors are the difference between the experienced and predicted rewards. They play a key role in a specific method of reinforcement learning called temporal difference learning (TD learning) (Schultz *et al*, 1997; Sutton & Barto, 1998; Glimcher, 2011; Steinberg *et al*, 2013). In addition to rapid responses encoding reward prediction errors, dopamine also slowly ramps up when approaching a reward (Howe *et al*, 2013; Hamid *et al*, 2016; Mohebi *et al*, 2019). Importantly for the present study, a recent experiment demonstrated that dopaminergic ramps represent a temporal derivative-like computation on expected reward (Kim *et al*, 2020). Kim et al. used virtual reality to manipulate the rate of change of expected reward as mice moved towards a target, and found that mice responded to the temporal derivative of expected reward, rather than to its absolute value.

To better understand the dopamine circuit on the system level, it is important to unify, in a single conceptual framework, both its reward-derivative and its motion control functions (Berke, 2018; Shadmehr & Ahmed, 2020). Why does dopamine play two important roles at the same time: invigorate movements and encode the derivative of expected reward? Theoretical studies have analyzed this question from the perspectives of learning (Friston, 2010; Bogacz, 2020) and cost-benefit theories (Yoon *et al*, 2018; Shadmehr & Ahmed, 2020). Here, we consider the function of the circuit from the point of view of navigation, a connection anticipated by early work on TD learning (Montague *et al*, 1995).

For this purpose, we develop a mechanistic model of the dopamine circuit, and show that this

circuit can be mathematically mapped to the navigational circuit that guides bacterial

chemotaxis, a foundational circuit in systems biology. We show that experimental observations

on dopaminergic regulation can be explained by a circuit property known as fold-change

detection *(FCD)*, in which the output of the circuit responds to *relative* rather than absolute

changes in the input (Shoval *et al*, 2010; Adler & Alon, 2018). FCD was discovered in bacterial

chemotaxis and was then found to be widespread in sensory and navigational circuits (Adler &

Alon, 2018). In the case of dopamine, the input to the circuit is expected reward. Additionally,

just as chemotaxis controls bacterial runs and tumbles, we build on measurements of animal

movement to propose a model where high dopamine results in prolonged bouts of vigorous and

correlated movements ("runs") whereas low dopamine causes pauses and slow reorientations

("tumbles"). This model for dopaminergic regulation of movement indicates how the two roles

of dopamine, reward-derivative and motion-regulator, combine to provide a robust navigation

system that climbs reward gradients. We call this the 'reward-taxis' model for the dopamine

system. The model provides a mechanistic explanation for the well-established generalized

matching law of behavior, predicts search dynamics that are invariant to the scale of the

rewards, and provides testable etiologies for specific dopamine-related disorders.

Results

Dopamine release as fold-change detection of expected reward

We begin by developing a minimal model for midbrain dopamine regulation by environmental

cues (Figure 1A). Consider a behaving animal exploring an open field for a reward of

magnitude *u*, such as a food or drink. For simplicity, we assume a uniform response amongst

all dopaminergic neurons. In reality, there are important heterogeneities between and within

midbrain structures, and some dopaminergic neurons may be specialized to specific cues (Eshel

et al, 2016; Parker *et al*, 2016; da Silva *et al*, 2018; Lee *et al*, 2019; Engelhard *et al*, 2019). If

the animal is familiar with the environment, dopamine release *d* will increase as the animal

observes cues that indicate closer proximity to the reward (Howe *et al*, 2013; Hamid *et al*,

2016; Kim *et al*, 2020). This is due to the increase in expected reward *R*, which decays with

distance from the target (Howe *et al*, 2013; Gershman, 2014; Kim *et al*, 2020). Here expected

reward is defined based on TD learning: the expected temporally discounted sum of present

and future rewards (see Methods).

How does dopamine release, *d,* depend on expected reward, *R*? Recordings from VTA

dopaminergic neurons in mice reveal a strongly sublinear relationship between the magnitude

of received rewards u and the dopaminergic response above its baseline, $\Delta d(u)$ (Eshel *et al*,

2015, 2016) (Figure 1B). The strong sublinear relationship indicates that dopamine neurons

may be (at least in some magnitude range) responding to the logarithm of expected reward, i.e.,

that: $\Delta d(u) = \mu \log(R(u))$, where $R(u) = au + b$ is the expected reward, where a is a scaling

factor and b is a magnitude-independent component of the expected reward. The parameter μ

is the gain of the dopaminergic response. Logarithmic responses are consistent with widespread

logarithmic coding in the brain (Dehaene, 2003; Nieder & Miller, 2003; Shen, 2003; Dehaene

et al, 2008; Nieder & Dehaene, 2009; Laughlin, 1989) as well as with economic notions of

utility (Bernoulli, 1968; Rubinstein, 1977).

To test this, we fit the function $\mu \log(au + b)$ to the average dopaminergic response to a

variable water reward in mice (Eshel *et al*, 2016), finding an excellent fit ($r^2 = 0.999$), with a

gain of $\mu = 4.94 \pm 0.45$.

In addition to activation by expected reward, dopaminergic neurons in the VTA are inhibited

by the output g of adjacent GABAergic neurons, and this inhibition is subtractive (Eshel *et al*,

2015, 2016). We therefore propose the following minimal description of dopamine release:

$$d = C + \mu \log R - \alpha g \quad [1]$$

where C is the baseline activity of the dopaminergic neurons when $\log R = g = 0$, μ is the

gain, R is perceived expected reward, and α is the effectiveness of inhibition by the GABAergic

output g. Since our model focuses on the regulation of behavior, rather than on learning or

representation, we will assume that the log-transformed expected reward $\log R$ is an input

signal that is provided to the circuit. Additionally, while subtractive inhibition was established

for VTA dopaminergic neurons, we assume that similar regulation is shared among all

midbrain dopaminergic neurons.

To complete the model requires a minimal description of the dynamics of GABAergic output

g. The mechanisms of interaction between GABAergic and dopaminergic neurons are

complex, and there are many local and remote interactions (Morales & Margolis, 2017; Cox &

Witten, 2019). However, there are several experimental observations that impose constraints

on these interactions (Figure 1CDE). First, upon presentation of a reward-predicting cue -

equivalent to a step increase in *R(t)* - dopamine rapidly increases and then drops and adapts

precisely to its baseline on a sub-second timescale (Schultz *et al*, 1997; Tobler *et al*, 2005;

Cohen *et al*, 2012) (Figure 1C). Second, dopamine responses are scaled relative to the expected

reward baseline: when the expected reward is of magnitude u and the received reward is of

magnitude λu for a constant λ, dopamine responses are nearly identical and independent of u

(Tobler *et al*, 2005) (Figure 1D). Third, dopamine release represents a temporal-derivative-like

computation of *R(t)* (Kim *et al*, 2020) (Figure 1E). Finally, GABAergic activity tracks *R(t)*

(Cohen *et al*, 2012) (Figure 1C).

These experimental observations are the hallmarks of a circuit feature known in systems

biology as *fold-change detection (FCD)* (Shoval *et al*, 2010; Adler & Alon, 2018) (Figure 1A).

FCD is a property where the circuit output depends only on relative changes in the input, rather

than absolute changes. FCD circuits output the temporal (logarithmic) derivative of low-

frequency input signals, whereas high frequency inputs are normalized by their baseline (Tu *et*

al, 2008; Adler *et al*, 2014; Lang & Sontag, 2016).

FCD circuits are ubiquitous in biological sensing, as reviewed in (Adler & Alon, 2018). They

can be implemented by a handful of specific feed-forward and feedback mechanisms (Adler *et*

al, 2017). In all FCD mechanisms, a slow internal variable (*g*) encodes the background input

level and acts to normalize the output accordingly. Taken together, the FCD properties can

explain all of the experimental observations on dopamine responses described in Figure 1.

Since we do not know the exact mechanistic implementation of the FCD property in the

dopamine circuit, we will make the simple assumption of a negative feedback loop, and choose

the simplest design that provides FCD. In this design, inhibitory neuron activity g is given by

an integral-feedback equation with respect to dopamine release d:

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$$\dot{g} = \omega \left(\frac{d}{d_0} - 1 \right) \quad [2]$$

Or, more generally, $\dot{g} = F(d)$ where $F(d)$ increases with d and has a single zero at $d = d_0$.

This feedback loop generates exact adaptation of d: the only steady state solution is $d = d_0$.

d_0 is the homeostatic activity level of dopaminergic neurons, which is about ~5 spikes/s in

mice (Robinson *et al*, 2004; Eshel *et al*, 2016). The parameter ω determines the adaptation

time of the dopaminergic neurons after a change in *R*(t). This timescale is on the order of

hundreds of milliseconds. For the GABAergic neurons, after a step change in *R(t)*, the firing

rate in the model increases proportionally to the logarithm of *R(t)*, such that $g = \frac{C - d_0 + \mu \log R}{\alpha}$

(this is because GABAergic output integrates dopaminergic activity). Taken together, Eqs. 1-

2 provide a minimal model for dopamine responses to expected reward inputs *R(t)*.

Figure 1. Dopamine responses as fold-change detection of expected reward. (A) Minimal circuit for dopamine responses. Dopamine *(d)* is activated by the logarithm of expected reward ($\log R$) and is inhibited by negative feedback from GABAergic input (*g*). See *Methods* for equations and parameters. (B) The response of VTA dopaminergic neurons to a water reward of variable volume (black squares, mean ± SEM, taken from Figure 1C in (Eshel *et al*, 2015)) is well-described by the logarithmic relation $\Delta d = \mu \log(aR + b)$ ($r^2 = 0.999$, best-fit parameters $a = 0.5, b = 1.5, \mu = 4.9$). (C) Simulation of dopamine and GABA responses to a step increase in expected reward input which corresponds to the presentation of a reward-predicting cue. The step is given by $R(t) = R_0 + \lambda\theta(t - t_0)$ where $\theta(t - t_0)$ is a unit step function, $R_0 = 1$ and $\lambda = 7$ for the black line (large reward) and $\lambda = 1.8$ for the gray line (small reward). *Insets*. Average change in firing rates from dopaminergic (type I) and GABAergic (type II) VTA neurons, in response to reward-predicting cues for a small reward (gray) or a large reward (black). Data from Figure 2D of (Cohen *et al*, 2012). (D) Dopamine responses in Macaque monkeys to variable size liquid reward, either without a preceding cue (left panels), or with a preceding visual cue that predicts reward delivery with 50% probability. *Upper*. The expected-reward input following the reward-predicting cue is $R = 0.5(b + \lambda u)$ (where λ is proportional to reward magnitude), which is doubled following reward delivery, $R = b + \lambda u$, where u is the reward magnitude (we use $b = 2$ and $\lambda = 10ml^{-1}$). *Middle*. Experimentally measured average dopaminergic responses, adapted from Fig. 2A and Fig. 4B of (Tobler *et al*, 2005). When the reward is delivered without a cue, dopaminergic responses increase with reward magnitude (left panel). When it is given after a cue that predicts reward delivery with 50% probability, dopaminergic responses to reward delivery are identical (right panel), as predicted by the FCD property. This is despite the 10-fold difference in reward magnitude. *Lower*. Simulations of the dopamine model capture the experimentally observed dynamics. (E) Dopamine output to movement in a reward gradient given by $R(x) = e^{-\gamma x^h}$, with $h = 1.5, x_0 = 1, v_0 = 1$ and $\gamma = 0.04$, and perturbations as described

173 in Figure 2 of (Kim *et al*, 2020). *Insets.* Corresponding dopaminergic outputs from VTA neurons (measured by calcium imaging) from Figure
174 2CGKO in (Kim *et al*, 2020), smoothed using a Savitzky–Golay filter. All simulations were performed with parameters provided in Table 1.

A reward-taxis model for dopamine regulation of behavior

To link the dopamine circuit to animal behavior, we provide an additional equation as a
minimal model for dopamine control of motion. The equation is motivated by the well-
established role of dopamine in the regulation of action vigor and locomotor activity (Niv *et*
al, 2007; Beierholm *et al*, 2013; Panigrahi *et al*, 2015; Ek *et al*, 2016; da Silva *et al*, 2018).
We focus on stochastic exploratory motion, as opposed to directed and deliberate motion along
a specific path. We therefore model movement as a random-walk process, which is a standard
approach for modelling animal movement (Blackwell, 1997; Codling *et al*, 2008; Smouse *et*
al, 2010; Mwaffo *et al*, 2015; Hanks *et al*, 2015; Michelot *et al*, 2019). While the model is
presented in the context of physical spatial motion of an animal, it can be readily generalized
to other aspects of behavior, as we will detail below.
We assume that the navigating animal can be in one of two states – arrested or moving. We
also assume that when the animal is moving, it runs in a straight line, and when the animal is
arrested, it chooses its subsequent movement direction at random (allowing the new direction
to be correlated with the previous direction does not affect the conclusions). Such a model can
capture, for example, the swimming behavior of zebrafish (Budick & O'Malley, 2000; Mwaffo
et al, 2015), or the movement of mice in an open field (da Silva *et al*, 2018). The random walk
has two parameters: movement speed v (units of length/time), and run duration, which is
exponentially distributed with mean ϕ (units of time).
Dopamine increases movement vigor, which may be due to increasing v or increasing ϕ. In
zebrafish, the dopamine agonist apomorphine increased the speed of swimming bouts at low
doses (higher v), and the duration of swimming bouts (higher ϕ) at high doses (Ek *et al*, 2016).
In a recent study, da Silva et al. performed sub-second measurements of SNc dopaminergic
neurons in freely behaving mice in an open field (da Silva *et al*, 2018). The neuronal
measurements were coupled with high-resolution motion measurements. Dopamine activity
prior to movement increased the probability and vigor of subsequent movement (da Silva *et al*,
2018). Both these effects may again manifest in higher movement speed v, or in correlated
movement patterns that correspond to higher ϕ.
Based on these observations, we propose two possibilities for dopamine regulation of
movement. In the first possibility, dopamine increases run duration:

$$\phi(d) = \tau \frac{d}{d_0} \quad [3]$$

Where τ is the average run duration in an environment without reward cues. Dopamine d in

Eq. 2 corresponds to the (normalized) dopaminergic activity in the brain, so loss of

dopaminergic neurons (as occurs in Parkinson's disease) reduces d proportionately, which

effectively reduces run duration τ. We assume a linear relation between run duration and d,

which is consistent with the gradual increase in movement vigor with dopaminergic activity

(see Fig. 1j-k in da Silva et al. (da Silva *et al*, 2018)). The important aspect is that high

dopamine results in directed motion, whereas low dopamine decorrelates motion.

We can also consider an alternative model where dopamine increases running speed:

$$v(d) = v_0 \frac{d}{d_0} \quad [4]$$

In this case, loss of dopamine neurons effectively decreases the average running speed v_0. Such

a speed-control model provides the same conclusions as a run-duration-control model with Eq.

3 (Methods). However, in order to emphasize the analogy with chemotaxis, we use Eq. 3. Note

that in both cases, we focus on the sub-second regulation of movement parameters by

dopamine, as observed by da Silva et al. (da Silva *et al*, 2018).

Eqs. 1-3 define a biased random walk that is guided by the gradient of the logarithm of the

expected reward (Figure 2B), which we call *reward-taxis*.

This biased random walk is analogous to bacterial chemotaxis: bacteria such as *E. coli* use a

run-and-tumble navigation strategy to climb gradients of chemical attractants (Figure 2CDE)

(Berg & Brown, 1972; Tu *et al*, 2008; Shoval *et al*, 2010; Sourjik & Wingreen, 2012). The

chemotaxis signaling network is based on an FCD circuit that controls run duration (Shoval *et*

al, 2010; Lazova *et al*, 2011). It therefore maps onto dopamine release dynamics, where

expected-reward inputs play the role of chemoattractant concentration in chemotaxis. Both

systems respond to relative input levels. They both respond to the derivative of low-frequency

input components and respond proportionally to high-frequency input components (Tu *et al*,

2008), and show exact adaptation to step changes in input. Thus, the dopamine circuit can allow

animals to climb gradients of reward expectation.

The reward-taxis model was presented for whole-body spatial movement, but its assumptions

are general and may potentially extend to other aspects of behavior. One such aspect is

hippocampal replay, the activation of hippocampal place cells during sharp-wave ripples

(Gomperts *et al*, 2015; Ólafsdóttir *et al*, 2018; Stella *et al*, 2019). Hippocampal replay consists

of a firing sequence of neurons that represents temporally compressed motion trajectories (Lee

& Wilson, 2002; Pfeiffer & Foster, 2013; Ólafsdóttir *et al*, 2018). It can occur either during

sleep or rest ("offline") or when the animal is awake and engaged in a task ("online"). Online

replay plays an important role in planning and navigation (Pfeiffer & Foster, 2013). The

activation of the place neurons corresponds to stochastic movement trajectories with a

characteristic speed (Davidson *et al*, 2009) that are biased towards the location of rewards

(Pfeiffer & Foster, 2013); when foraging is random, the trajectories are diffusive, resembling

Brownian motion (Stella *et al*, 2019). Hippocampal activity during online replay is tightly

coordinated with reward-associated dopamine transmission (Gomperts *et al*, 2015). To map

reward taxis to hippocampal replay requires that dopamine transmission modulate the

stochastic trajectories of hippocampal replay, for example through the modulation of

reorientation frequency.

Figure 2. Dopamine regulation of behavior in the model is analogous to bacterial chemotaxis. (A) The behavior of an animal in an open
field is modelled as a series of directed movements (runs). The direction of each run is chosen at random (or, more generally, stops between
runs decorrelate motion direction), and the duration of each run increases with dopamine level. (B) Dopamine is controlled by an FCD circuit
activated by expected reward. (CD) The reward-dopamine-behavior circuit is analogous to the chemotaxis circuit that underlies bacterial
navigation towards chemo-attractants. Bacterial motion is composed of a series of runs. The direction of each run is randomized by tumbling
events, and run duration increases with the inactivation of a receptor-kinase complex, which is controlled by an FCD circuit activated by
chemoattractant concentration. (E) Table detailing the mapping between the dopamine system and the chemotaxis system.

Reward-taxis quantitatively provides the matching law of operant behavior

We next ask how the reward-taxis model compares with features of animal behavior. We find

that the model provides the *general matching law of operant behavior*, one of the best-

established behavioral phenomena (Herrnstein, 1970; Baum, 1974, 1981; Baum *et al*, 1999;

Lau & Glimcher, 2005; McDowell, 2013). If an animal has a choice of two rewards with

expected magnitude r_1, r_2 at two locations x_1, x_2, the long-term average of its response rates

(the relative amount of time it chooses each reward (Baum & Rachlin, 1969)) $P(x_1), P(x_2)$

goes as a power β of the ratio of rewards:

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$$\frac{P(x_1)}{P(x_2)} = \left(\frac{r_1}{r_2}\right)^\beta \quad [5]$$

The matching law holds both for varying reward magnitude and varying reward probability

(Davison & McCarthy, 1988). The matching law was originally proposed with perfect

matching $\beta = 1$ (Herrnstein, 1961). A large number of studies in various vertebrate species

under different experimental conditions observed that β can be somewhat variable, showing

slight undermatching ($\beta < 1$) and overmatching ($\beta > 1$), with the former more commonly

observed (Baum, 1974; William, 1979; Baum, 1981; Davison, 1991; Baum *et al*, 1999; Lau &

Glimcher, 2005; McDowell, 2013). Matching is a robust property which holds over orders of

magnitude of reward ratios (up to ~1:500 in pigeons (Baum *et al*, 1999)). The overall

robustness of the matching law has led authors to suggest that it reflects intrinsic properties of

the vertebrate nervous system (McDowell, 2013).

While previous explanations for the general matching law focused on underlying choice

processes (Herrnstein & Prelec, 1991; Soltani & Wang, 2006; Simen & Cohen, 2009), here we

show that matching is a robust emergent property of the dopamine system, and that β is an

intrinsic property of dopamine neurons that can be directly estimated from neuronal recordings.

To test whether the model provides the matching law, we model the dynamics of the location

of a behaving animal as a stochastic process. Let $R(x)$ be the input field, which is the expected

reward R as a function of location x. At long time- and length-scales, run-and-tumble motion

resembles Brownian motion (Berg, 1993), with a diffusion coefficient $D \approx m^{-1} v^2 \tau$, where v

is the average run speed, τ is the average run duration, and m is the dimension (we assume that

d is close to the adapted level d_0). Brownian motion in the model is biased by longer runs

when the agent is moving up the gradient. To account for this, one can add to the diffusion

process an advection term that is proportional to the logarithmic gradient: $\chi \nabla \log R(x)$ (Si *et*

al, 2012; Dufour *et al*, 2014; Menolascina *et al*, 2017). Taken together, the stochastic dynamics

of the agent are approximated by a Langevin equation similar to the classic Keller-Segel

equation used to model chemotaxis (Keller & Segel, 1971):

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$$dx = \chi \nabla \log R(x)\, dt + \sqrt{2D}\, dW \quad [6]$$

where W is an m-dimensional Wiener process (see Supplementary Information 1 for the

derivation of Eq. 6). For relatively short runs compared with the adaptation time (sub-second

order), the advection parameter χ, also called chemotactic drift, is given by: $\chi \approx m^{-1}v^2\tau \cdot$

μd_0^{-1} (Si *et al*, 2012), and thus rises with velocity v, gain μ and mean run duration τ. It is

important to note that Eq. 6 holds also for the model variant in which dopamine modulates

movement speed (see Supplementary Information 2).

Eq. 6 captures how animal movement depends on the parameters of the dopamine circuit, as

well as on movement parameters τ, v. Decreasing the average run duration τ or average

movement speed v (as in Parkinson's disease) decreases both diffusivity D and advection χ,

resulting in slower effective motion and gradient climbing. Gradient climbing efficiency

(chemotactic drift) increases with μd_0^{-1}, which is the gain of dopamine neurons normalized by

their baseline activity. Other circuit parameters do not affect movement dynamics.

Eq. 6 is equivalent to the Langevin Monte Carlo algorithm, a common algorithm from

statistical physics for sampling probability distributions (Roberts & Tweedie, 1996; Neal,

2011; Girolami & Calderhead, 2011; Dalalyan, 2014). The steady-state distribution can be

readily solved, using standard methods of statistical physics, similar to a Boltzmann

distribution in a potential field. The motion samples a probability distribution *P(x)* proportional

to a power β of the expected-reward distribution:

$$P(x) \propto e^{\beta \log R(x)} = R(x)^\beta \quad [7]$$

where the power-law β equals the normalized gain of the dopaminergic neurons: $\beta = \frac{\chi}{D} =$

μd_0^{-1}. From Eq. 7 we infer that for any two rewards r_1, r_2 the response rates $P(x_1), P(x_2)$ obey

the general matching law presented in Eq. 5.

The reward-taxis model therefore predicts operant matching with $\beta = \mu d_0^{-1}$. Thus, β is equal

to the average ratio of gain to baseline activity in dopaminergic neurons. As mentioned above,

these values can be estimated from the neuronal measurements of Eshel et al. (Eshel *et al*,

2016), $\mu \approx 5$ spikes/s and $d_0 \approx 5$ spikes/s. These values yield $\beta \approx 1$, in agreement with the

matching law. Similar parameters are found also in primates (Supplementary Information 3).

The agreement is striking since there is no a-priori reason for the gain and baseline to be similar;

normalized gain μd_0^{-1} could in principle have a wide range of values including $\mu d_0^{-1} \ll 1$ or

$\mu d_0^{-1} \gg 1$.

The matching exponent β only depends on parameters that are intrinsic to the dopamine

neurons; β is independent of movement speed v or run duration τ, which may vary depending

on animal physiology and the environmental context, as well as on the number of dopaminergic

neurons. This may explain the robustness of the matching phenomena across species and

experimental conditions. Taken together, this suggests that the reward-taxis model may provide

a physiological mechanism underpinning operant matching.

Figure 3. Reward-taxis model provides the generalized matching law. (A) The generalized matching law, amply supported experimentally,

is a power-law relation between the frequency of rewards and the frequency of responses $\frac{P(x_1)}{P(x_2)} = \left(\frac{r_1}{r_2}\right)^{\beta}$, r_1, r_2 are rewards at locations x_1, x_2.

Perfect matching occurs for $\beta = 1$, while undermatching/overmatching are due to variations in β. Such a power-law relation arises directly

from the dynamics of the model where $\beta = \mu d_0^{-1}$. (B) Simulation of the dopamine model for choice between two rewards r_1, r_2. Expected

reward is the sum of two Gaussians: $R(x) = r_1 e^{\left(\frac{x-x_1}{b}\right)^2} + r_2 e^{\left(\frac{x-x_2}{b}\right)^2}$ (the exact distribution is not important for matching) with $\tau = 50ms, \mu =$

$5, d_0 = 5, v = 40cm\ s^{-1}, x_1 = +30cm, x_2 = -30cm, b = 20cm$ ($\beta = \mu d_0^{-1} = 1$, Methods). The model was simulated for different ratios of

r_1/r_2, and the response ratio was estimated by the ratio of the time spent $\pm 2.5cm$ from x_1, x_2.

Reward-taxis and pathologies of the dopamine system

Variation and defects in the dopamine system can be classified by their effects on the

parameters of the reward-taxis model. Variation in intrinsic neuronal parameters can affect

gain μ and baseline firing rate d_0. Such changes are predicted to affect the matching law

behavior by changing the exponent β. For example, ADHD is associated both with impaired

sensitivity to reinforcement (corresponding to low β) (Kollins *et al*, 1997; Luman *et al*, 2010)

and impaired dopaminergic gain (corresponding to low μ) (Tripp & Wickens, 2008; Hauser *et*

al, 2016). Reduced gain can also cause reduced sensitivity to reward gradients so that only

strong reward gradients produce continuous focused climbing, as shown by work in bacterial

chemotaxis (Long *et al*, 2017). Such hypotheses are in principle testable by detailed behavioral

experiments and by experimental manipulation of neuronal gain and baseline properties using

drugs or genetic tools.

Other disorders can be due to reduction in the number of functional dopaminergic neurons, as

in Parkinson's disease, where SNc dopaminergic neurons are lost. Such damage is predicted to

change mean run duration τ and/or mean motion vigor v_0. If the damage does not sizably affect

the intrinsic properties of surviving neurons, such as gain and baseline, they are not expected

to change β in matching law experiments.

The analogy to chemotaxis can also be used to explore the role of variability in parameters.

Variability can occur over time in the same individual (state), or between individuals (trait). In

E. coli chemotaxis the basal tumbling rate and adaptation time show large non-genetic

variability between bacteria that lasts a generation time (Spudich & Koshland, 1976; Levin *et*

al, 1998; Levin, 2003; Frankel *et al*, 2014; Keegstra *et al*, 2017; Salek *et al*, 2019). This

variation is consequential for gradient climbing performance. Lower basal tumbling frequency

improves navigation towards distant targets, whereas high basal tumbling frequency is

preferable for nearby targets (Frankel *et al*, 2014). More recently, it was discovered also that

pathway gain varies between individual bacteria. Theoretical work indicates that high pathway

gain is beneficial for localizing around a single attractant peak, whereas low pathway gain

allows efficient crossing between attractant peaks (Karin & Alon, 2021).

Mapping these findings to the dopamine model suggests that state and trait variation can affect

reward-taxis. Trait-like variation may be due to developmental or genetic factors. State-like

variation in parameters, on the other hand, can potentially be implemented by neuromodulators

such as epinephrine and serotonin. Movement vigor shows individuality in humans (Reppert

et al, 2018), which is in some ways analogous to individuality in baseline tumbling frequency.

The reward-taxis model therefore provides testable predictions for the effects of such variations

on movement and search behavior.

<u>Discussion</u>

This study unifies the two main effects of dopamine, encoding reward-derivative and

increasing movement vigor, by mapping them to a reward-taxis navigational circuit. The circuit

is analogous to the bacterial chemotaxis circuit, where in the dopamine case navigation is along

gradients of expected reward. The mapping is based on mathematical analogies at both the

physiological and behavioral levels. At the physiological level, experiments on the regulation

of dopamine transmission can be parsimoniously explained as fold-change detection (FCD) of

expected reward. At the behavioral level, dopamine increases the probability and vigor of

movements, thus increasing the length of correlated motion ("runs") compared with

reorientations ("tumbles"). Both aspects map to the well-characterized chemotaxis navigation

circuit in bacteria.

FCD is a recurring motif in biological sensory systems (Adler & Alon, 2018). Therefore, the

model of dopamine transmission as FCD of expected reward is in line with the

conceptualization of dopamine neurons as sensory neurons for reward (Watabe-Uchida *et al*,
2017). FCD includes the classic Weber's law of sensory systems, which posits that the maximal
response to a change in input is normalized by the background level of the signal (Ekman,
1959). FCD is more general than Weber's law in that the entire dynamics of the output,
including amplitude and duration, is normalized by the background input level. FCD allows
the system to function in a scale-invariant manner across several decades of background input
(Shoval *et al*, 2010). It also provides a common scale to compare different types of sensory
inputs, by referring to their relative (rather than absolute) changes (Hart *et al*, 2013).
While the model focused on the average activity of dopaminergic neurons, the proposed
mechanism for FCD (inhibition from neighboring GABAergic neurons) may apply at the level
of individual dopaminergic neurons or groups of neurons. This raises the possibility of
heterogeneity between dopaminergic neurons – different dopaminergic neurons could become
adapted to different expected-reward levels at the same time. This would be consistent with a
recent study that demonstrated that a single reward can simultaneously elicit positive and
negative prediction errors in different neurons (Dabney *et al*, 2020).
Our model places vertebrate motion regulation in the wider family of run-and-tumble stochastic
navigation circuits, which includes motion regulation in bacteria, algae, and simple animals
(Pierce-Shimomura *et al*, 1999; Polin *et al*, 2009; Luo *et al*, 2014; Kirkegaard *et al*, 2016).
Reward-taxis was anticipated in the early work on TD learning, where Montague et al. showed
that run-and-tumble dynamics driven by reward prediction errors can explain observations on
bee foraging (Montague *et al*, 1995). Run-and-tumble navigation with FCD guarantees
effective sampling of the rewards distributed in the environment, by implementing a search
algorithm (Eq. 6) analogous to the widely used Langevin Monte Carlo (LMC) algorithm for
sampling probability distributions (Roberts & Tweedie, 1996; Neal, 2011; Girolami &
Calderhead, 2011; Dalalyan, 2014) and for global optimization (Chiang *et al*, 1987; Gelfand &
Mitter, 1991; Lee *et al*, 2018; Erdogdu *et al*, 2018; Ma *et al*, 2019; Chen *et al*, 2020). The
animal samples a probability distribution given by a power of the distribution of expected
rewards. This explains the generalized matching law of operant behavior. Furthermore, run-
and-tumble navigation provides benefits beyond the Langevin Monte Carlo algorithm by
boosting gradient climbing on sufficiently steep reward gradients due to the positive feedback
between behavior and sensing. The positive feedback occurs since running along the gradient
provides increasing input that further enhances the run duration (Long *et al*, 2017).
The model provides a quantitative and mechanistic explanation of the generalized matching
law of operant behavior, with the matching exponent given by the ratio of dopaminergic gain

to basal activity. The logarithmic derivative property is crucial for obtaining the matching law.
A non-logarithmic derivative, or absolute responses, do not provide matching (Supplementary
Information 3).

The model predicts that manipulating the relative gain of dopamine neurons μd_0^{-1} will change
the reward sensitivity parameter β in the matching law. This prediction can be tested by
measuring μd_0^{-1} in genetically modified animals where matching behavior is different from
wild-type. One such case is mice that are deficient in the cannabinoid type-1 receptor (CB$^{-/-}$),
which have β that is lower by ~30% compared with wild-type mice (Sanchis-Segura et al,
2004). In agreement with the model prediction, CB$^{-/-}$ mice also have deficient dopaminergic
responses relative to baseline (Li et al, 2009).

FCD also causes navigation to be invariant (i.e., not affected by) multiplying the input field by
a scalar (Shoval et al, 2010) – in other words, by multiplying expected rewards across space
by a constant factor λ. The FCD model predicts that the entire temporal dynamics of the search
process, including the distribution of search times, the distribution of run lengths, etc., are
invariant to the scale λ of reward magnitude. In fact, FCD is a necessary and sufficient
condition for scale invariance of a search mechanism (Shoval et al, 2010). Scale invariance is
prevalent across aspects of perception and behavior (Chater & Brown, 1999; Maylor et al,
2001; Stewart et al, 2006; Hart et al, 2018; Han et al, 2020). This property may be crucial for
the dopamine circuit, since rewards can vary widely in scale and magnitude.

Together with the similarities there are also differences between bacterial chemotaxis and the
reward-taxis model for dopamine. The value of β in bacterial chemotaxis is much larger than
in the dopamine reward-taxis model, with $\beta > 10$ in $E.$ $coli$ (Hu & Tu, 2014; Karin & Alon,
2021) and $\beta \sim 1$ estimated for the dopamine system. The high β value in bacteria indicates a
strong preference for higher rewards, akin to an optimization for accumulation near attractant
peaks. A value of $\beta \sim 1$ (which results in the matching law) allows a greater range for
exploration of submaximal rewards.

While the presentation of reward-taxis in this study focused on spatial movement, its
assumptions are general and it may therefore apply to other behavioral systems. We discussed
above the possibility that it applies to hippocampal replay. Another potential system is eye
movements. Eye movements are modulated by dopamine and impaired in Parkinson's disease
(Kori et al, 1995; Chan et al, 2005; Pretegiani & Optican, 2017), and their vigor is modulated
by reward prediction errors (Sedaghat-Nejad et al, 2019). Since eye movements are commonly

studied in behavioral experiments such as reward matching, they may be a good candidate to
test the reward-taxis model.

The present minimal model does not include several important aspects including goal-directed
behavior and planning. Such aspects are likely to complement stochastic navigation with more
directed movement. Such a combination of navigation mechanisms is evident in simple
organisms that employ run-and-tumble navigation. For example, in *C. elegans* thermotaxis,
run-and-tumble navigation is combined with biased reorientations in order to navigate towards
an optimal temperature range (Luo *et al*, 2014). Formally, while run-and-tumble navigation
resembles Langevin-based sampling, directed reorientations are more closely related to
gradient descent, which is efficient for local optimization but poor for global optimization (Ma
et al, 2019; Raginsky *et al*, 2017; Xu *et al*, 2018). Understanding how animals combine
stochastic navigation with other behavioral strategies can therefore improve our understanding
of the algorithmic basis of animal behavior.

Methods

Model equations and fold-change detection

The equations for dopamine (d) and GABAergic inhibition (g) are provided by:

472
$$\dot{d} = \omega_d(C + \mu \log R - \alpha g - d) \quad [8]$$

473
$$\dot{g} = \omega\left(\frac{d}{d_0} - 1\right) \quad [9]$$

For the dopamine equation, ω_d determines the dopamine degradation rate, μ is dopamine gain,

R is expected reward (defined in the next Methods section), and α is GABAergic inhibition

strength. For the GABAergic inhibition equation, ω determines the adaptation rate and d_0 is

the adapted steady-state of dopamine. As is common in FCD circuits, we assume that dopamine

dynamics are faster than the dynamics of adaptation due to g (i.e., ω_d is large compared with

ω) so we take:

480
$$d = C + \mu \log R - \alpha g \quad [10]$$

Eqs. 8,9,10 can represent the average activity of individual neurons, or the total activity of

many neurons. We therefore used the same equations both to model average individual neuron

recordings (as in Figure 1), and to model the effect of dopamine on movement, which is likely

to be the sum of the activity of many neurons.

Consider now a constant input $R = R_0$, so that after some time the system reaches steady-state.

To find the steady state, we solve Eqs. 8,9, taking $\dot{d} = 0, \dot{g} = 0$, which yields the steady-state

solutions $d_{st} = d_0$ and $g_{st} = \alpha^{-1}(C - d_0 + \mu \log R_0)$. The observation that $d_{st} = d_0$

regardless of R_0 and other circuit parameters is an important circuit feature from systems

biology known as *exact adaptation* (Barkai & Leibler, 1997; Alon *et al*, 1999; Ma *et al*, 2009;

Ferrell Jr, 2016; Alon, 2019). This feature is essential for explaining why dopamine activity

returns precisely to baseline after a step increase in expected reward, while GABAergic activity

increases in a way that tracks expected reward.

Beyond exact adaptation, the system has an even stronger property of *fold-change detection*

(FCD). FCD is defined as dynamics of dopamine (d) in response to an input $\lambda R(t)$ that are

independent of λ, starting from initial conditions at steady-state for $\lambda R(0)$. To show this we

replace $g = g' + \alpha^{-1}\mu \log \lambda$:

497
$$\dot{d} = \omega_d(C + \mu \log \lambda R - \alpha g - d) = \omega_d(C + \mu \log R - \alpha g' - d) \quad [11]$$

498
$$\dot{g}' = \omega\left(\frac{d}{d_0} - 1\right) \quad [12]$$

Eqs. 11, 12 are completely independent of λ, and their steady-state $g'_{st} = \alpha^{-1}(C - d_0 +$

$\mu \log R_0)$ and $d_{st} = d_0$ is also independent of λ. This means that the dynamics of the system

have the FCD property. The FCD property is essential for explaining the scale invariance of

the dopaminergic responses to rewards in Fig. 1D – the response only depends on the fold-

change of expected reward (two-fold change upon reception of reward at p=0.5) but not on

reward magnitude.

While Eqs. 8,9,10 provide FCD, they are not the only possible model that provides FCD for

this system. In particular, a feed-forward model where expected reward activates g is also

possible, i.e.:

$$\dot{d} = \omega_d(d_0 + C + \mu \log R - \alpha g - d) \quad [13]$$

$$\dot{g} = \omega\left(\frac{C + \mu \log R}{\alpha} - g\right) \quad [14]$$

510   For this circuit, the steady state for a constant input $R = R_0$ is $g_{st} = \frac{a + \mu \log R_0}{\alpha}$ and $d_{st} = d_0$.

511   FCD can also be analogously shown. Given an input $\lambda r(t)$, we can take $g = g' + \alpha^{-1}\mu \log \lambda$,

512   which again provides equations and steady-state that are independent of λ:

$$\dot{d} = \omega_d(d_0 + C + \mu \log \lambda R - \alpha g - d) = \omega_d(d_0 + C + \mu \log R - \alpha g' - d) \quad [15]$$

$$\dot{g}' = \omega\left(\frac{C + \mu \log \lambda R}{\alpha} - g\right) = \omega\left(\frac{C + \mu \log R}{\alpha} - g'\right) \quad [16]$$

Providing FCD. Parameters used in simulations are provided in Table 1.

While this simple log-linear model captures various important experimental observations, it is

important to note that it has some clear limitations. One limitation is that both d and g can in

principle reach negative values when R is small. Measurements of dopamine responses in

monkeys indeed show deviations from sub-linearity for small rewards (Stauffer *et al*, 2014).

Future studies may build on improved measurements and better mechanistic characterization

of the dopamine circuit to refine this model.

Definition of expected reward and relation between the circuit and TD learning

Here we will define the input to the circuit, which is the logarithmic expected reward $\log R$,

and present it in the context of the temporal difference (TD) learning theory of dopamine

function (Schultz *et al*, 1997; Sutton & Barto, 1998). We first define the expected temporally

discounted sum of future rewards V (also known as the value function in TD learning):

$$V(t_0) = \mathbb{E}\left[\sum_{t=t_0}^{\infty} \gamma^t r(t) dt\right] \quad [17]$$

Where $\gamma < 1$ is a "future discounting" factor and $r(t)$ is the reward received at time t into the

future (here for simplicity we take discrete time; for equivalent formulation for continuous

time, see Doya et al. (Doya, 2000)). It is possible to think of V as a function of the current state

s of the agent, which may include for example its location in space x. This is known as the

Markovian setting, where we denote the value function as $V(s)$. The value function plays an

important role in decision making - learning the value function is a principal focus of

reinforcement learning algorithms (Schultz *et al*, 1997; Sutton & Barto, 1998). However, for

our purposes, we assume that V is already learned and can be provided to the circuit as an input.

In our model, the input to the circuit for an agent moving into a state s at time t is defined using

the expected reward R:

$$R(t, s) = r(t) + V(s) = r(t) + V(t) \quad [18]$$

As an example, consider the setting of Figure 1D, where a reward of size $r = y$ is delivered

with probability p at Δt time-units into future: the expected reward would in this case be

$R(0) \approx p\gamma^{\Delta t} y$. An actual delivery of the reward would then increase R to $R(\Delta t) \approx y$, so the

ratio $\frac{R(\Delta t)}{R(0)} = \frac{1}{p\gamma^{\Delta t}}$ is independent of reward magnitude.

Note that due to discounting and uncertainty, R decays with the distance from a location where

a reward is delivered, as in Figure 1E (Gershman, 2014).

We will now show that our model is consistent with the TD learning theory of dopamine

function. We will first briefly present the TD learning algorithm. In the reinforcement learning

setting, the agent usually does not know V and needs to learn it from experience. This presents

a computational problem, since V is an infinite sum over unknown future events. A way to get

around this is to update the learned V using dynamic programming (Barto *et al*, 1989). The key

insight is that Eq. 17 can be rewritten as:

$$V(t_0) = \mathbb{E}\left[\sum_{t=t_0}^{\infty} \gamma^t r(t) dt\right] = \mathbb{E}[r(t_0)] + \gamma V(t_0 + 1) \quad [19]$$

552 The above equation implies that V can be estimated iteratively with a simple update rule, which

553 is at the heart of TD learning. If the agent is at state s at time t, and at state s' at time $t+1$, the

554 update rule is:

$$V_{t+1}(s) \leftarrow V_t(s) + \alpha \underbrace{\left(r(t) + \gamma V_t(s') - V_t(s)\right)}_{prediction\ error} \quad [20]$$

Where $V_t(s)$ is the computed estimate of the expected reward at state s at time t, α is the

learning rate, $r(\text{t})$ is the reward delivered at time t and γ is the discounting factor. There is an

extensive literature demonstrating correspondence between TD learning and midbrain

dopamine function (reviewed by (Glimcher, 2011)); specifically, experiments show convincing

correspondence between phasic dopamine secretion and the prediction error term of Eq. 20

(Schultz *et al*, 1997; Glimcher, 2011), in the sense that positive or negative firing of dopamine

neurons relative to baseline corresponds positive and negative predictions errors in TD models

of learning.

The FCD model proposed here is consistent with the correspondence between dopamine

function and TD learning. To see this, notice that learning the logarithm of V is equivalent to

learning V. Also note from Eq. 19:

$$\log V(t_0) = \log\big(\mathbb{E}[r(t_0)] + \gamma V(t_0 + 1)\big) \approx \mathbb{E}\big[\log\big(r(t_0) + \gamma V(t_0 + 1)\big)\big] \quad [21]$$

The last approximation holds when r is either narrowly distributed, or not very large compared

with $\gamma V(t_0 + 1)$. Thus, the following update rule is approximately equivalent to the TD update

rule, but for $\log V$:

$$\log V_{t+1}(s) \leftarrow \log V_t(s) + \alpha \underbrace{\big(\log\big(r(t) + \gamma V_t(s')\big) - \log\big(V_t(s)\big)\big)}_{prediction\ error} \quad [22]$$

We note that in the continuous limit, and in the absence of reward, the error term is

approximately proportional the logarithmic derivative of the value function. This corresponds

to the output of our proposed FCD model for low frequency signals. The output of the circuit

to a delivered reward in a transition from a state s to a state s' is also similar to the above error

term:

$$d(t + dt) - d_0 = C + \mu \log R(t + dt) - \alpha g(t + dt) - d_0$$

$$= \mu\big(\log\big(r(t) + V(s')\big)\big) - \log(V(s)) \quad [23]$$

The final equality is due to the fact that prior to reward delivery, GABAergic output adapts to

$\alpha^{-1}(C - d_0 + \mu \log V(s))$. The FCD model is therefore compatible with the TD learning

theory of dopamine function.

We conclude by noting that since the value that is learned is $\log V$, providing the input to the

circuit $\log\big(r(t) + V(t)\big)$ requires a rather complex calculation; however, this can be simplified

by using the approximation $\log(r + V) \approx \max(\log r, \log V)$, which is a much simpler

calculation for neural circuits (Gawne & Martin, 2002; Knoblich *et al*, 2006).

Table 1

Parameter	Value (mouse experiments)	Value (primate experiments)
ω_d	50 s^{-1}	100 s^{-1}
ω	15 s^{-1}	10 s^{-1}
C	15 spike s^{-1}	15 spike s^{-1}
μ	6 spike s^{-1}	6 spike s^{-1}
α	0.7	0.7
d_0	5 spike s^{-1}	5 spike s^{-1}

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