

1 **Punctual ecological changes rather than global factors drive species diversification and**  
2 **the evolution of wing phenotypes in *Morpho* butterflies.**

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22

23

24 **Abstract**

25 Assessing the relative importance of geographical and ecological drivers of evolution is  
26 paramount to understand the diversification of species and traits at the macroevolutionary  
27 scale. Here, we use an integrative approach, combining phylogenetics, biogeography,  
28 ecology, and quantified phenotypes to investigate the drivers of both species and phenotypic  
29 diversification of the iconic Neotropical butterfly genus *Morpho*. We generated a time-  
30 calibrated phylogeny for all known species and inferred historical biogeography. We fitted  
31 models of time-dependent (accounting for rate heterogeneity across the phylogeny) and  
32 paleoenvironment-dependent diversification (accounting for global effect on the phylogeny).  
33 We used geometric morphometrics to assess variation of wing size and shape across the tree,  
34 and investigated their dynamics of evolution. We found that the diversification of *Morpho* is  
35 best explained when considering multiple independent diversification dynamics across the  
36 tree, possibly associated with lineages occupying different microhabitat conditions. First, a  
37 shift from understory to canopy was characterized by an increased speciation rate partially  
38 coupled with an increasing rate of wing shape evolution. Second, the occupation of dense  
39 bamboo thickets accompanying a major host-plant shift from dicotyledons towards  
40 monocotyledons was associated with a simultaneous diversification rate shift and an  
41 evolutionary “jump” of wing size. Our study points to a diversification pattern driven by  
42 punctual ecological changes instead of a global driver or biogeographic history.

43

44 **Keywords**

45 Species diversification, phenotypic diversification, wing size, wing shape, geometric  
46 morphometrics, butterflies, *Morpho*.

## 47 **Introduction**

48           Investigating the rates of phenotypic evolution and the relationships between  
49 phenotypes and species ecology can shed light on the drivers of time and geographic patterns  
50 of diversity. Previous studies have demonstrated that rates of both species and phenotypic  
51 diversification vary widely through time and among clades at all taxonomic scales (e.g.  
52 Venditti *et al.*, 2011; Eastman *et al.*, 2011; Rabosky & Adams, 2012; Rabosky *et al.*, 2013;  
53 Rabosky *et al.*, 2014; Cooney & Thomas, 2020). These variations have resulted in the  
54 striking heterogeneity in species and phenotypic diversity observed across the tree of life.  
55 Such variations may eventually be coupled, indicating an interaction between the processes  
56 of species and phenotypic diversifications. Studies investigating such coupling have yielded  
57 contrasted results. Some of them support an association between specific and phenotypic  
58 diversification (e.g. in salamanders: Rabosky & Adams, 2012; fish: Rabosky *et al.*, 2013;  
59 vertebrates: Cooney & Thomas, 2020), while others found no support for this relationship  
60 (e.g. in lizards: Rabosky *et al.*, 2014; squirrels: Zelditch *et al.*, 2015; reef fishes: Price *et al.*,  
61 2015; snakes: Lee *et al.*, 2016). For example in squirrels, Zelditch *et al.* (2015) suggested that  
62 species diversification was geographically driven while phenotypic diversification was  
63 ecologically driven, resulting in a decoupling of the two dynamics.

64           A correlation between species and phenotypic diversification rates is notably expected  
65 in some specific cases. For example, adaptive radiations – rapid adaptive diversification in a  
66 variety of ecological niches – are expected to produce bursts of diversification and  
67 phenotypic evolution especially during the initial stages of diversification (Schluter, 2000;  
68 Gavrilets & Losos, 2009). Speciation rate increases when a large number of ecological niches  
69 are vacant while phenotypes rapidly evolve in response to the diversity of ecological  
70 opportunities. Strong correlation between speciation rates and phenotypic diversification may  
71 also be found when the focal trait directly drives reproductive isolation. For example, the

72 evolution of male genitalia, involved in mating, may facilitate reproductive isolation between  
73 populations (see Langerhans *et al.*, 2016 for a review). Correlated dynamics leading to a  
74 lower rate of diversification can also be predicted. For example, if extinction probability is  
75 biased with respect to phenotype leading to a non-random loss of variation in a particular  
76 clade, both species and morphological diversity should show a correlated drop down (Foote,  
77 1997). In this study, we assess the role of multiple ecological causes of variations in rates of  
78 species and wing diversification and the extent to which these variations are coupled, by  
79 focusing on the case of the butterfly genus *Morpho* (Nymphalidae).

80         The genus *Morpho* comprises 30 species (Blandin & Purser, 2013), which are  
81 amongst the largest butterflies in the Neotropics and are well known for their blue iridescent  
82 wing coloration. Several ecological factors have already been suggested as potential drivers  
83 of diversification and phenotypic evolution. Previous biogeographic estimations suggested  
84 that *Morpho* butterflies originated and started diversifying in the Andes (Penz *et al.*, 2012,  
85 Blandin & Purser 2013), before spreading across the Neotropics. There is also evidence that  
86 *Morpho* lineages separated early in their history into two microhabitats (DeVries *et al.*, 2010;  
87 Chazot *et al.*, 2016). One clade is composed of species that tend to fly high, often above the  
88 forest canopy, with some species typically harbouring gliding flight behaviour such as *M.*  
89 *cisseis* and *M. hecuba*. The remaining species mostly fly within the first meters above ground  
90 in the understory (DeVries *et al.*, 2010; Chazot *et al.*, 2016). Finally, according to Cassildé *et*  
91 *al.* (2010) and Penz *et al.* (2012), the genus *Morpho* was ancestrally feeding on  
92 monocotyledons, and two major host-plant shifts occurred during its diversification: after the  
93 first divergence event, one of the two clades shifted to dicotyledon host-plants and, within  
94 this clade, a subclade subsequently reversed to the monocotyledons.

95         Here we focus on the wings of *Morpho*, which are at the crossroad of multiple  
96 selective pressures and tightly linked to species diversification. Typically, wing colour

97 patterns can be involved in camouflage, aposematism or courting behaviours (Naisbit *et al.*,  
98 2001; Merrill *et al.*, 2011). Wings also allow flight, enabling dispersal, foraging, predator  
99 escape, mating or host-plant searching (Dudley, 2002). Hence butterfly wings are under  
100 strong natural and/or sexual selection and may be associated with variations of speciation rate  
101 (Ortiz-Acevedo *et al.*, 2020). Both size and shape are important aspects of wing morphology.  
102 They both strongly affect the performance of flight behaviours (Dudley, 2002; Le Roy *et al.*,  
103 2019) and therefore might be closely associated to habitat use, dispersal strategies or host-  
104 plant searching. Besides, fore and hind wings can be functionally differentiated, for example  
105 during flight (Grodnitsky *et al.*, 1994; Le Roy *et al.*, 2020), which may lead to uncorrelated  
106 patterns of diversification.

107 To investigate whether species and phenotypic diversification dynamics are coupled  
108 and to identify potential drivers of variations, we inferred a time-calibrated molecular  
109 phylogeny of the genus that we combined to a dataset of geographical distributions and  
110 morphometric measurements of wing size and shape. We applied an integrative approach and  
111 addressed the following questions: (1) Have rates of phenotypic diversification varied across  
112 the tree? We investigated potential variations in rate of phenotypic diversification among  
113 clades using phenograms and models of trait evolution to compare evolutionary rates for  
114 wing size and shape. (2) Is species diversification better explained by global processes or  
115 clade-specific (ecological) factors? First, we fitted different models of species diversification  
116 testing for global drivers of diversification, specifically past temperatures and Andean  
117 orogeny. Second, we compared these global drivers to models in which species  
118 diversification varied according to clade-specific ecological factors (microhabitat and major  
119 shifts of host-plants) and/or variations of phenotypic diversification identified in the first step.  
120 (3) Can we explain the variations in diversification rates by historical biogeography? We  
121 performed ancestral areas estimation in order to assess whether variations in phenotypic

122 evolutionary rates or species diversification rates may be associated with specific  
123 biogeographic events.

124

## 125 **Material and methods**

### 126 *Time-calibrated phylogeny*

127 Phylogenetic relationships and divergence time were inferred with Bayesian inference. We  
128 concatenated DNA data for one mitochondrial (COI) and four nuclear genes (CAD, EF-1 $\alpha$ ,  
129 GAPDH and MDH) using published sequences (Cassildé *et al.*, 2012; Penz *et al.*, 2012;  
130 Chazot *et al.*, 2016) retrieved from GenBank, generating a molecular dataset of a total length  
131 of 5001 nucleotides. Our dataset includes all *Morpho* species (i.e. 30 species *sensu* Blandin,  
132 2007). *Morpho helenor*, which harbours many subspecies, is distributed throughout the entire  
133 Neotropical region, resulting in unresolved biogeographic reconstructions in preliminary  
134 analyses. To help resolving the biogeographic inferences, *M. helenor* was represented in the  
135 biogeographic analyses by six subspecies that each occupies a distinct Neotropical area. For  
136 all other analyses, we pruned all subspecies of *M. helenor* but one in order to keep a single  
137 branch for the species. We also included 11 outgroups to root and calibrate the tree (see  
138 Supporting Information S1) on the basis of the most comprehensive nymphalid phylogeny to  
139 date (Wahlberg *et al.*, 2009).

140 To simultaneously estimate the topology and branching times of the phylogeny we  
141 used a Bayesian relaxed-clock approach as implemented in *BEAST* 1.8.2 (Drummond *et al.*,  
142 2012). To choose the best partitioning strategy and the corresponding substitution models, we  
143 ran *PartitionFinder* 1.1.1 (Lanfear *et al.*, 2012) allowing all possible partitions and models  
144 implemented in *BEAST*. Three subsets were defined: the first included position 1 and 2 of all  
145 genes and followed a GTR+I+ $\Gamma$  model, the second included position 3 of all nuclear  
146 fragments and followed a GTR+ $\Gamma$  model, and the third including the position 3 of the

147 mitochondrial fragment and followed a TrN+ $\Gamma$  model. We implemented an uncorrelated  
148 lognormal relaxed clock model. Given the lack of *Morpho* fossil we relied on secondary  
149 calibrations to calibrate the molecular clock. Penz *et al.* (2012) calibrated the divergence  
150 between *Morpho* and its sister groups using a unique calibration point from Wahlberg *et al.*  
151 (2009), and a normal distribution for the corresponding prior. However, Sauquet *et al.* (2012)  
152 showed that using a single secondary calibration prior could yield biased estimates. Hence,  
153 we used a set of seven calibrations defined by uniform priors bounded by the 95% credibility  
154 intervals (95% CI) estimated by Wahlberg *et al.* (2009) (see Supporting Information S1). We  
155 implemented a Yule process for the tree prior, and we ran the phylogenetic analyses for 30  
156 million Markov chain Monte Carlo (MCMC) generations. We checked for chain convergence  
157 using *Tracer* 1.6, as indicated by effective sample size (ESS) values. Finally, we used  
158 *TreeAnnotator* 1.8.2 (Drummond *et al.*, 2012) to select the maximum clade credibility (MCC)  
159 tree with median age values calculated from the posterior distribution of branch lengths,  
160 applying a 20% burn-in.

161

### 162 ***Morphological data***

163 Our morphological dataset (published by Chazot *et al.* 2016) consists in the size and shape of  
164 the fore and hind wings, as assessed by morphometric measurements. A total of 911  
165 collection specimens of both sexes and representing all *Morpho* species were photographed.  
166 Wing shape was described using landmarks and semi-landmarks placed at vein intersections  
167 and wing margins, respectively (see Chazot *et al.*, 2016 for details), which were  
168 superimposed with *tpsRelw* (Rohlf, 1993). Wing size was measured using the log-  
169 transformed mean centroid size per species. Importantly, for analyses involving wing shape  
170 we used the residuals of a multivariate regression of species mean Procrustes coordinates on  
171 species mean centroid size (log-transformed), which allows focusing on the non-allometric

172 shape variation. Similar analyses were performed separately on the fore and hind wings. All  
173 analyses were performed on males and females separately. As we found divergent patterns  
174 among sexes, we show the results for males and females separately. No female *M. niepelti*  
175 was available. This species was therefore pruned from the tree for all analyses involving  
176 female data.

177

### 178 *Dynamics of phenotypic diversification*

179 We investigated whether the evolutionary rates of wing size and shape have varied among  
180 subclades across the phylogeny.

181 *Wing size* – We first visualized the evolution of traits through time using the  
182 *phenogram* function in *PHYTOOLS* 0.5-20 (Revell, 2012), which represents the trait values  
183 inferred at each node along a time axis. Second, we investigated the dynamics of wing size  
184 evolution across lineages using the method implemented in the function *rjmcmbm* available  
185 in *GEIGER* 2.0.6 (Harmon *et al.*, 2008; Eastman *et al.*, 2011) for univariate traits. This  
186 method uses Bayesian analyses and reversible-jump MCMC to infer the number and the  
187 location of shifts of morphological diversification dynamics. We fitted and compared three  
188 different models of trait evolution: (1) a single-rate Brownian model (BM); (2) a relaxed  
189 model of Brownian evolution in which a trait evolved according to distinct Brownian-motion  
190 models across the tree (rBM); and (3); a model in which trait evolution can also occur at  
191 punctual “jumps”, i.e. brief periods of rapid evolution at any branch in the phylogeny (jBM).  
192 We ran models on both the MCC and a posterior distribution of trees. For the MCC tree  
193 analysis, we ran for each model one MCMC of 30 million generations, sampling every 3,000  
194 generations. We checked for convergence of each run using *CODA* (Plummer *et al.* 2020),  
195 and computed the ESS. We applied a 25% burn-in and compared the three models using the  
196 Akaike's Information Criterion for MCMC samples *aicm* and *aicw* implemented in *GEIGER*



197 (Supporting Information S2). To assess the robustness of the inferences to branch length  
198 uncertainties, we repeated the analysis on a posterior distribution of trees and summarized the  
199 results. We sampled 100 trees with a topology identical to that of the MCC tree from the  
200 posterior distribution. For each tree, we ran the three models but reduced the MCMC to 10  
201 million generations, calculated the *aicm* score, the mean *aicm* pairwise differences between  
202 models and the position of rate shifts and jumps. We summarized the results by calculating  
203 the frequency of shifts and jumps at nodes across the posterior distribution. These results are  
204 hereafter referred to as shift/jump posterior tree frequencies.

205 *Wing shape* –Some authors have used the scores on the first PC-axis as a univariate  
206 shape measure (e.g. Rabosky *et al.*, 2014; Thacker, 2014) to investigate shifts in evolutionary  
207 rates for multidimensional traits, but this may lead to spurious results (Uyeda *et al.*, 2014).  
208 We rather investigated variations in rates of shape evolution across the phylogeny in a  
209 multivariate way using the function *compare.evol.rate* from *GEOMORPH* (Adams, 2014;  
210 Denton & Adams, 2015). It allows testing whether species assigned to different ecological  
211 factors have significantly different rates of shape evolution, by comparing the ratio between  
212 the rates of each group to a null distribution of ratios obtained through simulation of a unique  
213 neutral evolutionary rate (two or more factors can be tested). When more than two factors are  
214 included, the function performs a global test for the significance of the multiple rates model  
215 compared to a one-rate model, but also assesses the significance of differences among each  
216 pair of factors. We used this factor assignment to define monophyletic subgroups with  
217 potentially divergent evolutionary rate from the background rate. We first tested all models  
218 with one shift, i.e. all species belonging to one subclade (each subclade had a minimum of  
219 three species) are assigned to one group, and the rest of the species assigned to another group.  
220 If two or more subclades were identified as having a rate of evolution significantly different  
221 from that of the background, we identified the subclade with the highest ratio (hence the

222 greatest shift). Then we ran again *compare.evol.rate* on all possible combinations of two  
223 shifting subclades that include the first identified shift. A two-shift model was considered  
224 significant if at least the two shifting subclades showed a significant difference with the  
225 background rate when considering the pairwise comparisons. Given the relatively small size  
226 of our phylogeny, we limited our analysis to two shifts (Supporting Information S3-S4). As  
227 for wing size, we repeated the analysis on both the MCC tree and a posterior distribution of  
228 trees with identical topologies. We summarized the results from the posterior distribution by  
229 calculating the frequency of significant shifts at nodes across the trees, and refer to these as  
230 posterior tree frequencies.

231

### 232 ***Dynamics of species diversification***

233 We compared two types of species diversification models: (1) diversification rates varying  
234 according to global factors, i.e. factors virtually affecting all lineages, and (2) diversification  
235 rates varying at specific clades characterized by clade-specific ecological factors. For each  
236 type we investigated different factors (see below). All models were compared using their AIC  
237 scores to identify the model that best explains the diversification of the genus *Morpho*.

238 *Global drivers of diversification* – We tested the role of temperature fluctuations and  
239 of the paleo-elevation of the Andes on species diversification by using birth-death models  
240 that allow speciation and extinction rates to vary according to a past environmental variable  
241 itself varying through time (Condamine *et al.*, 2013). For each paleoenvironmental variable,  
242 we designed three models to be tested: (i) the speciation rate varies exponentially with the  
243 environment and the extinction rate is constant, (ii) the speciation rate is constant and the  
244 extinction rate varies exponentially with the environment, and (iii) both speciation and  
245 extinction rates vary exponentially with the environment. We repeated these three models  
246 with a linear dependence to the environmental variable, instead of exponential dependence.

247 For temperature we relied on the well-known Cenozoic temperature dataset published by  
248 Zachos *et al.* (2008). The orogeny of the Andes is a highly complex process, with important  
249 differences in uplift tempo and mode from the south of Central Andes to Northern Andes  
250 (Blandin & Purser, 2013, and references therein). Several general phases have been identified  
251 from the late Eocene to present, but they are difficult to synthesize in a unique model. As  
252 Blandin & Purser (2013) suggested that the early diversification of the *Morpho* occurred  
253 along the proto-Central Andes, we used the model of surface uplift inferred by Leier *et al.*  
254 (2013) for the eastern cordillera of the southern Central Andes to test the possible influence  
255 of Andean orogeny on the diversification of the *Morpho*. We used the R-package *PSPLINE*  
256 1.0-17 to reconstruct smooth lines of the paleo-data for each environmental variable. The  
257 smooth line is introduced in the birth-death model to represent the variation of the  
258 environment through time. Given the dated phylogeny, the model then estimates speciation  
259 and extinction rates, as well as their respective variations according to the environment  
260 (Condamine *et al.*, 2013). These analyses were performed on 200 trees randomly sampled  
261 from the posterior distribution generated by BEAST.

262 *Clade-specific drivers of diversification* – We assessed whether the diversification  
263 rates across the genus *Morpho* have varied among specific clades using models of time-  
264 dependent diversification. To do so we used the method developed by Morlon *et al.* (2011),  
265 which allows partitioning diversification rates into independent dynamics (a backbone and  
266 different subclades). We compared different partitioning schemes according to three events:  
267 (1) the microhabitat change (from understory to canopy), (2) the shift of wing shape  
268 evolutionary rate, and (3) the reverse shift to monocotyledon host-plants (also identified as a  
269 punctual evolutionary jump of wing size at the stem). Because the evolutionary rate shift of  
270 wing shape is nested within the microhabitat shift (see Results), we could not test both  
271 combined. Instead, each of those shifts was combined to the monocotyledon host-plant shift

272 with a two-shift model of diversification rate. For each subclade and the remaining backbone,  
273 we fitted the following models: (i) constant speciation rate and no extinction, (ii) time-  
274 dependent speciation rate and no extinction, (iii) constant speciation and extinction rates, (iv)  
275 time-dependent speciation rate and constant extinction rate, (v) constant speciation rate and  
276 time-dependent extinction rate, and (vi) time-dependent speciation and extinction rates. Time  
277 dependency was modelled using an exponential function of time. The stem branch of each  
278 subclade was included in the subclades and excluded from the backbones but we kept the  
279 node of the divergence (speciation event) of the subclade within the backbones. The root of  
280 the tree was excluded from the analyses. The analysis was performed on the MCC tree, since  
281 partitioning the tree requires defining clades *a priori*, which entails a fixed topology.

282

### 283 ***Historical biogeography***

284 To assess where and when diversification occurred, we estimated ancestral areas using the  
285 dispersal-extinction-cladogenesis (DEC, Ree and Smith, 2008) model as implemented in the  
286 R-package *BioGeoBEARS* 0.2.1 (Matzke, 2014). The analyses were performed using the  
287 MCC tree (outgroups removed) and included six subspecies of *M. helenor* (each subspecies  
288 was assigned to its current distribution).

289 The distribution of *Morpho* is restricted to South America and Central America (all  
290 Neotropics except the Caribbean Islands). A geographic model was incorporated to include  
291 operational areas, defined as geographic ranges shared by at least two or more species and  
292 delimited by geological, oceanic or landscape features, which may have acted as barriers to  
293 dispersal. The model comprised 7 component areas: (A) Central America, (B) trans-Andean  
294 South-America, (C) slopes of northern Andes, (D) eastern slopes of central Andes, Orinoco-  
295 Amazonian basin north of the Amazon, including the Guyanas, (E) Amazonian basin, south  
296 of the Amazon River, and (F) Atlantic forest.

297 An adjacency matrix was designed whilst taking into account the geological history  
298 and the biological plausibility of combined ranges (Supporting Information S6).  
299 Distributional data were compiled from monographies (Blandin, 2007). We excluded  
300 distribution margins overlapping with adjacent areas. For example, *M. marcus* and *M.*  
301 *eugenia* are mainly found in lowlands but their distributions reach the Andean slopes up to  
302 altitudes of 700-800m. Nevertheless, we did not consider these as species occupying the  
303 Andean biogeographic areas. By contrast, a species such as *M. sulkowskyi*, which occurs  
304 between 1500 and 3500m high in the Andes was considered as an Andean species. We also  
305 set a maximum of 3 areas per node to be constitutive of an ancestral range. We fitted two  
306 different DEC models, one that assumed equal dispersal probabilities among all areas and one  
307 that included time-stratified matrices of varying dispersal probabilities (Supporting  
308 Information S6). We compared the likelihoods of both reconstructions to select the model  
309 best explaining the current pattern of species distribution.

310

## 311 **Results**

### 312 *Divergence times*

313 We estimated that the genus *Morpho* diverged (stem age) from its sister genus *Caerois* 38.05  
314 Ma (95% CI=35.48-39.20 Ma) and the first event (crown age) of diversification was  
315 recovered at 28.12 Ma (95% CI=25.22-31.24 Ma; Supporting Information S1). These  
316 divergence time estimates are slightly older than those estimated by Penz *et al.* (2012) and  
317 Chazot *et al.* (2019) who found an average divergence from *Caerois* around 32.00 Ma and  
318 29.08 Ma respectively. This difference probably results from prior choices for calibrating the  
319 trees (see Material and Methods).

320

### 321 *Dynamics of phenotypic diversification*

322           *Wing size* – Both analyses on the MCC and posterior distribution of trees found  
323 similar results. We found no support for any shift in rate of wing size diversification.  
324 However, we found support for an evolutionary jump. For females the model jBM was highly  
325 supported for both wings, with a highly probable (posterior tree frequency [PF] of 0.99,  
326 Supporting Information S2) evolutionary jump at the root of the clade including the species  
327 *M. absoloni*, *M. aurora*, *M. zephyritis*, *M. rhodopteron*, *M. sulkowskyi*, *M. lympharis*, *M.*  
328 *aega*, and *M. portis* (subclade *portis*). Phenograms show that in this subclade, female wings  
329 are on average 34% smaller than in the other *Morpho* species for both fore and hind wings  
330 (Fig. 1). This is all the more striking as the sister clade (including *M. amathonte*, *M.*  
331 *menelaus*, and *M. godartii*) contains some of the largest species of the genus (e.g. *M.*  
332 *amathonte* has a wingspan of 10–15 cm). For males, the *portis* clade exhibits the same trend,  
333 but the support for the evolutionary jump is lower than for females (PF<sub>forewing</sub>=0.76,  
334 PF<sub>hindwing</sub>=0.71, respectively, Supporting Information S2). Males wings in the *portis* clade  
335 were on average 30 and 32% smaller for fore and hind-wing respectively.

336           *Wing shape* – We found support for two shifts of evolutionary rate for male hindwing,  
337 in both cases towards lower rate of evolution. These subclades encompass *M. helenor*, *M.*  
338 *achilles*, and *M. granadensis* (Fig. 2, Supporting Information S3) on one side, and *M.*  
339 *godartii*, *M. menelaus*, and *M. amathonte* on the other. This result was supported by the  
340 analyses with the MCC tree. The analyses performed on the posterior tree distribution found  
341 a moderate support for these shifts, with PF of 0.62 and 0.77, respectively. For females and  
342 for both wings the subclade encompassing *M. theseus*, *M. amphitryon*, *M. telemachus* and *M.*  
343 *hercules* exhibited the greatest shift (highest ratio) (Fig. 2, Supporting Information S4). This  
344 shift corresponds to a large increase in rate of evolution (forewing ratio=181.74, hindwing  
345 ratio=184.49 in the MCC analysis), i.e. wing shape evolving faster within this group than the

346 other *Morpho*. This result was strongly supported by posterior distribution analyses, with PF  
347 of 0.96 and 0.99 for fore and hind-wing, respectively.

348

### 349 ***Dynamics of species diversification***

350 *Global drivers of diversification* – In the best model accommodating for Central  
351 Andean paleo-altitudes, speciation rates were negatively dependent on the paleo-altitude and  
352 extinction rates were constant (Table 2a). This model leads to a continuous decrease in  
353 speciation rate towards the present, suggesting that *Morpho* diversification was high during  
354 the early stages of the orogeny but the rise in altitude did not lead to any increased  
355 opportunities for speciation over time. We also found a significant correlation between  
356 *Morpho* diversification and temperature compared to a null model (Table 2b). The best fitting  
357 paleoclimatic model indicates that speciation rate was positively correlated with temperature  
358 variation while extinction remained constant. This means that speciation rate was high during  
359 the initial stages of diversification when the temperatures were warmer but globally  
360 decreased during the last 14 million years as the Earth cooled down (Zachos *et al.*, 2008).

361 *Clade-specific dynamics of diversification* – The best-partitioned models included a  
362 shift of diversification rate for the host-plant shift and for the canopy shift (Table 3,  
363 Supporting Information S5). Under this configuration, the diversification of the clade that  
364 shifted to monocotyledon host-plants was best modelled by a speciation rate decreasing  
365 through time combined with no extinction, and the diversification of the canopy clade was  
366 best modelled by a constant speciation rate with no extinction (Table 3, Fig. 3). For the  
367 remaining backbone lineages the best fitting model was a time-dependent speciation and  
368 extinction. The resulting net diversification rate (speciation minus extinction) of this  
369 backbone was high during the very early stages of diversification but rapidly decreased  
370 through time and became negative *ca.* 25 Ma, implying a declining diversity (Fig. 3). Around

371 22 Ma, the net diversification rate became positive again and reached zero at the present. This  
372 model of partitioned dynamics of diversification outperformed any model involving a global  
373 driver of diversification. Indeed, the multi-rate time-dependent model better fit the  
374 diversification of *Morpho* (AICc=191.69) than the temperature-dependent model  
375 (AICc=197.3,  $\Delta$ AIC=5.61) and the altitude-dependent model (AICc=199.0,  $\Delta$ AIC=7.31).

376

### 377 ***Historical biogeography***

378 The model of biogeographic estimation with user-specified dispersal probabilities yielded a  
379 worse fit than the model with equal dispersal probabilities (likelihood with time-stratified  
380 dispersal multipliers:  $DEC_{strat}=-143.41$ ; likelihood without time-stratified dispersal  
381 multipliers  $DEC_{null}=-140.75$ ) and the ancestral state estimations involved some important  
382 differences. In both reconstructions the root state was highly unresolved. In the  $DEC_{null}$   
383 model (highest likelihood), the area with the highest probability at the root was the southern  
384 part of the Amazonian Basin, *ca.* 28.1 Ma. The early divergence of the clade containing *M.*  
385 *marcus* and *M. eugenia* was accompanied by a colonization of the northern part of the  
386 Amazonian Basin (Fig. 4). The ancestor of the remaining group of *Morpho* occupied the  
387 Central Andes. This lineage then diverged into an Andean and an Amazonian lineage. This  
388 event (21.8 Ma) was also accompanied by a shift in microhabitat use: flight in low forest  
389 strata (understory) for the Andean lineage, and flight high above ground up to the canopy for  
390 the Amazonian lineage. The Andean lineage began a long-term occupation of the Central  
391 Andes with local diversification (12 nodes inferred occupying the Central Andes after the  
392 initial dispersal event). Around 11-12 Ma, cis-Andean (east of the Andes) recolonizations of  
393 Amazonia and the Atlantic Forest happened in three lineages. *M. polyphemus* is an intriguing  
394 case as it diverged 20.8 Ma from an Andean ancestor, but nowadays occupies Central  
395 America, whose connection to South America is often considered to be only completed



396 during the last 4-3 million years. This implies either an earlier dispersal route of emerging  
397 Central America or a more recent dispersal with a joint extinction in the South American  
398 landmass. Overall, Northern Amazonia and the Northern Andes appear to have been  
399 colonized recently, during the last 5 million years (Fig. 4).

400

## 401 **Discussion**

402 In this study we aimed at investigating the large-scale patterns of diversification of the  
403 *Morpho* butterflies by jointly evaluating the dynamics of species and phenotypic  
404 diversification, to assess whether they are coupled or not and to test whether they correlate  
405 with clade-specific factors and/or biogeographic events. Our results show that ecological  
406 idiosyncrasies predominantly explain the pattern of diversification, instead of global (tree  
407 wide) factors. These ecological changes affected to a large extent both species and  
408 phenotypic diversification, leading to the partial coupling of both dynamics. Based on the  
409 amount of information currently available on the ecology of *Morpho* we discuss the potential  
410 role of several ecological and biogeographic events as well as the correlation with phenotypic  
411 diversification in explaining these variations among groups.

412

### 413 *Study limitations*

414 A number of limitations have to be mentioned before discussing our results. Focusing on a  
415 small clade allowed us to combine multiple ecological, morphological and historical  
416 components thereby providing a deep understanding of the *Morpho* history. Although we  
417 sampled all known species for both the molecular phylogeny and morphological traits, our  
418 comparative analyses probably lack power as a result of both the small number of taxa (30  
419 species) and the phylogenetic distribution of the traits of interest. Both microhabitat shift and  
420 host-plant shift (towards monocotyledons) are single events happening at the root of a single

421 clade each and we lack phylogenetically independent similar shifts. Typically, we found an  
422 evolutionary jump in wing size to be associated with a shift from dicotyledons to  
423 monocotyledons host-plants. Further work addressing this pattern at a larger phylogenetic  
424 scale will be necessary to assess the generality of our finding. Furthermore, the reliability of  
425 birth-death models to assess the diversification dynamics from phylogenies of extant taxa is  
426 debated (e.g. Quental & Marshall, 2010; Louca & Pennell, 2020). We thus remain cautious  
427 with our estimation of the diversification dynamics and the interpretation of the different  
428 models tested. In particular, we avoided interpreting the speciation and extinction rates  
429 independently to focus only on the net diversification dynamics. Finally, the timing and  
430 magnitude of the Andean surface uplift is also controversial (see for example Evenstar *et al.*,  
431 2015, and references therein; Fiorella *et al.*, 2015). We based our test on the reconstruction  
432 proposed by Leier *et al.* (2013) that focused only on the eastern cordillera of the Central  
433 Andes where the *Morpho* diversity is the highest, but had a large uncertainty in their paleo-  
434 altitude estimations. The Andean orogeny was spatially and temporally heterogeneous  
435 (Horton, 2018), which makes the use and interpretation of the paleoaltitude-dependent  
436 diversification model difficult (Condamine *et al.*, 2018). Those limitations should thus be  
437 kept in mind throughout the following discussion of the drivers of diversification, and the  
438 signal of declining diversity in particular.

439

#### 440 *Early Andean diversification not directly driven by Andean uplift*

441 The diversification of the genus *Morpho* in the Andes could have happened either  
442 simultaneously with the uplift – a scenario where speciation is driven by the increasing  
443 heterogeneity of ecological conditions with new altitudes (Lagomarsino *et al.*, 2016) – or  
444 decoupled from orogenesis – a scenario where a clade radiates across a range of altitudes  
445 already established through adaptations to ecological conditions (e.g. climate, host-plants,

446 predators). Our results support the second hypothesis. We found that a model of  
447 diversification rate responding to paleo-altitude performed worse than the clade-specific  
448 diversification models (Tables 2 and 3), which means that neither global speciation nor  
449 extinction rate variations are well explained by the paleo-altitudes of the Central Andes. From  
450 a biogeographic point of view, 16 extant species (over 30) are almost restricted to the  
451 lowlands, while only six extant species have a distribution strictly restricted to the Andes.  
452 Yet, from the Oligocene-Miocene boundary to middle Miocene periods (23.5 to 11.6 Ma), 11  
453 nodes out of 14 were inferred to be at least in the Central Andes from our biogeographic  
454 estimation (Fig. 4). Combined to the hypothesis that the *Morpho* probably originated in the  
455 foothills of the proto-Central-Andes, it is undeniable that the Central Andes played an  
456 important role in the early diversification of *Morpho*. During the second half of their  
457 evolutionary history, these lineages dispersed and diversified out of the Central Andes.

458 In contrast with the pattern of Central Andean diversification described above, the  
459 Northern Andes appear to have played only a minor role: while Northern Andean uplift likely  
460 established a barrier in three instances, resulting in cis- and trans-Andean *Morpho* lineages  
461 (Fig. 4), no major diversification was associated with the periods of Northern Andean uplift  
462 (Blandin & Purser, 2013). This absence of local diversification in the Northern Andes is a  
463 major difference compared to other butterflies such as the Ithomiini in which several groups  
464 repeatedly diversified at a high rate in the Northern Andes such as the genera *Napeogenes*  
465 (Elias *et al.*, 2009), *Oleria* (De-Silva *et al.*, 2016), *Hypomenitis* (Chazot *et al.*, 2016) or  
466 *Pteronymia* (De-Silva *et al.*, 2017).

467 Diversification driven by host-plant evolution may be an alternative explanation for  
468 the early diversification of *Morpho*. Penz and DeVries (2002) and Cassildé *et al.* (2010)  
469 suggested that monocotyledons were the ancestral host-plants of the genus *Morpho*, probably  
470 because at the time it was admitted that *M. marcus* larvae feed on monocotyledons

471 (Constantino, 1997). However, we now know that *M. marcus* very probably feeds on  
472 Fabaceae (e.g. *Inga auristellae*; Ramírez-García *et al.*, 2014; Vásquez Bardales *et al.*, 2017),  
473 and *M. eugenia* certainly feed on Caesalpiniaceae (Bénélu, 2016). Therefore, since groups  
474 closely related to *Morpho*, notably the sister genus *Caerois*, are known to only feed on  
475 monocotyledon host-plants (Beccaloni *et al.*, 2008), it is likely that the divergence of the  
476 *Morpho* was associated with an initial shift to dicotyledons. This host-plant shift at the root of  
477 Morphos created the conditions for an early rapid diversification of the group.

478

479 *A shift towards the canopy driving phenotypic and diversification changes*

480 We found a shift of species diversification associated with a single shift from the understory  
481 to the canopy (DeVries *et al.* 2010; Chazot *et al.* 2016). We also found strong indications that  
482 female wing shape evolution in the canopy clade is different from a neutral evolution. An  
483 increasing rate of shape evolution for both fore- and hind-wings was supported in the  
484 subclade nested in the canopy clade and including *M. theseus*, *M. niepelti*, *M. amphytrion*, *M.*  
485 *telemachus*, and *M. hercules*. Chazot *et al.* (2016) showed that both male and female wing  
486 shapes in the canopy clade are significantly different from wing shapes in understory species.  
487 Here we show that this microhabitat change associated with different vegetation structure,  
488 microclimatic conditions and predator community may have also affected the rate of female  
489 wing shape evolution in addition to shape *per se*. However, we note that the highest rate shift  
490 was not placed at the root of the canopy clade, suggesting that other factors may have caused  
491 this rapid phenotypic evolution. This increased rate of wing shape evolution was not found in  
492 males. Instead, in males we found two significant slowdowns in rate at different small  
493 subclades, only in the case of hindwings. The lack of more precise information on these  
494 species ecology unfortunately prevents speculating on the factors involved in such changes in  
495 wing shape evolutionary rate.

496

497 *A second change in microhabitat conditions associated with a host-plant, phenotypical and*  
498 *diversification shifts*

499 Published information in the *portis* clade (Heredia & Alvarez, 2007; Beccaloni *et al.*,  
500 2008; Montero Abril & Ortiz Perez, 2010) indicate that four *Morpho* species (*M. portis*, *M.*  
501 *aega*, *M. sulkowskyi* and *M. rhodopteron*) feed on Neotropical woody bamboos (Poaceae,  
502 tribe Bambuseae), notably on *Chusquea* species (subtribe Chusqueinae), in particular  
503 *Chusquea* aff. *scandens* for *M. sulkowskyi* that occurs at cloud forest elevations (Heredia &  
504 Alvarez, 2007). Recent observations indicate that *M. zephyritis* also feeds on woody  
505 bamboos (Roberto Maravi, pers. comm.). For the other species of the *portis* clade, there are  
506 only field observations indicating that they live in areas with important bamboo vegetation  
507 (Purser & Lacomme, 2016; pers. obs. in Peru, Daniel Lacomme pers. com.).

508 If, as observations indicate, the *portis* clade diversified after an initial shift back to  
509 monocotyledon host-plants, this reversal evolutionary event is a strong support for the  
510 “oscillation hypothesis” (Janz *et al.*, 2006). This hypothesis was proposed to explain the  
511 pattern of nymphalid butterflies with respect to host-plant use (Janz *et al.*, 2006) and states  
512 that the ability to recolonize “lost” hosts should be conserved over long evolutionary times,  
513 leading to recurrent recolonization events. Compared to the speciation rate of the backbone,  
514 species diversification within the *portis* clade proceeded at a higher rate, and rapidly  
515 decreased through time to reach almost zero at present. Adaptive radiations, here following a  
516 major host-plant shift, predict this rapid dampening of speciation rate as a result of niche  
517 filling (Schluter, 2000; Gavrilets & Losos, 2009).

518 Interestingly, we found that an evolutionary jump – a fast punctual event of evolution  
519 – toward smaller wing sizes also coincided with the host-plant shift. Chazot *et al.* (2016) did  
520 not identify any driver of this wing size evolution. To our knowledge, there is no clear

521 expectation or evidence supporting a specific relationship between body size and monocot  
522 *versus* dicot feeders but this question has rarely been addressed (but see Garcia-Barros 2000).  
523 The jump toward smaller sizes also cannot be associated with any altitudinal change because  
524 some species of the clade only occur at low to mid altitudes (200-1500 m), while others occur  
525 at higher altitudes (1500-3500 m) (Blandin, 2007; Gayman *et al.*, 2016).

526 Therefore, other hypotheses need to be explored, in particular that of a second  
527 possible change of microhabitat conditions. Many Bambusinae, in particular *Chusquea*  
528 species, form dense thickets, twigs and leaves creating inextricable tangles as a result from  
529 abundant vegetative branching at each node (Fisher, 2011; Fisher *et al.*, 2014). Observational  
530 data on the behaviour of the bamboo feeding *Morpho* is scarce, but observations on *M.*  
531 *rhodopteron* (Montero Abril and Ortiz Perez, 2010; Purser and Lacomme, 2016), *M.*  
532 *sulkowskyi* (Heredia and Alvarez-Lopez 2007), and *M. aega* (Otero & Marigo, 1990) suggest  
533 that females are more often resting inside the *Chusquea* thickets while males are flying  
534 around (males, when resting, also stand in the vegetation). Moreover, Heredia & Alvarez-  
535 Lopez (2007) noted that *M. sulkowskyi* females having light and dark alternating stripes on  
536 wings ventral side are difficult to detect inside *Chusquea* thickets. More or less contrasted  
537 similar patterns exist in males and females of other species, except in *M. absoloni*. Therefore,  
538 we hypothesize that size reduction, associated to a more or less striped appearance of the  
539 ventral side, could be an adaptation to the microhabitat structure of dense woody bamboo  
540 thickets, highlighting once again the importance of the microhabitat conditions on species  
541 and trait evolution.

542

#### 543 *Declining diversity in the Neotropical Morpho*

544 When accounting for heterogeneity in diversification rates (isolating the two shifting  
545 subclades), the diversification dynamics for the remaining lineages was characterized by a

546 negative net diversification rate, indicative of a declining diversity, mainly during the  
547 Miocene. Whether diversity decline can be accurately estimated only from phylogenies of  
548 extant species is a matter of debate (e.g. Quental & Marshall, 2010; but see Morlon *et al.*,  
549 2011). In the case of *Morpho*, this pattern may explain why some branches in the tree (such  
550 as the stem branch of *M. marcus* and *M. eugenia* or the branches leading to *M. anaxibia*, *M.*  
551 *deidamia*, or *M. polyphemus*) are surprisingly long. Extinct lineages may also explain why *M.*  
552 *polyphemus*, which diverged from its sister clade 20 Ma, is found in Central America, while  
553 colonization of Central America is often expected to be much more recent (but see Montes *et*  
554 *al.*, 2015, Farris *et al.*, 2011). Major landscape transformations during the Miocene in western  
555 Amazonia may explain this decline. Between 23-10 Ma, Western Amazonia transformed into  
556 a large wetland of lakes, swamps and shallow water, called the Pebas System (Wesselingh  
557 *et al.*, 2001; Hoorn *et al.*, 2010). The exact nature of the Pebas System is still under  
558 discussion but it was most likely unsuitable for terrestrial fauna (Salas-Gismondi *et al.*,  
559 2015). Evidence of extinction has been found from a west Amazonian fossil record, in  
560 particular with a major decrease of mammalian diversity at the transition between the  
561 Oligocene and the Miocene (Antoine *et al.*, 2016), which is in line with the beginning of the  
562 diversity decline in *Morpho* (Fig. 3).

563

#### 564 *Conclusion*

565 Our results support a prevailing ecological basis for both species and phenotypic  
566 diversification in *Morpho* butterflies: (1) a major host-plant shift, which punctually affected  
567 wing size evolution and greatly affected species diversification dynamics (pattern of adaptive  
568 radiation), and (2) a microhabitat shift affecting species diversification and partially wing  
569 shape diversification. Therefore, to a large extent, the dynamics of species diversification and  
570 phenotypic diversification are coupled in *Morpho*, most likely as a result of two major

571 ecological events. More importantly, we show that both species and phenotypic  
572 diversification in *Morpho* butterflies are better explained by multiple clade-specific factors  
573 instead of global abiotic drivers. Current methods for identifying drivers of diversification,  
574 based on model comparisons, are unable to test for potential interactions between drivers.  
575 Hence, our results do not exclude the possibility that the Andes played a role in  
576 diversification, but rather suggest that their effect on the shape of the phylogenetic tree was  
577 less significant than other factors. Nevertheless, the extent to which the effects of these  
578 ecological drivers can be generalised is unknown given the scale of our dataset. In particular  
579 future work at a larger phylogenetic scale should shed light on the importance of major host-  
580 plant transitions on the evolution of body size and the dynamics of diversification. Our study  
581 also highlights that both phenotypic and ecological information are of key relevance for  
582 understanding macroevolutionary patterns of diversification.

583

584



585 **Table 1.** Summary results obtained from fitting three models of trait evolution on 100 trees,  
586 using the *rjmcmbm* function as implemented in the R package GEIGER on a) males and b)  
587 females. *bm*=single Brownian rate, *rbm*=relaxed Brownian rates, *jbm*=jumps of Brownian  
588 rates. AIC<sub>bm</sub>, AIC<sub>rbm</sub>, AIC<sub>jbm</sub> = mean AIC score across the 100 trees for all three models.  
589  $\Delta$ AIC<sub>bm-rbm</sub>,  $\Delta$ AIC<sub>bm-jbm</sub>,  $\Delta$ AIC<sub>rbm-jbm</sub> = pairwise AIC differences between models for each  
590 tree.

591

592 **a) Males**

	AIC <sub>bm</sub>	AIC <sub>rbm</sub>	AIC <sub>jbm</sub>	$\Delta$ AIC <sub>bm-rbm</sub>	$\Delta$ AIC <sub>bm-jbm</sub>	$\Delta$ AIC <sub>rbm-jbm</sub>
Forewing	-23.92 (18.72)	-27.78 (10.33)	13.41 (26.32)	3.85 (21.70)	-37.34 (34.54)	-41.20 (30.02)
Hindwing	-14.43 (20.28)	-18.62 (14.58)	14.94 (24.28)	4.18 (24.02)	-29.37 (29.76)	-33.56 (28.21)

593

594 **b) Females**

	AIC <sub>bm</sub>	AIC <sub>rbm</sub>	AIC <sub>jbm</sub>	$\Delta$ AIC <sub>bm-rbm</sub>	$\Delta$ AIC <sub>bm-jbm</sub>	$\Delta$ AIC <sub>rbm-jbm</sub>
Forewing	-22.36 (10.44)	-17.74 (9.40)	-34.70 (1.60)	-4.61 (15.73)	12.33 (10.98)	16.95 (9.56)
Hindwing	-15.44 (17.06)	-13.30 (22.83)	-32.80 (1.17)	-2.14 (29.62)	17.36 (16.88)	19.50 (23.01)

595 **Table 2.** Paleoenvironmental-dependent diversification analyses using paleoaltitude (a) and  
 596 Cenozoic temperature (b) data. Mean parameter and standard error estimates are presented  
 597 for each model. Best-fitting model, as determined via a combination of the lowest AIC and  
 598  $\Delta$ AIC (see main text) highlighted in bold. In our best-fit paleoaltitude-dependent model,  
 599 speciation is negatively correlated to Andean orogeny over time (adding extinction as a  
 600 parameter did not improve the model fit). Likewise, speciation is positively correlated to  
 601 temperature variation over time (allowing extinction to vary with temperature did not  
 602 improve the likelihood).  $\lambda$  = speciation rate (in events/Myr/lineage);  $\mu$  = extinction rate (in  
 603 events/Myr/lineage);  $\alpha$  = rate of variation of the speciation according to the relevant  
 604 paleoenvironmental variable;  $\beta$  = rate of variation of the extinction according to the  
 605 paleoenvironmental variable; NP = number of parameters in each model.

606  
 607  
 608  
 609

**a) Paleoaltitude models**

Models	Dependency	NP	logL	AIC	$\Delta$ AIC	$\lambda$	$\alpha$	$\mu$	$\beta$
$\lambda$ Alti. and no $\mu$	Linear	2	-97.50 $\pm 0.085$	199.00 $\pm 0.171$	0.00	0.190 $\pm 0.004$	-3.60E-05 $\pm 1.39E-06$	-	-
$\lambda$ Alti. and no $\mu$	Exponential	2	-97.68 $\pm 0.083$	199.37 $\pm 0.166$	0.37	0.210 $\pm 0.004$	-3.06E-04 $\pm 5.76E-06$	-	-
$\lambda$ Alti. and $\mu$ constant.	Linear	3	-97.49 $\pm 0.085$	200.99 $\pm 0.170$	1.99	0.190 $\pm 0.0003$	-3.47E-05 $\pm 8.75E-07$	5.15E-04 $\pm 4.17E-04$	-
$\lambda$ Alti. and $\mu$ constant	Exponential	3	-97.69 $\pm 0.083$	201.37 $\pm 0.166$	2.37	0.220 $\pm 0.004$	-3.11E-04 $\pm 5.68E-06$	9.09E-07 $\pm 7.91E-07$	-
$\lambda$ constant and $\mu$ Alti	Exponential	3	-97.75 $\pm 0.104$	201.51 $\pm 0.207$	2.51	0.080 $\pm 3.16E-04$	-	42038.92 $\pm 11873.33$	-7.48E-03 $\pm 2.62E-04$
$\lambda$ Alti. and $\mu$ Alti	Exponential	4	-96.94 $\pm 0.094$	201.87 $\pm 0.187$	2.87	0.880 $\pm 0.160$	-5.00E-04 $\pm 2.00E-05$	6.820 $\pm 1.159$	-1.89E-03 $\pm 4.72E-05$
$\lambda$ Alti. and $\mu$ Alti.	Linear	4	-97.42 $\pm 0.086$	202.83 $\pm 0.171$	3.84	0.210 $\pm 0.004$	-3.83E-05 $\pm 1.18E-06$	0.070 $\pm 0.007$	-2.38E-05 $\pm 2.32E-06$
$\lambda$ constant and $\mu$ Alti.	Linear	3	-98.56 $\pm 0.099$	203.12 $\pm 0.198$	4.12	0.080 $\pm 0.001$	-	0.030 $\pm 0.005$	-1.14E-05 $\pm 1.76E-06$

610

611  
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613

**b) Paleoclimate models**

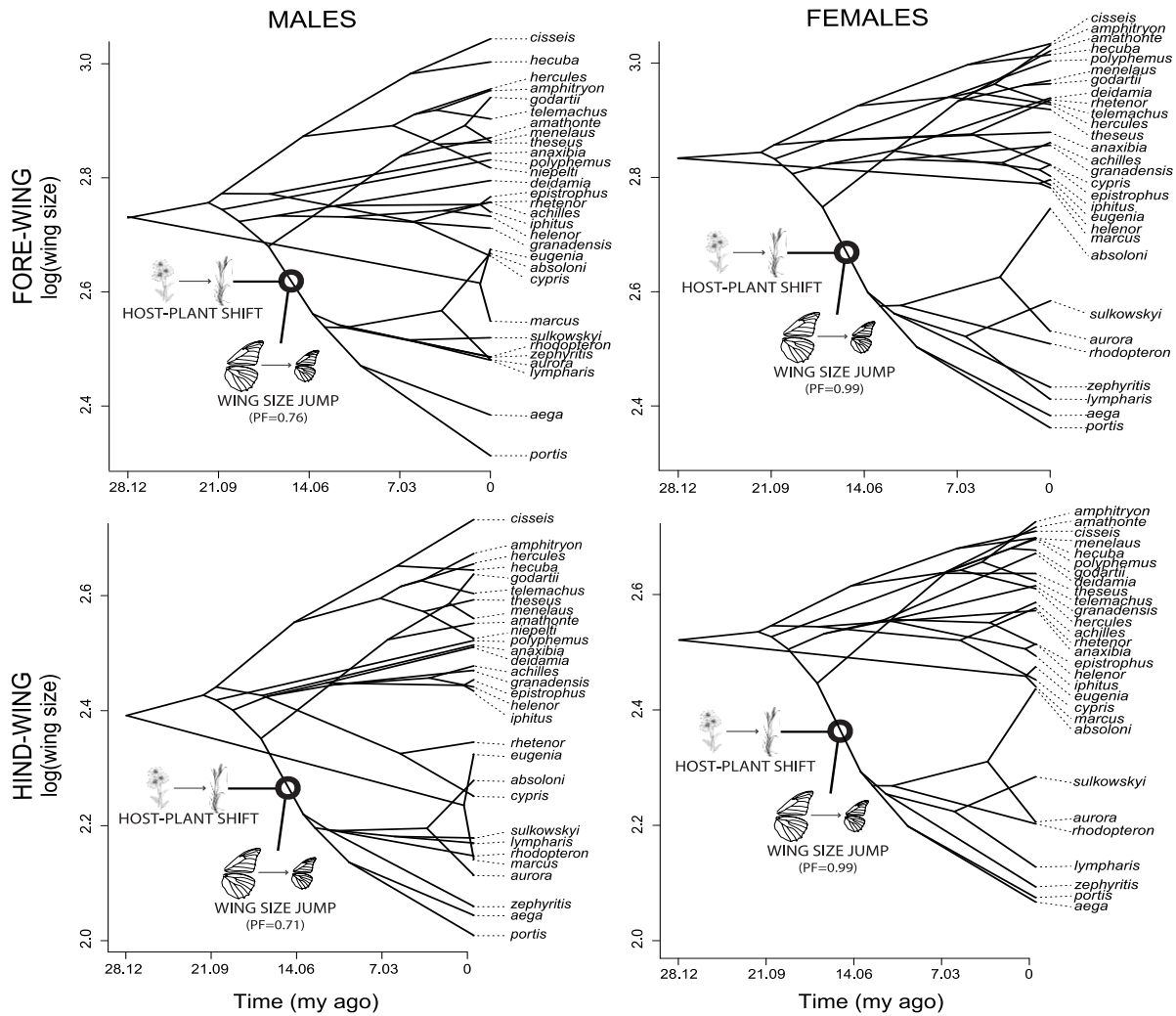
Models	Dependency	NP	logL	AIC	$\Delta$ AIC	$\lambda$	$\alpha$	$\mu$	$\beta$
$\lambda$ Temp. and no $\mu$	Exponential	2	-96.65 $\pm 0.095$	197.30 $\pm 0.189$	0.00	0.030 $\pm 4.13E-04$	0.179 $\pm 1.91E-03$	-	-
$\lambda$ Temp. and no $\mu$	Linear	2	-96.72 $\pm 0.097$	197.44 $\pm 0.195$	0.13	0.015 $\pm 5.50E-04$	0.013 $\pm 1.17E-04$	-	-
$\lambda$ Temp. and $\mu$ constant	Exponential	3	-96.58 $\pm 0.092$	199.15 $\pm 0.184$	1.85	0.030 $\pm 3.93E-04$	0.210 $\pm 3.62E-03$	0.043 $\pm 0.004$	-
$\lambda$ Temp. and $\mu$ constant	Linear	3	-96.67 $\pm 0.097$	199.33 $\pm 0.195$	2.03	0.023 $\pm 1.48E-03$	0.021 $\pm 1.34E-03$	0.046 $\pm 0.007$	-
$\lambda$ Temp. and $\mu$ Temp.	Linear	4	-96.42 $\pm 0.095$	200.85 $\pm 0.189$	3.54	0.017 $\pm 1.11E-03$	0.027 $\pm 8.07E-04$	0.290 $\pm 0.012$	-0.037 $\pm 0.002$
$\lambda$ Temp. and $\mu$ Temp.	Exponential	4	-96.55 $\pm 0.092$	201.10 $\pm 0.183$	3.80	0.032 $\pm 7.99E-04$	0.205 $\pm 4.57E-03$	0.156 $\pm 0.040$	-0.164 $\pm 0.019$
$\lambda$ constant and $\mu$ Temp.	Exponential	3	-98.57 $\pm 0.103$	203.14 $\pm 0.205$	5.84	0.081 $\pm 2.96E-04$	-	4.47E-08 $\pm 3.08E-09$	0.004 $\pm 2.19E-04$
$\lambda$ constant and $\mu$ Temp.	Linear	3	-98.57 $\pm 0.103$	203.14 $\pm 0.205$	5.84	0.081 $\pm 2.97E-04$	-	1.09E-04 $\pm 1.08E-04$	-1.37E-05 $\pm 1.37E-05$

614

615 **Table 3.** Results of model comparison for the five time-dependent diversification analyses  
 616 presented, with mean parameter estimates for each model.  $\lambda$  = speciation rate (in  
 617 events/Myr/lineage);  $\alpha$  = parameter of rate variation for speciation;  $\mu$  = extinction rate (in  
 618 events/Myr/lineage);  $\beta$  = parameter of rate variation for extinction; NP = number of  
 619 parameters in each model; AICc = corrected Akaike information criterion; logL = log-  
 620 likelihood.

621

Clade partition	Models	NP	logL	AIC	$\lambda$	$\alpha$	$\mu$	$\beta$	Joint logL	Joint AIC
background	BVAR DVAR	4	-35.68	79.36	0.063	0.237	0.079	0.228		
monocots	BVAR	2	-21.78	47.55	0.014	0.213	-	-	-88.84	191.69
canopy	BCST	1	-31.39	64.77	0.083	-	-	-		
background	BCST	1	-50.91	103.83	0.072	-	-	-		
monocots	BVAR	2	-21.78	47.55	0.014	0.213	-	-	-92.83	193.66
shape shift	BCST	1	-20.14	42.28	0.095	-	-	-		
background	BCST	1	-73.75	149.49	0.081	-	-	-	-95.52	197.05
monocots	BVAR	2	-21.78	47.55	0.014	0.213	-	-		
whole	BCST	1	-98.40	198.81	0.081	-	-	-	-98.40	198.81
background	BCST	1	-78.18	158.36	0.078	-	-	-	-98.32	200.63
shape shift	BCST	1	-20.14	42.28	0.095	-	-	-		
background	BCST	1	-67.01	136.03	0.080	-	-	-	-98.40	200.80
canopy	BCST	1	-31.39	64.77	0.083	-	-	-		



622

623

**Figure 1.** Phenograms for wing size (log scale) for males (left panels) and females (right

624

panels). The top panels are the forewings, and the bottom panels are the hindwings. Wing

625

size values are reconstructed at the nodes and plotted on a time scale. Phylogenetic

626

relationships are projected into the phenogram. The position (branch) where the main host-

627

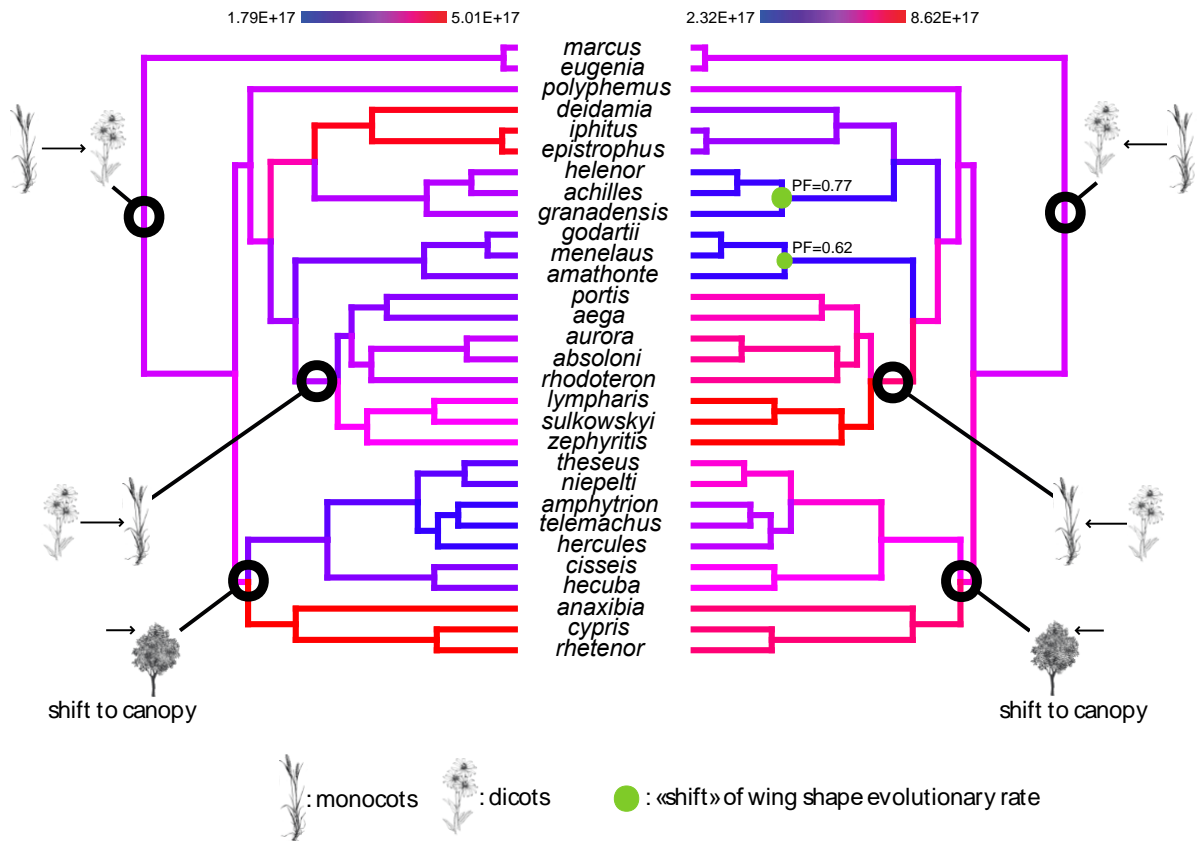
plant shift and significant wing size jump happened is also shown. PF values indicate the

628

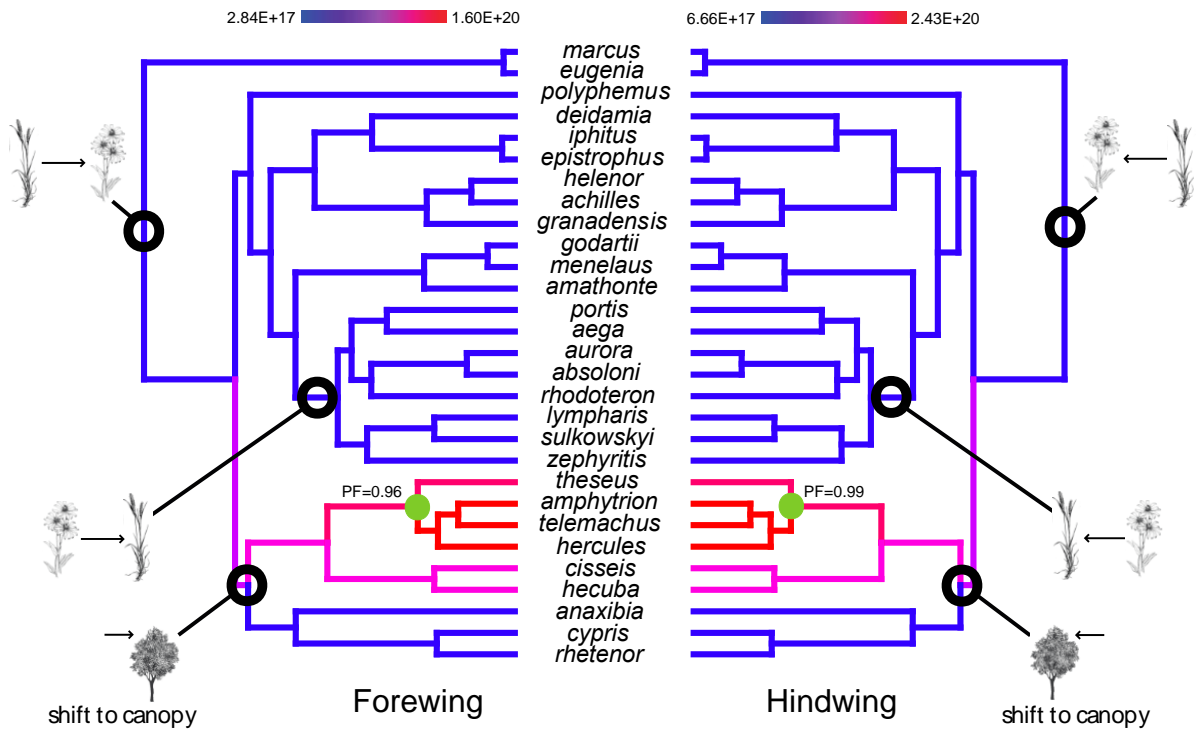
frequency at which each jump was found across the posterior distribution of trees.

629

a) Males

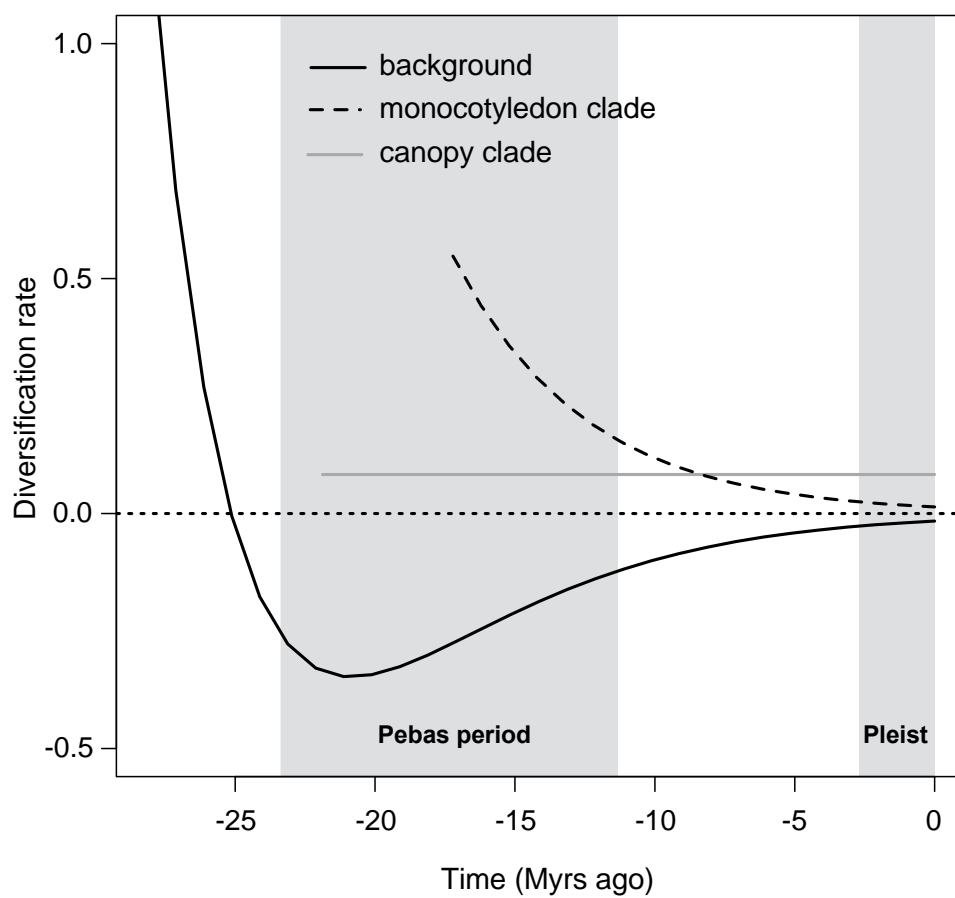


b) Females



631 **Figure 2.** Rate of wing shape diversification for a) males and b) females. Branches of the  
632 phylogenies are coloured according to the evolutionary rate inferred at the nodes using the R  
633 package GEOMORPH. Green points indicate the changes in the rate of wing shape evolution  
634 and black points the evolutionary jumps of wing size. Only shifts with a posterior tree  
635 frequency higher than 0.5 are shown. PF values indicate the frequency at which each shift  
636 was found across the posterior distribution of trees. On these phylogenies some major  
637 evolutionary events including important host-plant shifts and microhabitat shifts are also  
638 indicated.

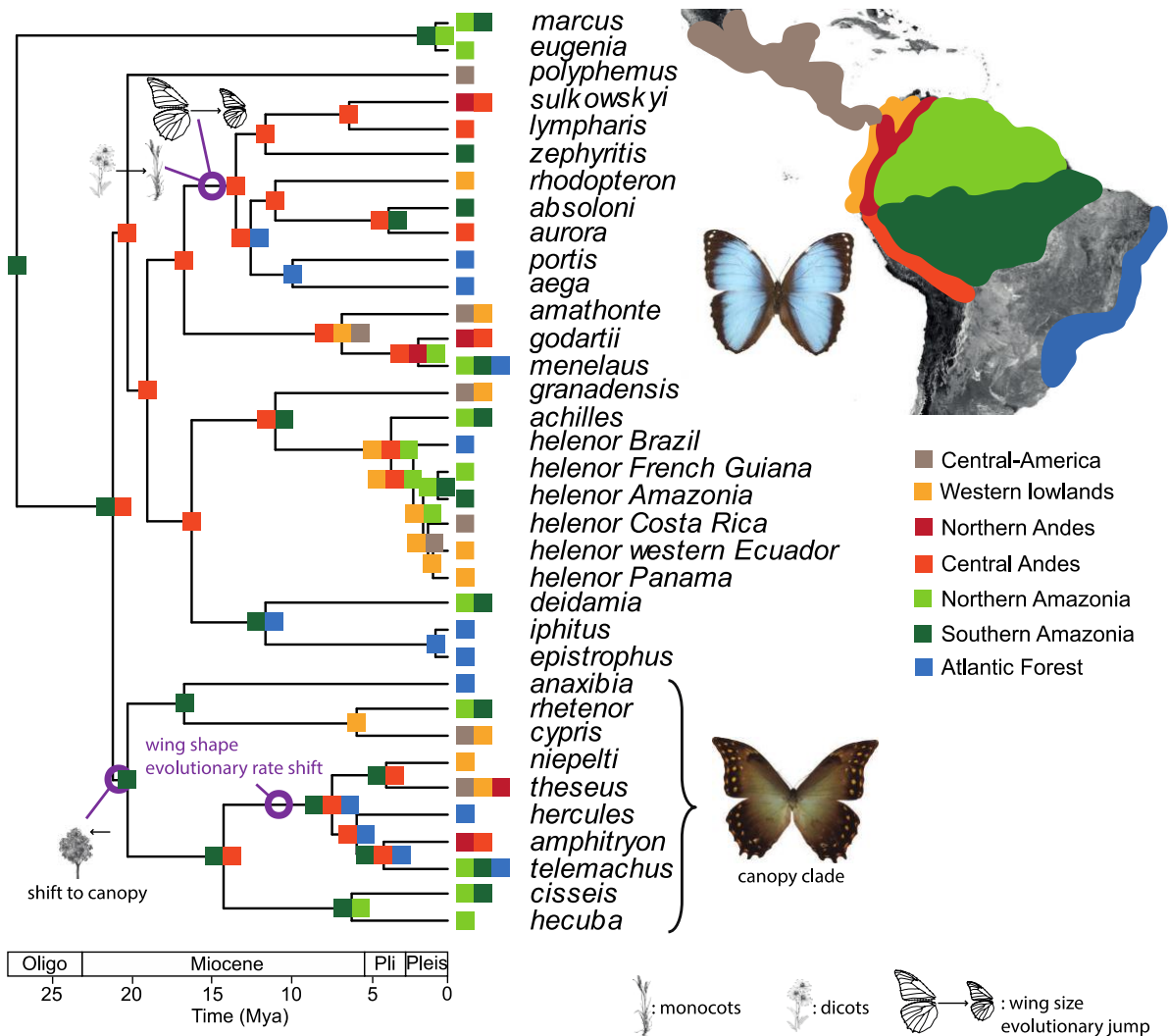
639



640



641 **Figure 3.** Estimation of the temporal dynamics of diversification for the genus *Morpho*.  
642 Diversification rates (speciation minus extinction) for the best models identified for the  
643 different subclades (canopy and monocotyledon) and the remaining lineages (background).  
644 The early background diversification is elevated and decreases through time until it becomes  
645 negative in the early Miocene. The canopy clade has constant rates of diversification, while  
646 the monocotyledon clade conforms to an early-burst pattern with high rates that decrease  
647 toward the present.  
648



649

650 **Figure 4.** Historical biogeography inferred for the genus *Morpho*. The most likely states are

651 indicated at the nodes. The different clade-specific ecological factors are also indicated on the

652 tree. The two pictures of *Morpho* depict the typical wing shapes associated with each

653 microhabitat – top: short rounded wings characteristic of the understory species, bottom:

654 elongated wings toward the apex characteristic of the canopy clade.

655

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