1	Testing the Tea Bag Index as a potential indicator for assessing litter decomposition in aquatic
2	ecosystems
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15	Abstract
16	The Tea Bag Index (TBI) approach is a standardized method for assessing litter decomposition in
17	terrestrial ecosystems. This method allows determination of the stabilized portion of the hydrolysable
18	fraction during the decomposition process, and derivation of a decomposition constant (k) using single
19	measurements of the mass-loss ratios of green and rooibos teas. Although this method is being applied
20	to aquatic systems, it has not been validated in these environments, where initial leaching tends to be
21	higher than in terrestrial ecosystems. Here, we first validated a critical assumption of the TBI method
22	that green tea decomposition plateaus during the standard incubation period of 90 days, and then tested
23	the accuracy of a TBI-based asymptote model using a second model obtained from fitting actual
24	decomposition data. Validation data were obtained by incubating tea bags in water samples taken from
25	a stream, a pond, and the ocean in Kumamoto, Japan. We found that green tea decomposition did not
26	plateau during the 90-day period, contradicting a key assumption of the TBI method. Moreover, the
27	TBI-based asymptote models disagreed with actual decomposition data. Subtracting the leachable
28	fraction from the initial tea mass improved the TBI-based model, but discrepancies with the actual
29	decomposition data remained. Thus, we conclude that the TBI approach, which was developed for a
30	terrestrial environment, is not appropriate for aquatic ecosystems. However, the use of tea bags as a
31	standard material in assessments of aquatic litter decomposition remains beneficial.
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33	Keywords: aquatic environment; asymptote model; decomposition; decomposition constant k;
34	stabilization factor S; Tea Bag Index
35	
36	Introduction

Litter decomposition is an important process in aquatic systems (Graça et al., 2015), playing a major role in the global cycling of carbon and nutrients (Battin et al., 2009; Gessner et al., 1999; Graça et al., 2015). Three main factors control aquatic litter decomposition: litter quality (Jabiol et al., 2019; Neiff et al., 2006); environmental factors such as temperature, nutrient availability, salinity, acidity, and oxygen concentration (Almeida Júnior et al., 2020; Ferreira et al., 2015; Gomes et al., 2018; Griffiths and Tiegs, 2016; Woodward et al., 2012; Young et al., 2008); and decomposer community composition, including macroinvertebrates, fungi, and bacteria (Balibrea et al., 2020; Hieber and Gessner, 2002).

Litter bags filled with litter from the local area are often used to assess litter decomposition rates in aquatic systems. However, understanding the impacts of environmental changes on litter decomposition rates at large geographical scales requires a standardized method (Keuskamp et al., 2013; Mori et al., 2021a), because high local variation in litter quality may negate the influence of environmental effects. A standardized method applicable to both aquatic and terrestrial ecosystems would be beneficial (Seelen et al., 2019), because integrated models of both terrestrial and aquatic systems remain poor (García-Palacios et al., 2016).

51 The Tea Bag Index (TBI) approach proposed by Keuskamp et al. (2013) is a standardized 52 method for assessing litter decomposition that was developed in a terrestrial ecosystem. This method 53 uses two types of commercially available tea bags (green and rooibos teas, Lipton) as standard 54 materials to calculate the TBI, which consists of two parameters: a stabilization factor S (the stabilized 55 portion of the hydrolysable fraction during decomposition) and the decomposition constant k of an 56 asymptote model (Keuskamp et al., 2013). Due to its cost-effectiveness and ability to collect 57 comparable globally distributed data (Keuskamp et al., 2013), multiple studies have used the TBI 58 (Becker and Kuzyakov, 2018; Fanin et al., 2020; Fujii et al., 2017; Mori et al., 2021b; Mueller et al., 59 2018; Petraglia et al., 2019).

60 Calculation of the TBI rests on several assumptions. S is calculated as the ratio between the 61 mass of hydrolysable fraction of green tea remaining at the end of the 90-day standard incubation 62 period and the entire hydrolysable fraction of green tea (0.842; Keuskamp et al. 2013); this assumes 63 that the decomposition of the acid-insoluble fraction of green tea is negligible, and that the 64 undecomposed hydrolysable fraction stabilizes and transforms to a recalcitrant fraction within the 65 incubation period. Thus, the decomposition curve of green tea must reach a plateau before the end of 66 the incubation period, given that the decomposition and stabilization of the hydrolysable fraction 67 should be completed within this period. Next, the decomposition constant (k) is determined by fitting 68 an asymptote model to rooibos tea decomposition data using the following formula:

69
$$W_r(t) = a_r * e^{-kt} + (1 - a_r)$$

eq. 1

where $W_r(t)$ is the ratio of the remaining mass of rooibos tea after the incubation period *t* relative to the initial mass, *k* is a decomposition constant, a_r is the decomposable fraction, and *l*- a_r is the undecomposable fraction of rooibos tea (Keuskamp et al., 2013). The TBI approach allows

73 parameterization of the asymptote model without fitting the model to time series data of rooibos tea 74 decomposition by determining a_r using green tea decomposition data. Because the TBI approach 75 assumes that the acid-insoluble fraction does not degrade within a 90-day period, a_r represents the 76 decomposable hydrolysable fraction (1 – acid-insoluble fraction) of rooibos tea. Under the assumption 77 that the ratio of the decomposable hydrolysable fraction to the total hydrolysable fraction of rooibos 78 tea is the same as that of green tea, a_r can be calculated as the total hydrolysable fraction (0.552; 79 Keuskamp et al. 2013) multiplied by (1 - S). The decomposition constant k can be determined by 80 substituting the values obtained from a single measurement of rooibos tea decomposition for the 81 parameters in eq. 1 (i.e., t is 90 and $W_r(t)$ is the remaining-mass ratio of rooibos tea at the end of the 82 incubation period).

83 Although the TBI method was developed in a terrestrial ecosystem, it has increasingly been 84 adopted in aquatic environments (Hunter et al., 2019), including mangrove systems (Mueller et al., 85 2018), marshes (Mueller et al., 2018; Yousefi Lalimi et al., 2018), and streams (Peralta-Maraver et al., 86 2019). In these studies, the TBI was calculated following the protocol of Keuskamp et al. (2013), 87 which assumes that the index can be applied equivalently in terrestrial and aquatic ecosystems. 88 However, the initial decomposition phase, i.e., leaching, is much more rapid in aquatic environments 89 (Webster and Benfield, 1986), which could cause the TBI-based asymptote model to deviate from real-90 world decomposition (Seelen et al., 2019). In addition, the assumption that green tea decomposition 91 plateaus during the incubation period has not been verified in an aquatic ecosystem (Keuskamp et al., 92 2013).

We first aimed to validate whether green tea decomposition does plateau during the 90-day incubation period, and then tested the accuracy of a TBI-based asymptote model by comparison with a model derived from actual decomposition data. Seelen et al. (2019) suggested that correcting the initial tea weights by subtracting the easily leachable fraction may provide a more accurate TBI in aquatic systems. However, the validity of this corrected approach was not evaluated using actual data. Therefore, we also evaluated a TBI-based model corrected by subtraction of the easily leachable fraction.

100

101 Materials and methods

102 Incubation experiment

We evaluated application of the TBI approach to aquatic ecosystems by monitoring the mass-loss ratios of green tea bags submerged in water. A microcosm approach (Santschi et al., 2018), rather than an *in-situ* approach, was used to control environmental variability as much as possible. We collected water samples for our experiment from three different sources: a stream running through an evergreen conifer plantation dominated by *Cryptomeria japonica* (Linnaeus f.) D. Don and *Chamaecyparis obtusa* (Sieb. et Zucc.) Endl. in Yamaga City, an artificial pond located at the Forestry and Forest

Products Research Institute (FFPRI) in Kyushu, and the ocean in Uki City. All three sites are located 110 in Kumamoto Prefecture, Japan. Because the purpose of our study did not include investigating factors 111 leading to differences in decomposition rates among the water samples, we did not perform chemical 112 analyses of the water samples. Each of the three water samples was placed into eight plastic bottles 113 (6.5 cm in diameter); four were assigned to green tea (EAN: 87 22700 05552 5, non-woven mesh, 114 Lipton) and four to rooibos tea (EAN: 87 22700 18843 8, non-woven mesh, Lipton). We placed 200 115 mL of water in each bottle and submerged five tea bags therein. We then covered the bottles with a 116 polyethylene sheet to prevent evaporation (Mori et al., 2013). The bottles were incubated at 25°C in 117 the dark. One tea bag per replicate was retrieved at 3, 11, 27, 55, and 91 days after the start of the incubation period, whereupon weights were determined by oven drying at 70°C for 72 h. 118 119 120 Calculating TBI 121 The TBI was calculated following Keuskamp et al. (2013). S was calculated as: 122 $S = 1 - a_g / H_g$ eq. 2 123 where a_g is the mass loss of green tea during the 90-day period (note that we used 91 days) and H_g 124 (0.842) is the hydrolysable fraction of green tea (Keuskamp et al. 2013). Assuming that the S value of 125 rooibos tea is the same as that of green tea, the decomposable fraction of rooibos tea was calculated 126 as:

127 $a_r = H_r * (1 - S)$ eq. 3

128 where H_r (0.552) is the hydrolysable fraction of rooibos tea (Keuskamp et al. 2013). The 129 decomposition constant (k) was determined from an asymptote model of rooibos tea decomposition: $W_r(t) / W_r(0) = a_r * e^{-kt} + (1 - a_r)$ 130 eq. 1'

131 where $W_r(t)$ and $W_r(0)$ are the mass of rooibos tea remaining after the incubation time and that before 132 the incubation time t, respectively.

133

109

134 Leaching factor correction

135 Seelen et al. (2019) proposed correction of the TBI using a leaching factor, which was defined as the 136 ratio of the easily leachable fraction to the total tea weight. Initial tea weight can then be corrected by 137 multiplying the initial tea mass by (1 - the leaching factor). Accordingly, all remaining mass values 138 and hydrolysable fractions of both the green and rooibos teas (H_g and H_r) were corrected by subtracting 139 the leaching factor and dividing the result by (1 - the leaching factor). The TBI and the TBI-based 140 asymptote model were then calculated using the corrected data. 141 Seelen et al. (2019) suggested that a leaching factor be determined using mass loss data of

142 both green and rooibos teas after submersion in water for 3 h. We chose to calculate a corrected TBI 143 using several scenarios with different combinations of leaching factors for green and rooibos teas, 144 rather than experimentally determining leaching loss in the manner suggested. We chose this approach

145 because it has been reported that leaching can occur for up to 72 and 48 hours in green and rooibos 146 teas, respectively (Edwartz, 2018), and a long-term leaching experiment could overestimate the 147 leaching factor due to microbial decomposition. In addition, solute concentration may influence initial 148 leaching losses in tea, leading to differences between ecosystems and potential interactions with the 149 teas' initial chemical composition. If the TBI is to be corrected by a leaching factor, then a global 150 leaching factor would be most appropriate. Our approach therefore represents the first step toward a 151 global leaching factor applicable to all aquatic environments. We created multiple scenarios, with a 152 maximum leaching factor based on the mass loss of the teas on day 3 of incubation, assuming that 153 leaching would be complete within 3 days and that the minimum leaching factor is 0, and intermediate 154 values between the maximum leaching factor and zero, in our calculations of a corrected TBI. 155 Asymptote models were constructed using the corrected TBI. We note that if a leaching factor is 156 applied in assessments of aquatic litter decomposition, the results will not be comparable to those in 157 terrestrial systems due to inherent differences in initial chemical compositions.

158

159 Leaching experiments

160 We assessed tea leaching in deionized water and a tea solution to understand the influence of solute 161 concentration on leaching. We used a tea solution rather than a salt solution to avoid any potential 162 chemical reactions that could influence the leaching process. Solutions of both green and rooibos teas 163 were prepared by submerging 10 tea bags in 600 mL of deionized water overnight. The deionized 164 water and tea solutions were autoclaved at 120°C for 20 min for sterilization purposes. The autoclaved 165 solutions were then placed in an incubator overnight at 25°C. Green and rooibos tea bags were then 166 submerged into either the deionized water or tea solution, where green and rooibos tea bags were 167 submerged in green and rooibos solutions, respectively, for 20 min or 9 h. The dry weights of the tea 168 bags were then determined following oven drying at 70°C for 72 h.

169

170 TBI-based asymptote model in terrestrial systems

As a reference, we calculated a decomposition curve based on a TBI-based asymptote model in a terrestrial ecosystem using published data (Keuskamp et al., 2013). These data, representing the outcomes of a laboratory incubation study conducted at 25°C, were obtained from Keuskamp et al. (2013) using the Data Thief 3.0 program (Tummers, 2006). We compared the TBI-based model to an asymptote model (eq. 1) that had been fitted to the time series data. Given that Keuskamp et al. (2013) did not collect remaining-mass observations on day 90 of incubation, the remaining mass of both green and rooibos teas at 90 days was calculated using the fitted asymptote model with a *t* of 90.

178

179 *Statistical analyses*

180 All statistical analyses were performed using R software (R Core Team, 2019). The asymptote model

181 was fitted using nonlinear regression. Additionally, we fitted a double exponential model to the 182 experimental data obtained from the stream- and pond-water samples using nonlinear regression. The 183 model was expressed as:

184 $W(t) = a * e^{-kt} + b * e^{-k2t}$ eq. 4

185 where W(t) is the mass remaining after t, k1 and k2 are decomposition constants, and a and b represent 186 organic matter fractions differing in decomposability. A one-way ANOVA followed by Tukey's post 187 hoc test was used to compare the remaining-mass ratios of the teas among the different water samples. 188 Shapiro-Wilk tests showed that the data were not significantly different from a normal distribution, 189 and log-transformation did not improve data fit; therefore, a normal distribution was assumed. The 190 leaching losses of the two teas were analyzed using a two-way (time vs. solution) ANOVA. Data were 191 log-transformed prior to this analysis.

192

193 **Results and Discussion**

194 Effects of solute concentration on the teas' leaching losses

195 We found that solute concentration, as well as soaking time, affected the leaching loss of both the 196 green and rooibos teas (Fig. 1). The leaching loss of both teas was lower in tea solution compared to 197 deionized water, and increased with increasing soaking time (Fig. 1a). This indicates that any leaching 198 factor determined by submerging tea bags for several hours will be affected by solution concentration, 199 including parameters such as dissolved organic carbon and salt concentrations. Therefore, it is difficult 200 to determine an adequate leaching factor for the TBI method using a leaching experiment. We 201 confirmed that our approach to determining leaching factors was appropriate, i.e., the use of multiple 202 scenarios with different combinations, because leaching factors determined following several hours of 203 submersion will likely change depending on solute concentration.

204

205 Evaluating the stabilization factor in aquatic environments

206 The remaining-mass ratios of the green tea samples at the end of the incubation period ranged from 207 0.3 to 0.6 (Fig. 2a-c), values within the range of those reported from aquatic environments (Peralta-208 Maraver et al., 2019; Seelen et al., 2019; Lalimi et al., 2018). The asymptote model did not fit the 209 time-series green tea decomposition data well, because green tea decomposition did not plateau during 210 the incubation period, at least not in the stream and pond water samples (Fig. 2a, b). Rather, green tea 211 mass continued to decline until the end of the incubation period (Fig. 2). This result disagrees with 212 those from a terrestrial system, where decomposition was found to plateau within the same period (Fig. 213 3a, Keuskamp et al. 2013). The continuous decline in mass therefore violates the plateau assumption 214 of the TBI. This continuous decline was likely not due to slower overall decomposition rates in aquatic 215 systems. In fact, the mass loss ratios of green tea in both the stream and pond samples were higher 216 than those reported from terrestrial ecosystems (Djukic et al., 2018). We did not aim to investigate the

potential causes of the differences in green tea decomposition curves between aquatic and terrestrial systems, and thus cannot speculate further on this point. However, we clearly demonstrated that the standard protocol for the TBI method is not suitable for determining *S* in aquatic environments. The inclusion of a leaching factor provides a negligible improvement in model fit, because it does not eliminate the issue of a decreasing trend in green tea mass loss (Fig. S1).

222

223 Evaluating the decomposition constant and TBI-based asymptote models in aquatic environments

224 We found that k values determined using the TBI approach (0.0081, 0.013, and 0.0088 in stream, pond, 225 and ocean samples, respectively) were at least one order of magnitude lower than those determined by 226 fitting actual decomposition data (0.58, 0.33, and 0.72 in stream, pond, and ocean samples, 227 respectively), indicating substantial disagreements between the TBI-based asymptote models and the 228 models constructed using actual data (Fig. 2d-f). These results contrast with those obtained in a 229 terrestrial ecosystem (Fig. 3b, Keuskamp et al. 2013), where these two models produced nearly 230 identical results. We suggest that this discrepancy is the product of a violation of the assumption that 231 the ratio of the decomposable to the total hydrolysable fraction is the same for rooibos and green tea. 232 The decomposable hydrolysable fraction (a_r) values determined by the TBI approach (0.46, 0.40, and 233 0.29 in stream, pond, and ocean samples, respectively) were much larger than those obtained from the 234 actual data-based model (0.21, 0.24, and 0.13 in stream, pond, and ocean samples, respectively). The 235 fact that green tea decomposition did not reach a plateau, such that S may have been overestimated, 236 was not the reason for the discrepancy, because the overestimation of S should cause an 237 underestimation of a_r (see eq. 2). Therefore, compensating for any overestimation of S would widen 238 the differences between the TBI-based model and actual-data-based model. Subtracting the leachable 239 fraction from the initial tea mass, as proposed by Seelen et al. (2019), improved the TBI-based 240 asymptote model (Fig. 4). A larger leaching factor reduced the discrepancy between the TBI-based 241 model and actual-data-based model for rooibos tea (Fig. 4). However, excluding some cases with 242 ocean water samples, wherein the leaching factors were 80% and 100% of the maximum (Fig. 4k, l), 243 the discrepancy remained. Therefore, the TBI approach may overestimate the hydrolysable fraction of 244 rooibos tea, leading to an underestimation of k in asymptote models. We note that the asymptote model 245 did not fit the real-world data well, and the discrepancy between the two model types may therefore 246 have been overestimated. However, considering that we could not determine S, and that under- or over 247 estimation of S leads to incorrect estimation of k (Mori et al., 2021b), we concluded that the standard 248 protocol used for the TBI method, which was developed in a terrestrial ecosystem, is not applicable to 249 aquatic ecosystems.

250

251 Future prospects for the use of the TBI method in aquatic environments

252 Although the TBI may be an unsuitable index in aquatic ecosystems, this does not necessarily negate

253 the benefits of using tea bags as a standard material in assessments of litter decomposition rates in 254 aquatic systems. For example, the remaining raw mass of both tea types could provide information on 255 decomposition rates that is comparable on a global scale (Djukic et al., 2018). We found that the 256 remaining mass of both teas following 91 days of incubation reflected the differences among the three 257 water sample types used in this study (Fig. 5). This indicates that tea bags are sensitive enough to 258 reflect differences in chemical composition and microbial community, both of which control litter 259 decomposition (temperature was controlled in our study). Moreover, patterns in the remaining-mass 260ratios among the water sample types differed between the two tea types. The ratio was lowest in stream 261 water for green tea and lowest in pond water for rooibos tea, although it was highest in ocean water 262 for both types, likely because the high salt concentration inhibited decomposition (Contreras et al., 263 2017). Combining these two tea types could be advantageous for detecting differences in 264 decomposition processes among different aquatic ecosystems.

265 In addition, tea bags could still be used as standard materials to obtain time-series data and 266 fit models in assessments of decomposition rates. In this scenario, an alternative model type could also 267 be selected, as the asymptote model may not fit such data best. For example, we fitted a double 268 exponential model, which is a generalized version of an asymptote model (i.e., an asymptote model is 269 a special case of the double exponential model where $k^2 = 0$), and found improved fit relative to an 270 asymptote model (Fig. 6). The AIC values resulting from double exponential models (-26.0, -28.3, -271 33.3, and -36.9 in green tea-stream, green tea-pond, rooibos tea-stream, and rooibos tea-pond, 272 respectively) were lower than those from the asymptote model (-10.5, -12.9, -25.5, and -21.5 in green 273 tea-stream, green tea-pond, rooibos tea-stream, and rooibos tea-pond, respectively). More complicated 274models, using more data with more frequent sampling and longer incubation periods, such as triple 275 exponential models considering the decomposition of the acid-insoluble fraction, may provide 276 important further information. This approach would negate a major advantage of the TBI method, i.e., 277 only a single measurement is required to determine an asymptote model, but it is still beneficial 278 because tea bags reduce the labor of preparing litter bags and the materials are highly standardized, 279 which enables comparison between studies. In conclusion, we have demonstrated that the TBI 280approach is not applicable to aquatic environments, but we nevertheless suggest that tea bags are 281 beneficial for assessing aquatic litter decomposition, so their potential application should be further 282 assessed.

284 References

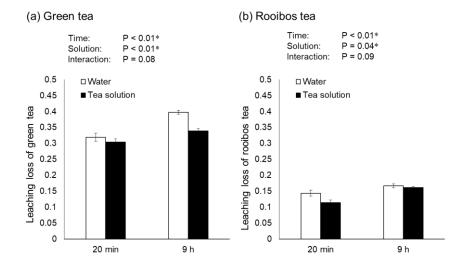
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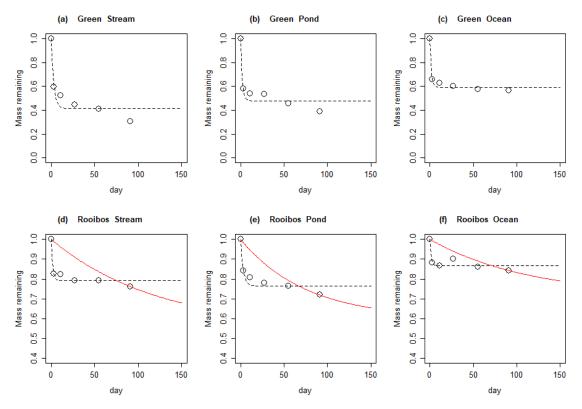
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Fig. 1. Relationships between solute concentration and the leaching ability of (a) green tea and (b) rooibos tea. Leaching losses were compared between deionized water and a tea solution for each tea type. Error bars indicate the standard error of three replicates.

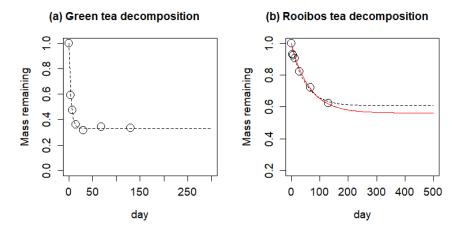
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Fig. 2. Relative remaining masses of green (a–c) and rooibos (d–f) tea bags in water samples taken from a stream (a, d), pond (b, e), and ocean (c, f) in Kumamoto, Japan. Open circles represent the average of four replicates. Tea bags were incubated in the dark and retrieved at 0, 3, 11, 27, 55, and 91 days after the start of the incubation. Dashed lines indicate the associated asymptote model: W(t) = a $*e^{-kt} + (1-a)$, where W(t) is the mass remaining after incubation time *t*, *k* is a decomposition constant, *a* is the decomposable and *1-a* is the un-decomposable fraction of the teas. Solid red lines indicate the asymptote models describing rooibos tea decomposition, as determined by the TBI.

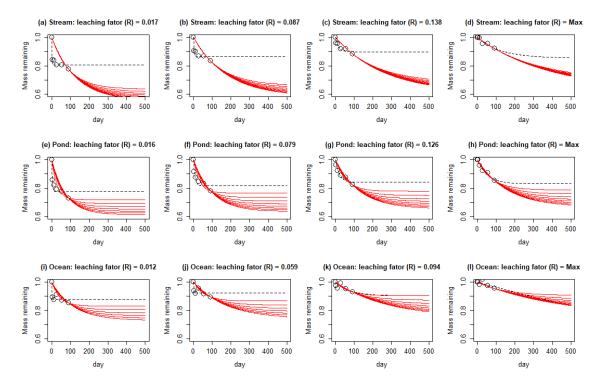
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402 Fig. 3. Relative remaining masses of (a) green and (b) rooibos teas in a terrestrial system. Data were 403 obtained from Keuskamp et al. (2013) using Data Thief 3.0 (Tummers, 2006). Tea bags were incubated 404 at 25°C in the dark for 0, 4, 7, 14, 30, 68, and 130 days. Dashed lines indicate asymptote models 405 (reported by Keuskamp et al. 2013) see Fig. 2. The solid red line indicates the TBI-based model.

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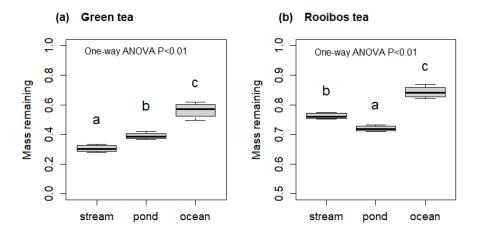
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409 Fig. 4. Leaching factor correction scenarios for the TBI-based asymptote model. Multiple scenarios 410 with different leaching factors were tested. The remaining-mass values of rooibos tea at 0, 3, 11, 27, 411 55, and 91 days after the start of incubation are shown, assuming 10% (a, e, i), 50% (b, f, j), 80% (c, 412 g, k), and 100% (d, h, l) of the maximum leaching factor of rooibos tea. Dashed lines indicate 413 asymptote models. Solid red lines indicate TBI-based asymptote models, assuming 0%, 12.5%, 25%, 414 37.5%, 50%, 62.5%, 75%, 87.5%, and 100% of the maximum leaching factor of green tea. Water 415 samples were obtained from a stream (a-d), pond (e-h), and ocean (i-l) in Kumamoto Prefecture, 416 Japan. R indicates the leaching factor for rooibos tea.

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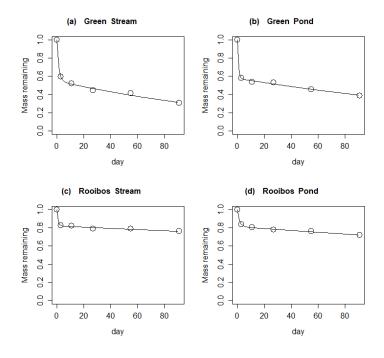


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421 Fig. 5. Boxplots representing the remaining mass values of (a) green and (b) rooibos tea bags at the

422 end of a 91-day incubation period. Amounts relative to initial weights are shown. Letters indicate

423 significant differences among groups, as determined by Tukey's post hoc tests.



426 **Fig. 6.** Time series data of green (a, b) and rooibos (c, d) teas in water samples taken from a stream (a, 427 c) and pond (b, d) in Kumamoto, Japan. Solid lines indicate double exponential models: $W(t) = a * e^{-428}$ 428 $k^{1t} + b * e^{-k2t}$, where W(t) is the mass remaining after incubation time *t*, *k1* and *k2* are decomposition 429 constants, and *a* and b represent organic matter fractions differing in decomposability. Each open circle 430 represents the average of four replicates. Tea bags were incubated in the dark and retrieved at 0, 3, 11, 431 27, 55, and 91 days after the start of the incubation.

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436 Supplemental materials

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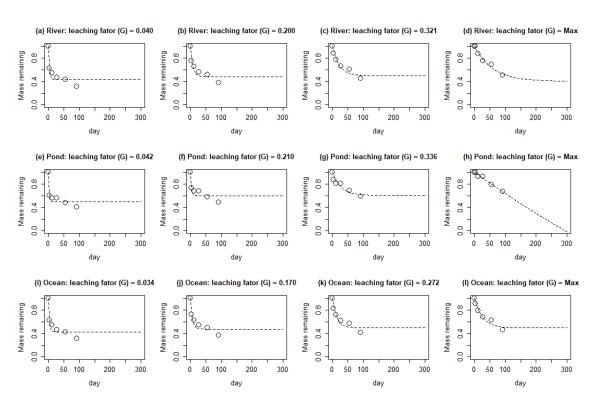


Fig. S1. Leaching factor scenarios applied to time-series green tea decomposition data. The remainingmass values of green tea at 0, 3, 11, 27, 55, and 91 days after the start of the incubation are shown, assuming 10% (a, e, i), 50% (b, f, j), 80% (c, g, k), and 100% (d, h, l) of the maximum leaching factor of green tea. Dashed lines indicate asymptote models. Data were obtained from a stream (a–d), pond (e–h), and ocean (i–l) in Kumamoto Prefecture, Japan. G indicates the leaching factor of green tea.

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