

1 **Proprioceptive Genes as a Source of Genetic Variation Underlying**  
2 **Robustness for Flight Performance in *Drosophila***

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9

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20 **Article Summary**

21 We sought to understand the genetic architecture of robustness (variation in a trait  
22 caused by non-genetic factors) for flight performance. We used 197 Drosophila Genetic  
23 Reference Panel (DGRP) lines to find significant individual variants and pairs of  
24 epistatic interactions, many of which were involved in proprioception. Additionally, we  
25 validated significant genes identified from a prior study for the mean of flight  
26 performance, showing genes affecting trait means may also affect trait robustness.

27 **ABSTRACT**

28

29 A central challenge of quantitative genetics is partitioning phenotypic variation into  
30 genetic and non-genetic components. These non-genetic components are usually  
31 interpreted as environmental effects; however, variation between genetically identical  
32 individuals in a common environment can still exhibit phenotypic variation. A trait's  
33 resistance to variation is called robustness, though the genetics underlying it are poorly  
34 understood. Accordingly, we performed an association study on a previously studied,  
35 whole organism trait: robustness for flight performance. Using 197 of the *Drosophila*  
36 Genetic Reference Panel (DGRP) lines, we surveyed variation across single nucleotide  
37 polymorphisms, whole genes, and epistatic interactions to find genetic modifiers  
38 robustness for flight performance. There was an abundance of genes involved in the  
39 development of sensory organs and processing of external stimuli, supporting previous  
40 work that processing proprioceptive cues is important for affecting variation in flight  
41 performance. Additionally, we tested insertional mutants for their effect on robustness  
42 using candidate genes found to modify flight performance. These results suggest  
43 several genes involved in modulating a trait mean are also important for affecting trait  
44 variance, or robustness, as well.

## 45 INTRODUCTION

46

47 Evolution acts on the genetic variation underlying phenotypic variation among  
48 individuals and populations. While many research programs focus on understanding  
49 genetic factors that contribute to phenotypic variation, fewer focus on non-genetic  
50 factors. The phenomenon of non-genetic (micro-environmental) variation describes the  
51 phenotypic variation that occurs in the absence of genetic variation, best studied in  
52 genetically identical individuals. Non-genetic variation can arise from external  
53 (environmental) or internal (developmental) factors. Phenotypic variation across  
54 different environmental conditions (e.g. temperature) in genetically homogenous  
55 organisms is termed phenotypic plasticity. However, significant phenotypic variation can  
56 also arise among genetically homogeneous organisms in the absence of explicit  
57 environmental variation (MORGANTE *et al.* 2015; VOGT 2015). Here, internal factors spur  
58 developmental noise in stochastic molecular processes, such as important transcripts or  
59 signals in very low abundance, which can result in varying levels of developmental  
60 stability (ALBAYRAK *et al.* 2016; SCHOR *et al.* 2017; KLINGENBERG 2019). The processes  
61 or ability for organisms to maintain a consistent phenotype in the presence of these  
62 perturbations is termed buffering, while the resulting phenotype is deemed robustness  
63 (KLINGENBERG 2019).

64

65 Developmental noise can affect an organism's developmental trajectory, which may  
66 impact the efficacy of natural selection by altering the association between genotype  
67 and phenotype. While it is difficult to directly observe developmental noise, deviations

68 from an expected phenotype provide an adequate lens for study (MORGANTE *et al.* 2015;  
69 VOGT 2015). An example of this is deviations in bilateral symmetry (fluctuating  
70 asymmetry) (VALEN 1962; SOTO *et al.* 2008), which are hypothesized to be negatively  
71 associated with fitness in the case of facial symmetry (QUINTO-SANCHEZ *et al.* 2018;  
72 LAJUS *et al.* 2019). Some genetic safeguards exist to buffer against developmental noise  
73 and maintain phenotypic robustness in the presence of these stressors. Chaperonins  
74 (HSP90) do so by maintaining a protein's structure during stressful times (RUTHERFORD  
75 AND LINDQUIST 1998; CHEN AND WAGNER 2012), as does the mitochondrial unfolded  
76 protein response in maintaining homeostasis and promoting longevity (PELLEGRINO *et al.*  
77 2013; JOVAISAITE *et al.* 2014). In contrast, certain neurodevelopmental cell-cell adhesion  
78 molecules (e.g. DSCAMs, cadherins, and teneurins) leverage developmental noise to  
79 create more robust neural networks. In doing so, they drive repeatable non-genetic  
80 phenotypic variation in behavioral responses to serve as a bet hedging strategy (VOGT  
81 *et al.* 2008; AYROLES *et al.* 2015; HIESINGER AND HASSAN 2018; HONEGGER AND DE BIVORT  
82 2018).

83  
84 Genes that modulate a system's ability to resist developmental noise or a stressor are  
85 hypothesized evolutionary targets (WAGNER 2008; VOGT 2015; MENEZES *et al.* 2018) and  
86 subject to natural selection. And yet, these sources of non-genetic phenotypic variation  
87 are poorly understood. Previous studies employed a Genome Wide Association Study  
88 (GWAS) framework on trait robustness, demonstrating the strategy's feasibility to  
89 identify significant genetic modifiers (KAIN *et al.* 2012; AYROLES *et al.* 2015; MORGANTE  
90 *et al.* 2015; MENEZES *et al.* 2018; ROMAN *et al.* 2018). Similarly, we sought to elucidate

91 these genetic factors by studying the robustness of flight performance in a GWAS  
92 framework. We turned to the *Drosophila* Genetics Reference Panel (DGRP) lines, a  
93 collection of 205 genetically distinct and inbred lines of *D. melanogaster* that represent a  
94 snapshot of natural variation in a wild population (MACKAY *et al.* 2012; HUANG *et al.*  
95 2014). Using a flight column to assay flies' ability to react and respond to an abrupt drop  
96 (BENZER 1973; BABCOCK AND GANETZKY 2014), we tested 197 DGRP lines for their  
97 mean-normalized standard deviation (coefficient of variation) in flight performance. The  
98 natural log-transformed coefficient of variation serves as a more normally-distributed  
99 proxy for studying phenotypic robustness for genetically distinct groups comprised of  
100 genetically identical individuals. In this study, we identified significant individual variants  
101 and epistatic interactions, while also exploring the top hits from a whole gene screen  
102 across four sex-based phenotypes (males, females, and the average (sex-average) and  
103 difference (sex-difference) between sexes). We also used a panel of insertional  
104 mutations in several candidate genes (*bru1*, *CadN*, *CG15236*, *CG32181/Adgf-A/Adgf-*  
105 *A2*, *CG3222*, *flippy/CG9766*, *CREG*, *Dscam4*, *flapper/CG11073*, *Form3*, *fry*,  
106 *Lasp/CG9692*, *Pde6*, *Snoo*), detected in a previous study though they were not  
107 significant in the current one (SPIERER *et al.* 2021). The successful validation of these  
108 genes hints at the dual importance of genes modulating a trait mean and its variance,  
109 and it highlights how there are still many more genetic modifiers that affect robustness  
110 of flight performance. Across these analyses, we found consistent evidence for the  
111 development and function of sensory organs that process external stimuli, including  
112 those involved in touch, sight, smell, and sound. Together, these genes highlight the  
113 importance of processing proprioceptive cues for robust flight performance.

## 114 **METHODS**

115

### 116 *Drosophila Stocks and Husbandry*

117 197 *Drosophila* Genetic Reference Panel (DGRP) lines (HUANG *et al.* 2014) and 24  
118 stocks used in the validation experiment were obtained from Bloomington *Drosophila*  
119 Stock Center (Table S1; <https://bdsc.indiana.edu/>). Flies were grown on a standard  
120 cornmeal media (MOSSMAN *et al.* 2016) at 25° under a 12h:12h light-dark cycle. Two to  
121 three days post-eclosion, flies were sorted by sex under light CO<sub>2</sub> anesthesia and given  
122 five days to recover before assaying flight performance. All flies scored for robustness,  
123 whether in the initial phenotyping screen or in the validation screen, were reared under  
124 the same conditions.

125

126

### 127 *Flight performance assay*

128 We tested approximately 100 flies of each sex from 197 DGRP genotypes (Table S1)  
129 using a refined protocol (BABCOCK AND GANETZKY 2014) for measuring flight  
130 performance (BENZER 1973). For each sex-genotype combination, groups of 20 flies in  
131 five glass vials were knocked down, uncorked, and rapidly inverted down a 25 cm  
132 chute. The vials traveled until they reached a stop, at which point flies were ejected into  
133 a 100 cm long by 13.5 cm wide tube. Freefalling flies instinctively attempt to right  
134 themselves and land. A transparent acrylic sheet coated in TangleTrap<sup>®</sup> adhesive lined  
135 the inside of the tube and immobilized flies at their respective landing height. The sheet,  
136 was removed, pinned to a white poster board, and photographed using a Raspberry Pi

137 (model 3 B+) and PiCamera (V2). The positional coordinates were extracted using  
138 ImageJ/FIJI's (SCHINDELIN *et al.* 2012) 'Find Maxima' feature with options for a light  
139 background and noise tolerance of 30. The distributions of landing heights for each sex-  
140 genotype combination were used to calculate the mean distance traveled and standard  
141 deviation. The coefficient of variation represents the standard deviation normalized by  
142 the mean distance traveled. Finally, we performed a natural log-transformation on each  
143 genotype score to make the data more normally distributed. Thus, the natural log-  
144 transformed coefficient of variation serves as our phenotype proxy for robustness of  
145 flight performance.

146

147

#### 148 Genome wide association mapping

149 Robustness phenotypes (Table S2) were submitted to the DGRP2 webserver (reference  
150 genome FB5.57) for the association analysis (<http://dgrp2.gnets.ncsu.edu/>) (MACKAY *et*  
151 *al.* 2012; HUANG *et al.* 2014). This computational pipeline returned single variant results  
152 for four sex-based phenotypes: males, females, average between sexes (sex-average)  
153 and difference between sexes (sex-difference). We refer to this analysis and its  
154 respective results as the individual variant analysis, since this analysis is the standard  
155 when working with the DGRP lines. We analyzed 1,901,174 common variants (minor  
156 allele frequency  $\geq 0.05$ ) using a mixed effect model to account for *Wolbachia* infection  
157 status and presence of five major inversions. Since certain inversions covaried with the  
158 robustness phenotype (Table S4), only significance scores from a linear mixed model



159 accounting for Wolbachia status and the presence of five major inversions were  
160 considered.

161

162

### 163 Validating candidate genes

164 Candidate genes (Table S1B) were selected if they were identified from variants  
165 identified in the sex-average, individual variant screen for mean landing height and if  
166 there were publicly available lines containing a *Minos* enhancer trap (*Mi{ET1}*)  
167 mutational insertion (METAXAKIS *et al.* 2005) generated by the Drosophila Gene  
168 Disruption Project (BELLEN *et al.* 2011). Experimental and control lines were derived  
169 from common isoparental crosses for each candidate gene stock backcrossed for five  
170 generations to the respective  $w^{1118}$  or  $y^1w^{67c23}$  background. Isoparental crosses between  
171 the resulting heterozygous offspring were partitioned for absence (control line) or  
172 presence (experimental line) of the *Mi{ET1}* construct. Experimental lines were verified  
173 for homozygosity if all progeny contained the insertion after several rounds of culturing.  
174 Validations were conducting in the flight performance assay described above. The  
175 distributions in landing heights were assessed for significance if they passed a  $P \leq 0.05$   
176 significance threshold in a Kolmogorov-Smirnov test comparing control and mutant  
177 genotypes (SPIERER *et al.* 2021).

178

179

### 180 Calculating gene-score significance

181 Gene-level significance scores (gene-score) were determined using `PEGASUS_flies`  
182 (`SPIERER et al. 2021`), a Drosophila-optimized method for the human-based platform  
183 Precise, Efficient Gene Association Score Using SNPs (`PEGASUS`) (`NAKKA et al. 2016`).  
184 This analysis calculates gene-scores for each gene as a test of whether the distribution  
185 of individual variants within a gene (accounting for linkage disequilibrium) deviates from  
186 a null chi-squared distribution. Variants from the individual variant association screen  
187 were considered and mapped onto gene annotations and linkage disequilibrium files  
188 available with the `PEGASUS_flies` package—derived initially from the DGRP2  
189 webserver. Because no variants passed the strict Bonferroni significance threshold ( $P =$   
190  $3.13E-6$ ), we explored the top five genes for each sex-based phenotype.

191

192

### 193 Screening for epistatic interactions

194 Epistatic hub variants, corresponding with variants more likely to interact with other  
195 variants, were identified using Marginal ePIstasis Test (`MAPIT`) (`CRAWFORD et al. 2017`).  
196 This approach tests the marginal effect of each variant against a focal phenotype.  
197 `MAPIT` requires a complete genotype-phenotype matrix so the DGRP genome was  
198 imputed for missing variants using `BEAGLE 4.1` (`BROWNING AND BROWNING 2007`;  
199 `BROWNING AND BROWNING 2016`) and filtered for  $MAF \geq 0.05$  using `VCFtools` (v.0.1.16)  
200 (`DANECEK et al. 2011`).

201

202 `MAPIT` was run using the 'Davies' method on the raw phenotype scores, 1,952,233  
203 `BEAGLE`-imputed and filtered variants, and the DGRP2 webserver's relatedness and

204 covariate status files. Since none of the epistatic hub variant  $P$ -values passed the strict  
205 Bonferroni significance threshold ( $P < 2.56e-8$ ) in any of the sex-based phenotypes, we  
206 used the 15 most significant variants as a focused subset for targeted pairwise epistasis  
207 testing against the unimputed variants ( $n = 1,901,174$ ). Epistatic interactions were  
208 calculated using the '-epistasis' test in a '-set-by-all' framework in PLINK (v.1.90)  
209 (PURCELL *et al.* 2007). Significant epistatic interactions were considered if they passed a  
210 Bonferroni threshold ( $P < 1.75E-9$ ).

211

212

### 213 Data availability

214 All phenotype data required to run the outlined analyses are available in Table S2 or  
215 using the DGRP2 webserver (<http://dgrp2.gnets.ncsu.edu/>).

## 216 **RESULTS and DISCUSSION**

217

218 We sought to identify the genetic modifiers of robustness in a whole organism  
219 phenotype: flight. Using the Drosophila Genetic Reference Panel (DGRP) lines, we  
220 identified several individual variants, validated a previously identified subset of genes for  
221 robustness, and two pairs of significant epistatic interactions. While we didn't find any  
222 significant whole genes, some of the most significant genes corresponded with  
223 modifiers of trans-regulatory gene expression and detecting external stimuli. In the  
224 sections that follow we describe the variant-based analysis, gene-based analysis,  
225 epistatic analysis, and validation of candidate genes.

226

### 227 *Variation in flight performance across the DGRP*

228 We screened 197 DGRP lines (Table S1A) for their flight ability in response to an abrupt  
229 drop (Figure 1A-B). Qualitative observations made in a previous study of strong,  
230 intermediate, and weak genotypes in the flight assay suggests stronger genotypes react  
231 faster and respond more effectively than weaker one (SPIERER *et al.* 2021). The mean  
232 and standard deviation in landing height were calculated for each sex-genotype  
233 combination, though the standard deviation was related to the mean landing height  
234 (males:  $R = 0.72$ ,  $P < 1E-32$ ; females:  $R = 0.68$ ,  $P < 1E-28$ ). To study variation in the  
235 absence of the mean, we chose to use the coefficient of variation. Additionally, we  
236 natural log-transformed the coefficient of variation to make the data more normally  
237 distributed (Figure S1). An earlier pre-print of this work calculated the coefficient of  
238 variation as the standard deviation normalized by the mean landing height from the

239 bottom of the column (SPIERER AND RAND 2020), though this created a negative  
240 association between our metric and robustness so we chose to normalize the standard  
241 deviation by the mean distance fallen in the column (Table S2). Thus, the natural log-  
242 transformed coefficient of variation served as our metric for robustness.

243

244 In this study, genotypes with a lower coefficient of variation (more consistent) were  
245 more robust for flight performance (KLINGENBERG 2019). On average, flight performance  
246 was more robust in males than females (males:  $-0.45 \text{ A.U.} \pm 0.19 \text{ SD}$  vs. females:  $-0.58$   
247  $\text{A.U.} \pm 0.19 \text{ SD}$ ; Figures 1B). There was a significant relationship in robustness between  
248 sexes ( $R = 0.41$ ;  $P < 5E-9$ ; Figure 1C), suggesting the genetic architecture of  
249 robustness in flight performance is similar between the sexes. However, the magnitude  
250 of the regression coefficient suggests robustness is somewhat sexually dimorphic.

251

252 We tested our phenotype in both males and females against those publicly available on  
253 the DGRP2 webserver to determine whether robustness of flight performance was a  
254 unique trait. We found no significant relationship after imposing a significance threshold  
255 of  $P \leq 1.8E-3$  to account for multiple testing (Table S3), suggesting our phenotype is  
256 unique.

257

### 258 *Several variants of large effect associate with robustness in flight performance*

259 We performed a Genome Wide Association Study (GWAS) to calculate each variant's  
260 significance, and subsequently whole gene significance scores. We analyzed the effects  
261 of 1,901,174 common variants ( $\text{MAF} \geq 0.05$ ) across for four sex-based phenotypes

262 (males, females, the sex-average, and sex-difference; Figures 1D and S2-4). Although  
263 none of the major inversions or presence of Wolbachia covaried with our phenotype  
264 scores (Table S4), we still used a mixed effects model to minimize extraneous sources  
265 of variation.

266

267 We performed a GWAS on the robustness phenotype using the DGRP2 webserver  
268 pipeline. Only one variant (2L\_5852054\_SNP;  $P = 6.24E-9$ ) in the sex-difference screen  
269 passed a strict Bonferroni significance threshold ( $P = 2.63E-8$ ). This variant mapped to  
270 an intron of *TrissinR*, a neuropeptide receptor that binds *Trissin* and acts as a G-protein  
271 coupled receptor. *TrissinR* was previously identified to be important for neuronal  
272 communication in Olfactory Receptor Neurons (ORN) and Ionotropic Receptors (IR)  
273 (MCLAUGHLIN *et al.* 2021). The gene's importance in flight was previously documented in  
274 our previous study on the genetic modifiers of the mean of flight performance where it  
275 was a significant epistatic interactor with the chemo- and mechanosensing gene *ppk23*  
276 (SPIERER *et al.* 2021).

277

278 Applying the individual variant, DGRP association threshold ( $P \leq 1E-5$ ), we identified 69  
279 unique variants across 41 genes (Table S5). No variant corresponded with protein  
280 coding changes, suggesting variation in complex traits is driven by modulation of gene  
281 regulation rather than changes to protein coding sequence (MACKAY *et al.* 2012;  
282 MACKAY AND HUANG 2018). Seventeen of these genes were identified from several  
283 different analyses in our prior study: *app*, *CG10362*, *CG15270*, *CG17839*, *CG32264*,  
284 *CG43313*, *cv-c*, *dpr2*, *ec*, *Eip75B*, *Gmap*, *ju*, *Kdm4B*, *ncd*, *ppk8*, *TrissinR*, *X11Lbeta*.

285 This overlap is suggestive that genes affecting a trait mean may also be important for  
286 affecting variation in the same trait.

287

288 In addition to direct overlaps in genes, we identified four paralogous genes shared  
289 between the present and prior study. In the present study, *Dscam2* is a paralog with  
290 *Dscam4*, which was identified in the individual variant analysis as a Bonferroni variant  
291 and validated for its role in mean flight performance. Dscam genes are also paralogs  
292 with defective proboscis response (dpr) genes, like *dpr2*, which was also identified here.  
293 Finally, two pickpocket genes (*ppk8* and *ppk27*) were paralogous with *ppk23*, a highly  
294 significant epistatic hub gene that is likely involved in relaying proprioceptive  
295 information.

296

297

### 298 *Analyses of whole-gene effects identifies distinct factors affecting robustness*

299 The individual variant screen takes a minSNP approach, deeming a gene significant if  
300 its most significant variant passes a significance threshold. However, this approach is  
301 biased toward longer genes and does not account for linkage between sites. To  
302 counteract these biases, we employed PEGASUS\_*flies* (SPIERER *et al.* 2021), a  
303 *Drosophila* version of the human-focused PEGASUS platform (NAKKA *et al.* 2016). This  
304 method takes a gene-specific approach; assessing a whole gene's significance by  
305 testing the distribution of variants within a gene against a null chi-squared distribution of  
306 SNP *P*-values. Thus, it can detect significant genes of moderate effect, as well as genes  
307 that may be missed in a minSNP approach.

308

309 We failed to identify any significant genes across the four sex-based phenotypes using  
310 a strict Bonferroni threshold ( $P \leq 3.43E-6$ ). Since this threshold is overly conservative,  
311 we looked at the top five genes from each of the four sex-based analyses and identified  
312 18 unique genes using `PEGASUS_flies` (Table S6 and Figure S5). Of these genes,  
313 only one had a single  $P$ -value exceed the individual significance threshold ( $P = 1E-5$ ; *ju*  
314 in sex-average), demonstrating how `PEGASUS_flies` is capable of expanding the list  
315 of potential candidate genes in GWAS-type studies.

316

317 Of the top five genes in each sex-based phenotype, four corresponded with trans-  
318 regulatory factors (*CG2034*, *CG4565*, *CG42526*, *Wdr82*) (GAUDET *et al.* 2011), which  
319 supports our earlier observation that variants identified through the individual variant  
320 analysis were in non-coding regions. Additionally, we identified genes involved in  
321 sensing the external environment through the chaeta development (hair-like structures  
322 responsible for chemo- and mechanosensation; *ju*) and the development of chordontal  
323 organs (stretch receptor organs; *btv*) (EBERL *et al.* 1997; EBERL *et al.* 2000; SHAPIRA *et*  
324 *al.* 2011). While these genes were not significant under a strict Bonferroni significance  
325 threshold, they still support an important role for variation in proprioception and  
326 receiving external stimuli in modulating the robustness of flight performance.

327

328

329 *Association of epistatic hub and pairwise epistatic variants with robustness in flight*  
330 *performance*



331 Epistatic, or pairwise, interactions play an outsized role as context-specific effectors in  
332 complex traits (HUANG *et al.* 2012). Traditional epistasis analyses face large  
333 computational and statistical hurdles, so we turned to MArginal ePIstasis Test (MAPIT)  
334 to focus the exhaustive pairwise search and identify epistatic hub variants with a greater  
335 likelihood of interacting with other variants (CRAWFORD *et al.* 2017). These hub variants  
336 were then used as a subset in a set-by-all pairwise epistasis search against all variants  
337 considered in the individual variant association analysis.

338

339 We failed to identify any epistatic hub variants that passed a strict Bonferroni  
340 significance threshold ( $P = 2.56E-8$ ). Since we were using MAPIT to narrow our search  
341 space for epistatic variants, we decided to focus on the 15 most significant variants in  
342 each sex-based phenotype to inform our search for epistatic variants instead. Doing so  
343 yielded two pairs of epistatic interactions, one in each the female and sex-difference  
344 analyses though none leading to changes in the protein coding sequence (Table S7).

345

346 The female interaction was between SNP pairs X\_14165625\_SNP and  
347 2R\_3523428\_SNP. The former corresponded with a synonymous coding site in *narrow*  
348 *abdomen* (*na*), an ion channel involved in locomotor rhythm and mechanosensation  
349 (NASH *et al.* 2002; LEAR *et al.* 2013). The latter corresponded with two separate genes,  
350 Myosin-7a binding protein (*M7BP*; intron) and *antisense RNA:CR45131* (704 bp  
351 upstream). Interestingly, *M7BP* localizes to actin-bundles in sensory organs in  
352 *Drosophila*, as well as the Johnston's organ, which is used for auditory sensation  
353 (KIEHART *et al.* 2004; TODI *et al.* 2005; TODI *et al.* 2008; LIU *et al.* 2021). The connection

354 between *na* and *M7BP* supports the importance of sensory hairs in proprioception and  
355 receiving external stimuli during flight that may modulate robustness.

356

357 The sex-difference interaction was between SNP pairs 3L\_7643140\_SNP and  
358 3R\_16731290\_SNP. The first SNP lies 508 bp upstream of *CG32373*. It is expressed in  
359 the Johnston's organ and is hypothesized to aid in synaptic formation (KURUSU *et al.*  
360 2008; SENTHILAN *et al.* 2012). Additionally, it is hypothesized to work with *nmo*,  
361 previously identified in flight performance (SPIERER *et al.* 2021), in ommatidial rotation  
362 (MUNOZ-SORIANO *et al.* 2013). Meanwhile, 3R\_16731290\_SNP falls within or near two  
363 genes: *Turandot X (TotX)* and *Grik*. *TotX* is a stress response gene in the JAK-STAT  
364 pathway best known in the context of heat stress (MANENTI *et al.* 2018), though it has  
365 been documented to have some connection to auditory processing (IMMONEN AND  
366 RITCHIE 2012). While it is possible that it interacts with *CG32373*, it is far more likely that  
367 the main epistatic interaction is with *Grik*, a glutamate receptor involved in synaptic  
368 transmission in the adult brain and visual system (GAUDET *et al.* 2011; KARUPPUDURAI *et*  
369 *al.* 2014). It is orthologous to *glutamate ionotropic receptor kainate type subunits 1-3*  
370 (*GRIK1-3*), involved in the development of intellectual disability and Huntington's  
371 disease in humans (MACDONALD *et al.* 1999). Together, it would follow that *CG32373*  
372 and *Grik* might work together in the *Drosophila* flight system to process visual and/or  
373 auditory signals that are important in the robustness of flight performance.

374

375 *Functional validation of candidate genes supports a role for neurodevelopment affecting*  
376 *robustness of flight performance*

377 Finally, we sought to test whether genes that modify the mean flight performance  
378 phenotype also modify the robustness in flight performance. To do so, we tested 24  
379 independent insertional mutations in candidate genes identified from an earlier study on  
380 mean flight performance (SPIERER *et al.* 2021). Of these, 21 constructs fell in unique  
381 genes while three constructs were used as independent replicates of different highly  
382 significant genes in the mean flight phenotype (*CadN*, *Dscam4*, *flap* (CG11073))  
383 (SPIERER *et al.* 2021). Of the 21 unique genes, all but one (*CREG*) were strongly  
384 significant in the mean flight performance paper's list of top variants (SPIERER *et al.*  
385 2021). Despite their significance in the other study, none of these genes were  
386 significantly associated with robustness in any of the four sex-based phenotypes in the  
387 present study. Thus, we were also able to test whether there were significant genes  
388 affecting robustness that we were unable to detect due to a lack of power.  
389  
390 Of these 21 genes, there was a significant difference in robustness for 13 constructs  
391 using a comparison of genotypes carrying an insertional mutation in a candidate gene of  
392 interest against their backcross-control genotypes. We found statistical significance with  
393 a Kolmogrov-Smirnov test for 11 candidate genes where the construct inserted within  
394 single genes (*bru1*, *CadN*, *flip* (CG9766), *CG15236*, *CREG*, *Dscam4*, *flap* (CG11073),  
395 *form3*, *fry*, *Pde6*, and *Snoo*) and two where the construct inserted in multiple genes  
396 (*Adgf-A/Adgf-A2/CG32181* and *CG9692/Lasp*) (Figure 2 ; Table S8). These genes were  
397 also previously validated in the mean flight performance screen (SPIERER *et al.* 2021),  
398 suggesting that genes likely play dual roles modifying the ability and variability of flight  
399 performance. These analyses using insertional mutations showed that while natural

400 variation in this set of 21 candidate genes for mean flight performance do not pass  
401 robustness of flight GWA thresholds for significance, specific mutations in those genes  
402 are capable of impacting robustness in 13 of these 21 genes.

403

404 Interestingly, *CadN* and *Dscam4* are important genes contributing to type IV dendritic  
405 arborization sensory neurons. These genes are known to contribute to robustness as  
406 they connect sensory structures (e.g. chaete) to the peripheral nervous system. They  
407 also work with teneurins (e.g. *Ten-a*), which are known to affect robustness of locomotor  
408 handedness (BUCHANAN *et al.* 2015). *CadN* and *Dscam4* also work with *fry* and *Snoo*,  
409 which develop and pattern chaete and campaniform sensilla on the wing, and are likely  
410 useful in mechanosensation and proprioception during flight (EMOTO *et al.* 2004; NEVES  
411 *et al.* 2004; SOBA *et al.* 2007; FUERST AND BURGESS 2009; QUIJANO *et al.* 2010;  
412 MATSUBARA *et al.* 2011).

413

414 Our findings suggest that our experimental design is sufficient to identify individual  
415 variants affecting robustness. However, we were limited in our power to detect whole  
416 gene or epistatic interactions affecting robustness. While we could not comprehensively  
417 detect all genetic modifiers of flight robustness, the fact that mutations in genes  
418 affecting mean flight performance can affect robustness implies that many other genes  
419 affecting robustness likely exist. Even using all but eight of the available DGRP lines,  
420 we lacked the power to detect many genes. Therefore, we suggest that future studies  
421 exploring the mean and robustness for traits with the DGRP lines should supplement  
422 the core panel with other sources of genetic variation, such as the Global Diversity

423 Panel (GDP) or an Advanced Intercross Population (AIP) (GRENIER *et al.* 2015; MACKAY  
424 AND HUANG 2018).

425

#### 426 Conclusions

427 We present results from four analyses across four sex-based phenotypes surveying  
428 different facets of the genetic architecture of robustness for flight performance. The  
429 individual variant analysis was the most fruitful for identifying novel genetic modifiers of  
430 robustness in flight performance, while the screen for epistatic interactions found two  
431 pairs of genes that were both involved in processing external cues (mechano-, audio-  
432 and visuosensory sensory) that are also likely important for proprioception. A whole  
433 gene screen did not meet strict significance thresholds though the most significant  
434 genes in the analysis indicated trans-regulatory genes and some genes involved in the  
435 development of proprioceptive structures were important. Finally, we validated several  
436 genes roles in contributing to robustness of flight performance that were not detected in  
437 this study. This result suggests that despite our current findings, there are many more  
438 genetic modifiers of robustness left to identify. These genetic modifiers likely require  
439 additional genotypic and phenotypic variation to detect, so we suggest future studies  
440 supplement the DGRP with other panels of flies (GDP or an AIP) to counteract these  
441 limitations.

442

443 Future studies in other phenotypes should consider evaluating both the mean and  
444 standard deviation or coefficient of variation for their focal phenotype to better  
445 understand modifiers affecting robustness in a specific complex trait, as well as

446 robustness in complex traits more generally. Doing so would provide a better survey of  
447 the genetic modifiers of robustness as a phenotype and allow for greater insights into  
448 the mechanisms of evolutionary change.

449

#### 450 **Author contributions**

451 DMR and ANS conceived the idea and designed the experiment. ANS performed  
452 experiments and analyses. DMR and ANS wrote and revised the manuscript.

453

#### 454 **Data accessibility statement**

455 All phenotype data required to run the outlined analyses are available in Table S1 or  
456 using the DGRP2 webserver (<http://dgrp2.gnets.ncsu.edu/>). Supplemental tables and  
457 supplemental figures are hosted by Dataverse: <https://doi.org/10.7910/DVN/MV7QA4>.

458

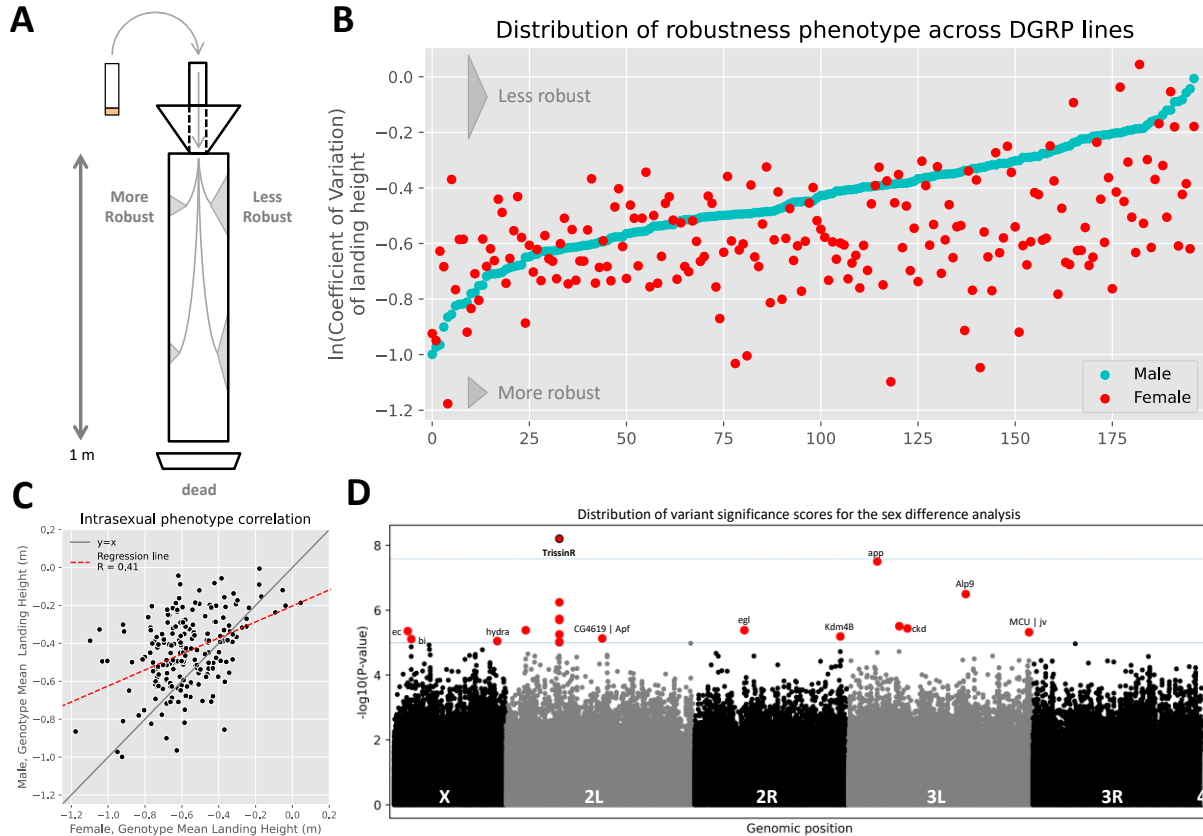
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462

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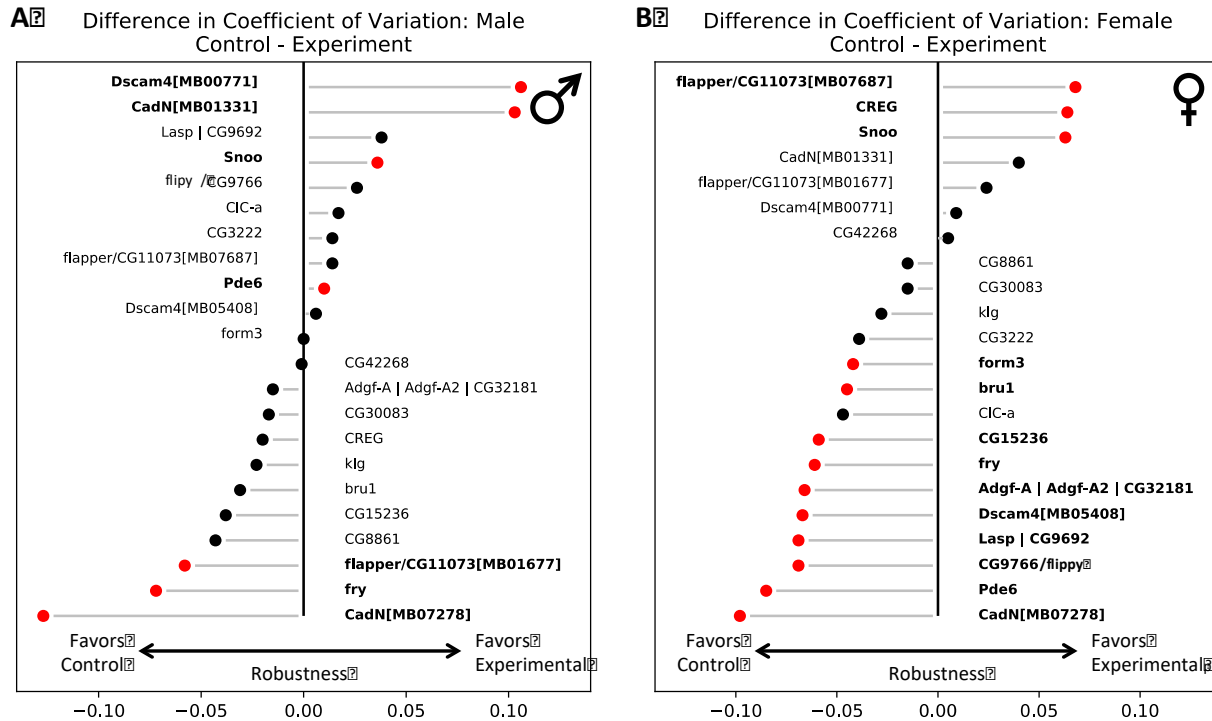
464 This work and ANS are supported by National Institutes of Health R01 GM067862 (to  
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**Figure 1. The *Drosophila* Genetic Reference Panel lines demonstrate variation for robustness in flight performance across genotypes and sexes.** (A) Flies were assayed for flight performance using a meter-long flight column (BABCOCK AND GANETZKY 2014). The natural log-transformed coefficient of variation (mean-normalized standard deviation) is a proxy for robustness; more robust genotypes have less variation in landing height around the mean. Flies that passed through the column were excluded from the analysis. (B) The phenotypic distribution of sex-genotype pairs, ordered by increasing male score, demonstrates the DGRP lines have variation in their robustness for flight performance. Genotypes demonstrated phenotypic variation for robustness in both sexes. More negative values correspond with increased robustness. (C) Males were generally more robust than females, though the two were related ( $R = 0.41$ ,  $P < 5E-9$ ; regression line in red). Sexual dimorphism is observed by the intersection of the regression line and  $y = x$  line (gray). (D) Individual variants in the sex-difference analysis, visualized as a function of the  $-\log_{10}$  of variants'  $P$ -value illustrates several variants (red) passed the suggestive DGRP significance threshold ( $P \leq 1E-5$ ; blue solid line), and one (red with black outline) passed Bonferroni significance threshold ( $P \leq 2.63E-8$ , blue dashed line). Variants that did not pass the significance threshold are colored in black or gray by chromosome. Other sex-based phenotype Manhattan plots are available in Figure S4.

467

## Functional validation of candidate genes



**Figure 2. Several genes validated for robustness of flight performance.** Flies homozygous for *Mi{ET1}* insertion constructs inserted in candidate genes (experiment) were tested against their background control (control). Comparisons between control and experiment lines were assessed for significance using a Kolmogrov-Smirnoff test ( $P \leq 0.05$ ; red points and bold text). Values to the left of the midline suggest control genotypes were more robust than experimental lines, while the opposite is true for values to the right of the line. (A) Seven constructs were significant in males, (B) while 13 were significant in females. Some candidate genes were tested more than once (*CadN*, *Dscam4*, and *flap*) because they were strongly significant in the sex-average, individual variant association screen. Separate constructs are denoted by a suffix containing a 'MB' code.

468



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