The genome sequence of the avian vampire fly (*Philornis downsi*), an invasive nest parasite of Darwin's finches in Galápagos

Melia Romine[1], Sarah A. Knutie[2,3], Carly M. Crow[4], Grace J. Vaziri[2], Jaime Chaves[5,6], Jennifer A.H. Koop[4*] & Sangeet Lamichhaney[1,7*#]

[1]School of Biomedical Sciences, Kent State University, Kent, OH, USA

[2]Department of Ecology and Evolutionary Biology, University of Connecticut, Storrs, CT USA

[3]Institute for Systems Genomics, University of Connecticut, Storrs, CT USA

[4]Department of Biological Sciences, Northern Illinois University, DeKalb, IL, USA

[5]Department of Biology, San Francisco State University, San Francisco, CA, USA

[6]Colegio de Ciencias Biológicas y Ambientales, Universidad San Francisco de Quito, Quito, Ecuador

[7]Department of Biological Sciences, Kent State University, Kent, OH, USA

[*]co-senior authors

[#]corresponding author (slamichh@kent.edu)

Abstract

The invasive avian vampire fly (*Philornis downsi*) is considered one of the greatest threats to the unique and endemic avifauna of the Galápagos Islands, Ecuador. The fly parasitizes nearly every passerine species, including Darwin's finches, in the Galápagos. The fly is thought to have been introduced from mainland Ecuador, although the full pathway of invasion is not yet known. The majority of research to date has focused on the effects of the fly on the fitness of avian hosts and explorations of mitigation methods. A lag in research related to the genetics of this invasion demonstrates, in part, a need to develop full-scale genomic resources with which to address further questions within this system. In this study, an adult *P. downsi* collected from San Cristobal Island within the Galápagos archipelago was sequenced to generate a high-quality genome assembly. We examined various features of the genome (e.g., coding regions, non-coding transposable elements) and carried out comparative genomics analysis against other dipteran genomes. We identified lists of gene families that are significantly expanding/contracting in *P. downsi* that are related to insecticide resistance, detoxification, and potential feeding ecology and counter defense against host immune responses. The *P. downsi* genome assembly provides an important foundational resource for studying the molecular basis of its successful invasion in the Galápagos and the dynamics of its population across multiple islands. The findings of significantly changing gene families associated with insecticide resistance and immune responses highlight the need for further investigations into the role of different gene families in aiding the fly's successful invasion. Furthermore, this genomic resource will also better help inform future research studies and mitigation strategies aimed at minimizing the fly's impact on the birds of the Galápagos.

Keywords: Avian vampire fly genome, parasitic invasion, Galápagos, insecticide resistance, Darwin's finches

Introduction

The invasive avian vampire fly (*Philornis downsi*) (Figure 1a) is considered among the greatest threats to the unique and endemic avifauna of the Galápagos Islands, Ecuador (Causton *et al.*, 2013). The fly parasitizes eleven species of Darwin's finches (Figure 1b) as well as nearly every other Galápagos passerine species (Causton *et al.*, 2013; Fessl *et al.*, 2018). The earliest record of adult flies from the Galápagos occurred in 1964, but the fly was not observed in the nests of birds until 1997 (Fessl and Tebbich, 2002; Causton *et al.*, 2006). The fly has now been found on all major islands within the archipelago except Española, Genovesa, Darwin, and Wolf (Causton *et al.*, 2013). The impact of parasitism by the fly has been severe in some populations of birds in Galápagos and while the effects are variable, some studies have reported near-total nest failure rates due to parasitism (Dudaniec *et al.*, 2007; Koop *et al.*, 2011; Koop, Le Bohec, *et al.*, 2013; O'Connor *et al.*, 2014; Knutie *et al.*, 2016a; Heimpel *et al.*, 2017; Addesso *et al.*, 2020). The fly has also been implicated in the decline of the medium tree finch (*Camarhyncus pauper*), the warbler finch (*Certhidia olivacea*), and the mangrove finch (*Camarhyncus heliobates*) (Dvorak *et al.*, 2004; Grant *et al.*, 2005; Cunninghame *et al.*, 2017; Peters and Kleindorfer, 2018; Bulgarella *et al.*, 2019). Furthermore, the potential for population-level declines of even relatively prominent bird species, e.g., the medium ground finch, have also been demonstrated using predictive models (Koop *et al.*, 2015).

The genus *Philornis* includes approximately 50 species found primarily in the Neotropics and into North America (Dodge, 1955, 1963; Spalding *et al.*, 2002; Dudaniec and Kleindorfer, 2006; Couri *et al.*, 2007). Of the described species within *Philornis*, only two species (including *P. downsi*) are free-living ectoparasites within the nests of their hosts (Fessl *et al.*, 2006), while most others are subcutaneous on their hosts. Several *Philornis* species are found in mainland

Ecuador (Bulgarella *et al.*, 2015, 2017), but *P. downsi* remains the only recorded species of the genus present, to date, in the Galápagos. *P. downsi* is thought to have been introduced from mainland Ecuador, though the full pathway of invasion is not yet known (Fessl *et al.*, 2018). Preliminary population genetics studies show that flies within the archipelago have a high degree of relatedness relative to those on the mainland, which supports the hypothesis of a relatively recent invasion and also the possibility of continued gene flow between populations in the Galápagos (Dudaniec *et al.*, 2008; Koop *et al.*, 2021).

A collaborative research effort has been made to continue monitoring the effects of the avian vampire fly on bird populations in the Galápagos. These efforts aim to identify source population(s), the pathway and mechanism of invasion, and possible long-term mitigation methods, including the Sterile Insect Technique and biocontrol (Causton *et al.*, 2013, 2019). Current stopgap mitigation efforts in the Galápagos include the direct application of insecticide to bird nests. Newly hatched nestlings are removed, and a permethrin solution is sprayed inside the nest and allowed to dry, at which point the nestlings are placed back in the nest. This method has been shown to effectively reduce parasites in the nest and increase fledging success (Koop *et al.*, 2011). A continuation of this method relies on birds incorporating permethrin-treated cotton into their nests, achieving similar increases in fledging success (Knutie *et al.*, 2014). Both methods, with subtle variations, are being used in attempts to increase the nesting success of the critically endangered mangrove finch, currently one of the most endangered birds in the world (Fessl *et al.*, 2018). The same method is also being used successfully to protect endangered bird species against their respective parasites in Australia (Alves *et al.*, 2020).

Despite the robust number of studies that explored the effects of these flies on hosts in the Galápagos, questions remain about the underlying ecological and evolutionary mechanisms of

94 their successful invasion in the Galápagos. This knowledge gap demonstrates the need to develop
95 a full-scale genomic resource of the fly as it provides a critical knowledge base from which to
96 explore these questions, similar to other parasites of concern (Scott *et al.*, 2020). In this study,
97 we generated a high-quality draft genome of the avian vampire fly, which is expected to become
98 an important resource of future molecular studies in this system. We further carried out
99 comparative genomics analysis with additional published dipteran genomes that identified
100 evidence of significantly changing gene families associated with insecticide resistance,
101 detoxification, and possible counter defense against host immune responses. These results serve
102 as the first step toward investigations of this fly's ability to rapidly evolve traits associated with
103 its successful invasion in the Galápagos. From an applied aspect, these genomic resources will
104 also help inform future research studies and mitigation strategies aimed at minimizing the fly's
105 impact on the birds of the Galápagos.
106

107 **Materials and Methods**

108 **Sampling and DNA extraction for genome sequencing**

109 Flies were collected in Jardín de las Opuntias on San Cristobal Island, Galápagos,
110 Ecuador (-0.9491651°, -89.5528454°) in March-April of 2019. Adult flies were reared from
111 pupae collected in the nests of small ground finches (*Geospiza fuliginosa*). When the nests were
112 empty because nestlings died or fledged, all larvae and pupae were extracted from the nest and
113 placed in a falcon tube with a modified lid that allowed airflow. After flies eclosed from their
114 pupal case, they were placed in the freezer, then preserved in 95% ethanol. Preserved flies were
115 transported to the University of Connecticut, then shipped to Northern Illinois University for
116 further processing. DNA was extracted using DNeasy Blood and Tissue Kit (Qiagen, Valencia,

117 California, USA) on whole fly samples after wings and legs were removed using forceps. All
118 samples were treated with Monarch® RNase A (New England Biolabs) to remove RNA from
119 genomic DNA samples. Multiple samples were extracted and those with low yields of DNA or
120 contamination were discarded from further processing. Quantification of double-stranded DNA
121 was done using QuBit® and samples were run on a gel to assess fragmentation.
122
123 **Estimate of genome size and ploidy**
124 We prepared a 10X Chromium GEM library from the extracted DNA above according to
125 the manufacturer's recommended protocols. The library was sequenced using the Illumina 10X
126 platform to generate paired-end 150 bp reads. We used a k-mer based approach to estimate
127 genome size, heterozygosity, and repeat content from unprocessed short sequencing reads
128 (Vurture *et al.*, 2017). We also used the k-mer distribution to extract heterozygous k-mer pairs to
129 assess the ploidy level in *P. downsi* (Ranallo-Benavidez *et al.*, 2020). These estimates of genome
130 size and ploidy were then used to choose parameters for the downstream genome assembly
131 pipeline.
132
133 **Genome assembly and annotation**
134 The 10x Chromium linked reads generated above were used to generate a reference
135 genome using Supernova.v.2.1.1 (Weisenfeld *et al.*, 2017) with default parameters. The genome
136 contiguity statistics such as scaffold N_{50} and the total number of scaffolds etc. were calculated
137 using custom Perl scripts. We further compared the draft genome assembly against a set of
138 conserved genes in Diptera using BUSCO (Waterhouse *et al.*, 2017) to assess gene-space
139 completeness. We also mapped the short sequencing reads back to the draft genome assembly

140 using BWA (Li and Durbin, 2009) to estimate how many short-sequence reads were used to
141 build the reference genome and infer genome completeness. The annotation of the *P. downsi*
142 genome was done using the MAKER annotation pipeline (Cantarel *et al.*, 2008). We combined
143 protein homology evidence (using publicly available protein datasets from major dipteran
144 lineages, downloaded from the Ensemble database (Cunningham *et al.*, 2019) and *ab-initio* gene
145 predictions models to annotate the genome.
146
147 **Transposable elements (TE) in *P. downsi***
148 We combined homology-based and de novo approaches using Repeatmasker (Smit
149 Hubley P., 2013; Flynn *et al.*, 2019) to characterize TEs in *P. downsi*. We used two different sets
150 of repeat libraries, (a) reference repeat library downloaded from the Repbase database (Bao *et*
151 *al.*, 2015) and (b) *de novo* custom-built species-specific repeat library generated for *P. downsi*
152 using RepeatModeler (Flynn *et al.*, 2019). The usage of the *de novo* species-specific repeat
153 library increased the accuracy of detection and annotation of transposable elements. For an
154 unbiased comparison of repeat landscapes among all species, we used similar approaches to
155 detect TEs in the Housefly *(M. domestica)*, Stable fly *(S. calcitrans)*, Tsetse fly *(G. morsitans)*,
156 Mediterranean fruit fly *(C. capitata)*, Fruit fly *(D. melanogaster)* and Yellow fever mosquito *(A.*
157 *aegypti)*, together with an outgroup, Postman butterfly *(H. melpomene)*.
158
159 **Orthologs to other dipteran genomes**
160 We inferred orthogroups and orthologs by comparative analysis of proteomes in *P.*
161 *downsi* and six additional dipteran species (*M. domestica, S. calcitrans, G. morsitans, C.*
162 *capitata, D. melanogaster, and A. aegypti),* together with an outgroup, *H melpomene.* This

163 analysis used the draft proteome of *P. downsi* generated from our genome annotation pipeline
164 described above. Complete proteomes of each of the remaining six species were downloaded
165 from the Ensemble database (Cunningham *et al.*, 2019). The orthologs inference was carried out
166 using Orthofinder (Emms and Kelly, 2015).
167
168 **Gene family evolution**
169 To explore the evolution of gene families in *P. downsi* to other Dipterans, we first
170 constructed a maximum likelihood phylogeny using 3,070 single-copy orthologs among these
171 eight species used for inferring orthologues (Figure 3a). We then used this phylogeny to analyze
172 the changes in gene family size across lineages leading to each of these eight species using a
173 maximum likelihood approach (CAFÉ) (De Bie *et al.*, 2006) that uses a birth and death process
174 to model gene gain and loss across a phylogenetic tree.
175
176 **Results**
177 **Genome sequencing and assembly**
178 We sampled an adult *P. downsi* fly in Jardín de las Opuntias on San Cristobal Island,
179 Galápagos. DNA library was prepared using 10X Chromium linked-read approach (Zheng *et al.*,
180 2016) and sequenced using Illumina 10X platform to generate paired-end 150 bp reads which
181 resulted in ~479 million read pairs (~72 Gb of raw sequence data). We first generated a k-mer
182 distribution based on these short-sequencing reads for preliminary characterization of genome
183 structure in *P. downsi* (e.g., genome size, an abundance of repetitive elements, rate of
184 heterozygosity) (Supplementary Table 1, Supplementary Figure 1) that further allowed us to
185 make informed decisions on parameters needed for building a reference genome. We also

186 utilized the k-mer distribution to estimate the ploidy level in *P. downsi,* which indicated it to be a
187 diploid species (Supplementary Figure 2).
188 Based on genome size estimates using k-mer distributions from short sequence reads
189 (Supplementary Table 1), we had ~ 91X sequence coverage for generating the draft *de-novo*
190 genome assembly. The total estimated length of the draft assembly was 971.6 Mb. The assembly
191 contained 41,176 total scaffolds (minimum 1000 bp to maximum 8.6 Mb) with scaffold N_{50} of
192 1.3 Mb. We further assessed genome contiguity and gene-space completeness using
193 benchmarking universal single-copy orthologs (BUSCO) (Waterhouse *et al.*, 2017). Among
194 3,258 genes highly conserved across Diptera, 3,147 (95.8%) full-length genes were detected in
195 the *P. downsi* draft genome assembly. Partial sequences of 71 genes were identified (2.2%) and
196 only 67 (2.0%) were missing, indicating a high degree of contiguity in the *P. downsi* genome. In
197 addition, 99.16% of total short sequencing reads aligned back to the draft genome indicating a
198 high degree of genome completeness as well.
199
200 **Genome annotation**
201 We combined protein homology-based evidence and *ab-initio* gene prediction models
202 using the MAKER genome annotation pipeline (Cantarel *et al.*, 2008) which identified 15,774
203 protein-coding genes in the *P. downsi* genome. We compared the genome annotation of *P.*
204 *downsi* with two other published and highly curated dipteran genomes, *Musca domestica* (Scott
205 *et al.*, 2014) and *D. melanogaster* (Adams *et al.*, 2000). We annotated fewer genes in the *P.*
206 *downsi* genome (15,774) compared to that of *M. domestica* (17,283) or *D. melanogaster* (17,468)
207 (Table 1). The average length of genes is similar among *P. downsi* and *D. melanogaster,* whereas

208 *M. domestica* typically has longer genes. The mean number of exons per gene is fewer in *P.*

209 *downsi* compared to the other two species.

210 **Table 1: Statistics of genomic features among three fly genomes**

Genome statistics	*P. downsi*	*M. domestica (Scott et al. 2014)*	*D. melanogaster (Adams et al. 2000)*
Genome size (Gb)	0.97	0.69	0.18
Total genes	15,774	17,283	17,468
Avg. gene length (bp)	4,789	13,747	5,830
Total exons	56,595	123,831	188,405
Avg. exon length (bp)	367	453	485
Number of Exons per gene	3.59	7.16	10.79

211

212 **Transposable elements in *P. downsi***

213 Mobile transposable elements (TE) are key features of eukaryotic genomes, being major

214 determinants of genome size variation (Kapusta *et al.*, 2017; Lamichhaney *et al.*, 2021), and

215 important contributors to the evolutionary potential of an organism (Pourrajab and

216 Hekmatimoghaddam, 2021). We characterized the transposable elements in the *P. downsi*

217 genome using homology-based (Smit Hubley P., 2013) and *de-novo* approaches (Flynn *et al.*,

218 2019). More than half of the genome (51.7%) of *P. downsi* consists of transposable elements

219 (Supplementary Table 2). Among these sequences, 9.3% of the genome are retroelements (7.7%

220 LINEs and 1.6% LTRs) and 23.4% DNA transposons. Short interspersed nuclear elements

221 (SINEs), a major category of retroelements, were not detected in the *P. downsi* genome.

222 We also used similar methods to detect, annotate, and compare the repeat content across

223 several other species including the house fly *(Musca domestica)*, stable fly *(Stomoxys calcitrans)*,

224 tsetse fly *(Glossina morsitans)*, Mediterranean fruit fly *(Ceratitis capitata)*, fruit fly *(Drosophila*

225 *melanogaster)* and yellow fever mosquito *(Aedes aegypti)*. The postman butterfly *(Heliconius*

226 *melpomene*) was used as an outgroup. Transposable elements are known to be highly correlated

227 with genome size across the tree of life (Kidwell, 2002; Lynch, 2007), and our results across
228 various dipteran genomes are consistent with this pattern (Figure 2a).
229 We also compared different categories of transposable elements across these seven
230 genomes (Figure 2b). *P. downsi* had the highest proportion of DNA transposons (23.4%) among
231 all taxa analyzed. In comparison, only 6.36% of the *S. calcitrans* genome and 15.96% of the *M.*
232 *domestica* genome consisted of DNA transposons. Although *P. downsi* and *S. calcitrans* have
233 similar genome sizes, *S. calcitrans* had slightly higher amounts of repeat content in its genome
234 (58.3%), compared to *P. downsi* (51.7%) (Figure 2b). *S. calcitrans* had higher amounts of long
235 interspersed nuclear elements (LINEs) (23.8%), compared to *P. downsi* (7.7%). We observed
236 that most Diptera (except *Aedes*) have only a few SINE elements, one of the major classes of
237 transposable elements (Figure 2b). LTR elements were relatively common in *D. melanogaster*,
238 whereas other dipterans had a low amount of LTR elements (including *P. downsi*).
239
240 **Orthologs to other dipteran genomes and outgroup *Heliconius***
241 Comparative genomics analysis of 113,047 protein sequences from *P. downsi* and seven
242 other species (listed above) identified 11,112 orthogroups. A total of 95,567 proteins (out of
243 113,047, 86.3%) were assigned to these orthogroups. The mean size of an orthogroup was 8.8
244 genes/species, and 3,069 orthogroups had singly copy genes in each species. A total of 5,754
245 orthogroups were shared among all eight species. Only a few orthogroups (minimum 496 in *A.*
246 *aegypti* and a maximum of 1,445 in *M. domestica*) were present in fewer than four species
247 (Figure 3a). The number of unique orthologs in each species was consistent with their
248 phylogenetic relationships (e.g., *A. aegypti* and the outgroup *H. melpomene* had the highest
249 number of unique orthogroups). The number of shared orthologs among each pair of species is

250 presented in Supplementary Table 3. The number of orthologs identified in *P. downsi* is
251 consistent with other published dipteran genomes (Figure 3a, Supplementary Table 3), indicating
252 no major bias in the genome assembly and annotation pipeline used in this study.
253 Within *P. downsi*, 13,706 out of 15,774 (86.9%) annotated genes were assigned to
254 orthogroups. We expect the remaining missing genes to either be the most recently evolved
255 orphan genes in the branch leading to the *P. downsi* lineage or the consequence of a lack of
256 inclusion of enough closely related species of *P. downsi* in the analysis. The distribution of these
257 unique genes in the *P. downsi* genome is random and does not show specific clustering patterns
258 across various locations of the genome.
259 We also compared the number of pairwise orthogroups that are uniquely shared among
260 all eight species (Figure 3b). A total of 993 orthogroups were unique to Diptera (after excluding
261 the outlier *H. melpomene),* 79 orthogroups were shared only between *M. domestica* and *P.*
262 *downsi* and 12 putative gene families were unique to only *P. downsi.* These 12 gene families
263 consisted of 27 genes and the great majority had the best BLAST hits against "uncharacterized"
264 or "hypothetical" proteins in other related species (Supplementary Table 4). This result indicates
265 that these gene families that appeared "unique" in *P. downsi* are likely due to the lack of proper
266 gene annotation in other species.
267 We further examined the gene ontology terms of the 993 orthogroups unique to Diptera
268 using the PANTHER gene ontology database (Mi *et al.*, 2021). The common biological
269 processes of these genes included localization, locomotion, immune system processes, response
270 to stimulus, and reproduction (Supplementary Figure 3), many of which are likely key genes for
271 the overall development and function of dipterans.
272

273 **Gene family evolution**

274 We used a maximum likelihood approach to analyze changes in gene family size among
275 the same eight species used for inferring orthologs (De Bie *et al.*, 2006). This method uses a
276 statistical approach to model gene gain and loss, accounting for phylogenetic history and assess
277 the significance of the observed gene family size differences among taxa. Across 11,112
278 orthogroups we identified before among eight taxa; 101 gene families were significantly
279 expanding/contracting ($p < 0.01$) across the phylogenetic tree. 25 out of these 101 gene families
280 were identified in the branch leading to *P. downsi*. The list of these gene families in *P. downsi*
281 included those associated with insecticide resistance or detoxification and host defense or
282 immunity proteins (Table 2).

283

284 **Table 2:** *List of* significantly expanding/contracting ($p < 0.01$) *gene families in P. downsi*

Functional Categories	Gene Family
Insecticide Resistance and Detoxification	Cytochrome P450
	Glutathione S Transferase
	Cuticular Protein
Defense against host Immunity	Fibrinogen C-terminal Domain-Containing Protein
	Scp Domain-Containing Protein

285

286 **Gene families associated with insecticide resistance and detoxification**

287 *Cytochrome P450 gene family*

288 We examined the number of genes in the cytochrome P450 (CYP450) family across the
289 seven Diptera and the *H. melpomene* outgroup. *P. downsi and M. domestica* have an expanded
290 CYP450 gene family in comparison to their most recent common ancestor with *G. morsitans*
291 (family Glossinidae) (Figure 4a). For example, The *P. downsi* CYP450 family is composed of
292 102 genes in comparison to 66 in *G. morsitans.* An even greater level of expansion was observed
293 in two other members of Muscidae *(M. domestica:* 143 genes and *S. calcitrans:* 193 genes).

294 Compared to *G. morsitans*, the expansion of CYP450 genes is mainly found in CYP4, CYP6,
295 and CYP28 genes (Figure 4b). The CYP6 subfamily in *P. downsi* is composed of 25 genes,
296 almost doubling the number from the 14 genes present in *G. morsitans.* We also found an
297 expansion in the CYP4 subfamily from 15 genes in *G. morsitans* to 31 genes in *P. downsi.* A
298 similar expansion was seen in the CYP28 subfamily (4 genes in *G. morsitans* to 8 genes in *P.*
299 *downsi*).
300
301 *Glutathione S-transferase gene family*
302 The *P. downsi* Glutathione S-transferase family consists of 25 genes and the pattern
303 across the dipterans is similar to observations of the Cytochrome P450 gene family expansion
304 (Figure 4c). For example, in comparison to *G. morsitans* (15 genes), *P. downsi and M. domestica*
305 have an expanded number of genes (25 genes in *P. downsi,* 31 genes in *M. domestica*). The
306 GSTs are grouped into six subclasses (Delta, Epsilon, Omega, Sigma, Theta, and Zeta). The
307 expansion of GSTs in *P. downsi* occurred mainly in the Delta, Epsilon, and Zeta subclasses
308 (Figure 4d).
309
310 *Cuticular Protein*
311 The *P. downsi* cuticular gene family consists of 214 genes and the pattern across the
312 dipterans is similar to observations of the Cytochrome P450 and GST gene family expansions
313 (Supplementary Figure 4). For example, the *Muscidae* family (represented here by *P. downsi, M.*
314 *domestica, and S. calcitrans)* has an expanded gene family in comparison to their most recent
315 common ancestor with *G. morsitans* (115 genes). An even greater level of expansion was

316 observed by two other members of *Muscidae (M. domestica* – 357 genes and *S. calcitrans* – 270
317 genes).
318
319 **Gene families associated with immunity**
320 Two gene families (Fibrinogen C-terminal Domain-Containing Protein and Scp Domain-
321 Containing Protein) associated with immunity were also identified as significantly changing in P.
322 downsi (Table 2). Opposite of genes associated with insecticide resistance and detoxification
323 (Cytochrome P450 and Glutathione S-transferase), these gene families showed a reduction in the
324 number of genes in *P. downsi* compared to the most recent common ancestor with *G. morsitans*
325 (Supplementary Table 5). They also show contraction in comparison with two included members
326 of the Family Muscidae *(M. domestica and S. calcitrans).*
327
328 **Discussion**
329 In this study, we report the first genome sequence of the avian vampire fly, a highly
330 invasive parasitic nest fly that threatens endemic avifauna of the Galápagos Islands (Fessl *et al.*,
331 2018; Causton *et al.*, 2019). This genome is meant to serve as an important resource to research
332 efforts aimed at characterizing the molecular mechanisms of the fly's successful invasion in the
333 Galápagos. The genome size of *P. downsi* is 971.6 Mb and its high quality is reflected both in
334 terms of genome contiguity (scaffold N_{50} of 1.3 Mb) and completeness (98% BUSCO gene-space
335 score). Interestingly, the total number of annotated genes in *P. downsi* is slightly lower compared
336 to other published fly genomes (*M. domestica* and *D. melanogaster*) (Table 1). The low gene
337 count likely reflects the lack of *P. downsi* specific transcriptome data in this study, which
338 perhaps led to reduced gene predictions. We aim to improve the *P. downsi* genome annotations

339 in the future using additional transcriptomic resources using additional flies collected from the
340 Galápagos.
341 Transposable elements (TE) are typically non-coding sequences that can insert themselves in
342 various places of the genome, often with neutral or deleterious phenotypic consequences
343 (Bourque *et al.*, 2018). The role of TEs, as well as their evolution across insect genomes, is still
344 an area of major research, but they are thought to be important drivers of genomic architecture
345 depending on the location of the genome to which they insert themselves (i.e., coding versus
346 non-coding regions). Furthermore, TE may also be a critical mechanism of adaptive evolution, as
347 has been shown in an invasive ant species (Schrader *et al.*, 2014). Analysis of transposable
348 elements in *P. downsi* and across other dipteran genomes showed a strong positive correlation
349 between genome size and repeat content (Figure 2a), consistent with similar findings across other
350 taxa (Lynch, 2007; Lamichhaney *et al.*, 2021). Interestingly, *P. downsi* had a higher number of
351 DNA transposons (Class II TEs) than any other compared genome, including *M. domestica, S.*
352 *calcitrans,* and *G. morsitans*. While long terminal repeat (LTR) transposons, LINEs, and SINEs)
353 were present in the species studied, *P. downsi* had no SINEs, a finding consistent with a study by
354 Petersen and colleagues (Petersen *et al.*, 2019) showing that SINEs contribute less than 1% to the
355 TE content of dipterans. However, it is important to note that some SINEs may be present in *P.*
356 *downsi* but are currently masked as unclassified. Future research should explore the role of TEs,
357 especially DNA transposons, in aiding the invasion of *P. downsi* to the Galápagos.
358 Comparative genomics analysis of *P. downsi* and other additional dipteran genomes
359 allowed us to identify gene families that were significantly expanded/contracted ($p < 0.01$) in *P.*
360 *downsi*. The list of these gene families in *P. downsi* included those associated with insecticide
361 resistance or detoxification and host defense or immunity proteins, and we predict that these

362 gene families are associated with a successful invasion of *P. downsi* in the Galápagos (Table 1).
363 E.g. CYP450 mono-oxygenases are a diverse superfamily of proteins, including enzymes,
364 associated with catabolism and anabolism of xenobiotics and endogenous compounds. These
365 monooxygenase-mediated metabolisms have allowed numerous insect species to develop
366 insecticide resistance and detoxification (Scott, 1999; Wen *et al.*, 2011). The expansion of
367 CYP450 genes in *P. downsi* is mainly found in CYP4 and CYP6 gene subfamilies (Figure 4b),
368 which has also been shown in other species from the family Muscidae (Scott *et al.*, 2014;
369 Olafson *et al.*, 2021). Interestingly, in higher Diptera, many of the genes within the CYP6
370 subfamily (e.g., CYP6A, CYP6G, CYP6D) are associated with insecticide resistance
371 (Feyereisen, 2012). For example, the Cyp6g1 gene is involved in resistance to the insecticide
372 Dichlorodiphenyltrichloroethane (DDT) in *D. melanogaster* (Festucci-Buselli *et al.*, 2005). The
373 CYP4 subfamily can also influence the breakdown of synthetic insecticides (Iga and Kataoka,
374 2012). Cyp4d4v2, Cyp4g2, and Cyp6a38 can be co-up-regulated in-house flies that are resistant
375 to the insecticide permethrin, which is used to control *P. downsi* in the Galapagos (Zhu *et al.*,
376 2008). Overall, the expansion of CYP4 and CYP6 subfamilies may indicate the evolution of
377 insecticide resistance in *P. downsi* over macroevolutionary time (i.e., before arriving in the
378 Galápagos), which could have facilitated its invasion to the Galápagos and might affect its
379 management on the islands.
380 Glutathione S-transferase (GSTs) was another major expanding gene family in *P. downsi.*
381 GSTs are a highly conserved, large family of dimeric enzymes associated with detoxification of
382 endogenous and/or xenobiotic compounds, such as insecticides (Ketterman *et al.*, 2011). The
383 GST family is further grouped into six subclasses (Delta, Epsilon, Omega, Sigma, Theta, and
384 Zeta), with Delta and Epsilon being specific subclasses found in the class Insecta. We observed

385 an expansion of GST genes in *P. downsi* relative to *G. morsitans*, but fewer than were found in
386 *M. domestica and S. calcitrans*. The major expansions of the GST family in *P. downsi* were
387 observed in Delta and Epsilon subclasses (Figure 4d). Insecticides, such as permethrin, are used
388 to experimentally manipulate *P. downsi* abundance in bird nests to study the effects of the
389 parasite on the health of the birds (Fessl *et al.*, 2010; Koop *et al.*, 2011; Koop, Le Bohec, *et al.*,
390 2013; Koop, Owen, *et al.*, 2013; Knutie *et al.*, 2014, 2016b; O'Connor *et al.*, 2014; McNew *et*
391 *al.*, 2020; Addesso *et al.*, 2020). Increased expression of GSTs following permethrin exposure
392 has been documented in several insect species including oriental fruit flies (*Bactrocera dorsalis*)
393 (Hu *et al.*, 2008). Expansion of the GST gene family in *P. downsi* may be a result of such
394 permethrin exposure. However, gene expression studies that further explore the role of the GST
395 gene family in insecticide resistance and detoxification are needed across insect taxa including *P.*
396 *downsi*. It is important to note that the expansion of CYP450 and GST families observed in *P.*
397 *downsi* may be an artifact of phylogenetic relationships rather than ecological adaptations. Still,
398 given the observed and predicted impacts of *P. downsi* on native endemic host populations, it is
399 important to consider the implications of expanded gene families related to detoxification and
400 insecticide resistance.

401 Previous studies have shown that Darwin's finch species can increase specific antibody
402 responses to parasitism (Koop, Owen, *et al.*, 2013). This response is most prominent in adult
403 females that are likely parasitized while brooding nestlings or eggs on the nest. However, little is
404 known about the ability of host bird immunological responses to effectively reduce fly fitness, in
405 part, because so little is known about the fly itself. One of the most prominent questions is
406 whether the *P. downsi* possesses the ability to counter defend against host immune responses.
407 We identified a reduction in the size of two additional gene families (Fibrinogen C-terminal

408 Domain-Containing and SCP domain-containing gene family), both with immune function
409 properties in *P.downsi* (Table 1, Supplementary Table 5). Fibrinogen plays a key role in blood
410 clot formation through the conversion of fibrinogen to insoluble fibrin (Weisel and Litvinov,
411 2017) and the C-terminal domain of fibrinogen is the primary binding site of platelets
412 (Hanington and Zhang, 2011). The Sperm-coating glycoprotein (Scp) family contains, among
413 other proteins, antigen 5 (Ag5), which is associated with the venom secretory ducts of stinging
414 insects (Gibbs *et al.*, 2008). However, the interpretation of these results is difficult without
415 further investigation into whether these genes are associated with innate immune responses of *P.*
416 *downsi* toward their hosts or whether they might be important components of their feeding
417 ecology. The goal of our study was not to make such inferences, but rather to highlight
418 promising avenues of future research.
419 The invasion of *P. downsi* has had dramatic negative effects on the endemic avifauna of
420 the Galápagos, including Darwin's finches. As researchers work to better understand the
421 pathway of invasion and the ecological and evolutionary processes that may have facilitated its
422 invasion to the Galápagos, the need for a high-quality whole genome sequence has grown. The
423 addition of this resource is therefore meant to provide the foundation for further investigations
424 using genomics tools in this system. These genomic resources will further allow us to understand
425 the evolution of the *P. downsi* defense in response to host defenses or disease resistance itself.
426 Gene expression studies could shed light on the development of larvae and at what stage they are
427 most vulnerable to host defenses. Further population-scale resequencing of various populations
428 of *P. downsi* will also allow us to explore mechanisms of local adaptation of the parasite to the
429 environment across islands.
430

Data availability

All raw data generated in this study (raw short sequence reads and draft genome assembly) has been deposited to NCBI, under accession number PRJNAXXX. The final genome assembly and annotation can be found under the accession number GCA_XXX. Supplementary material is available on figshare.

Acknowledgment

We would like to thank Lauren Albert, Taylor Verrett, and Corrine Arthur for field assistance and the Galápagos Science Center and the Galápagos National Park for logistical support. We also thank Noah K. Whiteman for his helpful comments on the manuscript. The sampling was done under Galápagos National Parks permits PC 28-19 and Genetic Access permit MAE-DNB-CM-2016-0041. This project was supported by the Department of Biological Sciences, Kent State University to SL, and the University of Connecticut to SAK.

Competing interest statement

The authors declare no competing interests.

Author contributions

SL and JAHK conceived the idea. SAK, GJV, and JC collected the specimens. CMC and JAHK carried out the laboratory work. MR and SL designed and performed the bioinformatic analyses with the support of CMC and JAHK. SL and JAHK prepared the manuscript, and all authors edited and approved the final version.

455

456 **List of Figures**

457 **Figure 1:** The avian vampire fly, Philornis downsi **(a)**, parasitizes many endemic bird species of
458 the Galapagos Islands, including the medium ground finch, Geospiza fortis **(b)**. The fly is
459 parasitic in its larval forms **(c**, bottom three) when it feeds on the blood and other fluids of its
460 avian hosts. The larva then pupates **(c**, second from top) and eclose **(c**, top) as adult flies.

461

462 **Figure 2:** Landscape of Transposable elements in *P. downsi* **(a)** Comparison of repeat content
463 and genome size across Diptera and its outgroup **(b)** Repeat statistics on various classes of
464 transposable elements across dipteran genomes.

465

466 **Figure 3:** Orthogroups in *P. downsi* **(a)** Phylogenetic relationship between *P. downsi* and other
467 seven published Diptera genomes, estimated using alignments from 3,069 orthogroups had
468 singly copy orthogroups in each species. Horizontal bars for each species show number of
469 orthogroups that are single-copy orthologs in all species, present in all species, present in the
470 majority of species, present in few species, and unique to the species **(b)** Number of shared
471 orthologs among all species.

472

473 **Figure 4:** Gene family evolution **(a)** Number of genes in Cytochrome P450 gene family across
474 Diptera **(b)** Number of genes in various subfamilies of Cytochromes P450 gene family in *G.*
475 *morsitans* and *P. downsi, M. domestica* and *D. melanogaster* **(c)** Number of genes in Glutathione
476 S-transferase gene family across Diptera **(d)** Number of genes in various subfamilies of

477 Glutathione S-transferase gene family in *G. morsitans* and *P. downsi, M. domestica* and *D.*

478 *melanogaster.*

479

480 **References**

481 Adams MD, Celniker SE, Holt RA, Evans CA, Gocayne JD, Amanatides PG, *et al.* (2000). The
482 genome sequence of Drosophila melanogaster. *Science* **287**: 2185–95.

483 Addesso AM, Harvey JA, Vaziri GJ, Verrett TB, Albert L, Arthur C, *et al.* (2020). Effect of
484 introduced parasites on the survival and microbiota of nestling cactus finches (Geospiza
485 scandens) in the Galápagos Islands. *Journal of Ornithology* **161**: 1011–1019.

486 Alves F, Langmore N, Heinsohn R, Stojanovic D (2020). 'Self-fumigation' of nests by an
487 endangered avian host using insecticide-treated feathers increases reproductive success
488 more than tenfold. *Animal Conservation*.

489 Bao W, Kojima KK, Kohany O (2015). Repbase Update, a database of repetitive elements in
490 eukaryotic genomes. *Mobile DNA* **6**: 11.

491 Bourque G, Burns KH, Gehring M, Gorbunova V, Seluanov A, Hammell M, *et al.* (2018). Ten
492 things you should know about transposable elements. *Genome Biol* **19**: 199–199.

493 Bulgarella M, Quiroga MA, Boulton RA, Ramirez IE, Moon RD, Causton CE, *et al.* (2017). Life
494 Cycle and Host Specificity of the Parasitoid Conura annulifera (Hymenoptera:
495 Chalcididae), a Potential Biological Control Agent of Philornis downsi (Diptera:
496 Muscidae) in the Galapagos Islands. *Ann Entomol Soc Am* **110**: 317–328.

497 Bulgarella M, Quiroga MA, Brito vera GA, Dregni JS, Cunningham F, Mosquera Munoz DA, *et*
498 *al.* (2015). Philornis downsi (Diptera: Muscidae), an avian nest parasite invasive to the
499 Galapagos islands, in mainland Ecuador. *Annals of the Entomological Society of*
500 *America*.

501 Bulgarella M, Quiroga MA, Heimpel GE (2019). Additive negative effects of Philornis nest
502 parasitism on small and declining Neotropical bird populations. *Bird Conservation*
503 *International* **29**: 339–360.

504 Cantarel BL, Korf I, Robb SMC, Parra G, Ross E, Moore B, *et al.* (2008). MAKER: an easy-to-
505 use annotation pipeline designed for emerging model organism genomes. *Genome Res*
506 **18**: 188–196.

507 Causton CE, Cunninghame F, Tapia W (2013). *Management of the avian parasite Philornis*
508 *downsi in the Galapagos Islands: a collaborative and strategic action plan*. GNPS,
509 GCREG, CDF, and GC: Puerto Ayora, Galapagos, Ecuador.

510 Causton CE, Moon RD, Cimadom A, Boulton RA, Cedeño D, Lincango MP, *et al.* (2019).
511 Population dynamics of an invasive bird parasite, Philornis downsi (Diptera: Muscidae),
512 in the Galapagos Islands. *PLoS One* **14**: e0224125.

513 Causton CE, Peck SB, Sinclair BJ, Roque-Albelo L, Hodgson CJ, Landry B (2006). Alien
514 insects: Threats and implications for conservation of Galápagos Islands. *Annals of the*
515 *Entomological Society of America* **99**: 121–143.

516 Couri MS, de Carvalho CJB, Lowenberg-Neto P (2007). Phylogeny of Philornis Meinert species
517 (Diptera: Muscidae). *Zootaxa*: 19–26.

518 Cunningham F, Achuthan P, Akanni W, Allen J, Amode MR, Armean IM, *et al.* (2019).
519 Ensembl 2019. *Nucleic Acids Research* **47**: D745–D751.

520 Cunninghame F, Fessl B, Sevilla CR, Young GR, La Greco N (2017). *Manejo de la*
521 *conservacion a largo plazo para salvar al pinzon de manglar (Camarhynchus heliobates)*
522 *en peligro critico de extincion.* DPNG, CGREG, FCD and GC: Puerto Ayora, Galapagos,
523 Ecuador.

524 De Bie T, Cristianini N, Demuth JP, Hahn MW (2006). CAFE: a computational tool for the
525 study of gene family evolution. *Bioinformatics (Oxford, England)* **22**: 1269–1271.

526 Dodge HR (1955). New Muscid flies from Florida and the West Indies (Diptera: Muscidae). *The*
527 *Florida Entomologist* **38**: 147–151.

528 Dodge HR (1963). A new Philornis with coprophagous larva, and some related species (Diptera:
529 Muscidae). *Journal of the Kansas Entomological Society* **36**: 239–247.

530 Dudaniec RY, Fessl B, Kleindorfer S (2007). Interannual and interspecific variation in intensity
531 of the parasitic fly, Philornis downsi, in Darwin's finches. *Biological Conservation* **139**:
532 325–332.

533 Dudaniec RY, Gardner MG, Donnellan S, Kleindorfer S (2008). Genetic variation in the invasive
534 avian parasite, Philornis downsi (Diptera, Muscidae) on the Galapagos archipelago. *BMC*
535 *Ecology* **8**: 13.

536 Dudaniec RY, Kleindorfer S (2006). Effects of the parasitic flies of the genus Philornis (Diptera :
537 Muscidae) on birds. *Emu* **106**: 13–20.

538 Dvorak M, Vargas H, Fessl B, Tebbich S (2004). On the verge of extinction: a survey of the
539 mangrove finch Cactospiza heliobates and its habitat on the Galápagos Islands. *Oryx* **38**:
540 1–9.

541 Emms DM, Kelly S (2015). OrthoFinder: solving fundamental biases in whole genome
542 comparisons dramatically improves orthogroup inference accuracy. *Genome Biology* **16**:
543 157.

544 Fessl B, Heimpel G, Causton C (2018). Invasion of an avian nest parasite, Philornis downsi, to
545 the Galapagos Islands: colonization history, adaptations to novel ecosystems, and
546 conservation challenges. In: Parker P (ed) *Disease Ecology, Social and Ecological*
547 *Interactions in the Galapagos Islands*, Springer International Publishing.

548 Fessl B, Sinclair BJ, Kleindorfer S (2006). The life-cycle of Philornis downsi (Diptera :
549 Muscidae) parasitizing Darwin's finches and its impacts on nestling survival.
550 *Parasitology* **133**: 739–747.

551 Fessl B, Tebbich S (2002). Philornis downsi - a recently discovered parasite on the Galápagos
552 archipelago - a threat for Darwin's finches? *Ibis* **144**: 445–451.

553 Fessl B, Young GH, Young RP, Rodríguez-Matamoros J, Dvorak M, Tebbich S, *et al.* (2010).
554 How to save the rarest Darwin's finch from extinction: the mangrove finch on Isabela
555 Island. *Philos Trans R Soc Lond B Biol Sci* **365**: 1019–1030.

556 Festucci-Buselli RA, Carvalho-Dias AS, de Oliveira-Andrade M, Caixeta-Nunes C, Li HM,
557 Stuart JJ, *et al.* (2005). Expression of Cyp6g1 and Cyp12d1 in DDT resistant and
558 susceptible strains of Drosophila melanogaster. *Insect Mol Biol* **14**: 69–77.

559 Feyereisen R (2012). Insect CYP genes and P450 enzymes. In: Gilbert LI, editor. *Insect*
560 *Molecular Biology and Biochemistry*: 236–316.

561 Flynn JM, Hubley R, Goubert C, Rosen J, Clark AG, Feschotte C, *et al.* (2019).
562 RepeatModeler2: automated genomic discovery of transposable element families.
563 *bioRxiv*: 856591.

564 Gibbs GM, Roelants K, O'Bryan MK (2008). The CAP Superfamily: Cysteine-Rich Secretory
565 Proteins, Antigen 5, and Pathogenesis-Related 1 Proteins—Roles in Reproduction,
566 Cancer, and Immune Defense. *Endocrine Reviews* **29**: 865–897.

567 Grant PR, Grant BR, Petren K, Keller LF (2005). Extinction behind our backs: the possible fate
568 of one of the Darwin's finch species on Isla Floreana, Galápagos. *Biological*
569 *Conservation* **122**: 499–503.

570 Hanington PC, Zhang S-M (2011). The primary role of fibrinogen-related proteins in
571 invertebrates is defense, not coagulation. *J Innate Immun* **3**: 17–27.

572 Heimpel GE, Hillstrom A, Freund D, Knutie SA, Clayton DH (2017). Invasive Parasites and the
573 Fate of Darwin's Finches in the Galapagos Islands: The Case of the Vegetarian Finch
574 (Platyspiza crassirostris). *Wilson J Ornithol* **129**: 345–349.

575 Hu J, Zhang JL, Nardi F, Zhang RJ (2008). Population genetic structure of the melon fly,
576 Bactrocera cucurbitae (Diptera: Tephritidae), from China and Southeast Asia. *Genetica*
577 **134**: 319–24.

578 Iga M, Kataoka H (2012). Recent Studies on Insect Hormone Metabolic Pathways Mediated by
579 Cytochrome P450 Enzymes. *Biological and Pharmaceutical Bulletin* **35**: 838–843.

580 Kapusta A, Suh A, Feschotte C (2017). Dynamics of genome size evolution in birds and
581 mammals. *Proceedings of the National Academy of Sciences* **114**: E1460 LP-E1469.

582 Ketterman AJ, Saisawang C, Wongsantichon J (2011). Insect glutathione transferases. *Drug*
583 *Metabolism Reviews* **43**: 253–265.

584 Kidwell MG (2002). Transposable elements and the evolution of genome size in eukaryotes.
585 *Genetica* **115**: 49–63.

586 Knutie SA, McNew SM, Bartlow AW, Vargas DA, Clayton DH (2014). Darwin's finches
587 combat introduced nest parasites with fumigated cotton. *Current Biology* **24**: R355–
588 R356.

589 Knutie SA, Owen JP, McNew SM, Bartlow AW, Arriero E, Herman JM, *et al.* (2016a).
590 Galapagos mockingbirds tolerate introduced parasites that affect Darwin's finches.
591 *Ecology* **97**: 940–950.

592 Koop JAH, Causton CE, Bulgarella M, Cooper E, Heimpel GE (2021). Population structure of a
593 nest parasite of Darwin's finches within its native and invasive ranges. *Conservation*
594 *Genetics* **22**: 11–22.

595 Koop JAH, Huber SK, Laverty SM, Clayton DH (2011). Experimental demonstration of the
596 fitness consequences of an introduced parasite of Darwin's finches. *PLoS ONE* **6**: e19706
597 doi:10.1371/journal.pone.0019706.

598 Koop JAH, Kim PS, Knutie SA, Adler F, Clayton DH (2015). Introduced parasitic fly may lead
599 to local extinction of Darwin's finch populations. *Journal of Applied Ecology*.

600 Koop JAH, Le Bohec C, Clayton DH (2013). Dry year does not reduce invasive parasitic fly
601 prevalence or abundance in Darwin's finch nests. *Reports in Parasitology* **3**: 11–17.

602 Koop JAH, Owen JP, Knutie SA, Aguilar MA, Clayton DH (2013). Experimental demonstration
603 of a parasite-induced response in wild birds: Darwin's finches and introduced nest flies.
604 *Ecology and Evolution* **3**: 2514–2523.

605 Lamichhaney S, Catullo R, Keogh JS, Clulow S, Edwards SV, Ezaz T (2021). A bird-like
606 genome from a frog: Mechanisms of genome size reduction in the ornate burrowing frog,
607 Platyplectrum ornatum. *Proceedings of the National Academy of Sciences* **118**:
608 e2011649118.

609 Li H, Durbin R (2009). Fast and accurate short read alignment with Burrows–Wheeler transform.
610 *Bioinformatics* **25**: 1754–1760.

611 Lynch M (2007). *The Origins of Genome Architecture*, 1 edition. Sinauer Associates Inc:
612 Sunderland.

613 McNew SM, Knutie SA, Clayton DH (2020). No evidence of sex ratio manipulation by
614 Galápagos mockingbirds in response to environment. *Journal of Avian Biology* **51**.

615 Mi H, Ebert D, Muruganujan A, Mills C, Albou L-P, Mushayamaha T, *et al.* (2021). PANTHER
616 version 16: a revised family classification, tree-based classification tool, enhancer regions
617 and extensive API. *Nucleic Acids Research* **49**: 394–403.

618 O'Connor JA, Robertson J, Kleindorfer S (2014). Darwin's finch begging intensity does not
619 honestly signal need in parasitised nests. *Ethology* **120**: 228–237.

620 Olafson PU, Aksoy S, Attardo GM, Buckmeier G, Chen X, Coates CJ, *et al.* (2021). The genome
621 of the stable fly, Stomoxys calcitrans, reveals potential mechanisms underlying
622 reproduction, host interactions, and novel targets for pest control. *BMC Biology* **19**: 41.

623 Peters KJ, Kleindorfer S (2018). Avian population trends in Scalesia forest on Floreana Island
624 (2004-2013): Acoustical surveys cannot detect hybrids of Darwin's tree finches
625 Camarhynchus spp. *Bird Conservation International* **28**: 319–335.

626 Petersen M, Armisén D, Gibbs RA, Hering L, Khila A, Mayer G, *et al.* (2019). Diversity and
627 evolution of the transposable element repertoire in arthropods with particular reference to
628 insects. *BMC Evol Biol* **19**: 11–11.

629 Pourrajab F, Hekmatimoghaddam S (2021). Transposable elements, contributors in the evolution
630 of organisms (from an arms race to a source of raw materials). *Heliyon* **7**: e06029–
631 e06029.

632 Ranallo-Benavidez TR, Jaron KS, Schatz MC (2020). GenomeScope 2.0 and Smudgeplot for
633 reference-free profiling of polyploid genomes. *Nature Communications* **11**: 1432.

634 Schrader L, Kim JW, Ence D, Zimin A, Klein A, Wyschetzki K, *et al.* (2014). Transposable
635 element islands facilitate adaptation to novel environments in an invasive species. *Nature*
636 *Communications* **5**: 5495.

637 Scott JG (1999). Cytochromes P450 and insecticide resistance. *Insect Biochem Mol Biol* **29**:
638 757–77.

639 Scott MJ, Benoit JB, Davis RJ, Bailey ST, Varga V, Martinson EO, *et al.* (2020). Genomic
640 analyses of a livestock pest, the New World screwworm, find potential targets for genetic
641 control programs. *Communications Biology* **3**: 424–424.

642 Scott JG, Warren Wc Fau - Beukeboom LW, Beukeboom Lw Fau - Bopp D, Bopp D Fau - Clark
643 AG, Clark Ag Fau - Giers SD, Giers Sd Fau - Hediger M, *et al.* (2014). Genome of the
644 house fly, Musca domestica L., a global vector of diseases with adaptations to a septic
645 environment.

646 Smit Hubley P. AFA R& Green (2013). RepeatMasker Open-4.0,.

647 Spalding MG, Mertins JW, Walsh PB, Morin KC, Dunmore DE, Forrester DJ (2002). Burrowing
648 fly larvae (Philornis porteri) associated with mortality of eastern bluebirds in Florida.
649 *Journal of Wildlife Diseases* **38**: 776–783.

650 Vurture GW, Sedlazeck FJ, Nattestad M, Underwood CJ, Fang H, Gurtowski J, *et al.* (2017).
651 GenomeScope: fast reference-free genome profiling from short reads. *Bioinformatics* **33**:
652 2202–2204.

653 Waterhouse RM, Seppey M, Simao FA, Manni M, Ioannidis P, Klioutchnikov G, *et al.* (2017).
654 BUSCO applications from quality assessments to gene prediction and phylogenomics.
655 *Molecular biology and evolution*.

656 Weisel JW, Litvinov RI (2017). Fibrin Formation, Structure and Properties. *Subcell Biochem* **82**:
657 405–456.

658 Weisenfeld NI, Kumar V, Shah P, Church DM, Jaffe DB (2017). Direct determination of diploid
659 genome sequences. *Genome Research* **27**: 757–767.

660 Wen Z, Zhang X, Zhang Y (2011). P450—mediated Insecticide Detoxification and Its
661 Implication in Insecticide Efficacy. In: Liu T, Kang L (eds) *Recent Advances in*
662 *Entomological Research: From Molecular Biology to Pest Management*, Springer Berlin
663 Heidelberg: Berlin, Heidelberg, pp 229–245.

664 Zheng GXY, Lau BT, Schnall-Levin M, Jarosz M, Bell JM, Hindson CM, *et al.* (2016).
665 Haplotyping germline and cancer genomes with high-throughput linked-read sequencing.
666 *Nature Biotechnology* **34**: 303–311.

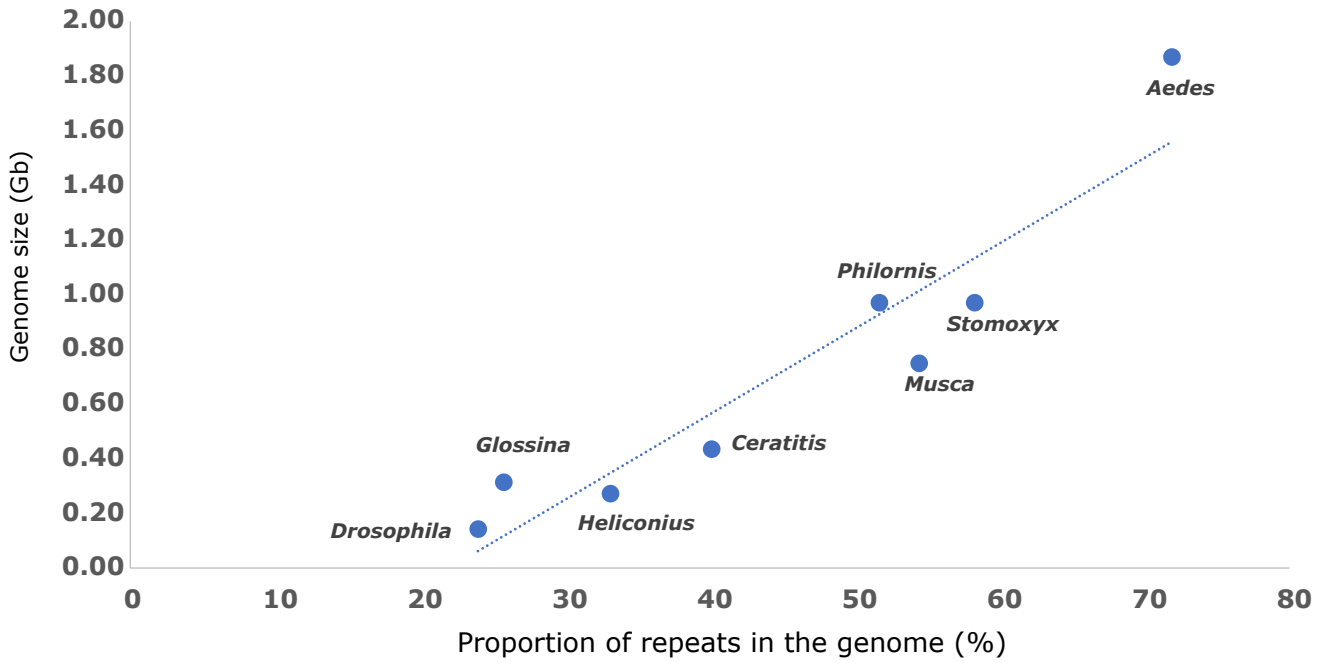
667 Zhu F, Li T, Zhang L, Liu N (2008). Co-up-regulation of three P450 genes in response to
668 permethrin exposure in permethrin resistant house flies, Musca domestica. *BMC Physiol*
669 **8**: 18.

670

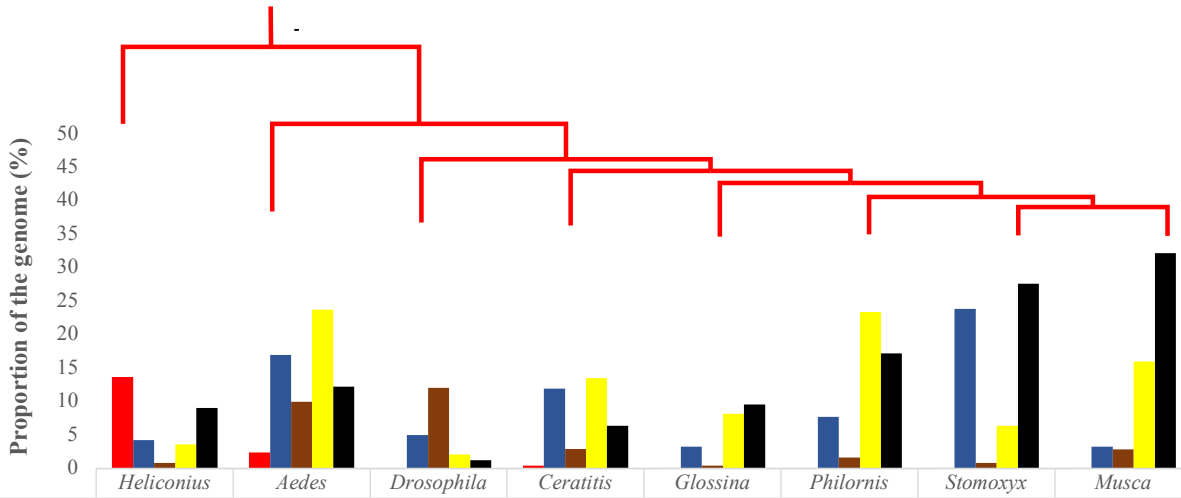
671

Figure 1

a



b



	Heliconius	Aedes	Drosophila	Ceratitis	Glossina	Philornis	Stomoxyx	Musca
SINE	13.67	2.36	0	0.41	0.01	0	0.02	0.02
LINE	4.22	16.92	4.95	11.91	3.22	7.67	23.81	3.23
LTR elements	0.81	9.97	12.05	2.89	0.43	1.62	0.82	2.82
DNA transposons	3.59	23.72	2.08	13.47	8.15	23.39	6.36	15.96
Unclassified	9	12.18	1.2	6.34	9.53	17.15	27.6	32.15

Figure 2

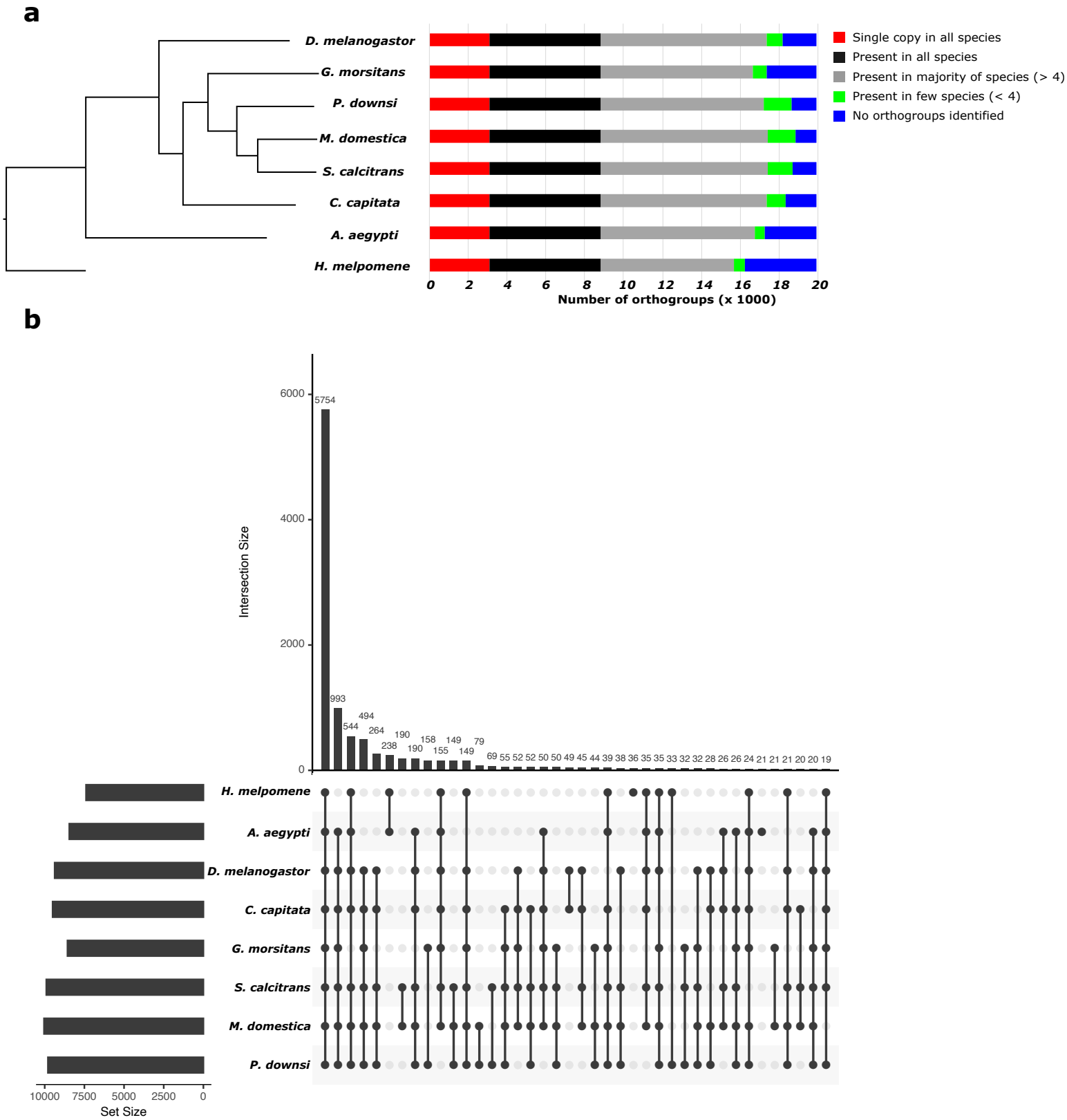

a
D. melanogastor
G. morsitans
P. downsi
M. domestica
S. calcitrans
C. capitata
A. aegypti
H. melpomene
0 2 4 6 8 10 12 14 16 18 20
Number of orthogroups (x 1000)
Single copy in all species
Present in all species
Present in majority of species (> 4)
Present in few species (< 4)
No orthogroups identified
b
Intersection Size
6000
4000
2000
0
5754 993 544 494 264 238 190 190 158 155 149 149 79 69 55 52 52 50 50 49 45 44 39 38 36 35 35 33 32 32 28 26 26 24 21 21 21 20 20 19
H. melpomene
A. aegypti
D. melanogastor
C. capitata
G. morsitans
S. calcitrans
M. domestica
P. downsi
10000 7500 5000 2500 0
Set Size

Figure 3

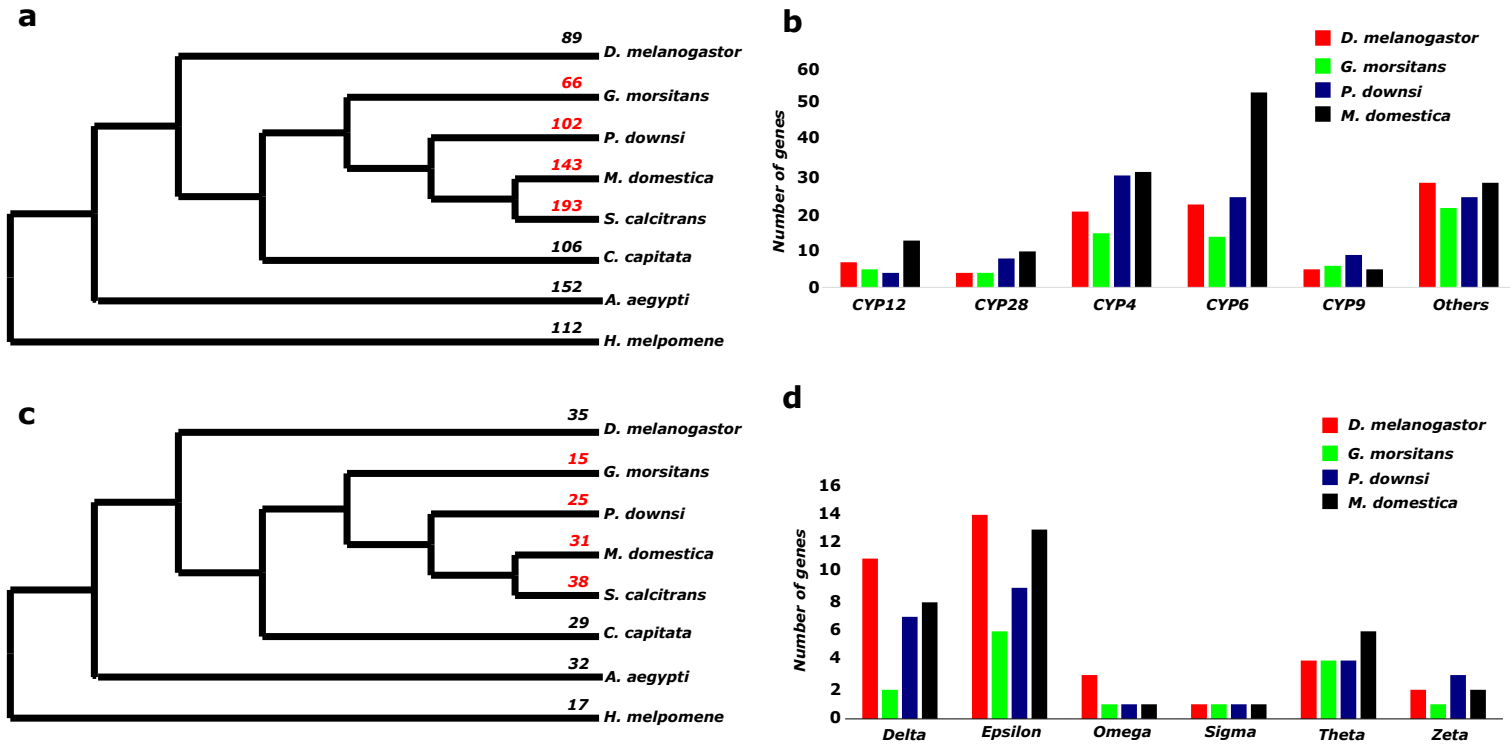

a
89
D. melanogastor
66
G. morsitans
102
P. downsi
143
M. domestica
193
S. calcitrans
106
C. capitata
152
A. aegypti
112
H. melpomene
b
D. melanogastor
G. morsitans
P. downsi
M. domestica
Number of genes
60
50
40
30
20
10
0
CYP12
CYP28
CYP4
CYP6
CYP9
Others
c
35
D. melanogastor
15
G. morsitans
25
P. downsi
31
M. domestica
38
S. calcitrans
29
C. capitata
32
A. aegypti
17
H. melpomene
d
D. melanogastor
G. morsitans
P. downsi
M. domestica
Number of genes
16
14
12
10
8
6
4
2
0
Delta
Epsilon
Omega
Sigma
Theta
Zeta

Figure 4

Supplementary Materials

The genome sequence of the avian vampire fly (Philornis downsi), an invasive nest parasite of Darwin's finches

Supplementary Table 1: *Genome Statistics of P. downsi estimated from short-sequencing reads*

Property	***min***	***max***
Heterozygosity	1.40879%	1.42194%
Genome Haploid Length	796,797,589 bp	799,050,413 bp
Genome Repeat Length	232,405,721 bp	233,062,812 bp
Genome Unique Length	564,391,869 bp	565,987,600 bp
Model Fit	95.7713%	98.7012%
Read Error Rate	1.228%	1.228%

Supplementary Table 2: *Repeat statistics in P. downsi genome*

sequences: 41,176
total length: 971,6346,46 bp
GC level: 35.10 %
bases masked: 502,200,627 bp (51.69 %)

	number of elements*	length occupied	percentage of sequence
Retroelements	224657	90253146 bp	9.29 %
SINEs:	**58**	**6761 bp**	**0.00 %**
Penelope	3187	661022 bp	0.07 %
LINEs:	**196268**	**74550945 bp**	**7.67 %**
CRE/SLACS	0	0 bp	0.00 %
L2/CR1/Rex	43733	18550153 bp	1.91 %
R1/LOA/Jockey	15852	7502835 bp	0.77 %
R2/R4/NeSL	834	673762 bp	0.07 %
RTE/Bov-B	102395	29493616 bp	3.04 %

L1/CIN4	1415	94740 bp	0.01 %
LTR elements:	**28331**	**15695440 bp**	**1.62 %**
BEL/Pao	12565	7084057 bp	0.73 %
Ty1/Copia	1561	775084 bp	0.08 %
Gypsy/DIRS1	9951	7481940 bp	0.77 %
Retroviral	2481	143387 bp	0.01 %
DNA transposons	621139	227262774 bp	23.39 %
hobo-Activator	8749	1542960 bp	0.16 %
Tc1-IS630-Pogo	576750	212960682 bp	21.92 %
En-Spm	0	0 bp	0.00 %
MuDR-IS905	0	0 bp	0.00 %
PiggyBac	1280	395680 bp	0.04 %
Tourist/Harbinger	4060	1924044 bp	0.20 %
Other	275	11451 bp	0.00 %
Rolling-circles	19653	4597029 bp	0.47 %
Unclassified:	**828801**	**166508339 bp**	**17.14 %**
Total interspersed repeats:		484024259 bp	49.82 %
Small RNA:	7148	2907047 bp	0.30 %
Satellites:	796	124775 bp	0.01 %
Simple repeats:	213281	8909635 bp	0.92 %
Low complexity:	34278	1638267 bp	0.17 %

Supplementary Table 3: *Number of shared orthogroups among various dipteran species and their outgroup*

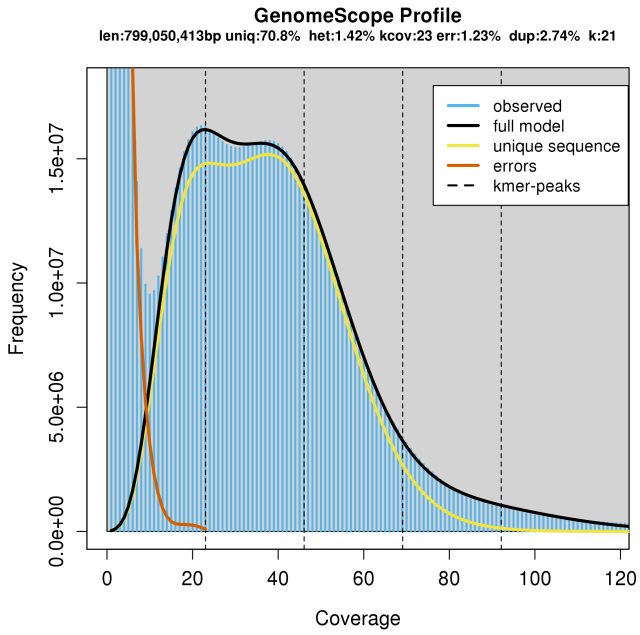
	H.melpomene	*A.aegypti*	*C.capitata*	*S.calcitrans*	*M.domestica*	*P.downsi*	*G.morsitans*	*D.melanogaster*
H.melpomene		6,978	6,874	6,883	6,910	6,800	6,337	6,859
A.aegypti	6,978		8,015	8,008	8,018	7,796	7,228	7,995
C.capitata	6,874	8,015		9,123	9,186	8,855	8,014	9,096
S.calcitrans	6,883	8,008	9,123		9,650	9,197	8,087	9,041
M.domestica	6,910	8,018	9,186	9,650		9,295	8,144	9,117
P.downsi	6,800	7,796	8,855	9,197	9,295		8,103	8,770
G.morsitans	6,337	7,228	8,014	8,087	8,144	8,103		7,940
D.melanogaster	6,859	7,995	9,096	9,041	9,117	8,770	7,940	

Supplementary Table 4: *Gene families unique only to P. downsi*

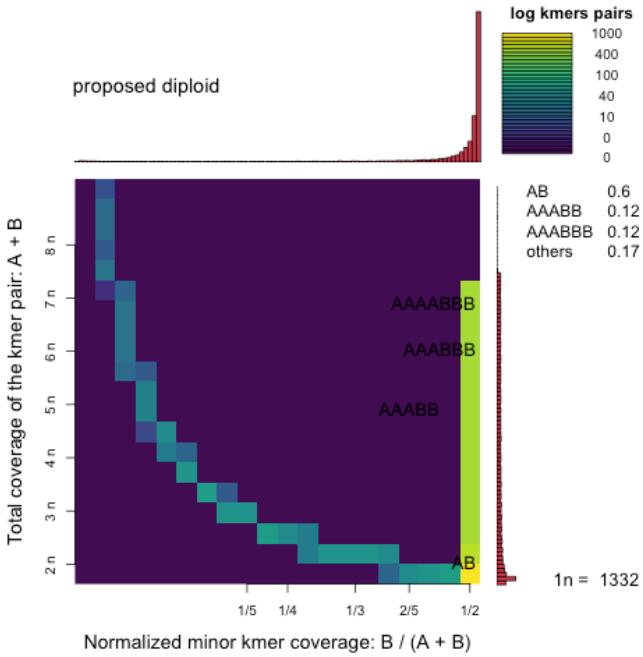
EGT47331	hypothetical protein CAEBREN_08836 [Caenorhabditis brenneri]
GBO13109	hypothetical protein AVEN_233885-1 [Araneus ventricosus]
GBP44057	PiggyBac transposable element-derived protein 4 [Eumeta japonica]
KMQ83311	transposable element tc3 transposase [Lasius niger]
KNC20639	hypothetical protein FF38_06891, partial [Lucilia cuprina]
KNC20966	hypothetical protein FF38_09018 [Lucilia cuprina]
KNC26906	hypothetical protein FF38_10930 [Lucilia cuprina]
KXJ68891	hypothetical protein RP20_CCG001210 [Aedes albopictus]
OAF68034	hypothetical protein A3Q56_04227 [Intoshia linei]
PCG68510	hypothetical protein B5V51_5157, partial [Heliothis virescens]
XP_011295145	PREDICTED: tigger transposable element-derived protein 6-like isoform X2 [Musca domestica]
XP_011295374	PREDICTED: tigger transposable element-derived protein 6 [Musca domestica]
XP_013109704	PREDICTED: uncharacterized protein LOC106088638 [Stomoxys calcitrans]
XP_017475229	PREDICTED: uncharacterized protein LOC108365650 [Rhagoletis zephyria]
XP_017478109	PREDICTED: uncharacterized protein LOC108367917 [Rhagoletis zephyria]
XP_017478991	PREDICTED: uncharacterized protein LOC108368617 [Rhagoletis zephyria]
XP_017479715	PREDICTED: uncharacterized protein LOC108369194 [Rhagoletis zephyria]
XP_017481195	PREDICTED: RNA-directed DNA polymerase from mobile element jockey-like [Rhagoletis zephyria]
XP_019891578	PREDICTED: ATP-binding cassette sub-family A member 3-like [Musca domestica]
XP_019894716	PREDICTED: uncharacterized protein LOC105262305 isoform X1 [Musca domestica]
XP_021704105	protein ALP1-like [Aedes aegypti]
XP_022823959	piggyBac transposable element-derived protein 4-like [Spodoptera litura]
XP_022834134	uncharacterized protein LOC111361914 [Spodoptera litura]
XP_033325321	uncharacterized protein LOC117219890 [Megalopta genalis]
XP_036214104	trypsin zeta-like [Bactrocera oleae]
XP_036337749	uncharacterized protein LOC118747737 isoform X3 [Rhagoletis pomonella]
XP_036342696	uncharacterized protein LOC118751975 [Rhagoletis pomonella]

Supplementary Table 5: Number of genes in *Fibrinogen C-Terminal Domain-Containing* gene family and *SCP domain-containing* gene family

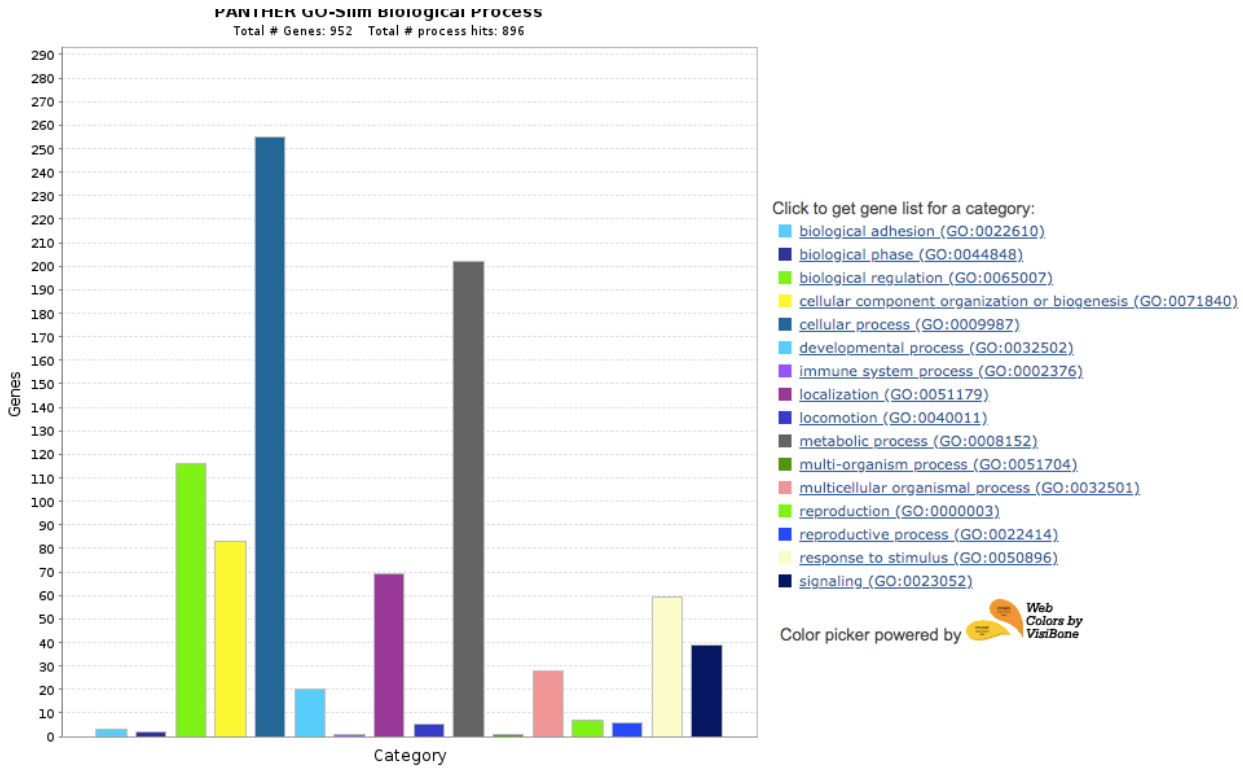
Species	Fibrinogen C-Terminal Domain-Containing gene family	SCP domain-containing gene family
H.melpomene	1	2
A.aegypti	33	20
C.capitata	10	6
S.calcitrans	45	17
M.domestica	32	25
P.downsi	5	7
G.morsitans	7	7
D.melanogaster	11	15



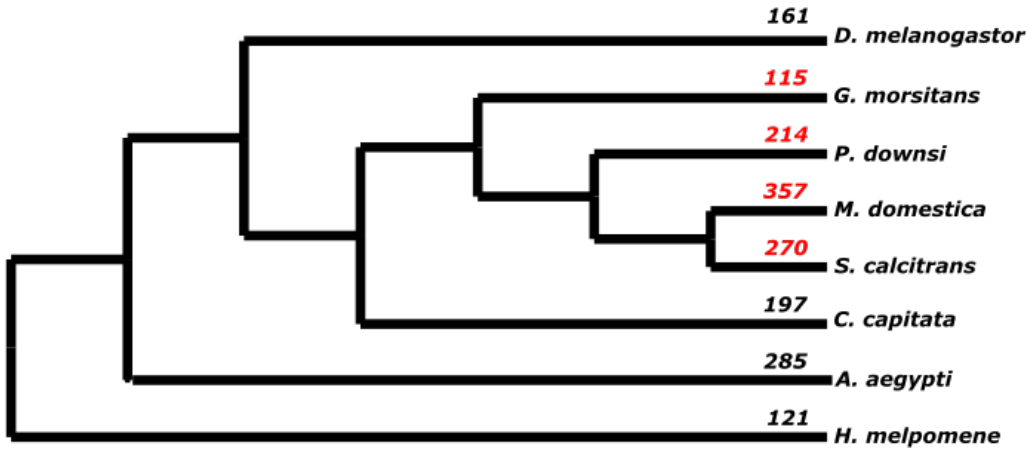
Supplementary Figure 1: *k-mer spectrum and fitted modelling used for estimating genome parameters of P. downsi from short sequencing reads*



Supplementary Figure 2: *Distribution of the total coverage of the k-mer pair (y-axis) against relative minor k-mer coverage (x-axis) providing evidence of diploidy in P. downsi.*



Supplementary Figure 3: *Gene ontology terms associated with 993 gene families unique to diptera*



Supplementary Figure 4: *Number of genes in Cuticular gene family across dipterans*