

1 **Multiple parasitoid species enhance top-down control, but**
2 **parasitoid performance is context-dependent**

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9 **Running Head:** Trophic interactions depend on context

10 **Data availability:** All raw data used for this study are available from the Zenodo database:

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12 **Abstract.** Ecological communities are composed of a multitude of interacting species, forming
13 complex networks of interactions. Current global changes are altering community composition and
14 we thus need to understand if the mechanisms structuring species interactions are consistent across
15 different species compositions. However, it is challenging to explore which aspects of species
16 interactions are primarily driven by community structure and which by species identity. Here we
17 compared the outcome of host-parasitoid interactions across four community modules that are
18 common in host-parasitoid communities with a laboratory experiment using a pool of three
19 *Drosophila* host and three larval parasitoid species, resulting in nine different species assemblages.
20 Our results show general patterns of community structure for host-parasitoid interactions. Multiple
21 parasitoid species enhanced host suppression without general antagonistic effects between
22 parasitoid species. Presence of an alternative host species had no general effects on host
23 suppression nor on parasitoid performance, therefore showing no evidence of indirect interactions
24 between host species nor any host switching behavior. However, effects of community structure
25 on parasitoid performance were species-specific and dependent on the identity of co-occurring
26 species. Consequently, our findings highlight the importance of both the structure of the
27 community and its species composition for the outcome of interactions.

28 **Key words:** community modules, multiple predator effects, community composition, interaction
29 modification, *Drosophila*

30

31 **Introduction**

32 In nature, species interact in a variety of ways, forming complex ecological networks (Fontaine et
33 al. 2011, Kéfi et al. 2012, 2015, Piloosof et al. 2017, García-Callejas et al. 2018, Miele et al. 2019).
34 How species interact depends on the structure of the community, but also on identity of species in
35 the assemblage (Bográn et al. 2002). With environmental changes such as climate warming,
36 species are shifting their ranges and phenology (Parmesan and Yohe 2003). But species show
37 differences in their sensitivity and responsiveness to these changes, disrupting historical patterns
38 of interactions and co-occurrences, with communities of new species composition (Alexander et
39 al. 2015). We thus need to determine which aspects of species interactions are primarily driven by
40 community structure, and which are driven by species identity to accurately forecast the ecological
41 consequences of changes in the biotic environment induced by global changes.

42 Together, trophic and non-trophic interactions, and their modifications by other co-occurring
43 species act in combinations to shape communities (Thierry et al. 2019), and their dynamics
44 (Kawatsu et al. 2021). A predator-prey interaction can be weakened by the presence of another
45 predator via exploitative competition, interference or intraguild predation, enhanced via
46 facilitation, or unchanged if predators have additive effects on prey suppression (Sih et al. 1998).
47 Top-down control might also be driven by a single influential predator species independently of
48 predator diversity (Letourneau et al. 2009), or be enhanced if predators show some degree of niche
49 differentiation (Bográn et al. 2002, Pedersen and Mills 2004, Snyder et al. 2006, 2008). A predator
50 might switch prey species with the presence of a competing predator, or with change in relative
51 prey abundances (Siddon and Witman 2004, Randa et al. 2009). In addition, two competing species
52 are able to coexist in nature, in part, because of indirect interactions through a shared natural
53 enemy, which reduces the frequency of the dominant competitor that would otherwise exclude less

54 competitive species (i.e., apparent competition) (Bonsall and Hassell 1999, Singh and Baruah
55 2020). Most studies looking at complex networks are observational (e.g., Tylianakis et al. 2007,
56 Jeffs et al. 2021), and typically unable to disentangle the potential mechanisms driving species
57 interactions described above. Experimental systems are thus needed to disentangle the mechanisms
58 structuring networks of interacting species. For this purpose, community modules (i.e., a small
59 number of species interacting in a specified pattern; Holt 1997, also referred to as “motifs” in the
60 literature; Milo et al. 2002) represent a powerful tool to isolate certain key interactions that
61 structure complex networks. They are the building blocks of natural communities (Gilman et al.
62 2010), and thus allow us to disentangle the mechanisms structuring them. Common community
63 modules in food webs are pair of predator-prey, two prey species sharing a common natural enemy
64 (i.e., apparent competition or mutualism; hereafter apparent competition module), or two predator
65 species attacking the same prey (i.e., exploitative competition, interference, or facilitation;
66 hereafter exploitative competition module). But experimental studies investigating the
67 mechanisms structuring interactions with community modules rarely consider potential variations
68 due to species-specific effects (but see Bográn et al. 2002, Snyder et al. 2006). Thus, it is unclear
69 whether the mechanisms structuring interactions are consistent when looking at community
70 modules of different species compositions (Cusumano et al. 2016).

71 Experiments manipulating interactions in different community contexts with different species
72 assemblages are still rare, and usually manipulate species assemblage of one trophic level at a time.
73 For instance, Bográn et al. (2002) revealed competitive interactions among predator species in
74 some, but not all the predator assemblages studied. However, the study used only one prey species.
75 Snyder et al. (2006) found varying strength in the effect of predator species diversity on aphid
76 suppression depending on the aphid species considered, but did not vary species composition in

77 multiple predator treatment. Understanding how the identity of co-occurring species at both trophic
78 levels affects the outcome of consumer-resource interactions is of particular importance in the
79 context of current global changes.

80 Here, we investigated the mechanisms structuring consumer-resource interactions using a host-
81 parasitoid system. Parasitoids are a diverse group of insects that use other arthropods as a nursery
82 for their offspring, killing their host to complete development (Godfray 2004). Parasitoids are
83 important for top-down control in agricultural and natural ecosystems, and widely used as
84 biological control agents. Interactions between hosts and parasitoids are easily observed, and host-
85 parasitoid communities thus represent a good model system to study how the structure and
86 composition of communities influence species interactions. We used a set of three *Drosophila*
87 species and three of their larval parasitoids from a natural tropical community in Australia (Jeffs
88 et al. 2021) in a laboratory experiment to isolate direct and indirect interactions within host-
89 parasitoid communities. We aimed to uncover general effects of community modules in our
90 *Drosophila*-parasitoid system, and detect any species-specific effects depending on the co-
91 occurring species identity (using 9 species assemblages for each of the four common community
92 modules in host-parasitoid networks: host-parasitoid pair, exploitative competition module,
93 apparent competition module, and both exploitative and apparent competition module).
94 Specifically, we tested the following hypotheses: (i) host suppression will be higher with the
95 presence of multiple parasitoid species (i.e., exploitative competition module) because of increased
96 chances to have an efficient parasitoid species (Pedersen and Mills 2004, Letourneau et al. 2009),
97 (ii) despite potential multiparasitism events and therefore a decrease in parasitoid performance
98 (Harvey et al. 2013). (iii) Pairwise interaction between a focal host-parasitoid pair will weaken
99 with the presence of an alternative host in the apparent competition module because of trait- and

100 density- mediated indirect effects (Werner and Peacor 2003). (iv) Combined effects of exploitative
101 competition among parasitoids and apparent competition among hosts in the four-species module
102 will differ from three-species modules depending on the identity of the co-occurring species
103 because of species-specific effects (Bográn et al. 2002, Sentis et al. 2017).

104 **Methods**

105 *Study system*

106 The experiment used cultures of *Drosophila* species and their associated parasitoids collected from
107 two tropical rainforest locations in North Queensland Australia: Paluma (S18° 59.031' E146°
108 14.096') and Kirrama Range (S18° 12.134' E145° 53.102') (< 100 m above sea level) (Jeffs et al.
109 2021). *Drosophila* and parasitoid cultures were established between 2017 and 2018, identified
110 using both morphology and DNA barcoding, and shipped to the Czech Republic under permit no.
111 PWS2016-AU-002018 from Australian Government, Department of the Environment. All cultures
112 were maintained at 23°C on a 12:12 hour light and dark cycle at Biology Centre, Czech Academy
113 of Sciences. Three host species (*Drosophila birchii*, *D. simulans* and *D. pallidifrons*), and three
114 larval parasitoid species *Asobara* sp. (Braconidae: Alysiinae; strain KHB, reference voucher no.
115 USNMENT01557097, reference sequence BOLD process ID:DROP043-21), *Leptopilina* sp.
116 (Figitidae: Eucolinae; strain 111F, reference voucher no. USNMENT01557117, reference
117 sequence BOLD process ID:DROP053-21), and *Ganaspis* sp. (Figitidae: Eucolinae; strain 84BC,
118 reference voucher no. USNMENT01557102 and USNMENT01557297, reference sequence
119 BOLD process ID:DROP164-21) were used (for more details on the parasitoid strains used see
120 Lue et al. 2021). *Drosophila* isofemale lines were kept on standard *Drosophila* medium (corn flour,
121 yeast, sugar, agar and methyl-4-hydroxybenzoate) for approximately 45 to 70 non-overlapping
122 generations. To revive genetic variation, four to seven lines from each host species were combined

123 to establish two population cages per species of mass-bred lines prior the start of the experiment.
124 Single parasitoid isofemale lines were used, and maintained for approximately 25 to 40 non-
125 overlapping generations prior to the start of the experiment by providing them every week with 2-
126 days-old larvae of *Drosophila melanogaster*. This host species was not used in the experiment,
127 thus avoiding potential bias due to maternal effects.

128 *Experimental design*

129 To investigate the effects of community structure and species composition on host-parasitoid
130 interactions, we used four community modules, and 9 different species assemblages each (6 host
131 and 6 parasitoid assemblages, from the pool of three host species and three parasitoid species)
132 replicated 6 times (Figure 1). Each replicate was represented by a set of two vials in one box, for
133 a total of 216 boxes. Either conspecific (Figure 1a and c) or heterospecific (Figure 1b and d)
134 parasitoids were used. The two vials contained *Drosophila* larvae from either the same host species
135 (Figure 1a and b) or different host species (Figure 1c and d). We also included control treatments
136 for each host species to acquire baseline levels of survival in the absence of parasitoids (replicated
137 8 times).

138 To initiate the experiment, twenty-five eggs of each host species were placed in a single glass
139 vial with 10mL of food media. To collect *Drosophila* eggs, an egg-wash protocol was developed
140 based on Nouhaud et al. (2018). The day before the egg-washed protocol was conducted, two egg-
141 laying mediums (petri dishes with agar gel topped with yeast paste) were introduced in each
142 population cage for flies to laying eggs overnight. We used a #3 round paintbrush and distilled
143 water to rinse out the yeast paste and collect the eggs on a fine black mesh that allowed only yeast
144 and water to filter through, leaving the eggs on the surface. Eggs were transferred into petri dishes
145 containing PBS (1 mM Calcium chloride dihydrate, 0.5 mM Magnesium Chloride Hexahydrate,

146 pH = 7.4), and collected under microscope using a pipette to transfer eggs from the petri dish with
147 PBS to the experimental vials.

148 After 48 hours, two vials with *Drosophila* second instar larvae (initially eggs) were placed in a
149 hermetically sealed plastic box (15x11x19 cm) with four 3-to-5-days-old parasitoids (1:1 sex
150 ratio). Twenty-four hours later, parasitoids were removed, and vials were removed from the boxes
151 and plugged for rearing (Figure S1). Every vial was checked daily for emergences until the last
152 individual emerged (up to 41 days for the species with the longest developmental time). We
153 stopped collecting host emergences after 5 consecutive days without emergences to avoid collecting the
154 second generation. All emergences were collected, identified, sexed, and stored in 95% ethanol. A
155 total of 11,400 host eggs were collected across 456 experimental vials, of which 7,494 (65.7%)
156 successfully emerged as adults (3,702 hosts and 3,792 parasitoids).

157 *Data analysis*

158 We characterized the outcome of host-parasitoid interactions by a combination of degree of
159 infestation (DI) for each host species (i.e., the probability of a larvae to be parasitized and die),
160 and successful parasitism rate (SP) for each host-parasitoid pair representing parasitoid
161 performance (i.e., the probability of a parasitized host to give rise to an adult parasitoid) (Carton
162 and Kitano 1981, Boulétreau and Wajnberg 1986). Degree of infestation (DI) was calculated as
163 the proportion of host attacked (the difference between adult hosts emerging from the controls
164 without parasitoids and from the experiment) among the total of hosts (set to 0 if the number of
165 hosts emerging from the experiment was greater than the controls). Successful parasitism rate (SP)
166 was calculated as the proportion of parasitoid adults emerging among the number of hosts attacked
167 (Carton and Kitano 1981, Boulétreau and Wajnberg 1986). If no parasitoid emerged or if the
168 number of hosts attacked was estimated to be zero, SP was set to 0. If the number of parasitoids

169 that emerged was greater than the estimated number of hosts attacked, SP was set to 1. For
170 treatments with one parasitoid species, we assumed that each of the two parasitoid individuals
171 were attacking hosts equally, therefore the number of parasitoid adults emerging was divided by
172 two to allow comparison of parasitism rates between single and multiple parasitoid species.

173 Data were analyzed using generalized linear mixed-effects models (GLMMs). Model
174 assumptions were verified with the *DHARMA* package (Hartig 2019). To correct for overdispersion
175 of the residuals and zero inflation, data were modeled using zero-inflation models with a beta
176 binomial error distribution and a logit link function using the *glmmTMB* function from the *TMB*
177 package (Lüdecke et al. 2019). Three model types were used to investigate general effects of
178 community modules, species-specific responses, and effects of community composition for each
179 focal species. (i) “Community module models” used two explanatory variables and their two-way
180 interaction to account for the fully-factorial design of the experiment that resulted in four
181 community modules (exploitative competition treatment with two levels: presence or not of a
182 parasitoid heterospecific, and apparent competition treatment with two levels: presence or not of
183 an alternative host species). Box ID (214 levels) was included as a random factor to remove the
184 variation between the different species assemblages and thus extract general effects of community
185 modules. Host species (three levels) for DI, and host-parasitoid pairs for SP were also included as
186 random factors to remove the variation between different species. (ii) “Species-specific
187 community module models” used the same explanatory variables than previously described, and
188 Box ID as a random factor, but host species and host-parasitoid pairs were included as fixed factors
189 to test if effects varied depending on the focal species. All three and two-way interactions between
190 treatments (exploitative and apparent competition), host species, and host-parasitoid pairs were
191 tested and kept in our models if found to be significant based on backward model selection using

192 Likelihood-ratio tests. Models for SP were also run for each host-parasitoid pair separately to
193 quantify differences in the sign and magnitude of the effects of community structure on pairwise
194 interaction depending on the focal species. (iii) “Community composition models” used species
195 assemblages rather than community modules as explanatory variables (host species assemblage: 6
196 levels, and parasitoid species assemblage: 6 levels). The two-way interaction between host and
197 parasitoid assemblages was always kept in the models to account for the fully-factorial design of
198 the experiment. Models for DI were run for each host species, and models for SP were run for each
199 host-parasitoid pair separately. Blocks (6 levels) were included in all models as a random effect.
200 Significance of the effects was tested using Wald type III analysis of deviance with Likelihood-
201 ratio tests. Factor levels of community modules and species assemblages were compared to the
202 reference module and species assemblages of the host-parasitoid pair in isolation by Tukey's HSD
203 *post hoc* comparisons of all means, using the *emmeans* package (Lenth 2018). All analyses were
204 performed using R 4.0.2 (Team 2017).

205 **Results**

206 *Effects of community structure on host suppression*

207 The presence of multiple parasitoid species in the module significantly increased the probability
208 of host being infested (DI) by 48% (CI 26-70%) (community module model: $\chi^2_{(1)} = 7.08$, $P =$
209 0.008 ; Post Hoc Odds Ratio (OR) exploitative competition module/pairwise interaction = 1.58, $P =$
210 0.076 ; OR exploitative and apparent competition module/pairwise interaction = 1.32, $P = 0.376$).
211 However, DI did not significantly change with the presence of an alternative host species ($\chi^2_{(1)} =$
212 0.56 , $P = 0.452$; OR apparent competition module/pairwise interaction = 0.96, $P = 0.984$), and the
213 two-way interaction between apparent and exploitative competition treatments had no significant
214 effect (community module model: $\chi^2_{(1)} = 0.22$, $P = 0.638$) (Figure 2a).

215 *Effects of host species and community composition on host suppression*

216 Host DI did not differ significantly across host species (species-specific community module
217 model: $\chi^2_{(2)} = 0.07$, $P = 0.965$). The directionality of the effect of parasitoid diversity did not vary
218 depending on species assemblages (community composition models; Figure S2 and Appendix S2).

219 *Effects of community structure on parasitoid performance*

220 Community modules had no general effects of successful parasitism rates (SP) (community
221 module model; Figure 2b), but the effects significantly varied across host-parasitoid pairs (species-
222 specific community module model; three-way interaction: $\chi^2(8) = 36.81$, $P < 0.0001$; Table 1).
223 The interaction between exploitative and apparent competition treatments had a significant effect
224 on SP for one out of the nine host-parasitoid pairs (*Ganaspis sp.* on *D. simulans*). SP of two other
225 host-parasitoid pairs significantly decreased with exploitative competition between parasitoid
226 species (*Ganaspis sp.* on *D. birchii* and on *D. pallidifrons*). SP of one host-parasitoid pair
227 significantly increased with apparent competition (*Asobara sp.* on *D. simulans*). SP for the rest of
228 the host-parasitoid pairs did not significantly changed between community modules when
229 compared to the host-parasitoid pair in isolation (Table 1).

230 *Effects of community composition on parasitoid performance*

231 Effects of an alternative host and a parasitoid competitor on parasitoid performance varied
232 depending on co-occurring species identity, both in term of magnitude and direction of their
233 response (community composition models). The interaction between host and parasitoid species
234 assemblages had a significant effect on SP for four out of the nine host-parasitoid pairs: *Asobara*
235 *sp.* on *D. simulans*, *Leptopilina sp.* on *D. birchii*, and *Ganaspis sp.* on *D. birchii* and on *D.*
236 *simulans*. Effects of species assemblages on SP for each host-parasitoid pair are summarized in
237 Table 2 and Figure 3, and presented in Appendix S3.

238 **Discussion**

239 Our results confirm some general effects of community structure on consumer-resource
240 interactions over a number of species combinations, but also reveal important species-specific
241 effects linked to the identity of species composing the community. Specifically: (i) the presence
242 of multiple parasitoid species consistently increased host suppression, showing the prevalence of
243 synergistic effects between consumer species in our system. On the contrary, (ii) the presence of
244 an alternative host had no general effect on host suppression, but increased or decreased successful
245 parasitism rate depending on host-parasitoid pairs and co-occurring species identity.

246 *Positive effects of consumer diversity on top-down control*

247 The presence of multiple parasitoid species generally increased host suppression. An increase in
248 top-down control compared to single-species treatments with one consumer species is predicted
249 when the different natural enemies present a certain degree of niche differentiation, therefore
250 complementing each other (Pedersen and Mills 2004), which have been reported in several
251 experimental studies (reviewed in Letourneau et al. 2009). Here, we observed a general positive
252 effect of consumer diversity on top-down control independently of the number of host species
253 present in the community. No general effects were detected on parasitoid performance, suggesting
254 no general difference between intra- and interspecific competition between consumers. Moreover,
255 the positive effects of parasitoid diversity on host suppression were mainly driven by the presence
256 of the most efficient parasitoid species for the focal host (e.g., presence of *Ganaspis sp.* for *D.*
257 *simulans*). These results match the sampling effect model suggesting that an increase in top-down
258 control with an increase in consumer diversity is explained because of an increasing probability
259 that a superior natural enemy species will be present in the community (Myers et al. 1989).

260 *No indirect interactions detected among prey species*

261 Contrary to the presence of an additional parasitoid species, we did not detect any effect of an
262 additional host species on host suppression, revealing no host switching behavior nor any indirect
263 interactions between host species over this one generation experiment. Another empirical study
264 failed to uncover any evidence of indirect interactions in natural host-parasitoid food webs
265 (Kaartinen and Roslin 2013). Indirect interactions between prey, mediated by a shared natural
266 enemy, is supposedly common in nature (e.g., Morris et al. 2001), and an important mechanism
267 for prey species coexistence via density- and trait-mediated effects (Holt 1977, Fleury et al. 2004,
268 van Veen et al. 2005, McPeck 2019). But direct exploitative competition between host species,
269 which was not allowed in our experiment, might have a stronger effect on host population and
270 community structure than apparent competition (Jones et al. 2009). Furthermore, frequency-
271 dependent attack rate, in which a predator switches between two prey species depending on their
272 relative densities, through aggregative behavior (Bonsall and Hassell 1999), and through learning
273 (Ishii and Shimada 2012), is another important mechanism determining the strength of predator-
274 prey interactions (van Veen et al. 2005), but was not tested in the present study. Varying host and
275 non-host densities could also change parasitoid foraging behavior (Kehoe et al. 2016). In our study,
276 potential effects of an alternative host (i.e., apparent competition module and both exploitative and
277 apparent competition module) on host-parasitoid interactions were tested over a single generation
278 and with constant host density. Therefore, experimental studies manipulating alternative prey
279 density over several generations might be needed to detect indirect interactions between host
280 species.

281 *Importance of community composition for consumer-resource interactions*

282 Our study was based on a particular set of interacting species, yet even the relatively small number
283 of species used in our experiment allowed us to uncover species-specific response within a given
284 community module. Community modules have been extensively used as a tool to study the
285 mechanisms structuring and stabilizing complex natural communities (Bascompte and Melián
286 2005, Rip et al. 2010), yet the effects of species identity in such studies is often ignored. Our results
287 highlight the variation in directionality and magnitude of the effects of a particular community
288 module on host-parasitoid interactions depending on the species assemblage considered.

289 Successful parasitism rate increased in modules with a parasitoid competitor compared to the
290 pairwise interaction in 6 species combinations (out of the 67 species combinations representing
291 host-parasitoid pairs in modules with either or both exploitative and apparent competition),
292 suggesting that some parasitoid species benefit from presence of an heterospecific. According to
293 a recent review on interspecific interactions among parasitoids (Cusumano et al. 2016), and to the
294 best of our knowledge, only one study showed facilitation between two parasitoid species on
295 cabbage white caterpillars (Poelman et al. 2014). Our case seems to be different because successful
296 parasitism rates did not increase for both parties. Here, modules with the pairwise interaction in
297 isolation had two parasitoid conspecifics, and our results therefore suggest that in these 6 cases,
298 interspecific competition between parasitoids was weaker than intraspecific competition.
299 Parasitoids can compete both as adults for oviposition and as larvae within an host (Harvey et al.
300 2013). Extrinsic competition would have negative effects on parasitoid attack rates, linked to
301 search efficiency and handling time, leading to a potential decrease in host mortality (Xu et al.
302 2016), which seems to not be the case in our study. Intrinsic competition is the result of a super-
303 or multiparasitism event when two parasitoids (conspecifics or heterospecific, respectively)

304 parasitize the same host individual. It is usually detrimental for the host survival, and therefore the
305 most likely interaction between parasitoids happening in our experiment that would explain an
306 increase in host suppression. Furthermore, parasitoids can inflict non-reproductive effects on their
307 hosts (i.e., ovipositor probing and egg laying without successful parasitism) that can lead to host
308 death (reviewed in Abram et al. 2019). This could also explain that host suppression increased
309 with multiple parasitoids while successful parasitism rate did not generally increase. Our
310 contrasting results on successful parasitism rate depending on the host-parasitoid pair and the other
311 parasitoid species present in the community are likely due to differences in traits (e.g., immune
312 response of the hosts, and oviposition behavior and virulence of the parasitoids; Carton et al. 2008).
313 The different trait combinations and trade-offs across host and parasitoid species are likely an
314 important mechanism driving species interactions and co-occurrences in natural communities
315 (Wong et al. 2019).

316 *Conclusion*

317 Our study is the first to our knowledge to investigate the effects of community module of different
318 species assemblages on consumer-resource interactions at both trophic levels. Studies looking at
319 interaction between predator species on single prey (e.g., Valente et al. 2019, Ortiz-Martínez et al.
320 2019), or studies looking at the effect of an alternative prey with single predator species (e.g., Ishii
321 and Shimada 2012, De Rijk et al. 2016), will overlook important mechanisms present in nature
322 where a number of species co-occur. With current global changes such as climate warming, the
323 structure and composition of communities is changing, either via direct effects on species
324 performance (Thierry et al. 2021), via effects on interactions (Hance et al. 2007, Thierry et al.
325 2019), or due to shift in ranges and phenology, and an increase in invasive species. It is therefore
326 imperative to take into account the context in which species interact, both abiotic and biotic, as all

327 those factors are likely to act together in influencing the outcome of the interaction between focal
328 species (Gilman et al. 2010).

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334 **Author contributions**

335 MT conceived the project; NP and JH contributed to the experimental design; MT, NP, MG, and
336 GP collected the data; MT analyzed the data. All authors contributed critically to the draft and
337 gave final approval for publication.

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488 **Table 1.** Odds ratios of having a successful parasitism event in each community module (exploitative competition, apparent competition,
489 and both exploitative and apparent competition) compared to the host-parasitoid pair in isolation for each pair (host abbreviations: b: *D.*
490 *birchii*, p: *D. pallidifrons*, s: *D. simulans*, and parasitoid abbreviations: A: *Asobara sp.*, L: *Leptopilina sp.*, G: *Ganaspis sp.*). Odds Ratios
491 superior or inferior to 1 translate an increased or a decreased probability of having successful parasitism, respectively. Results come
492 from the species-specific community module models run for each host-parasitoid pair separately. Significant Odds Ratios are highlighted
493 in bold.

494

Module	b-A		p-A		s-A		b-L		p-L		s-L		b-G		p-G		s-G	
Expl. comp.	1.49	(ns)	0.63	(ns)	1.56	(ns)	0.85	(ns)	1.36	(ns)	0.00	(ns)	0.30	*	0.19	*	0.67	(ns)
App. comp.	0.95	(ns)	1.03	(ns)	4.29	***	1.16	(ns)	1.65	(ns)	0.00	(ns)	0.94	(ns)	0.92	(ns)	0.83	(ns)
Expl. + app. comp.	0.65	(ns)	0.38	(ns)	0.76	(ns)	1.44	(ns)	1.82	(ns)	0.00	(ns)	0.73	(ns)	0.45	(ns)	4.12	***
Df residuals	65		65		65		65		65		65		64		64		65	

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497

498 **Table 2.** Effects of community composition on the probability to have a successful parasitism event for each host-parasitoid pair. Effects
 499 are shown by the summary of Likelihood-ratio chi-square tests on the community composition models with the effects of host and
 500 parasitoid species assemblages (3 levels each) (host abbreviations: b: *D. birchii*, p: *D. pallidifrons*, s: *D. simulans*, and parasitoid
 501 abbreviations: A: *Asobara sp.*, L: *Leptopilina sp.*, G: *Ganaspis sp.*). For p-A and s-L models contain only host species assemblage as a
 502 fixed effect due to convergence problem with the full model. Degrees of freedom (Df) are given for each factor and for the residuals. χ^2
 503 values are presented with the significance of the effect: (***) $P < 0.001$, (**) $P < 0.01$, (*) $P < 0.05$, (ns) $P > 0.05$.
 504

Effects	Df	b-A	p-A	s-A	b-L	p-L	s-L	b-G	p-G	s-G
Host sp.	2	11.14 **	1.12 (ns)	15.56 ***	4.80 (ns)	4.83 (ns)	10.08 **	34.14 ***	2.23 (ns)	19.71 ***
Parasitoid sp.	2	6.63 *	-	0.11 (ns)	38.32 ***	4.36 (ns)	-	2.57 (ns)	1.73 (ns)	13.81 **
Host x para	4	7.80 (ns)	-	24.19 ***	40.57 ***	7.81 (ns)	-	25.22 ***	9.01 (ns)	20.51 ***
Df residuals	60	66	60	60	60	66	59	59	60	

505

506 **Figure legends**

507

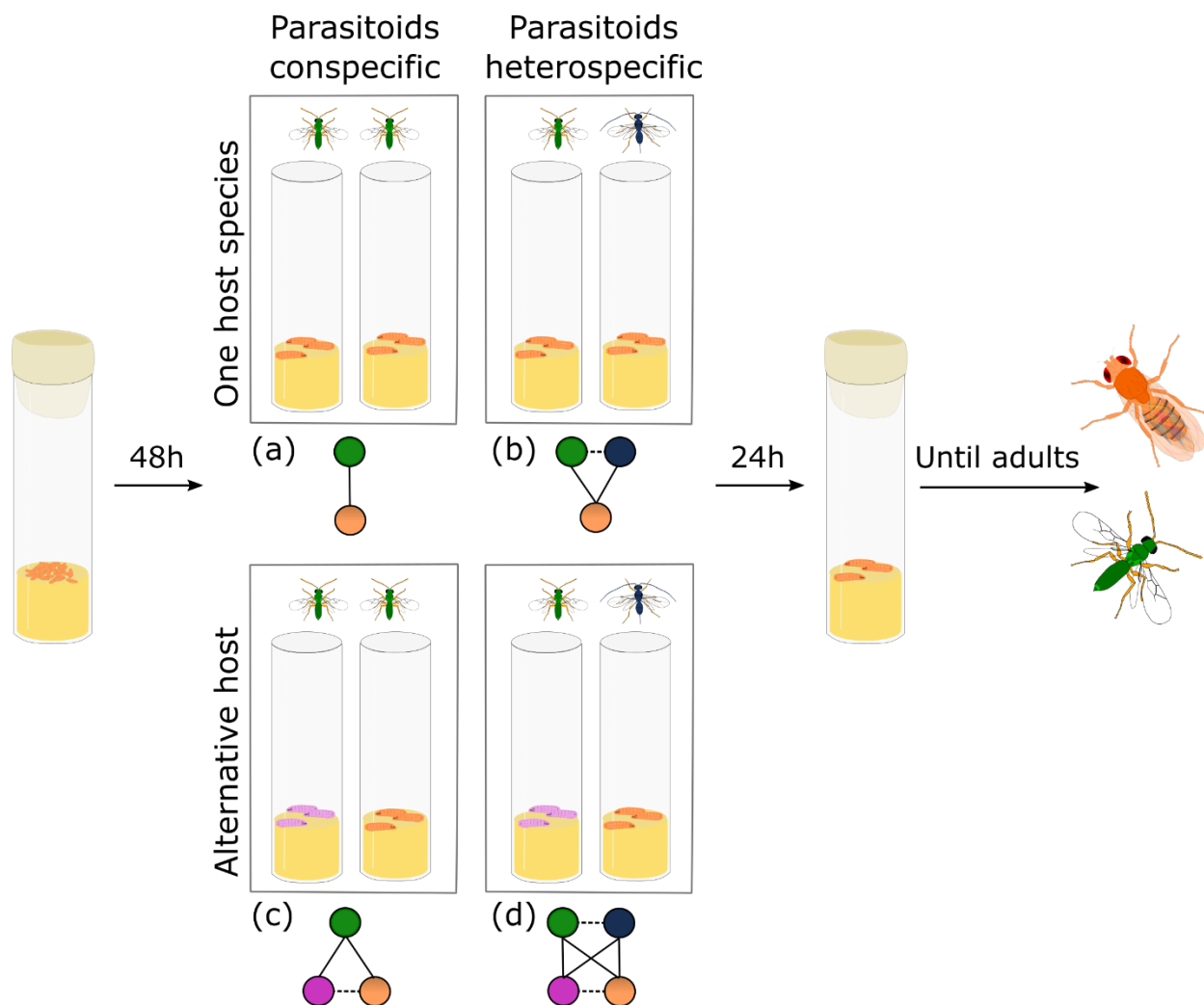
508 **Figure 1.** Schematic representation of the experimental treatments with the potential direct and
509 indirect interactions in each community module. Orange and pink nodes and larvae represent
510 different *Drosophila* host species, and green and blue nodes and wasps represent different
511 parasitoid species, assembled in a fully factorial design in four different community modules
512 represented schematically below their corresponding experimental box: a) host-parasitoid pair
513 (one host and one parasitoid species), b) exploitative competition module (one host and two
514 parasitoid species), c) apparent competition module (two host and one parasitoid species) and, d)
515 both exploitative and apparent competition module (two host and two parasitoid species). In the
516 community module schemas, solid lines represent trophic interactions, and dashed lines represent
517 non-trophic interactions (in b) either exploitative competition, interference, or facilitation between
518 parasitoids, c) either apparent competition or mutualism between hosts, and d) potential for all the
519 above). Direct interaction between host species were not allowed. See Thierry et al. (2019) for a
520 detailed description of each interaction type

521

522 **Figure 2.** Effects of community structure (represented by the community module bellow each
523 treatment) on (a) degree of infestation and on (b) successful parasitism rate. Different capital letters
524 denote significant differences between community structure from the community module models.
525 The small points represent the observed values, and the large points represent the predicted values
526 with their 95% confidence intervals

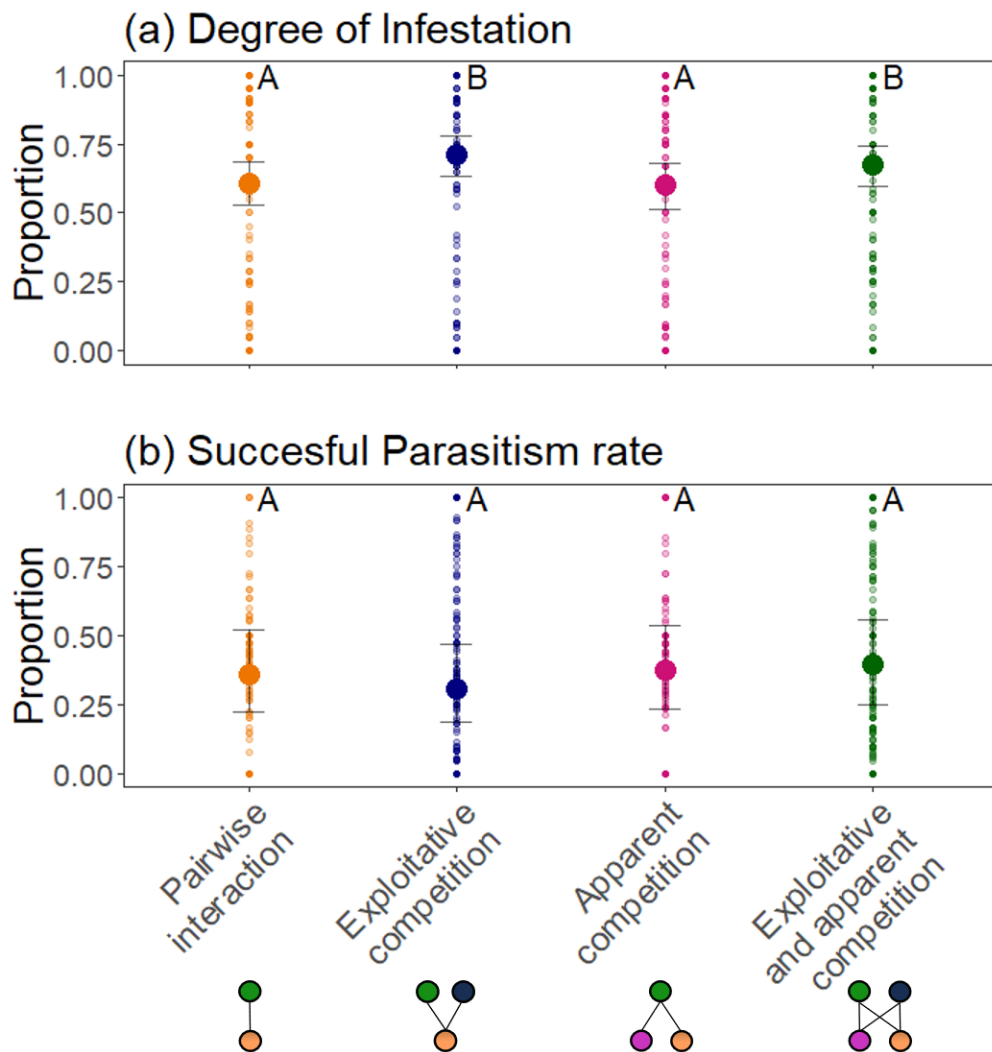
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528 **Figure 3.** Effects of community composition (identity of the alternative host and the parasitoid
529 heterospecific) on the successful parasitism rate of each parasitoid species on each host species
530 [rows are host species (host abbreviations: b: *D. birchii*, p: *D. pallidifrons*, s: *D. simulans*), and
531 columns are parasitoid species (parasitoid abbreviations: A: *Asobara sp.*, L: *Leptopilina sp.*, G:
532 *Ganaspis sp.*)]. Host assemblages are represented by the different colors, and parasitoid
533 assemblages are on the x axis. For SP p-A and SP s-L, only effect of host assemblages was analysed
534 due to convergence problem with the full model, and are represented for all parasitoid assemblages
535 combined. The small points represent the observed values, and the large points represent the
536 predicted values with their 95% confidence intervals from the community composition models



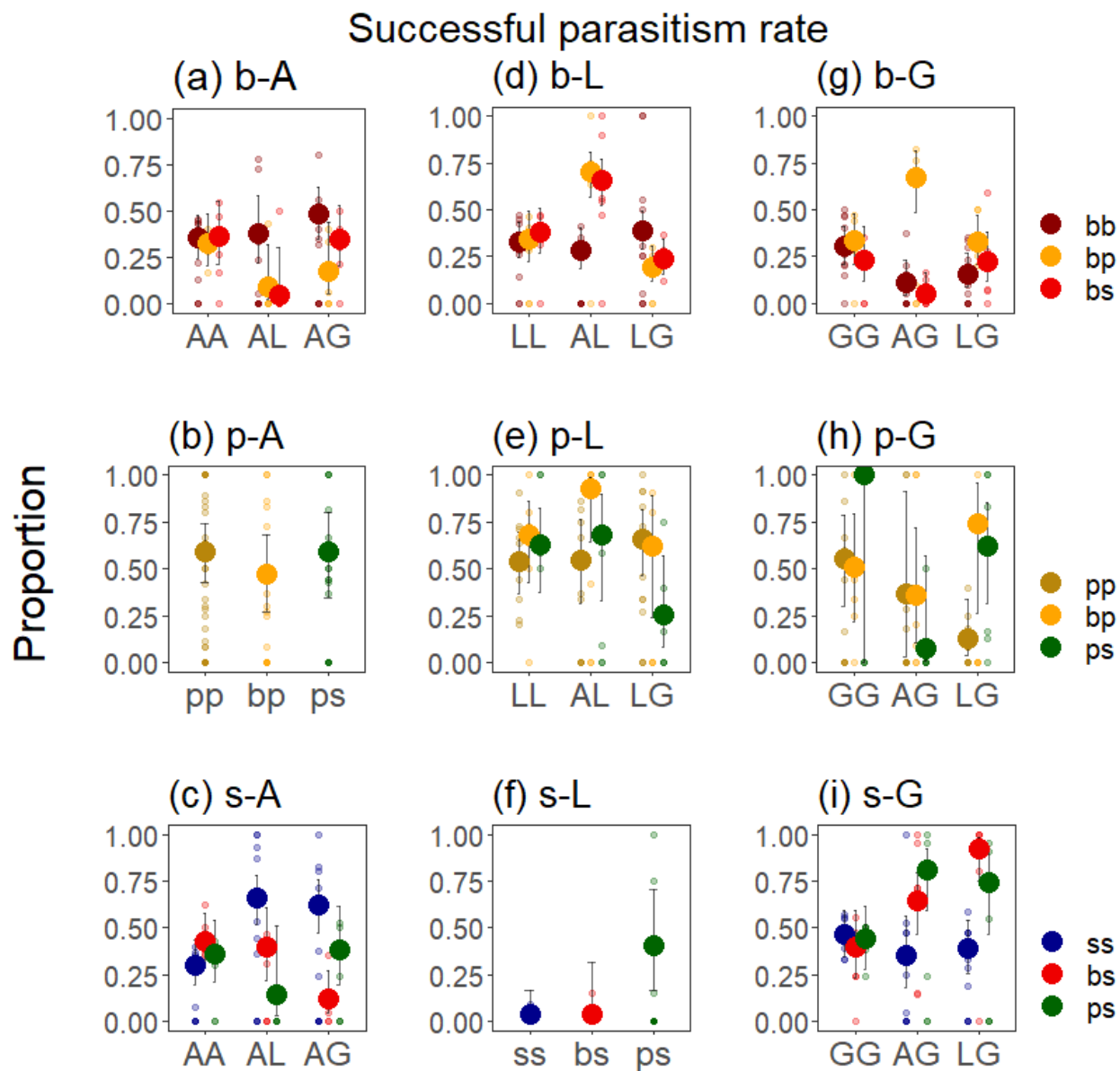
537

538 **Figure 1.**



539

540 **Figure 3.**



541

542 **Figure 3.**