

1 **Phylo-taxonomgenomics of 182 strains of genus *Leuconostoc* elucidates its robust taxonomy and**  
2 **biotechnological importance**

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18

19 **Abstract**

20 Genus *Leuconostoc* is a group of a diverse range of lactic acid bacteria (LAB) majorly found in dairy,  
21 food and environmental ecology. These microbes are commercially important for several industries  
22 due to their intrinsic genomic attributes such as bacteriocins, carbohydrate-active enzymes, plasmids  
23 etc. Even though the species of *Leuconostoc* are commercially significant, their taxonomy is largely  
24 based on old, low resolution traditional methods. There have been several taxonomic reclassifications  
25 in the past which are inadequate for microbiologist and food industry professionals to truly demarcate  
26 any new strain of genus *Leuconostoc*. The current taxonomy of the genus is largely based on classical  
27 approaches, which are in utmost need of reinvestigation by whole genome-based approaches. In the  
28 present study, the taxono-phylogenomic analysis clearly depicted sixteen species including three novel  
29 genomospecies in addition to several reshufflings across the species namely, *L. mesenteroides*, *L.*  
30 *pseudomesenteroides*, *L. gelidum* and *L. lactis*. The presence of a wide range of carbohydrate-active  
31 enzymes, type III polyketide synthase and vector plasmids suggested the biotechnological potential of

32 constituent strains of the genera. Further, the absence of antibiotic gene clusters reaffirms their utility  
33 in industries such as food and dairy. Such large-scale in-depth genome-based study can shed light on  
34 the nature of the genome dynamics of the species and help to obtain a more robust taxonomic  
35 classification.

36 **Keywords:** *Leuconostoc*, reclassification, taxonogenomic, phylogenomics, genomospecies,  
37 CAZymes, type III PKS.

### 38 **Abbreviations**

39 GH: Glycoside Hydrolase; CE: Carbohydrate Esterase; PL: Polysaccharide Lyase; GT:  
40 Glycosyltransferase; AA: Auxiliary Activities; CBM: Carbohydrate-Binding Module; ANI: Average  
41 Nucleotide Identity; dDDH: digital DNA-DNA hybridization; LAB: Lactic Acid Bacteria; Type III  
42 PKS: Type III polyketide synthase

### 43 **Introduction**

44 The genus *Leuconostoc* [1] belongs to the family *Leuconostocaceae* [2] which is one among the widely  
45 used group of lactic acid bacteria (LAB). *Leuconostoc* was first isolated by Cienkowski from a slime  
46 outbreak in a sugar factory in 1878 [3]. The isolate was named *Ascococcus mesenteroides* which  
47 produces dextran slime in sucrose solution [3, 4]. In 1911, an aroma bacteria "X" was isolated from a  
48 creamery starter which was later named *Leuconostoc* in 1930 [3, 5]. Historically, the taxonomy of  
49 *Leuconostoc* is a long tug of war across the several families of LAB. *Leuconostoc* sp. often occurs in  
50 similar habitats as *Lactobacillus* and *Lactococcus*, and was considered as an intermediate between  
51 *Streptococcus* and *Lactobacillus*. Genome based investigation of *Lactobacillaceae* and  
52 *Leuconostocaceae* suggests union of these two families [6]. In accordance with the latest standing in  
53 nomenclature of *Leuconostoc* (<https://lpsn.dsmz.de/genus/leuconostoc>), there are 16 valid species and  
54 along with 11 synonyms. Species of genus *Leuconostoc* are facultatively anaerobic, Gram-positive,  
55 nonmotile, catalase-negative, asporogenous, psychrotolerant or psychrotrophic bacteria with optimum  
56 growth temperature of 25-30 °C with an average GC content of 37.5% [2]. These are primarily  
57 associated with plant matter, fermenting vegetables, industries such as dairy, food and pharma etc. [4,  
58 7-10]. Few reports of their presence in chilled stored meats and human blood are a matter of concern  
59 [11, 12]. *Leuconostoc* have been known as a component of starter cultures in dairy since the 1920s,  
60 but its factual information about dairy is not very well documented [13]. These microbes are especially,  
61 known for their probiotic properties and ability to catalyse the production of several biotechnologically  
62 important products [14, 15]. *L. mesenteroides* is the type species of genus *Leuconostoc* [1] which is  
63 explored quite extensively in industries. *L. mesenteroides* species are used as a probiotic candidates  
64 which enables them to survive and grow under various stress conditions present in the gastrointestinal  
65 tract [16]. Species of *Leuconostoc* is an ideal fermenter of simple carbohydrates and have the capacity

66 to metabolize a wide range of carbohydrates, sugar alcohols and gluconate [17, 18]. Although the  
67 *Leuconostoc* sp. has been identified for safe use in the food industry and accorded "generally  
68 recognized as safe" (GRAS) status, there are several reports of its disease-causing nature [19, 20].  
69 Interestingly, the strains of *L. mesenteroides* follow dual lifestyles [16] causing disease in plants and  
70 humans [21-24].

71 Even though the species of the genus *Leuconostoc* are widely used for several purposes, the taxonomy  
72 is largely unexplored. All the earlier attempts to improve the taxonomy of genus *Leuconostoc* were  
73 based on limited phenotypic based methods or lower resolution phylogenetic approaches including  
74 housekeeping genes such as 16S rRNA, *rpoB*, *recA* gene etc. [25-28]. As these are based on very  
75 limited information, such methods do not provide a robust classification [29]. These studies largely  
76 lack a whole genome-based approach to obtain a robust taxonomic, identification of the key functional  
77 attributes across the several species of genus *Leuconostoc*. Although, there are some genome based  
78 study such as species of *Leuconostoc gasicomitatum* [30] was emended to *Leuconostoc gelidum* subsp.  
79 *gasicomitatum* [31]. Later *Leuconostoc mesenteroides* subsp. *suionicum* [32] was emended to  
80 *Leuconostoc suionicum* [33]. Most recently genome-based identification of *L. falkenbergense* sp. nov.  
81 [34] using whole genome-based sequence information from previously reported strains of *L.*  
82 *pseudomesenteroides*. Another recent study suggests the reclassification of *L. mesenteroides* MTCC  
83 10508 as a strain of *L. suionicum* [35]. All the studies Are focusing on limited strains from some  
84 species of *Leuconostoc* and not considering the whole genus *Leuconostoc*.

85 To obtain robust taxonomy of *Leuconostoc* sp., we reinvestigated and re-evaluated the phylogeny,  
86 genetic relatedness, and genomic determinants of the species within the present genus *Leuconostoc*  
87 (<https://lpsn.dsmz.de/genus/leuconostoc>). Implementation of these methods resulted in a robust  
88 taxonomic framework. We have identified three novel genomospecies namely, GS1 GS2 and GS3.  
89 We observed several large reclassifications across the strains of species *L. gelidum* subsp.  
90 *gasicomitatum*, *L. pseudomesenteroides*, *L. lactis* and *L. mesenteroides*. We have also identified  
91 species-specific unique genes in the pangenome of the genus *Leuconostoc*. We identified the presence  
92 of type III polyketide synthase system in all representative strains *Leuconostoc*, which signifies the  
93 antimicrobial ability against pathogenic bacteria. The abundance of the carbohydrate-active enzyme  
94 was also identified which includes majorly glycoside hydrolase and glycosyltransferase responsible  
95 for degradation, modification, and creation of glycosidic bonds. Furthermore, we believe such a large  
96 whole genome-based investigation can help in the estimation and evaluation of new potential strains  
97 or species belonging to the genus *Leuconostoc*. In recent days, the whole genome-based taxonomic  
98 evaluation became one of the most robust approaches [6]. Our investigation is an attempt to understand  
99 the genus *Leuconostoc* whose industrial importance is widely known but have underexplored  
100 taxonomic classification.

## 101 **Methods**

### 102 **Genome dataset procurement, initial QC and annotation**

103 All the genomes affiliated to genus *Leuconostoc* available in the public repository as of 31 March 2021  
104 were procured from NCBI (<https://www.ncbi.nlm.nih.gov/genome/?term=leuconostoc>) and  
105 EzBioCloud (<https://www.ezbiocloud.net/taxonomy>). Procured genomes were quality assessed using  
106 checkM v1.1.0 [36] with completeness and contamination bar of 95% and less than 3% respectively.  
107 There were altogether 183 including an outgroup genome resources were taken in the study (Table 1).  
108 Genome sequences for the type strains of the species *L. lactis* [25], *L. miyukkimchii* [37], *L. palmae* [38],  
109 were not included in the analyses due to the absence or poor-quality data in public databases. Also,  
110 synonym species/subspecies were strains of the *L. amelibiosum*, *L. argentinum*, *L. durionis*, *L.*  
111 *ficulneum*, *L. fructosum*, *L. oeni*, *L. paramesenteroides*, *P. pseudoficulneum* were not included due to  
112 their unavailability in the public repository. All the genomes were annotated using prokka v1.12 [39].

### 113 **Phylogenetic and Phylogenomic analysis**

114 16S rRNA gene predicted using barnap v0.8 (<https://github.com/tseemann/barnap>) was used to fetch  
115 16S rRNA gene sequences from all 183 genomes. Multiple sequence alignment of 16S rRNA was  
116 performed using clustalw [40]. A phylogenetic tree based on maximum likelihood (ML) was constructed  
117 using mega 7 [41] with a bootstrap replication of 1000. *Weissella viridescens* DSM 20410 was used as  
118 an outgroup. Whole genome-based phylogeny was implemented using PhyloPhlAn v3.0 [42] which  
119 used more than 400 most conserved genes across the strains under investigation. PhyloPhlAn uses  
120 usearch [43] for searching the gene cluster, muscle [44] for performing multiple sequence alignment  
121 and fasttree [45] for phylogenetic tree generation. A core genome-based tree was also generated using  
122 roary v3.1.2 [46] with 70% (inter-species similarity) cut off from the core genome alignment obtained  
123 with mafft [47]. All the phylogenetic tree generated was annotated using iTOL (<https://itol.embl.de/>)  
124 [48] for more lucid representation.

### 125 **Genome similarity assessment**

126 Genome similarity assessment was performed using several methods. Average nucleotide identity (ANI)  
127 was calculated using orthoANI v1.2 [49, 50] which uses usearch v8.1 [43] to find the orthologous cluster  
128 across the genomes for similarity comparison. A genome similarity assessment cut-off of 96% for  
129 orthoANI was used. Another tool fastANI v1.32 [51] (<https://github.com/ParBLiSS/FastANI>) was  
130 implemented to calculate nucleotide identity. fastANI uses a novel algorithm, which utilizes Mashmap  
131 with an intra and inter-species demarcation of >96% and <83% [52]. Digital DNA-DNA hybridization  
132 (dDDH) [53] was implemented using an online web portal of genome-to-genome distance calculator 2.1  
133 (GGDC) (<http://ggdc.dsmz.de/ggdc.php#>). Heatmap of the genome similarity assessment was generated  
134 using GENE-E (<https://software.broadinstitute.org/GENE-E/>).

## 135 **Pangenome and core genome assessment**

136 Pangenome analysis was implemented using roary v3.1.2 [46] with gff files obtained from prokka with  
137 blastp percentage identity of 85%. The flower plot indicates the core gene, unique gene and species  
138 specific to genes (Figure 3A). fasttree v2.1.10 [45] was implemented on the core gene alignment to  
139 obtain the core gene phylogeny. Core genome obtained was annotated with eggNOG-mapper [54]  
140 (<http://eggno-mapper.embl.de/>).

## 141 **Plasmid identification, antibiotic resistance and unique functional genomic attributes**

142 To obtain the presence of plasmid in the strains of genus *Leuconostoc*, abricate v1.0.1  
143 (<https://github.com/tseemann/abricate>) was implemented using plasmidFinder v2.1 database [55].  
144 Antibiotic resistance genes were also checked using the card module v3.08 [56], ResFinder v4.1 [57]  
145 and ARG-ANNOT [58] of abricate. Genomic attributes for the prediction of biosynthetic gene clusters  
146 were identified using antiSMASH v6 [59]. All the type strains and the novel genomospecies were  
147 checked for the presence of a biosynthetic gene cluster.

## 148 **Carbohydrate active enzyme analysis**

149 Carbohydrate-active enzymes (CAZy enzymes) (<http://www.cazy.org/>) among all the type strains and  
150 the novel genomospecies obtained from genome similarity analysis were identified using dbCAN2 [60].  
151 dbCAN2 scans the genome using Hidden Markov model (HMM) profile which uses HMMdb v7 [61]  
152 (e-value of  $< 1e-15$ , coverage  $> 0.35$ ), DIAMOND [62] (e-value  $< 1e-102$ ) and Hotpep [63] (frequency  
153  $> 2.6$ , hits  $> 6$ ) to improve the prediction. The genes were annotated by at least two of the methods were  
154 taken for further analysis. The detailed carbohydrate-active enzyme family information was obtained on  
155 the CAZyme (<http://www.cazy.org/>). Identified CAZymes were broadly classified as glycoside  
156 hydrolase (GHs), glycosyltransferases (GTs), carbohydrate esterases (CEs), carbohydrate-binding  
157 enzymes (CBM), auxiliary active enzymes (AAs), and polysaccharide lyases (PLs).

## 158 **Result and Discussion**

### 159 **Genomic features of the genus *Leuconostoc***

160 A total of 182 genomes of the genus *Leuconostoc* were used for the present study. Their average genome  
161 size ranged from 1 mb to 2.5 mb with an average GC content of 37%. The genomic attributes including  
162 #tRNA, #rRNA, #CDS etc. are summarized in table 1.

### 163 **Phylogenomics of the genus *Leuconostoc* reveals major phylogroups**

164 Due to high similarity 16S rRNA based phylogeny could not resolve the constituent species of the genus  
165 *Leuconostoc*. For instance, *L. inhae* and *L. gelidum* subsp. *gasicomitatum*; *L. rapi* and *L. kimchii*; *L.*  
166 *falkenbergense* and few strains of *L. pseudomesenteroides*; *L. mesenteroides*, *L. suionicum*, *L. litchi* and

167 *L. fallax* could not form distinct clade (Supplementary Figure 1). To get more accurate phylogeny, we  
168 obtained core genome-based phylogeny using roary and PhyloPhlAn (Figure 1, Supplementary Figure  
169 2). Both of these approaches resulted in major reshuffling across the *Leuconostoc* sp. Strains of major  
170 species of genus *Leuconostoc* namely, *L. mesenteroides*, *L. pseudomesenteroides*, *L. gelidum* and *L.*  
171 *lactis* were not forming species-specific clades. Rather, each of these four species were splitting into  
172 two distinct phylogroups.

173 Out of 76 strains of *L. mesenteroides* 74 formed a distinct phylogroup along with the *L. mesenteroides*  
174 subsp. *mesenteroides* ATCC8293<sup>T</sup> [1] confirming their species status. However, two of the remaining  
175 isolates namely, *L. mesenteroides* subsp. *mesenteroides* CBA3607 and *L. mesenteroides* CBA3628  
176 formed a distinct phylogroup. Similarly, out of 29 strains of *L. pseudomesenteroides* 4 strains including  
177 *L. pseudomesenteroides* LMG11482<sup>T</sup> [26] could be distinguished from the remaining 25 strains.  
178 Interestingly, these 25 strains of *L. pseudomesenteroides* formed a phylogroup with *L. falkenbergense*  
179 LMG10779<sup>T</sup> [34]. Further, out of 12 strains of *L. gelidum* only one strain *L. gelidum* subsp. *gelidum* JB7  
180 was forming phylogroup with *L. gelidum* subsp. *gelidum* KCTC3527<sup>T</sup> [64]. Whereas, remaining 10  
181 strains (*L. gelidum* subsp. *gasicomitatum*) were forming a phylogroup with the *L. inhae* KCTC3774<sup>T</sup>  
182 [65]. Similarly, 13 strains of *L. lactis* were also forming two distinct phylogroups comprising of 10 and  
183 3 strains respectively. However, due to the poor genome quality of the *L. lactis* type strain, we could not  
184 include in the present analysis. Overall, we could deduce 16 distinct phylogroups including 4 distinct  
185 unary strains which are representatives of previously described species namely, *L. fallax* ATCC700006<sup>T</sup>  
186 [66], *L. rapi* DSM27776<sup>T</sup> [27], *L. holzapfelii* CCUG54536<sup>T</sup> [67], *L. litchii* MB7<sup>T</sup> [67]

### 187 **Taxonogenomic assessment led to the identification of the novel genomospecies and reassignment** 188 **of strains**

189 Earlier taxonomic classification of species of *Leuconostoc* was mostly based on the classical taxonomy  
190 [1]. Description of the major species of genus *Leuconostoc* such as *L. mesenteroides* [1], *L. citreum* [26],  
191 *L. carnosum* [64], *L. inhae* [65] etc. was devoid of the whole genome-based approach such as ANI,  
192 dDDH and AAI etc., due to which, the current taxonomic classification of the *Leuconostoc* suggests  
193 them as a valid species (<https://lpsn.dsmz.de/genus/leuconostoc>). Robust phylogenomics of the genus  
194 *Leuconostoc* revealed several major reshufflings which need to be further confirmed by  
195 taxonogenomics.

196 Our whole genome-based taxonomic evidence using ANI (OrthoANI and fastANI) and dDDH  
197 correlated with the phylogenomics. For instance, three phylogroups consisting of earlier defined *L. lactis*  
198 and *L. mesenteroides* strains were found to be forming three novel genomospecies (GS1, GS2 and GS3)  
199 (Figure 2, Supplementary Figure 3). In addition to novel genomospecies, some of the previously defined  
200 strains of *L. pseudomesenteroides* and *L. gelidum* were identified as *L. falkenbergense* and *L. inhae*

201 respectively. The remaining species stands valid in accordance with the standing in nomenclature  
202 (<https://lpsn.dsmz.de/genus/leuconostoc>) (LPSN link, table 2).

### 203 **Pan genome-based identification of core genome and species-specific genes**

204 Pan-genome analysis using roary resulted in a comparatively small core genome (270 genes) and a large  
205 pan genome (22906 genes) which reveals a wide diversity of gene content within genus *Leuconostoc*.  
206 Here, we could get unique genes specific to the 16 species identified using taxonogenomics and  
207 phylogenomics (Figure 3A). *L. mesenteroides* (3822 genes) and *L. citreum* (2497 genes) largely  
208 contributed to the diversity of the pangenome. While, GS3 (134 genes), *L. suionicum* (148 genes) and  
209 *L. gelidum* (165 genes) had the least species-specific genes. Interestingly, amongst the unary species  
210 members, *L. fallax* showed the highest diversity with 1309 genes. The gene clusters obtained suggested  
211 a high number of accessory genes (shell and cloud) of 22,559 depicting a high degree of diversity across  
212 the species of genus *Leuconostoc*. Pan-genome matrix as well depicted the presence of species-specific  
213 genes (Figure 3C). Functional classification of the core genes suggested the majority of the information  
214 storage and processing (44%) and metabolism (32%) related genes (Figure 3B).

### 215 **Unique genomic attributes across spp. of *Leuconostoc*.**

216 High diversity in the genus *Leuconostoc* is evident from the gene content analysis. On the contrary, they  
217 were found to harbour pLCK4 (12183 bp) and LkipL4719 (21924 bp) plasmids in 97 and 86 genomes  
218 respectively. Plasmid pLCK4 was originally derived from *L. citreum* KM20 [68], which is one among  
219 the four high copy number plasmids of *L. citreum* KM 20 [68]. Plasmid pLCK4 which is considered to  
220 be a part of pMBLT00 [69], is considered to be a shuttle vector for inducing overproduction of D-lactate  
221 in *Leuconostoc* and *Lactococcus lactis* strains [69]. LkipL4719 is first reported in the *Leuconostoc*  
222 *kimchii* IMSNU 11154 [70] which harbours several metal transport gene families, suggesting its intrinsic  
223 nature across the species of *Leuconostoc* [70]. Additionally, 8 different plasmids namely pKLC4  
224 (36602) in 11 genomes, pIGMS32 (or ColRNAI\_1) (9294 bp) in 6 genome, pJB01 (2235 bp) in genome  
225 and pLFE1 (4031 bp) in 2 genomes. Whereas plasmid pYSI8 (4973 bp), pMG828-1 (Col(MG828)\_1)  
226 (1902 bp), pJE1 (5149 bp) and pBI143 (2747 bp) are present in one genome each (Figure 1). In addition  
227 to low diversity in the plasmid, antibiotic resistance genes were also present but only in 4 genomes out  
228 of 182 *Leuconostoc* strains analysed using CARD, resfinder and arg-annot database integrated in  
229 abricate (supplementary table 2). However, genomic analysis of the *Leuconostoc* human pathogens is  
230 limited due to a lack of genomic resources.

231 Whereas, we found Type III polyketide synthase (PKSs) in all the type strains of 16 species identified  
232 (Figure 4 A). Type III polyketide synthase (PKSs) are homodimers of ketosynthase which catalyze the  
233 condensation of one or several molecules of extender substrate onto a starter substrate through an  
234 iterative decarboxylative Claisen condensation reactions [71, 72]. Microbial type III PKSs seem to use  
235 an acyl–acyl carrier protein (ACP) as a starter substrate, [73], and in some cases, type III PKS genes

236 form a cluster with ACP or fatty acid biosynthetic genes [74, 75]. Future studies on type III PKSs will  
237 provide important insights into the properties of these enzymes and their role in the biosynthesis of  
238 natural products. Moreover, strains of genus *Leuconostoc* is reported to harbour several bacteriocins  
239 belonging to class IIa bacteriocins, leucocin H, mesentericin etc. [76-79] imparting antimicrobial  
240 activity against foodborne pathogens [80]. In-depth genomic investigation of bacteriocins and  
241 antimicrobial peptides are the need of the hour to explore the potential of the species of genus  
242 *Leuconostoc*.

### 243 **Carbohydrate-Active Enzyme (CAZymes)**

244 Among type strains of 16 phylogroups identified using genome similarity and phylogenomic assessment  
245 reveals the presence of CAZyme groups of GH, GT, CR, AA, CBM and PL using dbCAB2. These  
246 CAZymes groups are the enzymes involved in the degradation, modification, and creation of glycosidic  
247 bonds. Altogether 1344 protein-coding genes were detected by one of three methods namely hotpep,  
248 Diamond and HMMER (Figure 4 B). We found all the major CAZyme groups distributed across the  
249 strains represented with a heatmap (Figure 4 B) reveals the differential presence of all the classes of  
250 CAZyme in the strains of *Leuconostoc*. Most of the identified gene clusters were distributed across GH  
251 (52%), GT (40%), CE (4.75%), CMB (2.2%) and PL (1%) families. (Summarized in supplementary table  
252 1). We found diverse GHs namely GH13 (7.07%), GH70 (5.49%), GH73 (5.38%), GH1 (5.12%), GH  
253 32 (4.85%), GH25 (4.54%), GH65 (2.95%) across *Leuconostoc* sp. GHs catalyse the cleavage of O-  
254 glycosidic bonds linking the carbohydrate moieties or carbohydrate and non-carbohydrate moieties.  
255 GH13 family of CAZymes reveals the presence of amylases, beta-glucosidases, beta-xylosidases, beta-  
256 galactosidase gene cluster across the species of *Leuconostoc*. Similarly, the GH70 family includes  
257 transglucosylases which catalyse the intra- or intermolecular replacement of glycoside molecules at the  
258 anomeric position, giving rise to new glycoside molecules, such as oligosaccharides [81]. GH73 family  
259 includes N-acetylmuramidase which cleaves the  $\beta$ -1,4-glycosidic bond between the N-  
260 acetylglucosaminyl (GlcNAc) and N-acetylmuramyl (MurNAc) moieties in bacterial peptidoglycan  
261 [82]. Glycosyl transferases are primarily involved in the formation of glycosidic bonds by transfer of  
262 sugar moiety from the activated sugar donor to the acceptor molecule. We identified three major GTs  
263 namely GT4 (13.5%), GT2 (12.6%) and GT51 (4.3%). GT4 a glycosyltransferase family 4 protein is  
264 responsible for accessory Sec system glycosyltransferase GtfA. GT2 glycosyltransferase family 2  
265 protein is responsible for undecaprenyl-diphospho-muramoyl pentapeptide and beta-N-  
266 acetylglucosaminyltransferase and putative glycosyltransferase, exosortase G system-associated. GT51  
267 is responsible for PBP1A family penicillin-binding protein and murein polymerases. These are majorly  
268 involved in the synthesis of the peptidoglycan cell wall and play crucial roles in maintaining the integrity  
269 of the cell wall [83]. Likewise, CAZymes present under CE, CBM and PL categories are CE9 (2.11%),  
270 CE1 (2.1%), CBM50 (1.79%) and PL1 (1%).



## 271 Conclusion

272 Several taxonomic reclassifications were found by the present study inferring the true phylogenetic  
273 positioning of the strains of genus *Leuconostoc* (Table 2). True phylogeny will help future research to  
274 better identify and reclassify the strains of the genus belonging to biotechnologically important genera.  
275 In-depth genomic investigations of the strains suggest the presence of several determinants such as  
276 plasmid and type III PKS system etc. and the absence of antibiotic gene cluster also approves GRAS  
277 status. However, minor reports of *Leuconostoc* strains as human pathogens have raised concerns about  
278 their biotechnological implications. The present phylo-taxonomomic study provides a robust  
279 taxonomic framework of the genus which will be valuable in the identification of these clinically  
280 relevant strains. Such future genomic investigations of strains from diverse niches including nosocomial,  
281 environment, food and industry will shed light on their niche-specific attributes. This will be critical in  
282 surveillance to demarcate safe to use strains for food and industry from their pathogenic counterparts.  
283 Thus, our work is expected to promote research on the biotechnological important genus which is long  
284 overlooked and to better understand the intrinsic property of these important microbes.

## 285 Figure and table legends

286 **Figure 1: Whole genome-based core genome phylogenomic tree obtained using roary:** Phylogenetic  
287 tree was obtained using fasttree on the core gene alignment using mafft in roary run. For circular  
288 representation iTOL was used and labelled in accordance to the species group. Bootstrap values are  
289 represented with blue dots. *Weissella viridescens* DSM 20410 was used as an outgroup. Genomospecies  
290 1, genomospecies 2 and genomospecies 3 represent the novel group of bacteria within the genus  
291 *Leuconostoc*. The plasmid was labelled in the genome with coloured dots at the circumference.

292 **Figure 2: Whole genome-based similarity assessment using OrthoANI and dDDH.** A). All is to all  
293 ANI similarity heatmap showing separate groups obtained by using a cut-off of 96%. B). Heatmap of  
294 digital DNA-DNA hybridization all across the type strains with the cut-off of 70% genome similarity.

295 **Figure 3: Pan-genome analysis:** A). Pan-genome representation of all the gene clusters obtained from  
296 roary (22906). Core gene (270), and species-specific genes (unique gene) along with the number of  
297 strains in the group is marked in each group of genera *Leuconostoc*. B) Distribution of COG-based  
298 functional categories of core genes of the *Leuconostoc* genus. Here, the x-axis represents the number of  
299 genes and y-axis represents the functional categories. The count of gene cluster is indicated across  
300 classes in the box plot. C). Pan-genome plot representing the gene presence-absence across the strains.  
301 Core genome phylogroups are represented in the left isolates.

302 **Figure 4: Type III PKS and CAZymes:** A). The presence of type III PKS gene cluster was present in  
303 all the type strains in the study. The presence of the core biosynthetic gene in almost all the type strains  
304 was identified. Other genes such as additional biosynthetic genes, transporter genes etc. were also

305 identified. B). Heatmap showing all the different classes of the carbohydrate-active enzyme classified  
306 under GH: Glycoside Hydrolase; CE: Carbohydrate Esterase; PL: Polysaccharide Lyase; GT:  
307 Glycosyltransferase; AA: Auxiliary Activities; CBM: Carbohydrate-Binding Module. Venn diagram  
308 revealing carbohydrate-active enzyme identified by three different approaches of HMMER, diamond  
309 and hotpep. Heatmap was generated based on the presence of the carbohydrate-active enzyme by at least  
310 two approaches.

311 **Supplementary Figure 1: 16S rRNA tree:** A phylogenetic tree based on ML-based obtained by 16S  
312 rRNA gene sequences of all the strains under study. Bootstrap values are represented by blue dots. Strain  
313 name of some of the complete genome which resulted in multiple copies of 16S rRNA were appended  
314 with 1, 2, 3 etc.

315 **Supplementary Figure 2: Phylogenetic tree obtained using PhyloPhlAn:** A phylogenetic tree based  
316 on more than 400 conserved genes. Bootstrap values are marked with blue dots.

317 **Supplementary Figure 3: fastANI:** Heatmap obtained using genome similarity matrix obtained using  
318 fastANI.

319 **Table 1:** Metadata of the strains used in the study. A detailed description of all the strains used in the  
320 study along with the assembly statistics such as assembly size, %GC content, number of CDS, etc.  
321 general identification of the strains such as strain Id, NCBI accession number and source etc. are also  
322 summarized.

323 **Table:2:** A descriptive list of the earlier classification and robust reclassification of the strain based on  
324 this study.

325 **Supplementary Table 1:** A descriptive list of antibiotic resistance genes predicted using several  
326 databases such as using CARD, resfinder and arg-annot.

327 **Supplementary Table 2:** A descriptive list of CAZyme identified using dbCAN2.

## 328 References

- 329 1. Van Tiegham, P.J.A.d.s.n.B.B., *Sur la gomme de sucrerie*. Annales des sciences naturelles.  
330 Botanique. Botanique, 1878. **6**(7): p. 180.
- 331 2. Schleifer, K.H.J.B.s.M.o.S.o.A. and Bacteria, *Leuconostocaceae fam. nov.* 2015: p. 1-1.
- 332 3. Hucker, G. and C.S.J.B. Pederson, *Studies on the Coccaceae XVI. The genus Leuconostoc.*  
333 *Tech.* 1930. **167**.
- 334 4. Devoyod, J. and F.J.L.I. Poullain, *Les Leuconostocs. Propriétés: leur rôle en technologie*  
335 *laitière*. 1988. **68**(3): p. 249-279.
- 336 5. Lundstedt, E.J.C.D.P.J., *Some reflections on the development of starters for the cultured dairy*  
337 *products industry [in USA and Europe]*. 1983.
- 338 6. Zheng, J., et al., *A taxonomic note on the genus Lactobacillus: Description of 23 novel genera,*  
339 *emended description of the genus Lactobacillus Beijerinck 1901, and union of*  
340 *Lactobacillaceae and Leuconostocaceae*. International journal of systematic evolutionary  
341 microbiology, 2020. **70**(4): p. 2782-2858.
- 342 7. Cogan, T. and S.J.T.I.m.p. Gilliland, *In Bacterial Starter Cultures for Foods*. 1985, CRC Press  
343 Boca Raton, FL. p. 25-40.
- 344 8. Kekkonen, R.A., et al., *Probiotic Leuconostoc mesenteroides ssp. cremoris and Streptococcus*  
345 *thermophilus induce IL-12 and IFN- $\gamma$  production*. 2008. **14**(8): p. 1192.
- 346 9. Ogier, J.-C., et al., *Safety assessment of dairy microorganisms: the Leuconostoc genus.*  
347 International journal of food microbiology, 2008. **126**(3): p. 286-290.
- 348 10. Duan, P., et al., *Whole-cell biosynthesis of 2-O- $\alpha$ -D-glu-copyranosyl-sn-glycerol by*  
349 *recombinant Bacillus subtilis*. 2020. **36**(9): p. 1918-1928.
- 350 11. Borch, E., M.-L. Kant-Muermans, and Y.J.I.j.o.f.m. Blixt, *Bacterial spoilage of meat and cured*  
351 *meat products*. 1996. **33**(1): p. 103-120.
- 352 12. Handwerger, S., et al., *Infection due to Leuconostoc species: six cases and review*. 1990.  
353 **12**(4): p. 602-610.
- 354 13. Thunell, R.J.J.o.d.s., *Taxonomy of the Leuconostocs*. 1995. **78**(11): p. 2514-2522.
- 355 14. Saravanan, C., et al., *Production, partial characterization and antioxidant properties of*  
356 *exopolysaccharide  $\alpha$ -D-glucan produced by Leuconostoc lactis KC117496 isolated from an idli*  
357 *batter*. 2019. **56**(1): p. 159-166.
- 358 15. Abid, Y., et al., *Rheological and emulsifying properties of an exopolysaccharide produced by*  
359 *potential probiotic Leuconostoc citreum-BMS strain*. 2021. **256**: p. 117523.
- 360 16. de Paula, A.T., et al., *The two faces of Leuconostoc mesenteroides in food systems*. 2015.  
361 **31**(2): p. 147-171.
- 362 17. Lonvaud-Funel, A.J.L.a.b.G., *metabolism and applications, Lactic acid bacteria in the quality*  
363 *improvement and depreciation of wine*. 1999: p. 317-331.
- 364 18. Kandler, O.J.A.v.L., *Carbohydrate metabolism in lactic acid bacteria*. 1983. **49**(3): p. 209-224.
- 365 19. Cappelli, E.A., et al., *Leuconostoc pseudomesenteroides as a cause of nosocomial urinary*  
366 *tract infections*. 1999. **37**(12): p. 4124-4126.
- 367 20. Franco-Cendejas, R., et al., *Leuconostoc mesenteroides periprosthetic knee infection, an*  
368 *unusual fastidious Gram-positive bacteria: a case report*. BMC infectious diseases, 2017.  
369 **17**(1): p. 1-5.
- 370 21. Friedland, I., M. Snipelisky, and M.J.J.o.c.m. Khoosal, *Meningitis in a neonate caused by*  
371 *Leuconostoc sp.* 1990. **28**(9): p. 2125-2126.
- 372 22. Conn, K., et al., *Leuconostoc mesenteroides subsp. mesenteroides, the first report of a*  
373 *coccoid bacterium causing a plant disease*. 1995. **85**(5): p. 593-599.
- 374 23. Khudaier, B.Y.J.J.o.T.-Q.U.n.V., *Leuconostoc mesenteroides cause Nosocomial UTI At a*  
375 *tertiary care center in North India*. 2011.
- 376 24. Lee, M.-R., et al., *Healthcare-associated bacteraemia caused by Leuconostoc species at a*  
377 *university hospital in Taiwan between 1995 and 2008*. 2011. **78**(1): p. 45-49.
- 378 25. Garvie, E.I.J.J.o.D.R., *The genus Leuconostoc and its nomenclature*. 1960. **27**(2): p. 283-292.

- 379 26. FARROW, J.A., et al., *Nucleic acid homologies of some vancomycin-resistant leuconostocs*  
380 *and description of Leuconostoc citreum sp. nov. and Leuconostoc pseudomesenteroides sp.*  
381 *nov.* 1989. **39**(3): p. 279-283.
- 382 27. Lyhs, U., et al., *Leuconostoc rapi sp. nov., isolated from sous-vide-cooked rutabaga.* 2015.  
383 **65**(8): p. 2586-2590.
- 384 28. Ricciardi, A., et al., *Analysis of rpoB polymorphism and PCR-based approaches for the*  
385 *identification of Leuconostoc mesenteroides at the species and subspecies level.* 2020. **318**: p.  
386 108474.
- 387 29. Chun, J., et al., *Proposed minimal standards for the use of genome data for the taxonomy of*  
388 *prokaryotes.* International journal of systematic evolutionary microbiology, 2018. **68**(1): p.  
389 461-466.
- 390 30. Björkroth, K.J., et al., *Characterization of Leuconostoc gasicomitatum sp. nov., associated*  
391 *with spoiled raw tomato-marinated broiler meat strips packaged under modified-*  
392 *atmosphere conditions.* 2000. **66**(9): p. 3764-3772.
- 393 31. Rahkila, R., et al., *Reclassification of Leuconostoc gasicomitatum as Leuconostoc gelidum*  
394 *subsp. gasicomitatum comb. nov., description of Leuconostoc gelidum subsp. aenigmaticum*  
395 *subsp. nov., designation of Leuconostoc gelidum subsp. gelidum subsp. nov. and emended*  
396 *description of Leuconostoc gelidum.* 2014. **64**(4): p. 1290-1295.
- 397 32. Gu, C.T., et al., *Leuconostoc mesenteroides subsp. suionicum subsp. nov.* 2012. **62**(7): p.  
398 1548-1551.
- 399 33. Jeon, H.H., et al., *A proposal of Leuconostoc mesenteroides subsp. jongsajibkimchii subsp.*  
400 *nov. and reclassification of Leuconostoc mesenteroides subsp. suionicum (Gu et al., 2012) as*  
401 *Leuconostoc suionicum sp. nov. based on complete genome sequences.* 2017. **67**(7): p. 2225-  
402 2230.
- 403 34. Wu, Y. and C.T. Gu, *Leuconostoc falkenbergense sp. nov., isolated from a lactic culture,*  
404 *fermentating string beans and traditional yogurt.* International Journal of Systematic  
405 Evolutionary Microbiology, 2021. **71**(1): p. 004602.
- 406 35. Kaushal, G. and S.P.J.G. Singh, *Comparative genome analysis provides shreds of molecular*  
407 *evidence for reclassification of Leuconostoc mesenteroides MTCC 10508 as a strain of Leu.*  
408 *suionicum.* 2020. **112**(6): p. 4023-4031.
- 409 36. Parks, D.H., et al., *CheckM: assessing the quality of microbial genomes recovered from*  
410 *isolates, single cells, and metagenomes.* 2015. **25**(7): p. 1043-1055.
- 411 37. Lee, S.H., et al., *Leuconostoc miyukkimchii sp. nov., isolated from brown algae (Undaria*  
412 *pinnatifida) kimchi.* 2012. **62**(Pt\_5): p. 1098-1103.
- 413 38. Ehrmann, M.A., et al., *Leuconostoc palmae sp. nov., a novel lactic acid bacterium isolated*  
414 *from palm wine.* 2009. **59**(5): p. 943-947.
- 415 39. Seemann, T., *Prokka: rapid prokaryotic genome annotation.* Bioinformatics, 2014. **30**(14): p.  
416 2068-2069.
- 417 40. Thompson, J.D., D.G. Higgins, and T.J. Gibson, *CLUSTAL W: improving the sensitivity of*  
418 *progressive multiple sequence alignment through sequence weighting, position-specific gap*  
419 *penalties and weight matrix choice.* Nucleic acids research, 1994. **22**(22): p. 4673-4680.
- 420 41. Kumar, S., G. Stecher, and K. Tamura, *MEGA7: Molecular Evolutionary Genetics Analysis*  
421 *version 7.0 for bigger datasets.* Mol Biol Evol, 2016: p. msw054.
- 422 42. Asnicar, F., et al., *Precise phylogenetic analysis of microbial isolates and genomes from*  
423 *metagenomes using PhyloPhlAn 3.0.* 2020. **11**(1): p. 1-10.
- 424 43. Edgar, R.C., *Search and clustering orders of magnitude faster than BLAST.* Bioinformatics,  
425 2010. **26**(19): p. 2460-2461.
- 426 44. Edgar, R.C.J.N.a.r., *MUSCLE: multiple sequence alignment with high accuracy and high*  
427 *throughput.* 2004. **32**(5): p. 1792-1797.
- 428 45. Price, M.N., P.S. Dehal, and A.P.J.P.o. Arkin, *FastTree 2—approximately maximum-likelihood*  
429 *trees for large alignments.* 2010. **5**(3): p. e9490.

- 430 46. Page, A.J., et al., *Roary: rapid large-scale prokaryote pan genome analysis*. *Bioinformatics*,  
431 2015. **31**(22): p. 3691-3693.
- 432 47. Katoh, K., J. Rozewicki, and K.D.J.B.i.b. Yamada, *MAFFT online service: multiple sequence*  
433 *alignment, interactive sequence choice and visualization*. 2019. **20**(4): p. 1160-1166.
- 434 48. Letunic, I. and P.J.N.a.r. Bork, *Interactive Tree Of Life (iTOL) v4: recent updates and new*  
435 *developments*. 2019. **47**(W1): p. W256-W259.
- 436 49. Yoon, S.-H., et al., *A large-scale evaluation of algorithms to calculate average nucleotide*  
437 *identity*. 2017. **110**(10): p. 1281-1286.
- 438 50. Lee, I., et al., *OrthoANI: an improved algorithm and software for calculating average*  
439 *nucleotide identity*. *International journal of systematic and evolutionary microbiology*, 2016.  
440 **66**(2): p. 1100-1103.
- 441 51. Jain, C., et al., *High throughput ANI analysis of 90K prokaryotic genomes reveals clear species*  
442 *boundaries*. 2018. **9**(1): p. 1-8.
- 443 52. Jain, C., et al. *A fast approximate algorithm for mapping long reads to large reference*  
444 *databases*. in *International Conference on Research in Computational Molecular Biology*.  
445 2017. Springer.
- 446 53. Auch, A.F., et al., *Digital DNA-DNA hybridization for microbial species delineation by means*  
447 *of genome-to-genome sequence comparison*. *Standards in genomic sciences*, 2010. **2**(1): p.  
448 117-134.
- 449 54. Huerta-Cepas, J., et al., *Fast genome-wide functional annotation through orthology*  
450 *assignment by eggNOG-mapper*. *Molecular biology and evolution*, 2017. **34**(8): p. 2115-  
451 2122.
- 452 55. Carattoli, A., et al., *In silico detection and typing of plasmids using PlasmidFinder and plasmid*  
453 *multilocus sequence typing*. 2014. **58**(7): p. 3895-3903.
- 454 56. Jia, B., et al., *CARD 2017: expansion and model-centric curation of the comprehensive*  
455 *antibiotic resistance database*. 2016: p. gkw1004.
- 456 57. Zankari, E., et al., *Identification of acquired antimicrobial resistance genes*. 2012. **67**(11): p.  
457 2640-2644.
- 458 58. Gupta, S.K., et al., *ARG-ANNOT, a new bioinformatic tool to discover antibiotic resistance*  
459 *genes in bacterial genomes*. 2014. **58**(1): p. 212-220.
- 460 59. Blin, K., et al., *antiSMASH 5.0: updates to the secondary metabolite genome mining pipeline*.  
461 2019. **47**(W1): p. W81-W87.
- 462 60. Zhang, H., et al., *dbCAN2: a meta server for automated carbohydrate-active enzyme*  
463 *annotation*. 2018. **46**(W1): p. W95-W101.
- 464 61. Potter, S.C., et al., *HMMER web server: 2018 update*. 2018. **46**(W1): p. W200-W204.
- 465 62. Buchfink, B., C. Xie, and D.H.J.N.m. Huson, *Fast and sensitive protein alignment using*  
466 *DIAMOND*. 2015. **12**(1): p. 59-60.
- 467 63. Busk, P.K., et al., *Homology to peptide pattern for annotation of carbohydrate-active*  
468 *enzymes and prediction of function*. 2017. **18**(1): p. 1-9.
- 469 64. Shaw, B., C.J.I.J.o.S. Harding, and E. *Microbiology, Leuconostoc gelidum sp. nov. and*  
470 *Leuconostoc carnosum sp. nov. from chill-stored meats*. *International Journal of Systematic*  
471 *Evolutionary Microbiology*, 1989. **39**(3): p. 217-223.
- 472 65. Kim, B., et al., *Leuconostoc inhae sp. nov., a lactic acid bacterium isolated from kimchi*. 2003.  
473 **53**(4): p. 1123-1126.
- 474 66. Martinez-Murcia, A. and M.J.F.m.l. Collins, *A phylogenetic analysis of an atypical*  
475 *leuconostoc: description of Leuconostoc fallax sp. nov.* 1991. **82**(1): p. 55-59.
- 476 67. De Bruyne, K., et al., *Leuconostoc holzapfelii sp. nov., isolated from Ethiopian coffee*  
477 *fermentation and assessment of sequence analysis of housekeeping genes for delineation of*  
478 *Leuconostoc species*. 2007. **57**(12): p. 2952-2959.
- 479 68. Kim, J.F., et al., *Complete genome sequence of Leuconostoc citreum KM20*. 2008. **190**(8): p.  
480 3093-3094.

- 481 69. Chae, H.S., et al., *Use of a novel Escherichia coli-Leuconostoc shuttle vector for metabolic*  
482 *engineering of Leuconostoc citreum to overproduce D-lactate*. 2013. **79**(5): p. 1428-1435.
- 483 70. Oh, H.-M., et al., *Complete genome sequence analysis of Leuconostoc kimchii IMSNU 11154*.  
484 2010. **192**(14): p. 3844-3845.
- 485 71. Katsuyama, Y. and Y.J.M.i.e. Ohnishi, *Type III polyketide synthases in microorganisms*. 2012.  
486 **515**: p. 359-377.
- 487 72. Abe, I. and H.J.N.p.r. Morita, *Structure and function of the chalcone synthase superfamily of*  
488 *plant type III polyketide synthases*. 2010. **27**(6): p. 809-838.
- 489 73. Grünschow, S., et al., *Substrate profile analysis and ACP-mediated acyl transfer in*  
490 *Streptomyces coelicolor type III polyketide synthases*. 2007.
- 491 74. Hayashi, T., et al., *Fatty Acyl-AMP Ligase Involvement in the Production of Alkylresorcylic Acid*  
492 *by a Myxococcus xanthus Type III Polyketide Synthase*. 2011. **12**(14): p. 2166-2176.
- 493 75. Funai, N., et al., *A novel quinone-forming monooxygenase family involved in modification of*  
494 *aromatic polyketides*. 2005. **280**(15): p. 14514-14523.
- 495 76. Papathanasopoulos, M.A., et al., *Multiple bacteriocin production by Leuconostoc*  
496 *mesenteroides TA33a and other Leuconostoc/Weissella strains*. 1997. **35**(6): p. 331-335.
- 497 77. Revol-Junelles, A.M., et al., *Leuconostoc mesenteroides subsp. mesenteroides FR52*  
498 *synthesizes two distinct bacteriocins*. 1996. **23**(2): p. 120-124.
- 499 78. Blom, H., et al., *Characterization, production, and purification of leucocin H, a two-peptide*  
500 *bacteriocin from Leuconostoc MF215B*. 1999. **39**(1): p. 43-48.
- 501 79. Héchard, Y., et al., *Characterization and purification of mesentericin Y105, an anti-Listeria*  
502 *bacteriocin from Leuconostoc mesenteroides*. 1992. **138**(12): p. 2725-2731.
- 503 80. Woo, C., et al., *Bacteriocin production by Leuconostoc citreum ST110LD isolated from organic*  
504 *farm soil, a promising biopreservative*. Journal of Applied Microbiology, 2021.
- 505 81. Bissaro, B., et al., *Glycosynthesis in a waterworld: new insight into the molecular basis of*  
506 *transglycosylation in retaining glycoside hydrolases*. 2015. **467**(1): p. 17-35.
- 507 82. Lipski, A., et al., *Structural and biochemical characterization of the  $\beta$ -N-*  
508 *acetylglucosaminidase from Thermotoga maritima: toward rationalization of mechanistic*  
509 *knowledge in the GH73 family*. 2015. **25**(3): p. 319-330.
- 510 83. Sauvage, E., et al., *The penicillin-binding proteins: structure and role in peptidoglycan*  
511 *biosynthesis*. FEMS microbiology reviews, 2008. **32**(2): p. 234-258.

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519 **Table 1:** Metadata of the strains used in the study. A detailed description of all the strains used in the study along with the assembly statistics such as assembly  
520 size, %GC content, number of CDS, etc. general identification of the strains such as strain Id, NCBI accession number and source etc. are also summarized.

Strains ID	Accession	BioSample	BioProject	Assembly Level	Genome Size(Mb)	GC%	#CDS	#Contigs	#rRNA	#tRNA	#tmRNA	completeness/ contamination	Source
<b>L. carnosum CBA3620</b>	GCA_007954525.1	SAMN11843679	PRJNA544551	Complete	1.70133	37.3348	1618	3	12	67	1	99.48/1.05	kimchi
<b>L. carnosum DSM5576 T</b>	GCA_014207505.1	SAMN14908560	PRJNA632261	Scaffold	1.82066	37	1740	16	3	49	1	99.48/1.05	chill-stored meats
<b>L. carnosum JB16</b>	GCA_000300135.1	SAMN02603179	PRJNA174298	Complete	1.77347	37.091	1655	5	12	67	1	99.48/1.05	Kimchi, Korean traditional fermented food
<b>L. carnosum MFPA29A1405</b>	GCA_900322425.1	SAMEA104699785	PRJEB25279	Contig	1.66684	37.2698	1598	22	4	48	1	99.48/1.05	Beef carpaccio
<b>L. carnosum MFPC16A2803</b>	GCA_900322415.1	SAMEA104699786	PRJEB25280	Scaffold	1.81202	36.9587	1770	50	3	34	1	99.48/1.05	Beef carpaccio
<b>L. carnosum WC0318</b>	GCA_008690985.1	SAMN11618753	PRJNA542256	Scaffold	1.74508	37.2	1655	11	5	51	1	99.48/1.05	Cooked ham
<b>L. carnosum WC0319</b>	GCA_008690925.1	SAMN11618754	PRJNA542256	Scaffold	1.69968	37.1	1610	8	5	49	1	99.48/1.05	Cooked ham
<b>L. carnosum WC0320</b>	GCA_008690965.1	SAMN11618755	PRJNA542256	Contig	1.81154	37.1	1752	14	5	51	1	99.48/1.05	Cooked ham
<b>L. carnosum WC0321</b>	GCA_008690875.1	SAMN11618756	PRJNA542256	Scaffold	1.80189	37.1	1735	24	5	52	1	99.48/1.05	Cooked ham
<b>L. carnosum WC0322</b>	GCA_008690865.1	SAMN11618757	PRJNA542256	Scaffold	1.85285	37	1806	13	5	50	1	99.48/1.05	Cooked ham
<b>L. carnosum WC0323</b>	GCA_008690845.1	SAMN11618758	PRJNA542256	Scaffold	1.77288	37.2	1700	17	5	52	1	99.48/1.05	Cooked ham
<b>L. carnosum WC0324</b>	GCA_008690825.1	SAMN11618759	PRJNA542256	Contig	1.76563	37.1	1699	12	5	49	1	99.48/1.05	Cooked ham
<b>L. carnosum WC0325</b>	GCA_008690785.1	SAMN11618761	PRJNA542256	Scaffold	1.8298	37.2	1773	24	5	52	1	99.48/1.05	Cooked ham
<b>L. carnosum WC0326</b>	GCA_008689765.1	SAMN11618763	PRJNA542256	Contig	1.76965	37.2	1679	19	5	49	1	99.48/1.05	Sausage
<b>L. carnosum WC0327</b>	GCA_008689755.1	SAMN11618764	PRJNA542256	Contig	1.76938	37.2	1678	18	5	49	1	99.48/1.05	Sausage
<b>L. carnosum WC0328</b>	GCA_008689695.1	SAMN11618767	PRJNA542256	Scaffold	1.81458	37.1	1745	27	5	51	1	98.95/1.05	Sausage
<b>L. carnosum WC0329</b>	GCA_008689675.1	SAMN11618768	PRJNA542256	Scaffold	1.65083	37.2	1562	13	5	52	1	99.48/1.05	Cooked ham
<b>L. citreum 1301 LGAS</b>	GCA_001077275.1	SAMN03197274	PRJNA267549	Scaffold	1.82335	39	1715	37	9	60	1	99.89/0.19	Host: Homo sapiens
<b>L. citreum CBA3621</b>	GCA_007954785.1	SAMN11843662	PRJNA544551	Complete	1.90325	38.9138	1826	2	12	69	1	99.89/0.56	kimchi
<b>L. citreum CBA3623</b>	GCA_007954705.1	SAMN11843666	PRJNA544551	Complete	1.96941	38.9123	1848	6	12	69	1	99.33/0	kimchi
<b>L. citreum CBA3624</b>	GCA_007954565.1	SAMN11843677	PRJNA544551	Complete	1.89477	38.9513	1778	5	12	69	1	99.89/0.56	kimchi
<b>L. citreum CBA3627</b>	GCA_008033195.1	SAMN11843672	PRJNA544551	Complete	1.90325	38.9138	1824	2	12	69	1	99.89/0.56	kimchi
<b>L. citreum CW28</b>	GCA_002591805.1	SAMN06343424	PRJNA375706	Contig	1.98292	38.7	1873	9	12	69	1	99.33/0	corn fermented beverage
<b>L. citreum DmW 111</b>	GCA_002115685.1	SAMN06767174	PRJNA383560	Contig	1.82988	38.8	1768	19	7	53	1	99.33/0.75	from a built environment(Host: Drosophila)



<b>L. citreum EFEL 2700</b>	GCA_002804045.1	SAMN08014125	PRJNA418065	Chromosome	1.92383	38.964	1808	5	12	69	1	99.33/0	(host:Brassica rapa subsp. pekinensis)
<b>L. citreum F192-5</b>	GCA_008326505.1	SAMD00166981	PRJDB8200	Scaffold	2.06178	38.4	1989	42	3	47	1	99.89/0	
<b>L. citreum G4</b>	GCA_006381855.1	SAMN09099490	PRJNA470672	Contig	1.83552	38.8	1770	4	7	52	1	99.89/0	sourdough
<b>L. citreum KM20</b>	GCA_000026405.1	SAMN02603482	PRJNA16062	Complete	1.89661	38.8804	1777	5	12	69	1	99.89/0	
<b>L. citreum LBAE C10</b>	GCA_000239895.2	SAMEA2272656	PRJEA78993	Contig	1.93443	38.8	1850	76	3	49	1	99.89/0.08	
<b>L. citreum LBAE C11</b>	GCA_000239915.2	SAMEA2272136	PRJEA78995	Contig	1.96996	38.7	1848	83	3	49	1	99.89/0	
<b>L. citreum LBAE E16</b>	GCA_000239935.2	SAMEA2272523	PRJEA78997	Contig	1.8016	38.9	1737	45	3	49	1	99.89/0	
<b>L. citreum NBRC102476</b>	GCA_007989545.1	SAMD00166019	PRJDB6146	Contig	1.78757	38.9	1703	25	3	29	1	99.89/0	
<b>L. citreum NRIC1776 T</b>	GCA_008326545.1	SAMD00166982	PRJDB8200	Scaffold	1.80678	38.9	1737	8	3	47	1	99.89/0	Honey dew of rye ear
<b>L. citreum NRRL B-1299</b>	GCA_000820985.2	SAMEA2668124	PRJEB5537	Scaffold	1.75381	39	1769	1	9	53	1	99.33/0	
<b>L. citreum NRRL B-742</b>	GCA_000820965.2	SAMEA2668125	PRJEB5535	Scaffold	1.7181	39.2	1720	1	9	67	1	99.25/0	
<b>L. citreum TMW21194</b>	GCA_009792915.1	SAMN07739528	PRJNA401313	Contig	1.73532	38.9	1639	8	5	51	1	99.89/0	food
<b>L. citreum TR116</b>	GCA_004359915.1	SAMN07629511	PRJNA404070	Contig	1.83094	38.8	1764	31	5	51	1	99.89/0	yellow pea flour sourdough
<b>L. citreum TR153</b>	GCA_006406175.1	SAMN09099486	PRJNA470672	Contig	1.89839	38.7	1844	39	5	51	1	99.89/0	sourdough
<b>L. citreum WiKim0101</b>	GCA_009707745.1	SAMN13267039	PRJNA588925	Chromosome	2.01927	38.9102	1861	6	12	69	1	99.16/0.19	kimchi
<b>L. falkenbergense LMG10779 T</b>	GCA_014634805.1	SAMD00244137	PRJDB7793	Contig	1.96383	39.1	1876	94	3	50	1	100/0.18	fermentating string beans and traditional yogurt
<b>L. fallax ATCC700006 T</b>	GCA_004354645.1	SAMN08555278	PRJNA434383	Scaffold	1.66052	37.7	1589	16	15	86	1	99.3/0.52	sauerkraut
<b>L. gelidum subsp gasicomitatum C120c</b>	GCA_900009505.1	SAMEA3718674	PRJEB12226	Scaffold	1.99276	36.5	1889	31	3	42	1	99.44/0	vegetable salad
<b>L. gelidum subsp gasicomitatum C122c</b>	GCA_900016145.1	SAMEA3718675	PRJEB12227	Scaffold	2.00836	36.6	1914	22	3	43	1	98.88/0.19	vegetable salad
<b>L. gelidum subsp gasicomitatum KG16-1</b>	GCA_001536305.1	SAMEA3608619	PRJEB11303	Complete	2.05073	36.8506	1963	4	12	67	1	100/0.11	
<b>L. gelidum subsp gasicomitatum KSL4-2</b>	GCA_900016165.1	SAMEA3718676	PRJEB12228	Scaffold	2.08949	36.6	1993	23	3	44	1	99.33/0.11	Acetic acid preserved herring with carrots
<b>L. gelidum subsp gasicomitatum LMG18811 T</b>	GCA_000196855.1	SAMEA2272393	PRJEA48123	Complete	1.95408	36.7	1862	1	12	68	1	99.44/0	modified-atmosphere packaged, tomato-marinated broiler meat strips
<b>L. gelidum subsp gasicomitatum MFPA44A1401</b>	GCA_900218135.1	SAMEA104233143	PRJEB21908	Contig	1.89091	36.8	1798	50	2	48	1	100/0	beef carpaccio
<b>L. gelidum subsp gasicomitatum PB1a</b>	GCA_900016185.1	SAMEA3718677	PRJEB12229	Scaffold	2.08363	36.6	1987	29	3	48	1	99.44/0.56	carrot

<b>L. gelidum subsp gasicomitatum PB1e</b>	GCA_900016175.1	SAMEA3718678	PRJEB12230	Scaffold	2.08518	36.5	2014	22	3	43	1	99.44/0	carrot
<b>L. gelidum subsp gasicomitatum PL111</b>	GCA_900016205.1	SAMEA3718679	PRJEB12231	Scaffold	2.0581	36.5	1960	29	3	50	1	99.44/0	carrot
<b>L. gelidum subsp gasicomitatum TMW 2.1619</b>	GCA_009296125.1	SAMN05756392	PRJNA342276	Complete	1.92438	36.8534	1814	4	12	58	1	99.64/0.56	meat
<b>L. gelidum subsp gelidum JB7</b>	GCA_000298875.1	SAMN02603178	PRJNA174297	Complete	1.8935	36.7	1768	1	12	67	1	99.44/0.19	kimchi, Korean traditional fermented food
<b>L. gelidum subsp gelidum KCTC3527 T</b>	GCA_000166715.2	SAMN02470198	PRJNA60229	Contig	1.95728	36.6	1845	43	3	47	1	99.44/0.19	vacuum-packed meats
<b>L. holzapfelii CCUG54536 T</b>	GCA_012396485.1	SAMN14517846	PRJNA622446	Contig	1.75843	44	1668	18	5	50	1	99.37/0.28	coffee fermentation
<b>L. inhae KCTC3774 T</b>	GCA_000166735.2	SAMN02472184	PRJNA60231	Contig	2.29809	36.4	1622	893	3	48	1	95.81/6.07	kimchi
<b>L. kimchii C2</b>	GCA_000219785.1	SAMN02603135	PRJNA67911	Complete	1.87727	37.9	1792	1	12	68	1	100/0	kimchi
<b>L. kimchii IMSNU11154 T</b>	GCA_000092505.1	SAMN02603394	PRJNA40837	Complete	2.10179	37.9101	2025	6	12	68	1	100/0	kimchi, a traditional Korean food
<b>L. kimchii NKJ218</b>	GCA_004551615.1	SAMN10887832	PRJNA521423	Complete	1.99344	37.7797	1892	4	12	68	1	100/0	kimchi
<b>L. lactis 1300 LCIT</b>	GCA_001062635.1	SAMN03197273	PRJNA267549	Scaffold	1.80711	43.1	1687	58	9	63	1	99.44/0.26	Host: Homo sapiens
<b>L. lactis aa 0143</b>	GCA_004167235.1	SAMN10239554	PRJNA496358	Contig	1.73814	43.2	1637	38	3	46	1	99.44/0	stool(host:Homo sapiens)
<b>L. lactis BIOML-A1</b>	GCA_009678855.1	SAMN11946164	PRJNA544527	Contig	1.748	43.1	1632	30	4	46	1	99.44/0	(host:Homo sapiens)
<b>L. lactis CBA3625</b>	GCA_007954605.1	SAMN11843673	PRJNA544551	Complete	1.79161	43.3467	1714	2	12	68	1	99.44/0.56	kimchi
<b>L. lactis CBA3626</b>	GCA_007954665.1	SAMN11843668	PRJNA544551	Complete	1.83981	43.1491	1742	2	12	70	1	99.44/1.12	kimchi
<b>L. lactis CCK940</b>	GCA_002287365.1	SAMN07482638	PRJNA397634	Contig	1.74151	43.3	1657	2	12	69	1	100/0	kimchi
<b>L. lactis KACC91922</b>	GCA_000709265.1	SAMN02730106	PRJNA244961	Contig	1.68817	43.4	1619	35	3	50	1	99.44/0.67	Kimchi
<b>L. lactis KCTC3773</b>	GCA_000179875.1	SAMN02470199	PRJNA52539	Contig	1.72068	42.9	1663	98	3	48	1	99.44/0	
<b>L. lactis KFRI01</b>	GCA_001998805.1	SAMN05219201	PRJNA325075	Complete	1.71519	43.3281	1613	3	12	69	1	99.44/0	kimchi
<b>L. lactis LN19</b>	GCA_002092595.1	SAMN05981500	PRJNA352459	Contig	1.72439	42.9	1633	47	6	51	1	99.44/0.14	dairy
<b>L. lactis LN24</b>	GCA_002092695.1	SAMN05981502	PRJNA352459	Contig	1.72466	42.9	1641	48	6	52	1	99.44/0.14	dairy
<b>L. lactis NBRC 12455</b>	GCA_006539105.1	SAMD00097163	PRJDB6024	Contig	1.64349	43.5	1531	49	3	39		99.44/0	
<b>L. lactis WiKim40</b>	GCA_001698145.1	SAMN05300424	PRJNA327015	Complete	1.78807	43.1131	1654	4	12	68	1	100/0.56	Kimchi
<b>L. litichii MB7 T</b>	GCA_008107645.1	SAMN10786216	PRJNA516297	Scaffold	1.86586	35.8	1789	13	3	48	1	100/0	Litchi chinensis
<b>L. mesenteroides CBA3628</b>	GCA_007954745.1	SAMN11843664	PRJNA544551	Complete	1.86456	36.9975	1757	4	12	71	1	100/0	kimchi
<b>L. mesenteroides subsp cremoris ATCC19254 T</b>	GCA_000160595.1	SAMN00139187	PRJNA34629	Scaffold	1.73507	38.5	1511	29	3	49	1	100/0	Hansen's dried starter powder

<b>L. mesenteroides subsp cremoris LMGCF01</b>	GCA_002092045.1	SAMN05981463	PRJNA352459	Contig	1.64067	38	1449	115	5	51	1	99.82/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF02</b>	GCA_002092135.1	SAMN05981464	PRJNA352459	Contig	1.72228	37.9	1570	112	5	53	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF03</b>	GCA_002092145.1	SAMN05981465	PRJNA352459	Contig	1.66544	38	1494	116	5	53	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF04</b>	GCA_002092155.1	SAMN05981466	PRJNA352459	Contig	1.64019	38	1446	124	5	52	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF05</b>	GCA_002092085.1	SAMN05981467	PRJNA352459	Contig	1.63731	38	1452	118	5	53	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF07</b>	GCA_002092215.1	SAMN05981469	PRJNA352459	Scaffold	1.63929	38.1	1431	109	5	53	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF09</b>	GCA_002092105.1	SAMN05981471	PRJNA352459	Contig	1.62909	38.1	1468	105	5	53	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF10</b>	GCA_002092245.1	SAMN05981472	PRJNA352459	Contig	1.61642	38	1452	109	5	51	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF11</b>	GCA_002092315.1	SAMN05981473	PRJNA352459	Contig	1.67437	38.2	1488	86	5	53	1	99.87/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF12</b>	GCA_002092385.1	SAMN05981474	PRJNA352459	Contig	1.66343	38.1	1474	113	4	54	1	100/0.53	dairy
<b>L. mesenteroides subsp cremoris LMGCF13</b>	GCA_002092465.1	SAMN05981475	PRJNA352459	Contig	1.63644	38.3	1451	219	4	50	1	100/1.06	dairy
<b>L. mesenteroides subsp cremoris LMGCF14</b>	GCA_002092335.1	SAMN05981476	PRJNA352459	Contig	1.62752	38	1462	105	5	52	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF16</b>	GCA_002092515.1	SAMN05981478	PRJNA352459	Contig	1.61323	38	1448	124	5	52	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF17</b>	GCA_002092425.1	SAMN05981479	PRJNA352459	Contig	1.62949	38	1457	101	5	52	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF18</b>	GCA_002092495.1	SAMN05981480	PRJNA352459	Contig	1.62399	38	1456	105	5	52	1	100/0	dairy

<b>L. mesenteroides subsp cremoris LMGCF19</b>	GCA_002092415.1	SAMN05981481	PRJNA352459	Contig	1.64533	38	1517	113	5	53	1	100/0	dairy
<b>L. mesenteroides subsp cremoris LMGCF20</b>	GCA_002092455.1	SAMN05981482	PRJNA352459	Contig	1.63087	38	1474	109	5	53	1	99.74/0	dairy
<b>L. mesenteroides subsp cremoris LN07</b>	GCA_002092615.1	SAMN05981497	PRJNA352459	Contig	1.59621	38.1	1424	107	5	52	1	100/0	dairy
<b>L. mesenteroides subsp cremoris T26</b>	GCA_000686485.1	SAMN02767914	PRJNA205545	Scaffold	1.83247	38.4	1690	123	3	56	1	100/10.23	mesophilic cheese starter cultures
<b>L. mesenteroides subsp cremoris TIFN8</b>	GCA_000447945.1	SAMN02472108	PRJNA175676	Contig	1.71009	38.2	1452	173	7	61		99.21/0.26	propagated starter culture used in cheese manufacturing
<b>L. mesenteroides subsp dextranicum DSM20484 T</b>	GCA_001047695.1	SAMN03105774	PRJNA263697	Complete	1.85473	38.0389	1692	2	12	71	1	100/0.79	Cheese
<b>L. mesenteroides subsp dextranicum FAM 18356</b>	GCA_005864365.1	SAMN11653949	PRJNA543085	Contig	1.87153	37.8	1707	40	5	53	1	100/0	
<b>L. mesenteroides subsp dextranicum FM06</b>	GCA_002148235.1	SAMN06061940	PRJNA355067	Complete	1.94688	37.9649	1746	4	12	71	1	100/0	10 weeks old 45+ Samsø cheese
<b>L. mesenteroides subsp dextranicum LbT16</b>	GCA_001184265.1	SAMN03463677	PRJNA280520	Contig	1.90636	37.8	1733	65	3	51	1	99.47/0	Taleggio cheese
<b>L. mesenteroides subsp dextranicum LN05</b>	GCA_002092565.1	SAMN05981496	PRJNA352459	Contig	2.00187	38	1835	250	16	57	1	98.53/2.95	dairy
<b>L. mesenteroides subsp dextranicum LN08</b>	GCA_002092625.1	SAMN05981498	PRJNA352459	Contig	1.96519	37.8	1866	106	5	54	1	100/0	dairy
<b>L. mesenteroides subsp dextranicum LN32</b>	GCA_002092705.1	SAMN05981505	PRJNA352459	Contig	1.84828	37.8	1696	68	5	54	1	100/0	dairy
<b>L. mesenteroides subsp dextranicum LN34</b>	GCA_002092775.1	SAMN05981506	PRJNA352459	Contig	1.87513	37.7	1719	110	5	53	1	100/0	dairy
<b>L. mesenteroides subsp dextranicum NCTC10817</b>	GCA_900452955.1	SAMEA4362418	PRJEB6403	Contig	1.97325	37.8	1860	3	12	71	1	100/0.53	
<b>L. mesenteroides subsp dextranicum OG2</b>	GCA_002276945.1	SAMN06718482	PRJNA375758	Contig	1.71628	37.9	1583	146	7	51	1	100/0	kefir
<b>L. mesenteroides subsp dextranicum P45</b>	GCA_000756355.1	SAMN03032191	PRJNA260860	Contig	1.87419	37.5	1726	6	9	52	1	100/0	fermented pulque

<b>L. mesenteroides</b> subsp <b>dextranicum</b> <b>WC0331</b>	GCA_008690745.1	SAMN11618762	PRJNA542256	Scaffold	2.01813	37.6	1923	10	5	53	1	100/0.53	Cooked ham
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii B5</b>	GCA_004745755.1	SAMN10484942	PRJNA507123	Contig	2.12682	37.5	2037	26	5	53	1	100/0.18	Mike cake
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>BD3749</b>	GCA_001583825.1	SAMN04508169	PRJNA312878	Complete	1.98657	37.8	1898	1	12	71	1	100/0	fermeted vegetable
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>CBA7131</b>	GCA_003255835.1	SAMN07206913	PRJNA390217	Complete	2.08731	37.6375	1986	4	12	71	1	99.47/0	Korean adult feces
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>DRC1506 T</b>	GCA_001886915.1	SAMN04198570	PRJNA299366	Complete	1.98392	37.6424	1787	4	12	71	1	99.89/0.53	Kimchi
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii KFRI-</b> <b>MG</b>	GCA_000512955.1	SAMN02641526	PRJNA18679	Complete	1.89641	37.7	1796	1	12	69	1	100/0	
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>KIBGE-IB22</b>	GCA_005049065.1	SAMN11431368	PRJNA532993	Scaffold	1.94823	37.5	1880	13	3	50	1	100/0.18	cabbage(host:Vegetables)
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>SRCM102735</b>	GCA_009913935.1	SAMN08707610	PRJNA438180	Complete	2.05844	37.5883	1982	4	12	71	1	100/0	Soybean paste (Chonggugjang)
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>SRCM103356</b>	GCA_004102585.1	SAMN10742283	PRJNA515114	Complete	2.03939	37.5826	1984	4	12	71	1	100/0.18	food
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>SRCM103453</b>	GCA_004103675.1	SAMN10754493	PRJNA515383	Complete	2.1826	37.6453	2090	5	12	71	1	100/0	food
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>SRCM103460</b>	GCA_004194375.1	SAMN10743321	PRJNA515149	Complete	2.08721	37.5882	1996	5	12	71	1	100/0	food
<b>L. mesenteroides</b> subsp <b>jonggajibkimchii</b> <b>WC0333</b>	GCA_008689685.1	SAMN11618766	PRJNA542256	Contig	2.08859	37.5	2031	35	5	53	1	100/0	Sausage

<b>L. mesenteroides</b> subsp jonggajibkimchii Wikim33	GCA_003433375.1	SAMN07156024	PRJNA387486	Complete	1.97006	37.6362	1883	4	12	71	1	100/0	Baechu-kimchi
<b>L. mesenteroides</b> subsp mesenteroides CBA3607	GCA_009676745.1	SAMN13258970	PRJNA544551	Complete	1.85432	37.008	1750	3	12	71	1	100/0	kimchi
<b>L. mesenteroides</b> subsp mesenteroides 213M0	GCA_001570665.1	SAMD00042708	PRJDB4328	Contig	2.03285	37.7	1946	58	3	49	1	99.93/0.53	
<b>L. mesenteroides</b> subsp mesenteroides 406	GCA_001515385.1	SAMD00042709	PRJDB4329	Contig	2.00431	37.7	1932	69	3	49	1	100/1.06	
<b>L. mesenteroides</b> subsp mesenteroides ATCC8293 T	GCA_000014445.1	SAMN02598482	PRJNA315	Complete	2.07576	37.6586	1967	2	12	71	1	100/0.53	fermenting olives
<b>L. mesenteroides</b> subsp mesenteroides AtHG50	GCA_004368765.1	SAMN08777814	PRJNA441539	Scaffold	2.03285	37.7	1960	60	3	52	1	100/0.53	
<b>L. mesenteroides</b> subsp mesenteroides BD1710	GCA_002117185.1	SAMN04530070	PRJNA314170	Complete	2.12404	37.6	2014	1	12	71	1	100/0.53	kefir
<b>L. mesenteroides</b> subsp mesenteroides CECT9217	GCA_900290445.1	SAMEA104569583	PRJEB24702	Contig	2.02219	37.7	1932	13	5	53	1	100/0	
<b>L. mesenteroides</b> subsp mesenteroides CECT9266	GCA_900312985.1	SAMEA104569776	PRJEB24710	Contig	2.10613	37.7	2032	44	5	53	1	99.82/0	
<b>L. mesenteroides</b> subsp mesenteroides CECT9268	GCA_900290155.1	SAMEA104569777	PRJEB24712	Contig	2.07557	37.7	2002	26	5	53	1	99.82/0	
<b>L. mesenteroides</b> subsp mesenteroides DE0394	GCA_007673005.1	SAMN11792554	PRJNA543692	Scaffold	1.93918	37.7	1816	358	3	48	1	100/1.6	environmental
<b>L. mesenteroides</b> subsp mesenteroides DRC0211	GCA_002009375.1	SAMN04198273	PRJNA299363	Complete	2.1241	37.5981	1962	5	12	71	1	100/0	Kimchi
<b>L. mesenteroides</b> subsp mesenteroides GL1	GCA_001541125.1	SAMN04231183	PRJNA300958	Scaffold	1.81597	38.1	1649	11	2	35	1	100/0.79	Dromedary milk
<b>L. mesenteroides</b> subsp mesenteroides J18	GCA_000234825.3	SAMN02603136	PRJNA75083	Complete	2.01726	37.6784	1883	5	12	71	1	100/0	
<b>L. mesenteroides</b> subsp mesenteroides KMB608	GCA_003346165.1	SAMN09398940	PRJNA474823	Contig	1.95951	37.7	1877	81	3	46	1	100/0	sheep milk

<b>L. mesenteroides subsp mesenteroides KMB609</b>	GCA_003346155.1	SAMN09398941	PRJNA474823	Contig	2.01223	37.7	1930	84	3	47	1	100/0.53	bryndza cheese
<b>L. mesenteroides subsp mesenteroides KMB611</b>	GCA_003346345.1	SAMN09398943	PRJNA474823	Contig	2.10341	37.5	2037	33	3	46	1	100/0.53	bryndza cheese
<b>L. mesenteroides subsp mesenteroides LbE15</b>	GCA_001184245.1	SAMN03463642	PRJNA280507	Contig	2.00792	37.6	1923	63	3	51	1	100/0	Taleggio cheese
<b>L. mesenteroides subsp mesenteroides LbE16</b>	GCA_001184255.1	SAMN03463665	PRJNA280516	Contig	2.03609	37.5	1964	85	3	51	1	100/0.53	Taleggio cheese
<b>L. mesenteroides subsp mesenteroides LK-151</b>	GCA_002370415.1	SAMD00065790	PRJDB5233	Complete	2.13737	37.7366	2017	4	9	68	1	100/0	
<b>L. mesenteroides subsp mesenteroides LN25</b>	GCA_002092735.1	SAMN05981503	PRJNA352459	Contig	2.2776	37.5	2180	70	6	54	1	100/1.06	dairy
<b>L. mesenteroides subsp mesenteroides LN27</b>	GCA_002092745.1	SAMN05981504	PRJNA352459	Contig	2.2246	37.5	2123	54	6	54	1	100/1.06	dairy
<b>L. mesenteroides subsp mesenteroides M11</b>	GCA_009497095.1	SAMN13018494	PRJNA577063	Scaffold	1.97939	37.8	1863	79	3	52	1	100/1.06	(host:Oreochromis niloticus)
<b>L. mesenteroides subsp mesenteroides NBRC102481</b>	GCA_007989505.1	SAMD00166021	PRJDB6187	Contig	1.69543	37.8	1630	109	3	45	1	98.41/0	
<b>L. mesenteroides subsp mesenteroides NBRC3832</b>	GCA_006538765.1	SAMD00093693	PRJDB6025	Contig	1.90981	37.6	1789	31	3	45	1	100/0.53	
<b>L. mesenteroides subsp mesenteroides SRCM102733</b>	GCA_009913915.1	SAMN08707608	PRJNA438180	Complete	2.05873	37.7211	1967	3	12	71	1	100/0	Gochujang
<b>L. mesenteroides subsp mesenteroides TR154</b>	GCA_006406185.1	SAMN09099487	PRJNA470672	Contig	1.85995	37.8	1756	43	5	53	1	100/0	sourdough
<b>L. mesenteroides subsp mesenteroides WC0330</b>	GCA_008690805.1	SAMN11618760	PRJNA542256	Scaffold	1.76569	37.9	1637	41	4	54	1	100/0.26	Cooked ham
<b>L. mesenteroides subsp mesenteroides WC0332</b>	GCA_008689715.1	SAMN11618765	PRJNA542256	Contig	1.8553	37.7	1744	39	5	54	1	100/0	Sausage
<b>L. mesenteroides subsp mesenteroides Wikim17</b>	GCA_000787735.1	SAMD00022975	PRJDB3305	Contig	1.85932	37.8	1748	41	4	53	1	100/0.26	

<b>L. mesenteroides subsp mesenteroides YL48</b>	GCA_002886025.1	SAMN06298812	PRJNA371414	Contig	2.0584	37.6	1975	154	6	54	1	100/0	Root Surface(host:Carrot)
<b>L. pseudomesenteroides 1159</b>	GCA_000686465.1	SAMN02767913	PRJNA205546	Scaffold	2.03869	39	1969	100	3	49	1	100/0	mesophilic cheese starter cultures
<b>L. pseudomesenteroides 4882</b>	GCA_000297375.1	SAMEA3138421	PRJNA84285	Contig	2.00859	39.1	1936	106	5	59	1	93.12/0.18	
<b>L. pseudomesenteroides AMBR10</b>	GCA_901830415.1	SAMEA5665507	PRJEB32716	Contig	2.29673	39.2	2248	338	7	56	1	100/0.7	Human adenoid
<b>L. pseudomesenteroides BM2</b>	GCA_002092535.1	SAMN05981461	PRJNA352459	Contig	2.02255	39	1997	101	7	53	1	100/0	dairy
<b>L. pseudomesenteroides CBA3630</b>	GCA_008033175.1	SAMN11843675	PRJNA544551	Complete	2.31609	38.9999	2171	4	12	70	1	99.82/0.18	kimchi
<b>L. pseudomesenteroides HPK01</b>	GCA_002092075.1	SAMN05981330	PRJNA352459	Contig	1.95326	39.1	1853	92	5	51	1	100/0	dairy starter
<b>L. pseudomesenteroides KMB610</b>	GCA_003346375.1	SAMN09398942	PRJNA474823	Contig	1.9824	39	1919	57	3	43	1	100/0	bryndza cheese
<b>L. pseudomesenteroides LMG11482 T</b>	GCA_014634745.1	SAMD00239684	PRJDB7793	Contig	2.09581	38.9	1971	29	4	50	1	99.82/0	cane juice
<b>L. pseudomesenteroides LMGCF06</b>	GCA_002092035.1	SAMN05981468	PRJNA352459	Contig	1.95205	39.1	1848	83	5	51	1	100/0	dairy
<b>L. pseudomesenteroides LMGCF08</b>	GCA_002092165.1	SAMN05981470	PRJNA352459	Contig	2.19336	39.1	2069	267	5	53	1	100/11.2	dairy
<b>L. pseudomesenteroides LMGCF15</b>	GCA_002092375.1	SAMN05981477	PRJNA352459	Contig	1.92427	39.1	1833	78	5	51	1	100/0	dairy
<b>L. pseudomesenteroides LMGH100</b>	GCA_002072505.1	SAMN05981487	PRJNA352459	Contig	2.06006	39.1	1903	238	4	52	1	98.7/2.61	dairy
<b>L. pseudomesenteroides LMGH278</b>	GCA_002072495.1	SAMN05981488	PRJNA352459	Contig	1.9401	39.1	1852	106	5	51	1	100/0	dairy
<b>L. pseudomesenteroides LMGH280</b>	GCA_002072555.1	SAMN05981489	PRJNA352459	Contig	1.98015	39.1	1861	85	4	55	1	100/0.71	dairy



<b>L. pseudomesenteroides LMGH284</b>	GCA_002072575.1	SAMN05981490	PRJNA352459	Scaffold	2.07199	39	1952	222	4	52	1	99.21/2.03	dairy
<b>L. pseudomesenteroides LMGH61</b>	GCA_002072515.1	SAMN05981483	PRJNA352459	Scaffold	2.00878	39.1	1901	62	4	54	1	100/0	dairy
<b>L. pseudomesenteroides LMGH83</b>	GCA_002072475.1	SAMN05981484	PRJNA352459	Contig	2.05883	39.1	1885	201	4	53	1	98.4/1.41	dairy
<b>L. pseudomesenteroides LMGH95</b>	GCA_002072565.1	SAMN05981485	PRJNA352459	Contig	1.98982	39	1948	171	5	51	1	100/0.18	dairy
<b>L. pseudomesenteroides LMGH97</b>	GCA_002072585.1	SAMN05981486	PRJNA352459	Contig	1.98975	39	1946	132	5	50	1	100/0	dairy
<b>L. pseudomesenteroides LMGTW1</b>	GCA_002092235.1	SAMN05981491	PRJNA352459	Contig	1.9692	39	1924	125	5	52	1	100/0	dairy
<b>L. pseudomesenteroides LMGTW3</b>	GCA_002092355.1	SAMN05981492	PRJNA352459	Contig	1.89805	39.1	1834	87	5	52	1	100/0	dairy
<b>L. pseudomesenteroides LMGTW6</b>	GCA_002092255.1	SAMN05981493	PRJNA352459	Contig	1.88144	39.1	1821	90	5	52	1	100/0	dairy
<b>L. pseudomesenteroides LMGTW8</b>	GCA_002092295.1	SAMN05981494	PRJNA352459	Contig	1.98207	39	1936	118	5	52	1	100/0	dairy
<b>L. pseudomesenteroides LN02</b>	GCA_002092555.1	SAMN05981495	PRJNA352459	Contig	1.91243	39.1	1803	88	6	53	1	100/0	dairy
<b>L. pseudomesenteroides LN12</b>	GCA_002092635.1	SAMN05981499	PRJNA352459	Contig	1.86409	39.1	1762	85	6	52	1	100/0	dairy
<b>L. pseudomesenteroides LN23</b>	GCA_002092645.1	SAMN05981501	PRJNA352459	Contig	1.99273	39.1	1936	126	5	47	1	100/0.18	dairy
<b>L. pseudomesenteroides MGYG-HGUT-00096</b>	GCA_902363165.1	SAMEA5849597	PRJEB33885	Scaffold	2.29682			33	4	52	1	99.82/0	human gut
<b>L. pseudomesenteroides PS12</b>	GCA_000686505.1	SAMN02767912	PRJNA205547	Scaffold	1.93438	39.1	1863	91	3	48	1	100/0	undefined mesophilic cheese starter culture
<b>L. pseudomesenteroides TR070</b>	GCA_006382035.1	SAMN09099485	PRJNA470672	Contig	2.23055	38.8	2162	76	5	52	1	99.82/0	sourdough
<b>L. rapi DSM27776 T</b>	GCA_016908715.1	SAMN17619917	PRJNA695649	Contig	1.92491	38	1843	24	3	49	1	100/1.24	rutabaga

<b>L. suionicum CECT 8486</b>	GCA_900289255.1	SAMEA104569300	PRJEB24691	Contig	2.07002	37.3	1968	20	5	53	1	100/0.53	
<b>L. suionicum CECT 9216</b>	GCA_900290145.1	SAMEA104569335	PRJEB24693	Contig	2.06953	37.3	1972	18	5	53	1	100/0.53	
<b>L. suionicum CECT8484</b>	GCA_900289205.1	SAMEA104569299	PRJEB24690	Contig	2.06903	37.3	1974	18	5	53	1	100/0.53	
<b>L. suionicum DSM20241 T</b>	GCA_001891125.1	SAMN04811211	PRJNA318320	Complete	2.04883	37.5936	1923	2	12	71	1	100/0	Source unknown

522 **Table:2:** A descriptive list of the earlier classification and robust reclassification of the strain based on this study.

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Strains ID	Earlier taxonomy	Taxonomy status	Proposed taxonomy
<i>L. carnosum</i> CBA3620	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> DSM5576 T	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> JB16	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> MFPA29A1405	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> MFPC16A2803	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0318	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0319	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0320	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0321	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0322	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0323	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0324	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0325	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0326	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0327	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0328	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. carnosum</i> WC0329	<i>L. carnosum</i>	Correct	<i>L. carnosum</i>
<i>L. citreum</i> 1301 LGAS	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> CBA3621	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> CBA3623	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> CBA3624	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> CBA3627	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> CW28	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> DmW 111	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> EFEL 2700	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> F192-5	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> G4	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> KM20	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<i>L. citreum</i> LBAE C10	<i>L. citreum</i>	Correct	<i>L. citreum</i>

<b><i>L. citreum</i> LBAE C11</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> LBAE E16</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> NBRC102476</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> NRIC1776 T</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> NRRL B-1299</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> NRRL B-742</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> TMW21194</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> TR116</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> TR153</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. citreum</i> WiKim0101</b>	<i>L. citreum</i>	Correct	<i>L. citreum</i>
<b><i>L. falkenbergense</i> LMG10779 T</b>	<i>L. falkenbergense</i>	Correct	<i>L. falkenbergense</i>
<b><i>L. fallax</i> ATCC700006 T</b>	<i>L. fallax</i>	Correct	<i>L. fallax</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> C120c</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> C122c</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> KG16-1</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> KSL4-2</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> LMG18811 T</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> MFPA44A1401</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> PB1a</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> PB1e</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> PL111</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gasicomitatum</i> TMW 2.1619</b>	<i>L. gelidum</i> subsp <i>gasicomitatum</i>	Revisited	<i>L. inhae</i>
<b><i>L. gelidum</i> subsp <i>gelidum</i> JB7</b>	<i>L. gelidum</i> subsp <i>gelidum</i>	Correct	<i>L. gelidum</i>
<b><i>L. gelidum</i> subsp <i>gelidum</i> KCTC3527 T</b>	<i>L. gelidum</i> subsp <i>gelidum</i>	Correct	<i>L. gelidum</i>
<b><i>L. holzapfelii</i> CCUG54536 T</b>	<i>L. holzapfelii</i>	Correct	<i>L. holzapfelii</i>
<b><i>L. inhae</i> KCTC3774 T</b>	<i>L. inhae</i>	Correct	<i>L. inhae</i>
<b><i>L. kimchii</i> C2</b>	<i>L. kimchii</i>	Correct	<i>L. kimchii</i>
<b><i>L. kimchii</i> IMSNU11154 T</b>	<i>L. kimchii</i>	Correct	<i>L. kimchii</i>
<b><i>L. kimchii</i> NKJ218</b>	<i>L. kimchii</i>	Correct	<i>L. kimchii</i>
<b><i>L. lactis</i> 1300 LCIT</b>	<i>L. lactis</i>	Revisited	GS2
<b><i>L. lactis</i> aa 0143</b>	<i>L. lactis</i>	Revisited	GS2
<b><i>L. lactis</i> BIOML-A1</b>	<i>L. lactis</i>	Revisited	GS2
<b><i>L. lactis</i> CBA3625</b>	<i>L. lactis</i>	Revisited	GS2
<b><i>L. lactis</i> CBA3626</b>	<i>L. lactis</i>	Revisited	GS2

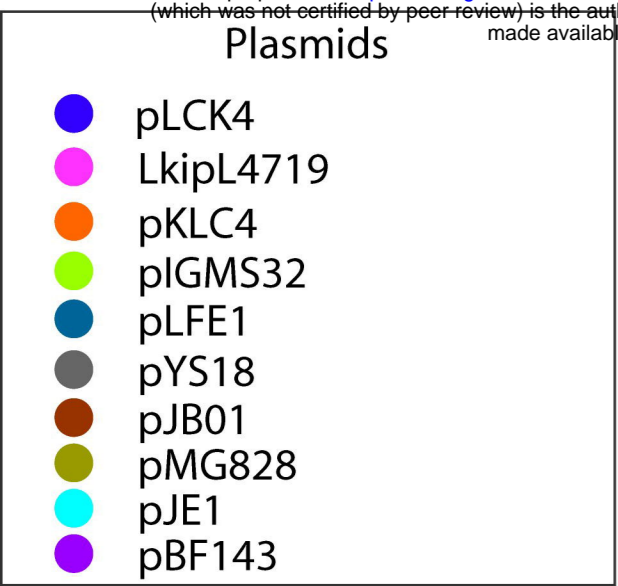
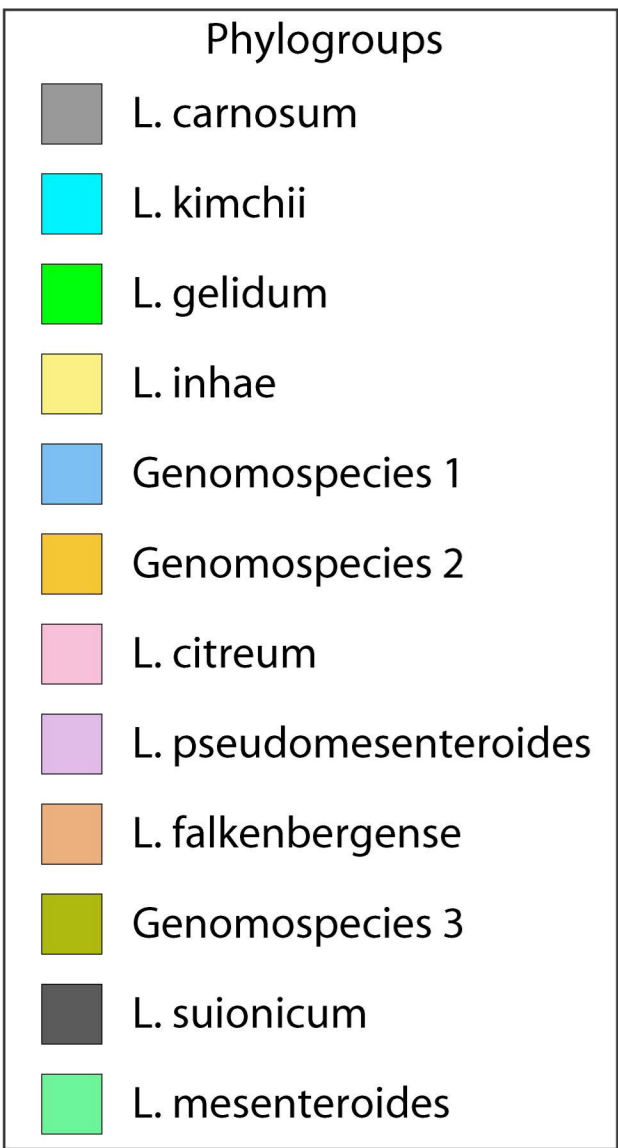
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<b><i>L. lactis</i> KCTC3773</b>	<i>L. lactis</i>	Revisited	GS1
<b><i>L. lactis</i> KFRI01</b>	<i>L. lactis</i>	Revisited	GS2
<b><i>L. lactis</i> LN19</b>	<i>L. lactis</i>	Revisited	GS1
<b><i>L. lactis</i> LN24</b>	<i>L. lactis</i>	Revisited	GS1
<b><i>L. lactis</i> NBRC 12455</b>	<i>L. lactis</i>	Revisited	GS2
<b><i>L. lactis</i> WiKim40</b>	<i>L. lactis</i>	Revisited	GS2
<b><i>L. litchii</i> MB7 T</b>	<i>L. litchii</i>	Correct	<i>L. litchii</i>
<b><i>L. mesenteroides</i> CBA3628</b>	<i>L. mesenteroides</i>	Revisited	GS3
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> ATCC19254 T</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF01</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF02</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF03</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF04</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF05</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF07</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF09</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF10</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF11</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF12</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF13</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF14</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF16</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF17</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF18</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF19</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LMGCF20</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> LN07</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> T26</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>cremoris</i> TIFN8</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> DSM20484 T</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> FAM 18356</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> FM06</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>

<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> LbT16</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> LN05</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> LN08</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> LN32</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> LN34</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> NCTC10817</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> OG2</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> P45</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>dextranicum</i> WC0331</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> B5</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> BD3749</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> CBA7131</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> DRC1506 T</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> KFRI-MG</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> KIBGE-IB22</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> SRCM102735</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> SRCM103356</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> SRCM103453</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> SRCM103460</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> WC0333</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>jonggajibkimchii</i> WiKim33</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> CBA3607</b>	<i>L. mesenteroides</i>	Revisited	GS3
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> 213M0</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> 406</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> ATCC8293 T</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> AtHG50</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> BD1710</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> CECT9217</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> CECT9266</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> CECT9268</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> DE0394</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> DRC0211</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> GL1</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> J18</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>

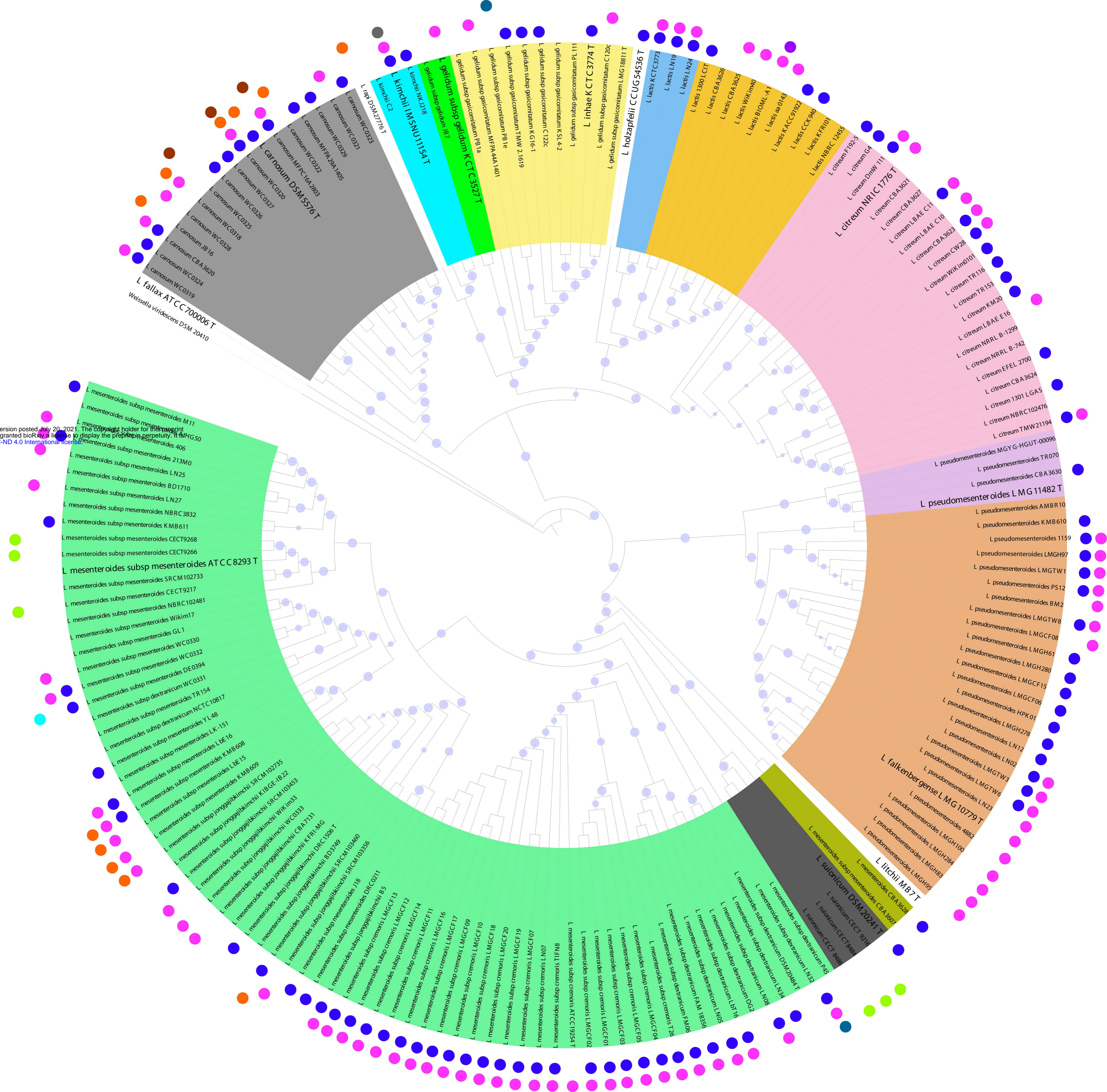
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<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> KMB609</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> KMB611</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> LbE15</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> LbE16</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> LK-151</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> LN25</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> LN27</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> M11</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> NBRC102481</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> NBRC3832</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> SRCM102733</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> TR154</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> WC0330</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> WC0332</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> Wikim17</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. mesenteroides</i> subsp <i>mesenteroides</i> YL48</b>	<i>L. mesenteroides</i>	Correct	<i>L. mesenteroides</i>
<b><i>L. pseudomesenteroides</i>1159</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> 4882</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> AMBR10</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> BM2</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> CBA3630</b>	<i>L. pseudomesenteroides</i>	Correct	<i>L. pseudomesenteroides</i>
<b><i>L. pseudomesenteroides</i> HPK01</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> KMB610</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> LMG11482 T</b>	<i>L. pseudomesenteroides</i>	Correct	<i>L. pseudomesenteroides</i>
<b><i>L. pseudomesenteroides</i> LMGCF06</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGCF08</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGCF15</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGH100</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
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<b><i>L. pseudomesenteroides</i> LMGH284</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGH61</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGH83</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falkenbergense</i>

<b><i>L. pseudomesenteroides</i> LMGH95</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGH97</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGTW1</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGTW3</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGTW6</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> LMGTW8</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> LN02</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> LN12</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> LN23</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> MGYG-HGUT-00096</b>	<i>L. pseudomesenteroides</i>	Correct	<i>L. pseudomesenteroides</i>
<b><i>L. pseudomesenteroides</i> PS12</b>	<i>L. pseudomesenteroides</i>	Revisited	<i>L. falckenbergense</i>
<b><i>L. pseudomesenteroides</i> TR070</b>	<i>L. pseudomesenteroides</i>	Correct	<i>L. pseudomesenteroides</i>
<b><i>L. rapi</i> DSM27776 T</b>	<i>L. rapi</i>	Correct	<i>L. rapi</i>
<b><i>L. suionicum</i> CECT 8486</b>	<i>L. suionicum</i>	Correct	<i>L. suionicum</i>
<b><i>L. suionicum</i> CECT 9216</b>	<i>L. suionicum</i>	Correct	<i>L. suionicum</i>
<b><i>L. suionicum</i> CECT8484</b>	<i>L. suionicum</i>	Correct	<i>L. suionicum</i>
<b><i>L. suionicum</i> DSM20241 T</b>	<i>L. suionicum</i>	Correct	<i>L. suionicum</i>

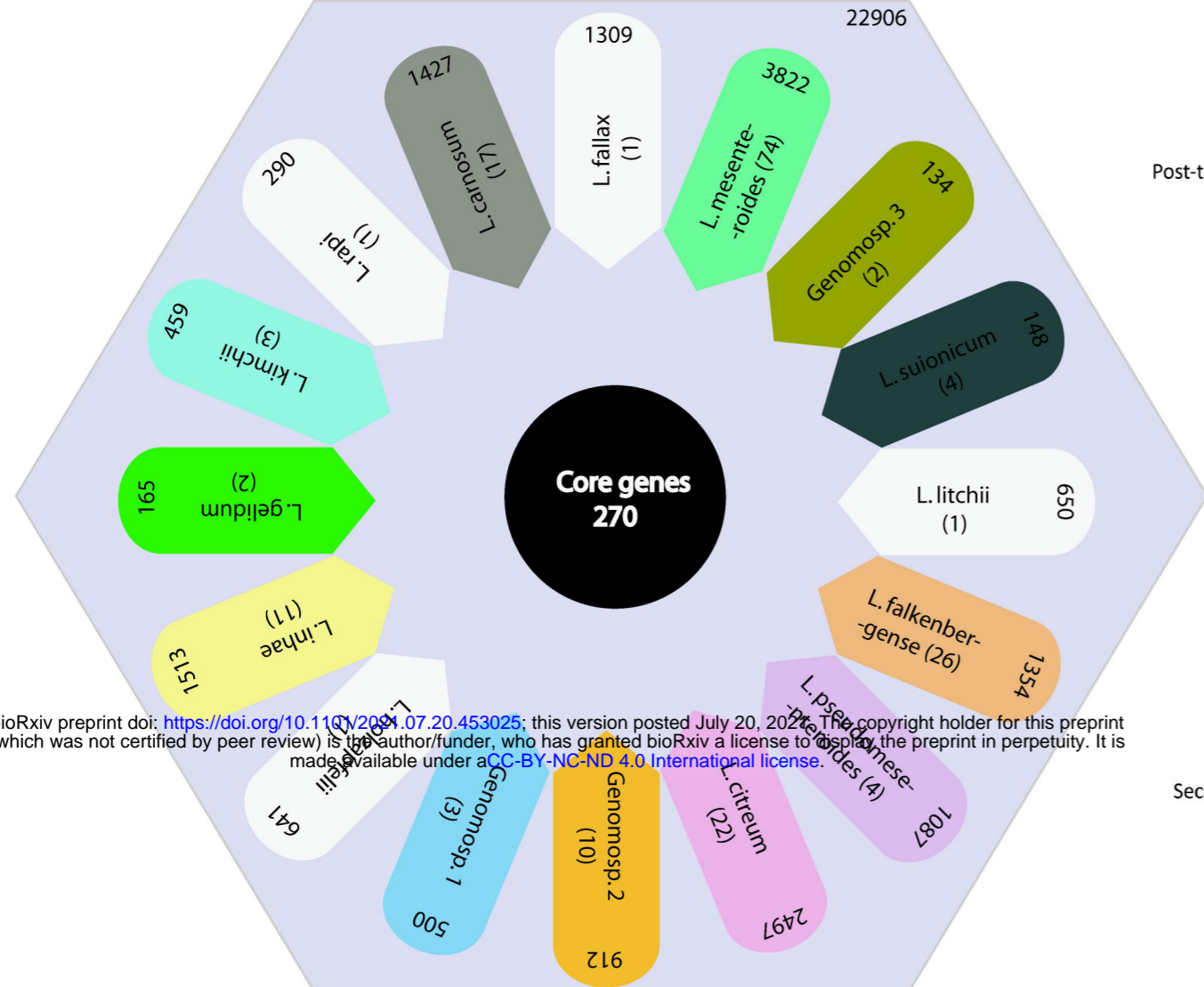




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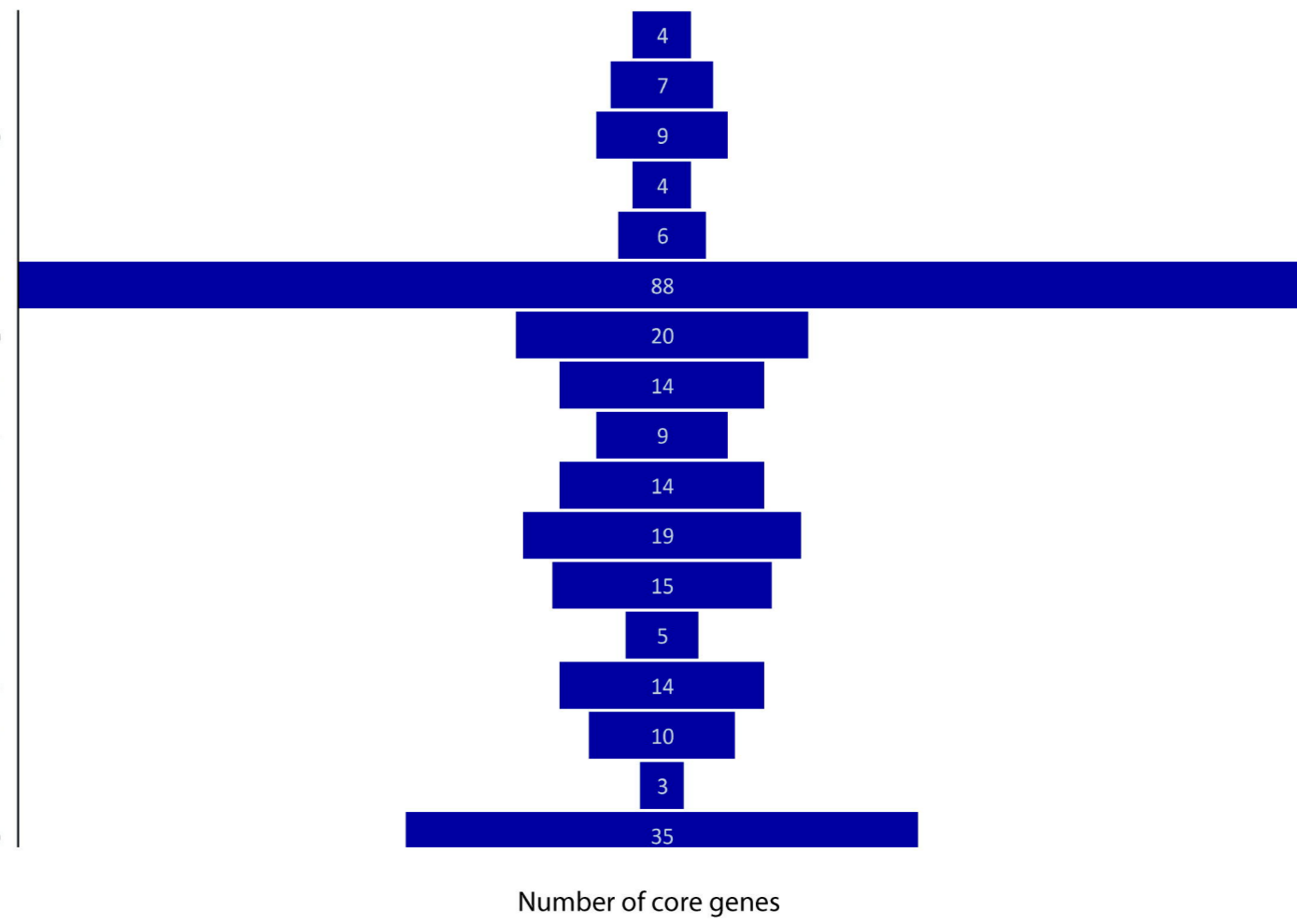




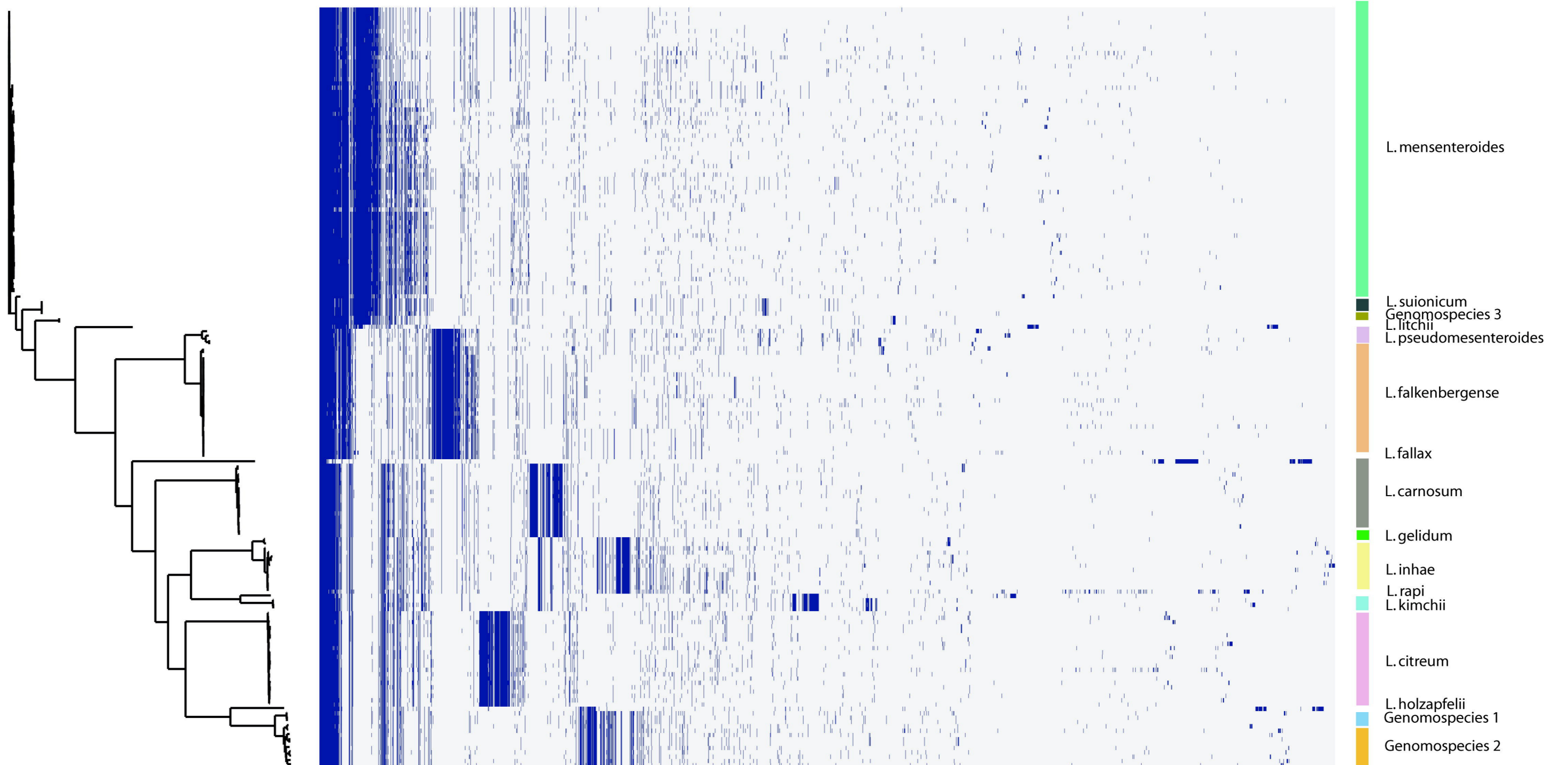
bioRxiv preprint doi: <https://doi.org/10.1101/2024.07.20.453025>; this version posted July 20, 2024. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

(A)

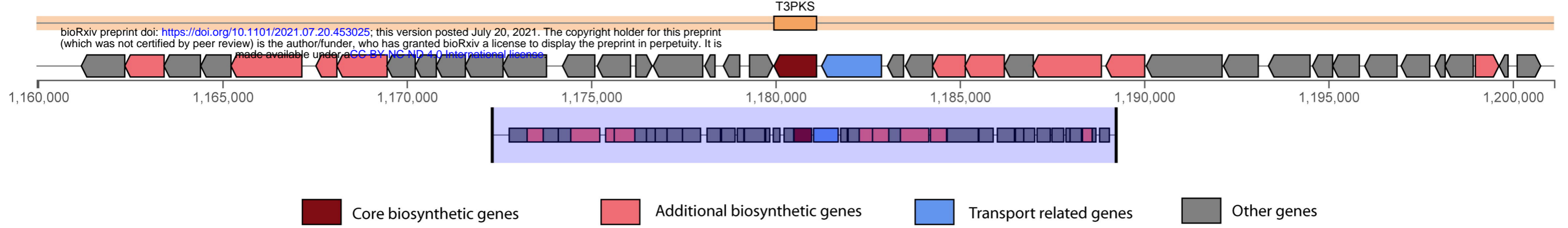
Cell cycle control, cell division, chromosome partitioning (D)  
 Cell wall/membrane/envelope biogenesis (M)  
 Post-translational modification, protein turnover, and chaperones (O)  
 Signal transduction mechanisms (T)  
 Intracellular trafficking, secretion, and vesicular transport (U)  
 Translation, ribosomal structure and biogenesis (J)  
 Transcription (K)  
 Replication, recombination and repair (L)  
 Energy production and conversion (C)  
 Amino acid transport and metabolism (E)  
 Nucleotide transport and metabolism (F)  
 Carbohydrate transport and metabolism (G)  
 Coenzyme transport and metabolism (H)  
 Lipid transport and metabolism (I)  
 Inorganic ion transport and metabolism (P)  
 Secondary metabolites biosynthesis, transport, and catabolism (Q)  
 Function unknown (S)



(B)

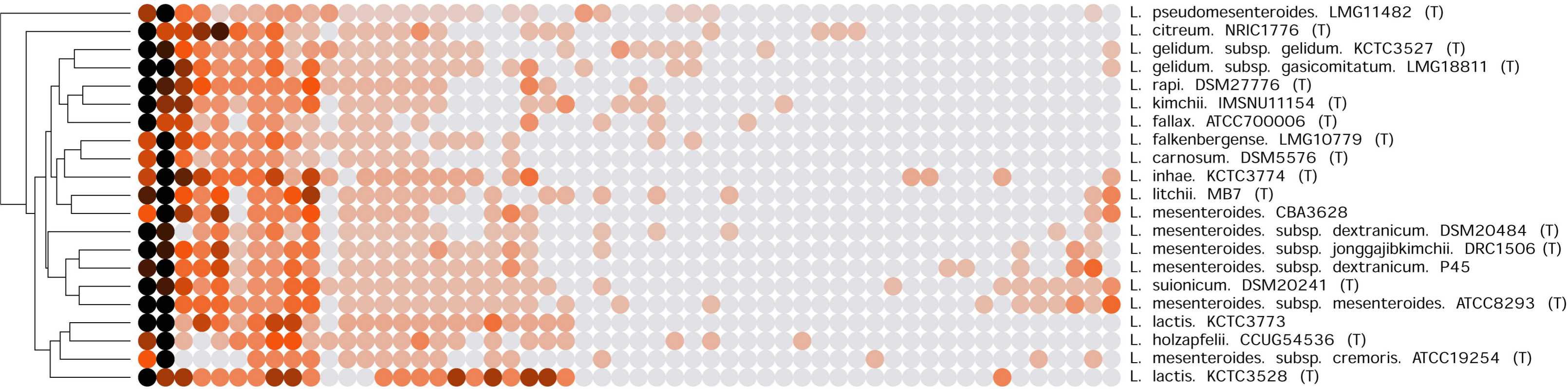
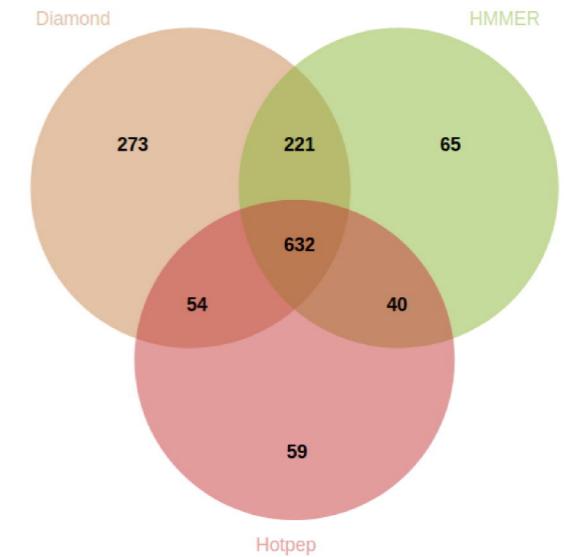
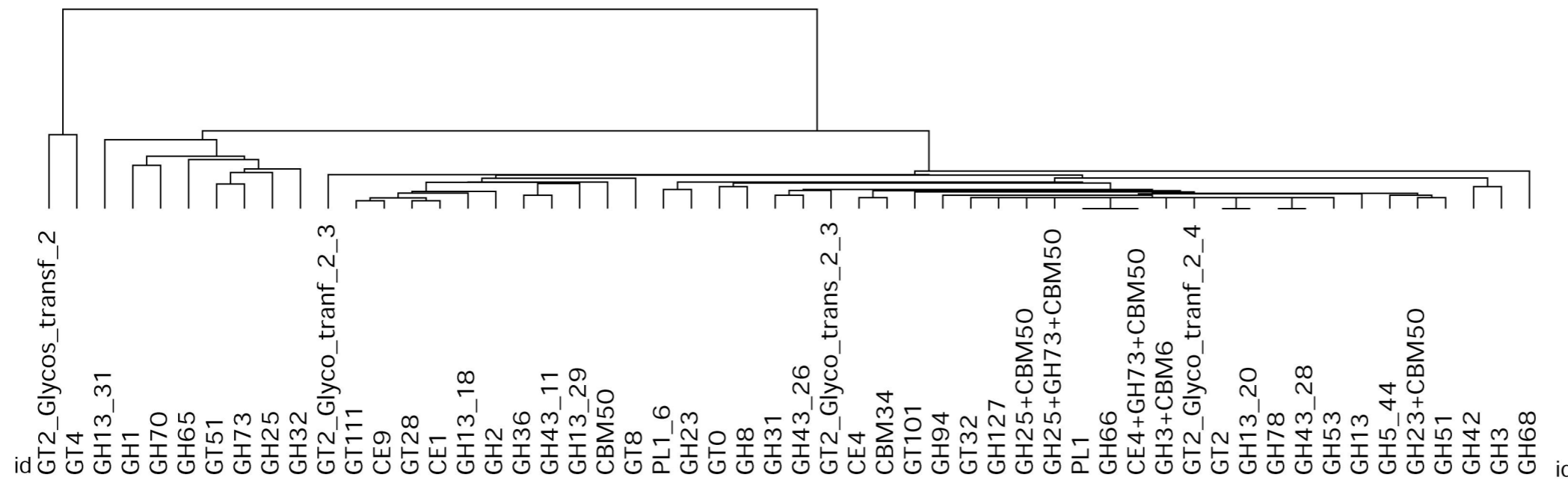


(C)



(A)

row min row max



(B)