Disentangling key species interactions in diverse and heterogeneous communities: A Bayesian sparse modeling approach

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Running Title: Sparse modeling of community interactions

Keywords: parameter shrinkage, pairwise interactions, environmental gradients, species diversity, coexistence, plant fecundity

Type of Article: Method

Number of Words: Abstract: 145, Main text: 4987 words, Text Box: 527

Number of References: 60

Number of Figures: 4

Number of Tables: 1

Number of Text Boxes: 1

Data accessibility statement: Upon acceptance, all data will be archived on Dryad and the data DOI will be included at the end of the article. Model code is available on GitHub, with the URL included in the manuscript. Upon acceptance, model code with be archived on Zenodo and the URL will be updated with the Zenodo link.

Statement of authorship: CWL and CMW contributed equally. CWL developed and implemented the statistical approach with input from CMW, CHB, OG and LGS. CMW developed and implemented the simulations. Data was contributed by MM. CWL, CMW, LMH, and LGS wrote the first manuscript draft. The manuscript was developed through conversations with the entire author list and all authors contributed to manuscript edits.

1 Abstract

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Modeling species interactions in diverse communities traditionally requires a prohibitively large number of 2 species-interaction coefficients, especially when considering environmental dependence of parameters. We 3 implemented Bayesian variable selection via sparsity-inducing priors on non-linear species abundance mod-4 els to determine which species-interactions should be retained and which can be represented as an average 5 heterospecific interaction term, reducing the number of model parameters. We evaluated model performance 6 using simulated communities, computing out-of-sample predictive accuracy and parameter recovery across 7 different input sample sizes. We applied our method to a diverse empirical community, allowing us to dis-8 entangle the direct role of environmental gradients on species' intrinsic growth rates from indirect effects 9 via competitive interactions. We also identified a few neighboring species from the diverse community that 10 had non-generic interactions with our focal species. This sparse modeling approach facilitates exploration 11 of species-interactions in diverse communities while maintaining a manageable number of parameters. 12

2 Introduction

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Understanding what maintains the diversity of life—where and how species abundances change through 14 time—has long fascinated and challenged ecologists. It is widely accepted that community composition in 15 any given time and place is driven by the interplay of species interactions, responses to environmental con-16 ditions, and feedbacks between local and regional dynamics (Chesson, 2000; HilleRisLambers et al., 2012; 17 Vellend, 2020). However, given the myriad of biotic interactions that may, themselves, be mediated by under-18 lying environmental conditions (Bulleri et al., 2016; Germain et al., 2018; Letten et al., 2018), feasibility and 19 model overfitting concerns quickly arise when trying to incorporate observed levels of diversity. Arguably, 20 the magnitude of this methodological limitation has even shaped our historical theoretical frameworks and 21 empirical tests. For example, classic species trait trade-offs, such as the competition-colonization trade-off, 22 apply for species pairs (Levins & Culver, 1971; Tilman, 1982). Similarly, while modern coexistence theory 23 (Chesson, 2000) can be applied to any level of species richness (Spaak & De Laender, 2020), the vast ma-24 jority of empirical studies focus on pairwise species comparisons (e.g. Kraft et al. 2015; Wainwright et al. 25 2019) and the effect of environmental variation on these comparisons (Bimler et al., 2018; Lanuza et al., 26 2018). Yet nonlinearity, higher-order interactions, and intransitivity in diverse systems may yield complex 27 dynamics that dramatically alter population growth and coexistence dynamics (Allesina & Levine, 2011; Li 28 et al., 2021; May & Leonard, 1975; Mayfield & Stouffer, 2017). The further development and empirical 29 testing of these theories thus requires a statistical approach that is applicable in diverse communities and is 30 capable of identifying and incorporating key species interactions and environmental covariates. 31

To date, empirical studies of population dynamics and species coexistence frequently take one of two 32 approaches for dealing with parameterization limitations that arise in diverse communities and varied envi-33 ronments. In the first approach, experimental studies focus on a few focal species. For example, Wainwright 34 et al. (2019) examined coexistence based on pairwise interaction coefficients between four annual forbs in 35 two locations and across two water availability treatments—a lofty number of species interaction coefficients 36 to estimate—but still a relatively small subset of the community's full diversity $(10 - 14 \text{ species in } 0.09 \text{ m}^2)$; 37 Dwyer et al. 2015). Finer-scale environmental variation can further limit the number of species that can be 38 feasibly incorporated: in a study of grass and forb coexistence under variable rainfall regimes, Hallett et 39

al. (2019) considered four rainfall conditions, requiring estimates of eight distinct species interaction coef ficients even with only two species. Isolating species interactions across environmental conditions is a high

barrier in species rich communities, even in laboratory and microcosm based studies (Letten et al., 2018).

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In the second approach, often used to interpret observational data, species are grouped into broad cat-43 egories. At the most extreme, a single interaction coefficient is then calculated between the focal species 44 and all heterospecific individuals—regardless of their identity (Clark et al., 2020a; Uriarte et al., 2004). 45 Heterospecifics may also be grouped more finely, for example, according to their taxonomic relationship 46 (Uriarte et al., 2004) or their origin status and life form (e.g. native versus exotic and grasses versus forbs) 47 (Martyn et al., 2020). Alternatively, functional groups can be created by grouping species according to their 48 traits (e.g. specific leaf area, canopy height, seed number) (Kühner & Kleyer, 2008; Uriarte et al., 2004). 49 However, this methodological approach often necessitates a priori knowledge of the system and makes an 50 underlying assumption that species grouped together will interact similarly with each other and with the 51 focal species. These assumptions are often not met (Mayfield & Levine, 2010), suggesting a need for a more 52 parsimonious and robust methodology that would allow the data to inform species groupings. 53

Various alternative statistical approaches have been proposed to assess species interactions using obser-54 vational data. For example, joint species distribution modeling has become a common approach to infer 55 species interactions from co-occurrence patterns (Legendre & Gauthier, 2014; Ovaskainen et al., 2019, 56 2017b). However, in addition to species interactions, patterns of co-occurrence may result from environ-57 mental sorting (Barner et al., 2018), or dispersal patterns (Schamp et al., 2015). Further, co-occurrence 58 patterns are scale dependent and regional analyses are not suited to assessing local-scale species interactions 59 (König et al., 2021). Recognizing a need to directly estimate species interaction coefficients, recent work has 60 expanded multivariate autoregressive models for use in more diverse communities (Picoche & Barraquand, 61 2020), including examining which linear combinations of species abundances best predict future growth 62 rates (Ovaskainen et al., 2017a). This approach is effective for binning species based on their competitive 63 effects, but does not account for variation in the environment. Clark et al. (2020b) recently developed a 64 state-space hierarchical Bayesian model to assess the effect of environmental gradients on nonlinear species 65 abundance patterns, incorporating environment responses in species' density-independent growth rates, but 66 not in species interactions (Clark et al., 2020b). Lastly, García-Callejas et al. (2020) developed a method to 67

incorporate environment responses in species' density-dependent growth rates but without the flexibility of Bayesian approaches. Independently, these different methodological developments each address one of the largest hurdles for modeling species abundances in diverse communities: (1) identifying important species interactions and (2) accounting for the mediating effect of the environment (here referred to as speciesenvironment interactions). Addressing these two aspects simultaneously would solidify a path forward for characterizing species interactions in diverse communities and across environmental gradients.

Here, we present an approach for modeling dynamics in diverse communities and across environmental 74 gradients. The approach balances realism and complexity without extensive experimental manipulation or 75 a priori assumptions regarding species groupings. Our method is based on two innovations to standard 76 population and community ecology models. First, we define heterospecific species interaction coefficients 77 as linear combinations of the average interaction strength and species-specific deviations from this average. 78 In parallel, we allow environmental covariates to modify species intrinsic growth rates and the strength 79 of biotic interactions—both the average and species deviation terms. We implement this approach using 80 a Beverton-Holt model of community dynamics (Beverton & Holt, 1957) within a single growing season, 81 although the method can easily be adapted to other models of population abundance (e.g. Mayfield & 82 Stouffer 2017; Ricker 1954) or incorporate additional dynamics such as seed banks or dispersal (Levine 83 & HilleRisLambers, 2009; Thompson et al., 2020). Second, we extend Bayesian statistical methods for 84 variable selection via sparsity-inducing priors in linear models (such as Lasso and Ridge regression; Hastie 85 et al. 2015; Piironen et al. 2017) to our non-linear abundance model, thereby reducing the number of terms 86 included in the final model fit, yielding a 'sparse model.' By coupling these two modeling approaches, we 87 can identify heterospecific species that deviate in their interaction strength, and how environmental gradients 88 alter species' density-independent growth rates and biotic interactions. We explore model effectiveness 80 using simulated data and apply the model to empirical data from a highly diverse (45 species) annual plant 90 community. 91

3 Methods

3.1 Deconstructing species interaction coefficients and fecundity

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⁹⁴ Models of community dynamics incorporate species-specific interaction coefficients for each species pair, ⁹⁵ commonly denoted as $\alpha_{i,j}$, the effect of species *j* on species *i*, resulting in a large number of parameters ⁹⁶ required to model diverse communities or environmental relationships. To reduce the number of parameters ⁹⁷ required to model diverse communities and incorporate environmental variation, we start with a partitioning ⁹⁸ approach. We first define these interaction terms as

$$ln(\alpha_{e,i,j}) = \bar{a}_{0,i} + \hat{a}_{0,i,j} + (\bar{a}_{e,i} + \hat{a}_{e,i,j})X_e$$
(1)

where $\alpha_{e,i,j}$ is the effect of species j on species i in environment e with $i \neq j$. In Eqn. 1, $\overline{a}_{0,i}$ is the effect 99 of an average heterospecific individual on individuals of species *i*, $\hat{a}_{0,i,j}$ is the deviation from this average 100 effect associated with species j, $\bar{a}_{e,i}$ is the average slope of species *i*'s interaction coefficients with environ-101 mental covariate X_e , and $\hat{a}_{e,i,j}$ is the deviation from this slope associated with species j. Upon first glance, 102 using Eqn. 1 may seem counter productive as it increases the number of parameters compared to traditional 103 interaction coefficients. However, in the next section we describe how coupling this approach with spar-104 sity inducing priors in a Bayesian context can dramatically reduce the number of required parameters by 105 identifying only the necessary species-specific terms ($\hat{a}_{0,i,j}$ and $\hat{a}_{e,i,j}$) for accurately modeling population 106 dynamics of species i. 107

While intraspecific competition ($\alpha_{e,i,i}$) could in principle be modeled according to Eqn. 1, we instead define it separately as:

$$ln(\alpha_{e,i,i}) = a_{0,i,i} + a_{e,i,i}X_e \tag{2}$$

where $a_{0,i,i}$ and $a_{e,i,i}$ are the intercept and slope for the effect of intraspecific individuals. As both theoretical expectations (Chesson, 2000) and empirical results (Adler *et al.*, 2018) point to the importance of intraspecific competition, we use Eqn. 2 to explicitly exclude the intraspecific terms from the sparsity inducing process defined in the next section. These terms, therefore, will always be included in the final model fit.

Interaction coefficients (Eqns. 1 & 2) can be incorporated in many different models of community dynamics. We use the Beverton-Holt model due to its legacy in studies of annual plant communities and coexistence theory (e.g. Godoy & Levine 2014; Kraft *et al.* 2015). We emphasize, however, that our general statistical approach can be adapted to other population models. In the Beverton-Holt model, the fecundity $F_{e,i}$ of a focal species *i* in environment *e* is modeled as:

$$F_{e,i} = \frac{\lambda_{e,i}}{1 + \alpha_{e,i,i}N_{e,i} + \sum_{j \neq i}^{S} \alpha_{e,i,j}N_{e,j}}$$
(3)

Fecundity depends on a species' intrinsic growth rate (i.e. density-independent seed production; $\lambda_{e,i}$) and the competitive effects of all *S* species in the community ($\alpha_{e,i,j}$ terms as defined by Eqns. 1 &2) scaled by each species' abundance ($N_{e,j}$) (Levine & HilleRisLambers, 2009; Pérez-Ramos *et al.*, 2019; Shoemaker & Melbourne, 2016). To incorporate environmental variation in intrinsic growth rates, we model $\lambda_{e,i}$ as:

$$ln(\lambda_{e,i}) = b_{0,i} + b_{e,i}X_e \tag{4}$$

where $b_{0,i}$ is the intercept of the intrinsic growth rate and $b_{e,i}$ its slope with environmental covariate X_e . We use Eqn. 3 to model observed fecundity within a single growing season of both simulated and empirical data.

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3.2 Incorporating sparsity-inducing priors

By deconstructing interaction coefficients into a combination of species-specific and generic terms, we can determine which, if any, species-specific terms are necessary for the final model. Allowing only a subset of parameters to take non-zero values is referred to as 'sparse modeling,' and various techniques exist to induce sparsity in linear models (Hastie *et al.*, 2015; O'Hara *et al.*, 2009).

To extend a sparse modeling approach to our non-linear model of fecundity (Eqn. 3), we employ sparsityinducing priors which act to shrink all but a subset of parameters to 0, thus producing a sparsely parameterized model. Specifically, we model $\hat{a}_{0,i,j}$ and $\hat{a}_{0,i,j}$, the species-specific intercepts and slopes of the inter-

specific interaction coefficients (Eqn. 1), with regularized horseshoe priors which more accurately estimate 134 large parameter values compared to other sparsity-inducing priors (Bhadra et al., 2019; Carvalho et al., 2009; 135 Piironen *et al.*, 2017; Van Erp *et al.*, 2019). Parameters $\hat{a}_{i,j}$ and $\hat{a}_{e,i,j}$, are given priors Normal $(0, \tau \beta_{0,j})$ and 136 Normal $(0, \tau \beta_{e,j})$ respectively. (Note that since we fit the model for a single focal species, we drop the *i* 137 subscript from the priors for simplicity.) In these priors, τ defines the global tendency towards sparsity 138 through its effect on the priors' standard deviations. In other words, with smaller values of τ , the priors for 139 all $\hat{a}_{i,j}$ and $\hat{a}_{e,i,j}$ parameters become more tightly centered on 0. Conversely, the β terms allow specific pa-140 rameters to escape this global trend towards sparsity. As an individual $\tilde{\beta}$ term becomes large, its associated 141 prior becomes wider, and that species-specific term is more likely to be included in the final model. In the 142 regularized horseshoe prior, these $\tilde{\beta}$ terms are defined as: 143

$$\widetilde{\beta}_{j} = \frac{c\beta_{j}}{\sqrt{c^{2} + \tau\beta_{j}^{2}}}$$

$$\beta_{j} \sim \text{half-Cauchy}(0, 1) \tag{5}$$

$$c^{2} \sim \text{inverse-gamma}(\frac{\nu}{2}, \frac{\nu s^{2}}{2})$$

¹⁴⁴ Defining $\hat{\beta}_j$ as the combination of a half-Cauchy and inverse-gamma distribution causes large coefficients to ¹⁴⁵ be shrunk towards 0 by a Student's *t* distribution with *v* degrees of freedom and a scale of s^2 (Piironen *et al.*, ¹⁴⁶ 2017; Van Erp *et al.*, 2019). Following the recommendations of Piironen and Vehtari (2017), we set *v* to 4 ¹⁴⁷ and s^2 to 2. Rather than setting the global shrinkage parameter τ to a fixed value, we give it a half-Cauchy ¹⁴⁸ prior with scale parameter equal to 1 ($\tau \sim$ half-Cauchy(0,1)) and allow the data to inform the posterior ¹⁴⁹ distribution of τ (Piironen *et al.*, 2017; Van Erp *et al.*, 2019).

We employ a hybrid approach in which we first fit the full model with regularized horseshoe priors to induce sparsity in the species-specific terms; we subsequently fit a final model using traditional, non-sparse methods. From the preliminary model fit, we identify which species-specific terms have sufficient evidence to be included in the final model fit. We calculate credible intervals (CIs) for each species-specific term in the preliminary model and include in the final model only those terms whose intervals do not overlap 0. By

using this approach, we can directly adjust how conservative we wish to be in including model parameters, 155 balancing model prediction, the proportion of variance explained, and simplicity depending on modeling 156 goals (Tredennick et al., 2021) (i.e. using a 50% CI will lead to models including more parameters than if 157 we use a 95% CI). Then, for the final model fit, the included species-specific terms $(\hat{a}_{0,i,j} \text{ and } \hat{a}_{e,i,j})$ are given 158 standard normal priors (i.e. Normal(0,1)). In both preliminary and final model fits, the terms defining $\lambda_{e,i}$ 159 $(b_{0,i} \text{ and } b_{e,i}; \text{Eqn 4})$ are also given standard normal priors. The intercept and slope terms defining intraspe-160 cific competition ($a_{0,i,i}$ and $a_{e,i,i}$; Eqn.2) and the generic intercept and slope defining interspecific competition 161 $(\overline{a}_{0,i} \text{ and } \overline{a}_{e,i}; \text{Eqn. 1})$ are both given weakly informative priors in each model fit, matching the expected scale 162 of these interaction coefficients: $a_{0,i,i} \sim \text{Normal}(-6,3)$, $\overline{a}_{0,i} \sim \text{Normal}(-6,3)$, $a_{e,i,i} \sim \text{Normal}(0,0.5)$, and 163 $\bar{a}_{e,i} \sim \text{Normal}(0, 0.5)$. All models were fit using the stan language with the rstan package (version 2.18.2; 164 Stan Development Team 2018 in R (version 3.5.3; R Core Team 2019). All code for the analyses and 165 simulations presented here can be found at https://github.com/tpweiss06/SparseInteractions. 166

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3.3 Simulation tests of model performance

To test our ability to predict changes in population size and recover true parameter values, we first paired 168 our Bayesian sparse modeling approach with simulated Beverton-Holt data using Eqns. 1-4. For the simu-169 lations, we generated communities of 15 species in different plots, where each plot was a unique run of the 170 simulation for a given community with a given environmental condition X_e . We aimed to generate popula-171 tion growth rates comparable to those found in a community adapted to its environment. Each species was 172 assigned an intrinsic growth rate $\lambda_{e,i}$ following Eqn 4 (Table 1), pairwise species competitive interactions 173 $\alpha_{e,i,j}$ were composed of the generic competition term $\overline{a}_{0,i}$ with small amounts of variation and a generic envi-174 ronmental response $\overline{a}_{e,i}$. Seven randomly selected species also had a non-generic competition term through 175 species-specific deviations from $\overline{a}_{0,i}$ ($\hat{a}_{0,i,j}$). Seven separately selected species had a non-generic environ-176 mental response through species-specific deviations $\hat{a}_{e,i,j}$. Intraspecific competition $\overline{a}_{0,i,i}$ was set as a fixed 177 value higher than interspecific competition to minimize extinction in the simulations (Table 1). Each plot 178 simulation was run deterministically for 20 time steps with each time step $N_{t+1} = F_t N_t$ using F_t from equa-179 tion 3. This resulted in some subset of the 15 species remaining with populations greater than zero in each 180

plot. Then each population was perturbed by drawing from a normal distribution with mean and standard deviation equal to the previous population size, truncated at 0 to prevent negative population sizes. This perturbed state and the following time step generated our simulated 'full-community' data. In addition to 500 full-community plots, we simulated 500 'no-competition' treatments with a single phytometer individual of the focal species per plot, running the Beverton-Holt function for one time step. This simulated treatment matches methods commonly used in experimental studies to parse intrinsic growth rates from competition parameters (Hallett *et al.*, 2019; Wainwright *et al.*, 2019). Simulation details are included in Supplement 1.

We used these simulations to measure our sparse modeling approach's ability to predict population growth in diverse communities and recover underlying parameters. We selected one focal species and tested our model's performance using varying numbers of full-community and no-competition plots. We tested out-of-sample predictions on 200 full-community plots not used to fit the model. We then calculated the posterior distribution of the root-mean-square error (RMSE) of model predictions compared to true values for each model fit. This allowed us to quantify the gain in predictive accuracy resulting from including more data.

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3.4 Empirical application

We additionally applied our model to species interactions and their environmental dependencies in the an-196 nual plant understory of the York gum (Eucalyptus loxophleba Benth) - jam (Acacia acuminata) woodlands 197 of southwestern Western Australia. This community is highly diverse and heterogeneous, with local compo-198 sition of annual forbs and grasses influenced by gradients in soil nutrients and shade from York gum and jam 199 trees (Dwyer et al., 2015; Lai et al., 2015). We focused on two York gum-jam woodland remnants: West 200 Perenjori Nature Reserve (29°47'S, 116°20'E) and Bendering Nature Reserve (32°23'S, 118°22'E). Both 201 sites experience a Mediterranean climate with mild winters and long, dry summers (Suppiah et al., 2007) 202 and have high overlap in annual species composition, sharing several dominant species. Data used for this 203 study were originally collected as part of a larger experiment described in full in Wainwright et al. (2019). 204 We focus on two species used as focal species in the original study and common to both reserves: Waitzia 205 *acuminata*, an abundant native annual forb, and *Arctotheca calendula*, a prevalent exotic annual forb. 206

We used data from 11 experimental blocks in Bendering Nature Reserve and 18 blocks in West Perenjori 207 Nature Reserve. Each block was $\approx 15 \text{ x} 15 \text{ m}$, a size selected to account for previously identified soil-nutrient 208 turn-over rates (Dwyer et al., 2015). Each block was split into 50 x 50 cm plots and each plot was further 209 subdivided into four 25 x 25 cm quadrats. One individual of either focal species near the center of each 210 quadrat was assigned as the focal individual for that quadrat. Which focal species were in a given quadrat 211 depended on the natural distribution of individuals. This experiment employed five thinning treatments at 212 the plot level to manipulate local community compositions (individual focal individuals with no competitors, 213 native dominated competitors, exotic dominated competitors, monocultures with only conspecific competi-214 tors, and unmanipulated plots) (Wainwright et al., 2019). This ensured a range of observed densities of both 215 species and the background communities to inform model estimates of competition coefficients and intrinsic 216 growth rates. Across both reserves we used data from 129 focal individuals in 69 plots interacting with 45 217 neighbouring species for W. acuminata and 95 focal individuals in 54 plots interacting with 40 species for 218 A. calendula. 219

We applied our sparse modeling framework to quantify the effect of the competitive environment on 220 fecundity in W. acuminata and A. calendula under different environmental conditions. Fecundity $F_{e,i}$ was 221 measured as the number of flowers produced by each focal individual. The competitive environment was 222 characterized as the number of individuals of each interacting species in the quadrat after the experimental 223 treatment had been applied $(N_{e,i})$. We considered two aspects of the physical environment X_e : percent 224 overhead tree canopy cover, measured at the plot scale, and soil Colwell P (mg/kg), measured at the block 225 scale. Both environmental covariates were standardized for inclusion in the model. We ran a separate model 226 for each focal species and environmental covariate, for a total of four model fits. To account for regional 227 differences between the Bendering and Perenjori reserves, we incorporated a fixed effect for the two different 228 reserves into our sparse modeling approach by allowing λ_i , $\lambda_{e,i}$, $\hat{\alpha}_{i,j}$, and $\hat{\alpha}_{e,i,j}$ to differ between reserves. 229 Using this approach, we quantified $\lambda_{e,i}$ and $\alpha_{e,i,j}$ for both species across both environmental gradients in the 230 York gum-jam woodland communities. 231

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4 Results

4.1 Simulations

Our model accurately predicted growth rates for simulated communities even with relatively low sample 234 sizes (Fig. 1) and across different model formalizations (Box 1). With only 10 full-community and 10 235 no-competition plots, the model predicted growth rates with a root-mean-square error (RMSE) of 0.495 236 (credible interval, CI: 0.353-0.665). While increasing sample size further increased model accuracy (RMSE 237 of 0.315 (CI: 0.211-0.520) for 50 plots and 0.227 (CI 0.195-0.288) for 200 plots), these results indicate the 238 model can accurately predict species' realized growth rates using limited data. Furthermore, species' growth 239 rates can be accurately predicted using observed competitive communities paired with no-competition plots, 240 rather than necessitating common manipulative experimental designs where each possible species combina-241 tion is paired across a gradient of densities (Hallett et al., 2019; Kraft et al., 2015). 242

Our model was also able to accurately predict individual parameter estimates for simulated communities 243 (Fig. 1). In particular, estimates of the intercept and slope parameters for intrinsic growth rate $(b_0, i \text{ and } b_0)$ 244 $b_{e,i}$ respectively) dramatically increased in accuracy from 10 to 200 data points (Fig. 1a,e). The accuracy 245 of the estimates for the slope and intercept of intraspecific competition ($a_{0,i,i}$ and $a_{e,i,i}$ respectively) also 246 increased with more data, but less dramatically than the terms defining intrinsic growth rate. Parameters 247 associated with interspecific competition ($\overline{a}_{0,i}$ representing the intercept and $\overline{a}_{e,i}$ for the slope) also increased 248 in accuracy with increasing data, although there was more variance in this relationship. This is likely because 249 the model correctly identified a larger number of species-specific terms with more data, which decreased the 250 total number of species contributing to the estimation of $\overline{a}_{0,i}$ and $\overline{a}_{e,i}$. When fit to only 10 simulated plots, 251 the model did not identify any species-specific terms $(\hat{a}_{0,i,j} \text{ or } \hat{a}_{e,i,j})$ and only used $\overline{a}_{0,i}$ and $\overline{a}_{e,i}$. The model 252 identified two species-specific terms within a single species when fit to 50 plots and eight species-specific 253 terms across six species when fit to 200 plots. In general, the estimates of species-specific terms were highly 254 accurate; only two out of the eight estimated species-specific interaction terms did not include the true value 255 in their 95% credible intervals. 256

4.2 Empirical application

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Our method identified environmental dependencies in intrinsic growth rates (Fig. 3a,b; 4, a,b), relative strengths of intraspecific competition and average interspecific competition, along with competition-environment interactions (Fig. 3c,d; 4, c,d), all of which differed between our two focal species. Additionally, our model highlighted three species with deviations from the average interspecific effects on native *W. acuminata*, but no such species when fit to data on exotic *A. calendula*.

W. acuminata and A. calendula's intrinsic growth rates differed in their relationship with the environmen-263 tal gradients and reserves. The intrinsic growth rate of W. acuminata across both environmental gradients 264 varied between the Bendering and Perenjori reserves (Fig. 3a,b). In contrast, $\lambda_{e,i}$ for A. calendula was quite 265 similar between the two reserves as it varied with both phosphorous and canopy cover (Fig. 4a,b). This 266 could reflect local adaptation in regional populations of the native W. acuminata but not in the newly intro-267 duced A. calendula. Importantly, the intrinsic growth rate of W. acuminata declined with high phosphorous 268 (marginally in Bendering, but substantially in Perenjori) while A. calendula's intrinsic growth rate increased 269 with phosphorous, potentially explaining the high prevalence of invasive species in areas with increased 270 phosphorous (Dwyer et al., 2015). 271

Relative effects of competition between conspecifics versus heterospecifics also differed between the two 272 focal species. For W. acuminata, the relationship between intraspecific competition and average interspecific 273 competition varied with the underlying environmental gradients. At low levels of phosphorous and high 274 levels of canopy cover, intraspecific competition in W. acuminata was greater than average interspecific 275 competition (Fig. 3c,d). However, at high levels of phosphorous and low levels of canopy cover, intra-276 and interspecific competition converged to similar values. On the other hand, intraspecific competition 277 for A. calendula was similar to or lower than generic interspecific competition across both environmental 278 gradients (Fig. 4c,d). This likely contributes to the invasive status of A. calendula in this ecosystem, whereas 279 W. acuminata populations self-regulate under certain environmental conditions—a necessary component of 280 stable coexistence. 281

Our model highlighted multiple species with competitive effects on *W. acuminata* that differed from the generic interaction term. Across the observed gradient in phosphorous, *Hyalosperma glutinosum* had

a higher than average effect on W. acuminata in the Perenjori reserve while Schoenus nanus had a lower 284 than average effect in the Bendering reserve (Fig. 3c). Across the observed gradient in canopy cover, 285 Hypochaeris glabra had a much higher than average effect on W. acuminata in Bendering (Fig. 3d). In 286 contrast, all heterospecific interactive effects on A. calendula remained grouped in the generic competition 287 term. The lack of species with unique effects on A. calendula (Fig. 4c,d) could be due to its exotic status 288 (Lai et al., 2015). With no shared evolutionary history with any other community members, A. calendula 289 could be experiencing a form of competitive release, wherein the identity of competitor species matters less 290 than simply the presence of additional individuals. 291

5 Discussion

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Given the inherent complexity of ecological communities, ecologists are often forced to rely on simplify-293 ing assumptions in order to perform tractable analyses, such as limiting the number of species considered 294 or ignoring environmental variation. The sparse modeling approach presented here provides an alternative 295 method to analyze community data without requiring extensive additional data or sacrificing complexity. 296 This approach enabled us to accurately predict population growth rates with limited data and identify how 297 species' demographic rates and competitive interactions depend on the environment. Our results identify en-298 vironment by species interactions that deviate from the species-averaged community effects without making 299 a priori assumptions about species groupings (Figure 3c, d). This information and output from the sparse 300 modeling approach generates concrete, testable hypotheses about species interactions and environmental 301 conditions. We see broad potential for this method's implementation in community ecology, from theory 302 development to management applications. 303

The sparse modeling approach's flexibility in modeling populations and communities allows easy adjustments for the best match between underlying model structure and the given study system and research questions. As we show in Box 1 and Fig. 2, these models can successfully be applied to different forms of species-environment interactions and be modified to be more or less complex based on underlying ecological questions and data availability. For example, the functional form of the relationship between intrinsic growth rate and the environment likely depends on a study's spatial scale. For localized studies, a sim-

ple monotonic relationship (Figure 2a) might be appropriate to capture species' expected responses across 310 a small range of environmental variation. However, studies over larger spatial scales might require a func-311 tional form with optimal intrinsic growth reached at an intermediate environmental value and declining away 312 from that value (Figure 2b), mimicking expected patterns of adaptation across species' ranges (Angert et al., 313 2020). Additionally, while we used a Beverton-Holt framework in our examples (Beverton & Holt, 1957), 314 the sparse approach is agnostic to the underlying ecological model. Thus, it could be used with different 315 functional forms of competition (García-Callejas et al., 2020) with models incorporating both competitive 316 and facilitative interactions (Stachowicz, 2001) or different underlying demography such as seed banks. 317

With this flexibility, sparse modeling has the potential to be a powerful tool to accelerate the develop-318 ment of community ecology theory and practice. It can provide important insights into the covariation of 319 environmental conditions, species' demographic rates, and competitive effects-critical aspects of modern 320 coexistence theory (Chesson, 2000). This includes quantifying the relative strengths of intra- versus inter-321 specific competition, which is a key condition for stable coexistence (Adler et al., 2018; Chesson, 2000). 322 Furthermore, the approach elucidates the effect of environmental conditions on species' density-independent 323 growth rates versus competitive interactions, potentially allowing for quantification of variation-dependent 324 coexistence mechanisms, such as the storage effect, in diverse communities (Chesson, 2000). Similarly, out-325 put from our sparse modeling approach across environmental gradients can be used to quantify the relative 326 importance of environmental (abiotic) filtering, biotic interactions, and the joint effect on species occurrence 327 (Cadotte & Tucker, 2017). Applying such an approach is especially exciting for linking community theory 328 to global change predictions, depending on the underlying environmental gradient of interest. 329

In addition to expanding theory, we see exciting potential for sparse modeling to address questions in 330 applied contexts and generate new hypotheses from existing datasets that inform management strategies. 331 This includes quantifying how environmental modifications can be used in conjunction with community 332 manipulations to control invasive species or promote native species. For example, our results from the York 333 gum-jam woodlands of Western Australia suggest the native W. acuminata experiences declining fitness 334 with increasing levels of phosphorous, particularly in the Perenjori reserve (Fig. 3a). At the same time, 335 the model identified H. glutinosum as having a stronger than average competitive impact on W. acuminata 336 in Perenjori (Fig. 3c). Taken together, these results suggest that reserve managers could help maintain or 337

expand populations of *W. acuminata* by mitigating phosphorous run-off while simultaneously removing *H. glutinosum* in key locals. In contrast, our results for the invasive *A. calendula* suggest that neighbor species
 identity is unimportant (Fig. 4c,d) and management strategies focusing solely on environmental factors
 would be most impactful.

Beyond the implementation of the sparse modeling approach presented here, the underlying model struc-342 ture can be further adjusted to align the model with focal management questions. For example, if a manage-343 ment goal only requires knowledge of species interactions within a community, the model could be simplified 344 to remove environmental covariates (Box 1). Alternatively, the global shrinkage parameter τ could be set at 345 a fixed value to induce more or less sparsity in the final model results. Such a change could allow users to 346 manually explore the trade-off between inclusion of species-specific terms and precision of parameter esti-347 mates, finding the balance that best suits their particular goals. For example, fixing τ to a higher value would 348 yield more estimates of species-specific parameters, which could help to inform future research priorities, 349 but those estimates would likely be less precise, limiting their utility in predicting community dynamics. 350 Small adjustments such as these empower ecologists and managers to match the tool to their questions and 351 aims. 352

In its current structure, the sparse modeling framework is most useful when applied to high-diversity 353 communities with limited available data. As we observed when analyzing simulated data, the number of 354 non-generic terms does not necessarily increase with sample size, and is limited by τ at higher sample sizes 355 (Figure 2d). As described above, there may be cases where manual adjustments to τ would be beneficial 356 depending on the available data and questions of interest. However, a traditional, non-sparse model in 357 which every interaction term is included may still be preferable in situations with abundant data, lower-358 diversity communities, or when answering questions requiring individual estimates of all potential species 350 interactions. In contrast, the sparse approach is particularly helpful with limited data and in cases where 360 traditional models often struggle to converge or provide overly broad parameter estimates. 361

The model we present here is currently analyzed for a single growing season and for use with traditional population dynamic models (e.g. Beverton-Holt or Lotka-Voltera models). Given the importance of temporal stochasticity to community dynamics (Shoemaker *et al.*, 2020) and the need to predict community responses to changing anthropogenic pressures (Ma *et al.*, 2017), sparse modeling with time series could provide invaluable insight into the importance of species-specific interactions through time as well as space. Further,
 extending the approach to a wider range of input data beyond individual counts (e.g. percent cover or
 biomass) would allow for future uses across observational datasets and especially for perennial-dominated
 systems.

Sparse modeling approaches have proved immensely valuable in fields as diverse as genomics (Gianola 370 & Fernando, 2020) to economics (Fan et al., 2011). By dramatically reducing the parameter load required 371 to model diverse communities across environmental gradients, we show these sparse modeling approaches 372 can provide both theoretical and applied insights in community ecology as well. We demonstrate the flex-373 ibility of this approach across different ecological models and underlying biological assumptions, and are 374 excited to see it expanded and applied to a variety of ecological questions and applications. Although the 375 implementation of the sparse method requires an initial conceptual investment, the output results are easily 376 interpretable—a quality that is particularly important for linking models to practice. The sparse modeling 377 approach eliminates the need for a priori assumptions regarding species' groupings or the exclusion of all 378 but a handful of focal species, providing a critical method and step forward in expanding ecological theory 379 and linking models to observational and experimental datasets of diverse communities. 380

381

6 Acknowledgements

This paper is a joint effort of the working group sToration kindly supported by sDiv, the Synthesis Centre of the German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig, funded by the German Research Foundation (FZT 118, 02548816). CWL, CMW, LGS were supported by Modelscapes, NSF award #EPS-2019528. OG was supported by the Spanish Ministry of Economy and Competitiveness (MINECO) and by the European Social Fund through the Ramón y Cajal Program (RYC-2017- 23666); GB was supported by the Swedish Research Council (Vetenskapsrådet), grant 2017-05245. MMM was supported by the Australian Research Council (DP140100574).

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Parameter			Distribution	Unique to
S		regional species pool	15 species	single value for community
<i>N</i> _{<i>i</i>,0}		initial population	Gaussian (mean 80, sd 50) truncated at 0	species, plot
X _e		plot environment	Gaussian (mean 0, sd 1)	plot
$\lambda_{e,i}$		intrinsic growth with mono- tonic environmental response	$e^{b_{0,i}+b_{e,i}X_e}$	species
	$b_{0,i}$	environmentally-independent growth rate	uniform (0 to 1.5)	species
	$b_{e,i}$	environmental response in λ	Gaussian (mean 0, sd 0.5)	species, plot
$\alpha_{e,i,j}$		competitive interactions with environmental effects	$e^{\overline{a}_{0,i}+\hat{a}_{0,i,j}+\left(\overline{a}_{e,i}+\hat{a}_{e,i,j} ight)X_e}$	species pair, plot
	$\overline{a}_{0,i}$	generic interspecific competi- tion	Gaussian (mean -7, sd 0.1)	species pair
	$\hat{a}_{0,i,j}$	deviation in competition	uniform $\pm(1.5 \text{ to } 3.2)$	7 species j
	$\overline{a}_{0,i,i}$	intraspecific competition	-3.5	constant
	$\overline{a}_{e,i}$	generic environmental varia- tion in competition	Gaussian (mean 0, sd 0.3)	single value for community
	$\hat{a}_{e,i,j}$	specific environmental varia- tion in competition	uniform $\pm (0.5 \text{ to } 1.5)$	7 species j

Table 1: Parameter components and distributions used in simulations

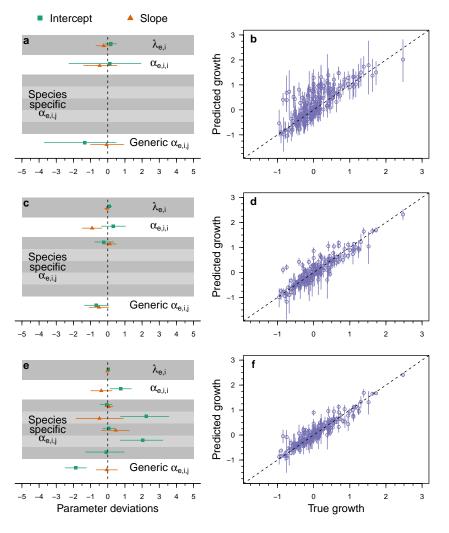


Figure 1: Fitted model parameter estimates and predicted growth rates. We fit the model to simulated data with 10 (a and b), 50 (c and d), and 200 (e and f) full-community and no-competition plots. The left column (a, c, and e) shows the deviation of parameter values from the true value used in the simulations (points are posterior means and lines are 95% credible intervals). The right column (b, d, and f) shows model accuracy of the focal species' growth rate for 200 simulated full-community plots not included in the model fitting. Growth rates were calculated as $ln(\frac{N_{t+1}}{N_t})$. The dashed line is the 1-1 line indicating a perfect match. Points show mean estimates and lines are 95% CIs.

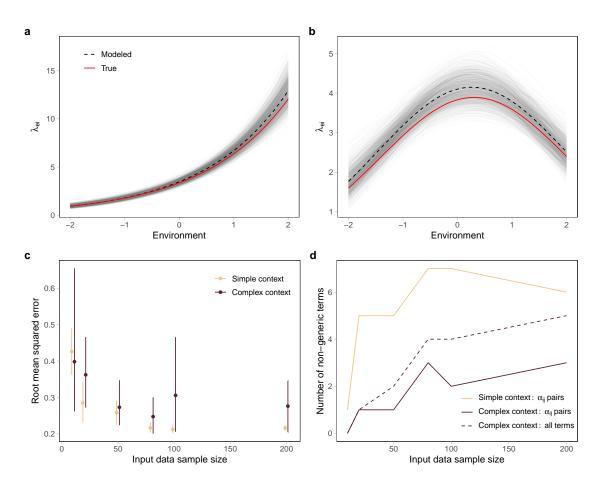


Figure 2: Estimates for $\lambda_{e,i}$ for simulated data with (A) a monotonic response to the environment and (B) an environmental optimum, with the true values as a solid red line, model means as a dashed black line, and individual model posterior draws as thin grey lines. Both models were run with 50 full community plots and 50 no-competition plots. All growth rate parameters fell within the 95% credible intervals for parameters in both models. (C) Simulated contexts with only $\alpha_{i,j}$ species pair intercepts compared to contexts with $\alpha_{e,i,j}$ species pair by environment slopes. Performance of sparse models matching these simulation contexts was measured as RMSE between the model predictions and the true value of 200 out-of-sample data points. Both models predict true growth rates similarly well at low sample sizes (10 samples) but the model of the simpler simulation converges closer to the true values. (D) Number of non-generic terms identified by models on the different α contexts, which is constrained at larger sample sizes by the global shrinkage parameter τ . The model of the species pair intercept only simulation identifies more non-generic $\alpha_{i,j}$ pairs at all sample sizes. The model of the simulation with both non-generic species pair $\alpha_{0,i,j}$ intercepts and species pair by environment $\alpha_{e,i,j}$ slopes requires more data to identify non-generic terms, and those are split between the intercept and slope terms.

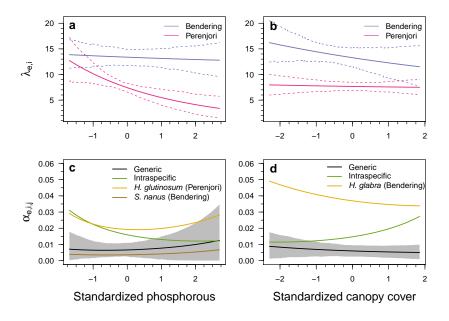


Figure 3: Model estimates for *W. acuminata*. Means (solid lines) and 95% CIs (dashed lines) are shown for $\lambda_{e,i}$ across a gradient of phosphorous (a) and canopy cover (b). Colors indicate the Bendering and Perenjori reserves. The mean (black line) and 95% CI for generic interspecific competition are shown across a phosphorous (c) and canopy cover (d) gradient. In both c and d, the mean intraspecific competition coefficient is shown in green and different species identified by the model as non-generic in each reserve are shown with other colors as indicated in the legends.

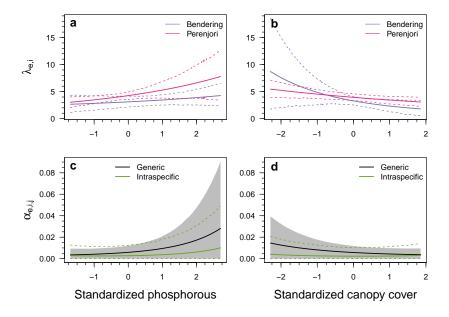


Figure 4: Model estimates for *A. calendula*. Means (solid lines) and 95% CIs (dashed lines) are shown for $\lambda_{e,i}$ across a gradient of phosphorous (a) and canopy cover (b). Colors indicate the Bendering and Perenjori reserves. The mean (black line) and 95% CI for generic interspecific competition are shown across a phosphorous (c) and canopy cover (d) gradient. In both c and d, the mean intraspecific competition coefficient is shown in green with dashed lines indicating the CI. The model did not identify any interspecific competition coefficient.

Box 1: Adapting the sparse modeling method to different ecological questions

This sparse modeling method is generalizable to a variety of underlying ecological models. The method's flexibility allows researchers to pick and choose which parameters to include and how to specify them as best fits with their study system and questions of interest.

For example, the relationship between species' growth rates and the environment can be modeled in multiple ways. A monotonic relationship would be appropriate for a study concentrated within a small spatial scale, while a humped-shape relationship would match expectations for a study over a broad environmental gradient. To demonstrate how our method can be modified for different underlying ecological models, we simulated environmental responses in growth rate two ways: with a monotonic relationship between species and the environmental conditions $\lambda_{e,i}$ and with a curved environmental optimum with a defined niche breadth for each species $\lambda_{e,i}^*$. This resulted in two model formulations:

$$\lambda_{e,i} = e^{b_{0,i} + b_{e,i} X_e} \tag{6}$$

in which $b_{0,i}$ is the mean intrinsic growth rate and $b_{e,i}$ is the slope of the environmental response, and

$$\lambda_{e,i}^* = b_{max,i} e^{-\left(\frac{z_i - X_e}{2\sigma_i}\right)^2} \tag{7}$$

⁵³⁷ in which $b_{max,i}$ is the maximum intrinsic growth rate, z_i is the environmental optimum, and σ_i is the environ-⁵³⁸ mental niche breadth (following the parameterization in Thompson *et al.* (2020)). We tested these models ⁵³⁹ using samples of 50 full community plots and 50 no-competition plots.

All growth rate parameters fell within the 95% credible intervals for parameters in both models. In the monotonic $\lambda_{e,i}$ model, both the intercept $b_{0,i}$ and the slope $b_{e,i}$ deviated from the true values by 3%. In the optimum $\lambda_{e,i}^*$ model, the maximum $b_{max,i}$ deviated from the true value by 6%, the niche breadth σ_i deviated by 1%, and the location of the environmental optimum z_i deviated by 13%, which is an absolute difference of 0.04 (Fig. 2a and b).

545 As a further example, the species interaction components of the model can be adjusted depending on the

main research questions of interest. For questions focused on species interactions, modeling interaction co-546 efficients independent of environmental conditions would optimize the number of non-generic species pairs 547 identified from a given sample size of data. We compared a simple simulation with competitive interac-548 tions independent of environmental conditions $\alpha_{i,i}$ to a more complex model with competitive interactions 549 dependent on the environmental conditions $\alpha_{e,i,j}$, and modeled each accordingly. We tested these models 550 using different sample sizes of full community data points and focal individuals. Both models predicted true 551 out-of-sample growth rates with average RMSE of ≈ 0.4 with only 10 full-community and no-competition 552 plots. With 80 or more full-community and no-competition plots the simpler model had a RMSE of ≈ 0.2 553 and the model of the more complex simulation had a RMSE of ≈ 0.25 (Figure 2c). The simpler model with-554 out species-environment interactions highlighted five non-generic $\hat{a}_{0,i,i}$ species pair interactions with just 50 555 full-community and no-competition plots. The more complex model highlighted only two non-generic terms 556 with 50 full-community and no-competition plots. At higher sample sizes the number of non-generic terms 557 was constrained by the global shrinkage parameter τ (Figure 2d) 558

We present these options as launching-off points for researchers to adapt the sparse modeling approach to their study systems and questions. Even more extensive modifications are possible; for example, replacing the Beverton-Holt community framework with a different underlying ecological model.