Untangling Microbiota Diversity and Assembly Patterns in the World's Largest Water Diversion Canal

Lu Zhang[1,2], Wei Yin[3], Chao Wang[3], Aijing Zhang[4], Hong Zhang[5], Tong Zhang[6] and Feng Ju[1,2*]

[1] Key Laboratory of Coastal Environment and Resources of Zhejiang Province, School of Engineering, Westlake University, 18 Shilongshan Road, Hangzhou 310024, Zhejiang Province, China

[2] Institute of Advanced Technology, Westlake Institute for Advanced Study, 18 Shilongshan Road, Hangzhou 310024, Zhejiang Province, China

[3] Changjiang Water Resources Protection Institute, 515 Qintai Street, Wuhan 430051, Hubei Province, China

[4] Construction and Administration Bureau of South-to-North Water Diversion Middle Route Project, 1 Yuyuantan South Road, Beijing 100038, China

[5] State Key Laboratory of Environmental Aquatic Chemistry, Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences, 18 Shuangqing Road, Beijing 100085, China

[6] Environmental Microbiome Engineering and Biotechnology Laboratory, Department of Civil Engineering, Pokfulam Road, The University of Hong Kong, Hong Kong 999077, China

*** Corresponding to:**

Dr. Feng Ju (Assistant Professor)

Address:

Environmental Microbiome and Biotechnology Lab (EMBLab)

Westlake University, 18 Shilongshan Road, Hangzhou 310024, China

Tel.: 571-87963205 (lab), 571-87380995 (office)

Fax: 0571-85271986

E-mail: jufeng@westlake.edu.cn

Abstract

Large water diversion projects are important constructions for reallocation of human-essential water resources. Deciphering microbiota dynamics and assembly mechanisms underlying canal water ecosystem services especially during long-distance diversion is the prerequisite for water quality monitoring, biohazard warning and sustainable management. Using a 1432-km canal of the South-to-North Water Diversion Projects as a model system, we answer three central questions: how bacterial and micro-eukaryotic communities spatio-temporally develop, how much ecological stochasticity contributes to microbiota assembly, and which immigrating populations better survive and navigate across the canal. We applied quantitative ribosomal RNA gene sequence analyses to investigate canal water microbial communities sampled over a year, as well as null model- and neutral model-based approaches to disentangle the microbiota assembly processes. Our results showed clear microbiota dynamics in community composition driven by seasonality more than geographic location, and seasonally dependent influence of environmental parameters. Overall, bacterial community was largely shaped by deterministic processes, whereas stochasticity dominated micro-eukaryotic community assembly. We defined a local growth factor (LGF) and demonstrated its innovative use to quantitatively infer microbial proliferation, unraveling taxonomically dependent population response to local environmental selection across canal sections. Using LGF as a quantitative indicator of immigrating capacities, we also found that most micro-eukaryotic populations (82%) from the source lake water sustained growth in the canal and better acclimated to the hydrodynamical water environment than bacteria (67%).

Taxa inferred to largely propagate include *Limnohabitans* sp. and *Cryptophyceae*, potentially contributing to water auto-purification. Combined, our work poses first and unique insights into the microbiota assembly patterns and dynamics in the world's largest water diversion canal, providing important ecological knowledge for long-term sustainable water quality maintenance in such a giant engineered system.

Key words: Engineered system; Bacterial community; Micro-eukaryote; Deterministic assembly; Stochastic assembly; Population growth

Introduction

Microorganisms are fundamental components of ecosystems and play vital roles in global biogeochemical cycling (Arrigo, 2005; Battin et al., 2016; Du et al., 2021). Knowledge about composition and dynamics of microbial communities across space and time is a prerequisite for predicting and manipulating the microorganisms in both natural and engineered systems (Bell et al., 2005; Bru et al., 2011; Ju and Zhang, 2015). Due to artificial system design and control, microbial communities in engineered ecosystems have received increasing attention to unravel the community dynamics underlying their eco-environmental or biotechnological services (Briones and Raskin, 2003; Ju et al., 2017; Ju et al., 2014; Vanwonterghem et al., 2014; Wu et al., 2019). Specifically, microorganisms in drinking water systems are closely linked with water auto-purification and eventually to public health (Berry et al., 2006). Thus, comprehensive microbiome monitoring is needed to differentiate baseline and disturbed microbial communities in engineered water systems to facilitate system management (Hull et al., 2019; Sagova-Mareckova et al., 2021).

The South-to-North Water Diversion (SNWD) Projects were built to aid in water supply in the drought-stricken north China, which represent the world's largest water diversion projects. The projects consist of the Eastern Route Project, the Middle Route Project (MRP) and the Western Route Project. The 1432-km MRP went into service in 2014 and has drawn 40 billion m^3 of water from the Danjiangkou Reservoir to He'nan and Hebei Provinces, and Beijing and Tianjin Municipalities, benefiting tens of millions of people. Despite their

important roles in controlling geochemical processes and indicating water quality (Chen et al., 2018; Dang et al., 2018), little has been known about the microbial constituents throughout the system (Luo et al., 2019), neither for other engineered long-distance water diversion canals. Especially, the MRP is a closed engineered system where microorganisms are subject to potentially less complex biotic and abiotic interactions, while the numerous hydraulic structures and engineered perturbations (e.g., inverted siphon, aqueduct and tunnels) along the main canal add to challenges to model microbial community dynamics, calling for comprehensive investigation of microbial community patterns in the MRP canal, to assist in establishing integrative assessment of water quality with microbial community monitoring (Sagova-Mareckova et al., 2021).

Understanding processes and mechanisms driving the community patterns underlying ecosystem services is one of the key issues in microbial ecology (Zhou et al., 2014). Microbial community composition in aquatic ecosystems can be affected deterministically by both abiotic or environmental factors, e.g., water temperature, pH, dissolved oxygen, and nutrient availability (Hu et al., 2017; Liu et al., 2020a; Zhang et al., 2019b), and biotic factors, such as competition and predation. In addition to deterministic mechanisms, stochastic processes, for instance, dispersal and ecological drift, also contribute to microbial community assembly. Efforts have been made to disentangle the relative importance of deterministic vs. stochastic processes based on a variety of null and neutral modeling methods (Ning et al., 2019; Sloan et al., 2006; Stegen et al., 2013). Depending on which ecosystems or organism types these methods are applied to, the relative importance of deterministic vs. stochastic

processes may greatly vary (Chen et al., 2019; Liu et al., 2018b; Wang et al., 2020; Yan et al., 2020; Niederdorfer et al., 2021). For instance, bacterial communities in lake water were revealed to be predominantly structured by determinism, whereas micro-eukaryotic communities largely by stochasticity (Logares et al., 2018). For the MRP, the closed channel, the environmental gradients and the variety of hydraulic constructions along the canal make it obscure to what extent ecological stochasticity determines the assembly of microbiota community, which is essential for an improved understanding of the ecosystem processes and microbial dynamics prediction.

Other emerging questions in such large water-diversion canal and the comparable systems, like river continuum (Gweon et al., 2021), include which immigrating microorganisms are adapted to local niches in the receiving water and how they functionally contribute to the ecological service. Especially in the MRP, how many of the microbial populations originated from a natural reservoir may survive in the contrastingly engineered canal remains an open question. Although population growth associated with microbial immigration has been quantified in other engineered water systems (Mei and Liu, 2019), such as activated sludge reactors (Mei et al., 2019), efforts in estimating microbial growth in long-distance and evolving systems are still lacking.

In the present study, we followed the spatio-temporal dynamics of prokaryotic and micro-eukaryotic community along the main canal of MRP of the SNWD project over a year to address: 1) How does microbial community develop with the water flow? 2) How is the

relative importance of deterministic and stochastic processes on shaping the community assembly in this closed turbulent engineered system? 3) How do the immigrating bacteria and micro-eukaryotes from water source survive, grow, or die off in the canal? Based on high-throughput marker gene (i.e., 16S and 18S) amplicon and metagenomic sequencing, bioinformatics, and statistical modeling of water microbiota, we elucidated the microbial diversity and population dynamics along the canal over four seasons and uncovered the key patterns and processes driving community assembly. By overcoming the limitations of widely used relative abundance measures with estimated absolute abundance (Barlow et al., 2020; Tkacz et al., 2018), we demonstrated the first use of local growth factor to predict growth capacity of the immigrating microbial populations along the water canal and revealed the divergent microbial fates and population lifestyles throughout the world's largest water diversion canal.

Materials and Methods

A full version of the Materials and methods are available in the Supplementary Information.

Study sites, sampling and physiochemical monitoring

The Middle Route Project (MRP) of the South-to-North Water Diversion (SNWD) Projects started from Danjiangkou reservoir and diverted water northwards until Beijing and Tianjin, which is an artificial system without direct connection with surface waters in the area. Fifty samples were collected from 19 sites (Fig. 1), including one at the Danjiangkou reservoir (S01) and 18 along the main canal of the MRP (S02-S18). The 18 sampling sites along the

canal were selected to capture the spatial dynamics of microbial communities from canal head to end across the subtropical and warm-temperate climate zones. The sites were grouped to three sections: S02-S08 (Section1), S09p-S14 (Section2) and S15-S18 (Section3). Canal water was sampled and filtered through 0.22-μm cellulose nitrate filters (Sartorius, Germany). Physiochemical properties, including water temperature (T), pH, dissolved oxygen (DO), total nitrogen (TN) and fluoride (F^-), were measured according to the environmental quality standard for surface water of China (GB3838-2002). Details about the sampling sites and physiochemical measurements are in the Supplementary Method S1.

DNA extraction, PCR amplification and sequencing

Genomic DNA was extracted from filters using FastDNA Spin Kit for Soil (MP Medicals, USA). Bacterial 16S rRNA gene V3-V4 region and eukaryotic 18S rRNA gene V4 region were amplified and sequenced on the Illumina platforms producing 250 bp paired-end reads at the Guangdong Magigene Biotechnology Co., Ltd. (Guangzhou, China). Real-time quantitative PCR (qPCR) was additionally performed to quantify the 16S rRNA gene copies (as a proxy for bacterial biomass) in each genomic DNA sample, as described in Ju et al. (2019). Detailed methodologies are provided in the Supplementary Method S2.

Each genomic DNA was also used for shot-gun metagenomic sequencing on the Illumina's Novaseq platform using a 150 bp paired-end sequencing strategy at the Novogene (Beijing, China). The sequencing data generated in this study have been deposited into CNGB Sequence Archive (CNSA) (Guo et al., 2020) of China National GeneBank DataBase

(CNGBdb) (Chen et al., 2020) with accession number CNP0001593.

Bioinformatics and statistical analyses of microbial diversity

The 16S and 18S rRNA gene amplicon sequence data were processed using Qiime2 pipeline (Bolyen et al., 2019) and DADA2 algorithm (Callahan et al., 2016) as described in the Supplementary Method S3, generating final abundance tables of amplicon sequence variants (ASVs) consisting of 6705 bacterial and 3217 eukaryotic ASVs, respectively. The alpha-diversity metrics of bacterial and micro-eukaryotic communities were further computed in Qiime2. All the univariate and multivariate statistics were performed in R (R Core Team, 2020). A set of spatial factors was generated for the sampling sites based on the geographic coordinates using the principal coordinates of neighboring matrices (PCNM) analysis (Borcard and Legendre, 2002).

The metagenomic data were pretreated and searched against the SILVA SSU132 database to identify rRNA gene reads. This information was further used to estimate the 16S rRNA and 18S rRNA gene copies detected in each metagenome based on the sum for the read coverage over the full length of each matched reference sequence in the database. Equation and details are available in the Supplementary Method S3. The copies of 18S rRNA genes in each water sample were estimated as 16S rRNA gene copies (determined by the aforementioned qPCR analysis) divided by ratios of 16S rRNA to 18S rRNA gene copy (Table S1) as sequenced and estimated in the metagenomic data.

Neutral and null model analyses of microbial community patterns

To disentangle the relative contribution of stochastic and deterministic processes to microbial community assembly, Sloan neutral model (Sloan et al., 2006) and null model-based NST approach (Ning et al., 2019) were applied to the 16S and 18S community data. Fitting of the model were performed in R according to Burns et al. (2016). Accordingly, ASVs were classified into three groups depending on whether their observed occurrence falling higher than ("above prediction"), within ("as predicted") or lower than ("below prediction") the 95% confidence interval of the neutral model predictions. Normalized stochasticity ratio (NST) was used to quantify ecological stochasticity in communities within canal sections (Ning et al., 2019). NST analysis was performed in R using the NST package (Ning et al., 2019). More details about the methodology are in the Supplementary Method S4.

Population lifestyle and local growth factor analyses

The absolute abundance of bacterial and micro-eukaryotic ASVs was calculated by multiplying relative abundance of each ASV computed from amplicon sequence data by the overall 16S and 18S rRNA gene copies detected in a given metagenome for all the Oct samples (see the Supplementary Method S5 for more details).

To characterize fate of microorganisms originated from natural reservoir after transported into the closed canal, we defined a local growth factor (LGF) to depict whether the populations grew or restrained, by comparing their mean absolute abundance in each canal

section to the abundance at the canal head, as follows:

$$\mathrm{LGF_x} = \log_{10} \frac{\mathrm{AA_{x,N}} + 1}{\mathrm{AA_{x,OctS02}} + 1} \quad (1)$$

where $AA_{x,\,N}$ is the average absolute abundance of ASV x in Section N of sampling sites and $AA_{x,\,OctS02}$ is the absolute abundance of ASV x at OctS02, the canal head. OctS02 was excluded from Section1 for the LGF calculation. An average LGF > 0 across the three sections was considered to indicate growth in the canal. The potentially pathogenic bacterial ASVs were identified referring to a list of potentially pathogenic genera (Table S2) summarized from Woolhouse et al. (2015), and LGFs of these ASVs were examined.

Results

Microbiota biomass and diversity shifted with seasonality

Microbial communities showed clear seasonal shifts (Fig. 2), whether for their biomass (A and B) or alpha (C-F) and beta diversity (Fig. 2G, 2H and Fig. S1). Both the bacterial and micro-eukaryotic biomass, as measured by 16S rRNA (A) and 18S rRNA (B) gene copies, were the highest in August and the lowest in October (Fig. 2). PERMANOVA showed that seasonality significantly (P < 0.001) influenced both the bacterial (F = 22.78) and micro-eukaryotic (F = 25.57) community composition. Moreover, the seasonal impacts were much more pronounced than the spatial effects along the canal, whether for bacteria (R^2 = 0.52 vs. 0.14) or micro-eukaryotes (R^2 = 0.55 vs. 0.14). Accordingly, Bray Curtis-based NMDS analyses of all canal water samples (n = 52) revealed strong clustering of bacterial

(Fig. 2G) and micro-eukaryotic (Fig. 2H) communities in relation to seasons, of which May and August had witnessed greater intra-season community variations along the canal. Seasonal variation was also observed for specific bacterial and micro-eukaryotic lineages (Fig. S2 and S3).

Microbiota composition and diversity changed along the canal

Both the source water from Danjiangkou reservoir (S01) and the canal water were dominated by phyla *Actinobacteria* (39.7 - 65.0%, mainly hgcI clade and CL500-29 marine group), *Proteobacteria* (17.9 - 38.3%), *Bacteroidetes* (0.9 - 29.5%), *Verrucomicrobia* (1.2 - 9.2%) and *Cyanobacteria* (0.2 - 8.6%, Fig. 3A). Overall, bacterial communities at the canal head (S02) were most structurally similar to the source water (Fig. 3A and Fig. S1A). However, compared with the source water, relative abundance of *Proteobacteria* at the canal head (i.e., S02) decreased whereas *Actinobacteria* and *Bacteroidetes* increased. Within the canal, both *Cyanobacteria* (mostly *Cyanobium*_PCC-6307, $F = 16.77$, $P < 0.001$) and *Bacteroidetes* ($F = 6.06$, $P < 0.05$) showed significant decrease in relative abundance with water flow (Fig. 3A). In contrast, *Limnohabitans* ($F = 19.87$) and *Candidatus* Methylopumilus ($F = 58.69$) within *Proteobacteria* became increasingly abundant along the canal ($P < 0.001$). Likewise, both the overall bacterial biomass (Fig. S4A) and Chao1 richness (Fig. S5A) exhibited highly dynamic variation patterns along the canal. The variation trend of Shannon diversity index largely agreed with (though relatively weaker than) that of Chao1 richness (Fig. S5B).

Unlike bacterial communities, micro-eukaryotic communities appeared more dynamic

along the canal water flow (Fig. 3B). The relative abundance of *Metazoa*, mostly *Calanoida*, was as high as 24.0% in Danjiangkou Reservoir. These organisms were drastically enriched by 2.3 times upon entering the canal head site, but largely declined in the downstream sites (S03 to S18). Two predominant lineages of *Cryptophyceae*, i.e., *Cryptomonas* (F = 4.11, P = 0.06) and *Teleaulax* (F = 9.50, P < 0.01), were more abundant in the source water and canal upstream (S02-S08) than downstream sites (S09p-S14), accompanied by their gradual replacement by two uncultured *Cryptophyceae* lineages (F = 30.53, P < 0.001; F = 24.60, P < 0.001, respectively), especially close to the canal end (S15-S18). The canal head communities were distinct from water source and the downstream communities (Fig. 3B and Fig. S1B). This special pattern was manifested by the dominance of *Calanoida* (72.0%) at the canal head site where micro-eukaryotic biomass was much higher (3.2×10^9 vs. average 5.9×10^8 18S rRNA gene copies L^{-1}, Fig. S4B) whereas alpha diversity metrics much lower compared with the other sites (Fig. S6), suggesting intriguing local micro-eukaryotic selection in the canal head.

Environmental factors influenced canal microbiota diversity and biomass

The influence of five environmental parameters, namely water temperature, pH, dissolved oxygen (DO), total nitrogen (TN) and fluoride (Table S3), and spatial factors (i.e., ten PCNM variables representing the quantitative spatial patterns, Table S4) on the microbial communities was tested by redundancy analysis. All the above-mentioned environmental variables and PCNM1, 2 and 4 were identified to exert significant impacts on bacterial

community composition ($P < 0.05$, Fig. 4A). These variables except for PCNM4 also represented the significant influential factors for micro-eukaryotic communities ($P < 0.05$; Fig. 4B). Seasonal variation of environmental factors appeared based on the RDA plots (Fig. 4A and 4B), for instance, fluoride concentration was especially higher in March, which was supported by the significant seasonal difference in the environmental variables by Kruskal-Wallis one-way ANOVA (Table S5). The abundant actinobacterial hgcI clade (r_s = 0.63, $P < 0.01$) and CL500-29 (r_s = -0.68, $P < 0.01$) marine group were among the bacterial lineages positively and negatively related to fluoride concentration, respectively (Table S6). For micro-eukaryotes, for instance, an ASV affiliated to *Mediophyceae* (r_s = 0.66, $P < 0.01$) was positively correlated with fluoride (Table S7).

The identified significant variables were employed in further variation partitioning analysis (VPA). The result showed clearly stronger influence of seasonality than that of spatial and examined environmental factors for both bacterial and eukaryotic communities (Figure 4C and 4D). Seasonality alone could explain 19% and 17% of the bacterial and micro-eukaryotic community variance, respectively, while only 6 - 9% of the variance could be explained exclusively by environmental or spatial factors. Notably high proportions of the community variance could be attributed to the co-effects of seasonality and environmental factors (29% for bacteria and 23% for micro-eukaryotes), implying their strong interplay in shaping microbial communities. Besides, temperature, DO and TN were found to be significantly correlated with bacterial biomass, while only DO for micro-eukaryotic biomass (as measured by 16S rRNA and 18S rRNA gene copies, Table S8).

Bacterial and micro-eukaryotic communities exhibited distinct and spatio-temporally dynamic stochasticity

The apparent dispersal of microorganisms in the canal water and engineered hydraulic manipulations (i.e., inverted siphon, aqueduct, and tunnels) along the main canal drove us to question on the relative contribution of stochastic vs. deterministic processes to the canal bacterial and micro-eukaryotic community assembly, which was first quantitatively assessed with a recently proposed metric, normalized stochasticity ratio (NST) (Ning et al., 2019). This new metric quantifies the relative importance of stochasticity in governing community assembly ranging from 0 to 1, with a value > 0.5 indicating more stochastic assembly and < 0.5 more deterministic assembly. NST analysis revealed contrasting trends between bacteria and micro-eukaryotes. In the upstream section, a highest NST of 0.78 was estimated for bacteria (Fig. 5A). This value decreased to 0.46 in the middle section, and to as low as 0.32 close to the canal end. On the contrary, NST was generally above 0.5 for micro-eukaryotes throughout the canal, with the highest value of 0.75 in the middle section (Fig. 5A).

While NST provides rigorous quantification of relative importance of stochasticity, the Sloan neutral model analysis (Sloan et al., 2006) allows for assessing how well the observed community patterns could be explained by neutral processes (e.g., birth, death, and immigration) and attributing the observed community-level assembly pattern to the divergent population behavior. According to seasonal comparison of the model fitness, neutral model could explain much higher proportions of community patterns in October (R^2 = 0.73 and 0.67

for bacteria and micro-eukaryotes, respectively) and March (R^2 = 0.77 and 0.72), but lowest in May (R^2 = 0.42 and 0.13, Fig. S7).

To link community patterns to the underlying population dynamics, bacterial and micro-eukaryotic ASVs were classified into three groups based on the neutral model analysis of the October samples. Depending on whether they empirically occurred more frequently than, as frequently as or less than predicted by the neutral model, ASVs were assigned to "above prediction", "as predicted" and "below prediction" groups that were different in abundance and composition (Fig. 5B-D). To fully capture the contribution of each group to the overall community, we estimated absolute abundance of ASVs by multiplying relative abundance of ASVs at each canal site with the estimated total copies of bacterial 16S rRNA (7.89×10^8 to 2.93×10^9 copies L^{-1}) or micro-eukaryotic 18S rRNA (3.62×10^8 to 3.19×10^9 copies L^{-1}) genes therein.

With the assistance of absolute abundance of ASVs, we found that 7.8% of the bacterial community richness (i.e., number of ASVs) accounted by above-prediction group corresponded to 65.5% in the total abundance (as 16S RNA gene copies), whereas the neutrally distributed group contributed 86.8% of the richness but only 24.9% of the abundance (Fig. 5B). Similar trend was detected for micro-eukaryotes (Fig. 5B). The three partitioning groups of ASVs also showed distinct taxonomic composition (Fig. 5C and 5D). *Cryptomonas* constituted 15.1% of the above-prediction group, but they were absent in the neutral and below-prediction groups. *Metazoa*, in contrast, accounted for only 0.4% of the

above-prediction group, but as high as 64.7% and 87.0% of the abundance of the neutral and below-prediction groups (Fig. 5D). Richness-based comparison produced a clearly distinct pattern for micro-eukaryotes (Fig. 5D). When perceiving at the taxon level, the relative contribution of the three groups to richness and abundance were largely taxonomically dependent (Fig. S8). Intriguingly, micro-eukaryotes showed higher overall occurrence (0.313±0.008 vs. 0.216±0.007; $P < 0.001$) while lower variability (2.879±0.033 vs. 3.055±0.032; $P < 0.001$) compared to bacteria.

Selective growth of immigrating bacterial and micro-eukaryotic populations along the canal

To resolve which populations of the water communities died off, sustained, or proliferated during dispersal over 1432-km canal, we proposed local growth factor (LGF) (see equation 1 in Methods) as a quantitative measure of variation in population abundance in each canal section compared to the abundance at the canal head, with an average LGF > 0 across the three sections indicating growth in the canal. LGF analysis was applied with the Oct sample dataset composed of 1389 bacterial and 1229 eukaryotic ASVs. In total, 67.1% of the bacterial ASVs (Table S9) and 82.3% of the micro-eukaryotic ASVs (Table S10) displayed positive average LGFs. The ASVs were further found to form clearly three clusters based on their LGFs in each section: LGF > 3, -3 < LGF < 3 and LGF < -3, which could indicate substantial growth, survival, and die-off of the microbial populations (Fig. 6). The survival group represented the largest proportion of the ASVs (45.6 - 60.2%), while only 4.1 - 21.7%

of the ASVs went extinction in the canal.

We also found that both the contrastingly high and low LGFs tended to be attributed to high-abundance microbial lineages (Fig. 6A and B). For example, the abundant *Luteolibacter* sp. (ASV106) and *Limnohabitans* sp. (ASV27) exhibited the highest LGF values in Section 2 and Section 3, respectively (Table S9). In stark contrast, ASV138 assigned to actinobacterial CL500-29 marine group and two *Cyanobium* PCC-6307 lineages (ASV144 and ASV250) were especially restrained in growth close to the canal end (Table S9). A total of 53 bacterial ASVs in the canal were related to 16 genera included in a published reference pathogen list (Table S2, summarized from Woolhouse et al., 2015). While LGF of most (39 out of 53) of these potentially pathogenic ASVs fell between -3 and 3, three *Acinetobacter*-associated ASVs (ASV794, ASV1108 and ASV 1261) showed LGFs ranging from -5.9 to -6.1, indicating notable depletion along the canal.

For micro-eukaryotes, two uncultured *Cryptophyceae* lineages (ASV54 and ASV80) as well as two ASVs related to *Acineta flava* (ASV224) and *Spumella* sp. (ASV59) were elevated throughout the canal (Table S10). We also compared the LGFs between the three groups of ASVs afore-defined by neutral model analysis, but no significant difference was detected (Fig. S9).

Discussion

Diversion canals widely built for reallocation of water resources represent an ideal engineered

system to model microbial communities fulfilling key eco-environmental services, as well as reflecting changes in ecosystem structure and function. Based on quantitative metagenomic approaches that enable absolute quantification of microbial taxon, we uncovered the microbiota spatio-temporal dynamics, key processes driving community assembly, and navigation of immigrating microorganisms through the world's largest water diversion project.

Microbiota composition and biomass are dynamic along the canal

Both bacterial and micro-eukaryotic communities in the source water showed marked variations in taxonomic composition upon entering the canal, which could be attributed to the readily comprehensible differences in the environmental conditions between the natural reservoir and its receiving engineered canal and potential influences exerted by the approach channel through which source water enters the canal. Meanwhile, we observed intriguingly decreased water microbial diversity after water transportation through an inverted siphon (S09 vs. S09p, Fig. S5 and S6). Taken together, these suggest that hydraulic structures along the canal apply considerable influence on the microbial community diversity and composition, which needs to be fully considered when setting baseline communities for long-term microbial monitoring in the water diversion canal.

The prevalence of uncultured actinobacterial hgcI clade and CL500-29 marine group echoed what was previously reported in other freshwater habitats (Keshri et al., 2018; Yang et al., 2017). While both groups of yet-to-be-cultured lineages have been shown to favor lower

level of dissolved organic carbon and temperature (Lindh et al., 2015), the hgcI clade was additionally implicated to be competitively advantageous in utilizing low-concentration organic carbon (Glöckner et al., 2000) and to harbor potentially photoheterotrophic species showing preferential utilization of nitrogen-rich organic compounds in freshwater (Ghylin et al., 2014), implying its potential role in the nutrients uptake and water auto-purification.

Unlike bacterial communities, micro-eukaryotic communities in the canal have not yet been explored before. By virtue of the high-throughput molecular analyses, we detected not only *Rotifea*, copepods and protozoa previously microscopically observed as abundant in the Danjiangkou Reservoir (Li et al., 2012), but also thousands of micro-eukaryotic populations (as ASVs), of which 6.7% are uncultured or unclassified even at the class level, revealing detection of numerous novel populations in the canal water. Notably, metazoan *Calanoida* was repeatedly found to show drastic increase in abundance in the micro-eukaryotic communities of canal head, compared with in the source water in both October and August. This big order of aquatic crustaceans belonging to *Copepoda* inhabiting marine and freshwater environments are found as the most successful among zooplankton colonizing pelagic habitats (Blanco-Bercial et al., 2011; Bradford-Grieve, 2002). We further noted that the SAR clade and *Cryptophyceae* showed substantially decreased relative abundance as a result of high enrichment of *Calanoida* compared with the source water. However, their absolute abundance or biomass, as we estimated as their 16S rRNA gene copies, remained comparable (2.78 vs. 3.43×10^8 16S-copies L^{-1} for the SAR clade, and 3.68 vs. 4.25×10^8 16S-copies L^{-1} for *Cryptophyceae*). This strong decoupling pattern between relative and

absolute abundance cannot have been resolved by currently popular sequence-based relative measures alone which have been shown to provide evidently misleading information on the variation of absolute abundance (Stämmler et al., 2016; Vieira-Silva et al., 2019). It also well demonstrates the power of our quantitative metagenomics approaches as an alternative to a more direct but difficult-to-achieve quantitative PCR assay approach (due to the lack of 18S rRNA gene universal primers) for estimating total micro-eukaryotic biomass.

Microbiota assembly is deterministically driven by seasonally dependent habitat filtering effects

Elucidating mechanisms and factors shaping biogeographic patterns has become a central goal in microbial ecology (Hanson et al., 2012). Environment exerts influence on community composition by selecting for populations that better adapted to the corresponding ecological niches. We found that temperature, pH, dissolved oxygen, total nitrogen and fluoride all showed significant effects on the community dynamics in the main canal. Fluoride affects bacterial growth and metabolic activity by reducing acid tolerance of bacteria through targeting enzyme inhibition (Liao et al., 2015). Fluoride has been reported to alter microbial community in high-fluoride groundwater (Zhang et al., 2019a) and to exhibit inhibitory effect on bacterial populations involved in various wastewater treatment processes (Ochoa-Herrera et al., 2009). Our findings showed that fluoride with concentration falling within the range of 0.14-0.22 mg L^{-1} (Table S3) could also affect microbial community structure in a drinking water diversion system, where fluoride could be mainly contributed from natural processes,

such as mineral weathering and karstification (Nong et al., 2019). Interestingly, we found positive response of the hgcI clade to elevated fluoride concentration. While the mechanisms of their fluoride resistance still await exploration, such evidence is in line with their favorable distribution in a wide range of water ecosystems.

Although bacterial and micro-eukaryotic communities are both significantly influenced by the examined environmental variables and spatial segregation, their overall variation were explained more by seasonality. Meanwhile, the strong co-influence of temporal and environmental factors revealed by variation partitioning analysis could be at least partially explained by seasonal variation of the examined variables (Table S5), suggesting season-driven habitat filtering effects on microbiota. These results together imply a strong interplay between temporal, spatial and environmental factors on influencing microbial communities in the canal water, as previous noticed in natural lakes (Hu et al., 2017). Although potential contribution from unmeasured environmental factors cannot be ruled out, the remaining unexplained variation could possibly be generalized and linked to the influence of stochastic processes.

Contrasting proportion of stochasticity contributes to bacterial and micro-eukaryotic community assembly

Microbial community assembly is typically governed by four processes: selection, drift, speciation and dispersal (Vellend and Agrawal, 2010). While selection is inherently deterministic and drift is stochastic, speciation and dispersal could involve both deterministic

and stochastic mechanisms (Chase and Myers, 2011). For the MRP main canal, an investigation period of one year is probably short for resolving speciation. Thus, selection (deterministic processes), drift (stochastic processes) and dispersal (involving both) are expected to shape the observed community assembly patterns, and their relative contributions could be disentangled to help answer when, where, why and how much each process contribute to the observed patterns (Hanson et al., 2012).

Based on NST estimation and neutral modeling, we found that relative importance of stochastic and deterministic processes in microbial community assembly differ between bacteria and micro-eukaryotes, and among seasons and the canal sections. Neutral processes explained ~67% to 76% of the community patterns in October and March, but notably less in May (~41% and 13% for bacteria and eukaryotes, respectively). Liu et al. (2018a) reported a prominent shortage in water storage of Danjiangkou Reservoir related to seasonal flood control in May, which could have possibly imposed higher environmental selection effect to the microbiota in the reservoir and in the canal, making the observed decreased community stochasticity (or increased determinism) plausible. Although ecological stochasticity adds to challenge to predict microbial communities, the new knowledge produced in this study undoubtedly contribute to an improved understanding on when the canal water microbiota is more prone to stochastic processes, or under which conditions stochasticity is of more relevance.

The NST values suggested that the relative contribution of deterministic processes is

larger to bacterial community assembly compared with micro-eukaryotic communities, where stochasticity prevail. Recently, Gweon et al. (2021) reported that stochastic processes played a more important role in the planktonic bacterial communities in the River Thames, presumably due to the influence of various natural and anthropogenic sources of the surface water. What we found showed that in a water ecosystem almost isolated from other sources than resource water, bacterial communities are predominantly shaped by deterministic abiotic and biotic processes. The micro-eukaryotes, however, possess structurally and functionally more complex cells than prokaryotes (Massana and Logares, 2013), which could facilitate niche adaptation and diminish the environmental filtering effect at the community level. This was supported by our observation that the micro-eukaryotes were distributed with higher occurrence frequency and lower variability across the canal compared to bacteria. The different controlling mechanisms between bacterial and micro-eukaryotic community assembly found here has been observed in other natural habitats, such as Antarctic (Logares et al., 2018) and Tibetan lakes (Liu et al., 2020b). Our comparative analyses between bacteria and micro-eukaryotes across canal sections and over seasons thus provide first informative insights into the domain-specific, and space- and season-dependent patterns in the canal water community assembly.

Divergent population lifestyles underly microbiota dynamics and assembly revealed by absolute quantification

A synthesis of population and community ecological perspectives can provide valuable

insights into how microorganisms drive ecosystem processes (Koch et al., 2018). To resolve the population dynamics underlying the afore-discussed community patterns and assembly processes, we employed absolute quantification of microbial populations by combining relative sequence abundance of microbial lineages with overall bacterial 16S and eukaryotic 18S rRNA gene copies. Microbial populations in this giant system displayed interestingly divergent lifestyles reflecting the difference in their adaptability to the canal environment. For instance, the integration of neutral prediction-based partitioning of microbiota populations (as proxied by ASVs) with their absolute abundance taxonomic distribution revealed higher proportion of non-neutral group in abundance for bacteria than micro-eukaryotes (Fig. 5B), consistent with higher determinism in bacterial community assembly.

In particular, we demonstrate the use of local growth factor (LGF) as a meaningful metric to infer growth of immigrating microbial taxa in the canal. This leads to the first estimation that 67.1% of bacterial and 82.3% of micro-eukaryotic populations were capable of sustaining growth (LGF > 0), implying again better acclimation of eukaryotes to the environmental conditions in the canal. *Limnohabitans* sp., ubiquitous in a wide variety of freshwater habitats, showed remarkable growth across the canal. Members of *Limnohabitans* typically display high growth rate as well as metabolic flexibility, and they actively utilize alga-derived dissolved organic substances and are vulnerable to protist grazing (Horňák et al., 2017; Šimek et al., 2013; Šimek et al., 2011). *Cryptophyceae* were among the micro-eukaryotes that displayed prominent growth throughout the canal, which are small flagellates as important food of many protist and micro-crustaceans (Horňák et al., 2017;

Posch et al., 2015), and are perceived as characteristic of clean water (Lackey, 1941). Thus, *Limnohabitans* and *Cryptophyceae* are potentially important constituents of water food webs in the investigated canal. Two cyanobacterial lineages (*Cyanobum* PCC-6307) and *Acinetobacter* spp., on the contrary, were clearly depressed in the canal downstream sections, demonstrating the elimination of these microorganisms of water security concern along the canal. *Acinetobacter* spp. are ubiquitous in nature of major ecological and clinical significance, featuring, for instance, multi-antimicrobial resistance and rapid resistance development (Jung and Park, 2015). The diminishing of the versatile *Acinetobacter* spp. along the canal could possibly be attributed to unfavorable environmental conditions (Al Atrouni et al., 2016) or composite intra- and inter-domain biological interactions. Further research efforts with metagenomic approaches are needed to elaborate the mechanisms behind such die-off patterns. It is noteworthy that due to the restriction in taxonomic resolution of the ribosomal RNA gene sequencing analysis employed in this study, species-level confirmation is still needed to evaluate the relevance of these potentially pathogenic microorganisms. Together, our results demonstrated the power of local growth factor to identify key players in the investigated system and facilitate predicting fate of the microbes traversing such thousand-kilometer water diversion canal.

Conclusions

The main canal of the Middle Route Project of the South-to-North Water Diversion projects is an engineered system where the aquatic ecosystem is still at its early forming stage (since

2014). Various aspects of hydrology, geochemistry and ecology need to be intricately elaborated to jointly contribute to sustained system's eco-environmental services, especially water auto-purification ensuring its quality and competence for designed social-economic aims. The present study uncovered the underlying microbial community assembly mechanisms and population dynamics in a giant engineered water diversion water canal, such as the sharp decline of potentially harm-producing microbes (e.g., *Cyanobacteria*) and potential pathogens (e.g., *Acinetobacter* spp.), and the favorable enrichment of alga-derived organic scavengers (e.g., *Limnohabitans*) as potential key microbial food-web players, demonstrating the valuable insights provided by microbial community and population assessments for water monitoring and their potential in indicating ecosystem disturbance. Furthermore, the bacterial community shift driven by combined environmental effects as well as the stochasticity in micro-eukaryotic community assembly needs to be considered in water quality monitoring along the canal, towards an appropriate microbe-based biomonitoring which enables deeper understanding of processes behind water quality parameters. Considering possibly occasional pollution into the canal water caused by, for example, traffic accidents on highway bridges across the canal, and the central roles microorganisms play in the water quality maintenance, future efforts are needed to explore the metabolic functions of microbial community contributing to water purification (e.g., carbon, nitrogen, and phosphorus removal) during diversion and sustaining water quality.

Acknowledgments

We would like to thank Yuanzhen He, Guoqing Zhang, Zhipeng Zhang, Shengguang Yuan, Aoxiang Fan and Qinglin Jia for the assistance in water sampling. This work was supported by National Natural Science Foundation of China (Grant No. 51908467 and No. U2040210), and Zhejiang Provincial Natural Science Foundation of China (Grant No. LQ20C030002).

References

Al Atrouni, A., Joly-Guillou, M.-L., Hamze, M. and Kempf, M. 2016. Reservoirs of non-baumannii *Acinetobacter* species. Frontiers in Microbiology 7, 49.

Arrigo, K.R. 2005. Marine microorganisms and global nutrient cycles. Nature 437(7057), 349-355.

Barlow, J.T., Bogatyrev, S.R. and Ismagilov, R.F. 2020. A quantitative sequencing framework for absolute abundance measurements of mucosal and lumenal microbial communities. Nature Communications 11(1), 2590.

Battin, T.J., Besemer, K., Bengtsson, M.M., Romani, A.M. and Packmann, A.I. 2016. The ecology and biogeochemistry of stream biofilms. Nature Reviews Microbiology 14(4), 251-263.

Bell, T., Newman, J.A., Silverman, B.W., Turner, S.L. and Lilley, A.K. 2005. The contribution of species richness and composition to bacterial services. Nature 436(7054), 1157-1160.

Berry, D., Xi, C. and Raskin, L. 2006. Microbial ecology of drinking water distribution systems. Current Opinion in Biotechnology 17(3), 297-302.

Blanco-Bercial, L., Bradford-Grieve, J. and Bucklin, A. 2011. Molecular phylogeny of the Calanoida (Crustacea: Copepoda). Molecular Phylogenetics and Evolution 59(1), 103-113.

Bolyen, E., Rideout, J.R., Dillon, M.R., Bokulich, N.A., Abnet, C.C., Al-Ghalith, et al. 2019. Reproducible, interactive, scalable and extensible microbiome data science using QIIME 2. Nature Biotechnology 37, 852-857.

Borcard, D. and Legendre, P. 2002. All-scale spatial analysis of ecological data by means of principal coordinates of neighbour matrices. Ecological Modelling 153(1), 51-68.

Bradford-Grieve, J.M. 2002. Colonization of the pelagic realm by calanoid copepods. Hydrobiologia 485(1), 223-244.

Briones, A. and Raskin, L. 2003. Diversity and dynamics of microbial communities in engineered environments and their implications for process stability. Current Opinion in Biotechnology 14(3), 270-276.

Bru, D., Ramette, A., Saby, N.P.A., Dequiedt, S., Ranjard, L., Jolivet, C., Arrouays, D. and Philippot, L. 2011. Determinants of the distribution of nitrogen-cycling microbial communities at the landscape scale. The ISME Journal 5(3), 532-542.

Burns, A.R., Stephens, W.Z., Stagaman, K., Wong, S., Rawls, J.F., Guillemin, K. and Bohannan, B.J.M. 2016. Contribution of neutral processes to the assembly of gut microbial communities in the zebrafish over host development. The ISME Journal 10, 655.

573 Callahan, B.J., McMurdie, P.J., Rosen, M.J., Han, A.W., Johnson, A.J.A. and Holmes, S.P. 2016.
574 DADA2: High-resolution sample inference from Illumina amplicon data. Nature Methods
575 13(7), 581-583.

576 Chase, J.M. and Myers, J.A. 2011. Disentangling the importance of ecological niches from
577 stochastic processes across scales. Philosophical Transactions of the Royal Society B:
578 Biological Sciences 366(1576), 2351-2363.

579 Chen, F.Z., You, L.J., Yang, F., Wang, L.N., Guo, X.Q., Gao, F., Hua, C., Tan, C., Fang, L., Shan, R.Q., Zeng,
580 W.J., Wang, B., Wang, R., Xu, X. and Wei, X.F. 2020. CNGBdb: China National GeneBank
581 DataBase. Yi Chuan 42(8), 799-809.

582 Chen, W., Ren, K., Isabwe, A., Chen, H., Liu, M. and Yang, J. 2019. Stochastic processes shape
583 microeukaryotic community assembly in a subtropical river across wet and dry seasons.
584 Microbiome 7(1), 138.

585 Chen, Z., Yuan, J., Sun, F., Zhang, F., Chen, Y., Ding, C., Shi, J., Li, Y. and Yao, L. 2018. Planktonic
586 fungal community structures and their relationship to water quality in the Danjiangkou
587 Reservoir, China. Scientific Reports 8(1), 10596.

588 Dang, C., Liu, W., Lin, Y., Zheng, M., Jiang, H., Chen, Q. and Ni, J. 2018. Dominant role of
589 ammonia-oxidizing bacteria in nitrification due to ammonia accumulation in sediments of
590 Danjiangkou reservoir, China. Applied Microbiology and Biotechnology 102(7), 3399-3410.

591 Du, M., Peng, X., Zhang, H., Ye, C., Dasgupta, S., Li, J., Li, J., Liu, S., Xu, H., Chen, C., Jing, H., Xu, H., Liu,
592 J., He, S., He, L, Cai, S., Chen, S. and Ni, J. 2021. Geology, environment, and life in the
593 deepest part of the world's oceans. The Innovation 2(2), 100109.

594 Ghylin, T.W., Garcia, S.L., Moya, F., Oyserman, B.O., Schwientek, P., Forest, K.T., et al. 2014.
595 Comparative single-cell genomics reveals potential ecological niches for the freshwater acI
596 Actinobacteria lineage. The ISME Journal 8, 2503.

597 Glöckner, F.O., Zaichikov, E., Belkova, N., Denissova, L., Pernthaler, J., Pernthaler, A. and Amann, R.
598 2000. Comparative 16S rRNA analysis of lake bacterioplankton reveals globally distributed
599 phylogenetic clusters including an abundant group of *Actinobacteria*. Applied and
600 Environmental Microbiology 66(11), 5053-5065.

601 Guo, X., Chen, F., Gao, F., Li, L., Liu, K., You, L., Hua, C., Yang, F., Liu, W., Peng, C., Wang, L., Yang, X.,
602 Zhou, F., Tong, J., Cai, J., Li, Z., Wan, B., Zhang, L., Yang, T., Zhang, M., Yang, L., Yang, Y., Zeng,
603 W., Wang, B., Wei, X. and Xu, X. 2020. CNSA: a data repository for archiving omics data.
604 Database (Oxford) 2020.

605 Gweon, H.S., Bowes, M.J., Moorhouse, H.L., Oliver, A.E., Bailey, M.J., Acreman, M.C. and Read, D.S.
606 2021. Contrasting community assembly processes structure lotic bacteria

607 metacommunities along the river continuum. Environmental Microbiology 23(1), 484-498.

608 Hanson, C.A., Fuhrman, J.A., Horner-Devine, M.C. and Martiny, J.B.H. 2012. Beyond biogeographic
609 patterns: processes shaping the microbial landscape. Nature Reviews Microbiology 10(7),
610 497-506.

611 Horňák, K., Kasalický, V., Šimek, K. and Grossart, H.P. 2017. Strain-specific consumption and
612 transformation of alga-derived dissolved organic matter by members of the Limnohabitans-C
613 and Polynucleobacter-B clusters of Betaproteobacteria. Environmental Microbiology 19(11),
614 4519-4535.

615 Hu, A., Ju, F., Hou, L., Li, J., Yang, X., Wang, H., Mulla, S.I., Sun, Q., Bürgmann, H. and Yu, C.-P. 2017.
616 Strong impact of anthropogenic contamination on the co-occurrence patterns of a riverine
617 microbial community. Environmental Microbiology 19(12), 4993-5009.

618 Hull, N.M., Ling, F., Pinto, A.J., Albertsen, M., Jang, H.G., Hong, P.-Y., Konstantinidis, K.T., LeChevallier,
619 M., Colwell, R.R. and Liu, W.-T. 2019. Drinking water microbiome project: Is it time?
620 Trends in Microbiology 27(8), 670-677.

621 Ju, F., Beck, K., Yin, X., Maccagnan, A., McArdell, C.S., Singer, H.P., Johnson, D.R., Zhang, T. and
622 Bürgmann, H. 2019. Wastewater treatment plant resistomes are shaped by bacterial
623 composition, genetic exchange, and upregulated expression in the effluent microbiomes. The
624 ISME Journal 13(2), 346-360.

625 Ju, F., Lau, F. and Zhang, T. 2017. Linking microbial community, environmental variables, and
626 methanogenesis in anaerobic biogas digesters of chemically enhanced primary treatment
627 sludge. Environmental Science and Technology 51(7), 3982-3992.

628 Ju, F., Xia, Y., Guo, F., Wang, Z. and Zhang, T. 2014. Taxonomic relatedness shapes bacterial
629 assembly in activated sludge of globally distributed wastewater treatment plants.
630 Environmental Microbiology 16(8), 2421-2432.

631 Ju, F. and Zhang, T. 2015. Bacterial assembly and temporal dynamics in activated sludge of a
632 full-scale municipal wastewater treatment plant. The ISME journal 9(3), 683.

633 Jung, J. and Park, W. 2015. *Acinetobacter* species as model microorganisms in environmental
634 microbiology: current state and perspectives. Applied Microbiology and Biotechnology 99,
635 2533-2548.

636 Keshri, J., Pradeep Ram, A.S., Nana, P.A. and Sime-Ngando, T. 2018. Taxonomical resolution and
637 distribution of bacterioplankton along the vertical gradient reveals pronounced
638 spatiotemporal patterns in contrasted temperate freshwater lakes. Microbial Ecology 76(2),
639 372-386.

640 Koch, B.J., McHugh, T.A., Hayer, M., Schwartz, E., Blazewicz, S.J., Dijkstra, P., van Gestel, N., Marks, J.C.,
641 Mau, R.L., Morrissey, E.M., Pett-Ridge, J. and Hungate, B.A. 2018. Estimating
642 taxon-specific population dynamics in diverse microbial communities. Ecosphere 9(1),
643 e02090.

644 Lackey, J.B. 1941. Two groups of flagellated algae serving as indicators of clean water. Journal
645 (American Water Works Association) 33(6), 1099-1110.

646 Li, Y.-Y., Zhang, Z., Yin, W., Hu, L., Liang, Z., Wang, F. and Lei, G. 2012. Zooplankton diversity in the
647 water resource area of the Mid-Line Project of South to North Water Division. Advanced
648 Materials Research 356-360, 199-203.

649 Liao, Y., Chen, J., Brandt, B.W., Zhu, Y., Li, J., van Loveren, C. and Deng, D.M. 2015. Identification
650 and functional analysis of genome mutations in a fluoride-resistant *Streptococcus mutans*
651 strain. PLOS ONE 10(4), e0122630.

652 Lindh, M.V., Lefébure, R., Degerman, R., Lundin, D., Andersson, A. and Pinhassi, J. 2015.
653 Consequences of increased terrestrial dissolved organic matter and temperature on
654 bacterioplankton community composition during a Baltic Sea mesocosm experiment. AMBIO
655 44(3), 402-412.

656 Liu, H., Yin, J. and Feng, L. 2018a. The dynamic changes in the storage of the Danjiangkou
657 Reservoir and the influence of the South-North Water Transfer Project. Scientific Reports 8(1),
658 8710.

659 Liu, J., Zhu, S., Liu, X., Yao, P., Ge, T. and Zhang, X.-H. 2020a. Spatiotemporal dynamics of the
660 archaeal community in coastal sediments: assembly process and co-occurrence relationship.
661 The ISME Journal 14(6):1463-1478.

662 Liu, K., Liu, Y., Hu, A., Wang, F., Chen, Y., Gu, Z., Anslan, S. and Hou, J. 2020b. Different community
663 assembly mechanisms underlie similar biogeography of bacteria and microeukaryotes in
664 Tibetan lakes. FEMS Microbiology Ecology 96(6).

665 Liu, T., Zhang, A.N., Wang, J., Liu, S., Jiang, X., Dang, C., Ma, T., Liu, S., Chen, Q., Xie, S., Zhang, T. and Ni,
666 J. 2018b. Integrated biogeography of planktonic and sedimentary bacterial communities
667 in the Yangtze River. Microbiome 6(1), 16.

668 Logares, R., Tesson, S.V.M., Canbäck, B., Pontarp, M., Hedlund, K. and Rengefors, K. 2018.
669 Contrasting prevalence of selection and drift in the community structuring of bacteria and
670 microbial eukaryotes. Environmental Microbiology 20(6), 2231-2240.

671 Luo, Z., Li, S., Hou, K. and Ji, G. 2019. Spatial and seasonal bacterioplankton community dynamics
672 in the main channel of the Middle Route of South-to-North Water Diversion Project. Research
673 in Microbiology 170(1), 24-34.

Massana, R. and Logares, R. 2013. Eukaryotic versus prokaryotic marine picoplankton ecology. Environmental Microbiology 15(5), 1254-1261.

Mei, R., Kim, J., Wilson, F.P., Bocher, B.T.W. and Liu, W.-T. 2019. Coupling growth kinetics modeling with machine learning reveals microbial immigration impacts and identifies key environmental parameters in a biological wastewater treatment process. Microbiome 7(1), 65.

Mei, R. and Liu, W.-T. 2019. Quantifying the contribution of microbial immigration in engineered water systems. Microbiome 7(1), 144.

Ning, D., Deng, Y., Tiedje, J.M. and Zhou, J. 2019. A general framework for quantitatively assessing ecological stochasticity. Proceedings of the National Academy of Sciences 116(34), 16892-16898.

Niderdorfer, R., Fragner, L., Yuan, L., Hausherr, D., Wei, J., Magyar, P., Joss, A., Lehmann, M.F., Ju, F. and Bürgmann, H. 2021. Distinct growth stages controlled by the interplay of deterministic and stochastic processes in functional anammox biofilms. Water Research 200, 117225.

Nong, X., Shao, D., Xiao, Y., Zhong and Hua 2019. Spatio-temporal characterization analysis and water quality assessment of the South-to-North Water Diversion Project of China. Int J Environ Res Public Health 16(12), 2227.

Ochoa-Herrera, V., Banihani, Q., León, G., Khatri, C., Field, J.A. and Sierra-Alvarez, R. 2009. Toxicity of fluoride to microorganisms in biological wastewater treatment systems. Water Research 43(13), 3177-3186.

Posch, T., Eugster, B., Pomati, F., Pernthaler, J., Pitsch, G. and Eckert, E. 2015. Network of interactions between ciliates and phytoplankton during spring. Frontiers in Microbiology 6, 14.

R Core Team 2020 R: A language and environment for statistical computing, R foundation for statistical computing.

Sagova-Mareckova, M., Boenigk, J., Bouchez, A., Cermakova, K., Chonova, T., Cordier, et al. 2021. Expanding ecological assessment by integrating microorganisms into routine freshwater biomonitoring. Water Research 191, 116767.

Šimek, K., Kasalický, V., Jezbera, J., Horňák, K., Nedoma, J., Hahn, M.W., Bass, D., Jost, S. and Boenigk, J. 2013. Differential freshwater flagellate community response to bacterial food quality with a focus on *Limnohabitans* bacteria. The ISME Journal 7(8), 1519-1530.

Šimek, K., Kasalický, V., Zapomělová, E. and Horňák, K. 2011. Alga-derived substrates select for

706 distinct betaproteobacterial lineages and contribute to niche separation in *Limnohabitan*
707 strains. Applied and Environmental Microbiology 77(20), 7307-7315.

708 Sloan, W.T., Lunn, M., Woodcock, S., Head, I.M., Nee, S. and Curtis, T.P. 2006. Quantifying the roles
709 of immigration and chance in shaping prokaryote community structure. Environmental
710 Microbiology 8(4), 732-740.

711 Stämmler, F., Gläsner, J., Hiergeist, A., Holler, E., Weber, D., Oefner, P.J., Gessner, A. and Spang, R.
712 2016. Adjusting microbiome profiles for differences in microbial load by spike-in bacteria.
713 Microbiome 4(1), 28.

714 Stegen, J.C., Lin, X., Fredrickson, J.K., Chen, X., Kennedy, D.W., Murray, C.J., Rockhold, M.L. and
715 Konopka, A. 2013. Quantifying community assembly processes and identifying features
716 that impose them. The ISME Journal 7, 2069.

717 Tkacz, A., Hortala, M. and Poole, P.S. 2018. Absolute quantitation of microbiota abundance in
718 environmental samples. Microbiome 6(1), 110.

719 Vanwonterghem, I., Jensen, P.D., Dennis, P.G., Hugenholtz, P., Rabaey, K. and Tyson, G.W. 2014.
720 Deterministic processes guide long-term synchronised population dynamics in replicate
721 anaerobic digesters. The ISME Journal 8(10), 2015-2028.

722 Vellend, M. and Agrawal, A. 2010. Conceptual synthesis in community ecology. The Quarterly
723 Review of Biology 85(2), 183-206.

724 Vieira-Silva, S., Sabino, J., Valles-Colomer, M., Falony, G., Kathagen, G., Caenepeel, C., Cleynen, I., van
725 der Merwe, S., Vermeire, S. and Raes, J. 2019. Quantitative microbiome profiling
726 disentangles inflammation- and bile duct obstruction-associated microbiota alterations
727 across PSC/IBD diagnoses. Nature Microbiology 4(11), 1826-1831.

728 Wang, W., Ren, K., Chen, H., Gao, X., Rønn, R. and Yang, J. 2020. Seven-year dynamics of testate
729 amoeba communities driven more by stochastic than deterministic processes in two
730 subtropical reservoirs. Water Research 185, 116232.

731 Woolhouse, M. E. J., Gowtage-Sequeria, S. and Evans, B. 2015. Quantitative analysis of the
732 characteristics of emerging and re-emerging human pathogens.
733 http://webarchive.nationalarchives.gov.uk/20121206154522/http://www.bis.gov.uk/assets/f
734 oresight/docs/infectious-diseases/t16.pdf 2015.

735 Wu, L., Ning, D., Zhang, B., Li, Y., Zhang, P., Shan, et al. 2019. Global diversity and biogeography of
736 bacterial communities in wastewater treatment plants. Nature Microbiology 4, 1183-1195.

737 Yan, L., Herrmann, M., Kampe, B., Lehmann, R., Totsche, K.U. and Küsel, K. 2020. Environmental
738 selection shapes the formation of near-surface groundwater microbiomes. Water Research

170, 115341.

Yang, C., Wang, Q., Simon, P.N., Liu, J., Liu, L., Dai, X., Zhang, X., Kuang, J., Igarashi, Y., Pan, X. and Luo, F. 2017. Distinct network interactions in particle-associated and free-living bacterial communities during a *Microcystis aeruginosa* bloom in a plateau lake. Frontiers in microbiology 8, 1202-1202.

Zhang, X., Gao, X., Li, C., Luo, X. and Wang, Y. 2019a. Fluoride contributes to the shaping of microbial community in high fluoride groundwater in Qiji County, Yuncheng City, China. Scientific Reports 9, 14488.

Zhang, X., Li, B., Xu, H., Wells, M., Tefsen, B. and Qin, B. 2019b. Effect of micronutrients on algae in different regions of Taihu, a large, spatially diverse, hypereutrophic lake. Water Research 151, 500-514.

Zhou, J., Deng, Y., Zhang, P., Xue, K., Liang, Y., Van Nostrand, J.D., Yang, Y., He, Z., Wu, L., Stahl, D.A., Hazen, T.C., Tiedje, J.M. and Arkin, A.P. 2014. Stochasticity, succession, and environmental perturbations in a fluidic ecosystem. Proceedings of the National Academy of Sciences 111(9), E836-E845.

Figure Legends

Fig. 1 Four sampling campaigns at 19 sites along the 1432-km main canal of the MRP of the SNWD project selected for microbial and physicochemical analyses. Colored fans indicate months of sampling at each site. The map was created using QGIS (version 3.10.5, www.qgis.org).

Fig. 2 Seasonal variation of bacterial and micro-eukaryotic communities in the canal. (A-F) Ribosomal RNA gene copies (A and B), Chao1 richness (C and D) and Shannon diversity index (E and F) across seasons. Different letters above boxes indicate statistically significant differences between months of sampling (Mann-Whitney *U* test with the Benjamini-Hochberg correction, $P < 0.05$). (G-H) Ordination analysis of communities using non-metric multidimensional scaling (NMDS) based on Bray-Curtis dissimilarities. Trajectories indicate locations of sampling sites along the canal.

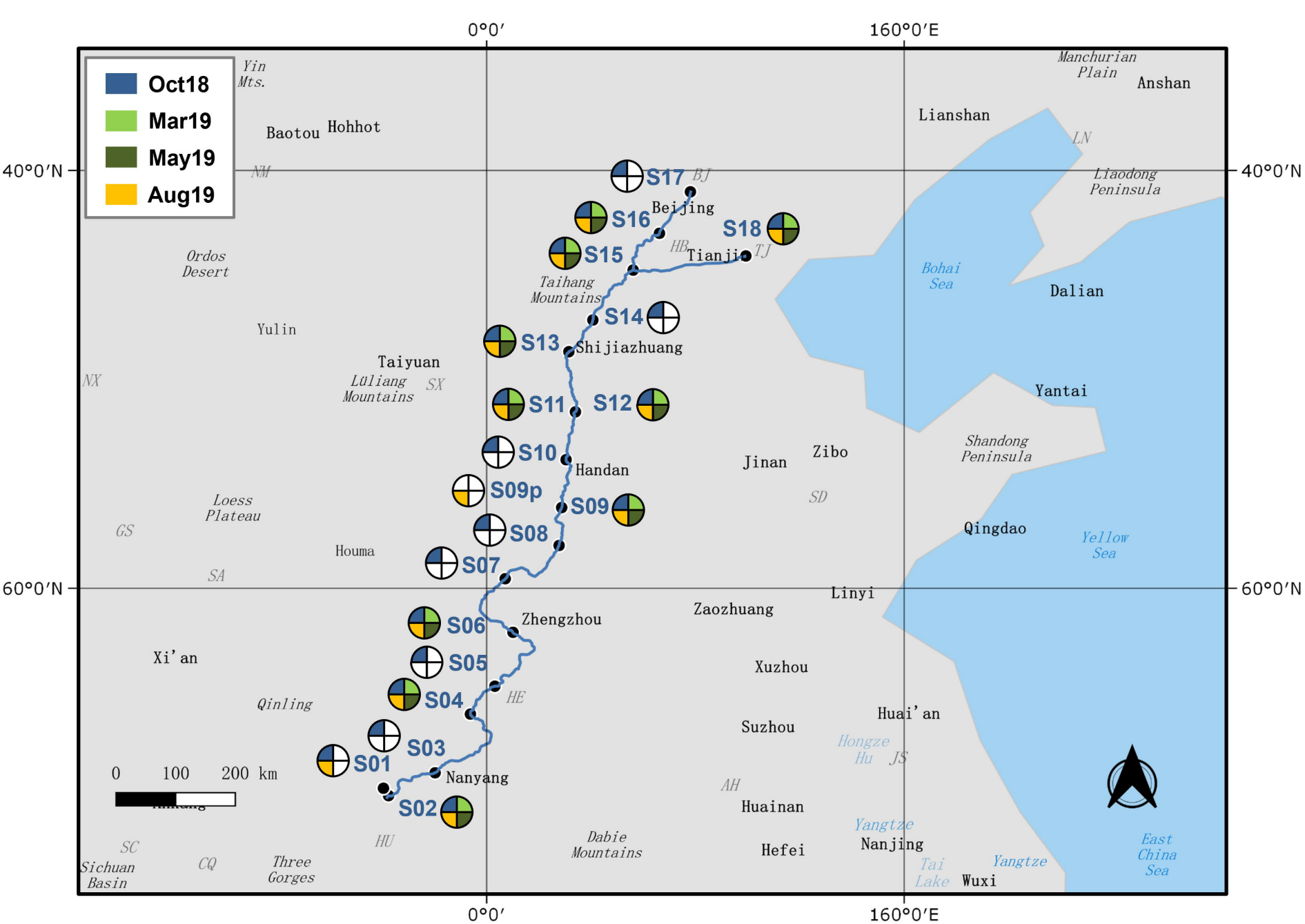
Fig. 3 Microbial community composition and diversity along the MRP main canal (~1432 km) of the SNWD project in October 2018. (A) Bacterial composition with abundant phyla (> 1%) and 10 most abundant genera shown. (B) Micro-eukaryotic composition with abundant phyla (> 1%) and 10 most abundant orders shown.

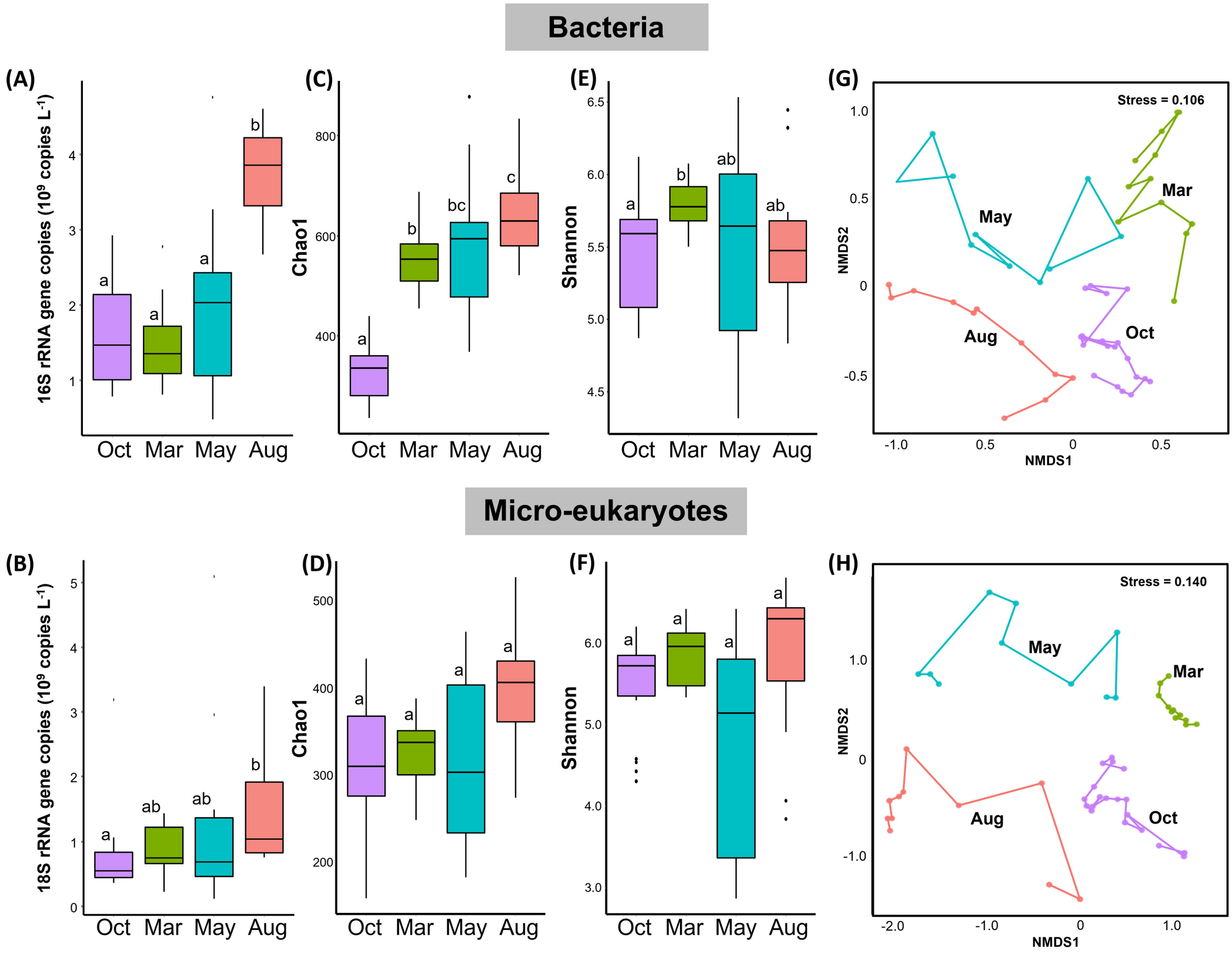
Fig. 4 Redundancy analysis (RDA) and variation partitioning analysis (VPA) demonstrating the relative influence of seasonality, spatial and environmental factors on the canal microbial communities. The RDA biplots show factors impacting bacterial (A) and micro-eukaryotic (B) communities. The VPA diagrams show the percentage contribution of seasonal, spatial and

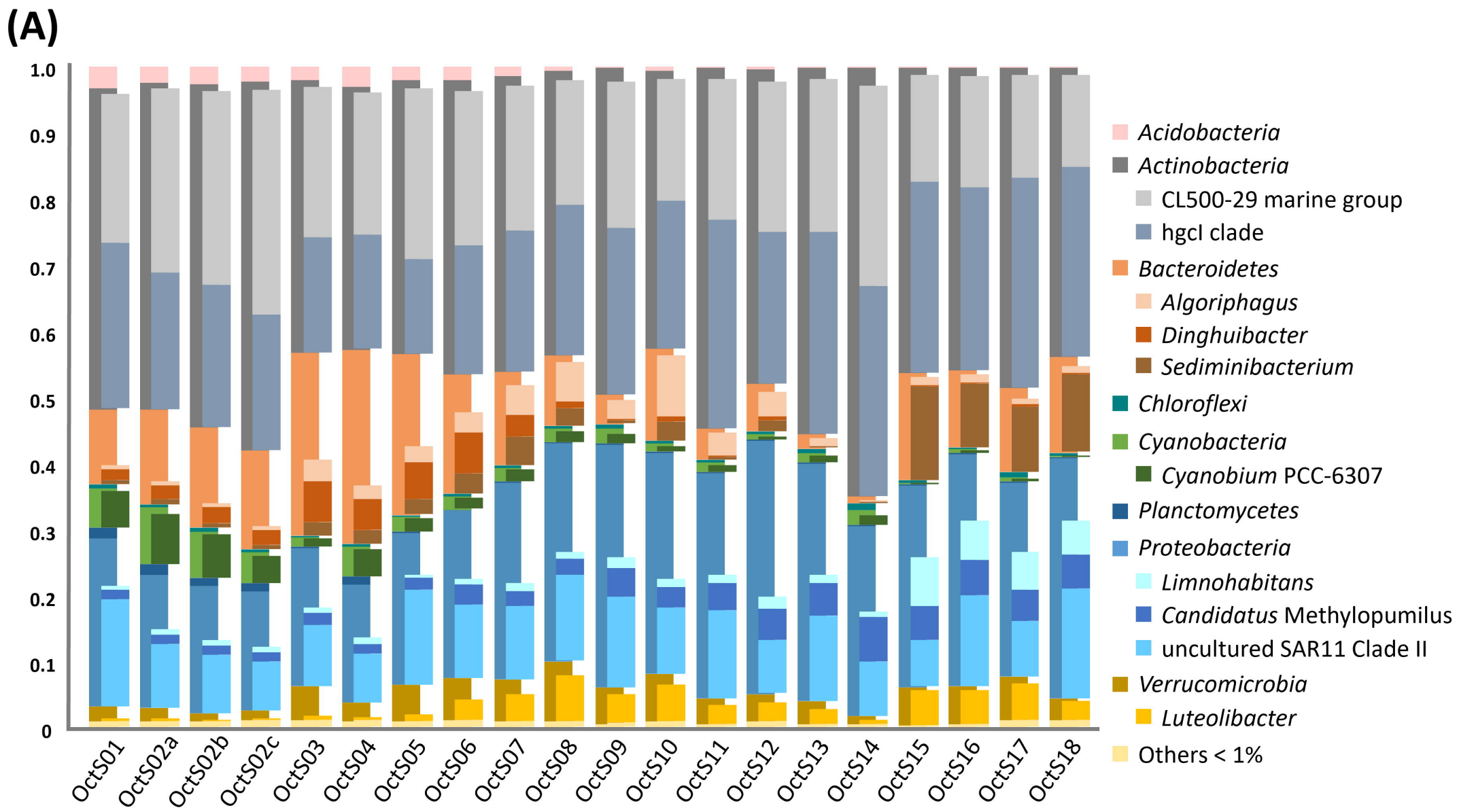
environmental factors to bacterial (C) and micro-eukaryotic (D) community variations. Only those factors identified to significantly influence bacterial and micro-eukaryotic communities are shown in the RDA and included in the VPA analyses. PCNMs, geographic factors generated by principal coordinate analysis of neighborhood matrices. T, temperature; F, fluoride; TN, total nitrogen; DO, dissolved oxygen.

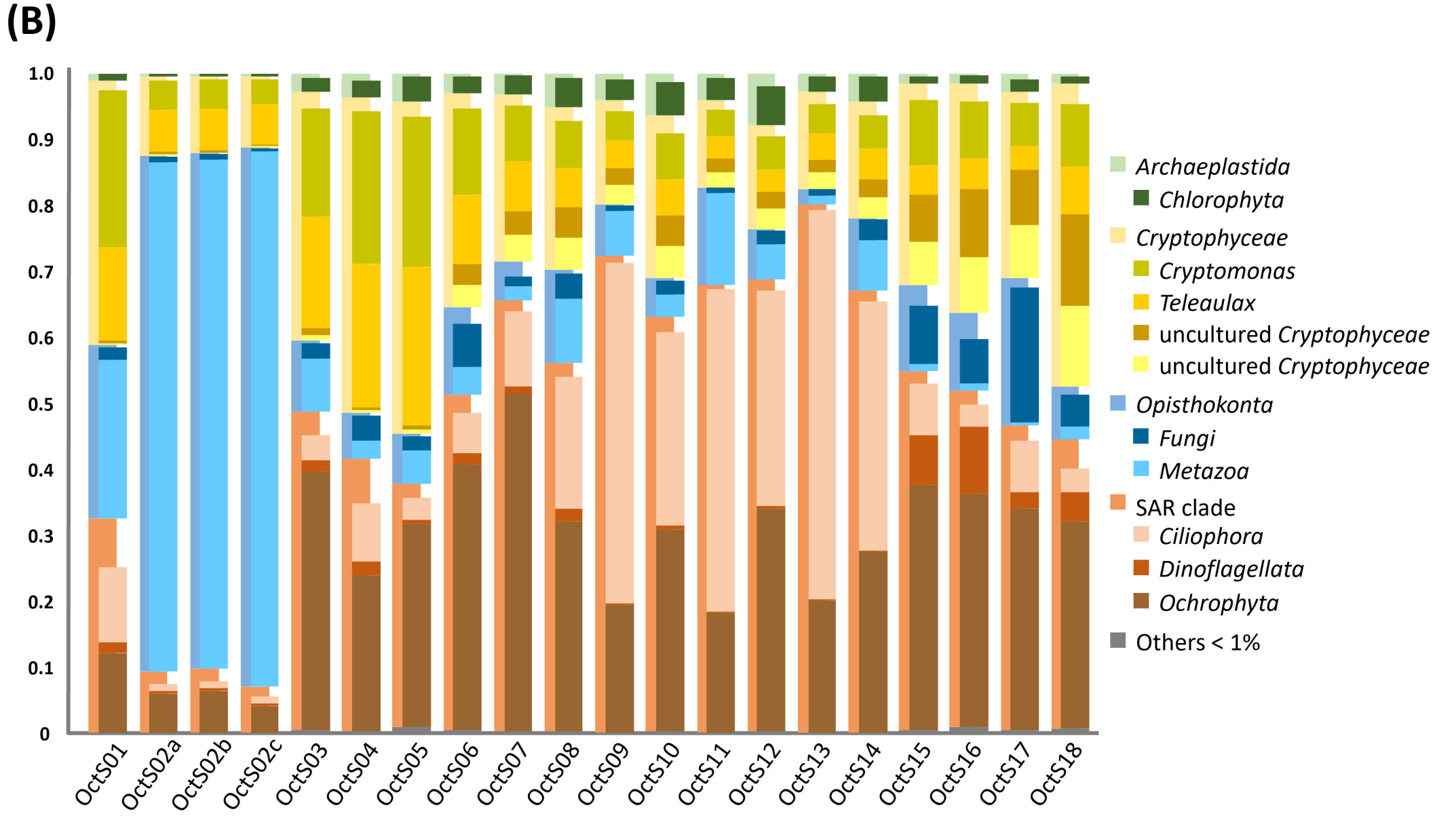
Fig. 5 Normalized stochasticity ratio (NST) estimation and microbiota partitioning by population compliance to the neutral model prediction. (A) NST estimated stochasticity in bacterial and micro-eukaryotic community assembly. (B) Proportions in richness (ASV number) and abundance (ribosomal RNA gene copies) of the three fitting groups (occurrence frequency above prediction, as predicted and below prediction) within the microbial communities to the Sloan neutral model. (C and D) Taxonomic composition of the three fitting partitions in richness (Ri) and abundance (Ab) for bacterial (C) and micro-eukaryotic (D) communities. Only abundant bacterial phyla (> 1%) and 10 most abundant micro-eukaryotic orders are shown.

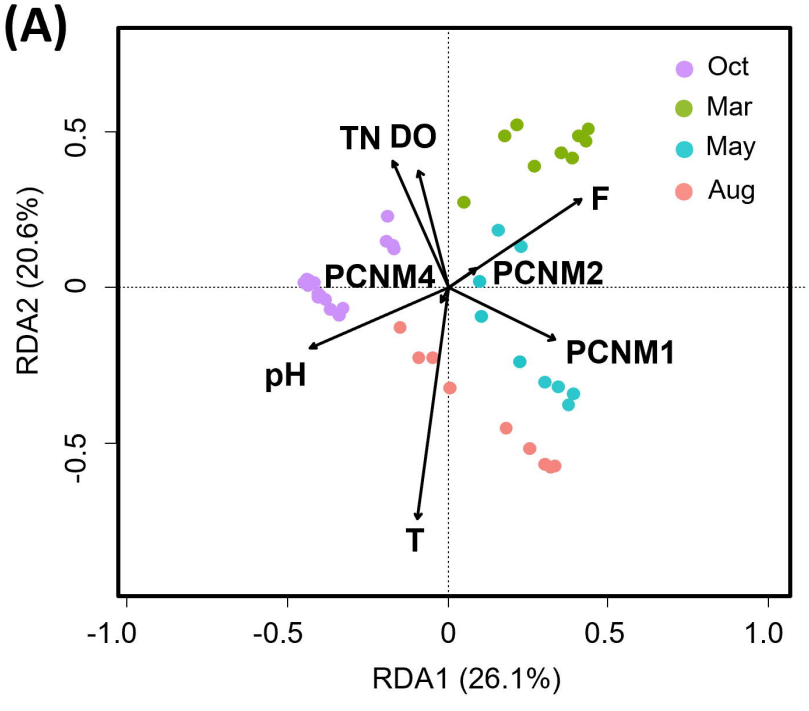
Fig. 6 Microbial population proliferation across canal sections evaluated by local growth factor analysis. Distribution patterns of population local growth factors in the different canal sections are shown with violin plots for bacteria (A) and micro-eukaryotes (B). Each dot represents an ASV with colors indicating its average abundance across the canal.

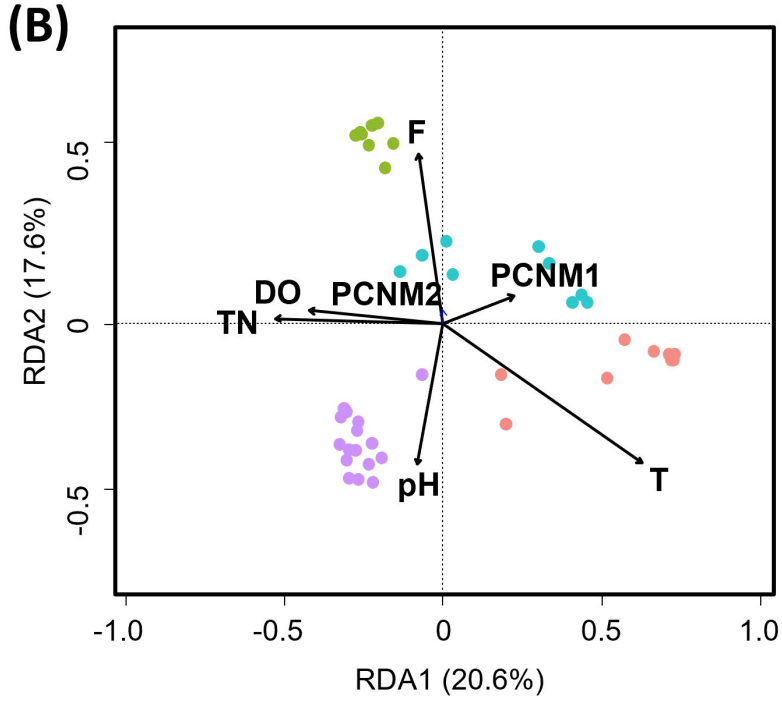

Oct18
Mar19
May19
Aug19
0°0′
160°0′E
40°0′N
60°0′N
Yin Mts.
Baotou
Hohhot
NM
Ordos Desert
Yulin
NX
Taiyuan
Lüliang Mountains
SX
Loess Plateau
GS
SA
Houma
Xi'an
Qinling
SC
Sichuan Basin
CQ
Three Gorges
HU
Manchurian Plain
Anshan
Lianshan
LN
Liaodong Peninsula
Bohai Sea
Dalian
Yantai
Shandong Peninsula
Jinan
Zibo
SD
Qingdao
Yellow Sea
Linyi
Zaozhuang
Xuzhou
Suzhou
Huai'an
Hongze Hu
JS
AH
Huainan
Hefei
Dabie Mountains
Yangtze
Nanjing
Tai Lake
Wuxi
East China Sea
Beijing
BJ
Tianjin
TJ
HB
Taihang Mountains
Shijiazhuang
Handan
Zhengzhou
HE
Nanyang
S01
S02
S03
S04
S05
S06
S07
S08
S09
S09p
S10
S11
S12
S13
S14
S15
S16
S17
S18
0 100 200 km

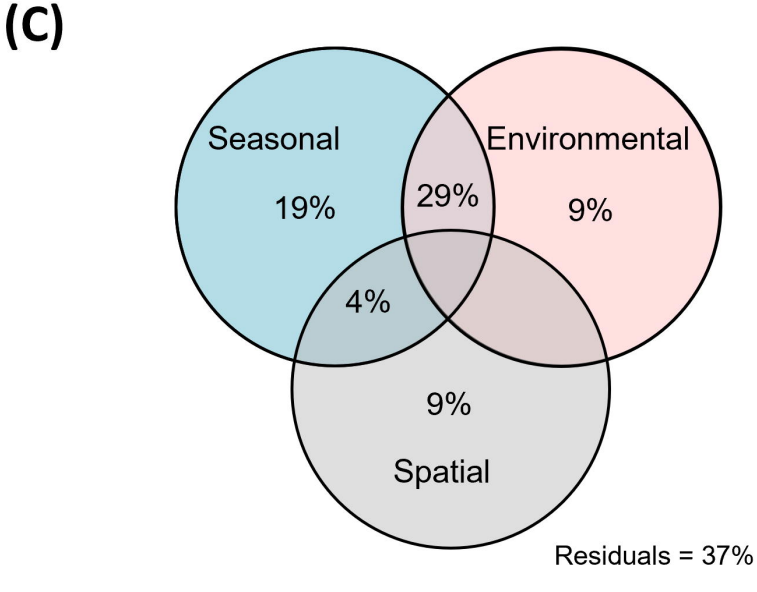

Bacteria
(A)
16S rRNA gene copies (10⁹ copies L⁻¹)
Oct Mar May Aug
(C)
Chao1
(E)
Shannon
(G)
Stress = 0.106
NMDS1
NMDS2
May
Mar
Aug
Oct
Micro-eukaryotes
(B)
18S rRNA gene copies (10⁹ copies L⁻¹)
(D)
(F)
(H)
Stress = 0.140

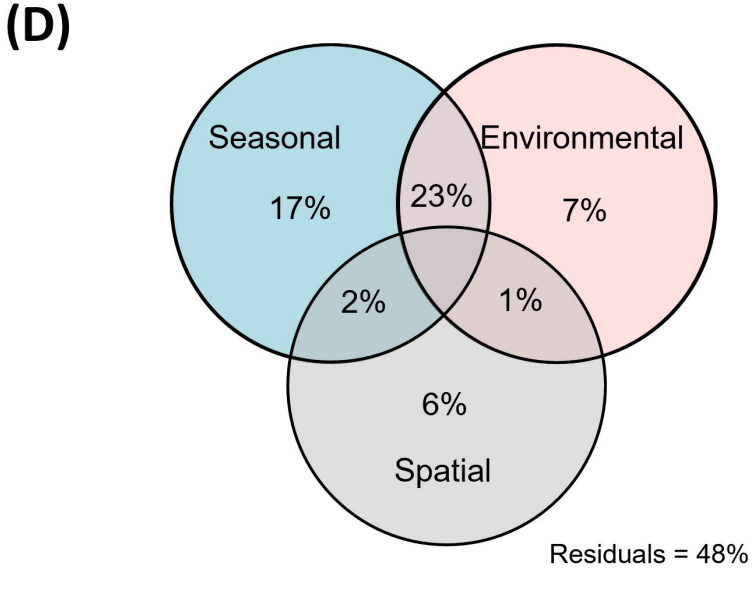

(A)
1.0
0.9
0.8
0.7
0.6
0.5
0.4
0.3
0.2
0.1
0
OctS01
OctS02a
OctS02b
OctS02c
OctS03
OctS04
OctS05
OctS06
OctS07
OctS08
OctS09
OctS10
OctS11
OctS12
OctS13
OctS14
OctS15
OctS16
OctS17
OctS18
Acidobacteria
Actinobacteria
CL500-29 marine group
hgcI clade
Bacteroidetes
Algoriphagus
Dinghuibacter
Sediminibacterium
Chloroflexi
Cyanobacteria
Cyanobium PCC-6307
Planctomycetes
Proteobacteria
Limnohabitans
Candidatus Methylopumilus
uncultured SAR11 Clade II
Verrucomicrobia
Luteolibacter
Others < 1%

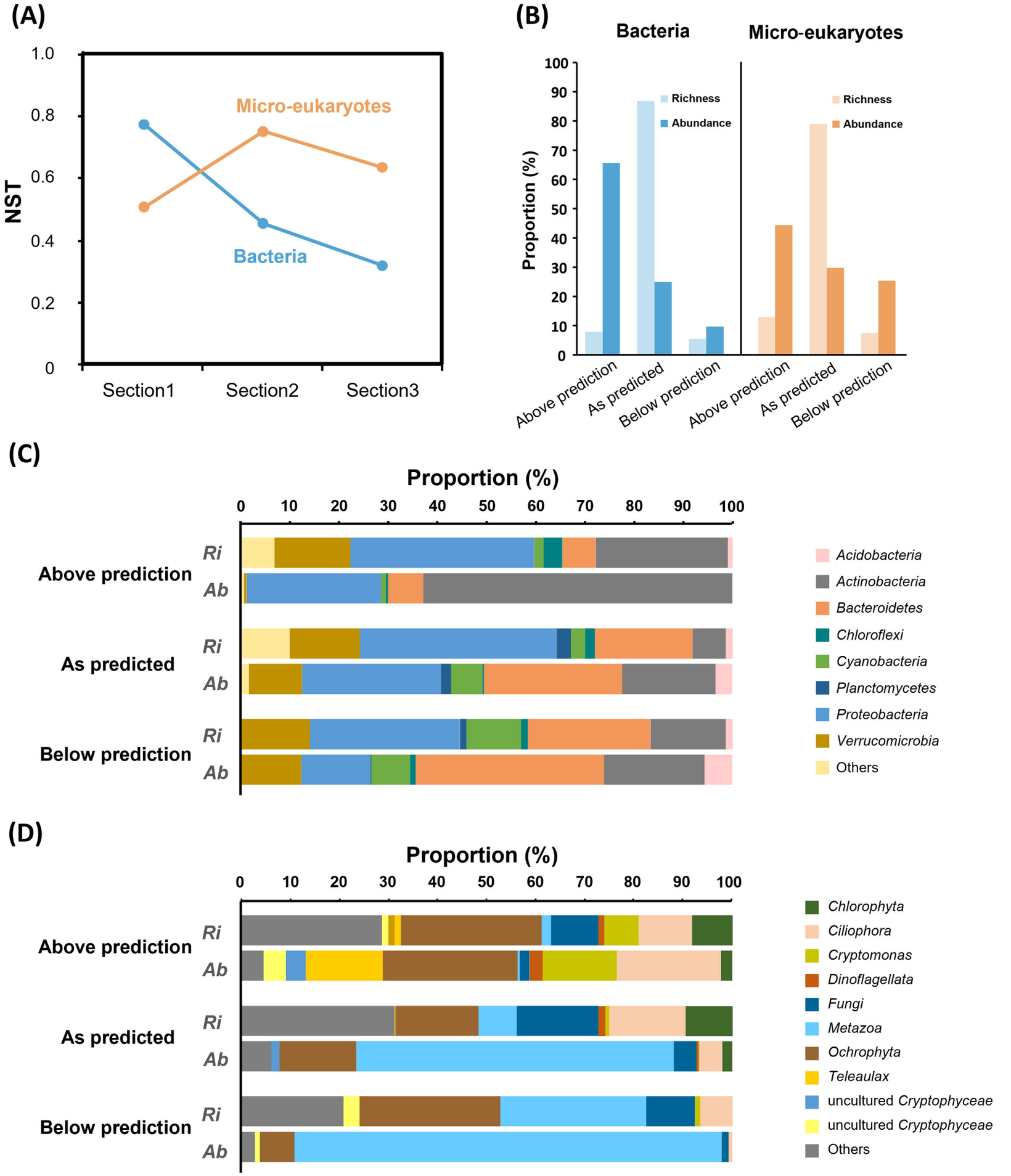

(B)
1.0
0.9
0.8
0.7
0.6
0.5
0.4
0.3
0.2
0.1
0
OctS01
OctS02a
OctS02b
OctS02c
OctS03
OctS04
OctS05
OctS06
OctS07
OctS08
OctS09
OctS10
OctS11
OctS12
OctS13
OctS14
OctS15
OctS16
OctS17
OctS18
Archaeplastida
Chlorophyta
Cryptophyceae
Cryptomonas
Teleaulax
uncultured Cryptophyceae
uncultured Cryptophyceae
Opisthokonta
Fungi
Metazoa
SAR clade
Ciliophora
Dinoflagellata
Ochrophyta
Others < 1%

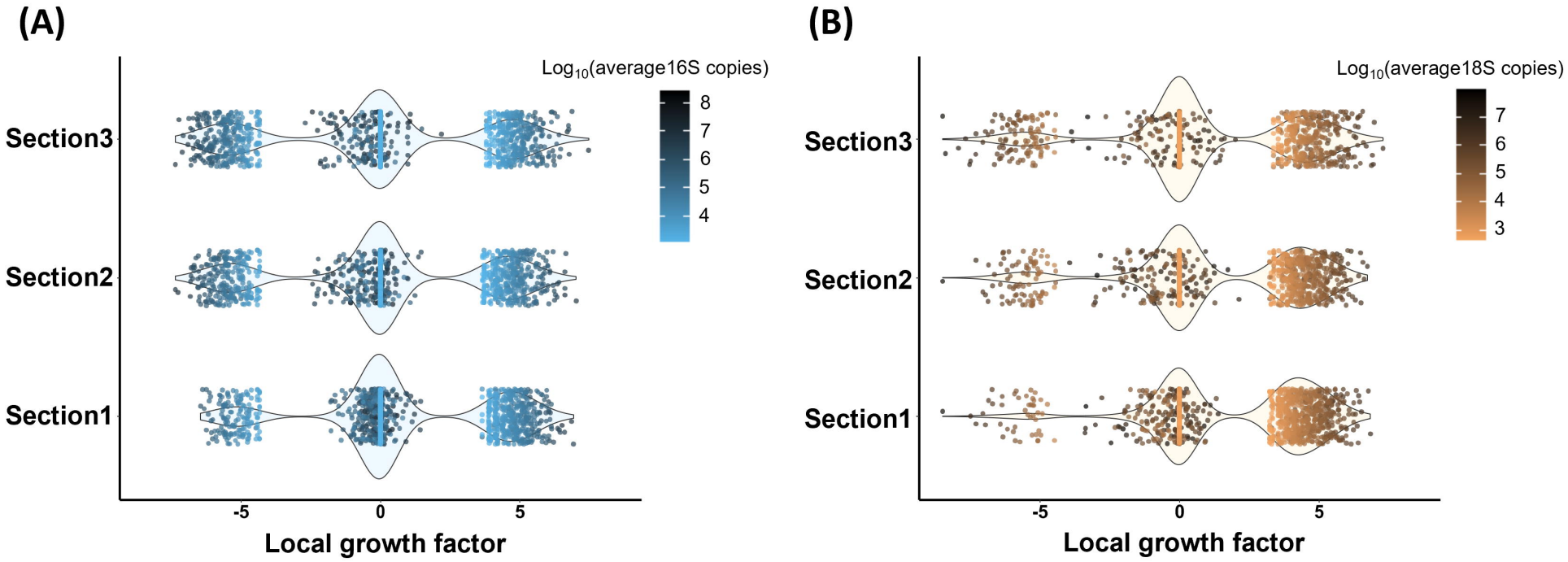

(A)
Oct
Mar
May
Aug
TN DO
F
PCNM4
PCNM2
PCNM1
pH
T
RDA2 (20.6%)
RDA1 (26.1%)
0.5
0
-0.5
-1.0
-0.5
0
0.5
1.0


(B)
F
DO
PCNM2
PCNM1
TN
pH
T
RDA2 (17.6%)
RDA1 (20.6%)
0.5
0
-0.5
-1.0
-0.5
0
0.5
1.0


(C)
Seasonal
19%
Environmental
9%
29%
4%
9%
Spatial
Residuals = 37%


(D)
Seasonal
17%
Environmental
7%
23%
2%
1%
6%
Spatial
Residuals = 48%


(A)
NST
1.0
0.8
0.6
0.4
0.2
0
Micro-eukaryotes
Bacteria
Section1
Section2
Section3
(B)
Bacteria
Micro-eukaryotes
Proportion (%)
Richness
Abundance
Above prediction
As predicted
Below prediction
(C)
Proportion (%)
Ri
Ab
Above prediction
As predicted
Below prediction
Acidobacteria
Actinobacteria
Bacteroidetes
Chloroflexi
Cyanobacteria
Planctomycetes
Proteobacteria
Verrucomicrobia
Others
(D)
Proportion (%)
Chlorophyta
Ciliophora
Cryptomonas
Dinoflagellata
Fungi
Metazoa
Ochrophyta
Teleaulax
uncultured Cryptophyceae
uncultured Cryptophyceae
Others


(A)
Log10(average16S copies)
8
7
6
5
4
Section3
Section2
Section1
-5
0
5
Local growth factor
(B)
Log10(average18S copies)
7
6
5
4
3
Section3
Section2
Section1
-5
0
5
Local growth factor