Soil fauna may buffer the negative effects of drought on alien plant invasion

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Abstract

Although many studies have tested the direct effects of drought on alien plant invasion,

less is known about how drought affects alien plant invasion indirectly via other groups of

organisms such as soil fauna. To test for such indirect effects, we grew single plant of nine

naturalized alien target species in pot-mesocosms with a native community of five native

grassland species under four combinations of two drought (well-watered *vs* drought) and two

soil-fauna-inoculation (with *vs* without) treatments. We found that drought decreased the

absolute and the relative biomass production of the alien plants, and thus reduced their

competitive advantage in the native community. Drought decreased the abundance of soil

fauna, particularly the soil mites, but did not affect abundance and richness of soil herbivores.

Soil-fauna inoculation increased the biomass of the native plant community and thereby

decreased the relative biomass production of the alien species. The increased invasion

resistance due to soil fauna, however, tended ($p = 0.09$) to be stronger for plants growing

under well-watered conditions than under drought. Our multispecies experiment thus shows

that soil fauna might help native resident communities to resist alien plant invasions, but that

this effect might be diminished by drought.

Key words: alien-native competition, biological invasion, climate change, exotic, plant-

environment interactions, soil animals

INTRODUCTION

With the rapid globalization, more and more plant species have been introduced to new regions outside their native range [1-3]. Some of these alien plants have become invasive, which could decrease native species diversity, change nutrient cycles, and thereby affect ecosystem functions [4-8]. Forecasts show that the number of alien plant species per continent may increase on average by 18% from 2005 to 2050 [9], indicating that the impacts of plant invasions on ecosystems may become even more severe. However, as there are many potential drivers of invasions, there are many uncertainties about how invasions and their impacts will develop in the future [10]. In this regard, particularly the potential effects of ongoing climate change on plant invasions have garnered interest [11, 12].

In many places, climate change is characterized by more frequent and more intense drought events [13-16]. Droughts do not only reduce water availability to the plants, but also decreases the nutrient absorption [17, 18]. Consequently, drought might affect the competition between alien and native plants [17, 19, 20], and thereby affect invasion success of alien plants [17, 21, 22]. For example, Manea et al. found that drought could reduce biomass production of native grasses, and consequently enhance the establishment success of alien plants [19]. However, many other studies also found that invasive plants may suffer more from drought than native species, indicating that drought could also suppress alien plant invasion [12, 20, 23-26]. One of the reasons for the mixed findings could be that most studies only considered direct effects of drought on alien plant invasion [12, 24, 26, 27]. Much less is known about the indirect effects, i.e. how drought affects alien plant invasion via other trophic levels.

Soil fauna include several important belowground trophic levels, which are increasingly recognized to affect plant competition [28-31]. Some of them could enhance nutrient mineralization, and thereby increase plant nutrient uptake [28, 32-37]. Given that invasive

plants frequently respond more positively to nutrient enrichment than native plants [12, 38],

the higher nutrient availability caused by soil fauna may increase the invasion success. In

addition, soil fauna can also change alien-native competition via herbivory effects [39, 40].

The enemy-release hypothesis poses that alien plants are released from most of their native

enemies [41-44]. Following this logic, alien plants would be damaged less than natives by

herbivorous soil fauna, and therefore soil fauna would promote alien plant invasion. However,

until now very few studies have tested how soil fauna affects alien plant invasion into

resident communities [39, 40].

It has been found that indirect effects of altered biotic interactions due to climate change

on animal populations are more pronounced than their direct effects [45], which may also be

the case for plant populations. For example, empirical studies indicated that belowground

trophic interactions could alter plant responses to drought [46-49]. Consequently, it is likely

that drought might indirectly affect alien plant invasion in resident communities via effects on

soil fauna. Indeed, drought can reduce the abundance and diversity of soil fauna [47, 50-53].

Given that soil fauna, in particular some of the root herbivores might affect competition

between native and alien plants, indirect effects of drought on alien plant invasion in resident

community would occur via the cascading effect of drought suppression on soil fauna [39,

40]. However, it has not been tested yet how drought could indirectly, via effects on soil

fauna, affect plant invasion.

To test the direct and indirect effects (i.e. via soil fauna) of drought on alien plant

invasion into a native resident community, we performed a mesocosm-pot experiment. We

grew single plants of nine alien target species in a community of five native grassland species

under four combinations of two drought (well-watered *vs* drought) and two soil-fauna-

inoculation (with *vs* without) treatments. By comparing the absolute aboveground biomass

production of the alien target species as well as their biomass production relative to the

biomass production of the native competitors, we addressed the following specific questions:

(1) Does drought suppress the absolute and relative biomass of alien species? (2) Does the

presence of soil fauna promote or suppress the absolute and relative biomass of alien species?

(3) Does the presence of soil fauna change the effect of drought on the absolute and relative

biomass of alien species?

MATERIAL AND METHODS

Study species

To test the effects of drought, the presence of soil fauna, and their interaction on alien plant invasion in a native grassland community, we chose nine naturalized alien species as targets and five native species as competitors from the herbaceous flora of China (see Table S1). We classified the species as naturalized alien or native to China based on information in the book "The Checklist of The Naturalized Plants in China" [54] and the Flora of China database (www.efloras.org). To cover a wide taxonomic breadth, the nine alien species were chosen from eight genera of four families. The five native species, used to create the native community, included two forbs and three grasses that are all very common and do co-occur in many grasslands of China. Seeds of all species, except one, whose seeds were bought from a commercial seed company, were collected from natural populations growing in grasslands (Table S1).

Soil-fauna collection

To provide a live soil-fauna community as inoculum for the pot mesocosms, we collected soil fauna from a grassland site, where the five native species used for present study also occur in, at the Northeast Institute of Geography and Agricultural Ecology, Chinese Academy of Sciences (125°24'30"E, 43°59'49"N) on 21 July 2020. In the grassland, we removed aboveground plant materials from each of 100 sampling locations (30cm × 30cm), and then collected from each location a soil sample of 1-L (10cm × 10cm× 10cm) using a shovel. Each sampling location was at least 10m apart from the others. Then, we brought the 100 soil samples back to the laboratory, where we extracted the soil fauna communities of each soil sample separately using Berlese-Tullgren extractors without heating (https://en.wikipedia.org/wiki/Tullgren_funnel). In brief, we put all soil samples on top of 100 stainless steel soil-sieves with a 2mm mesh size, and then waited 12 days so that many of

the soil organisms would fall through the holes, via stainless steel funnels, into plastic bottles

filled with 50 ml soil-fauna-free peat moss (Pindstrup Plus, Pindstrup Mosebrug A/S,

Denmark). On 2 August 2020, we finished the soil-fauna-community collection. We then

randomly chose ten of the 100 soil-fauna communities for soil-fauna investigation (see Table

S2), and used the remaining 90 soil-fauna communities for inoculating of the soil in the

experimental pot mesocosms.

Experimental set-up

To compare the growth performance of alien plants when growing in a resident native

grassland community under different drought and soil-fauna-inoculation treatments, we did a

mesocosm-pot experiment in a greenhouse of the Northeast Institute of Geography and

Agricultural Ecology, Chinese Academy of Sciences. We grew each of the nine alien species

in the centre of a matrix of the native community under two water availabilities (well-watered

vs drought) and two soil-fauna-inoculation (with *vs* without) treatments.

From 15 May to 5 July 2020, we sowed the seeds of each species separately into plastic

trays (195 mm × 146 mm × 65 mm) filled with peat moss as substrate (Pindstrup Plus,

Pindstrup Mosebrug A/S, Denmark). As previous experiments had shown that the time

required for germination differs among the species, we sowed the species on different dates

(Table S1) to obtain similarly sized seedlings at the start of the experiment. On 3 August 2020,

we filled 180 2.5-L circular plastic pots (top diameter × bottom diameter × height: 18.5 ×

12.5 × 15 cm, Yancheng Tengle Plastics Co., Ltd, China) with the same substrate as used for

germination. To avoid nutrient limitation during plant growth, we mixed each pot with 5 g

slow-release fertilizer (Osmocote® Exact Standard, Everris International B.V., Geldermalsen,

The Netherlands). To create the native community, we selected similarly sized seedlings from

each of the five native species, and transplanted one seedling of each native species at equal

distances in a circle (diameter = 11 cm) around the centre of each pot. We then planted into

the centre of each pot one seedling of one of the alien species. For each of the nine alien

species, we had 20 pots.

After transplanting, we randomly assigned two pots of each alien species to each of ten

plastic cages (150 cm × 90 cm × 100 cm; Figure 1). The ten cages were covered with nylon

nets (mesh size: 0.15 mm × 0.15 mm) to keep out small soil fauna. We put a plastic dish

under each pot, and regularly watered the pots before starting the drought treatment to ensure

that none of the plants were water limited. On 3 August 2020, we inoculated each pot in five

of the ten cages (i.e. 90 pots) with 50 ml of the soil-fauna inoculum we had collected. As the

soil-fauna inoculum also introduced another 50 ml of peat moss into pots, we also added 50

ml of the same peat moss (free of soil fauna) to each pot of the remaining five cages as a

control. We assigned the cages with and without soil-fauna inoculations to alternating

positions that were at least 0.5 m apart from each other (Figure 1). On 30 August 2020 (i.e. 27

days after the start of the experiment), we started the drought treatment. One of the two pots

of each alien target species in each cage served as a control, which was watered regularly to

keep the substrate moist throughout the entire experiment, while the other pots did not

receive water unless the plants were wilted. We daily checked all pots of the drought

treatment, and when some plants of the community in a pot were wilted (i.e. had lost leaf

turgor), we supplied the pot with 50 ml of water. So, for each of the nine alien species, we

had five replicates of each of the four drought × soil-fauna-inoculation treatment

combinations, resulting in a total of 180 pots (9 alien species × 2 soil-fauna inoculations

[with *vs* without] × 2 drought treatments [well-watered *vs* drought] × 5 replicates [i.e. cages]).

On 28 September 2020, 10 weeks after the start of the drought treatments, we harvested

the experiment. For each pot, we separately harvested the aboveground biomass of the alien

target species and of the native community. As the roots of the plants were too much

intertwined, we could not harvest the belowground biomass. All aboveground biomass was

dried for at least 72 h at 65 °C, and then weighed. Based on the aboveground biomass of the

alien and native species, we calculated the biomass proportion of the alien target species (the

biomass of the alien target species / [biomass of the alien target species + biomass of the

native community]) as a proxy of the dominance of the alien target species (for a similar

approach, see Refs. 38, 55).

To test the effect of the drought treatment on soil fauna, we investigated the soil fauna of

drought and well-watered pots after plant harvest. We first randomly selected one pot of each

target alien species under drought or well-watered conditions, and then brought all soil of

these pots back to the laboratory. In total, we had 18 soil samples for soil-fauna extraction (9

alien species × 2 drought treatments [well-watered *vs* drought] × 1 replicate). Using the

Berlese-Tullgren extractors without heating, we extracted the soil fauna communities of each

soil sample separately, and obtained of the total soil-fauna abundance, soil-mites abundance,

soil-herbivore abundance and soil-herbivore richness for each sample. As most soil-fauna

species were mites (*c.* 70.0%) for our samples, and it was difficult to identify all of them to

species level, we do not have data of soil-fauna richness. We also did soil-fauna investigation

for five pots that were randomly selected from the pots without soil-fauna inoculation, and

found no soil fauna in these pots, indicating that our soil-fauna inoculation treatment and the

isolation imposed by the cages were effective.

Statistical analysis

To analyze the effects of drought, soil-fauna-inoculation treatments and their interactions

on performance of the alien plants in the native community, we fitted linear mixed-effects

models using the lme function of the 'nlme' package [56] in R 4.0.3 [57]. Aboveground

biomass production of the alien target species, the native competitor species and biomass

proportion of the alien target species in each pot (i.e. target aboveground biomass/total

aboveground biomass) were the response variables. To meet the assumption of normality,

biomass production of the alien target species and the native competitor species were natural-

log-transformed, and biomass proportion of the target species was logit-transformed. We

included drought treatment (i.e. well-watered *vs* drought), soil-fauna-inoculation treatment

(i.e. with *vs* without addition) and their interactions as fixed effects in all models.

To account for non-independence of individuals of the same alien plant species and for

phylogenetic non-independence of the species, we included identity of the target species

nested within family as random factors in all models. In addition to account for non-

independence of plants within the same cage, we also included cage identity as a random

factor in all model. As the homoscedasticity assumption was violated in all models, we also

included variance structures to model different variances per species or per cage (based on

model selection) using the "*varIdent*" function in the R package 'nlme' [56]. We used log-

likelihood ratio tests to assess significance of the fixed effects drought treatment, soil-fauna-

inoculation treatment and their interaction [58]. These tests were based on comparisons of

maximum-likelihood models with and without the terms of interest, and the variance

components were estimated using the restricted maximum-likelihood method of the full

model [58]. To test the effect of drought on total soil-fauna abundance, soil-mites abundance,

soil-herbivore abundance and soil-herbivore richness, we applied Kruskal-Wallis test using

the kruskal.test function of the 'stats' package in R 4.0.3 [57].

RESULTS

The specific response patterns of each alien species to the drought and soil-fauna-inoculation treatments were more or less similar with the general patterns of alien target species (Figure 2 & S1). Averaged across the nine alien target species, drought significantly decreased the aboveground biomass production of alien target species (-58.6%; Table 1; Figure 2a), and the native community (-51.5%; Table 1, Figure 2b). As the biomass of the aliens decreased more strongly in response to drought than the biomass of the natives did, the biomass proportion of the alien target species in each pot decreased (-11.6%; Table 1; Figure 2c). Inoculation with soil fauna had no significant effect on aboveground biomass of the alien target plants, but had a significant positive effect on aboveground biomass of the native community (+40.1%; Table 1; Figure 2b). Consequently, the aboveground biomass proportion of the alien target species was decrease in the presence of soil fauna (-41.9%; Table 1; Figure 2c). Moreover, as the native community benefitted more from the presence of soil fauna under well-watered conditions (+44.0%) than under drought conditions (+32.5%; Table 1; Figure 2b), the negative effect of soil-fauna inoculation on biomass proportion of the alien target species tended to be stronger under well-watered conditions (-49.7%) than under drought conditions (-32.0%; marginally significant S × D interaction in Table 1, $p = 0.09$; Figure 2c). In addition, we found that drought significantly decreased the total abundance of soil fauna (-57.4%; Figure 3a) and soil mites (-54.6%; Figure 3b), but did not affect soil-herbivore abundance and richness (Figure 3c & 3d).

 DISCUSSION

Our multispecies experiment found that drought limited the absolute and the relative

biomass production of the alien target plants. This means that drought suppressed the growth

performance and thus reduced the competitive ability of the alien species in the native

community. In addition, we found that the presence of soil-fauna communities benefited the

native community and resulted in a decreased biomass proportion of the alien species. In

other words, the presence of soil fauna promoted the resistance of the native community

against invasion by the alien species. Moreover, the suppressive effect of drought on biomass

proportion of the alien plants tended to disappear (although this effect was only marginally

significant; $p = 0.09$) in the presence of soil fauna. This suggests that the presence of soil-

fauna communities might negate the negative effect of drought on alien plant invasion into a

resident community.

While drought is well known to reduce plant performance overall [59-61], recent studies

found that growth and reproduction were more strongly affected for invasive than for native

plant species [12, 23, 26]. Our results are consistent with these previous findings (see also the

total biomass production per pot in Figure S2), and suggest that the native competitors were

more tolerant to drought than the invasive alien species [12, 20, 23-26]. On the other hand, it

could also indicate that the invasive alien plants took more advantage of the well-watered

conditions than the native species did. This would be in line with the idea that invasive plants

show higher phenotypic plasticity, and capitalize more strongly on benign conditions than

native plants do (i.e. the Master-of-some strategy *sensu* Ref. 62). In any case, the negative

effect of drought on the biomass proportion of the alien plants suggests that the competitive

balance between invasive alien plants and native plants could be changed by drought in

favour of the resident community. Another recent study, however, showed that alien species

that are not invasive yet could benefit more from drought relative to resident plants [22],

which could imply a turn-over in invasive alien species with ongoing climate change.

Inoculation with soil fauna significantly increased the biomass production of the native

community. It is often suggested that soil fauna, such as collembolans and mites in our study

(Table S2), could enhance soil nutrient mineralization and consequently nutrient absorption

of plants [32, 63]. As a consequence, soil fauna frequently have positive effects on plant

performance [63-65]. However, we found that soil-fauna inoculation had no statistically

significant effect on growth of the alien target plants. Therefore, it is unlikely that soil fauna

promoted native plant growth by increasing nutrient availability, as we would have expected

the invasive alien species to benefit from it too. To confirm this, we further measured the total

nitrogen and alkaline hydrolyzable nitrogen (i.e. *plant-available* nitrogen) of 36 soil samples

(20% of total pots) from the drought treatment. This showed that soil-fauna inoculation did

not significantly affect both types of soil nitrogen (Figure S3).

The most likely explanation for why soil-fauna inoculation did not affect alien plant

growth is that the invasive alien species may have been released from their native enemies in

the study area [66, 67]. In contrast, native plants were most likely attacked by the soil

herbivores. The fact that they produced more aboveground biomass might indicate that the

presence of soil fauna resulted in a re-allocation of biomass from belowground to

aboveground structures or that compensatory growth actually resulted in overcompensation

[68, 69]. Indeed, it has been previously shown that collembolan, at intermediate densities,

could enhance plant growth through herbivory-induced overcompensation [70]. Although this

could explain why the soil fauna suppressed the dominance (i.e. biomass proportion) of alien

target plants, future explicitly experiments testing such soil-herbivory-induced

overcompensation are needed. Nevertheless, while it has been shown before that soil fauna

affects the composition of plant communities [28, 30, 39], this is one of the first studies to

document the ability of the soil fauna to provide resistance against alien plant invasion.

Although numerous studies have shown that climate change could affect interactions of

plants with aboveground organisms at other trophic levels [51, 71-73], only few studies have

addressed how climate change may affect interactions of plants with belowground organisms

at other trophic levels [47, 49, 51, 74]. To the best of our knowledge, no studies have

addressed how climate change interacts with resident soil fauna to affect the competitions

between native and invasive plant species. Many previous studies found that drought could

decrease the abundance of soil fauna [50-52, 75, 76]. Our study supports this, and further

showed that drought mainly decreased the abundance of soil mites rather than soil herbivores.

Given that drought also decreased the growth performance of native plants, the constant soil-

herbivore abundance would lead to a higher damage intensity, and thus the overcompensation

decreased in response to drought. This could explain why the growth promotion of native

plants induced by soil fauna was stronger under well-watered condition than under drought.

Our present finding is also consistent with previous studies, showing that drought can hinder

insect-induced-overcompensation of plants [68, 77].

Interestingly, we found tentative evidence that the decrease in dominance of alien target

plant caused by drought was larger in the absence of soil fauna than in its presence. In other

words, the presence of soil fauna might buffer against the negative effects of drought on alien

plant invasion in resident communities. This is because soil fauna did not mediate the drought

effects on the growth of alien plants which were most likely released from their native

enemies. On the other hand, it amplified the negative effect of drought on the growth of

native plants. Our study thus indicates that previous pot experiments that did not include soil

fauna might have overestimated the effects of drought on alien plant invasion. Therefore,

future studies testing effects of climate change on alien plant invasion should also consider

the role of other belowground trophic levels.

CONCLUSIONS

The findings of our multi-species experiment are in line with results of previous studies that drought can inhibit alien plant invasion into a native resident community. However, to the best of our knowledge, we here show for the first time that the presence of soil fauna might help the resident plant community to resist alien plant invasions. This soil-fauna mediated resistance may be partly negated by drought. This implies that with ongoing climate change, and more frequent droughts, invasive plants might be more likely to overcome the resistance provided by soil fauna.

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AUTHOR CONTRIBUTIONS

YL conceived the idea and designed the experiment. HJ and LC performed the experiment.

HJ and YL analyzed the data. HJ and YL wrote the first draft of the manuscript, with further

inputs from MvK and LC.

DATA ACCESSIBILITY

Should the manuscript be accepted, the data supporting the results will be archived in Dryad

and the data DOI will be included at the end of the article.

REFERENCES

[1] Seebens, H., Blackburn, T.M., Dyer, E.E., Genovesi, P., Hulmeh, P.E., Jeschke, J.M., Pagadl, S., Pyšekm, P., van Kleuneno, M., Winterq, M., et al. 2018 Global rise in emerging alien species results from increased accessibility of new source pools. *Proc. Natl. Acad. Sci. USA* **115**, 2264-2273. (doi:10.1073/pnas.1719429115).

[2] van Kleunen, M., Dawson, W., Essl, F., Pergl, J., Winter, M., Weber, E., Kreft, H., Weigelt, P., Kartesz, J., Nishino, M., et al. 2015 Global exchange and accumulation of non-native plants. *Nature* **525**, 100-103. (doi:10.1038/nature14910).

[3] van Kleunen, M., Essl, F., Pergl, J., Brundu, G., Carboni, M., Dullinger, S., Early, R., Gonzalez-Moreno, P., Groom, Q.J., Hulme, P.E., et al. 2018 The changing role of ornamental horticulture in alien plant invasions. *Biol. Rev.* **93**, 1421-1437. (doi:10.1111/brv.12402).

[4] Linders, T.E.W., Schaffner, U., Eschen, R., Abebe, A., Choge, S.K., Nigatu, L., Mbaabu, P.R., Shiferaw, H. & Allan, E. 2019 Direct and indirect effects of invasive species: biodiversity loss is a major mechanism by which an invasive tree affects ecosystem functioning. *J. Ecol.* **107**, 2660-2672. (doi:10.1111/1365-2745.13268).

[5] Vilà, M., Espinar, J.L., Hejda, M., Hulme, P.E., Jarošík, V., Maron, J.L., Pergl, J., Schaffner, U., Sun, Y. & Pyšek, P. 2011 Ecological impacts of invasive alien plants: a meta‐analysis of their effects on species, communities and ecosystems. *Ecol. Lett.* **14**, 702-708. (doi:10.1111/j.1461-0248.2011.01628.x).

[6] Pyšek, P., Jarošík, V., Hulme, P.E., Pergl, J., Hejda, M., Schaffner, U. & Vilà, M. 2012 A global assessment of invasive plant impacts on resident species, communities and ecosystems: the interaction of impact measures, invading species' traits and environment. *Glob. Change Biol.* **18**, 1725-1737. (doi:10.1111/j.1365-2486.2011.02636.x).

[7] Hejda, M., Pyšek, P. & Jarošík, V. 2009 Impact of invasive plants on the species richness, diversity and composition of invaded communities. *J. Ecol.* **97**, 393-403. (doi:10.1111/j.1365-

2745.2009.01480.x).

[8] Pyšek, P., Hulme, P.E., Simberloff, D., Bacher, S., Blackburn, T.M., Carlton, J.T., Dawson,

W., Essl, F., Foxcroft, L.C., Genovesi, P., et al. 2020 Scientists' warning on invasive alien

species. *Biol. Rev.* **95**, 1511-1534. (doi:10.1111/brv.12627).

[9] Seebens, H., Bacher, S., Blackburn, T.M., Capinha, C., Dawson, W., Dullinger, S.,

Genovesi, P., Hulme, P.E., van Kleunen, M., Kuhn, I., et al. 2020 Projecting the continental

accumulation of alien species through to 2050. *Glob. Change Biol.* **00**, 1-13.

(doi:10.1111/gcb.15333).

[10] Essl, F., Lenzner, B., Bacher, S., Bailey, S., Capinha, C., Daehler, C., Dullinger, S.,

Genovesi, P., Hui, C., Hulme, P.E., et al. 2020 Drivers of future alien species impacts: An

expert based assessment. *Glob. Change Biol.* **26**, 4880-4893. (doi:10.1111/gcb.15199).

[11] Bartz, R. & Kowarik, I. 2019 Assessing the environmental impacts of invasive alien

plants: a review of assessment approaches. *NeoBiota* **43**, 69-99.

(doi:10.3897/neobiota.43.30122).

[12] Liu, Y.J., Oduor, A.M.O., Zhang, Z., Manea, A., Tooth, I.M., Leishman, M.R., Xu, X.L.

& van Kleunen, M. 2017 Do invasive alien plants benefit more from global environmental

change than native plants? *Glob. Change Biol.* **23**, 3363-3370. (doi:10.1111/gcb.13579).

[13] Dai, A. 2013 Increasing drought under global warming in observations and models. *Nat.*

Clim. Chang. **3**, 52-58. (doi:10.1038/NCLIMATE1633).

[14] Spinoni, J., Vogt, J.V., Naumann, G., Barbosa, P. & Dosio, A. 2018 Will drought events

become more frequent and severe in Europe? *Int. J. Climatol.* **38**, 1718-1736.

(doi:10.1002/joc.5291).

[15] Diffenbaugh, N.S., Swain, D.L. & Touma, D. 2015 Anthropogenic warming has

increased drought risk in California. *Proc. Natl. Acad. Sci. USA* **112**, 3931–3936.

(doi:10.1073/pnas.1422385112).

[16] Yoon, J.H., Wang, S.Y.S., Gillies, R.R., Kravitz, B., Hipps, L. & Rasch, P.J. 2015

Increasing water cycle extremes in California and in relation to ENSO cycle under global

warming. *Nat. Commun.* **6**, 8657. (doi:10.1038/ncomms9657).

[17] Bueno, A., Pritsch, K. & Simon, J. 2020 Responses of native and invasive woody

seedlings to combined competition and drought are species-specific. *Tree Physiol.* **41**, 343-

357. (doi:10.1093/treephys/tpaa134).

[18] da Silva, E.C., Nogueira, R.J.M.C., da Silva, M.A. & de Albuquerque, M.B. 2011

Drought stress and plant nutrition. *Plant stress* **5**, 32-41.

[19] Manea, A., Sloane, D.R. & Leishman, M.R. 2016 Reductions in native grass biomass

associated with drought facilitates the invasion of an exotic grass into a model grassland

system. *Oecologia* **181**, 175-183. (doi:10.1007/s00442-016-3553-1).

[20] Werner, C., Zumkier, U., Beyschlag, W. & Máguas, C. 2010 High competitiveness of a

resource demanding invasive acacia under low resource supply. *Plant Ecol.* **206**, 83-96.

(doi:10.1007/s11258-009-9625-0).

[21] Ahanger, M.A., Morad□Talab, N., Abd-Allah, E.F., Ahmad, P. & Hajiboland, R. 2016

Plant growth under drought stress: significance of mineral nutrients. *Water Stress and Crop*

Plants: A Sustainable Approach **2**, 649-668. (doi:10.1002/9781119054450.ch37).

[22] Haeuser, E., Dawson, W. & van Kleunen, M. 2019 Introduced garden plants are strong

competitors of native and alien residents under simulated climate change. *J. Ecol.* **107**, 1328-

1342. (doi:10.1111/1365-2745.13101).

[23] Kelso, M.A., Wigginton, R.D. & Grosholz, E.D. 2020 Nutrients mitigate the impacts of

extreme drought on plant invasions. *Ecology* **101**, e02980. (doi:10.1002/ecy.2980).

[24] Copeland, S.M., Harrison, S.P., Latimer, A.M., Damschen, E.I., Eskelinen, A.M.,

Fernandez□Going, B., Spasojevic, M.J., Anacker, B.L. & Thorne, J.H. 2016 Ecological

effects of extreme drought on Californian herbaceous plant communities. *Ecol. Monogr.* **86**,

295-311. (doi:10.1002/ecm.1218).

[25] LaForgia, M.L., Spasojevic, M.J., Case, E.J., Latimer, A.M. & Harrison, S.P. 2018 Seed

banks of native forbs, but not exotic grasses, increase during extreme drought. *Ecology* **99**,

896-903. (doi:10.1002/ecy.2160).

[26] Valliere, J.M., Escobedo, E.B., Bucciarelli, G.M., Sharifi, M.R. & Rundel, P.W. 2019

Invasive annuals respond more negatively to drought than native species. *New Phytol.* **223**,

1647-1656. (doi:10.1111/nph.15865).

[27] Pintó-Marijuan, M., Cotado, A., Fleta-Soriano, E. & Munné-Bosch, S. 2017 Drought

stress memory in the photosynthetic mechanisms of an invasive CAM species*, Aptenia*

cordifolia. *Photosynth. Res.* **131**, 241-253. (doi:10.1007/s11120-016-0313-3).

[28] Eisenhauer, N., Sabais, A.C.W., Schonert, F. & Scheu, S. 2010 Soil arthropods

beneficially rather than detrimentally impact plant performance in experimental grassland

systems of different diversity. *Soil Biol. Biochem.* **42**, 1418-1424.

(doi:10.1016/j.soilbio.2010.05.001).

[29] Bardgett, R.D., Bowman, W.D., Kaufmann, R. & Schmidt, S.K. 2005 A temporal

approach to linking aboveground and belowground ecology. *Trends Ecol. Evol.* **20**, 634-641.

(doi:10.1016/j.tree.2005.08.005).

[30] Wardle, D.A., Bardgett, R.D., Klironomos, J.N., Setälä, H., van der Putten, W.H. & Wall,

D.H. 2004 Ecological linkages between aboveground and belowground biota. *Science* **304**,

1629-1633. (doi:10.1126/science.1094875).

[31] Eisenhauer, N. & Scheu, S. 2008 Earthworms as drivers of the competition between

grasses and legumes. *Soil Biol. Biochem.* **40**, 2650-2659. (doi:10.1016/j.soilbio.2008.07.010).

[32] Bardgett, R.D. & Chan, K.F. 1999 Experimental evidence that soil fauna enhance

nutrient mineralization and plant nutrient uptake in montane grassland ecosystems. *Soil Biol.*

Biochem. **31**, 1007-1014. (doi:10.1016/S0038-0717(99)00014-0).

[33] Barot, S., Ugolini, A. & Brikci, F.B. 2007 Nutrient cycling efficiency explains the long-

term effect of ecosystem engineers on primary production. *Funct. Ecol.* **21**, 1-10.

(doi:10.1111/j.1365-2435.2006.01225.x).

[34] Marinissen, J.C.Y. & de Ruiter, P.C. 1993 Contribution of earthworms to carbon and

nitrogen cycling in agro-ecosystems. *Agric. Ecosyst. Environ.* **47**, 59-74. (doi:10.1016/0167-

8809(93)90136-D).

[35] Cole, L., Dromph, K.M., Boaglio, V. & Bardgett, R.D. 2004 Effect of density and

species richness of soil mesofauna on nutrient mineralisation and plant growth. *Biol. Fertil.*

Soils **39**, 337–343. (doi:10.1007/s00374-003-0702-6).

[36] Smith, V.R. & Steenkamp, M. 1992 Soil macrofauna and nitrogen on a sub-Antarctic

island. *Oecologia* **92**, 201-206. (doi:10.1007/BF00317365).

[37] Xu, G.L., Mo, J.G., Zhou, G.Y. & Peng, S.L. 2003 Relationship of soil fauna and N

cycling and its response to N deposition. *Acta Ecologica Sinica* **23**, 2453-2463.

(doi:10.3321/j.issn:1000-0933.2003.11.030).

[38] Liu, Y.J., Liu, M., Xu, X.L., Tian, Y.Q., Zhang, Z. & van Kleunen, M. 2018 The effects

of changes in water and nitrogen availability on alien plant invasion into a stand of a native

grassland species. *Oecologia* **188**, 441-450. (doi:10.1007/s00442-018-4216-1).

[39] Bonkowski, M., Villenave, C. & Griffiths, B. 2009 Rhizosphere fauna: the functional

and structural diversity of intimate interactions of soil fauna with plant roots. *Plant Soil* **321**,

213-233. (doi:10.1007/s11104-009-0013-2).

[40] Korell, L., Schädler, M., Brandl, R., Schreiter, S. & Auge, H. 2019 Release from above-

and belowground insect herbivory mediates invasion dynamics and impact of an exotic plant.

Plants **8**, 544. (doi:10.3390/plants8120544).

[41] Keane, R.M. & Crawley, M.J. 2002 Exotic plant invasions and the enemy release

hypothesis. *Trends Ecol. Evol.* **17**, 164-170. (doi:10.1016/S0169-5347(02)02499-0).

[42] Liu, H. & Stiling, P. 2006 Testing the enemy release hypothesis: a review and meta-

analysis. *Biol. Invasions* **8**, 1535-1545. (doi:10.1007/s10530-005-5845-y).

[43] Mitchell, C.E. & Power, A.G. 2003 Release of invasive plants from fungal and viral

pathogens. *Nature* **421**, 625-627. (doi:10.1038/nature01317).

[44] Vilà, M., Maron, J.L. & Marco, L. 2005 Evidence for the enemy release hypothesis in

Hypericum perforatum. Oecologia **142**, 474-479. (doi:10.1007/s00442-004-1731-z).

[45] Ockendon, N., Baker, D.J., Carr, J.A., White, E.C., Almond, R.E.A., Amano, T., Bertram,

E., Bradbury, R.B., Bradley, C., Butchart, S.H., et al. 2014 Mechanisms underpinning

climatic impacts on natural populations: altered species interactions are more important than

direct effects. *Glob. Change Biol.* **20**, 2221-2229. (doi:10.1111/gcb.12559).

[46] Franco, A.L.C., Gherardi, L.A., de Tomasel, C.M., Andriuzzi, W.S., Ankrom, K.E., Bach,

E.M., Guan, P., Sala, O.E. & Wall, D.H. 2020 Root herbivory controls the effects of water

availability on the partitioning between above□and below□ground grass biomass. *Funct.*

Ecol. **34**, 2403-2410. (doi:10.1111/1365-2435.13661).

[47] Guyer, A., Hibbard, B.E., Holzkämper, A., Erb, M. & Robert, C.A.M. 2018 Influence of

drought on plant performance through changes in belowground tritrophic interactions. *Ecol.*

Evol. **8**, 6756-6765. (doi:10.1002/ece3.4183).

[48] Erb, M., Köllner, T.G., Degenhardt, J., Zwahlen, C., Hibbard, B.E. & Turlings, T.C.J.

2011 The role of abscisic acid and water stress in root herbivore-induced leaf resistance. *New*

Phytol. **189**, 308-320. (doi:10.1111/j.1469-8137.2010.03450.x).

[49] Wilschut, R.A. & van Kleunen, M. 2021 Drought alters plant□soil feedback effects on

biomass allocation but not on plant performance. *Plant Soil* **462**, 285-296.

(doi:10.1007/s11104-021-04861-9).

[50] Aupic-Samain, A., Baldy, V., Delcourt, N., Krogh, P.H., Gauquelin, T., Fernandez, C. &

Santonja, M. 2020 Water availability rather than temperature control soil fauna community

structure and prey☐predator interactions. *Funct. Ecol.* (doi:10.1111/1365-2435.13745).

[51] Eisenhauer, N., Cesarz, S., Koller, R., Worm, K. & Reich, P.B. 2012 Global change

belowground: impacts of elevated CO_2, nitrogen, and summer drought on soil food webs and

biodiversity. *Glob. Change Biol.* **18**, 435-447. (doi:10.1111/j.1365-2486.2011.02555.x).

[52] Makkonen, M., Berg, M.P., Van Hal, J.R., Callaghan, T.V., Press, M.C. & Aerts, R. 2011

Traits explain the responses of a sub-arctic collembola community to climate manipulation.

Soil Biol. Biochem. **43**, 377-384. (doi:10.1016/j.soilbio.2010.11.004).

[53] Wilschut, R.A. & Geisen, S. 2021 Nematodes as drivers of plant performance in natural

systems. *Trends Plant Sci.* **26**, 237-247. (doi:10.1016/j.tplants.2020.10.006).

[54] Yan, X.L., Wang, Z.H. & Ma, J.S. 2019 *The checklist of the naturalized plants in china*.

Shanghai, China, Shanghai Scientific and Technical Publishers.

[55] Parepa, M., Fischer, M. & Bossdorf, O. 2013 Environmental variability promotes plant

invasion. *Nat. Commun.* **4**, 1604. (doi:10.1038/ncomms2632).

[56] Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D. & R Core Team. 2020 nlme: linear and

nonlinear mixed effects models. (R package version 3.1-151.

[57] R Core Team. 2020 R: A language and environment for statistical computing. In *R*

Foundation for Statistical Computing (Vienna, Austria.

[58] Zuur, A.F., Ieno, E.N., Walker, N.J., Saveliev, A.A. & Smith, G.M. 2009 *Mixed effects*

models and extensions in ecology with R. New York, USA, Springer.

[59] Zlatev, Z. & Lidon, F.C. 2012 An overview on drought induced changes in plant growth,

water relations and photosynthesis. *Emir. J. Food Agric.* **24**, 57-72. (doi:10.1111/j.1439-

0426.2005.00664.x).

[60] Beierkuhnlein, C., Thiel, D., Jentsch, A., Willner, E. & Kreyling, J. 2011 Ecotypes of

European grass species respond differently to warming and extreme drought. *J. Ecol.* **99**,

703-713. (doi:10.1111/j.1365-2745.2011.01809.x).

[61] Gupta, A., Rico-Medina, A. & Caño-Delgado, A.I. 2020 The physiology of plant

responses to drought. *Science* **368**, 266-269. (doi:10.1126/science.aaz7614).

[62] Richards, C.L., Bossdorf, O., Muth, N.Z., Gurevitch, J. & Pigliucci, M. 2006 Jack of all

trades, master of some? On the role of phenotypic plasticity in plant invasions. *Ecol. Lett.* **9**,

981-993. (doi:10.1111/j.1461-0248.2006.00950.x).

[63] Lussenhop, J. & Bassirirad, H. 2005 Collembola effects on plant mass and nitrogen

acquisition by ash seedlings (*Fraxinus pennsylvanica*). *Soil Biol. Biochem.* **37**, 645-650.

(doi:10.1016/j.soilbio.2004.08.021).

[64] Mehring, A.S. & Levin, L.A. 2015 Potential roles of soil fauna in improving the

efficiency of rain gardens used as natural stormwater treatment systems. *J. Appl. Ecol.* **52**,

1445-1454. (doi:10.1111/1365-2664.12525).

[65] Partsch, S., Milcu, A. & Scheu, S. 2006 Decomposers (lumbricidae, collembola) affect

plant performance in model grasslands of different diversity. *Ecology* **87**, 2548-2558.

(doi:10.1890/0012-9658(2006)87[2548:DLCAPP]2.0.CO;2).

[66] Blossey, B. & Notzold, R. 1995 Evolution of increased competitive ability in invasive

nonindigenous plants: a hypothesis. *J. Ecol.* **83**, 887-889. (doi:10.2307/2261425).

[67] Chun, Y.J., van Kleunen, M. & Dawson, W. 2010 The role of enemy release, tolerance

and resistance in plant invasions: linking damage to performance. *Ecol. Lett.* **13**, 937-946.

(doi:10.1111/j.1461-0248.2010.01498.x).

[68] Ramula, S., Paige, K.N., Lennartsson, T. & Tuomi, J. 2019 Overcompensation: a 30-year

perspective. *Ecology* **100**, e02667. (doi:10.1002/ecy.2667).

[69] Garcia, L.C. & Eubanks, M.D. 2019 Overcompensation for insect herbivory: a review

and meta-analysis of the evidence. *Ecology* **100**, e02585. (doi:10.1002/ecy.2585).

[70] Gange, A. 2000 Arbuscular mycorrhizal fungi, collembola and plant growth. *Trends Ecol.*

Evol. **15**, 369-372. (doi:10.1016/S0169-5347(00)01940-6).

[71] de Sassi, C. & Tylianakis, J.M. 2012 Climate change disproportionately increases

herbivore over plant or parasitoid biomass. *PLoS One* **7**, e40557.

(doi:10.1371/journal.pone.0040557).

[72] Meza-Lopez, M.M. & Siemann, E. 2020 Warming alone increased exotic snail

reproduction and together with eutrophication influenced snail growth in native wetlands but

did not impact plants. *Sci. Total Environ.* **704**, 135271. (doi:10.1016/j.scitotenv.2019.135271).

[73] Nooten, S.S. & Hughes, L. 2014 Potential impacts of climate change on patterns of

insect herbivory on understorey plant species: a transplant experiment. *Austral Ecol.* **39**, 668-

676. (doi:10.1111/aec.12129).

[74] Classen, A.T., Sundqvist, M.K., Henning, J.A., Newman, G.S., Moore, J.A.M., Cregger,

M.A., Moorhead, L.C. & Patterson, C.M. 2015 Direct and indirect effects of climate change

on soil microbial and soil microbial-plant interactions: What lies ahead? *Ecosphere* **6**, 130.

(doi:10.1890/es15-00217.1).

[75] Kardol, P., Cregger, M.A., Campany, C.E. & Classen, A.T. 2010 Soil ecosystem

functioning under climate change: plant species and community effects. *Ecology* **91**, 767-781.

(doi:10.1890/09-0135.1).

[76] Lindberg, N. 2003 *Soil fauna and global change*. Niklas Lindberg, Uppsala, Sweden.

[77] Levine, M.T. & Paige, K.N. 2004 Direct and indirect effects of drought on compensation

following herbivory in Scarlet gilia. *Ecology* **85**, 3185-3191. (doi:10.1890/03-0748).

Table 1

Table 1 Results of linear mixed-effects models testing the effects of drought (well-watered *vs* drought), soil fauna (with *vs* without) and their

interactions on aboveground biomass production of the alien target plants, native competitor species, and biomass proportion of the alien target

species in each pot. Significant effects ($p < 0.05$) are in bold, and marginal significant effects ($0.05 < p < 0.1$) are underlined.

		Aboveground biomass production of the alien target species (natural-log-transformed)			Aboveground biomass production of the native competitor species (natural-log-transformed)			Aboveground biomass proportion of the target species in each pot (logit-transformed)	
Fixed effects	**Df**	χ^2	***p***	**Df**	χ^2	***p***	**Df**	χ^2	***p***
Soil fauna (S)	1	2.4990	0.1139	1	4.7722	**0.0289**	1	7.0803	**0.0078**
Drought (D)	1	63.8413	**<0.0001**	1	164.1043	**<0.0001**	1	5.6339	**0.0176**
S × D	1	0.1512	0.6974	1	5.6304	**0.0177**	1	2.8297	<u>0.0925</u>
Random effects		***SD***			***SD***			***SD***	
Family		0.004			0.002			0.004	
Species		0.970*			0.086			1.057*	
Cage		0.004			0.267*			0.187	
Residual		0.586			0.126			0.555	
	Marginal R^2		**Conditional R^2**	**Marginal R^2**		**Conditional R^2**	**Marginal R^2**		**Conditional R^2**
R^2 of the model	0.138		0.770	0.641		0.940	0.063		0.802

* Standard deviations for individual alien species or individual cage random effects for the saturated model are found in Table S3.

Figure 1

Figure 1 Graphical illustration of the experimental design.

Figure 2

Figure 2 Mean values (± SE) of aboveground biomass of the alien target species (a),

aboveground biomass production of the native competitor species (b), and biomass

proportion of the alien target species in each pot (c) under different drought (well-watered *vs*

drought) and soil-fauna-inoculation (with *vs* without) treatments. Significant parameters (i.e.

'S' [soil fauna], 'D' [drought] and 'S × D') are indicted with asterisks (*), the marginal

significant ones are indicated with daggers (†), and the non-significant ones are indicated

with "ns".

Figure 3

Figure 3 Total abundance of soil fauna (a), soil mites (b) and soil herbivores (c), as well as

the soil herbivore richness (c) under drought and well-watered conditions. The nine different

color points represent samples of the nine pots with different alien target species.

 SUPPORTING INFORMATION

571 **Table S1** Details of the study species used in the experiment.

Species	Family	Status	Species native distribution*	Seed sources	Sowing data
Bidens frondosa L.	Asteraceae	Alien	North America	Zhejiang, China	26/6/2020
Bidens pilosa L.	Asteraceae	Alien	America	Zhejiang, China	26/6/2020
Erigeron canadensis L.	Asteraceae	Alien	North America	Jilin, China	26/6/2020
Hibiscus trionum L.	Malvaceae	Alien	Africa, Eurasia, New Zealand and Australia	Jilin, China	5/7/2020
Lolium perenne L.	Poaceae	Alien	North Africa, the Azores, the Madeira Islands, the Canary Islands, Europe, West Asia, South Russia, Afghanistan and the Indian subcontinent	Greenwood flower seed industry, China	26/6/2020
Medicago sativa L.	Leguminosae	Alien	Asia, South Europe	Nei Mongol, China	26/6/2020
Paspalum notatum Flüggé	Poaceae	Alien	Mexico, the Caribbarean and South America	Green alliance ecological enterprise shop, China	15/5/2020
Solidago canadensis L.	Asteraceae	Alien	North America	Zhejiang, China	26/6/2020
Xanthium strumarium L.	Asteraceae	Alien	North America, the West Indies	Nei Mongol, China	26/6/2020
Chloris virgata Sw.	Poaceae	Native	China, Afghanistan, Bhutan, India, Myanmar, Nepal, Pakistan, Africa, America, Southwest Asia, Australia, Pacific Islands	Nei Mongol, China	26/6/2020
Digitaria sanguinalis (L.) Scop.	Poaceae	Native	China	Jilin, China	26/6/2020
Echinochloa crus-galli (L.) P.Beauv.	Poaceae	Native	China	Jilin, China	26/6/2020
Lepidium apetalum Willd.	Brassicaceae	Native	China, India, Japan, Kazakhstan, Korea, Mongolia, Nepal, Pakistan	Jilin, China	26/6/2020
Plantago asiatica L.	Plantaginaceae	Native	China, Bangladesh, Bhutan, India, Indonesia, Japan, Korea, Malaysia, Nepal, Sri Lanka	Nei Mongol, China	26/6/2020

572 * Species native distribution of each species was extracted from the book "The Checklist of The Naturalized Plants in China" and the Flora of
573 China database (www.efloras.org).
574

575   **Table S2** Soil-fauna taxa found in the soil used to inoculate the pot mesocosms.

Species	Taxonomy groups
Allonychiurus songi	Collembola
Entomobrya koreana	Collembola
Folsomia sp1	Collembola
Folsomides sp1	Collembola
Homidia phjongiangica	Collembola
Hypogastrura sp1	Collembola
Isotomiella minor	Collembola
Isotomodes sp1	Collembola
Orchesellides sinensis	Collembola
Proisotoma sp1	Collembola
Sminthurides sp1	Collembola
Thalassaphorura macrospinata	Collembola
Acrotritia ardua (Koch, 1841)	Mites
Antennoseius alexandrovi Bregetova, 1977	Mites
Galumna changchunensis Wen, 1987	Mites
Mesostigmata sp.1	Mites
Mesostigmata sp.2	Mites
Mesostigmata sp.3	Mites
Mesostigmata sp.4	Mites
Mesostigmata sp.5	Mites
Mesostigmata sp.6	Mites
Mesostigmata sp.7	Mites
mites nymph	Mites
Nothrus anauniensis Canestrini & Fanzago, 1877	Mites
Oppiella nova (Oudemans, 1902)	Mites
Prostigamata sp.1	Mites
Prostigamata sp.2	Mites
Scheloribates fimbriatus Thor, 1930	Mites
Tectocepheus velatus (Michael, 1880)	Mites
Trichogalumna nipponica (Aoki, 1966)	Mites
Veigaia slonovi Bregetova, 1961	Mites
Zygoribatula exilis (Nicolet, 1855)	Mites
Zygoribatula truncata (Aoki, 1961)	Mites
Araneae	Others
Chilopoda	Others
Coleoptera adult	Others
Coleoptera larvae	Others
Diptera larvae	Others
Formicidae	Others
Homoptera larvae	Others
Phytophthira	Others
Protura	Others
Scutigerellidae	Others

577 **Table S3** Standard deviations for individual alien species or individual cage random effects for metrics analyzed with models with a Gaussian

578 error distribution. The standard deviations given refer to the first species and cage respectively. For each species and cage, these should be

579 multiplied by the multiplication factors. The names of the alien species in the table are abbreviated using the first letter of the genus and species

580 epithet.

Metric	Species Standard Deviation	Multiplication factor standard deviation								
		LP	EC	HT	PN	SC	BF	BP	MS	XS
Aboveground biomass production of the alien target species	0.970	1.000	0.778	0.626	0.820	1.078	1.935	2.600	1.890	1.778
Aboveground biomass proportion of the target species in each pot	1.057	1.000	0.879	0.920	0.942	1.179	2.164	2.958	1.905	1.815

582
583

Metric	Cages Standard Deviation	Multiplication factor for standard deviation								
		Cage1	Cage2	Cage3	Cage4	Cage5	Cage6	Cage7	Cage8	Cage9	Cage10
Aboveground biomass production of the native competitor species	0.267	1.000	2.461	2.603	3.751	1.531	1.725	2.090	1.302	3.503	2.590

Figure S1

Figure S1 Mean values (± SE) of aboveground biomass of the specific alien target species (a)

and biomass proportion of the specific alien target species in each pot (b) under different

drought (well-watered vs drought) and soil-fauna-inoculation (with vs without) treatments.

Figure S2

Figure S2 Mean values (± SE) of the total aboveground biomass production per pot (i.e. alien

+ native) under different drought (well-watered *vs* drought) and soil-fauna-inoculation (with

vs without) treatments.

Figure S3

Figure S3 Mean values (± SE) of the soil total nitrogen (a) and soil alkaline hydrolyzable

nitrogen (b) in the pots with and without soil-fauna-inoculation treatments.