- 1 Selective logging shows no impact on the dietary breadth of the fawn leaf-nosed bat
- 2 (*Hipposideros cervinus*)
- 3 David R. Hemprich-Bennett^{1,2}, Victoria A. Kemp¹, Joshua Blackman¹, Owen T. Lewis²,
- 4 Matthew J. Struebig³, Henry Bernard⁴, Stephen J. Rossiter¹, Elizabeth L. Clare^{1,5}
- 5
- 6 ¹School of Biological and Chemical Sciences, Queen Mary University of London, Mile
- 7 End Road, London, UK E1 4NS
- 8 ²Department of Zoology, University of Oxford, 11a Mansfield Road, Oxford, UK, OX1
- 9 3SZ
- ³Durrell Institute of Conservation and Ecology, University of Kent, Canterbury, Kent,
 UK, CT2 7NZ
- 12 ⁴Institute for Tropical Biology and Conservation, Universiti Malaysia Sabah, Kota
- 13 Kinabalu, Sabah, Malaysia
- ⁵Department of Biology, York University, 4700 Keele Street, Toronto ON M3J 1P3
- 15 Correspondence: David Hemprich-Bennett: Department of Zoology, University of
- 16 Oxford, 11a Mansfield Road, Oxford, UK, OX1 3SZ, hemprich.bennett@gmail.com

17 Abstract

18 Logging activities degrade forest habitats across large areas of the tropics, but the impacts 19 on trophic interactions that underpin forest ecosystems are poorly understood. DNA 20 metabarcoding provides an invaluable tool to investigate such interactions, allowing 21 analysis at a far greater scale and resolution than has previously been possible. We 22 analysed the diet of the insectivorous fawn leaf-nosed bat *Hipposideros cervinus* across 23 a forest disturbance gradient in Borneo, using a dataset of ecological interactions from an 24 unprecedented number of bat-derived faecal samples. Bats predominantly consumed 25 insects from the orders Lepidoptera, Blattodea, Diptera and Coleoptera, and the 26 taxonomic composition of their diet remained relatively consistent across sites regardless 27 of logging disturbance. There was little difference in the richness of prey consumed in 28 each logging treatment, indicating potential resilience of this species to habitat 29 degradation. In fact, bats consumed a high richness of prey items, and intensive sampling 30 is needed to reliably compare feeding ecology over multiple sites regardless of the 31 bioinformatic procedures used.

32 Introduction

Logging is a common form of anthropogenic disturbance in forests, with over 90% of those in the tropics logged to some degree (Asner et al., 2009). Most logging undertaken in tropical forests is selective, which tends to favour removal of the largest, and highestquality trees. While this disturbance can have lasting effects on forest structure (Milodowski et al., 2021) selective logging tends to be much less destructive than clearfelling.

39 Forest modification through logging is especially pronounced on the island of Borneo, 40 which has lost half of its forest area since 1940 (Gaveau et al., 2014) and 62% of the 41 remaining forest is classified as 'degraded' or 'seriously degraded' (Gaveau et al., 2016). 42 Most studies of the impact this has on biodiversity have focussed on species composition 43 (e.g. Edwards et al., 2011; Slade et al., 2011; Kitching et al., 2013; Struebig et al., 2013; 44 Deere et al., 2018; Hayward et al., 2021). These often subtle changes to ecological 45 communities can result in changes to ecosystem functioning (Ewers et al., 2015) and the 46 structure of trophic networks (Hemprich-Bennett et al., 2020), indicating that selective 47 logging may alter resilience to future perturbations. Understanding the ecological shifts 48 that take place in degraded forest is of great importance for conservation, especially given 49 the vast scale at which forest is managed for timber extraction globally.

50

Animal diet can differ between individuals of a species depending on numerous intrinsic and environmental factors. In insectivorous bats for example, inter-individual variation in diet appears to correlate with multiple factors, including wing morphology (Oliveira et al., 2020), sex (Burgar et al., 2014), reproductive condition (Czenze et al., 2018), season (Andriollo et al., 2019; Kolkert et al., 2020), geographic location (Czenze et al., 2018;

Vallejo et al., 2019), and habitat (Aizpurua et al., 2018; Hemprich-Bennett et al., 2020;
Tournayre et al., 2021). Such variation is of interest when because intraspecific
differences in the feeding behaviour of consumers can alter the abundance, community
composition and ecological functioning of their prey (Des Roches et al., 2018).

60

61 Intraspecific variation in diet is also an important consideration for research design. The 62 analysis of diet in a highly generalist species requires many observations to obtain a 63 representative sample. This can be especially true when studying the dietary ecology of 64 insectivorous bats through metabarcoding, as the technique gives an unprecedented level 65 of taxonomic resolution (Clare et al., 2009), highlighting variation which would not have 66 been apparent with morphological study. Inter-individual variation in bat diet is however 67 often obscured by the use of samples collected from underneath roosts, where numerous 68 bats are defecating (hereafter 'roost-sourced' samples) (Clare et al., 2014; Andriollo et 69 al., 2019) and samples cannot be linked to an individual. Obtaining faecal samples from 70 individually identifiable animals (hereafter 'individual-sourced' samples) is labour-71 intensive due to the large trapping effort required, and so while many studies have used 72 individual-sourced samples (e.g. Czenze et al., 2018; Oliveira et al., 2020), their sample 73 sizes tend to be small. Mata et al (2018) used a dataset of individual-sourced samples to 74 analyse the importance of technical and biological replication on the dietary completeness 75 of Tadarida teniotis and reiterated the common rule of thumb that 20-50 such samples 76 per species is preferable, but stressed that higher sample sizes may be required for bat 77 species with greater dietary richness or intraspecific variation. The issue of sample size 78 is further complicated in networks generated from metabarcoding data because of 79 methodological considerations such as PCR primer bias and stochasticity (Alberdi et al.,

80 2018), and the influence of bioinformatic choices on the final data analysed (Hemprich-

- 81 Bennett et al., 2021).
- 82

83 Here we use an unprecedented number of individually-sourced insectivorous bat faecal 84 samples to test the hypothesis that selective logging alters the taxonomic composition and 85 species richness of bats' diet. We also assess how sample size and bioinformatic 86 parameters affect our inferences of insectivorous diet when using data derived from 87 metabarcoding. Our evaluation focuses on the fawn leaf-nosed bat, Hipposideros 88 cervinus - a cave-roosting insectivorous bat found throughout much of maritime 89 Southeast Asia to northeastern Australia. Using high-duty cycle (HDC) echolocation, it 90 is thought to use Doppler-shift compensation to detect the wingbeats of fluttering of prey 91 such as moths (Bell and Fenton, 1984) against a cluttered backdrop (Schnitzler and Kalko, 92 2001; Lazure and Fenton, 2011). Although some bat species are negatively affected by 93 logging, *H. cervinus* remains a dominant species in both old growth and logged forest in 94 Borneo (Struebig et al., 2013; Hemprich-Bennett et al., 2020). It is not known whether 95 bats such as *H. cervinus* respond to forest degradation by modifying their diets, or are 96 able to maintain stable diets through prey selection or behavioural changes in foraging. 97 We address three main predictions:

- 98 1) Taxonomic composition of the diet of *H. cervinus* is altered by rainforest99 degradation.
- 100 2) Individual bats are more specialised in logged forest sites than in primary forest.
- 101 3) Estimates of sampling completeness are heavily influenced by MOTU clustering
 102 threshold, quality-control methods used and the number of samples.

103 Methods

104	We sampled bats using six harp traps per night at four lowland tropical rainforest sites in
105	Sabah, Malaysia, each <500m above sea level and limited seasonality. Two sites comprise
106	mostly old growth rainforest (Danum Valley and Maliau Basin), and two sites have been
107	subject to substantial logging disturbance (the Sabah Biodiversity Experiment and the
108	Stability of Altered Forest Ecosystems Project) (Supplementary Table 1).

- Old growth rainforest:
- The Danum Valley Conservation Area (hereafter 'Danum') is a 438 km²
 region protected area of old growth rainforest in Sabah (Reynolds et al.,
 2011). Traps were erected in 2016 for ten nights in a 21-night period and
 2017 for ten nights in a 12-night period.
- The Maliau Basin Conservation Area (hereafter 'Maliau') is a 588 km²
 protected forest made up of lowland and hill forest, most of which has
 neither been logged nor inhabited in historical times. Traps were erected
 in 2016 and 2017 for ten nights in a 16-night period.
- Logged forest:

119	• The Stability of Altered Forest Ecosystems Project (hereafter 'SAFE') is
120	a large area of degraded forest being converted to oil palm plantation, with
121	fragments of forest retained for scientific study (Ewers et al., 2011). We
122	sampled in the blocks 'LFE', 'B' and 'C', within the Ulu Segama Forest
123	Reserve and Kalabakan area, during 2015, 2016 and 2017. Each block was
124	sampled for a 5-night period, and then resampled at least 5 weeks later.

The Sabah Biodiversity Experiment (Hector et al., 2011) (hereafter 'SBE')
is an area of forest which was logged once in the 1950s and once in the
2000s, and during the sampling period was in the early stages of
enrichment replanting (Hector et al., 2011). Sampling took place over a
total of 10 nights in a 20-night period in 2016.

Fieldwork, laboratory work and bioinformatics took place as previously described (Hemprich-Bennett et al., 2020). Briefly, bats were captured using harp traps erected along linear features such as streams and trails to target bat flyways. Sampling effort is summarised in Table 1. Faecal samples were processed by DNA extraction, PCR amplification of the CO1 gene using the primers described by Zeale et al (2011), and sequenced on an Illumina MiSeq. For complete methods see (Hemprich-Bennett et al., 2020).

137

138 **Bioinformatics pipeline**

Sequences were assembled into contigs using mothur (Schloss et al., 2009), and forward and reverse primers were removed using the galaxy web platform on the public server at usegalaxy.org (Afgan et al., 2016) sequence falling outside of a length of 155-159bp (2bp outside of the expected amplicon length) were excluded from analysis.

143

When processing the sequence data it is common to cluster sequences into MOTUs (Molecular Operational Taxonomic Units) (Floyd et al., 2002), on the basis of a given threshold of similarity, but the appropriate MOTU clustering thresholds required to bestrepresent the taxonomic diversity within metabarcoding samples are currently poorly

148 understood (Hemprich-Bennett et al., 2021). At high clustering thresholds routine 149 sequencing errors may be falsely designated as distinct MOTU, artificially inflating the 150 measured diversity and richness within a sample (Clare et al., 2016). Algorithms 151 implemented using software such as LULU (Frøslev et al., 2017) have been proposed as 152 a method of mitigating this, by combining probable duplicate MOTUs based on patterns 153 of sequence similarity and cooccurrence.

154

155 To assess the impact of clustering threshold on the datasets analysed (Hemprich-Bennett 156 et al., 2021) we generated datasets using MOTU clustering thresholds at ranges 91-98% 157 similarity, using the Uclust algorithm (Edgar, 2010) as implemented in the QIIME 158 platform (Caporaso et al., 2010). Representative sequences for each MOTU per clustering 159 level were then compared to one another using BLAST+ (Camacho et al., 2009), with the 160 resulting data being reduced in LULU (Frøslev et al., 2017) for quality control. All 161 resulting bat-MOTU adjacency lists were then transformed into adjacency matrices using 162 a custom perl script. These matrices were then split into multiple binary adjacency 163 matrices by site. Networks were created by pooling samples from multiple years. To test 164 prediction 2, separate analyses took place on networks both generated as composites of 165 multiple years, and as separate networks for each site and year (see Table 1). All 166 bioinformatic and statistical steps are recorded at

167 https://github.com/hemprichbennett/hice.

168 Prediction 1: Taxonomic composition of the diet of *H. cervinus* is altered by169 rainforest degradation

To analyse the prey taxa consumed by each bat, we used BLAST+ (Camacho et al., 2009)
to compare all MOTUs to a library of all arthropod CO1 genes identified to species level

172 using the Barcode of Life Database on 28/03/2018 (BOLD) (Ratnasingham and Hebert, 173 2007) (3,319,062 sequences), and assigned them taxonomy in MEGAN 6 (Huson et al., 174 2016) using the parameters in Salinas-Ramos et al. (2015). We then assigned MOTUs to 175 order and family level where possible, importing the resulting data into R for analysis, 176 and calculating the proportion of *H. cervinus* individuals per site consuming each 177 taxonomic order. To test the hypothesis that habitat type alters the order-level taxonomic 178 composition of the species' diet, we analysed the resulting values with a Chi-squared test. 179 The hypothesis was further tested using a permutational multivariate analysis of variance 180 test using distance matrices, and a non-metric multidimensional scaling ordination with 181 200 permutations using Bray-Curtis dissimilarity, both using the vegan package (Oksanen 182 et al., 2017) on datasets of the order-level diets of each individual bat. We also used a 183 similarity percentages analysis to identify the contribution of each taxonomic order to the 184 observed dissimilarity between sites and years, using Bray-Curtis dissimilarity.

185

We calculated correlations between the presence/absence of prey orders in faecal samples, using the r package 'corrplot' (Wei and Simko, 2017), to identify both potential significant correlations of prey consumption (e.g. bats that feed on Coleoptera may be more likely to feed on Blattodea), and any potential taxonomic bias in PCR.

190 Prediction 2: Individual bats are more specialised in logged forest sites than in old191 growth forest

We created binary bipartite networks for each sampling site and year at 95% similarity clustering and quality control using LULU. In the networks each individual bat and MOTU was classed as a distinct node. A criterion of 95% similarity was chosen for this and all following analyses because it provided a balance between over and under-splitting

MOTUs (Hemprich-Bennett et al., 2021). Using the R package 'bipartite' (Dormann, 2011) in R 3.4.4 (R Core Team, 2017) these networks were then analysed using the functions 'specieslevel', to calculate the degree of each bat ('degree' = the number of prey nodes a bat consumes). Differences between the degree of individuals were compared among sites using an ANOVA with Tukey's HSD test.

201 Prediction 3: Estimates of sampling completeness are heavily influenced by MOTU

202 clustering threshold and quality-control used

203 Using networks generated at each clustering threshold between 91 and 98% similarity,

both with and without quality-control using LULU (Frøslev et al., 2017), we estimated

- total MOTU richness and sampling completeness of the diet of *H. cervinus* at each site
- 206 and year using iNEXT (Hsieh et al., 2016), an R package for the interpolation and

207 extrapolation of species diversity using Hill numbers (Chao et al., 2014).

208

209 To assess how sample size affects assessments of bat diet, we generated multiple datasets

210 of *n* bats from each site, where *n* was a value of 10-100, increasing in increments of 10

211 (10, 20, 30, etc), with *n* bats taken at random from each site and the number of MOTUs

- 212 consumed in that sub-dataset calculated. This was repeated 100,000 times per site and
- 213 value of *n*, with the resulting data plotted in a violinplot.

214 **Results**

215 For the full sequencing run of multiple bat species (see Hemprich-Bennett et al., 2020) 216 18,737,930 contiguous reads were output when assembling the paired-end files. After 217 removing adapters and primers this was reduced to 10,064,815 sequences, which was 218 then further reduced to 932,459 haplotypes after collapsing to haplotype, removing 219 singletons and discarding sequences outside of 2bp of the expected read-length. For full 220 counts of MOTUs before and after clustering with LULU, see Supplementary information 221 2. Of these, 2,957,444 reads and 187,800 haplotypes were derived from *H. cervinus* 222 samples and included in this study.

Prediction 1: Taxonomic composition of the diet of *H. cervinus* is altered byrainforest degradation

225 The diet of the bat communities was dominated by insects from the orders Blattodea 226 (especially family Ectobiidae), Diptera (especially family Cecidomyiidae) and 227 Lepidoptera (Figure 1). The chi-squared test showed a non-significant effect of network 228 identity on the order-level composition of a bat populations' diet ($\chi 2 = 0.16$, df = 48, p > 229 0.05). The NMDS showed almost total overlap between the sites (Figure 2) with a stress 230 of 0.21, showing poor convergence. The permutational multivariate analysis of variance 231 test gave an R² of 0.014 for the explanatory power of site on bat diet. A total of 23 232 arthropod orders were eaten based on the combined diets of all bats, with Blattodea, 233 Coleoptera, Diptera and Lepidoptera collectively making up at least 79% of all MOTUs 234 identified at each site. Positive correlations were observed between the occurrences of 235 several taxa, with only Araneae and Hymenoptera being negatively correlated with the 236 presence of one another (Supplementary information 3). Blattodea was the only taxon 237 consistently observed to contribute significantly to inter-site dissimilarity scores (SAFE-

- 238 Maliau p<0.01, SAFE-SBE p = 0.014, Maliau-SBE p = 0.014, SBE-Danum p <0.01, see
- 239 Supplementary information 4). There was almost complete overlap between the different
- 240 years sampled at each site (Figure 2) and each site in 2016 (Figure 3).

241 Prediction 2: Individual bats will be more specialised in logged forest sites than in

- old growth forest
- When comparing networks with all years pooled together, significant differences (p<0.05) were only observed between Danum (old-growth) and SAFE (logged), and between SAFE (logged) and SBE (logged).
- 246 Prediction 3: Estimates of sampling completeness will be heavily influenced by

247 MOTU clustering threshold and quality-control used

- 248 None of the networks were estimated as near to fully sampled, with all estimates placing 249 completeness at under 54% (Figure 4), with completeness estimates varying between both 250 sites and years. The number of MOTUs expected increased markedly with clustering 251 threshold when not using LULU for quality control, but this effect was dramatically 252 reduced when using LULU. This algorithm increased estimated sampling completeness 253 by reducing observed and estimated MOTU richness, and lowered the estimated number 254 of samples required to sample the community. Full counts can be found in Supplementary 255 information 2.
- 256
- There was a positive correlation between the number of bats included in a dataset and thenumber of MOTUs detected (figure 5)

259 **Discussion**

Logging is widespread in tropical forests, yet the consequences of this structural disturbance for trophic interactions are poorly understood. Here we set out to assess how the diet of a generalist insectivorous bat differs between old-growth and degraded forest habitats. We observed broadly similar feeding habits in fawn leaf-nosed bats across forest type with bats consuming many arthropod orders, particularly Blattodea, Coleoptera, Diptera and Lepidoptera. Fawn leaf-nosed bats have extremely high dietary richness, with many hundreds of samples being required to fully capture their diet.

267

268 We observed very little alteration in the taxonomic composition of the diet of *H. cervinus*. 269 We saw no significant difference between the consumption of prey at the order-level 270 between sites or years. This suggests that while northeast Borneo may possess high beta-271 diversity of some insect species (Kitching et al., 2013), at coarse taxonomic levels there 272 is little spatial difference in the prey consumed by H. cervinus. Previous findings 273 suggested that, as high-duty cycle echolocators, H. cervinus primarily consumed flying 274 insects (Bell and Fenton, 1984; Link et al., 1986; Schnitzler and Kalko, 2001; Lazure and 275 Fenton, 2011), in particular Lepidoptera, Blattodea, Diptera and Coleoptera. The regular 276 presence of diverse families of spiders indicates a dietary contribution of these taxa 277 previously unknown in the Hipposideridae family of bats. Hipposiderids have been 278 observed gleaning stationary targets with fluttering wings (Bell and Fenton, 1984), but 279 the consumption of spiders would either suggest they are gleaning non-fluttering animals, 280 or taking them when ballooning as juveniles. Alternatively, the consumption of spiders 281 could be due to secondary predation: where the bat consumes a primary prey item which 282 has ingested a spider. This seems an unlikely explanation for our dataset, since predatory

283 arthropods other than Araneae are poorly represented in the MOTU dataset. In this study 284 we used one of the most reliable primer sets for amplification of a wide range of digested 285 arthropods (Zeale et al., 2011; Alberdi et al., 2018), but they are also reported to have 286 taxonomic biases towards Diptera and Lepidoptera. However, we found no significant 287 negative correlations between detecting Dipteran or Lepidopteran DNA in a sample, and 288 the detection of any other prey order. This indicates that amplification of dipteran or 289 lepidopteran DNA did not consistently inhibit the amplification of another taxonomic 290 order during PCR, and that sequencing depth is sufficient.

291

292 There was no clear pattern of degree differing between logged and old growth habitats. 293 This is in contrast to our previous findings in these study sites (Hemprich-Bennett et al., 294 2020), that the overall assemblage of bat species in these sites consistently had reduced 295 degree in logged forest than old growth. The diversity of the overall bats' diet is likely 296 due to the high diversity of prey available to them, and the lack of observed differences 297 in diet between sites may indicate highly flexible foraging, with low impact of land-use 298 change on their diets. Being able to forage adaptively, or fly long distances to viable 299 feeding sites (Struebig et al., 2009) may enable them to remain abundant despite selective 300 logging, while conspecific species experience population declines (Struebig et al., 2013). 301 This species may, as a result, provide ecological redundancy and continue to contribute 302 insectivory when more sensitive bat species have become locally extinct.

303

A crucial concern in network ecology is the minimum number of samples or observations
required to characterise reliably the structure and identity of the interactions within a
network (Nielsen and Bascompte, 2007; Rivera-Hutinel et al., 2012). This requirement is

307 complicated in studies utilising DNA metabarcoding as the number of nodes generated is 308 dependent on the bioinformatic choices used to generate them. While MOTU approaches 309 frequently apply a standard resolution to all nodes which helps control for variation in 310 identification, altering MOTU clustering threshold will change the number of nodes and 311 estimates of completeness, analogous to lumping taxonomy-based identifications to 312 higher levels, but without a biological equivalent. We tested MOTU clustering and the 313 use of LULU for quality-control and demonstrated that it was possible to alter estimates 314 of sampling completeness greatly (Figure 4). However, when generating networks with a 315 range of bioinformatics combinations, we observed that none exceeded an estimate of 316 50% completeness and thus regardless of parameters used, obtaining the full estimate of 317 *H. cervinus* diet would require several hundred samples per site, with the same likely true 318 of many ecologically similar species. Altering MOTU clustering parameters has 319 previously been shown to cause great variation in MOTU counts (Clare et al., 2016) and 320 changes in numerous measures of network-level architecture (Hemprich-Bennett et al., 321 2021). The reduction in number of estimated MOTUs provided by LULU (Frøslev et al., 322 2017) is expected to be of great use in future metabarcoding-based studies to reduce 323 spurious MOTU generation.

324

The dietary richness found here echoes previous studies (Clare et al., 2009; McCracken et al., 2012) but raises question about the capacity of bats to distinguish between prey types in detail (Neuweiler, 1990) and if this has implications for prey-choice. At the same time, our results highlight the substantial challenge of characterising the diets of this and other insectivorous bat species, especially in hyperdiverse ecosystems such as tropical rainforests. Their large dietary breadth is further highlighted by the fact that DNA extractions performed here were for pooled faecal samples from each individual bat, a

technique which Mata *et al.* (Mata et al., 2018) found underestimated the total richness
of the diet per bat. Previous intensive studies of arthropod diversity in lowland tropical
rainforest have failed to reach an asymptote (Novotný and Basset, 2000; Basset et al.,
2012), and if bats are foraging opportunistically it is perhaps unsurprising that the
taxonomic breadth of their diet is extremely large and nearly impossible to sample
completely.

338

339 We demonstrate the vast richness of prey consumed by insectivorous bats in tropical 340 rainforest and show that although quality-control steps in metabarcoding can reduce our 341 estimates of the number of distinct prey items in a site, many hundreds of samples are 342 required to collect a representative description of total diet. Although we focussed our 343 sampling on a single species of insectivorous bat, some inferences likely also apply to 344 similar species, and to other studies that use metabarcoding. The number of sites analysed 345 in this study was low, but it has been shown here that this Hipposiderid species has a 346 highly diverse diet; relying on cockroaches more than previously thought and potentially 347 having a strategy of gleaning non-fluttering prey previously unknown in the family. This 348 bat species is thus thought to exhibit low levels of dietary response to habitat degradation, 349 potentially indicating reasons for their known versatility in the face of landscape 350 modification.

351 Acknowledgements

This study was funded by the UK Natural Environment Research Council to SJR, OL and MJS (under the Human-Modified Tropical Forests programme, NE/K016407/1; http://lombok.nerc-hmtf.info/), a Royal Society grant (RG130793) to ELC, and a Bat Conservation International grant to DRHB. We used Queen Mary's Apocrita HPC facility, supported by QMUL Research-IT (http://doi.org/10.5281/zenodo.438045).

357 For assistance with data collection we thank Jamiluddin Jami, Arnold James, Mohd. 358 Mustamin, Ampat Siliwong, Sabidee Mohd. Rizan, Najmuddin Jamal, Genevieve 359 Durocher and Anne Seltmann. We thank the Sabah Biodiversity Council, Sabah Forest 360 Department, Yayasan Sabah, and Benta Wawasan Sdn. Bhd. for research permissions 361 (Access licenses: JKM/MBS.1000- 2/2 (374), JKM/MBS.1000-2/2 JLD.4 (23), 362 JKM/MBS.1000-2/2 JLD.4 (45), JKM/MBS.1000-2/2 JLD.4 (41), JKM/MBS.1000-2/2 363 JLD.4 (46), JKM/MBS.1000-2/2 JLD.5 (123), JKM/MBS.1000-2/2 JLD.5 (153), Export 364 licenses: JKM/MBS.1000-2/3 JLD.2(55), JKM/MBS.1000-2/3 JLD.2 (95). 365 JKM/MBS.1000-2/3 JLD.3 (31)) We thank Eleanor Slade and members of the LOMBOK 366 consortium for facilitating research in Sabah, and we are grateful to the Sabah 367 Biodiversity Council (Danum Valley access permits: YS/DVMC/2015/221, 368 YS/DVMC/2016/11, YS/DVMC/2015/222, YS/DVMC/2016/13, YS/DVMC/2017/42, 369 YS/DVMC/2017/41, Maliau Basin permits: access YS/MBMC/2015/186, 370 YS/MBMC/2016/23, YS/MBMC/2015/187, YS/MBMC/2016/25, YS/MBMC/2017/67, 371 YS/MBMC/2017/66)).

We thank Steven Le Comber, Hernani Oliveira, Joshua Potter, Sandra Álvarez Carretero
and Kim Warren for their analytical assistance, and Mark Brown and Darren Evans, for
helpful comments on earlier versions of this manuscript.

17

375 Works cited

376	Afgan, E., Baker, D., van den Beek, M., Blankenberg, D., Bouvier, D., Čech, M., et al.	
377	(2016). The Galaxy platform for accessible, reproducible and collaborative	
378	biomedical analyses: 2016 update. <i>Nucleic Acids Res.</i> 44, W3–W10.	
379	doi:10.1093/nar/gkw343.	
380	Aizpurua, O., Budinski, I., Georgiakakis, P., Gopalakrishnan, S., Ibañez, C., Mata, V.,	
381	et al. (2018). Agriculture shapes the trophic niche of a bat preying on multiple	
382	pest arthropods across Europe: Evidence from DNA metabarcoding. <i>Mol. Ecol.</i>	
383	27, 815–825. doi:https://doi.org/10.1111/mec.14474.	
384	Alberdi, A., Aizpurua, O., Gilbert, M. T. P., and Bohmann, K. (2018). Scrutinizing key	
385	steps for reliable metabarcoding of environmental samples. <i>Methods Ecol. Evol.</i>	
386	9, 134–147. doi:10.1111/2041-210X.12849.	
387	Andriollo, T., Gillet, F., Michaux, J. R., and Ruedi, M. (2019). The menu varies with	
388	metabarcoding practices: A case study with the bat <i>Plecotus auritus</i> . <i>PLOS ONE</i>	
389	14, e0219135. doi:10.1371/journal.pone.0219135.	
390	 Asner, G. P., Rudel, T. K., Aide, T. M., Defries, R., and Emerson, R. (2009). A	
391	Contemporary Assessment of Change in Humid Tropical Forests. <i>Conserv. Biol.</i>	
392	23, 1386–1395. doi:https://doi.org/10.1111/j.1523-1739.2009.01333.x.	
393	Basset, Y., Cizek, L., Cuénoud, P., Didham, R. K., Guilhaumon, F., Missa, O., et al.	
394	(2012). Arthropod Diversity in a Tropical Forest. <i>Science</i> 338, 1481–1484.	
395	doi:10.1126/science.1226727.	
396	Bell, G. P., and Fenton, M. B. (1984). The use of Doppler-shifted echoes as a flutter	
397	detection and clutter rejection system: the echolocation and feeding behavior of	
398	<i>Hipposideros ruber</i> (Chiroptera: Hipposideridae). <i>Behav. Ecol. Sociobiol.</i> 15,	
399	109–114. doi:10.1007/BF00299377.	
400	Burgar, J. M., Murray, D. C., Craig, M. D., Haile, J., Houston, J., Stokes, V., et al.	
401	(2014). Who's for dinner? High-throughput sequencing reveals bat dietary	
402	differentiation in a biodiversity hotspot where prey taxonomy is largely	
403	undescribed. <i>Mol. Ecol.</i> 23, 3605–3617. doi:10.1111/mec.12531.	
404	Camacho, C., Coulouris, G., Avagyan, V., Ma, N., Papadopoulos, J., Bealer, K., et al.	
405	(2009). BLAST+: architecture and applications. <i>BMC Bioinformatics</i> 10, 421.	
406	doi:10.1186/1471-2105-10-421.	
407	Caporaso, J. G., Kuczynski, J., Stombaugh, J., Bittinger, K., and Bushman, F. D.	
408	(2010). QIIME allows analysis of high-throughput community sequencing data.	
409	<i>Nat. Methods</i> 7, 335–336. doi:doi:10.1038/nmeth.f.303.	
410	Chao, A., Gotelli, N. J., Hsieh, T. C., Sande, E. L., Ma, K. H., Colwell, R. K., et al.	
411	(2014). Rarefaction and extrapolation with Hill numbers: a framework for	
412	sampling and estimation in species diversity studies. <i>Ecol. Monogr.</i> 84, 45–67.	

413	Clare, E. L., Chain, F. J. J., Littlefair, J. E., and Cristescu, M. E. (2016). The effects of		
414	parameter choice on defining molecular operational taxonomic units and		
415	resulting ecological analyses of metabarcoding data. <i>Genome</i> 59, 981–990.		
416	doi:10.1139/gen-2015-0184.		
417	Clare, E. L., Fraser, E. E., Braid, H. E., Fenton, M. B., and Hebert, P. D. N. (2009).		
418	Species on the menu of a generalist predator, the eastern red bat <i>Lasiurus</i>		
419	<i>borealis</i> : using a molecular approach to detect arthropod prey. <i>Mol. Ecol.</i> 18,		
420	2532–2542. doi:10.1111/j.1365-294X.2009.04184.x.		
421 422 423	Clare, E. L., Symondson, W. O. C., Broders, H., Fabianek, F., Fraser, E. E., MacKenzie, A., et al. (2014). The diet of <i>Myotis lucifugus</i> across Canada: assessing foraging quality and diet variability. <i>Mol. Ecol.</i> 23, 3618–3632. doi:10.1111/mec.12542.		
424	Czenze, Z. J., Tucker, J. L., Clare, E. L., Littlefair, J. E., Hemprich-Bennett, D. R.,		
425	Oliveira, H. F. M., et al. (2018). Spatiotemporal and demographic variation in		
426	the diet of New Zealand lesser short-tailed bats (<i>Mystacina tuberculata</i>). <i>Ecol.</i>		
427	<i>Evol.</i> 8, 7599–7610. doi:10.1002/ece3.4268.		
428	Deere, N. J., Guillera-Arroita, G., Baking, E. L., Bernard, H., Pfeifer, M., Reynolds, G.,		
429	et al. (2018). High Carbon Stock forests provide co-benefits for tropical		
430	biodiversity. J. Appl. Ecol. 55, 997–1008. doi:https://doi.org/10.1111/1365-		
431	2664.13023.		
432 433 434	Des Roches, S., Post, D. M., Turley, N. E., Bailey, J. K., Hendry, A. P., Kinnison, M. T., et al. (2018). The ecological importance of intraspecific variation. <i>Nat. Ecol. Evol.</i> 2, 57–64. doi:10.1038/s41559-017-0402-5.		
435 436	Dormann, C. F. (2011). How to be a specialist? Quantifying specialisation in pollination networks. <i>Netw. Biol.</i> 1, 1–20.		
437	Edgar, R. C. (2010). Search and clustering orders of magnitude faster than BLAST.		
438	<i>Bioinformatics</i> 26, 2460–2461. doi:10.1093/bioinformatics/btq461.		
439 440 441 442	Edwards, D. P., Larsen, T. H., Docherty, T. D. S., Ansell, F. A., Hsu, W. W., Derhé, M. A., et al. (2011). Degraded lands worth protecting: the biological importance of Southeast Asia's repeatedly logged forests. <i>Proc. R. Soc. B Biol. Sci.</i> 278, 82–90. doi:10.1098/rspb.2010.1062.		
443	Ewers, R. M., Boyle, M. J. W., Gleave, R. A., Plowman, N. S., Benedick, S., Bernard,		
444	H., et al. (2015). Logging cuts the functional importance of invertebrates in		
445	tropical rainforest. <i>Nat. Commun.</i> 6, 6836. doi:10.1038/ncomms7836.		
446	Ewers, R. M., Didham, R. K., Fahrig, L., Ferraz, G., Hector, A., Holt, R. D., et al.		
447	(2011). A large-scale forest fragmentation experiment: the Stability of Altered		
448	Forest Ecosystems Project. <i>Philos. Trans. R. Soc. B Biol. Sci.</i> 366, 3292–3302.		
449	doi:10.1098/rstb.2011.0049.		
450	Floyd, R., Abebe, E., Papert, A., and Blaxter, M. (2002). Molecular barcodes for soil		
451	nematode identification. <i>Mol. Ecol.</i> 11, 839–850. doi:10.1046/j.1365-		
452	294X.2002.01485.x.		

453	Frøslev, T. G., Kjøller, R., Bruun, H. H., Ejrnæs, R., Brunbjerg, A. K., Pietroni, C., et
454	al. (2017). Algorithm for post-clustering curation of DNA amplicon data yields
455	reliable biodiversity estimates. <i>Nat. Commun.</i> 8, 1188. doi:10.1038/s41467-017-
456	01312-x.
457	Gaveau, D. L. A., Sheil, D., Husnayaen, Salim, M. A., Arjasakusuma, S., Ancrenaz, M.,
458	et al. (2016). Rapid conversions and avoided deforestation: examining four
459	decades of industrial plantation expansion in Borneo. <i>Sci. Rep.</i> 6, 32017.
460	doi:10.1038/srep32017.
461	Gaveau, D. L. A., Sloan, S., Molidena, E., Yaen, H., Sheil, D., Abram, N. K., et al.
462	(2014). Four Decades of Forest Persistence, Clearance and Logging on Borneo.
463	<i>PLOS ONE</i> 9, e101654. doi:10.1371/journal.pone.0101654.
464	Hayward, R. M., Banin, L. F., Burslem, D. F. R. P., Chapman, D. S., Philipson, C. D.,
465	Cutler, M. E. J., et al. (2021). Three decades of post-logging tree community
466	recovery in naturally regenerating and actively restored dipterocarp forest in
467	Borneo. <i>For. Ecol. Manag.</i> 488, 119036. doi:10.1016/j.foreco.2021.119036.
468 469 470 471	 Hector, A., Philipson, C., Saner, P., Chamagne, J., Dzulkifli, D., O'Brien, M., et al. (2011). The Sabah Biodiversity Experiment: a long-term test of the role of tree diversity in restoring tropical forest structure and functioning. <i>Philos. Trans. R. Soc. Lond. B Biol. Sci.</i> 366, 3303–3315. doi:10.1098/rstb.2011.0094.
472	Hemprich-Bennett, D. R., Kemp, V. A., Blackman, J., Struebig, M. J., Lewis, O. T.,
473	Rossiter, S. J., et al. (2020). Altered structure and stability of bat-prey
474	interaction networks in logged tropical forests revealed by metabarcoding.
475	<i>bioRxiv</i> , 2020.03.20.000331. doi:10.1101/2020.03.20.000331.
476	Hemprich-Bennett, D. R., Oliveira, H. F. M., Comber, S. C. L., Rossiter, S. J., and
477	Clare, E. L. (2021). Assessing the impact of taxon resolution on network
478	structure. <i>Ecology</i> 102, e03256. doi:https://doi.org/10.1002/ecy.3256.
479 480 481	Hsieh, T. C., Ma, K. H., and Chao, A. (2016). <i>iNEXT: Interpolation and Extrapolation for Species Diversity</i> . Available at: http://chao.stat.nthu.edu.tw/blog/software-download/.
482	Huson, D. H., Beier, S., Flade, I., Górska, A., El-Hadidi, M., Mitra, S., et al. (2016).
483	MEGAN Community Edition - Interactive Exploration and Analysis of Large-
484	Scale Microbiome Sequencing Data. <i>PLOS Comput. Biol.</i> 12, e1004957.
485	doi:10.1371/journal.pcbi.1004957.
486 487 488 489	Kitching, R. L., Ashton, L. A., Nakamura, A., Whitaker, T., and Khen, C. V. (2013). Distance-driven species turnover in Bornean rainforests: homogeneity and heterogeneity in primary and post-logging forests. <i>Ecography</i> 36, 675–682. doi:10.1111/j.1600-0587.2012.00023.x.
490 491 492	Kolkert, H., Andrew, R., Smith, R., Rader, R., and Reid, N. (2020). Insectivorous bats selectively source moths and eat mostly pest insects on dryland and irrigated cotton farms. <i>Ecol. Evol.</i> 10, 371–388. doi:https://doi.org/10.1002/ece3.5901.

493 494	Lazure, L., and Fenton, M. B. (2011). High duty cycle echolocation and prey detection by bats. <i>J. Exp. Biol.</i> 214, 1131–1137. doi:10.1242/jeb.048967.	
495	Link, A., Marimuthu, G., and Neuweiler, G. (1986). Movement as a specific stimulus	
496	for prey catching behaviour in rhinolophid and hipposiderid bats. J. Comp.	
497	Physiol. A 159, 403–413. doi:10.1007/BF00603985.	
498	Mata, V. A., Rebelo, H., Amorim, F., McCracken, G. F., Jarman, S., and Beja, P.	
499	(2018). How much is enough? Effects of technical and biological replication on	
500	metabarcoding dietary analysis. <i>Mol. Ecol.</i> 0. doi:10.1111/mec.14779.	
501	McCracken, G. F., Westbrook, J. K., Brown, V. A., Eldridge, M., Federico, P., and	
502	Kunz, T. H. (2012). Bats Track and Exploit Changes in Insect Pest Populations.	
503	<i>PLoS ONE</i> 7, e43839. doi:10.1371/journal.pone.0043839.	
504	Milodowski, D. T., Coomes, D. A., Swinfield, T., Jucker, T., Riutta, T., Malhi, Y., et al.	
505	(2021). The impact of logging on vertical canopy structure across a gradient of	
506	tropical forest degradation intensity in Borneo. J. Appl. Ecol.	
507	doi:https://doi.org/10.1111/1365-2664.13895.	
508 509	Neuweiler, G. (1990). Auditory adaptations for prey capture in echolocating bats. <i>Physiol. Rev.</i> 70, 615–641. doi:10.1152/physrev.1990.70.3.615.	
510 511	Nielsen, A., and Bascompte, J. (2007). Ecological networks, nestedness and sampling effort. J. Ecol. 95, 1134–1141. doi:10.1111/j.1365-2745.2007.01271.x.	
512	Novotný, V., and Basset, Y. (2000). Rare species in communities of tropical insect	
513	herbivores: pondering the mystery of singletons. <i>Oikos</i> 89, 564–572.	
514	doi:10.1034/j.1600-0706.2000.890316.x.	
515 516 517	Oksanen, J., Blanchet, F. G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., et al. (2017). <i>vegan: Community Ecology Package</i> . Available at: https://CRAN.R-project.org/package=vegan.	
518	Oliveira, H. F. M. de, Camargo, N. F., Hemprich-Bennett, D. R., Rodríguez-Herrera, B.,	
519	Rossiter, S. J., and Clare, E. L. (2020). Wing morphology predicts individual	
520	niche specialization in <i>Pteronotus mesoamericanus</i> (Mammalia: Chiroptera).	
521	<i>PLOS ONE</i> 15, e0232601. doi:10.1371/journal.pone.0232601.	
522	R Core Team (2017). <i>R: A Language and Environment for Statistical Computing</i> .	
523	Vienna, Austria: R Foundation for Statistical Computing Available at:	
524	https://www.R-project.org/.	
525 526 527	Ratnasingham, S., and Hebert, P. D. (2007). BOLD: The Barcode of Life Data System (http://www.barcodinglife.org). <i>Mol. Ecol. Resour.</i> 7, 355–364. doi:10.1111/j.1471-8286.2007.01678.x.	
528	Reynolds, G., Payne, J., Sinun, W., Mosigil, G., and Walsh, R. P. D. (2011). Changes in	
529	forest land use and management in Sabah, Malaysian Borneo, 1990-2010, with a	
530	focus on the Danum Valley region. <i>Philos. Trans. R. Soc. B Biol. Sci.</i> 366,	
531	3168–3176. doi:10.1098/rstb.2011.0154.	

532	Rivera-Hutinel, A., Bustamante, R. O., Marín, V. H., and Medel, R. (2012). Effects of
533	sampling completeness on the structure of plant–pollinator networks. <i>Ecology</i>
534	93, 1593–1603. doi:10.1890/11-1803.1.
535	Salinas-Ramos, V. B., Herrera Montalvo, L. G., León-Regagnon, V., Arrizabalaga-
536	Escudero, A., and Clare, E. L. (2015). Dietary overlap and seasonality in three
537	species of mormoopid bats from a tropical dry forest. <i>Mol. Ecol.</i> 24, 5296–5307.
538	doi:10.1111/mec.13386.
539	Schloss, P. D., Westcott, S. L., Ryabin, T., Hall, J. R., Hartmann, M., Hollister, E. B., et
540	al. (2009). Introducing mothur: Open-Source, Platform-Independent,
541	Community-Supported Software for Describing and Comparing Microbial
542	Communities. <i>Appl. Environ. Microbiol.</i> 75, 7537–7541.
543	doi:10.1128/AEM.01541-09.
544	Schnitzler, HU., and Kalko, E. K. V. (2001). Echolocation by Insect-Eating Bats.
545	<i>BioScience</i> 51, 557–569. doi:10.1641/0006-
546	3568(2001)051[0557:EBIEB]2.0.CO;2.
547	Slade, E. M., Mann, D. J., and Lewis, O. T. (2011). Biodiversity and ecosystem
548	function of tropical forest dung beetles under contrasting logging regimes. <i>Biol.</i>
549	<i>Conserv.</i> 144, 166–174. doi:10.1016/j.biocon.2010.08.011.
550	Struebig, M. J., Kingston, T., Zubaid, A., Le Comber, S. C., Mohd-Adnan, A., Turner,
551	A., et al. (2009). Conservation importance of limestone karst outcrops for
552	Palaeotropical bats in a fragmented landscape. <i>Biol. Conserv.</i> 142, 2089–2096.
553	doi:10.1016/j.biocon.2009.04.005.
554	Struebig, M. J., Turner, A., Giles, E., Lasmana, F., Tollington, S., Bernard, H., et al.
555	(2013). Quantifying the Biodiversity Value of Repeatedly Logged Rainforests.
556	<i>Adv. Ecol. Res.</i> 48, 183–224. doi:10.1016/B978-0-12-417199-2.00003-3.
557	Tournayre, O., Leuchtmann, M., Galan, M., Trillat, M., Piry, S., Pinaud, D., et al.
558	(2021). eDNA metabarcoding reveals a core and secondary diets of the greater
559	horseshoe bat with strong spatio-temporal plasticity. <i>Environ. DNA</i> 3, 277–296.
560	doi:https://doi.org/10.1002/edn3.167.
561 562 563 564	 Vallejo, N., Aihartza, J., Goiti, U., Arrizabalaga-Escudero, A., Flaquer, C., Puig, X., et al. (2019). The diet of the notch-eared bat (<i>Myotis emarginatus</i>) across the Iberian Peninsula analysed by amplicon metabarcoding. <i>Hystrix Ital. J. Mammal.</i> 30, 59–64. doi:10.4404/hystrix-00189-2019.
565 566	Wei, T., and Simko, V. (2017). R package "corrplot": Visualization of a correlation matrix. Available at: https://github.com/taiyun/corrplot.
567	Zeale, M. R. K., Butlin, R. K., Barker, G. L. A., Lees, D. C., and Jones, G. (2011).
568	Taxon-specific PCR for DNA barcoding arthropod prey in bat faeces. <i>Mol. Ecol.</i>
569	<i>Resour.</i> 11, 236–244. doi:10.1111/j.1755-0998.2010.02920.x.
570	

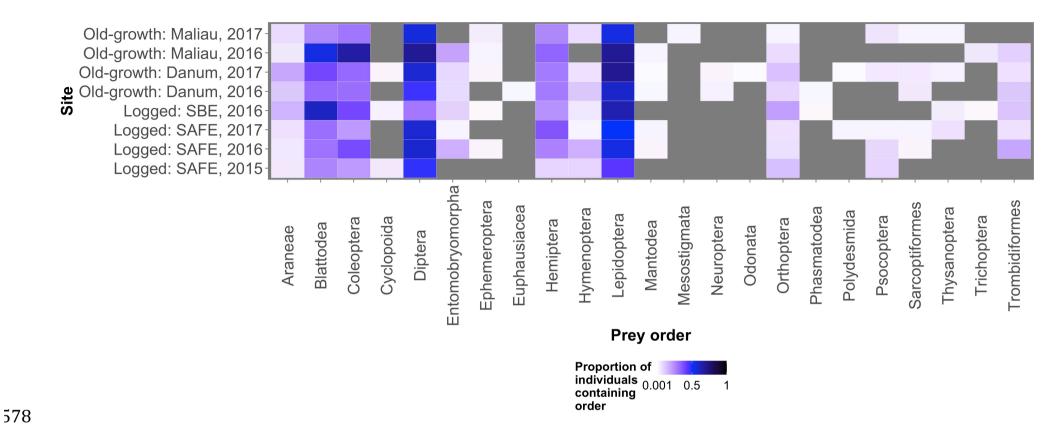
571 Tables

- 572 Table 1. Trapping effort per site, in harp trap nights. One harp trap night is a harp trap
- 573 erected for a single night. Six harp traps were used per night, so a single night's trapping
- 574 was equal to six harp trap nights.

Sample Site	2015	2016	2017
SAFE	216	180	180
Danum	0	60	60
Maliau	0	60	60
SBE	0	60	0

575

576 Figures



579 Figure 1. The proportion of all individual bats within a sampling event found to consume each potential prey order. Diptera, Lepidoptera and Blattodea
580 were the commonest prey items, with other prey orders being consumed rarely. The grey background shows locations in the plot where no arthropods of

581 that order were detected in any bats.

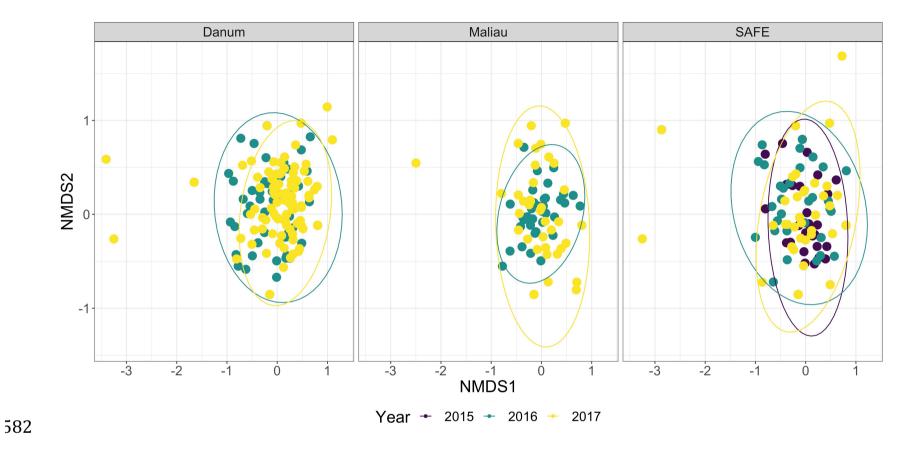


Figure 2. Non-Metric Multidimensional Scaling ordination of the order-level consumption of individual bats across multiple years. The ellipses of each
site show almost complete overlap. Stress was 0.21, indicating poor convergence. Danum and Maliau are old-growth sites, SAFE is a logged forest site.

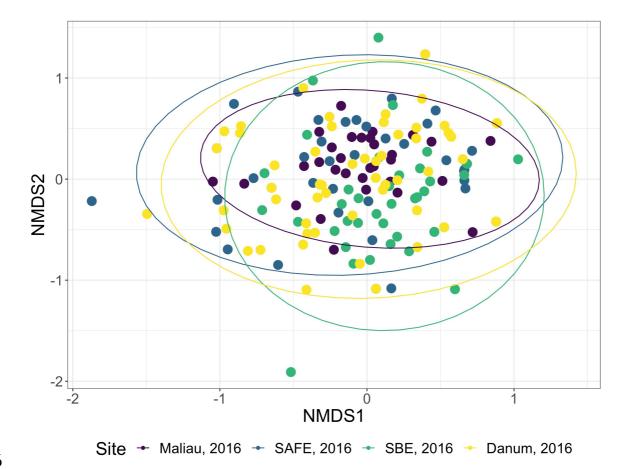
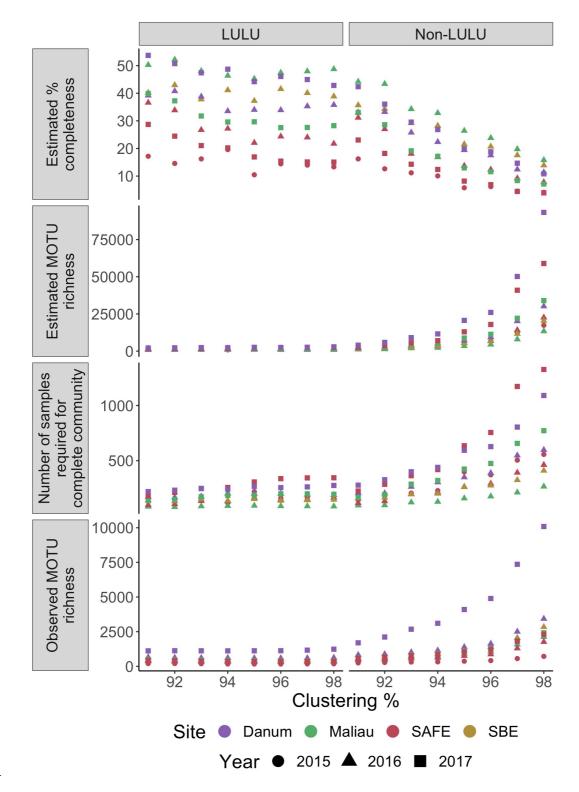




Figure 3. Non-Metric Multidimensional Scaling ordination of the order-level
consumption of individual bats in 2016. The ellipses of each site show almost complete
overlap. Stress was 0.22, indicating poor convergence. Danum and Maliau are old-growth
sites, SAFE and SBE are logged forest sites.



591

Figure 4. Completeness and richness for each network over a range of MOTU clustering
thresholds, with and without use of LULU for post-clustering quality-control. Number of
MOTUs is strongly positively correlated with clustering level when not using LULU for
quality-control, reducing the estimated completeness of each network.

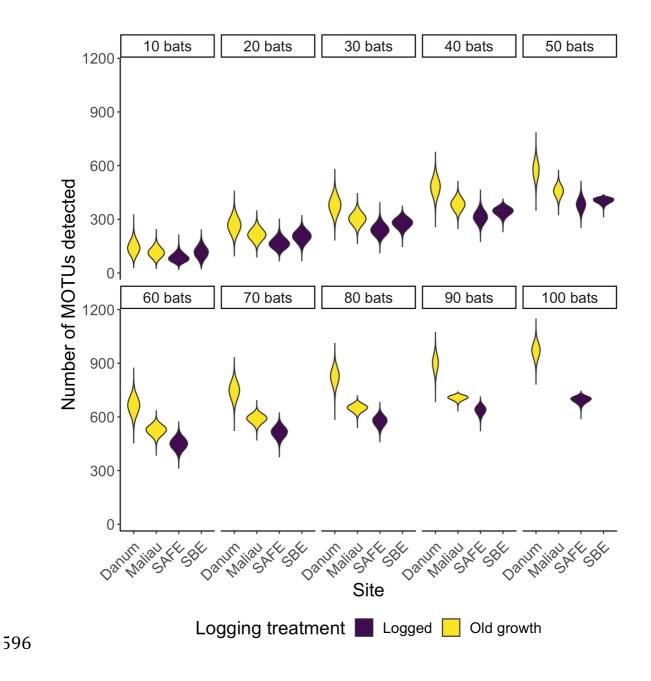


Figure 5: Violinplots showing the distribution of the number of MOTUs consumed when reducing a dataset to n bats. With small datasets, sites appear to be rather similar in MOTU richness, but differences emerge as sample sizes increase.