1 Broad-spectrum in vitro antiviral activity of ODBG-P-RVn: an orally-available, lipid-modified

2 monophosphate prodrug of remdesivir parent nucleoside (GS-441524)

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4 Michael K. Lo[a*]#, Punya Shrivastava-Ranjan[a], Payel Chatterjee[a], Mike Flint[a], James R. Beadle[b], Nadejda

5 Valiaeva[b], Robert T. Schooley[b], Karl Y. Hostetler[b], Joel M. Montgomery[a], and Christina Spiropoulou[a*]

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7 [a]Viral Special Pathogens Branch, Centers for Disease Control and Prevention, Department of Health and

8 Human Services, Atlanta, Georgia, USA

9 [b]Division of Infectious Diseases and Global Public Health, Department of Medicine, University of

10 California San Diego, La Jolla, California, USA

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13 *Address correspondence to:

14 #Michael K. Lo: mko2@cdc.gov and Christina F. Spiropoulou: ccs8@cdc.gov

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17 lipid prodrugs, ODBG, Vero E6 cells, Huh7 cells, NCI-H358 cells, human telomerase reverse-transcriptase

18 (hTERT) immortalized microvascular endothelial cells (TIME), and human small airway epithelial cells

19 (HSAEC1-KT)

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ABSTRACT

The intravenous administration of remdesivir for COVID-19 confines its utility to hospitalized patients. We evaluated the broad-spectrum antiviral activity of ODBG-P-RVn, an orally available, lipid-modified monophosphate prodrug of the remdesivir parent nucleoside (GS-441524) against viruses that cause diseases of human public health concern, including SARS-CoV-2. ODBG-P-RVn showed 20-fold greater antiviral activity than GS-441524 and had near-equivalent activity to remdesivir in primary-like human small airway epithelial cells. Our results warrant investigation of ODBG-P-RVn efficacy in vivo.

29 Remdesivir (RDV; Veklury, GS-5734) is an adenosine nucleotide analog phosphoramidate prodrug with
30 broad-spectrum antiviral activity in vitro and in vivo (1-8), and is currently the only therapeutic approved
31 by the FDA for treating coronavirus 19 disease (COVID-19) in hospitalized patients over the age of 12 (9).
32 While RDV did not significantly reduce COVID-19 mortality, it did shorten the time to recovery compared
33 to a placebo control group (10). The short half-life of RDV in human and animal plasma (1, 8, 11, 12),
34 alongside the in vivo efficacy of RDV parent nucleoside (GS-441524, RVn) against coronaviruses including
35 severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (13-16), have driven proposals to utilize
36 RVn instead of RDV to treat COVID-19 (17). A recent comparative pharmacokinetic study in non-human
37 primates, however, demonstrated higher nucleoside triphosphate (NTP) levels in lower respiratory tract
38 tissues of RDV-dosed animals than in RVn-dosed animals (8). A significant drawback of RDV is the
39 requirement for intravenous administration, which limits its use to hospital contexts. In an attempt to
40 develop an orally bioavailable form of remdesivir, we recently synthesized a 1-O-octadecyl-2-O-benzyl-
41 sn-glycerylester (ODBG) lipid-modified monophosphate prodrug of RVn (ODBG-P-RVn), which
42 demonstrated more favorable in vitro antiviral activity against SARS-CoV-2 compared to that of RVn and
43 RDV in Vero-E6 cells (18).

44 In this study, we extended our in vitro comparisons to include 14 viruses from across 7 virus families
45 responsible for causing diseases of significant human public health concern. These were *Filoviridae*:
46 Ebola virus (EBOV) and Marburg virus (MARV) (19, 20); *Paramyxoviridae*: Nipah virus (NiV), Hendra virus
47 (HeV), human parainfluenza virus 3 (hPIV3), measles virus (MV), mumps virus (MuV), and Sosuga virus
48 (SoSuV) (21-27); *Pneumoviridae*: respiratory syncytial virus (RSV) (28); *Flaviviridae*: yellow fever virus
49 (YFV); *Arenaviridae*: Lassa virus (LASV) (29); *Nairoviridae*: Crimean-Congo hemorrhagic fever virus
50 (CCHFV) (30); and *Coronaviridae*: SARS-CoV-2 (31). We utilized 3 previously described assays to compare
51 the antiviral activities of RVn, RDV, and ODBG-P-RVn against this panel of viruses: 1) directly measuring
52 fluorescence of a reporter protein expressed by recombinant viruses (REP) (2), (Figure 1A); 2)

53 quantitating focus-forming units (FFU) via fluorescent reporter imaging (32) (Figure 1B); and 3) indirectly
54 measuring cytopathic effect (CPE) based on cellular ATP levels (CellTiterGlo 2.0, Promega) (2) (Figure
55 1C), which was also used to evaluate compound cytotoxicity (Figure 1D). Assay conditions varied based
56 on virus replication kinetics and on the specific assay used; multiplicities of infection (MOI) ranged from
57 0.01–0.25, and endpoint measurements were conducted between 72-144 hours post-infection (hpi). We
58 initially conducted dose-response experiments using 8-point, 3-fold serial dilutions of RVn, RDV, and
59 ODBG-P-RVn against our panel of viruses in Vero-E6 cells, and showed that ODBG-P-RVn consistently
60 had greater antiviral activity than RVn and RDV against all viruses susceptible to RVn/RDV inhibition,
61 with effective concentration (EC_{50}) values ranging from 0.026 to 1.13 µM (Figure 1, Vero-E6 assays
62 represented in left column of panels A, B, C; Supplemental Figure S1; Table 1). RVn and ODBG-P-RVn
63 induced partial cytotoxicity but only at the highest concentration tested (100 µM) and without reaching
64 50% cytotoxicity (CC_{50}). We then compared these antivirals in human hepatoma (Huh7) and
65 bronchioalveolar carcinoma (NCI-H358) cell lines, which represent more relevant cell types targeted by
66 subsets of viruses used in our study. In both human cell lines, although ODBG-P-RVn showed EC_{50} values
67 remarkably similar to those observed in Vero-E6 cells and was 3- to 5-fold more active than RVn, it
68 consistently showed 6- to 20-fold less activity than RDV (Figure 1 [Huh7 and NCI-H358 assays
69 represented, respectively, in the middle and right columns of panels A, B, and C]; Supplemental Figures
70 S2, S3; Table 1). Whereas CC_{50} values for RDV in Huh7 and NCI-H358 cells were 54.2 and 77.2 µM,
71 respectively, ODBG-P-RVn was less cytotoxic in Huh7 cells (CC_{50} = 93.4 µM) and did not show
72 measurable cytotoxicity in NCI-H358 cells even at the highest concentration tested (100 µM) (Figure 1D,
73 right panel; Table 1).

74 To further evaluate cell type-specific effects on the antiviral activities of RVn, RDV, and ODBG-P-RVn, we
75 tested them against a smaller subset of filoviruses (EBOV-ZsG, MARV-ZsG) and a paramyxovirus (NiV-
76 ZsG) expressing ZsGreen reporter in primary-like human telomerase reverse transcriptase (hTERT)

77 immortalized human microvascular endothelial (TIME) cells (33, 34). In TIME cells, we observed a similar
78 trend in antiviral activity as in Huh7 and NCI-H358 cells, with ODBG-P-RVn showing 15- to 22-fold
79 greater activity than RVn, but 5- to 8-fold less activity than RDV in reporter-based assays (Figure 2A,
80 Table 2). To confirm this, we compared the respective abilities of RDV and ODBG-P-RVn to reduce
81 infectious yield of EBOV-ZsG and NiV-ZsG (MOI = 0.25) when cells were treated with each compound 2
82 hpi. Virus supernatants were collected at 72hpi and titered on Huh7 (for EBOV-ZsG) or NCI-H358 (for
83 NiV-ZsG) cells to determine 50% tissue culture infectious dose ($TCID_{50}$) by the method of Reed and
84 Muench (35). Both RDV and ODBG-P-RVn equivalently reduced infectious yield of EBOV-ZsG by up to 4
85 $\log_{10}$ and of NiV-ZsG by approximately 2 $\log_{10}$, in a dose-dependent manner, with EC_{50} values closely
86 mirroring values determined in reporter assays (Figure 2B, left and middle panels; Table 2). However,
87 RDV was more cytotoxic (CC_{50} = 17.2 µM) than ODBG-P-RVn (CC_{50} > 50 µM) (Figure 2B, right panel; Table
88 2), which is reflected in its biphasic inhibition of NiV-ZsG (Figure 2B, middle panel, cytotoxic inhibition by
89 RDV shown at 16.6 µM). Since the ODBG lipid modification has been shown to enhance in vivo lung
90 tissue distribution for a different orally administered nucleoside (36), we compared the activity of the 3
91 compounds against filoviruses, paramyxoviruses, and RSV in another primary-like, hTERT-immortalized
92 small airway epithelial cell (HSAEC1-KT) (37). Notably, the dose-response curves of RDV and ODBG-P-
93 RVn were strikingly similar, with EC_{50} values in the submicromolar range within a 3-fold range of each
94 other; EC_{50} values for some viruses were almost identical (Figure 2C; Supplemental Figure 4; Table 2).
95 Furthermore, RDV and ODBG-P-RVn equivalently reduced the infectious yields of EBOV-ZsG and NiV-ZsG
96 in HSAEC1-KT cells by by 5 $\log_{10}$ and 3 $\log_{10}$, respectively, and their EC_{50} values reflected the limited
97 differential in antiviral activity between them (Figure 2D, left and middle panels; Table 2). Although
98 ODBG-P-RVn was more cytotoxic (CC_{50} = 20.5) in HSAEC1-KT cells than RDV (CC_{50} > 100; Figure 2D, right
99 panel; Table 2), it also effectively reduced virus yields at non-cytotoxic concentrations.

100 In summary, our results demonstrate that ODBG-P-RVn has greater antiviral activity than RVn in all cell
101 lines tested and has cell-type dependent activity levels that range from moderately lesser than to nearly
102 equal to those of RDV. *In vivo* RDV is converted rapidly to RVn (1, 8, 11, 12), which has 0.5 to 2 $\log_{10}$ less
103 activity than RDV against most of the viruses tested. In contrast, ODBG-P-RVn is stable in plasma for >24
104 hours and at therapeutic plasma levels of ODBG-P-Rvn (above EC_{90} for SARS-CoV-2) after oral
105 administration of 16.9 mg/kg to Syrian hamsters; furthermore RVn was not observed at virologically
106 significant levels (38). Thus, one would predict sustained *in vivo* antiviral activity with ODBG-P-RVn
107 without substantial generation in plasma of RVn, the less active metabolite. Taken together, our results
108 strongly support investigation of in vivo efficacy of ODBG-P-RVn not only against SARS-CoV-2 but also
109 against other viruses significant to human health.

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253 FIGURE LEGENDS

254 Figure 1. Comparison of antiviral activities of RVn, RDV, and ODBG-P-RVn in African green monkey (Vero-
255 E6), human hepatoma (Huh7), and human bronchioalveolar carcinoma (NCI-H358) cell lines using
256 reporter-based, image-based, and cytopathic effect (CPE) assays. Representative dose-response
257 inhibition of viral replication and induction of cellular cytotoxicity by RVn (blue shapes), RDV (black
258 shapes), and ODBG-P-RVn (red shapes). A) Direct measurement of reporter fluorescence intensity by
259 recombinant Ebola virus (EBOV) expressing ZsGreen protein in Vero-E6 (left panel) and Huh7 (middle
260 panel) cells, and recombinant Nipah virus (NiV) expressing ZsGreen protein in NCI-H358 (right panel)
261 cells. B) Image-based counting of reporter fluorescence-positive cells infected with recombinant severe
262 acute respiratory syndrome coronavirus 2 (SARS-CoV-2) expressing mNeonGreen protein (Vero-E6 and
263 Huh7) and recombinant respiratory syncytial virus (RSV) expressing eGFP (NCI-H358). Infected cells
264 treated with DMSO were considered as 100% fluorescence intensity signal and 100% fluorescence-
265 positive cell counts. C) Compound-based inhibition of CPE induced by yellow fever virus (YFV) in Vero-E6
266 and Huh7 cells and by Hendra virus (HeV) in NCI-H358 cells determined by measuring cellular ATP levels
267 (CellTiterGlo 2.0). ATP levels in uninfected cells treated with DMSO were considered 100% CPE
268 inhibition. D) Compound cytotoxicity/cell viability measured by CellTiterGlo 2.0 assay. Dose-response
269 curves were fitted to the mean value of experiments performed in biological triplicate for each
270 concentration in the 8-point, 3-fold dilution series using a 4-parameter non-linear logistic regression
271 curve with variable slope. Data points and error bars indicate the mean value and standard deviation of
272 3 biological replicates; each colored shape/line in the legend represents an independent experiment
273 performed in biological triplicate. RVn and RDV used in this study was obtained from MedChemExpress
274 (Monmouth Junction, NJ USA).

275 Figure 2. Comparison of cell type-dependent antiviral activities of RVn, RDV, and ODBG-P-RVn in
276 primary-like hTERT-immortalized microvascular endothelial (TIME) cells and small airway epithelial cells

277 (HSAEC1-KT). A) Representative dose-response inhibition of recombinant EBOV, NiV, and Marburg virus
278 (MARV) expressing ZsGreen protein in TIME cells. B) Yield reduction of infectious EBOV-ZsG (left panel)
279 and NiV-ZsG (middle panel) by RDV and ODBG-P-RVn. Compound cytotoxicity/cell viability (right panel)
280 in TIME cells measured via CellTiterGlo 2.0 assay. C) Representative dose-response inhibition of
281 recombinant EBOV, NiV, and MARV expressing ZsGreen protein in HSAEC1-KT cells. D) Reduction of
282 infectious yield of EBOV-ZsG (left panel) and NiV-ZsG (middle panel) by RDV and ODBG-P-RVn in
283 HSAEC1-KT cells. Compound cytotoxicity/cell viability (right panel) in HSAEC1-KT cells measured via
284 CellTiterGlo 2.0 assay. Dose-response curves were fitted to the mean value of experiments performed in
285 biological triplicate for each concentration in the 8-point, 3-fold dilution series using a 4-parameter non-
286 linear logistic regression curve with variable slope. Data points and error bars indicate the mean value
287 and standard deviation of 3 or 4 biological replicates; each colored shape/line in the legend represents
288 an independent experiment performed in biological triplicate. Infectious yield reduction assays were
289 conducted once with biological quadruplicates.

290 SUPPLEMENTAL FIGURE LEGENDS

291 Supplemental Figure S1. Comparison of antiviral activities of RVn, RDV, and ODBG-P-RVn in African
292 green monkey (Vero-E6) cells using reporter-based, image-based, and CPE assays. Representative dose-
293 response inhibition of virus replication by RVn (blue shapes), RDV (black shapes), and ODBG-P-RVn (red
294 shapes). Signal from infected cells treated with DMSO served as 100% fluorescence intensity signal for
295 reporter assays and 100% fluorescence-positive cell counts for image-based assays. CPE inhibition was
296 measured by determining cellular ATP levels using CellTiterGlo 2.0 assay reagent. ATP levels in
297 uninfected cells treated with DMSO served as 100% CPE inhibition. Dose-response curves were fitted to
298 the mean value of experiments performed in biological triplicate for each concentration in the 8-point,
299 3-fold dilution series using a 4-parameter non-linear logistic regression curve with variable slope. Data
300 points and error bars indicate the mean value and standard deviation of 3 biological replicates; each

301 colored shape/line in the legend represents an independent experiment performed in biological
302 triplicate.

303 Supplemental Figure S2. Comparison of antiviral activities of RVn, RDV, and ODBG-P-RVn in Huh7 cells
304 using reporter-based, image-based, and CPE assays. Representative dose-response inhibition of virus
305 replication by RVn (blue shapes), RDV (black shapes), and ODBG-P-RVn (red shapes). Signal from
306 infected cells treated with DMSO served as 100% fluorescence intensity signal for reporter assays and
307 100% fluorescence-positive cell counts for image-based assays. CPE inhibition was measured by
308 determining cellular ATP levels using CellTiterGlo 2.0 assay reagent. ATP levels in uninfected cells
309 treated with DMSO served as 100% CPE inhibition. Dose-response curves were fitted to the mean value
310 of experiments performed in biological triplicate for each concentration in the 8-point, 3-fold dilution
311 series using a 4-parameter non-linear logistic regression curve with variable slope. Data points and error
312 bars indicate the mean value and standard deviation of 3 biological replicates; each colored shape/line
313 in the legend represents an independent experiment performed in biological triplicate.

314 Supplemental Figure S3. Comparison of antiviral activities of RVn, RDV, and ODBG-P-RVn in human
315 bronchioalveolar carcinoma (NCI-H358) cells using reporter-based, image-based, and CPE assays.
316 Representative dose-response inhibition of virus replication by RVn (blue shapes), RDV (black shapes),
317 and ODBG-P-RVn (red shapes). Signal in infected cells treated with DMSO served as 100% fluorescence
318 intensity signal for reporter assays and 100% fluorescence-positive cell counts for image-based assays.
319 CPE inhibition was measured by determining cellular ATP levels using CellTiterGlo 2.0 assay reagent. ATP
320 levels in uninfected cells treated with DMSO served as 100% CPE inhibition. Dose-response curves were
321 fitted to the mean value of experiments performed in biological triplicate for each concentration in the
322 8-point, 3-fold dilution series using a 4-parameter non-linear logistic regression curve with variable
323 slope. Data points and error bars indicate the mean value and standard deviation of 3 biological

324 replicates; each colored shape/line in the legend represents an independent experiment performed in
325 biological triplicate.

326 Supplemental Figure S4. Comparison of antiviral activities of RVn, RDV, and ODBG-P-RVn in primary-like
327 human small airway epithelial (HSAEC1-KT) cells using reporter-based, image-based, and CPE assays.
328 Representative dose-response inhibition of virus replication by RVn (blue shapes), RDV (black shapes),
329 and ODBG-P-RVn (red shapes). Signal in infected cells treated with DMSO served as 100% fluorescence
330 intensity signal for reporter assays and 100% fluorescence-positive cell counts for image-based assays.
331 CPE inhibition was measured by determining cellular ATP levels using CellTiterGlo 2.0 assay reagent. ATP
332 levels in uninfected cells treated with DMSO served as 100% CPE inhibition. Dose-response curves were
333 fitted to the mean value of experiments performed in biological triplicate for each concentration in the
334 8-point, 3-fold dilution series using a 4-parameter non-linear logistic regression curve with variable
335 slope. Data points and error bars indicate the mean value and standard deviation of 3 biological
336 replicates; each colored shape/line in the legend represents an independent experiment performed in
337 biological triplicate.

Figure 1.

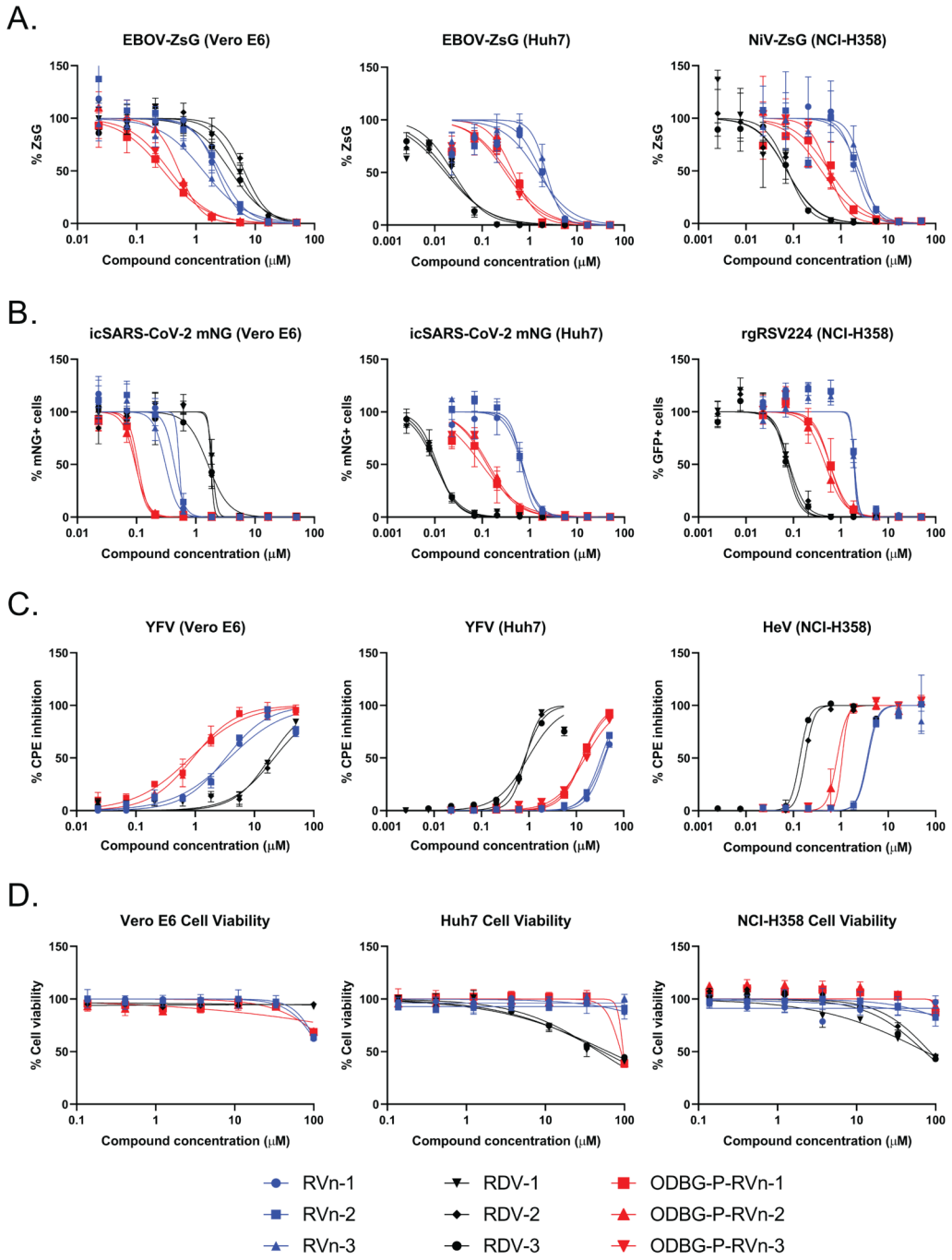

Figure 1. Comparison of antiviral activities of RVn, RDV, and ODBG-P-RVn in African green monkey (Vero E6), Human hepatoma (Huh7), and human bronchioalveolar carcinoma (NCI-H358) cell lines using reporter-based, image-based, and cytopathic effect assays. Representative dose response inhibition of virus replication and induction of cell cytotoxicity in by RVn (blue shapes), RDV (black shapes), and ODBG-P-RVn (red shapes). A) Direct measurement of green fluorescence reporter intensity by recombinant EBOV expressing ZsGreen protein in Vero E6 (left panel) and Huh7 (middle panel) cells, and recombinant NiV expressing ZsGreen protein in NCI-H358 (right panel) cells. B) Image-based counting of reporter fluorescence-positive cells infected with recombinant SARS-CoV-2 expressing mNeonGreen protein (Vero E6 and Huh7) and recombinant RSV expressing eGFP (NCI-H358). Infected cells treated with DMSO represented 100% fluorescence intensity signal and 100% fluorescence-positive cell counts. C) Inhibition of cytopathic effect (CPE) by YFV (Vero E6 and Huh7) and HeV (NCI-H358) measured by levels of cellular ATP (CellTiterGlo 2.0). Uninfected cells treated with DMSO served as 100% CPE inhibition. D) Compound cytotoxicity/cell viability measured by CellTiterGlo 2.0 assay. Dose response curves were fitted to the mean value of experiments performed in biological triplicate for each concentration in the 8-point 3-fold dilution series using a 4-parameter non-linear logistic regression curve with variable slope. Data points and error bars indicate the mean value and standard deviation of 3 biological replicates; each colored shape/line in the legend represents an independent experiment performed in biological triplicate.

Figure 2.

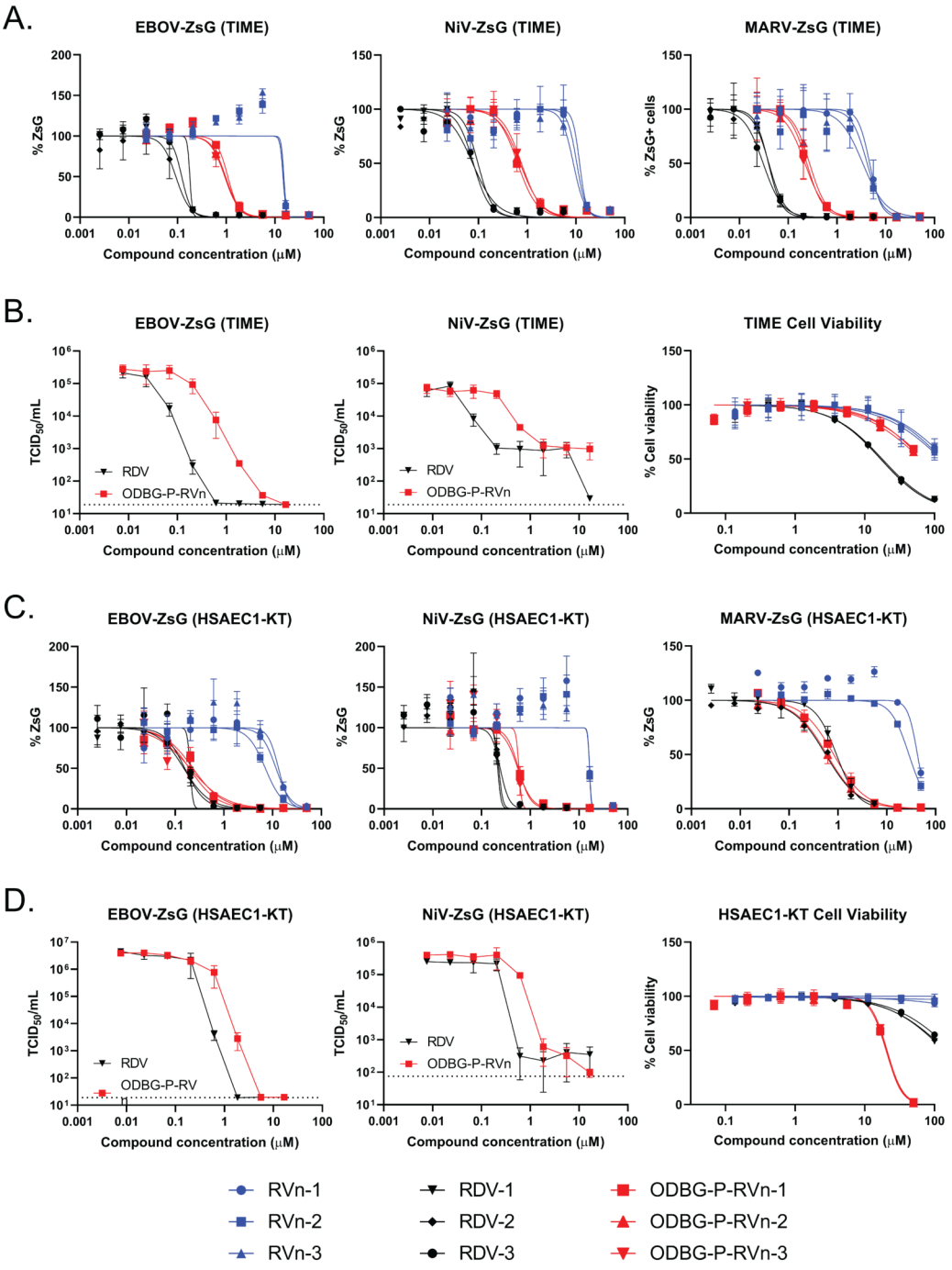

Figure 2. Comparison of cell-type dependent antiviral activities of RVn, RDV, and ODBG-P-RVn in primary-like hTERT-immortalized microvascular endothelial (TIME) and small airway epithelial cells (HSAEC1-KT). A) Representative dose response inhibition of recombinant EBOV, NiV, and MARV expressing ZsGreen protein in TIME cells. B) Infectious yield reduction by RDV and ODBG-P-RVn against EBOV-ZsG (left panel) and NiV-ZsG (middle panel). Compound cytotoxicity/cell viability (right panel) in TIME cells measured by CellTiterGlo 2.0 assay. C) Representative dose response inhibition of recombinant EBOV, NiV, and MARV expressing ZsGreen protein in HSAEC1-KT cells. (D) Infectious yield reduction by RDV and ODBG-P-RVn against EBOV-ZsG (left panel) and NiV-ZsG (middle panel) in HSAEC1-KT cells. Compound cytotoxicity/cell viability (right panel) in HSAEC1-KT cells measured by CellTiterGlo 2.0 assay. Dose response curves were fitted to the mean value of experiments performed in biological triplicate for each concentration in the 8-point 3-fold dilution series using a 4-parameter non-linear logistic regression curve with variable slope. Data points and error bars indicate the mean value and standard deviation of 3 or 4 biological replicates; each colored shape/line in the legend represents an independent experiment performed in biological triplicate. Infectious yield reduction assays were conducted once with biological quadruplicates.

Table 1. Mean antiviral activity of RVn, RDV, and ODBG-P-RVn in Vero E6, Huh7, and NCI-H358 cell lines

				Vero E6									Huh7/NCI-H358								
				RVn (GS-441524)			RDV (GS-5734)			ODBG-P-RVn			RVn (GS-441524)			RDV (GS-5734)			ODBG-P-RVn		
Virus Family	**Virus**	**Species/Variant**	**Assay**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: >100)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: >100)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: >100)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: >100/ >100)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: 54.2 ± 6.0/ 77.2 ± 5.3)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: 93.4 ± 3.0/ >100)**
Filoviridae	EBOV	Rec. Makona-ZsG	REP	2.03 ± 0.50	7.54 ± 1.09	49	5.15 ± 1.09	17.31 ± 0.89	>19	0.39 ± 0.10	1.71 ± 0.25	>258	1.84 ± 0.31	6.91 ± 1.79	>54	0.020 ± 0.003	0.16 ± 0.02	2710	0.37 ± 0.06	2.13 ± 0.37	251
	MARV	Rec. Bat371-ZsG	REP	0.96 ± 0.09	4.05 ± 1.42	104	2.16 ± 0.27	10.22 ± 2.02	>46	0.19 ± 0.04	0.81 ± 0.12	>521	1.92 ± 0.06	4.47 ± 0.48	>52	0.025 ± 0.002	0.075 ± 0.003	2128	0.33 ± 0.02	0.99 ± 0.09	285
Paramxyovirdae	NiV-M	Rec. Malaysia-ZsG	REP	1.10 ± 0.40	2.20 ± 1.05	73	5.87 ± 0.19	9.82 ± 0.43	>16	0.31 ± 0.04	0.78 ± 0.28	>196	2.43 ± 0.31	5.95 ± 1.10	>41	0.075 ± 0.001	0.31 ± 0.04	1026	0.50 ± 0.06	2.83 ± 1.39	>198
			CPE	0.48 ± 0.06	0.78 ± 0.19	207	3.34 ± 0.34	5.39 ± 0.29	>30	0.19 ± 0.01	0.30 ± 0.04	>522	ND	ND	N/A	ND	ND	N/A	ND	ND	N/A
	NiV-B	Bangladesh	CPE	0.52 ± 0.02	1.14 ± 0.02	192	2.84 ± 0.10	5.81 ± 0.44	>35	0.17 ± 0.01	0.38 ± 0.04	>599	3.42 ± 0.005	5.41 ± 0.29	>29	0.12 ± 0.0004	0.19 ± 0.01	661	0.82 ± 0.053	1.38 ± 0.05	>122
	HeV	1996	CPE	1.43 ± 0.17	12.06 ± 3.14	70	4.56 ± 0.20	17.58 ± 3.91	>22	0.37 ± 0.04	3.93 ± 1.98	>270	3.68 ± 0.08	6.33 ± 0.18	>27	0.16 ± 0.02	0.25 ± 0.03	491	0.95 ± 0.12	1.42 ± 0.03	>105
	MV	Rec. $rMV^{EZ}GFP(3)$	REP	0.58 ± 0.20	1.71 ± 0.07	172	4.97 ± 0.25	6.12 ± 0.3	>20	0.16 ± 0.03	0.21 ± 0.01	>609	0.88 ± 0.16	6.99 ± 1.90	>113	0.025 ± 0.007	0.13 ± 0.09	3074	0.12 ± 0.003	0.86 ± 0.22	>803
	hPIV3	Rec. JS-GFP	FFU	0.14 ± 0.01	0.28 ± 0.02	70	0.43 ± 0.09	0.90 ± 0.03	>232	0.026 ± 0.002	0.050 ± 0.002	>3896	1.43 ± 0.16	1.98 ± 0.05	>70	0.031 ± 0.002	0.052 ± 0.01	2458	0.22 ± 0.01	0.43 ± 0.02	>457
	MuV	Rec. IA2006-eGFP	FFU	5.11 ± 0.20	7.80 ± 0.64	18	16.81 ± 1.23	25.1 ± 1.97	>4.9	1.13 ± 0.04	2.53 ± 0.25	>56	9.3 ± 0.30	13.71 ± 0.24	>11	0.20 ± 0.003	0.24 ± 0.003	266	1.85 ± 0.11	2.24 ± 0.23	50
	SoSuV	Rec. 2012-ZsG	REP	1.00 ± 0.10	2.72 ± 0.62	100	5.31 ± 1.8	19.10 ± 9.31	>19	0.31 ± 0.089	0.80 ± 0.06	>325	2.06 ± 0.09	7.76 ± 1.11	>48	0.052 ± 0.01	0.13 ± 0.02	1042	0.52 ± 0.10	1.08 ± 0.15	180
Pneumoviridae	RSV	Rec. rgRSV0224 (A2)	FFU	0.49 ± 0.05	0.62 ± 0.01	206	1.80 ± 0.08	2.40 ± 0.27	>55	0.10 ± 0.02	0.22 ± 0.03	>997	1.93 ± 0.02	2.36 ± 0.08	>51	0.078 ± 0.004	0.17 ± 0.02	991	0.55 ± 0.057	1.41 ± 0.09	>180
Coronaviridae	SARS-CoV-2	Rec. icSARS-CoV-2 mNG (WA1)	FFU	0.42 ± 0.09	0.60 ± 0.06	236	1.77 ± 0.13	2.81 ± 0.78	>56	0.10 ± 0.005	0.16 ± 0.01	>997	0.69 ± 0.01	1.50 ± 0.20	>144	0.011 ± 0.001	0.035 ± 0.002	5073	0.12 ± 0.02	0.69 ± 0.07	778
Flaviviridae	YFV	17D	CPE	3.52 ± 0.24	30.25 ± 10.08	28	19.86 ± 1.73	>50	>5	0.87 ± 0.043	7.37 ± 1.59	>114	36.83 ± 2.85	>50	>2.7	0.88 ± 0.057	3.09 ± 1.47	62	14.11 ± 0.90	>50	6.6
Arenaviridae	LASV	Rec. Josiah-ZsG	REP	NI	NI	N/A	NI	NI	N/A	31.14 ± 7.79	>50	>3	NI	NI	N/A	2.87 ± 0.61	5.17 ± 0.33	19	NI	NI	N/A
Nairoviridae	CCHF	Rec. IbAr10200-ZsG	REP	NI	NI	N/A	NI	NI	N/A	NI	NI	N/A	NI	NI	N/A	NI	NI	N/A	NI	NI	N/A

EC_{50}, 50% effective inhibition concentration; EC_{90}, 90% effective inhibition concentration; CC_{50}, 50% cytotoxic concentration; SI, selective index = EC_{50}/CC_{50}; REP, reporter; CPE, cytopathic effect; FFU, focus-forming unit; ND, not determined; NI, no inhibition; N/A, not applicable; Rec, recombinant. Mean values with ± standard deviation values were derived from 3 independent experiments performed in biological triplicates except for NiV-B (NCI-H358), HeV (NCI-H358), and YFV (Vero E6)which were performed twice in biological triplicates. Data in red text derived from Huh7 cells, data in blue derived from NCI-H358 cells. REP/FFU/CPE assays were conducted between 72-144 hpi. EC_{50}, EC_{90}, and CC_{50} values were calculated using Graphpad Prism 9 software.

Table 2. Mean antiviral activity of RVn, RDV, and ODBG-P-RVn in primary-like hTERT-immortalized microvascular endothelial (TIME) and small airway epithelial (HSAEC1-KT) cell lines

				HSAEC1-KT									TIME								
				RVn (GS-441524)			RDV (GS-5734)			ODBG-P-RVn			RVn (GS-441524)			RDV (GS-5734)			ODBG-P-RVn		
Virus Family	**Virus**	**Species/Variant**	**Assay**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: >100)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: >100)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: 20.5 ± 0.29)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: >100)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: 17.2 ± 0.42)**	**EC_{50}**	**EC_{90}**	**SI (CC_{50}: >50)**
Filoviridae	EBOV	Rec. Makona-ZsG	REP	10.7 ± 2.62	21.79 ± 3.16	>9.3	0.17 ± 0.02	0.41 ± 0.14	>587	0.21 ± 0.02	1.06 ± 0.18	98	14.88 ± 0.28	17.24 ± 0.16	>3.36	0.13 ± 0.04	0.2 ± 0.01	132	0.99 ± 0.063	1.96 ± 0.043	>50
			VTR	ND	ND	N/A	0.11	0.82	>909	0.21	0.95	98	ND	ND	N/A	0.032	0.064	530	0.15	0.39	>324
	MARV	Rec. Bat371-ZsG	REP/FFU	35.53 ± 7.07	71.35 ± 1.28	>2.8	0.75 ± 0.19	2.92 ± 0.14	>133	0.71 ± 0.11	3.67 ± 0.49	29	5.2 ± 0.26	6.89 ± 0.86	>9.61	0.04 ± 0.003	0.086 ± 0.004	430	0.23 ± 0.036	0.66 ± 0.032	>213
Paramxyovirdae	NiV-M	Rec. Malaysia-ZsG	REP	16.46 ± 0.04	19.12 ± 0.05	>6.1	0.23 ± 0.01	0.31 ± 0.06	>440	0.57 ± 0.013	0.97 ± 0.21	36	13.53 ± 2.44	17.52 ± 0.77	>3.70	0.10 ± 0.01	0.20 ± 0.01	172	0.75 ± 0.05	2.01 ± 0.30	>66
			CPE	16.12 ± 4.21	78.1 ± 35.08	>6.2	0.31 ± 0.04	0.075 ± 0.004	>318	0.90 ± 0.07	10.22 ± 4.99	23	ND	ND	N/A	0.054	0.07	319	0.26	0.77	>195
			VTR	ND	ND	N/A	0.26	0.36	>379	0.47	0.77	44									
	NiV-B	Bangladesh	CPE	11.23 ± 0.63	33.6 ± 1.58	>8.9	0.21 ± 0.063	0.62 ± 0.20	>379	0.41 ± 0.039	1.71 ± 0.66	50									
	HeV	1994	CPE	11.52 ± 1.49	26.11 ± 4.44	>8.7	0.22 ± 0.04	0.65 ± 0.11	>463	0.42 ± 0.023	1.19 ± 0.061	49									
	MV	Rec. $rMV^{EZ}GFP(3)$	REP	4.98 ± 0.37	12.02 ± 2.7	>20	0.063 ± 0.02	0.128 ± 0.016	>1587	0.082 ± 0.026	0.29 ± 0.043	251									
	hPIV3	Rec. JS-GFP	FFU	4.96 ± 0.05	5.77 ± 0.06	>20	0.063 ± 0.001	0.074 ± 0.002	>1582	0.091 ± 0.009	0.20 ± 0.008	226									
Pneumoviridae	RSV	Rec. rgRSV0224 (A2)	FFU	4.92 ± 0.47	8.09 ± 0.68	>20	0.088 ± 0.026	0.21 ± 0.033	>1134	0.12 ± 0.008	0.34 ± 0.047	176									

EC_{50}, 50% effective inhibition concentration; EC_{90}, 90% effective inhibition concentration; CC_{50}, 50% cytotoxic concentration; SI, selective index = EC_{50}/CC_{50}; REP, reporter; CPE, cytopathic effect; FFU, focus-forming unit; VTR, virus titer reduction; ND, not determined; N/A, not applicable; Rec, recombinant. Mean values with ± standard deviation values were derived from a minimum of 3 independent experiments performed in biological triplicates. REP/FFU/CPE/VTR assays were conducted at 72 hpi. EC_{50}, EC_{90}, and CC_{50} values were calculated using Graphpad Prism 9 software.