Charting Brain Growth and Aging at High Spatial Precision

Saige Rutherford[1,2,3]*, **Charlotte Fraza**[1,2], **Richard Dinga**[1,2], **Seyed Mostafa Kia**[1,2,4], **Thomas Wolfers**[5,6], **Mariam Zabihi**[1,2], **Pierre Berthet**[5,6], **Amanda Worker**[7], **Serena Verdi**[8,9], **Derek Andrews**[10], **Laura Han**[11,12], **Johanna Bayer**[13,14], **Paola Dazzan**[7,15], **Phillip McGuire**[16], **Roel T. Mocking**[17], **Aaart Schene**[1,18], **Brenda W. Pennix**[11,12], **Chandra Sripada**[3], **Ivy F. Tso**[3], **Elizabeth R. Duval**[3], **Soo-Eun Chang**[3], **Mary Heitzeg**[3], **S. Alexandra Burt**[19], **Luke Hyde**[20], **David Amaral**[10], **Christine Wu Nordahl**[10], **Ole A. Andreasssen**[6,21], **Lars T. Westlye**[5,6,21], **Roland Zahn**[22], **Henricus G. Ruhe**[1,18]†, **Christian Beckmann**[1,2,23]†, **Andre F. Marquand**[1,2]*†

*For correspondence:
saige.rutherford@donders.ru.nl;
andre.marquand@donders.ru.nl

†These authors contributed equally to this work

[1]Donders Institute for Brain, Cognition, and Behavior, Radboud University, Nijmegen, the Netherlands; [2]Department of Cognitive Neuroscience, Radboud University Medical Center, Nijmegen, the Netherlands; [3]Department of Psychiatry, University of Michigan, Ann Arbor, MI, United States; [4]Department of Psychiatry, Utrecht University Medical Center, Utrecht, the Netherlands; [5]Department of Psychology, University of Oslo, Oslo, Norway; [6]Norwegian Center for Mental Disorders Research (NORMENT), University of Oslo, and Oslo University Hospital, Oslo, Norway; [7]Department of Psychological Medicine, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, United Kingdom; [8]8Centre for Medical Image Computing, Medical Physics and Biomedical Engineering, University College London, London, United Kingdom; [9]Dementia Research Centre, UCL Queen Square Institute of Neurology, London, United Kingdom; [10]The Medical Investigation of Neurodevelopmental Disorders (MIND) Institute and Department of Psychiatry and Behavioral Sciences, UC Davis School of Medicine, University of California Davis, Sacramento, CA, United States; [11]Amsterdam UMC, Vrije Universiteit, Psychiatry, Amsterdam Public Health Research Institute, Amsterdam, the Netherlands; [12]GGZ inGeest, Amsterdam Neuroscience, Amsterdam, the Netherlands; [13]Centre for Youth Mental Health, University of Melbourne, Melbourne, Australia; [14]Orygen Youth Health, Melbourne, Australia; [15]National Institute for Health Research Mental Health Biomedical Research Centre, South London and Maudsley National Health Service Foundation Trust and King's College, London, United Kingdom; [16]Department of Psychosis Studies, Institute of Psychiatry, King's College, London, United Kingdom; [17]Department of Psychiatry, Amsterdam UMC, Location AMC, Amsterdam, the Netherlands; [18]Department of Psychiatry, Radboud University Medical Center, Nijmegen, the Netherlands; [19]Department of Psychology, Michigan State University, East Lansing, MI, United States; [20]Department of Psychology, University of Michigan, Ann Arbor, MI, United States; [21]KG Jebsen Centre for Neurodevelopmental Disorders Research, Institute if Clinical Medicine, University of Oslo, Oslo, Norway; [22]Centre for Affective Disorders at the Institute of Psychiatry, King's College, London, United Kingdom; [23]Centre for Functional MRI of the Brain (FMRIB), Nuffield Department of Clinical Neurosciences, Wellcome Centre for Integrative Neuroimaging, University of Oxford, Oxford, United Kingdom

Abstract Defining reference models for population variation, and the ability to study individual deviations is essential for understanding inter-individual variability and its relation to the onset and progression of medical conditions. In this work, we assembled a reference cohort of neuroimaging data from 82 sites (N=58,836; ages 2-100) and use normative modeling to characterize lifespan trajectories of cortical thickness and subcortical volume. Models are validated against a manually quality checked subset (N=24,354) and we provide an interface for transferring to new data sources. We showcase the clinical value by applying the models to a transdiagnostic psychiatric sample (N=1,985), showing they can be used to quantify variability underlying multiple disorders whilst also refining case-control inferences. These models will be augmented with additional samples and imaging modalities as they become available. This provides a common reference platform to bind results from different studies and ultimately paves the way for personalized clinical decision making.

Introduction

Since their introduction more than a century ago, normative growth charts have become fundamental tools in pediatric medicine and also in many other areas of anthropometry (*Cole* (*2012*)). They provide the ability to quantify individual variation against centiles of variation in a reference population, which shifts focus away from group-level (e.g., case-control) inferences to the level of the individual. This idea been adopted and generalized in clinical neuroimaging and normative modelling is now established as an effective technique for providing inferences at the level of the individual in neuroimaging studies (*Marquand et al.* (*2016*, 2019)).

Although normative modelling can be used to estimate many different kinds of mappings – for example between behavioral scores and neurobiological readouts – normative models of brain development and aging are appealing considering that many brain disorders are grounded in atypical trajectories of brain development (*Insel* (*2014*)) and the association between cognitive decline and brain tissue in ageing and neurodegenerative diseases (*Jack et al.* (*2010*); *Karas et al.* (*2004*)). Indeed, normative modelling has been applied in many different clinical contexts, including charting the development of infants born pre-term (*Dimitrova et al.* (*2020*)) and dissecting the biological heterogeneity across cohorts of individuals with different brain disorders including schizophrenia, bipolar disorder, autism and attention deficit/hyperactivity disorder (*Bethlehem et al.* (*2020*); *Wolfers et al.* (*2021*); *Zabihi et al.* (*2019*)).

A hurdle to the widespread application of normative modelling is a lack of well-defined reference models to quantify variability across the lifespan and to compare results from different studies. Such models should: (i) accurately model population variation across large samples; (ii) be derived from widely accessible measures; (iii) provide the ability to be updated as additional data come on-line and (vi) be supported by easy-to-use software tools. In addition, they should quantify brain development and ageing at a high spatial resolution, so that different patterns of atypicality can be used to stratify cohorts and predict clinical outcomes with maximum precision. The purpose of this work is to introduce a set of reference models that satisfy these criteria.

To this end, we assemble a large neuroimaging dataset (Table 1) from 58,836 individuals across 82 scan sites covering the human lifespan (aged 2-100, Figure 1A) and fit normative models for cortical thickness and subcortical volumes derived from Freesurfer (version 6.0). We show the clinical utility of these models in a large transdiagnostic psychiatric sample (N=1,985). To maximize the utility of this contribution, we distribute model coefficients freely along with a set of software tools to enable researchers to derive subject-level predictions for new datasets against a set of common reference models.

Table 1. Sample Description and Demographics. mQC refers to the manual quality checked subset of the full sample. 'All' rows = Train + Test. Clinical refers to the transdiagnostic psychiatric sample (diagnostic details in Figure 2A).

		N (subjects)	N (sites)	Sex (%F, %M)	Age (Mean, s.d)
Full	All	58,836	82		
	Training set	29,418	82	51.1, 48.9	46.9, 24.4
	Test set	29,416	82	50.9, 49.1	46.9, 24.4
mQC	All	24,354	59		
	Training set	12,177	59	50.2, 49.8	30.2, 24.1
	Test set	12,177	59	50.4, 49.4	30.1, 24.2
Clinical	Test set	1,985	24	38.9, 61.1	30.5, 14.1

Results

We split the available data into training and test sets, stratifying the split by site, such that all sites are equally represented in the training and test sets (Table 1, Supplementary Table 2, Supplementary Table 3, and Supplementary Table 4). After careful automated and manual quality checking procedures (see methods), we then fit a normative model using a set of covariates (age, gender, and fixed effects for site) to predict cortical thickness and subcortical volume for each parcel in a high resolution atlas derived from the Freesurfer software package (*Destrieux et al.* (*2010*)). We employed a warped Bayesian linear regression model so as to accurately model both non-linear effects and non-Gaussian distribution of the imaging phenotype (*Fraza et al.* (*2021*)), whilst accounting for the well-known effects of different scanners on neuroimaging data (*Bayer et al.* (*2021*); *Kia et al.* (*2021*)). These models are summarized in Figure 3, Supplementary Table 5, Supplementary Table 6, Supplementary Table 7, and Supplementary Table 8.

We validate our models with several careful procedures: first, we report out of sample metrics; second, we perform a supplementary analysis on a subset of participants for whom input data had undergone manual quality checking by an expert rater (Table 1 - mQC). Third, each model fit was evaluated using metrics (Figure 3, Supplementary Table 5, Supplementary Table 6, Supplementary Table 7, and Supplementary Table 8) that quantify central tendency and distributional accuracy (*Dinga et al.* (*2021*); *Fraza et al.* (*2021*)). We also estimated separate models for males and females, which indicate that sex effects are adequately modeled using a global offset. Finally, to facilitate independent validation, we packaged pretrained models and code for transferring to new samples into an open resource for use by the community and demonstrate how to transfer the models to new samples (i.e., data not present in the initial training set).

Our models provide the opportunity for mapping the diverse trajectories of different brain areas. Several examples are shown in Figure 1 C and D which align with known patterns of development and aging (*Ducharme et al.* (*2016*); *Gogtay et al.* (*2004*); *Tamnes et al.* (*2010*)). Moreover, across the cortex and subcortex our model fits well, explaining up to 80% of the variance (minimum 12%) out of sample (Figure 3, Supplementary Table 5, Supplementary Table 6, Supplementary Table 7, and Supplementary Table 8 for full details).

A goal of this work is to develop normative models that can be applied to many different clinical conditions. To showcase this, we apply the model to a transdiagnostic psychiatric cohort (Table 1 – Clinical; Figure 2A) resulting in personalized, whole-brain deviation maps that can be used to understand inter-individual variability (e.g., for stratification) and to quantify group separation (e.g., case-control effects). To demonstrate this, for each clinical group, we summarized the individual deviations within that group by computing the proportion of subjects that have deviations in each region and compare to matched (same sites) controls in the test set (Figure 2B-C). Additionally, we performed case-control comparisons on the raw cortical thickness and subcortical volumes, and on the deviation maps (Figure 2D), again against a matched sample from the test set. This demonstrates the advantages of using normative models for investigating individual differences in

Figure 1. Normative Model Overview. A) Age distribution (x-axis) of each site (y-axis) in the full model train and test sets. B) Age distribution of each site in the clinical test set. C-D) Examples of lifespan trajectories of brain regions. Age is shown on x-axis and predicted thickness (or volume) values are on the y-axis. Centiles of variation are plotted for each region. In panel C, we show that sex differences between females (red) and males (blue) are most pronounced when modeling large scale features such as mean cortical thickness across the entire cortex or total gray matter volume. These sex differences manifest as a shift in the mean in that the shape of these trajectories is the same for both sexes, as determined by sensitivity analyses where separate normative models were estimated for each sex. The explained variance (in full test set) of the whole cortex and subcortex is highlighted inside the circle of panel D. All plots within the circle share the same color scale. E) The distribution of evaluation metrics in the full test set, separated into left and right hemispheres and subcortical regions, with the skew and excess kurtosis being measures that depict the accuracy of the estimated shape of the model, ideally both of these would be around zero.

psychiatry, i.e., quantifying clinically relevant information at the level of each individual. For most
diagnostic groups, the z-statistics derived from the normative deviations also provided stronger
case-control effects than the raw data. This shows the importance of accurate modeling of popula-
tion variance across multiple clinically relevant dimensions. The individual-level deviations provide
complimentary information to the group effects, which aligns with previous work (*Wolfers et al.*
(*2020*); *Zabihi et al.* (*2020*); *Wolfers et al.* (*2018*)). We note that a detailed description of the clinical
significance of our findings is beyond the scope of this work and will be presented separately.

Discussion

In this work we create lifespan brain charts of cortical thickness and subcortical volume derived
from structural MRI, to serve as reference models. Multiple datasets were joined to build a mega-
site lifespan reference cohort to provide good coverage of the lifespan. We applied the reference
cohort models to clinical datasets and demonstrate the benefits of normative modeling in addition
to standard case-control comparisons. All models, including documentation and code, are made
available to the research community.

We identify three main strengths of our approach. First, our large lifespan dataset provides
high anatomical specificity, necessary for discriminating between conditions, predicting outcomes,
and stratifying subtypes. Second, our models are flexible in that they can model non-Gaussian
distributions, can easily be transferred to new sites, and are built on validated analytical techniques
and software tools (*Fraza et al.* (*2021*); *Kia et al.* (*2021*); *Marquand et al.* (*2019*)). Third, we show
the general utility of this work in that it provides the ability to map individual variation whilst also
improving case control inferences across multiple disorders.

In recent work, a large consortium established lifespan brain charts that are complementary to

A) Clinical Validation Sample Description

	N (subjects)	N (sites)	Sex (F/M)	Age (m, s.d)
Attention Deficit Disorder (ADHD)	111	4	71/40	17.6, 12.4
Autism Spectrum Disorder (ASD)	450	6	389/61	19.1, 11.1
Early Psychosis (EP)	207	5	112/165	29.4, 10.7
Schizophrenia (SZ)	469	8	193/276	31.9, 9.5
Bipolar Disorder (BD)	249	4	138/111	34.5, 10.9
Major Depression Disorder (MDD)	499	9	260/239	40.2, 13.4

B) Percent of Individuals with Extreme Positive Deviations (Z-score > 2)

C) Percent of Individuals with Extreme Negative Deviations (Z-score < -2)

D) Control vs. Patient Univariate T-Tests

Figure 2. Normative Modeling in Clinical Cohorts. Reference brain charts were transferred to several clinical samples (panel A). Patterns of extreme deviations were summarized for each clinical group and compared to matched control groups (from the same sites). Panel B) shows extreme positive deviations (thicker/larger than expected) and panel C) shows the extreme negative deviation (thinner/smaller than expected) patterns. Panel D) shows the significant (FDR corrected p<0.05) results of classical case-control methods (mass-univariate t-tests) on the true cortical thickness data (top row) and on the deviations scores (bottom row). There is unique information added by each approach which becomes evident when noticing the maps in panels B-D are not identical. ADHD=Attention Deficit Hyperactive Disorder, ASD=Autism Spectrum Disorder, EP=Early Psychosis, SZ=Schizophrenia, BD=Bipolar Disorder, MDD=Major Depressive Disorder.

149   our approach (*Bethlehem et al.* (*2021*)). Benefits of their work include the use of a large sample
150   and good coverage of the peri-natal life stages. This was used to provide estimates of brain growth
151   in terms of global features (e.g., brain volume), which is valuable for applications where quantify-
152   ing global brain development or ageing is of interest but has limited spatial precision. In contrast,

Figure 3. Evaluation Metrics Across All Test Sets. The distribution of evaluation metrics in 3 different test sets (full, mQC, and Transfer) separated into left and right hemispheres and subcortical regions, with the skew and excess kurtosis being measures that depict the accuracy of the estimated shape of the model, ideally both of these would be around zero.

we focus on providing spatially specific estimates across the post-natal lifespan which provides
fine-grained anatomical estimates of deviation that may be valuable for understanding the biolog-
ical basis for mental disorders where individual patterns are widespread (e.g., not all individuals
deviate in the same regions).
We also identify limitations of this work. First, we view the word "normative" as problematic.
This language implies that there are normal and abnormal brains, a potentially problematic as-
sumption. As indicated in Figure 2, there is considerable individual variability and heterogeneity
among trajectories. We encourage the use of the phrase 'reference cohort' over 'normative model'.
Second, to provide coverage of the lifespan the curated dataset is based on aggregating existing
data, meaning there is unavoidable sampling bias. Race, education, and socioeconomic variables
were not fully available for all included datasets, however, given that data were compiled from
research studies, they are likely samples drawn predominantly from Western, Educated, Indus-
trialized, Rich, and Democratic (WEIRD) societies (*Henrich et al.* (*2010*)) and future work should
account for these factors. By sampling both healthy population samples and case-control studies,
we achieve a reasonable estimate of variation across individuals, however, downstream analyses
should consider the nature of the reference cohort and whether it is appropriate for the target
sample. Finally, although the models presented in this study are comprehensive, they are only
the first step, and we will augment our repository with more diverse data, different features and
modelling advances as these become available.

Methods and Materials

Data from 82 sites were combined to create the initial full sample. These sites are described in detail in Supplemental Table 5, including the sample size, age (mean and standard deviation), and sex distribution of each site. Many sites were pulled from publicly available data sets including ABCD, ABIDE, ADHD200, CAMCAN, CMI-HBN, HCP-Aging, HCP-Development, HCP-Early Psychosis, HCP-Young Adult, IXI, NKI-RS, Oasis, OpenNeuro, PNC, SRPBS, and UK Biobank. Other included data come from studies conducted at the University of Michigan (*Duval et al.* (*2018*); *Rutherford et al.* (*2020*); *Tomlinson et al.* (*2020*); *Tso et al.* (*2021*); *Weigard et al.* (*2021*); *Zucker et al.* (*1996*)), University of California Davis (*Nordahl et al.* (*2020*)), University of Oslo (*Nesvåg et al.* (*2017*)), King's College London (*Green et al.* (*2012*); *Lythe et al.* (*2015*)), and Amsterdam University Medical Center (*Mocking et al.* (*2016*)). Full details regarding sample characteristics, diagnostic procedures and acquisition protocols can be found in the publications associated with each of the studies. Equal sized training and testing data sets (split half) were created using scikit-learn's train_test_split function, stratifying on the site variable. It is important to stratify based on site, not only study (*Bethlehem et al.* (*2021*)), as many of the public studies (i.e., ABCD) include several sites, thus modeling study does not adequately address MRI scanner confounds.

The clinical validation sample consisted of a subset of the full data set (described in detail in Figure 2A and Supplemental Table 3). Studies (sites) contributing clinical data included: Autism Brain Imaging Database Exchange (ABIDE GU, KKI, NYU, USM), ADHD200 (KKI, NYU), CNP, SRPBS (CIN, COI, KTT, KUT, HKH, HRC, HUH, SWA, UTO), Delta (AmsterdamUMC), Human Connectome Project Early Psychosis (HCP-EP BWH, IU, McL, MGH), KCL, University of Michigan Schizophrenia Gaze Processing (UMich_SZG), and TOP (University of Oslo).

In addition to the sample-specific inclusion criteria, inclusion criteria for the full sample was based on participants having basic demographic information (age and sex), a T1-weighted MRI volume, and Freesurfer output directories that include summary files which represent left and right hemisphere cortical thickness values of the Destrieux parcellation and subcortical volumetric values (aseg.stats, lh.aparc.a2009s.stats, rh.aparc.a2009s.stats). Freesurfer image analysis suite (version 6.0) was used for cortical reconstruction and volumetric segmentation for all studies. The technical details of these procedures are described in prior publications (*Dale et al.* (*1999*); *Fischl and Dale* (*2000*); *Fischl et al.* (*2002*)). UK Biobank was the only study for which Freesurfer was not run by the authors. Freesurfer functions aparcstats2table and asegstats2table were run to extract cortical thickness from the Destrieux parcellation (*Destrieux et al.* (*2010*)) and subcortical volume for all participants into CSV files. These files were inner merged with the demographic files, using Pandas, and NaN rows were dropped.

Quality control (QC) is an important consideration for large samples and is an active research area (*Alfaro-Almagro et al.* (*2018*); *Klapwijk et al.* (*2019*); *Rosen et al.* (*2018*)). We consider manual quality checking of images both prior to and after preprocessing to be the gold standard. However, this is labor intensive and prohibitive for very large samples. Therefore, in this work we adopt a pragmatic and multi-pronged approach to QC. First, a subset of the full data set underwent manual quality checking (mQC, described in Supplemental Table 4) by author S.R. using Papaya, a JavaScript based image viewer. Each subject's T1w volume was viewed in 3D volumetric space, with the Freesurfer brain.finalsurfs file as an overlay, to check for obvious quality issues such as excessive motion, ghosting or ringing artifacts. Example scripts used for quality checking can be found on GitHub. The ABCD study data distributes a variable (freesqc01.txt; fsqc_qc==1/0) that represents manual quality checking (pass/fail) of the T1w volume and Freesurfer data, thus this data set was added into our manual quality checked data set bringing the sample size to 24,354 individuals passing manual quality checks. Although this has a reduced sample, we consider this to be a gold standard sample in that every single scan has been checked manually. All inferences reported in this manuscript were validated against this sample. Second, for the full sample, we adopted an automated QC procedure that quantifies image quality based on the Freesurfer Euler

Characteristic (EC), which has been shown to be an excellent proxy for manual labelling of scan quality (*Monereo-Sánchez et al.* (*2021*); *Rosen et al.* (*2018*)) and is the most important feature in automated scan quality classifiers (*Klapwijk et al.* (*2019*)). Since the distribution of the EC varies across sites, we adopt a simple approach that involves scaling and centering the distribution over the EC across sites and removing samples in the tail of the distribution (see (*Kia et al.* (*2021*)) for details).

Normative modeling was run using python 3.8 and the PCNtoolkit package (version 0.20). Bayesian Linear Regression (BLR) with likelihood warping was used to predict cortical thickness and subcortical volume from a vector of covariates (age, sex, site). For a complete mathematical description and explanation of this implementation see (*Fraza et al.* (*2021*)). Briefly, for each brain region of interest (cortical thickness or subcortical volume), y is predicted as:

$$y = \omega^{\mathsf{T}} \varphi(x) + \epsilon \tag{1}$$

Where ω^{T} is the estimated weight vector, $\varphi(x)$ is a basis expansion of the of covariate vector x, consisting of a B-spline basis expansion (cubic spline with 5 evenly spaced knots) to model non-linear effects of age, and $\epsilon = \eta(\theta, \beta)$ a Gaussian noise distribution with mean zero and noise precision term β (the inverse variance). A likelihood warping approach (*Rios and Tobar* (*2019*); *Snelson et al.* (*2003*)) was used to model non-Gaussian effects. This involves applying a bijective nonlinear warping function to the non-Gaussian response variables to map them to a Gaussian latent space where inference can be performed in closed form. We employed a 'sinarcsinsh' warping function, which is equivalent to the SHASH distribution commonly used in the generalized additive modeling literature (*Jones and Pewsey* (*2009*)) and which we have found to perform well in prior work (*Dinga et al.* (*2021*); *Fraza et al.* (*2021*)). Site variation was modeled using fixed effects, which we have shown in prior work provides relatively good performance (*Kia et al.* (*2021*)), although random effects for site may provide additional flexibility at higher computational cost. A fast numerical optimization algorithm was used to optimize hyperparameters (L-BFGS). Computational complexity of hyperparameter optimization was controlled by minimizing the negative log likelihood. Deviation scores (Z-scores) are calculated for the n_{t_h} subject, and d_{t_h} brain area, in the test set as:

$$Z_{n_d} = \frac{y_{n_d} - \hat{y_{n_d}}}{\sqrt{(\theta_d)^2 + (\theta_{*_d})^2}} \tag{2}$$

Where y_{n_d} is the true response, $\hat{y_{n_d}}$ is the predicted mean, θ_d^2 is the estimated noise variance (reflecting uncertainty in the data), and $\theta_{*_d}^2$ is the variance attributed to modeling uncertainty. Model fit for each brain region was evaluated by calculating the explained variance (which measures central tendency), the mean squared log-loss (MSLL, central tendency and variance) plus skew and kurtosis of the deviation scores (equation 2) which measures how well the shape of the regression function matches the data (*Dinga et al.* (*2021*)). Note that for all models, we report out of sample metrics (Figure 3, Supplementary Table 5, Supplementary Table 6, Supplementary Table 7, and Supplementary Table 8).

To provide a summary of individual variation within each clinical group, deviation scores were summarized for each clinical group (Figure 2B-C) by first separating them into positive and negative deviations, counting how many subjects had an extreme deviation (positive extreme deviation defined as $Z > 2$, negative extreme deviation as $Z < -2$) at a given ROI, and then dividing by the group size to show the percentage of individuals with extreme deviations at that brain area. Controls from the same sites as the patient groups were summarized in the same manner for comparison. We also performed classical case vs. control group difference testing on the true data and on the deviation scores (Figure 2D) and thresholded results at a Benjamini-Hochberg false discovery rate (FDR) of $p < 0.05$. Note that in both cases, we directly contrast each patient group to their matched controls to avoid nuisance variation confounding any reported effects (e.g., sampling characteristics, demographic differences).

All pretrained models and code are shared online with straightforward directions for transferring to new sites. Given a new set of data (e.g., sites not present in the training set), this is done by first applying the warp parameters estimating on the training data to the new dataset, adjusting the mean and variance in the latent Gaussian space, then (if necessary) warping the adjusted data back to the original space, which is similar to the approach outlined in (*Dinga et al.* (*2021*)). Note that to remain unbiased, this should be done on a held-out calibration dataset. To illustrate this procedure, we apply this approach to predicting a subset of sites in the 1000 functional connectomes project (*Biswal et al.* (*2010*)) that were not used during the model estimation step. These results are reported in Supplemental Figure 4 (bottom row). We also distribute scripts for this purpose in the GitHub Repository associated with this manuscript. Furthermore, to promote the use of these models and remove barriers to using them, we have set up access to the pretrained models and code for transferring to users' own data, using Google Colab, a free, cloud-based platform for running python notebooks. This eliminates the need to install python/manage package versions and only requires users to have a personal computer with stable internet connection.

Acknowledgments

This research was supported by grants from the European Research Council (ERC, grant "MENTAL-PRECISION" 10100118 and "BRAINMINT" 802998), the Wellcome Trust under an Innovator award ("BRAINCHART", 215698/Z/19/Z) and a Strategic Award (098369/Z/12/Z), the Dutch Organisation for Scientific Research (VIDI grant 016.156.415) the Research Council of Norway (223273, 249795, 298646, 300768, 276082), the South-Eastern Norway Regional Health Authority (2014097, 2015073, 2016083, 2019101), the KG Jebsen Stiftelsen, an Autism Center of Excellence grant awarded by the National Institute of Child Health and Development (NICHD) (P50 HD093079) as well as the National Institute of Mental Health (R01MH104438, R01MH103371). TW also gratefully acknowledges the Niels Stensen Fellowship as well as the European Union's Horizon 2020 research and innovation programme under the Marie Sklodowska-Curie Grant agreement No. 895011. RZ was funded by Medical Research Council grant (G0902304). IFT was funded by National Institute of Mental Health K23MH108823. SC was funded by National Institute on Deafness and other Communication Disorders (NIDCD/NIH) grant R01DC011277. CS was funded by the National Institute of Mental Health R01MH107741. MTwiNS was supported by the National Institute of Mental Health and the Office of the Director National Institute of Health, under Award Number UG3MH114249 and the Eunice Kennedy Shriver National Institute of Child Health & Human Development of the National Institutes of Health under Award Number R01HD093334 to SAB and LWH. RJTM was funded by an ABC Talent Grant. The ABCD Study is supported by the National Institutes of Health (NIH) award numbers U01DA041022, U01DA041028, U01DA041048, U01DA041089, U01DA041106, U01DA041117, U01DA041120, U01DA041134, U01DA041148, U01DA041156, U01DA041174, U24DA041123, and U24DA041147.

Conflicts of Interest

CFB is director and shareholder of SBGNeuro Ltd. OAA is a consultant for HealthLytix and received speaker's honorarium from Lundbeck and Sunovion. HGR received speaker's honorarium from Lundbeck and Janssen. The other authors report no conflicts of interest.

References

Alfaro-Almagro F, Jenkinson M, Bangerter NK, Andersson JLR, Griffanti L, Douaud G, Sotiropoulos SN, Jbabdi S, Hernandez-Fernandez M, Vallee E, Vidaurre D, Webster M, McCarthy P, Rorden C, Daducci A, Alexander DC, Zhang H, Dragonu I, Matthews PM, Miller KL, et al. Image processing and Quality Control for the first 10,000 brain imaging datasets from UK Biobank. NeuroImage. 2018 Feb; 166:400–424. https://linkinghub.elsevier.com/retrieve/pii/S1053811917308613, doi: 10.1016/j.neuroimage.2017.10.034.

Bayer JMM, Dinga R, Kia SM, Kottaram AR, Wolfers T, Lv J, Zalesky A, Schmaal L, Marquand A. Accommodating site variation in neuroimaging data using normative and hierarchical Bayesian models.

bioRxiv. 2021 May; p. 2021.02.09.430363. https://www.biorxiv.org/content/10.1101/2021.02.09.430363v2, doi: 10.1101/2021.02.09.430363, publisher: Cold Spring Harbor Laboratory Section: Confirmatory Results.

Bethlehem R, Seidlitz J, White SR, Vogel JW, Anderson KM, Adamson C, Adler S, Alexopoulos GS, Anagnostou E, Areces-Gonzalez A, Astle DE, Auyeung B, Ayub M, Ball G, Baron-Cohen S, Beare R, Bedford SA, Benegal V, Beyer F, Bae JB, et al. Brain charts for the human lifespan. bioRxiv. 2021 Jun; p. 2021.06.08.447489. https://www.biorxiv.org/content/10.1101/2021.06.08.447489v1, doi: 10.1101/2021.06.08.447489.

Bethlehem R, Seidlitz J, Romero-Garcia R, Trakoshis S, Dumas G, Lombardo MV. A normative modelling approach reveals age-atypical cortical thickness in a subgroup of males with autism spectrum disorder. Communications Biology. 2020 Sep; 3(1):486. doi: 10.1038/s42003-020-01212-9.

Biswal BB, Mennes M, Zuo XN, Gohel S, Kelly C, Smith SM, Beckmann CF, Adelstein JS, Buckner RL, Colcombe S, Dogonowski AM, Ernst M, Fair D, Hampson M, Hoptman MJ, Hyde JS, Kiviniemi VJ, Kotter R, Li SJ, Lin CP, et al. Toward discovery science of human brain function. Proceedings of the National Academy of Sciences. 2010 Mar; 107(10):4734–4739. http://www.pnas.org/cgi/doi/10.1073/pnas.0911855107, doi: 10.1073/pnas.0911855107.

Cole TJ. The development of growth references and growth charts. Annals of Human Biology. 2012 Sep; 39(5):382–394. doi: 10.3109/03014460.2012.694475.

Dale AM, Fischl B, Sereno MI. Cortical surface-based analysis. I. Segmentation and surface reconstruction. Neuroimage. 1999 Feb; 9(2):179–194. http://dx.doi.org/10.1006/nimg.1998.0395, doi: 10.1006/nimg.1998.0395.

Destrieux C, Fischl B, Dale A, Halgren E. Automatic parcellation of human cortical gyri and sulci using standard anatomical nomenclature. NeuroImage. 2010 Oct; 53(1):1–15. doi: 10.1016/j.neuroimage.2010.06.010.

Dimitrova R, Pietsch M, Christiaens D, Ciarrusta J, Wolfers T, Batalle D, Hughes E, Hutter J, Cordero-Grande L, Price AN, Chew A, Falconer S, Vecchiato K, Steinweg JK, Carney O, Rutherford MA, Tournier JD, Counsell SJ, Marquand AF, Rueckert D, et al. Heterogeneity in Brain Microstructural Development Following Preterm Birth. Cerebral Cortex (New York, NY: 1991). 2020 Jul; 30(9):4800–4810. doi: 10.1093/cercor/bhaa069.

Dinga R, Fraza CJ, Bayer JMM, Kia SM, Beckmann CF, Marquand AF. Normative modeling of neuroimaging data using generalized additive models of location scale and shape. Neuroscience; 2021.

Ducharme S, Albaugh MD, Nguyen TV, Hudziak JJ, Mateos-Pérez JM, Labbe A, Evans AC, Karama S, Brain Development Cooperative Group. Trajectories of cortical thickness maturation in normal brain development–The importance of quality control procedures. NeuroImage. 2016 Jan; 125:267–279. doi: 10.1016/j.neuroimage.2015.10.010.

Duval ER, Joshi SA, Block SR, Abelson JL, Liberzon I. Insula Activation is Modulated by Attention Shifting in Social Anxiety Disorder. Journal of anxiety disorders. 2018 May; 56:56–62. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5985215/, doi: 10.1016/j.janxdis.2018.04.004.

Fischl B, Dale AM. Measuring the thickness of the human cerebral cortex from magnetic resonance images. Proc Natl Acad Sci U S A. 2000 Sep; 97(20):11050–11055. http://dx.doi.org/10.1073/pnas.200033797, doi: 10.1073/pnas.200033797.

Fischl B, Salat DH, Busa E, Albert M, Dieterich M, Haselgrove C, van der Kouwe A, Killiany R, Kennedy D, Klaveness S, Montillo A, Makris N, Rosen B, Dale AM. Whole Brain Segmentation. Neuron. 2002; 33(3):341–355. http://dx.doi.org/10.1016/s0896-6273(02)00569-x, doi: 10.1016/s0896-6273(02)00569-x.

Fraza CJ, Dinga R, Beckmann CF, Marquand AF. Warped Bayesian Linear Regression for Normative Modelling of Big Data. bioRxiv. 2021 Apr; p. 2021.04.05.438429. https://www.biorxiv.org/content/10.1101/2021.04.05.438429v1, doi: 10.1101/2021.04.05.438429.

Gogtay N, Giedd JN, Lusk L, Hayashi KM, Greenstein D, Vaituzis AC, Nugent TF, Herman DH, Clasen LS, Toga AW, Rapoport JL, Thompson PM. Dynamic mapping of human cortical development during childhood through early adulthood. Proceedings of the National Academy of Sciences. 2004 May; 101(21):8174–8179.

Green S, Lambon Ralph MA, Moll J, Deakin JFW, Zahn R. Guilt-Selective Functional Disconnection of Anterior Temporal and Subgenual Cortices in Major Depressive Disorder. Archives of General Psychiatry. 2012 Oct; 69(10):1014–1021. https://doi.org/10.1001/archgenpsychiatry.2012.135, doi: 10.1001/archgenpsychiatry.2012.135.

364 **Henrich J**, Heine SJ, Norenzayan A. The weirdest people in the world? Behavioral and Brain Sciences.
365 2010 Jun; 33(2-3):61–83. https://www.cambridge.org/core/journals/behavioral-and-brain-sciences/article/abs/
366 weirdest-people-in-the-world/BF84F7517D56AFF7B7EB58411A554C17, doi: 10.1017/S0140525X0999152X,
367 publisher: Cambridge University Press.

368 **Insel TR**. Mental disorders in childhood: shifting the focus from behavioral symptoms to neurodevelopmental
369 trajectories. JAMA. 2014 May; 311(17):1727–1728. doi: 10.1001/jama.2014.1193.

370 **Jack CR**, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, Petersen RC, Trojanowski JQ. Hypothetical
371 model of dynamic biomarkers of the Alzheimer's pathological cascade. The Lancet Neurology. 2010 Jan;
372 9(1):119–128. doi: 10.1016/S1474-4422(09)70299-6.

373 **Jones MC**, Pewsey A. Sinh-arcsinh distributions. Biometrika. 2009; 96(4):761–780. https://www.jstor.org/stable/
374 27798865.

375 **Karas GB**, Scheltens P, Rombouts SaRB, Visser PJ, van Schijndel RA, Fox NC, Barkhof F. Global and local gray
376 matter loss in mild cognitive impairment and Alzheimer's disease. NeuroImage. 2004 Oct; 23(2):708–716.
377 doi: 10.1016/j.neuroimage.2004.07.006.

378 **Kia SM**, Huijsdens H, Rutherford S, Dinga R, Wolfers T, Mennes M, Andreassen OA, Westlye LT, Beckmann
379 CF, Marquand AF. Federated Multi-Site Normative Modeling using Hierarchical Bayesian Regression.
380 bioRxiv. 2021 May; p. 2021.05.28.446120. https://www.biorxiv.org/content/10.1101/2021.05.28.446120v1, doi:
381 10.1101/2021.05.28.446120.

382 **Klapwijk ET**, van de Kamp F, van der Meulen M, Peters S, Wierenga LM. Qoala-T: A supervised-learning
383 tool for quality control of FreeSurfer segmented MRI data. NeuroImage. 2019 Apr; 189:116–129. doi:
384 10.1016/j.neuroimage.2019.01.014.

385 **Lasagna CA**, McLaughlin MM, Deng WY, Whiting EL, Tso IF. Deconstructing eye contact perception: Measur-
386 ing perceptual precision and self-referential tendency using an online psychophysical eye contact detection
387 task. PLOS ONE. 2020 Mar; 15(3):e0230258. https://journals.plos.org/plosone/article?id=10.1371/journal.pone.
388 0230258, doi: 10.1371/journal.pone.0230258, publisher: Public Library of Science.

389 **Lythe KE**, Moll J, Gethin JA, Workman CI, Green S, Lambon Ralph MA, Deakin JFW, Zahn R. Self-blame–Selective
390 Hyperconnectivity Between Anterior Temporal and Subgenual Cortices and Prediction of Recurrent Depres-
391 sive Episodes. JAMA Psychiatry. 2015 Nov; 72(11):1119–1126. https://doi.org/10.1001/jamapsychiatry.2015.
392 1813, doi: 10.1001/jamapsychiatry.2015.1813.

393 **Marquand AF**, Kia SM, Zabihi M, Wolfers T, Buitelaar JK, Beckmann CF. Conceptualizing mental disor-
394 ders as deviations from normative functioning. Molecular Psychiatry. 2019 Oct; 24(10):1415–1424. doi:
395 10.1038/s41380-019-0441-1.

396 **Marquand AF**, Rezek I, Buitelaar J, Beckmann CF. Understanding Heterogeneity in Clinical Cohorts Using
397 Normative Models: Beyond Case-Control Studies. Biological Psychiatry. 2016 Oct; 80(7):552–561. doi:
398 10.1016/j.biopsych.2015.12.023.

399 **Lever-van Milligen BA**, Verhoeven JE, Schmaal L, van Velzen LS, Révész D, Black CN, Han LKM, Horsfall M,
400 Batelaan NM, van Balkom AJLM, van Schaik DJF, van Oppen P, Penninx BWJH. The impact of depression
401 and anxiety treatment on biological aging and metabolic stress: study protocol of the Mood treatment with
402 antidepressants or running (MOTAR) study. BMC Psychiatry. 2019 Dec; 19(1):425. https://doi.org/10.1186/
403 s12888-019-2404-0, doi: 10.1186/s12888-019-2404-0.

404 **Mocking RJT**, Figueroa CA, Rive MM, Geugies H, Servaas MN, Assies J, Koeter MWJ, Vaz FM, Wichers M, van
405 Straalen JP, de Raedt R, Bockting CLH, Harmer CJ, Schene AH, Ruhé HG. Vulnerability for new episodes in
406 recurrent major depressive disorder: protocol for the longitudinal DELTA-neuroimaging cohort study. BMJ
407 open. 2016 Mar; 6(3):e009510. doi: 10.1136/bmjopen-2015-009510.

408 **Monereo-Sánchez J**, de Jong JJA, Drenthen GS, Beran M, Backes WH, Stehouwer CDA, Schram MT, Linden DEJ,
409 Jansen JFA. Quality control strategies for brain MRI segmentation and parcellation: Practical approaches
410 and recommendations - insights from the Maastricht study. NeuroImage. 2021 Aug; 237:118174. doi:
411 10.1016/j.neuroimage.2021.118174.

412 **Nesvåg R**, Jönsson EG, Bakken IJ, Knudsen GP, Bjella TD, Reichborn-Kjennerud T, Melle I, Andreassen OA.
413 The quality of severe mental disorder diagnoses in a national health registry as compared to research
414 diagnoses based on structured interview. BMC Psychiatry. 2017 Mar; 17(1):93. https://doi.org/10.1186/
415 s12888-017-1256-8, doi: 10.1186/s12888-017-1256-8.

Nordahl CW, Iosif AM, Young GS, Hechtman A, Heath B, Lee JK, Libero L, Reinhardt VP, Winder-Patel B, Amaral DG, Rogers S, Solomon M, Ozonoff S. High Psychopathology Subgroup in Young Children With Autism: Associations With Biological Sex and Amygdala Volume. Journal of the American Academy of Child & Adolescent Psychiatry. 2020 Dec; 59(12):1353–1363.e2. https://linkinghub.elsevier.com/retrieve/pii/S0890856720300186, doi: 10.1016/j.jaac.2019.11.022.

Rios G, Tobar F. Compositionally-warped Gaussian processes. Neural Networks. 2019 Oct; 118:235–246. https://linkinghub.elsevier.com/retrieve/pii/S0893608019301856, doi: 10.1016/j.neunet.2019.06.012.

Rosen AFG, Roalf DR, Ruparel K, Blake J, Seelaus K, Villa LP, Ciric R, Cook PA, Davatzikos C, Elliott MA, Garcia de La Garza A, Gennatas ED, Quarmley M, Schmitt JE, Shinohara RT, Tisdall MD, Craddock RC, Gur RE, Gur RC, Satterthwaite TD. Quantitative assessment of structural image quality. NeuroImage. 2018 Apr; 169:407–418. doi: 10.1016/j.neuroimage.2017.12.059.

Rutherford S, Angstadt M, Sripada C, Chang SE. Leveraging big data for classification of children who stutter from fluent peers. bioRxiv. 2020 Oct; p. 2020.10.28.359711. https://www.biorxiv.org/content/10.1101/2020.10.28.359711v1, doi: 10.1101/2020.10.28.359711, publisher: Cold Spring Harbor Laboratory Section: New Results.

Snelson E, Rasmussen CE, Ghahramani Z. Warped Gaussian processes. In: *Proceedings of the 16th International Conference on Neural Information Processing Systems* NIPS'03, Whistler, British Columbia, Canada: MIT Press; 2003. p. 337–344.

Snoek L, van der Miesen MM, Beemsterboer T, van der Leij A, Eigenhuis A, Steven Scholte H. The Amsterdam Open MRI Collection, a set of multimodal MRI datasets for individual difference analyses. Scientific Data. 2021 Mar; 8:85. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7979787/, doi: 10.1038/s41597-021-00870-6.

Tamnes CK, Østby Y, Fjell AM, Westlye LT, Due-Tønnessen P, Walhovd KB. Brain Maturation in Adolescence and Young Adulthood: Regional Age-Related Changes in Cortical Thickness and White Matter Volume and Microstructure. Cerebral Cortex. 2010 Mar; 20(3):534–548. https://doi.org/10.1093/cercor/bhp118, doi: 10.1093/cercor/bhp118.

Tomlinson RC, Burt SA, Waller R, Jonides J, Miller AL, Gearhardt AN, Peltier SJ, Klump KL, Lumeng JC, Hyde LW. Neighborhood poverty predicts altered neural and behavioral response inhibition. NeuroImage. 2020 Apr; 209:116536. https://www.sciencedirect.com/science/article/pii/S1053811920300239, doi: 10.1016/j.neuroimage.2020.116536.

Tso IF, Angstadt M, Rutherford S, Peltier S, Diwadkar VA, Taylor SF. Dynamic causal modeling of eye gaze processing in schizophrenia. Schizophrenia Research. 2021 Mar; 229:112–121. https://www.sciencedirect.com/science/article/pii/S092099642030551X, doi: 10.1016/j.schres.2020.11.012.

Weigard AS, Brislin SJ, Cope LM, Hardee JE, Martz ME, Ly A, Zucker RA, Sripada C, Heitzeg MM. Evidence accumulation and associated error-related brain activity as computationally-informed prospective predictors of substance use in emerging adulthood. Psychopharmacology. 2021 Jun; https://doi.org/10.1007/s00213-021-05885-w, doi: 10.1007/s00213-021-05885-w.

Wolfers T, Arenas AL, Onnink AMH, Dammers J, Hoogman M, Zwiers MP, Buitelaar JK, Franke B, Marquand AF, Beckmann CF. Refinement by integration: aggregated effects of multimodal imaging markers on adult ADHD. Journal of Psychiatry & Neuroscience : JPN. 2017 Nov; 42(6):386–394. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5662460/, doi: 10.1503/jpn.160240.

Wolfers T, Beckmann CF, Hoogman M, Buitelaar JK, Franke B, Marquand AF. Individual differences v. the average patient: mapping the heterogeneity in ADHD using normative models. Psychological Medicine. 2020 Jan; 50(2):314–323. doi: 10.1017/S0033291719000084.

Wolfers T, Doan NT, Kaufmann T, Alnæs D, Moberget T, Agartz I, Buitelaar JK, Ueland T, Melle I, Franke B, Andreassen OA, Beckmann CF, Westlye LT, Marquand AF. Mapping the Heterogeneous Phenotype of Schizophrenia and Bipolar Disorder Using Normative Models. JAMA Psychiatry. 2018 Nov; 75(11):1146–1155. https://doi.org/10.1001/jamapsychiatry.2018.2467, doi: 10.1001/jamapsychiatry.2018.2467.

Wolfers T, Rokicki J, Alnaes D, Berthet P, Agartz I, Kia SM, Kaufmann T, Zabihi M, Moberget T, Melle I, Beckmann CF, Andreassen OA, Marquand AF, Westlye LT. Replicating extensive brain structural heterogeneity in individuals with schizophrenia and bipolar disorder. Human Brain Mapping. 2021 Jun; 42(8):2546–2555. doi: 10.1002/hbm.25386.

Yao B, Mueller SA, Grove TB, McLaughlin M, Thakkar K, Ellingrod V, McInnis MG, Taylor SF, Deldin PJ, Tso IF. Eye
Gaze Perception in Bipolar Disorder: Self-referential Bias but Intact Perceptual Sensitivity. Bipolar disorders.
2018 Feb; 20(1):60–69. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5807101/, doi: 10.1111/bdi.12564.

Zabihi M, Floris DL, Kia SM, Wolfers T, Tillmann J, Arenas AL, Moessnang C, Banaschewski T, Holt R, Baron-Cohen
S, Loth E, Charman T, Bourgeron T, Murphy D, Ecker C, Buitelaar JK, Beckmann CF, Marquand A, EU-AIMS LEAP
Group. Fractionating autism based on neuroanatomical normative modeling. Translational Psychiatry. 2020
Nov; 10(1):384. doi: 10.1038/s41398-020-01057-0.

Zabihi M, Oldehinkel M, Wolfers T, Frouin V, Goyard D, Loth E, Charman T, Tillmann J, Banaschewski T, Du-
mas G, Holt R, Baron-Cohen S, Durston S, Bölte S, Murphy D, Ecker C, Buitelaar JK, Beckmann CF, Mar-
quand AF. Dissecting the Heterogeneous Cortical Anatomy of Autism Spectrum Disorder Using Norma-
tive Models. Biological Psychiatry Cognitive Neuroscience and Neuroimaging. 2019 Jun; 4(6):567–578. doi:
10.1016/j.bpsc.2018.11.013.

Zucker RA, Ellis DA, Fitzgerald HE, Bingham CR, Sanford K. Other evidence for at least two alcoholisms II: Life
course variation in antisociality and heterogeneity of alcoholic outcome. Development and Psychopathol-
ogy. 1996; 8(4):831–848. http://www.cambridge.org/core/journals/development-and-psychopathology/article/
other-evidence-for-at-least-two-alcoholisms-ii-life-course-variation-in-antisociality-and-heterogeneity-of-alcoholic-outcome/
F477F2272522FCFB6F7A13011EB38BBB, doi: 10.1017/S0954579400007458, publisher: Cambridge Univer-
sity Press.

Explained Variance in Test Sets

Appendix 0 Figure 4. Comparison of the explained variance in cortical thickness across the different test sets. The patterns appear to be robust and consistent across the different test sets.

Appendix 0 Table 2. Full Sample Description, per site, in the train and test sets.

Site	Train N	%F/M	Age (m, s.d)	Test N	%F/M	Age (m, s.d)
ABCD_01	194	0.52, 0.48	9.95, 0.62	194	0.51, 0.49	9.85, 0.63
ABCD_02	271	0.47, 0.53	10.12, 0.6	271	0.46, 0.54	10.03, 0.64
ABCD_03	285	0.46, 0.54	9.86, 0.62	284	0.48, 0.52	9.9, 0.62
ABCD_04	315	0.47, 0.53	9.81, 0.64	316	0.5, 0.5	9.82, 0.65
ABCD_05	172	0.53, 0.47	9.9, 0.65	173	0.5, 0.5	9.89, 0.62
ABCD_06	282	0.5, 0.5	9.96, 0.59	282	0.51, 0.49	9.93, 0.59
ABCD_07	162	0.46, 0.54	9.87, 0.63	163	0.48, 0.52	9.87, 0.62
ABCD_08	168	0.42, 0.58	10, 0.62	168	0.54, 0.46	9.91, 0.62
ABCD_09	204	0.52, 0.48	9.95, 0.63	203	0.46, 0.54	9.98, 0.59
ABCD_10	287	0.47, 0.53	9.85, 0.63	288	0.5, 0.5	9.87, 0.62
ABCD_11	207	0.52, 0.48	9.84, 0.62	207	0.47, 0.53	9.79, 0.64
ABCD_12	81	0.41, 0.59	9.89, 0.56	80	0.55, 0.45	9.87, 0.62
ABCD_13	277	0.53, 0.47	9.81, 0.58	278	0.47, 0.53	9.82, 0.61
ABCD_14	292	0.45, 0.55	10.17, 0.56	291	0.47, 0.53	10.22, 0.57
ABCD_15	198	0.44, 0.56	9.87, 0.59	197	0.46, 0.54	9.94, 0.62
ABCD_16	460	0.45, 0.55	9.91, 0.65	461	0.45, 0.55	9.9, 0.65
ABCD_17	278	0.48, 0.52	9.79, 0.61	279	0.48, 0.52	9.84, 0.64
ABCD_18	171	0.48, 0.52	9.92, 0.61	170	0.46, 0.54	9.9, 0.65
ABCD_19	267	0.49, 0.51	10.04, 0.55	267	0.53, 0.47	10.08, 0.54
ABCD_20	320	0.51, 0.49	10.05, 0.5	320	0.49, 0.51	10.07, 0.48
ABCD_21	246	0.42, 0.58	9.92, 0.65	245	0.48, 0.52	9.91, 0.6
ABIDE_GU	27	0.41, 0.59	10.57, 1.68	27	0.59, 0.41	10.29, 1.75
ABIDE_KKI	93	0.34, 0.66	10.35, 1.19	94	0.35, 0.65	10.24, 1.2
ABIDE_NYU	68	0.15, 0.85	14.35, 6.01	67	0.27, 0.73	14.47, 6.62
ABIDE_USM	30	0.07, 0.93	23.12, 6.14	29	0.03, 0.97	20.98, 9.03
ADD200_KKI	31	0.45, 0.55	10.23, 1.28	30	0.43, 0.57	10.27, 1.29
ADD200_NYU	19	0.68, 0.32	10.04, 2.2	19	0.37, 0.63	10.38, 1.6
AOMIC_1000	464	0.51, 0.49	22.87, 1.71	464	0.53, 0.47	22.83, 1.71
AOMIC_PIPO2	104	0.59, 0.41	22.21, 1.91	105	0.56, 0.44	22.16, 1.69
ATV	39	0.21, 0.79	22.59, 1.96	38	0.24, 0.76	22.76, 2.02
CAMCAN	323	0.52, 0.48	55.11, 19.37	324	0.5, 0.5	53.28, 17.7
CIN	33	0.33, 0.67	48.64, 15.83	33	0.33, 0.67	47.15, 18.14
CMI-HBN_CBIC	99	0.35, 0.65	11.16, 3.73	100	0.38, 0.62	11.24, 3.74
CMI-HBN_RU	188	0.3, 0.7	10.37, 3.3	188	0.36, 0.64	10.69, 3.74
CMI-HBN_SI	53	0.32, 0.68	11.24, 3.83	53	0.4, 0.6	10.79, 3.64
CNP-35343.0	45	0.51, 0.49	31.91, 8.39	45	0.44, 0.56	31.98, 9.48
CNP-35426.0	10	0.6, 0.4	28.1, 7.17	10	0.5, 0.5	30.9, 8.75
COI	62	0.63, 0.37	51.92, 14.31	62	0.63, 0.37	51.81, 12.64
delta	24	0.42, 0.58	50.17, 9.27	25	0.32, 0.68	50.72, 8.44
ON_ds001734	54	0.52, 0.48	25.43, 3.75	54	0.59, 0.41	25.67, 3.45
ON_ds002236	43	0.42, 0.58	10.99, 1.75	43	0.47, 0.53	11.99, 2.21
ON_ds002330	33	0.58, 0.42	26.52, 4.15	33	0.55, 0.45	26.73, 4.52
ON_ds002345	104	0.63, 0.38	21.61, 4.12	103	0.64, 0.36	21.79, 5.26
ON_ds002731	29	0.66, 0.34	21.21, 1.57	30	0.3, 0.7	21.3, 1.34
ON_ds002837	43	0.56, 0.44	27.23, 10.63	43	0.42, 0.58	26.23, 9.55

Appendix 0 Table 2. Full Sample Description, per site, in the train and test sets.

Site	Train N	%F/M	Age (m, s.d)	Test N	%F/M	Age (m, s.d)
HCP_A_MGH	85	0.53, 0.47	61.41, 16.18	86	0.48, 0.52	58.13, 14.79
HCP_A_UCLA	62	0.58, 0.42	51.77, 11.36	62	0.56, 0.44	54.89, 14.03
HCP_A_UM	102	0.66, 0.34	63.71, 16.77	102	0.52, 0.48	59.48, 15.63
HCP_A_WU	89	0.64, 0.36	59.79, 14.34	89	0.62, 0.38	57.89, 12.54
HCP_D_MGH	108	0.53, 0.47	14.4, 3.94	108	0.48, 0.52	13.17, 3.72
HCP_D_UCLA	63	0.52, 0.48	14.3, 3.63	64	0.45, 0.55	13.98, 4.04
HCP_D_UM	78	0.53, 0.47	13.71, 3.69	78	0.56, 0.44	12.82, 3.55
HCP_D_WU	77	0.47, 0.53	14.21, 4.25	77	0.51, 0.49	13.73, 3.46
HCP_EP_BWH	4	0.25, 0.75	21.92, 2.07	4	0.25, 0.75	24.27, 4.28
HCP_EP_IU	12	0.42, 0.58	24.76, 4.21	13	0.46, 0.54	23.12, 3.27
HCP_EP_McL	7	0.29, 0.71	26.49, 4.29	6	0.33, 0.67	23.6, 2.33
HCP_EP_MGH	6	0.17, 0.83	26.89, 4.1	5	0.4, 0.6	28.48, 5.97
HCP_YA	557	0.55, 0.45	28.76, 3.73	556	0.53, 0.47	28.85, 3.66
HKH	15	0.47, 0.53	46.27, 7.69	14	0.71, 0.29	44.5, 11.42
HRC	25	0.76, 0.24	45.88, 12.74	24	0.71, 0.29	37.33, 8.7
HUH	33	0.55, 0.45	38.24, 13.66	34	0.59, 0.41	31.35, 11.45
IXI	279	0.56, 0.44	47.9, 16.57	279	0.56, 0.44	49.53, 16.39
KCL	20	0.65, 0.35	34.65, 14.13	21	0.57, 0.43	33.62, 13.44
KTT	64	0.28, 0.72	30.77, 9.7	64	0.36, 0.64	31.3, 8.88
KUT	79	0.39, 0.61	36.91, 13.35	80	0.44, 0.56	36.11, 13.89
NKI-RS	241	0.6, 0.4	42.63, 21.75	241	0.27, 0.73	42.63, 20.67
Oasis2	93	0.66, 0.34	75.92, 7.3	92	1.77, -0.77	77.86, 8.79
Oasis3	776	0.4, 0.6	69.41, 9.01	776	0.41, 0.59	70.16, 9
PNC	689	0.52, 0.48	14.16, 3.48	689	0.01, 0.99	14.28, 3.54
SWA	50	0.14, 0.86	27.34, 6.77	50	6.92, -5.92	29.62, 8.79
SWU_SLIM_ses1	274	0.59, 0.41	20.07, 1.3	274	0.57, 0.43	20.08, 1.25
TOP	146	0.45, 0.55	34.25, 9.68	146	0.47, 0.53	34.9, 9.43
UCDavis	69	0.49, 0.51	3.16, 0.57	67	0.42, 0.58	3.09, 0.55
ukb-11025.0	12493	0.52, 0.48	63, 7.51	12493	0.52, 0.48	62.98, 7.52
ukb-11027.0	4986	0.55, 0.45	64.35, 7.38	4986	0.54, 0.46	64.44, 7.47
UMich_CWS	14	0.57, 0.43	5.23, 1.15	15	0.6, 0.4	5.44, 1.16
UMich_IMPs	107	0.55, 0.45	12.91, 3.56	107	0.55, 0.45	12.83, 3.29
UMich_MLS	79	0.39, 0.61	19.94, 1.52	78	0.33, 0.67	20.14, 1.5
UMich_MTwins	300	0.47, 0.53	14.28, 1.97	300	0.45, 0.55	14.28, 2.13
UMich_SAD	57	0.4, 0.6	24.02, 4.75	57	0.35, 0.65	26.56, 9.02
UMich_SZG	23	0.43, 0.57	32.09, 11.09	22	0.55, 0.45	31.64, 8.63
UTO	101	0.56, 0.44	35.01, 17.31	101	0.49, 0.51	35.39, 16.53

Appendix 0 Table 3. Clinical Sample Description

Site	Patient N	%F/M	Age (m, s.d)
ABIDE_GU	47	0.17, 0.83	10.94, 1.53
ABIDE_KKI	77	0.25, 0.75	10.23, 1.5
ABIDE_NYU	127	0.13, 0.87	12.83, 6.86
ABIDE_USM	74	0.03, 0.97	21.61, 7.78

Continued on next page

Appendix 0 Table 3. Clinical Sample Description

Site	Patient N	%F/M	Age (m, s.d)
ADD200_KKI	22	0.45, 0.55	10.22, 1.56
ADD200_NYU	52	0.27, 0.73	10.05, 1.67
CNP-35343.0	63	0.48, 0.52	35.33, 9.75
CNP-35426.0	59	0.25, 0.75	34.63, 9.35
COI	69	0.58, 0.42	45.07, 12.62
delta	111	0.43, 0.57	45.23, 13.18
HCP_EP_BWH	23	0.35, 0.65	22.39, 4.26
HCP_EP_IU	59	0.37, 0.63	22.97, 3.84
HCP_EP_McL	31	0.48, 0.52	23.56, 3.46
HCP_EP_MGH	10	0.3, 0.7	20.32, 2.99
HKH	33	0.39, 0.61	44.82, 11.47
HRC	16	0.63, 0.38	40.5, 11.48
HUH	57	0.44, 0.56	43.33, 12.18
KCL	104	0.72, 0.28	31.44, 11.86
KTT	47	0.45, 0.55	37.89, 9.79
KUT	61	0.49, 0.51	41.7, 11.18
SWA	134	0.14, 0.86	33.62, 8.73
TOP	531	0.48, 0.52	32.42, 10.45
UMich_SZG	70	0.51, 0.49	32.77, 9.77
UTO	108	0.36, 0.64	36.1, 11.5

Appendix 0 Table 4. mQC Sample Description, per site, in the train and test sets.

Site	Train N	%F/M	Age (m, s.d)	Test N	%F/M	Age (m, s.d)
ABCD_01	180	0.53, 0.47	9.88, 0.62	179	0.47, 0.53	9.93, 0.63
ABCD_02	267	0.47, 0.53	10.12, 0.6	266	0.45, 0.55	10.02, 0.64
ABCD_03	282	0.49, 0.51	9.87, 0.61	282	0.45, 0.55	9.9, 0.62
ABCD_04	275	0.48, 0.52	9.87, 0.63	276	0.5, 0.5	9.79, 0.66
ABCD_05	169	0.51, 0.49	9.94, 0.65	170	0.52, 0.48	9.86, 0.6
ABCD_06	280	0.5, 0.5	9.97, 0.59	279	0.51, 0.49	9.93, 0.59
ABCD_07	160	0.43, 0.57	9.88, 0.6	160	0.51, 0.49	9.85, 0.65
ABCD_08	132	0.42, 0.58	9.98, 0.63	131	0.52, 0.48	10.01, 0.6
ABCD_09	197	0.5, 0.5	9.96, 0.6	197	0.5, 0.5	9.97, 0.63
ABCD_10	248	0.46, 0.54	9.9, 0.63	249	0.51, 0.49	9.83, 0.61
ABCD_11	203	0.44, 0.56	9.81, 0.63	203	0.55, 0.45	9.82, 0.63
ABCD_12	79	0.48, 0.52	9.97, 0.57	79	0.48, 0.52	9.79, 0.61
ABCD_13	240	0.5, 0.5	9.83, 0.62	240	0.51, 0.49	9.82, 0.59
ABCD_14	285	0.41, 0.59	10.2, 0.56	285	0.51, 0.49	10.21, 0.57
ABCD_15	188	0.49, 0.51	9.88, 0.59	188	0.41, 0.59	9.93, 0.61
ABCD_16	459	0.43, 0.57	9.9, 0.66	459	0.47, 0.51	9.91, 0.65
ABCD_17	264	0.51, 0.49	9.78, 0.62	265	0.47, 0.53	9.87, 0.64
ABCD_18	139	0.43, 0.57	9.99, 0.62	139	0.48, 0.52	9.92, 0.64
ABCD_19	254	0.5, 0.5	10.11, 0.55	254	0.52, 0.48	10.02, 0.54
ABCD_20	314	0.51, 0.49	10.08, 0.48	314	0.49, 0.51	10.04, 0.5
ABCD_21	239	0.47, 0.53	9.93, 0.62	239	0.44, 0.56	9.9, 0.62

Appendix 0 Table 4. mQC Sample Description, per site, in the train and test sets.

Site	Train N	%F/M	Age (m, s.d)	Test N	%F/M	Age (m, s.d)
CAMCAN	306	0.49, 0.51	52.83, 18.39	307	0.53, 0.53	53.56, 18.16
CMI-HBN_RU	170	0.35, 0.65	11.3, 3.4	170	0.38, 0.62	10.5, 3.7
CMI-HBN_SI	141	0.41, 0.59	12.1, 3.9	141	0.47, 0.53	11.4, 3.7
CNP-35343.0	44	0.39, 0.61	33.43, 9.5	43	0.58, 0.42	30.53, 8.26
CNP-35426.0	10	0.7, 0.3	28.7, 6.15	9	0.44, 0.56	29.33, 9.71
delta	24	0.42, 0.58	50.92, 8.48	23	0.3, 0.7	49.57, 9.44
ON_ds001734	54	0.57, 0.43	25.61, 3.8	54	0.54, 0.46	25.48, 3.4
ON_ds002236	37	0.46, 0.54	11.55, 2.02	37	0.51, 0.49	11.79, 2.17
ON_ds002330	33	0.52, 0.48	27.09, 4.24	33	0.61, 0.39	26.15, 4.39
ON_ds002345	104	0.66, 0.34	21.61, 4.36	103	0.6, 0.5	21.79, 5.06
ON_ds002731	29	0.48, 0.52	21.38, 1.7	30	0.47, 0.53	21.13, 1.17
ON_ds002837	43	0.53, 0.47	26.3, 9.05	43	0.44, 0.56	27.16, 11.06
HCP_A_MGH	85	0.56, 0.44	60.03, 15.91	85	0.44, 0.56	59.02, 14.68
HCP_A_UCLA	60	0.57, 0.43	55.35, 13.24	61	0.61, 0.39	50.91, 11.6
HCP_A_UM	101	0.59, 0.41	61.78, 16.25	100	0.58, 0.42	60.44, 15.59
HCP_A_WU	88	0.63, 0.38	58.93, 13.48	88	0.63, 0.38	58.25, 13.25
HCP_D_MGH	106	0.52, 0.48	13.94, 3.79	106	0.49, 0.51	13.74, 3.97
HCP_D_UCLA	63	0.46, 0.54	13.41, 3.76	62	0.5, 0.5	14.86, 3.85
HCP_D_UM	78	0.6, 0.4	13.61, 3.64	78	0.49, 0.51	12.93, 3.64
HCP_D_WU	76	0.43, 0.57	13.61, 3.74	75	0.53, 0.47	14.32, 3.94
HCP_EP_BWH	15	0.33, 0.67	22.59, 4	16	0.31, 0.69	22.55, 4.14
HCP_EP_IU	42	0.38, 0.62	23.12, 4.02	41	0.41, 0.59	23.43, 3.68
HCP_EP_McL	22	0.32, 0.68	23.87, 2.94	22	0.14, 0.36	24.2, 4.16
HCP_EP_MGH	10	0.3, 0.7	27.56, 5.58	11	1.09, 0.91	21.03, 2.99
HCP_YA	556	0.54, 0.46	28.96, 3.66	556	0.55, 0.45	28.65, 3.73
IXI	276	0.58, 0.42	48.7, 16.25	277	0.55, 0.45	48.76, 16.73
KCL	20	0.8, 0.2	32.8, 13.23	20	0.45, 0.55	34.75, 14.25
NKI-RS	229	0.66, 0.34	43.98, 20.32	228	0.63, 0.37	43.26, 20.56
Oasis3	199	0.36, 0.64	67.65, 10.93	198	0.3, 0.7	68.58, 10.07
PNC	631	0.52, 0.48	14.53, 3.38	631	0.52, 0.48	14.53, 3.42
TOP	187	0.52, 0.48	33.7, 9.5	188	0.36, 0.64	33.25, 9.72
ukb-11025.0	1867	0.53, 0.47	62.21, 7.45	1868	0.51, 0.49	62.43, 7.49
ukb-11027.0	1117	0.53, 0.47	63.24, 7.47	1117	0.56, 0.44	63.06, 7.39
UMich_CWS	14	0.64, 0.36	5.44, 1.18	15	0.53, 0.47	5.24, 1.13
UMich_IMPs	107	0.52, 0.48	12.96, 3.54	107	0.58, 0.42	12.78, 3.3
UMich_MTwins	300	0.48, 0.52	14.37, 2.04	300	0.44, 0.56	14.19, 2.06
UMich_SAD	57	0.35, 0.65	26.33, 7.65	57	0.4, 0.6	24.25, 6.82
UMich_SZG	22	0.45, 0.55	35.73, 9.82	23	0.52, 0.48	28.17, 8.54

Appendix 0 Table 5. Full Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
lh_G&S_frontomargin_thickness	0.27	-0.16	0.07	0.92
lh_G&S_occipital_inf_thickness	0.21	-0.11	0.10	0.30
lh_G&S_paracentral_thickness	0.21	-0.10	-0.05	0.25
			Continued on next page	

Appendix 0 Table 5. Full Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
lh_G&S_subcentral_thickness	0.26	-0.13	0.02	0.35
lh_G&S_transv_frontopol_thickness	0.25	-0.13	0.20	1.09
lh_G&S_cingul-Ant_thickness	0.28	-0.16	0.11	0.80
lh_G&S_cingul-Mid-Ant_thickness	0.23	-0.11	-0.44	0.90
lh_G&S_cingul-Mid-Post_thickness	0.33	-0.20	-0.13	0.64
lh_G_cingul-Post-dorsal_thickness	0.30	-0.15	-0.10	0.65
lh_G_cingul-Post-ventral_thickness	0.35	-0.21	0.02	0.18
lh_G_cuneus_thickness	0.39	-0.25	0.45	1.61
lh_G_front_inf-Opercular_thickness	0.32	-0.17	-0.14	0.65
lh_G_front_inf-Orbital_thickness	0.27	-0.14	-0.08	0.82
lh_G_front_inf-Triangul_thickness	0.35	-0.20	-0.05	0.68
lh_G_front_middle_thickness	0.39	-0.25	-0.26	0.82
lh_G_front_sup_thickness	0.39	-0.23	-0.18	0.62
lh_G_Ins_lg&S_cent_ins_thickness	0.13	-0.07	0.05	0.15
lh_G_insular_short_thickness	0.12	-0.06	-0.22	0.36
lh_G_occipital_middle_thickness	0.23	-0.13	-0.24	0.98
lh_G_occipital_sup_thickness	0.22	-0.12	0.17	0.41
lh_G_oc-temp_lat-fusifor_thickness	0.26	-0.12	-0.22	0.66
lh_G_oc-temp_med-Lingual_thickness	0.49	-0.35	0.19	0.57
lh_G_oc-temp_med-Parahip_thickness	0.14	-0.07	-0.21	0.24
lh_G_orbital_thickness	0.32	-0.19	-0.09	0.70
lh_G_pariet_inf-Angular_thickness	0.37	-0.23	-0.32	1.15
lh_G_pariet_inf-Supramar_thickness	0.34	-0.20	-0.21	0.65
lh_G_parietal_sup_thickness	0.35	-0.22	-0.14	0.55
lh_G_postcentral_thickness	0.22	-0.12	0.11	0.38
lh_G_precentral_thickness	0.23	-0.12	-0.50	1.04
lh_G_precuneus_thickness	0.34	-0.21	-0.09	0.43
lh_G_rectus_thickness	0.24	-0.11	-0.11	0.99
lh_G_subcallosal_thickness	0.23	-0.13	0.06	0.41
lh_G_temp_sup-G_T_transv_thickness	0.24	-0.13	0.16	0.24
lh_G_temp_sup-Lateral_thickness	0.23	-0.11	-0.08	0.36
lh_G_temp_sup-Plan_polar_thickness	0.14	-0.08	-0.27	0.41
lh_G_temp_sup-Plan_tempo_thickness	0.25	-0.12	-0.04	0.33
lh_G_temporal_inf_thickness	0.29	-0.14	-0.09	0.51
lh_G_temporal_middle_thickness	0.29	-0.17	-0.24	0.78
lh_Lat_Fis-ant-Horizont_thickness	0.16	-0.07	0.88	2.04
lh_Lat_Fis-ant-Vertical_thickness	0.21	-0.10	0.46	1.22
lh_Lat_Fis-post_thickness	0.30	-0.17	0.07	0.73
lh_Pole_occipital_thickness	0.15	-0.07	0.16	0.63
lh_Pole_temporal_thickness	0.18	-0.09	-0.31	0.77
lh_S_calcarine_thickness	0.51	-0.36	0.30	0.97
lh_S_central_thickness	0.30	-0.16	-0.03	0.63
lh_S_cingul-Marginalis_thickness	0.36	-0.22	-0.10	0.66
lh_S_circular_insula_ant_thickness	0.23	-0.10	0.02	0.60
lh_S_circular_insula_inf_thickness	0.28	-0.14	0.05	0.28
lh_S_circular_insula_sup_thickness	0.39	-0.23	-0.22	0.97

<div align="right">Continued on next page</div>

ROI	EV	MSLL	Skew	Kurtosis
lh_S_collat_transv_ant_thickness	0.11	-0.04	0.27	1.02
lh_S_collat_transv_post_thickness	0.19	-0.10	0.45	0.86
lh_S_front_inf_thickness	0.42	-0.25	-0.18	1.03
lh_S_front_middle_thickness	0.39	-0.23	0.08	1.40
lh_S_front_sup_thickness	0.43	-0.27	-0.29	1.11
lh_S_interm_prim-Jensen_thickness	0.16	-0.08	1.08	2.10
lh_S_intrapariet&P_trans_thickness	0.41	-0.25	-0.19	1.08
lh_S_oc_middle&Lunatus_thickness	0.27	-0.15	0.23	1.56
lh_S_oc_sup&transversal_thickness	0.32	-0.18	-0.06	0.67
lh_S_occipital_ant_thickness	0.19	-0.10	0.17	0.55
lh_S_oc-temp_lat_thickness	0.19	-0.10	0.22	0.77
lh_S_oc-temp_med&Lingual_thickness	0.32	-0.19	0.05	0.31
lh_S_orbital_lateral_thickness	0.24	-0.11	0.70	2.36
lh_S_orbital_med-olfact_thickness	0.20	-0.11	0.76	3.94
lh_S_orbital-H_Shaped_thickness	0.35	-0.19	0.11	0.75
lh_S_parieto_occipital_thickness	0.38	-0.22	-0.03	0.52
lh_S_pericallosal_thickness	0.10	-0.05	0.67	0.88
lh_S_postcentral_thickness	0.41	-0.25	-0.10	0.68
lh_S_precentral-inf-part_thickness	0.36	-0.21	-0.28	0.92
lh_S_precentral-sup-part_thickness	0.29	-0.17	-0.29	0.85
lh_S_suborbital_thickness	0.13	-0.06	0.64	1.53
lh_S_subparietal_thickness	0.29	-0.16	0.11	0.97
lh_S_temporal_inf_thickness	0.20	-0.11	0.14	0.79
lh_S_temporal_sup_thickness	0.41	-0.25	-0.22	0.68
lh_S_temporal_transverse_thickness	0.13	-0.07	0.49	0.20
lh_MeanThickness_thickness	0.46	-0.29	-0.30	0.62
rh_G&S_frontomargin_thickness	0.27	-0.13	0.36	1.60
rh_G&S_occipital_inf_thickness	0.12	-0.04	0.12	0.23
rh_G&S_paracentral_thickness	0.21	-0.10	0.05	0.40
rh_G&S_subcentral_thickness	0.25	-0.12	0.05	0.27
rh_G&S_transv_frontopol_thickness	0.29	-0.15	0.25	1.11
rh_G&S_cingul-Ant_thickness	0.36	-0.21	0.06	1.17
rh_G&S_cingul-Mid-Ant_thickness	0.25	-0.13	-0.36	0.92
rh_G&S_cingul-Mid-Post_thickness	0.31	-0.18	-0.16	0.63
rh_G_cingul-Post-dorsal_thickness	0.30	-0.17	-0.39	1.59
rh_G_cingul-Post-ventral_thickness	0.25	-0.14	0.00	0.31
rh_G_cuneus_thickness	0.50	-0.36	0.52	2.00
rh_G_front_inf-Opercular_thickness	0.33	-0.18	-0.14	0.55
rh_G_front_inf-Orbital_thickness	0.25	-0.13	0.00	0.66
rh_G_front_inf-Triangul_thickness	0.35	-0.19	-0.06	0.65
rh_G_front_middle_thickness	0.39	-0.25	-0.24	0.96
rh_G_front_sup_thickness	0.38	-0.24	-0.23	0.69
rh_G_Ins_lg&S_cent_ins_thickness	0.09	-0.04	0.02	0.02
rh_G_insular_short_thickness	0.13	-0.07	-0.01	0.11
rh_G_occipital_middle_thickness	0.25	-0.14	-0.21	1.03
rh_G_occipital_sup_thickness	0.26	-0.15	0.08	0.47

Continued on next page

Appendix 0 Table 5. Full Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
rh_G_oc-temp_lat-fusifor_thickness	0.20	-0.08	-0.11	0.47
rh_G_oc-temp_med-Lingual_thickness	0.54	-0.41	0.20	0.59
rh_G_oc-temp_med-Parahip_thickness	0.15	-0.07	-0.20	0.29
rh_G_orbital_thickness	0.29	-0.15	-0.04	0.47
rh_G_pariet_inf-Angular_thickness	0.33	-0.20	-0.30	0.91
rh_G_pariet_inf-Supramar_thickness	0.32	-0.19	-0.28	0.71
rh_G_parietal_sup_thickness	0.34	-0.20	-0.08	0.46
rh_G_postcentral_thickness	0.20	-0.11	0.09	0.49
rh_G_precentral_thickness	0.21	-0.10	-0.58	1.14
rh_G_precuneus_thickness	0.33	-0.20	0.00	0.42
rh_G_rectus_thickness	0.21	-0.09	0.07	1.04
rh_G_subcallosal_thickness	0.11	-0.05	0.39	0.48
rh_G_temp_sup-G_T_transv_thickness	0.21	-0.11	0.10	0.38
rh_G_temp_sup-Lateral_thickness	0.25	-0.12	-0.13	0.58
rh_G_temp_sup-Plan_polar_thickness	0.15	-0.07	-0.13	0.19
rh_G_temp_sup-Plan_tempo_thickness	0.27	-0.13	-0.02	0.40
rh_G_temporal_inf_thickness	0.25	-0.14	-0.17	0.50
rh_G_temporal_middle_thickness	0.31	-0.17	-0.26	0.95
rh_Lat_Fis-ant-Horizont_thickness	0.22	-0.10	0.46	1.08
rh_Lat_Fis-ant-Vertical_thickness	0.17	-0.08	0.68	1.71
rh_Lat_Fis-post_thickness	0.33	-0.19	0.04	1.10
rh_Pole_occipital_thickness	0.19	-0.10	0.20	0.40
rh_Pole_temporal_thickness	0.17	-0.08	-0.31	0.86
rh_S_calcarine_thickness	0.51	-0.36	0.32	1.14
rh_S_central_thickness	0.29	-0.16	0.04	0.70
rh_S_cingul-Marginalis_thickness	0.36	-0.21	-0.13	0.75
rh_S_circular_insula_ant_thickness	0.21	-0.09	0.12	0.42
rh_S_circular_insula_inf_thickness	0.26	-0.13	0.15	0.41
rh_S_circular_insula_sup_thickness	0.38	-0.23	-0.19	0.91
rh_S_collat_transv_ant_thickness	0.14	-0.05	0.57	1.52
rh_S_collat_transv_post_thickness	0.20	-0.10	0.28	0.73
rh_S_front_inf_thickness	0.41	-0.25	-0.26	1.47
rh_S_front_middle_thickness	0.43	-0.26	-0.13	1.32
rh_S_front_sup_thickness	0.42	-0.25	-0.28	1.26
rh_S_interm_prim-Jensen_thickness	0.25	-0.12	0.47	1.29
rh_S_intrapariet&P_trans_thickness	0.41	-0.25	-0.14	0.85
rh_S_oc_middle&Lunatus_thickness	0.23	-0.12	0.32	1.20
rh_S_oc_sup&transversal_thickness	0.31	-0.17	0.05	0.98
rh_S_occipital_ant_thickness	0.17	-0.09	0.24	0.39
rh_S_oc-temp_lat_thickness	0.17	-0.08	0.41	1.05
rh_S_oc-temp_med&Lingual_thickness	0.33	-0.20	0.08	0.59
rh_S_orbital_lateral_thickness	0.29	-0.16	0.48	2.19
rh_S_orbital_med-olfact_thickness	0.24	-0.13	0.49	2.50
rh_S_orbital-H_Shaped_thickness	0.39	-0.21	0.09	0.91
rh_S_parieto_occipital_thickness	0.34	-0.20	-0.05	0.53
rh_S_pericallosal_thickness	0.10	-0.05	0.82	1.07

Appendix 0 Table 5. Full Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
rh_S_postcentral_thickness	0.39	-0.23	-0.08	0.50
rh_S_precentral-inf-part_thickness	0.33	-0.19	-0.37	1.19
rh_S_precentral-sup-part_thickness	0.26	-0.15	-0.55	1.79
rh_S_suborbital_thickness	0.11	-0.06	0.31	0.21
rh_S_subparietal_thickness	0.27	-0.15	0.18	0.76
rh_S_temporal_inf_thickness	0.18	-0.10	0.20	1.13
rh_S_temporal_sup_thickness	0.41	-0.25	-0.18	0.70
rh_S_temporal_transverse_thickness	0.14	-0.07	0.25	0.18
rh_MeanThickness_thickness	0.45	-0.29	-0.24	0.59
Left-Lateral-Ventricle	0.40	-0.34	0.47	0.54
Left-Inf-Lat-Vent	0.30	-0.19	0.26	0.79
Left-Cerebellum-White-Matter	0.31	-0.20	0.32	2.15
Left-Cerebellum-Cortex	0.34	-0.20	-0.33	2.69
Left-Thalamus-Proper	0.58	-0.43	0.13	1.57
Left-Caudate	0.32	-0.19	0.06	1.24
Left-Putamen	0.46	-0.30	-0.22	2.02
Left-Pallidum	0.42	-0.29	-0.08	1.15
3rd-Ventricle	0.47	-0.37	0.68	1.15
4th-Ventricle	0.09	-0.05	0.35	0.48
Brain-Stem	0.37	-0.24	0.03	1.04
Left-Hippocampus	0.26	-0.15	-0.18	1.48
Left-Amygdala	0.28	-0.16	-0.16	1.40
CSF	0.28	-0.18	0.57	1.36
Left-Accumbens-area	0.60	-0.43	-0.20	1.23
Left-VentralDC	0.32	-0.19	0.10	0.55
Left-vessel	0.17	-0.09	0.12	0.72
Left-choroid-plexus	0.60	-0.42	-0.10	0.21
Right-Lateral-Ventricle	0.41	-0.35	0.46	0.56
Right-Inf-Lat-Vent	0.25	-0.14	0.14	0.71
Right-Cerebellum-White-Matter	0.29	-0.19	0.57	2.83
Right-Cerebellum-Cortex	0.32	-0.20	-0.31	2.77
Right-Thalamus-Proper	0.58	-0.42	0.08	0.94
Right-Caudate	0.33	-0.19	0.08	1.09
Right-Putamen	0.43	-0.28	-0.16	2.11
Right-Pallidum	0.35	-0.22	-0.03	0.80
Right-Hippocampus	0.25	-0.15	-0.14	1.02
Right-Amygdala	0.29	-0.17	-0.10	1.12
Right-Accumbens-area	0.42	-0.27	-0.04	1.02
Right-VentralDC	0.33	-0.19	0.12	0.64
Right-vessel	0.24	-0.11	0.08	1.40
Right-choroid-plexus	0.68	-0.45	-0.04	0.25
SubCortGrayVol	0.61	-0.42	-0.07	0.97
TotalGrayVol	0.60	-0.45	-0.16	1.40
SupraTentorialVol	0.35	-0.21	-0.03	0.52
SupraTentorialVolNotVent	0.37	-0.23	-0.05	0.54
avg_thickness	0.48	-0.31	-0.27	0.71

Appendix 0 Table 5. Full Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
EstimatedTotalIntraCranialVol	0.33	-0.20	-0.38	2.41

Appendix 0 Table 6. mQC Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
lh_G&S_frontomargin_thickness	0.26	-0.15	0.24	1.18
lh_G&S_occipital_inf_thickness	0.19	-0.11	0.06	0.41
lh_G&S_paracentral_thickness	0.29	-0.16	-0.02	0.48
lh_G&S_subcentral_thickness	0.27	-0.15	-0.01	0.48
lh_G&S_transv_frontopol_thickness	0.31	-0.16	0.27	1.43
lh_G&S_cingul-Ant_thickness	0.27	-0.15	0.09	0.80
lh_G&S_cingul-Mid-Ant_thickness	0.29	-0.17	-0.35	1.26
lh_G&S_cingul-Mid-Post_thickness	0.37	-0.23	-0.08	0.67
lh_G_cingul-Post-dorsal_thickness	0.33	-0.18	-0.04	0.50
lh_G_cingul-Post-ventral_thickness	0.37	-0.23	0.00	0.27
lh_G_cuneus_thickness	0.41	-0.26	0.24	0.58
lh_G_front_inf-Opercular_thickness	0.38	-0.24	-0.12	0.66
lh_G_front_inf-Orbital_thickness	0.34	-0.19	-0.06	0.90
lh_G_front_inf-Triangul_thickness	0.42	-0.26	-0.01	0.57
lh_G_front_middle_thickness	0.47	-0.32	-0.17	0.70
lh_G_front_sup_thickness	0.45	-0.30	-0.12	0.49
lh_G_Ins_lg&S_cent_ins_thickness	0.13	-0.07	0.04	0.15
lh_G_insular_short_thickness	0.15	-0.08	-0.24	0.33
lh_G_occipital_middle_thickness	0.27	-0.16	-0.32	1.45
lh_G_occipital_sup_thickness	0.27	-0.16	0.15	0.46
lh_G_oc-temp_lat-fusifor_thickness	0.26	-0.17	-0.15	0.62
lh_G_oc-temp_med-Lingual_thickness	0.50	-0.37	0.10	0.43
lh_G_oc-temp_med-Parahip_thickness	0.14	-0.08	-0.12	0.26
lh_G_orbital_thickness	0.36	-0.23	-0.01	0.49
lh_G_pariet_inf-Angular_thickness	0.44	-0.30	-0.36	1.37
lh_G_pariet_inf-Supramar_thickness	0.38	-0.24	-0.21	0.77
lh_G_parietal_sup_thickness	0.45	-0.30	-0.14	0.63
lh_G_postcentral_thickness	0.28	-0.16	0.15	0.64
lh_G_precentral_thickness	0.26	-0.14	-0.49	1.38
lh_G_precuneus_thickness	0.41	-0.26	-0.06	0.52
lh_G_rectus_thickness	0.24	-0.13	-0.18	0.86
lh_G_subcallosal_thickness	0.24	-0.15	0.03	0.43
lh_G_temp_sup-G_T_transv_thickness	0.30	-0.17	0.05	0.21
lh_G_temp_sup-Lateral_thickness	0.25	-0.12	-0.05	0.28
lh_G_temp_sup-Plan_polar_thickness	0.18	-0.12	-0.19	0.55
lh_G_temp_sup-Plan_tempo_thickness	0.27	-0.15	-0.04	0.40
lh_G_temporal_inf_thickness	0.25	-0.12	-0.14	0.53
lh_G_temporal_middle_thickness	0.34	-0.21	-0.20	0.94
lh_Lat_Fis-ant-Horizont_thickness	0.18	-0.08	0.73	1.73
lh_Lat_Fis-ant-Vertical_thickness	0.20	-0.10	0.45	1.21
				Continued on next page

Appendix 0 Table 6. mQC Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
lh_Lat_Fis-post_thickness	0.31	-0.18	0.01	0.65
lh_Pole_occipital_thickness	0.19	-0.10	0.10	0.77
lh_Pole_temporal_thickness	0.13	-0.06	-0.21	0.63
lh_S_calcarine_thickness	0.53	-0.39	0.19	0.35
lh_S_central_thickness	0.31	-0.18	-0.07	0.76
lh_S_cingul-Marginalis_thickness	0.42	-0.27	-0.15	0.77
lh_S_circular_insula_ant_thickness	0.22	-0.12	-0.14	0.73
lh_S_circular_insula_inf_thickness	0.30	-0.17	0.09	0.41
lh_S_circular_insula_sup_thickness	0.37	-0.23	-0.29	0.98
lh_S_collat_transv_ant_thickness	0.10	-0.05	0.46	1.59
lh_S_collat_transv_post_thickness	0.20	-0.11	0.40	1.20
lh_S_front_inf_thickness	0.40	-0.25	-0.35	1.29
lh_S_front_middle_thickness	0.36	-0.22	-0.04	1.11
lh_S_front_sup_thickness	0.40	-0.26	-0.36	1.40
lh_S_interm_prim-Jensen_thickness	0.13	-0.06	1.19	2.64
lh_S_intrapariet&P_trans_thickness	0.43	-0.27	-0.33	1.20
lh_S_oc_middle&Lunatus_thickness	0.24	-0.14	0.05	1.21
lh_S_oc_sup&transversal_thickness	0.32	-0.19	-0.17	0.71
lh_S_occipital_ant_thickness	0.18	-0.10	0.15	0.52
lh_S_oc-temp_lat_thickness	0.16	-0.08	0.30	0.91
lh_S_oc-temp_med&Lingual_thickness	0.33	-0.20	0.11	0.47
lh_S_orbital_lateral_thickness	0.21	-0.11	0.59	1.87
lh_S_orbital_med-olfact_thickness	0.21	-0.08	0.97	5.28
lh_S_orbital-H_Shaped_thickness	0.32	-0.23	0.14	0.62
lh_S_parieto_occipital_thickness	0.40	-0.25	-0.07	0.70
lh_S_pericallosal_thickness	0.11	-0.08	0.70	1.01
lh_S_postcentral_thickness	0.44	-0.28	-0.18	0.88
lh_S_precentral-inf-part_thickness	0.34	-0.21	-0.27	1.15
lh_S_precentral-sup-part_thickness	0.28	-0.17	-0.29	1.16
lh_S_suborbital_thickness	0.13	-0.05	0.57	1.45
lh_S_subparietal_thickness	0.30	-0.18	0.03	0.85
lh_S_temporal_inf_thickness	0.20	-0.11	0.32	1.32
lh_S_temporal_sup_thickness	0.42	-0.27	-0.19	0.79
lh_S_temporal_transverse_thickness	0.14	-0.07	0.57	0.49
lh_MeanThickness_thickness	0.50	-0.34	-0.35	0.88
rh_G&S_frontomargin_thickness	0.25	-0.14	0.22	1.07
rh_G&S_occipital_inf_thickness	0.13	-0.05	0.14	0.31
rh_G&S_paracentral_thickness	0.29	-0.16	0.03	0.57
rh_G&S_subcentral_thickness	0.26	-0.14	0.09	0.32
rh_G&S_transv_frontopol_thickness	0.33	-0.18	0.22	0.86
rh_G&S_cingul-Ant_thickness	0.33	-0.19	-0.02	1.00
rh_G&S_cingul-Mid-Ant_thickness	0.31	-0.18	-0.25	0.82
rh_G&S_cingul-Mid-Post_thickness	0.33	-0.20	-0.15	0.66
rh_G_cingul-Post-dorsal_thickness	0.31	-0.18	-0.39	1.41
rh_G_cingul-Post-ventral_thickness	0.24	-0.14	-0.04	0.43
rh_G_cuneus_thickness	0.51	-0.38	0.45	1.66

Appendix 0 Table 6. mQC Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
rh_G_front_inf-Opercular_thickness	0.39	-0.25	-0.06	0.49
rh_G_front_inf-Orbital_thickness	0.31	-0.18	0.01	0.64
rh_G_front_inf-Triangul_thickness	0.40	-0.25	-0.03	0.56
rh_G_front_middle_thickness	0.47	-0.32	-0.30	0.87
rh_G_front_sup_thickness	0.43	-0.28	-0.26	0.67
rh_G_Ins_lg&S_cent_ins_thickness	0.11	-0.05	0.05	0.04
rh_G_insular_short_thickness	0.14	-0.09	-0.02	0.10
rh_G_occipital_middle_thickness	0.32	-0.19	-0.24	1.28
rh_G_occipital_sup_thickness	0.33	-0.21	-0.02	0.47
rh_G_oc-temp_lat-fusifor_thickness	0.22	-0.12	-0.01	0.49
rh_G_oc-temp_med-Lingual_thickness	0.53	-0.42	0.25	0.90
rh_G_oc-temp_med-Parahip_thickness	0.15	-0.08	-0.08	0.22
rh_G_orbital_thickness	0.32	-0.17	0.04	0.50
rh_G_pariet_inf-Angular_thickness	0.40	-0.23	-0.29	1.00
rh_G_pariet_inf-Supramar_thickness	0.36	-0.22	-0.19	0.45
rh_G_parietal_sup_thickness	0.43	-0.28	-0.07	0.41
rh_G_postcentral_thickness	0.26	-0.14	0.16	0.61
rh_G_precentral_thickness	0.24	-0.14	-0.67	1.73
rh_G_precuneus_thickness	0.40	-0.25	-0.01	0.45
rh_G_rectus_thickness	0.20	-0.09	0.00	0.96
rh_G_subcallosal_thickness	0.11	-0.06	0.31	0.17
rh_G_temp_sup-G_T_transv_thickness	0.26	-0.14	0.07	0.41
rh_G_temp_sup-Lateral_thickness	0.26	-0.12	-0.08	0.42
rh_G_temp_sup-Plan_polar_thickness	0.17	-0.10	-0.04	0.27
rh_G_temp_sup-Plan_tempo_thickness	0.27	-0.15	0.03	0.41
rh_G_temporal_inf_thickness	0.23	-0.13	-0.05	0.48
rh_G_temporal_middle_thickness	0.36	-0.22	-0.20	0.70
rh_Lat_Fis-ant-Horizont_thickness	0.23	-0.12	0.34	1.04
rh_Lat_Fis-ant-Vertical_thickness	0.14	-0.07	0.65	1.48
rh_Lat_Fis-post_thickness	0.33	-0.20	-0.02	1.14
rh_Pole_occipital_thickness	0.24	-0.13	0.12	0.47
rh_Pole_temporal_thickness	0.14	-0.05	-0.15	0.54
rh_S_calcarine_thickness	0.52	-0.38	0.33	1.28
rh_S_central_thickness	0.33	-0.19	0.04	0.94
rh_S_cingul-Marginalis_thickness	0.40	-0.24	-0.20	0.75
rh_S_circular_insula_ant_thickness	0.19	-0.09	0.01	0.49
rh_S_circular_insula_inf_thickness	0.27	-0.15	0.20	0.64
rh_S_circular_insula_sup_thickness	0.35	-0.21	-0.24	0.92
rh_S_collat_transv_ant_thickness	0.12	-0.05	0.56	1.43
rh_S_collat_transv_post_thickness	0.21	-0.11	0.31	0.87
rh_S_front_inf_thickness	0.39	-0.24	-0.43	1.77
rh_S_front_middle_thickness	0.39	-0.24	-0.31	1.31
rh_S_front_sup_thickness	0.37	-0.24	-0.32	1.43
rh_S_interm_prim-Jensen_thickness	0.24	-0.13	0.57	1.72
rh_S_intrapariet&P_trans_thickness	0.43	-0.27	-0.21	0.92
rh_S_oc_middle&Lunatus_thickness	0.21	-0.12	0.14	0.74

Continued on next page

ROI	EV	MSLL	Skew	Kurtosis
rh_S_oc_sup&transversal_thickness	0.31	-0.18	-0.02	0.92
rh_S_occipital_ant_thickness	0.18	-0.09	0.28	0.60
rh_S_oc-temp_lat_thickness	0.13	-0.07	0.47	1.53
rh_S_oc-temp_med&Lingual_thickness	0.36	-0.25	0.16	0.44
rh_S_orbital_lateral_thickness	0.25	-0.14	0.13	1.71
rh_S_orbital_med-olfact_thickness	0.20	-0.08	0.70	3.90
rh_S_orbital-H_Shaped_thickness	0.33	-0.21	0.15	0.83
rh_S_parieto_occipital_thickness	0.39	-0.24	-0.12	0.61
rh_S_pericallosal_thickness	0.11	-0.09	0.87	1.37
rh_S_postcentral_thickness	0.42	-0.26	-0.08	0.76
rh_S_precentral-inf-part_thickness	0.30	-0.18	-0.43	1.41
rh_S_precentral-sup-part_thickness	0.24	-0.14	-0.57	1.78
rh_S_suborbital_thickness	0.09	-0.05	0.66	0.75
rh_S_subparietal_thickness	0.28	-0.17	0.07	0.71
rh_S_temporal_inf_thickness	0.17	-0.09	0.18	0.86
rh_S_temporal_sup_thickness	0.42	-0.26	-0.17	0.65
rh_S_temporal_transverse_thickness	0.15	-0.08	0.20	0.28
rh_MeanThickness_thickness	0.49	-0.34	-0.29	0.73
Left-Lateral-Ventricle	0.42	-0.43	0.60	0.71
Left-Inf-Lat-Vent	0.26	-0.17	0.30	0.86
Left-Cerebellum-White-Matter	0.25	-0.16	0.53	2.21
Left-Cerebellum-Cortex	0.31	-0.19	-0.40	4.09
Left-Thalamus-Proper	0.61	-0.54	0.23	1.15
Left-Caudate	0.32	-0.23	0.12	0.59
Left-Putamen	0.49	-0.38	-0.18	0.96
Left-Pallidum	0.42	-0.30	-0.04	0.73
3rd-Ventricle	0.50	-0.50	0.73	1.34
4th-Ventricle	0.08	-0.04	0.32	0.34
Brain-Stem	0.38	-0.26	0.08	0.36
Left-Hippocampus	0.26	-0.14	-0.08	0.80
Left-Amygdala	0.29	-0.16	-0.07	0.82
CSF	0.28	-0.20	0.69	1.97
Left-Accumbens-area	0.60	-0.49	-0.04	0.39
Left-VentralDC	0.29	-0.17	0.16	0.33
Left-vessel	0.14	-0.08	0.10	0.73
Left-choroid-plexus	0.57	-0.42	-0.02	0.43
Right-Lateral-Ventricle	0.43	-0.44	0.57	0.65
Right-Inf-Lat-Vent	0.20	-0.12	0.16	0.76
Right-Cerebellum-White-Matter	0.28	-0.19	0.69	2.88
Right-Cerebellum-Cortex	0.29	-0.17	-0.38	4.57
Right-Thalamus-Proper	0.58	-0.48	0.18	0.77
Right-Caudate	0.33	-0.23	0.17	0.57
Right-Putamen	0.45	-0.34	-0.08	0.81
Right-Pallidum	0.29	-0.18	0.05	0.60
Right-Hippocampus	0.25	-0.15	-0.06	0.61
Right-Amygdala	0.29	-0.17	-0.01	0.38

Appendix 0 Table 6. mQC Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
Right-Accumbens-area	0.36	-0.26	0.07	0.50
Right-VentralDC	0.30	-0.17	0.17	0.50
Right-vessel	0.23	-0.11	0.14	1.27
Right-choroid-plexus	0.61	-0.40	0.00	0.43
SubCortGrayVol	0.47	-0.34	0.00	0.23
TotalGrayVol	0.62	-0.49	-0.10	0.22
SupraTentorialVol	0.35	-0.20	0.00	0.18
SupraTentorialVolNotVent	0.37	-0.23	-0.01	0.17
avg_thickness	0.51	-0.36	-0.32	0.82
EstimatedTotalIntraCranialVol	0.36	-0.22	-0.28	1.54

Appendix 0 Table 7. Patient Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
lh_G&S_frontomargin	0.45	-0.94	0.69	3.04
lh_G&S_occipital_inf	0.38	-0.78	0.15	0.67
lh_G&S_paracentral	0.39	-0.37	0.00	0.32
lh_G&S_subcentral	0.46	-0.95	-0.05	0.76
lh_G&S_transv_frontopol	0.39	-0.52	0.75	2.31
lh_G&S_cingul-Ant	0.49	-1.05	0.29	1.35
lh_G&S_cingul-Mid-Ant	0.43	-0.35	0.00	1.07
lh_G&S_cingul-Mid-Post	0.58	-0.99	0.11	0.76
lh_G_cingul-Post-dorsal	0.53	-0.91	-0.11	1.35
lh_G_cingul-Post-ventral	0.48	-0.34	0.13	0.72
lh_G_cuneus	0.52	-0.41	0.66	1.85
lh_G_front_inf-Opercular	0.53	-1.10	-0.16	0.96
lh_G_front_inf-Orbital	0.41	-0.55	-0.10	1.09
lh_G_front_inf-Triangul	0.56	-0.96	-0.08	1.01
lh_G_front_middle	0.52	-1.12	-0.18	1.03
lh_G_front_sup	0.54	-1.02	-0.22	1.25
lh_G_Ins_lg&S_cent_ins	0.26	-0.14	-0.17	0.83
lh_G_insular_short	0.28	-0.18	-0.29	1.84
lh_G_occipital_middle	0.47	-0.79	-0.31	1.49
lh_G_occipital_sup	0.36	-0.42	0.32	0.78
lh_G_oc-temp_lat-fusifor	0.40	-0.97	-0.26	1.79
lh_G_oc-temp_med-Lingual	0.56	-0.38	0.56	2.02
lh_G_oc-temp_med-Parahip	0.24	-0.17	-0.41	0.89
lh_G_orbital	0.58	-0.79	-0.11	1.29
lh_G_pariet_inf-Angular	0.57	-1.06	-0.38	2.15
lh_G_pariet_inf-Supramar	0.55	-1.23	-0.28	1.27
lh_G_parietal_sup	0.48	-1.06	-0.07	1.17
lh_G_postcentral	0.37	-0.66	0.12	1.03
lh_G_precentral	0.43	-0.61	-0.59	1.59
lh_G_precuneus	0.58	-1.03	-0.06	0.69
lh_G_rectus	0.35	-0.65	0.89	5.08

Continued on next page

Appendix 0 Table 7. Patient Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
lh_G_subcallosal	0.25	-0.14	0.00	0.78
lh_G_temp_sup-G_T_transv	0.39	-0.44	0.11	0.13
lh_G_temp_sup-Lateral	0.41	-0.60	-0.27	1.14
lh_G_temp_sup-Plan_polar	0.33	-0.22	-0.49	1.06
lh_G_temp_sup-Plan_tempo	0.45	-0.86	0.03	0.46
lh_G_temporal_inf	0.31	-0.96	-0.33	1.41
lh_G_temporal_middle	0.45	-0.71	-0.42	1.50
lh_Lat_Fis-ant-Horizont	0.39	-0.64	0.72	1.41
lh_Lat_Fis-ant-Vertical	0.39	-0.90	0.60	1.42
lh_Lat_Fis-post	0.60	-1.04	0.15	0.38
lh_Pole_occipital	0.34	-0.26	0.74	3.35
lh_Pole_temporal	0.22	-0.50	-0.80	2.17
lh_S_calcarine	0.65	-0.69	0.79	2.83
lh_S_central	0.59	-0.96	0.27	1.98
lh_S_cingul-Marginalis	0.58	-1.21	0.20	0.92
lh_S_circular_insula_ant	0.49	-0.85	0.13	0.64
lh_S_circular_insula_inf	0.53	-0.57	0.04	0.66
lh_S_circular_insula_sup	0.65	-1.45	-0.02	0.49
lh_S_collat_transv_ant	0.32	-0.48	0.47	1.42
lh_S_collat_transv_post	0.37	-0.56	0.81	3.55
lh_S_front_inf	0.65	-1.69	0.08	0.84
lh_S_front_middle	0.53	-1.35	0.54	2.68
lh_S_front_sup	0.52	-1.55	-0.02	1.28
lh_S_interm_prim-Jensen	0.25	-0.52	1.16	2.69
lh_S_intrapariet&P_trans	0.62	-1.66	0.08	1.31
lh_S_oc_middle&Lunatus	0.41	-0.89	0.84	3.98
lh_S_oc_sup&transversal	0.50	-1.12	0.25	0.76
lh_S_occipital_ant	0.40	-0.75	0.79	5.01
lh_S_oc-temp_lat	0.37	-0.80	0.64	3.14
lh_S_oc-temp_med&Lingual	0.56	-0.75	0.06	0.84
lh_S_orbital_lateral	0.39	-0.80	1.03	3.80
lh_S_orbital_med-olfact	0.39	-0.41	1.73	8.42
lh_S_orbital-H_Shaped	0.53	-1.14	0.16	1.27
lh_S_parieto_occipital	0.64	-1.08	0.19	0.50
lh_S_pericallosal	0.24	-0.13	0.92	1.75
lh_S_postcentral	0.63	-1.53	0.09	0.51
lh_S_precentral-inf-part	0.58	-1.57	-0.01	0.83
lh_S_precentral-sup-part	0.48	-1.14	-0.21	0.90
lh_S_suborbital	0.32	-0.38	1.31	6.09
lh_S_subparietal	0.51	-1.12	0.56	2.81
lh_S_temporal_inf	0.45	-0.66	0.54	5.47
lh_S_temporal_sup	0.67	-1.45	-0.23	0.73
lh_S_temporal_transverse	0.30	-0.32	0.31	0.61
lh_MeanThickness	0.71	-1.60	-0.31	1.10
rh_G&S_frontomargin	0.42	-0.80	1.16	5.18
rh_G&S_occipital_inf	0.30	-0.53	0.21	0.49

ROI	EV	MSLL	Skew	Kurtosis
rh_G&S_paracentral	0.38	-0.35	0.18	0.76
rh_G&S_subcentral	0.45	-0.85	0.13	0.27
rh_G&S_transv_frontopol	0.42	-0.62	0.69	2.37
rh_G&S_cingul-Ant	0.62	-1.29	0.40	2.06
rh_G&S_cingul-Mid-Ant	0.53	-0.70	0.12	2.43
rh_G&S_cingul-Mid-Post	0.59	-1.07	0.18	0.79
rh_G_cingul-Post-dorsal	0.51	-0.99	-0.37	2.26
rh_G_cingul-Post-ventral	0.26	-0.16	0.11	0.49
rh_G_cuneus	0.46	-0.31	0.90	3.93
rh_G_front_inf-Opercular	0.51	-1.05	0.02	0.50
rh_G_front_inf-Orbital	0.39	-0.50	0.16	0.86
rh_G_front_inf-Triangul	0.51	-0.82	0.18	1.45
rh_G_front_middle	0.53	-1.00	-0.38	1.33
rh_G_front_sup	0.56	-0.99	-0.18	1.39
rh_G_Ins_lg&S_cent_ins	0.21	-0.11	-0.07	0.45
rh_G_insular_short	0.25	-0.14	-0.06	0.08
rh_G_occipital_middle	0.48	-0.91	-0.23	1.89
rh_G_occipital_sup	0.41	-0.46	0.05	0.95
rh_G_oc-temp_lat-fusifor	0.38	-0.71	-0.15	1.09
rh_G_oc-temp_med-Lingual	0.50	-0.35	0.66	2.67
rh_G_oc-temp_med-Parahip	0.27	-0.20	-0.42	0.77
rh_G_orbital	0.54	-0.68	0.05	0.70
rh_G_pariet_inf-Angular	0.53	-1.01	-0.38	1.72
rh_G_pariet_inf-Supramar	0.53	-1.16	-0.29	0.92
rh_G_parietal_sup	0.43	-0.98	-0.11	0.91
rh_G_postcentral	0.36	-0.53	0.19	0.40
rh_G_precentral	0.43	-0.52	-0.69	1.66
rh_G_precuneus	0.55	-0.98	-0.02	0.44
rh_G_rectus	0.35	-0.56	0.60	2.12
rh_G_subcallosal	0.12	-0.06	0.02	0.44
rh_G_temp_sup-G_T_transv	0.41	-0.47	0.12	0.17
rh_G_temp_sup-Lateral	0.43	-0.66	-0.29	1.16
rh_G_temp_sup-Plan_polar	0.37	-0.27	-0.19	0.81
rh_G_temp_sup-Plan_tempo	0.50	-1.00	0.09	0.25
rh_G_temporal_inf	0.36	-0.90	-0.37	1.54
rh_G_temporal_middle	0.43	-0.81	-0.38	1.23
rh_Lat_Fis-ant-Horizont	0.43	-0.83	0.62	1.68
rh_Lat_Fis-ant-Vertical	0.30	-0.64	0.84	2.23
rh_Lat_Fis-post	0.61	-1.07	0.04	0.35
rh_Pole_occipital	0.41	-0.26	0.51	1.54
rh_Pole_temporal	0.24	-0.46	-0.79	2.02
rh_S_calcarine	0.65	-0.72	0.66	1.99
rh_S_central	0.63	-0.92	0.28	1.03
rh_S_cingul-Marginalis	0.61	-1.33	0.08	0.77
rh_S_circular_insula_ant	0.42	-0.76	0.34	1.12
rh_S_circular_insula_inf	0.53	-0.58	0.29	0.60

ROI	EV	MSLL	Skew	Kurtosis
rh_S_circular_insula_sup	0.63	-1.44	0.11	1.12
rh_S_collat_transv_ant	0.33	-0.62	0.48	1.46
rh_S_collat_transv_post	0.40	-0.59	0.51	1.40
rh_S_front_inf	0.63	-1.60	0.39	2.23
rh_S_front_middle	0.58	-1.43	0.46	1.66
rh_S_front_sup	0.54	-1.49	-0.11	0.59
rh_S_interm_prim-Jensen	0.42	-0.99	0.44	1.51
rh_S_intrapariet&P_trans	0.60	-1.57	0.10	0.83
rh_S_oc_middle&Lunatus	0.37	-0.81	0.45	1.80
rh_S_oc_sup&transversal	0.49	-1.03	0.12	0.96
rh_S_occipital_ant	0.39	-0.75	0.12	1.65
rh_S_oc-temp_lat	0.36	-0.76	0.34	2.53
rh_S_oc-temp_med&Lingual	0.52	-0.57	0.05	0.72
rh_S_orbital_lateral	0.45	-0.98	1.28	7.26
rh_S_orbital_med-olfact	0.49	-0.63	0.79	3.40
rh_S_orbital-H_Shaped	0.55	-1.24	0.36	1.10
rh_S_parieto_occipital	0.62	-1.10	0.16	0.45
rh_S_pericallosal	0.27	-0.16	0.81	1.41
rh_S_postcentral	0.61	-1.38	0.14	0.37
rh_S_precentral-inf-part	0.57	-1.47	-0.06	0.57
rh_S_precentral-sup-part	0.45	-0.96	-0.34	1.31
rh_S_suborbital	0.25	-0.25	0.53	0.60
rh_S_subparietal	0.52	-1.04	0.24	0.54
rh_S_temporal_inf	0.42	-0.71	0.32	1.94
rh_S_temporal_sup	0.63	-1.57	-0.19	0.83
rh_S_temporal_transverse	0.32	-0.35	0.36	1.16
MeanThickness	0.71	-1.54	-0.18	0.84
Left-Lateral-Ventricle	0.37	-0.26	0.55	0.51
Left-Inf-Lat-Vent	0.27	-0.31	0.87	1.78
Left-Cerebellum-White-Matter	0.16	-0.22	-0.65	4.87
Left-Cerebellum-Cortex	0.27	-0.18	-1.75	14.02
Left-Thalamus-Proper	0.40	-0.22	-0.62	8.90
Left-Caudate	0.13	-0.07	-0.16	3.70
Left-Putamen	0.33	-0.18	-0.85	5.61
Left-Pallidum	0.47	-0.44	-0.57	5.08
3rd-Ventricle	0.38	-0.22	0.79	1.34
4th-Ventricle	0.01	0.00	0.30	1.42
Brain-Stem	0.13	-0.06	-0.64	6.30
Left-Hippocampus	0.35	-0.31	-1.54	8.88
Left-Amygdala	0.35	-0.28	-1.20	6.45
CSF	0.26	-0.16	0.50	2.88
Left-Accumbens-area	0.51	-0.53	-0.75	4.13
Left-VentralDC	0.40	-0.30	-0.64	7.99
Left-vessel	0.35	-0.28	0.00	1.58
Left-choroid-plexus	0.73	-0.55	0.20	1.13
Right-Lateral-Ventricle	0.39	-0.26	0.49	0.69

Appendix 0 Table 7. Patient Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
Right-Inf-Lat-Vent	0.24	-0.26	0.88	2.16
Right-Cerebellum-White-Matter	0.12	-0.16	-0.67	6.55
Right-Cerebellum-Cortex	0.25	-0.22	-1.80	16.41
Right-Thalamus-Proper	0.49	-0.32	-0.39	4.50
Right-Caudate	0.18	-0.10	0.09	2.29
Right-Putamen	0.36	-0.21	-0.91	4.40
Right-Pallidum	0.39	-0.38	-0.45	3.92
Right-Hippocampus	0.37	-0.31	-1.12	4.22
Right-Amygdala	0.36	-0.38	-0.94	3.38
Right-Accumbens-area	0.38	-0.23	-0.48	1.92
Right-VentralDC	0.40	-0.32	-0.31	5.74
Right-vessel	0.38	-0.25	-0.31	2.34
Right-choroid-plexus	0.80	-0.85	0.17	0.89
SubCortGrayVol	0.43	-0.23	-0.99	9.58
TotalGrayVol	0.54	-0.52	-0.37	2.99
SupraTentorialVolNotVent	0.33	-0.27	-0.06	1.69
avg_thickness	0.72	-1.66	-0.22	0.82
EstimatedTotalIntraCranialVol	0.26	-0.16	0.23	1.33

Appendix 0 Table 8. Transfer Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
lh_G&S_frontomargin_thickness	0.20	-0.62	-0.26	0.66
lh_G&S_occipital_inf_thickness	0.02	-0.43	0.66	2.95
lh_G&S_paracentral_thickness	0.26	-0.37	0.31	1.72
lh_G&S_subcentral_thickness	0.20	-0.44	0.22	0.61
lh_G&S_transv_frontopol_thickness	0.13	-0.15	0.47	0.74
lh_G&S_cingul-Ant_thickness	0.20	-0.42	0.52	1.24
lh_G&S_cingul-Mid-Ant_thickness	0.25	-0.28	0.29	0.04
lh_G&S_cingul-Mid-Post_thickness	0.24	-0.43	-0.16	0.55
lh_G_cingul-Post-dorsal_thickness	0.23	-0.34	-0.36	1.32
lh_G_cingul-Post-ventral_thickness	0.08	-0.08	0.38	0.79
lh_G_cuneus_thickness	0.61	-0.54	0.79	2.43
lh_G_front_inf-Opercular_thickness	0.33	-0.50	0.41	0.91
lh_G_front_inf-Orbital_thickness	0.20	-0.19	0.36	1.36
lh_G_front_inf-Triangul_thickness	0.19	-0.39	0.25	0.12
lh_G_front_middle_thickness	0.31	-0.58	0.38	1.08
lh_G_front_sup_thickness	0.50	-0.59	0.30	0.29
lh_G_Ins_lg&S_cent_ins_thickness	0.23	-0.10	-0.08	0.24
lh_G_insular_short_thickness	0.33	-0.16	-0.01	0.60
lh_G_occipital_middle_thickness	0.22	-0.51	0.10	-0.05
lh_G_occipital_sup_thickness	0.24	-0.27	0.58	3.04
lh_G_oc-temp_lat-fusifor_thickness	0.25	-0.73	0.02	0.48
lh_G_oc-temp_med-Lingual_thickness	0.52	-0.46	0.32	0.15
lh_G_oc-temp_med-Parahip_thickness	0.23	-0.12	0.21	0.32

ROI	EV	MSLL	Skew	Kurtosis
lh_G_orbital_thickness	0.42	-0.44	-0.15	0.29
lh_G_pariet_inf-Angular_thickness	0.19	-0.55	0.17	0.77
lh_G_pariet_inf-Supramar_thickness	0.17	-0.70	-0.02	0.62
lh_G_parietal_sup_thickness	0.26	-0.78	-0.06	1.43
lh_G_postcentral_thickness	0.14	-0.59	0.19	0.61
lh_G_precentral_thickness	0.46	-0.56	-0.75	4.87
lh_G_precuneus_thickness	0.22	-0.70	0.15	0.60
lh_G_rectus_thickness	0.25	-0.53	0.68	1.98
lh_G_subcallosal_thickness	0.07	0.00	0.03	0.48
lh_G_temp_sup-G_T_transv_thickness	0.14	-0.18	0.75	3.76
lh_G_temp_sup-Lateral_thickness	0.29	-0.31	-0.20	0.76
lh_G_temp_sup-Plan_polar_thickness	0.43	-0.28	-0.21	0.88
lh_G_temp_sup-Plan_tempo_thickness	0.19	-0.53	0.48	1.16
lh_G_temporal_inf_thickness	0.52	-1.24	0.35	0.45
lh_G_temporal_middle_thickness	0.23	-0.26	0.03	-0.15
lh_Lat_Fis-ant-Horizont_thickness	0.07	-0.31	0.72	0.82
lh_Lat_Fis-ant-Vertical_thickness	0.19	-0.41	0.53	3.39
lh_Lat_Fis-post_thickness	0.24	-0.35	0.25	0.39
lh_Pole_occipital_thickness	0.48	-0.33	0.51	2.73
lh_Pole_temporal_thickness	0.56	-0.60	0.12	0.00
lh_S_calcarine_thickness	0.48	-0.46	0.28	0.10
lh_S_central_thickness	0.40	-0.82	0.44	0.47
lh_S_cingul-Marginalis_thickness	0.28	-0.82	0.29	0.85
lh_S_circular_insula_ant_thickness	0.28	-0.51	0.37	0.90
lh_S_circular_insula_inf_thickness	0.17	-0.20	0.33	0.95
lh_S_circular_insula_sup_thickness	0.29	-0.63	0.35	0.59
lh_S_collat_transv_ant_thickness	0.30	-0.39	0.56	0.99
lh_S_collat_transv_post_thickness	0.10	-0.37	0.96	2.42
lh_S_front_inf_thickness	0.36	-1.20	0.18	0.66
lh_S_front_middle_thickness	0.26	-0.98	0.41	0.59
lh_S_front_sup_thickness	0.43	-1.03	0.19	0.64
lh_S_interm_prim-Jensen_thickness	0.00	-0.25	1.28	2.68
lh_S_intrapariet&P_trans_thickness	0.38	-1.30	-0.06	0.93
lh_S_oc_middle&Lunatus_thickness	0.18	-0.72	0.53	1.23
lh_S_oc_sup&transversal_thickness	0.18	-0.92	0.52	0.86
lh_S_occipital_ant_thickness	0.15	-0.55	0.49	0.95
lh_S_oc-temp_lat_thickness	0.22	-0.57	0.96	3.70
lh_S_oc-temp_med&Lingual_thickness	0.24	-0.25	1.03	3.51
lh_S_orbital_lateral_thickness	0.11	-0.48	0.88	1.31
lh_S_orbital_med-olfact_thickness	0.45	-0.36	0.73	1.45
lh_S_orbital-H_Shaped_thickness	0.22	-0.56	0.58	1.69
lh_S_parieto_occipital_thickness	0.29	-0.76	0.26	0.37
lh_S_pericallosal_thickness	0.13	-0.04	0.85	1.45
lh_S_postcentral_thickness	0.35	-1.12	0.38	0.68
lh_S_precentral-inf-part_thickness	0.30	-0.89	0.13	1.01
lh_S_precentral-sup-part_thickness	0.25	-0.55	-0.41	1.67

Appendix 0 Table 8. Transfer Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
lh_S_suborbital_thickness	0.03	-0.05	0.94	2.64
lh_S_subparietal_thickness	0.14	-0.71	0.28	0.22
lh_S_temporal_inf_thickness	0.30	-0.31	0.38	1.28
lh_S_temporal_sup_thickness	0.31	-0.86	0.30	0.52
lh_S_temporal_transverse_thickness	0.07	-0.11	0.43	-0.18
lh_MeanThickness_thickness	0.37	-1.00	0.10	0.53
rh_G&S_frontomargin_thickness	0.24	-0.75	0.62	1.55
rh_G&S_occipital_inf_thickness	0.14	-0.34	0.16	0.04
rh_G&S_paracentral_thickness	0.25	-0.38	0.27	1.05
rh_G&S_subcentral_thickness	0.20	-0.39	0.24	0.34
rh_G&S_transv_frontopol_thickness	0.21	-0.34	0.27	1.76
rh_G&S_cingul-Ant_thickness	0.38	-0.95	0.30	0.65
rh_G&S_cingul-Mid-Ant_thickness	0.33	-0.44	-0.04	0.76
rh_G&S_cingul-Mid-Post_thickness	0.25	-0.50	-0.01	0.95
rh_G_cingul-Post-dorsal_thickness	0.14	-0.46	-0.12	1.48
rh_G_cingul-Post-ventral_thickness	0.07	-0.03	0.22	0.85
rh_G_cuneus_thickness	0.56	-0.45	0.36	0.47
rh_G_front_inf-Opercular_thickness	0.29	-0.47	0.04	1.12
rh_G_front_inf-Orbital_thickness	0.14	-0.18	-0.16	1.11
rh_G_front_inf-Triangul_thickness	0.23	-0.38	0.28	0.31
rh_G_front_middle_thickness	0.33	-0.65	0.00	0.30
rh_G_front_sup_thickness	0.44	-0.62	0.22	0.80
rh_G_Ins_lg&S_cent_ins_thickness	0.13	-0.07	-0.09	-0.15
rh_G_insular_short_thickness	0.18	-0.06	0.09	0.11
rh_G_occipital_middle_thickness	0.22	-0.50	0.33	0.48
rh_G_occipital_sup_thickness	0.23	-0.37	0.26	1.06
rh_G_oc-temp_lat-fusifor_thickness	0.36	-0.59	0.43	0.67
rh_G_oc-temp_med-Lingual_thickness	0.46	-0.41	0.30	0.27
rh_G_oc-temp_med-Parahip_thickness	0.49	-0.31	0.17	0.46
rh_G_orbital_thickness	0.36	-0.46	0.18	0.33
rh_G_pariet_inf-Angular_thickness	0.26	-0.56	-0.17	1.04
rh_G_pariet_inf-Supramar_thickness	0.24	-0.62	0.07	0.85
rh_G_parietal_sup_thickness	0.22	-0.71	-0.13	0.82
rh_G_postcentral_thickness	0.19	-0.49	0.20	0.39
rh_G_precentral_thickness	0.40	-0.39	-1.28	6.69
rh_G_precuneus_thickness	0.24	-0.72	-0.17	1.05
rh_G_rectus_thickness	0.17	-0.55	0.06	0.76
rh_G_subcallosal_thickness	0.06	0.01	0.35	0.39
rh_G_temp_sup-G_T_transv_thickness	0.17	-0.27	-0.06	-0.04
rh_G_temp_sup-Lateral_thickness	0.31	-0.42	0.08	0.22
rh_G_temp_sup-Plan_polar_thickness	0.43	-0.23	-0.08	0.58
rh_G_temp_sup-Plan_tempo_thickness	0.16	-0.61	0.29	0.73
rh_G_temporal_inf_thickness	0.60	-1.13	0.00	0.29
rh_G_temporal_middle_thickness	0.44	-0.51	0.01	0.39
rh_Lat_Fis-ant-Horizont_thickness	0.25	-0.69	0.49	0.97
rh_Lat_Fis-ant-Vertical_thickness	0.10	-0.31	0.98	2.21

Appendix 0 Table 8. Transfer Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
rh_Lat_Fis-post_thickness	0.23	-0.38	0.22	0.00
rh_Pole_occipital_thickness	0.49	-0.34	0.36	1.42
rh_Pole_temporal_thickness	0.52	-0.55	-0.03	0.14
rh_S_calcarine_thickness	0.48	-0.44	0.12	0.47
rh_S_central_thickness	0.44	-0.79	0.08	0.61
rh_S_cingul-Marginalis_thickness	0.33	-0.91	0.19	1.28
rh_S_circular_insula_ant_thickness	0.25	-0.50	0.42	1.03
rh_S_circular_insula_inf_thickness	0.16	-0.18	0.49	1.98
rh_S_circular_insula_sup_thickness	0.32	-0.72	0.45	1.10
rh_S_collat_transv_ant_thickness	0.38	-0.36	0.55	0.76
rh_S_collat_transv_post_thickness	0.18	-0.35	0.49	1.90
rh_S_front_inf_thickness	0.27	-1.09	0.15	0.54
rh_S_front_middle_thickness	0.44	-1.37	0.23	0.37
rh_S_front_sup_thickness	0.43	-1.07	0.20	0.26
rh_S_interm_prim-Jensen_thickness	0.12	-0.52	0.61	1.22
rh_S_intrapariet&P_trans_thickness	0.42	-1.20	-0.47	1.95
rh_S_oc_middle&Lunatus_thickness	0.09	-0.50	0.95	3.38
rh_S_oc_sup&transversal_thickness	0.23	-0.80	0.32	0.45
rh_S_occipital_ant_thickness	0.20	-0.53	0.25	0.46
rh_S_oc-temp_lat_thickness	0.38	-0.52	0.14	2.55
rh_S_oc-temp_med&Lingual_thickness	0.27	-0.22	0.40	1.44
rh_S_orbital_lateral_thickness	0.22	-0.71	0.45	1.13
rh_S_orbital_med-olfact_thickness	0.27	-0.40	0.67	2.15
rh_S_orbital-H_Shaped_thickness	0.27	-1.14	0.40	0.61
rh_S_parieto_occipital_thickness	0.33	-0.78	-0.07	0.02
rh_S_pericallosal_thickness	0.20	-0.08	1.09	3.12
rh_S_postcentral_thickness	0.41	-1.06	0.12	0.46
rh_S_precentral-inf-part_thickness	0.30	-0.87	-0.14	2.32
rh_S_precentral-sup-part_thickness	0.26	-0.51	-0.68	3.83
rh_S_suborbital_thickness	0.15	-0.22	0.56	0.63
rh_S_subparietal_thickness	0.08	-0.60	-0.34	3.01
rh_S_temporal_inf_thickness	0.45	-0.50	0.72	1.74
rh_S_temporal_sup_thickness	0.32	-0.98	0.12	0.16
rh_S_temporal_transverse_thickness	0.12	-0.08	0.23	0.24
rh_MeanThickness_thickness	0.39	-1.05	0.03	0.52
Left-Lateral-Ventricle	0.14	-0.24	0.64	0.87
Left-Inf-Lat-Vent	0.08	-0.15	0.10	0.66
Left-Cerebellum-White-Matter	0.21	-0.19	-0.10	3.01
Left-Cerebellum-Cortex	0.38	-0.29	-0.84	2.33
Left-Thalamus-Proper	0.39	-0.29	0.12	0.11
Left-Caudate	0.19	-0.19	0.00	-0.05
Left-Putamen	0.36	-0.34	-0.22	0.95
Left-Pallidum	0.34	-0.23	0.19	-0.21
3rd-Ventricle	0.27	-0.29	0.96	3.49
4th-Ventricle	0.09	0.09	0.51	6.31
Brain-Stem	0.33	-0.17	0.06	0.59

Appendix 0 Table 8. Transfer Test Set Evaluation Metrics

ROI	EV	MSLL	Skew	Kurtosis
Left-Hippocampus	0.31	-0.19	0.14	0.05
Left-Amygdala	0.33	-0.23	0.33	0.41
CSF	0.08	-0.11	0.89	2.50
Left-Accumbens-area	0.35	-0.68	0.12	0.78
Left-VentralDC	0.36	-0.27	0.29	0.59
Left-vessel	0.00	0.03	-1.25	8.40
Left-choroid-plexus	0.08	-0.28	0.10	0.84
Right-Lateral-Ventricle	0.16	-0.26	0.76	1.64
Right-Inf-Lat-Vent	0.02	-0.03	-0.26	1.39
Right-Cerebellum-White-Matter	0.19	-0.19	-0.19	3.03
Right-Cerebellum-Cortex	0.41	-0.37	-0.80	2.55
Right-Thalamus-Proper	0.42	-0.26	0.04	1.34
Right-Caudate	0.20	-0.18	0.24	0.51
Right-Putamen	0.35	-0.31	-0.42	1.96
Right-Pallidum	0.24	-0.17	0.11	0.10
Right-Hippocampus	0.29	-0.19	0.13	0.00
Right-Amygdala	0.36	-0.16	0.50	4.22
Right-Accumbens-area	0.35	-0.49	0.20	0.52
Right-VentralDC	0.38	-0.28	0.41	1.70
Right-vessel	0.08	-0.02	-0.63	5.50
Right-choroid-plexus	0.15	-0.27	-0.37	1.01
SubCortGrayVol	0.43	-0.32	0.47	1.32
TotalGrayVol	0.51	-0.36	-0.23	0.49
SupraTentorialVol	0.39	-0.19	0.00	0.28
SupraTentorialVolNotVent	0.39	-0.18	0.03	0.31
avg_thickness	0.39	-1.06	0.06	0.56
EstimatedTotalIntraCranialVol	0.46	-0.37	-0.17	0.92