

1 **Rapid Screening Methods of Potato Cultivars for Low Glycemic Trait**

2 Rocio Rivas¹, Edward Dratz², Thomas Wagner³, Gary Secor⁴, Amanda Leckband¹, and David C.
3 Sands¹

4¹Department of Plant Sciences and Plant Pathology, Montana State University, Bozeman, MT,

5²Department of Chemistry and Biochemistry, Montana State University, ³Tater Seed, Everett,
6 WA, ⁴Department of Plant Pathology, North Dakota State University, Fargo, ND
7
8

9

10 **Abstract**

11 Potatoes are a dietary staple consumed by a significant portion of the world, providing
12 valuable carbohydrates and vitamins with minimal fat. Most commercially produced potatoes have
13 a high content of highly branched amylopectin starch, which generally results in a high glycemic
14 index (GI). Consumption of foods with high levels of amylopectin elicit a rapid spike in blood
15 glucose levels, which is undesirable for individuals who are pre-diabetic, diabetic, or obese. Some
16 cultivars of potatoes with lower amylopectin levels have previously been identified and are
17 commercially available in niche markets in some countries but are relatively unavailable in the
18 United States and Latin America. Among Native communities in North America and in the high
19 Andes countries of South America, some foods that include certain potato cultivars, may have
20 been used to help people mitigate what is now defined as the effects of blood sugar and obesity.
21 These cultivars are not widely available on a global market. This study utilizes three independent
22 analyses of potato starch: microscopic examination of granule structure, water absorption, and
23 spectrophotometry of iodine complexes to identify potato cultivars with low amylopectin that are
24 thought to have low GI potential. Differences among cultivars tested were detected by all three
25 types of analyses. Of the 60 potato cultivars evaluated the most promising are Huckleberry Gold,
26 Muru, Multa, Green Mountain, and an October Blue x Colorado Rose cross. Further work is
27 necessary to document the ability of these low amylopectin cultivars to reduce blood glucose spike
28 levels in human subjects.

29 **Keywords**

30 *Solanum tuberosum*; amylose, amylopectin, diabetes, microscopy; starch granules, starch swelling
31 capacity.

32 **Introduction**

33
34 Current lifestyles have led much of the general population to become relatively sedentary
35 and to adopt questionable eating habits, including high consumption of foods which appear to drive
36 an alarming rise in the incidence of obesity and Type 2 diabetes worldwide (1-3). This problem is
37 especially severe in low-income populations, where inexpensive high-energy foods tend to be
38 consumed, instead of more expensive foods with more balanced nutrition (4). Potatoes are a
39 popular staple food, available to consumers at a low cost per pound that can be easily stored. Potato
40 starch is also widely used in the food industry as a processed food ingredient (5,6). Most varieties
41 of potatoes have high levels of amylopectin starch, which is highly branched and rapidly digested.

42 High levels of amylopectin which causes a rapid spike in blood glucose level that occurs after
43 consumption of high glycemic index potatoes (2, 7–9).

44 Figure 1 (below) illustrates the difference in response of a typical individual’s blood sugar
45 levels to high and low glycemic index foods. Most varieties of potatoes are in the high glycemic
46 category of foods that people with prediabetes, diabetes, or obesity should seek to avoid. Not only
47 do high glycemic foods give a large spike in blood sugar levels, the body's response to try to bring
48 the level down tends to cause what is referred to as an undershoot, bringing levels below its
49 desirable amount. This low level of blood sugar tends to stimulate hunger and the desire to
50 consume more high glycemic foods. It follows that high glycemic foods tend to cause a “roller
51 coaster” effect on blood sugar and may cause excessive food consumption (10).

52 **Figure 1:** Blood plasma glucose response (mmol/l) from a high versus a low glycemic index (GI)
53 food. The high glycemic food tends to cause an “undershoot” in the blood glucose level below the
54 desirable level, which also tends to stimulate hunger and consumption of additional high glycemic
55 foods (10).

56 The high glycemic index of widely available potatoes presents a problematic “consumer’s
57 dilemma” for individuals and families that may not be able to afford a better-balanced, more
58 favorable diet (2, 7–9). The food industry has addressed this situation, particularly in Denmark and
59 Australia (8,11), by providing and promoting lower glycemic varieties of potatoes that are
60 relatively unavailable in the United States and most other countries. Some native communities in
61 the Andes (Bolivia, Chile, and Peru) reportedly embrace a tradition of providing low glycemic
62 tubers to people with obesity or diabetes (12).

63 Potato starch consists of two major types of carbohydrates, amylose and amylopectin (13).
64 Amylose, less abundant in most varieties of potato tubers, has a linear repeating glucose 1-4 linked
65 polysaccharide backbone structure that digests relatively slowly. Amylopectin shares the glucose
66 1-4 linked chains and additionally contains highly branched beta 1-6 side chains that produce a
67 broader structure (8,14–20) that digests much faster than amylose that can present metabolic
68 imbalances. Starch granule synthesis proceeds with the formation of amylose first and amylopectin
69 is layered on the outside of the granules. Typical high amylopectin potato starch granules have a
70 smooth rounded surface with concentric rings, formed by amylopectin (21,22).

71 Researchers have found that a higher ratio of amylose/amylopectin correlates with loss of
72 surface smoothness in starchy granules (9,23). We noticed when screening potato starch granules,
73 that some potato cultivars showed a highly disrupted starch granule surface, observed under the
74 bright field microscope at 400x, while granules from other cultivars have a smooth appearance.
75 The surface structure of the starch granules might be useful as an initial screen for identifying
76 promising varieties with higher amylose starch content, that would favor a low Glycemic Index
77 (GI) (9,16,20,25,26). In addition, the swelling capacity (water absorption capacity) of the starch
78 has also been reported to correlate with the amylose/amylopectin ratio (18,26–30).

79 Zhu et al. (31), observed that high amylose rice consumption had beneficial health effects
80 on obese rats, by reducing blood sugar levels and body weight, which would presumably also have
81 positive health benefits for humans (2,23,24,31). This might be consistent with beliefs and dietary
82 recommendations about types of plants among Native communities in Peru and Chile, but
83 unfortunately most of their traditional potato germplasm was lost during the colonization of the
84 region (12). However, potato varieties maintained in seed banks or that may already have been
85 commercialized might possess beneficial, low glycemic traits. We focused on screening 60 potato

86 varieties and selections in this study to investigate the relationship between starch granule
87 conformation, swelling power and amylose: amylopectin ratios, seeking to validate rapid screening
88 methods for low GI potential. The favorable lines revealed from this screening study can be used
89 for further agronomic, breeding, and human GI testing.

90 **Methods and Materials**

91 **Potato Material Tested**

92 A collection of 60 potato varieties and selections of potato germplasm were obtained from research
93 laboratories of Montana State University, the USDA potato collection (32), commercial seed
94 producers, and private breeder Tom Wagner, Everett, Washington. These varieties were tested for
95 low GI potential using starch granule morphology, water absorption capacity and amylose content,
96 as described below. The potato materials were chosen for their favorable agronomic traits and
97 increased in the MSU greenhouse for testing. Three tubers of each variety or selection were
98 analyzed according to the schematic flow chart shown in Figure 2.

99

100 **Figure 2:** Schematic flow chart illustrating starch analysis of 60 selected potato varieties and
101 selections.

102 **Morphological analysis of starch granules**

103 A cork-borer was used to remove a core (1cm x 1 cm) from the center medullary tissue of tubers
104 for observation. Tubers were assayed several months after harvest and storage. A subsample 5
105 mm² was pressed onto a microscope slide, covered with a cover slip and viewed with a bright
106 field microscope (Nikon Eclipse E440) at 400x magnification and photographed. Starch granules

107 from each variety or selection was numerically graded using a grading scale assessment (GSA).
108 The GSA was numerically graded according to the numerical scale described in figure 2 . Three
109 different tubers of each variety or selection were examined and photographed. Each variety was
110 given more than one rating score if the granules within the picture showed diversity, as explained
111 further in the results section.

112 **Figure 3:** Grading scale for starch granule assessment: 0= whole granules with smooth visible
113 hilum 1= whole smooth granules with dots 2= whole smooth granules with surface alterations
114 (wrinkles) 3= Whole granules with large wrinkles 4= Starch granule surface fractures 5= collapsed
115 granule

116 **Water absorption capacity of starch**

117 The swelling capacity (SC) or water absorption capacity (WAC) measures the ratio
118 between weights for the hydrated starch flour gel and dry flour samples. Samples with increased
119 amylose concentrations support lower water absorption after starch jellification (26–28,33–35).
120 WAC was evaluated using a procedure modified from Martin (34). Potato samples were freeze
121 dried and processed into powdered form in a coffee grinder to obtain potato flour
122 consistency. Thirty to forty mg of potato flour was weighed in pre-weighed 2 mL screw top glass
123 tubes and the total weight measured to four significant figures. To each tube 1.5 ml of distilled
124 water was added to the tubes capped, vortexed, placed in a thermo-mixer (Thermo Fisher) at 92
125 °C, and mixed at 800 RPM for 30 minutes. The tubes were removed and centrifuged at 1000g for
126 10 minutes at 4° C. The supernatant was aspirated from the tubes, carefully avoiding the gelatinous
127 layer, and the tubes reweighed. The capacity to absorb water was calculated by dividing the flour
128 plus water weight by the dry flour weight.

129 **Spectrophotometric Analysis of Starch Properties**

130 Rundle (36) first reported a marked difference in iodine inclusion between amylose and
131 amylopectin and developed a spectrophotometric method that allows quantification of both
132 components in mixtures. We used this spectrophotometric method, following modifications by
133 Jarvis and Walker (37) and Fajardo, et al (16), to quantify the amylose/ amylopectin ratios. 20 mg
134 of freeze-dried potato flour was suspended in 2ml of 80% ethanol in a 15 ml plastic tube and mixed
135 with a Vortex Shaker for a short period of time. Samples were centrifuged at room temperature for
136 30-60 seconds at top speed in a clinical centrifuge and the supernatant carefully removed. This
137 washing procedure was repeated twice. The resulting washed starch (pellet) was mixed with 5ml
138 of water and 5ml of 1 M KOH in the same tube and vortexed. It is important to use ultra-pure water
139 in the analysis for reproducible results and the absorbance value of the iodine solution at 550 nm
140 versus solvent is near 0.1, as the iodine solution is light and age sensitive (16). Precision in the
141 preparation of the iodine and the amylopectin/amylose solutions is crucial to getting reliable
142 amylose determinations. Since we are determining the proportion of amylose content rather than
143 the exact amount of amylose in the potato starch, there is no purification step for the isolation of
144 pure starch from the potato powder as presented by other determination methods. One ml of this
145 mixture was transferred into a 50 ml tube, neutralized with 5ml of 0.1M HCl, mixed with 0.5 ml
146 of Lugol solution (37), and adjusted to 50ml final volume with distilled water. The samples were
147 prepared in triplicate and allowed to stand for 15 min at room temperature, before measurement to
148 stabilize the inclusion of the iodine. Each sample was placed in a one cm plastic spectrophotometer
149 cuvette and absorption spectra recorded from 480-800 nm versus a distilled water blank using a
150 Genesis 10 UV Instrument (Thermo Fisher Scientific). The spectra were analyzed utilizing the
151 Vision Lite software (Thermo Fisher).

152

153 **Figure 4: Spectra of solutions of known concentrations of potato starch polysaccharide**
154 **mixtures of amylose (referred on the figure as amilosa) and amylopectin. The spectrum**
155 **marked Yodo is the iodine containing reagent solution blank.**

156 **Data Analysis**

157 The data obtained was processed using the statistical software Stat Plus: Mac LE 2009.

158 **Results Morphological Analysis of Starch Granules.**

159 We noticed that for most potato varieties and selections, virtually all the granules in each
160 sample were quite similar in appearance. In other cases, there was a high diversity of granular
161 appearance. For these cases, two scores were assigned, as shown in Table 1 with a rough
162 percentage of the granules the scores represented. Figure 5 shows that the granules in the common
163 commercial varieties Russet Burbank and Yukon Gold, known to have a high amylopectin content
164 (72% and 69% respectively), showed very smooth granules within a distribution in the sizes of
165 their granules. The previously shown grading scale in Figure 3 dictated that the granules in Figure
166 5 fit a GSA rating of 1.

167

168 Figure 5: Bright field microscopic photos of smooth starch granules of Yukon Gold (left) and
169 Russet Burbank (right) at 400x. These granules were given a GSA rating of 1.

170 In contrast, the starch granules in only a few potato varieties showed significant granular surfaces,
171 and had higher GSA values. Huckleberry Gold, Muru , and Mich-Oct have GSA ratings of 5, as
172 shown in Figure 6, where 90-95% of the total granules observed presented collapsed surfaces, the

173 other 5-10% correspond to granules with minor irregularities. The variety Multa graded 4 GSA
174 with 60-65% of the granules presenting a surface fracture level, and the 35-40% corresponded to
175 minor irregularities.

176

177 Figure 6: Starch granule images from left to right are Huckleberry Gold, Muru, Multa, Bzura, and
178 Michigan October.

179 The GSA ratings and percent starch granules in the GSA rating of 60 varieties and selections are
180 listed in Table 1.

Variety or Selection	GSA Rating	% Uniformity
Bjorna	5	55-60
Bzura	5	70-75
Huckleberry Gold	5	100
I 1035	5	90-95
Laram K'anchali	5	75-80
Muru	5	100
Olalla	5	75-80
Arma	4	45-50
Bolivian Blizzard	4	80-85
Bareroot River	4	85-90
I 1036	4	85-90
Multa	4	60-65
Green Mountain	3	60-65

Iker	3	45-50
Juice Valley	3	45-50
Lumper	3	50-55
Marble Gold	3	85-90
Monona	3	35-40
Oct Blue	3	85-90
Oct Blue x Col Rose	3	85-90
Pirola	3	65-70
Biddy Taro	2	65-70
Bison	2	90-95
C97007	2	85-90
Cherry red	2	80-85
Katelidan	2	45-50
Norland	2	75-80
Papa Amorga	2	85-90
Purple Valley	2	65-70
Red Pontiac	2	50-55
Allegany	1	20-30
Anolla	1	35-40
Asun	1	45-50
Bolivian Spring	1	90-95
Russet Burbank	1	90-95
Charlotte	1	90-95
Chella x Bulk Clover	1	90-95

Garnet Chile	1	90-95
Golden Anniversary	1	85-90
Goldra	1	80-85
Leona	1	85-90
Mich Oct x John T	1	85-90
NH x SPG	1	45-50
Nicola	1	45-50
Phytophyter	1	35-40
Picasso	1	45-50
Rush Share	1	45-50
Sangre	1	45-50
Sassy Lassy	1	85-90
Skagit Magic	1	90-95
Violet Butter	1	40-45
X gem	1	60-75
Yukon Gold	1	80—85
Arcilla-597779	0	100
Chelan	0	100
Chipitiquilla	0	100
Dark Red Norland	0	100
Fontenay	0	100
Rose Valley	0	100
Studebaker x Nordic Oct	0	100

182 Table 1: GSA rating and range of uniformity of starch granules from 60 potato varieties and
183 selections.

184 **Water absorption capacity of starch**

185 During the development of the assay to measure the water absorption capacity (WAC)
186 during starch gel formation, we found that it was essential to handle the samples carefully after the
187 final centrifugation, with minimum vibration, since removing the supernatant was difficult in some
188 samples. The WAC data for 60 varieties and selections, is shown with standard deviations in Table
189 2. The ten varieties with the lowest water absorption were the same as those with the highest GSA
190 scores. These included Huckleberry Gold, Muru, Bzura, Michigan October, and the F-1 hybrid
191 Michigan October X Colorado Rose (Figure 2). The October Blue x Colorado Rose hybrid was
192 ranked third lowest in its relative capacity to absorb water.

Variety	Water Absorption Capacity WAC		Standard Deviation
Monona	21.6	±	2.3
Chipitiquilla	21.5	±	
Skagit Magic	20.8	±	2.3
Picasso	20.5	±	
Cherry red	20.3	±	1.7
C97007	20	±	1.2
Katelidan	19.4	±	3.9
Purple Valley	18.3	±	4.5
Bison	18.2	±	2.3
Rose Valley	18	±	1.6

Garnet Chili	17.8	±	2.2
Chelan	17.4	±	
Nicola	17.3	±	2.1
Iker	17.2	±	1.6
Phytophyter	17	±	2.1
Dark Red Norland	16.9	±	4.1
Juice Valley	16.4	±	2.4
Bolivian Spring	16.1	±	3.7
Charlotte	15.8	±	2.8
Golden Anniversary	15.7	±	2.8
Goldra	15.7	±	2.7
Pirola	15.7	±	0.9
Asun	15.6	±	3
Anolla	15.5	±	0.9
Laram K'anchali	15.5	±	1.2
I 1035	15.4	±	5
Bjorna	15.4	±	2.1
Sassy Lassy	15.2	±	2.3
Chella x Bulk Clover	15.1	±	1.1
NH x SPG	14.9	±	1.2
Sangre	14.9	±	0.5
Violet Butter	14.7	±	1.3
Arma	14.5	±	2.6

I 1036	14	±	3.1
Studebaker x Nordic Oct	13.8	±	1.9
Bzura	13.7		
Rush Share	13.6	±	1.1
X Gem	13.6	±	0.8
Leona	13.4	±	1
Bolivian Blizzard	13.2	±	1.7
Olalla	13.2	±	1.6
Bareroot River	13.1	±	3.2
Arcilla-597779	12.9	±	1.2
Allegany	12.9	±	1.9
Green Mountain	12.8	±	1.3
Lumper	12.7	±	1.3
Multa	12.3	±	1.8
Biddy Taro	12.2	±	0.6
Fontenay	12.1	±	1.3
Marble Gold	12	±	1.1
Oct Blue	12	±	
Norland	11.5	±	0.8
Yukon Gold	11.2	±	2.1
Red Pontiac	10.9	±	2
Papa Amorga	10.6	±	0.7
Muru	10.5	±	0.6

Mich Oct x John T	10.4	±	0.3
Oct Blue x Col Rose	9.9	±	0.9
Huckleberry Gold	9.5	±	0.7
Russet Burbank	9.5	±	0.8

193 Table 2: Water absorption capacity (WAC) and standard deviations of 60 potato varieties and
194 selections.

195 **Determination of Amylose percentage**

196 We encountered a wide range of deviations when we assessed amylose percentage, using
197 the Jarvis and Walker approach (37). Fajardo et al (16) reported that using absorbance changes at
198 the optimum wavelengths can provide highly consistent results. Fajaro, et al (16) found that the
199 maximum absorbance of the amylose-iodine complex was at 620 nm and the maximum absorbance
200 of the amylopectin-iodine complex was at 550 nm. Thus the percent amylose can be determined:

201
$$\text{Amylose \%} = y = 107.7x - 77.4$$

202 Where $x = A_{620\text{nm}}/A_{550\text{nm}}$, measured against the reagent blank.

203 We Successfully screened 60 potato varieties in triplicate to present their respective
204 standard deviations. The amylose ratings of 60 potato varieties and selections are shown in Table
205 3. The potato varieties with a higher concentration of amylose are Green Mountain, Muru, Bzura,
206 Multa, Huckleberry Gold, and the F-1 cross October Blue X Colorado Rose. These samples also
207 had high surface alterations of their starch granules, which supports our initial hypothesis that this
208 granule feature was due to higher amylose.

209

Particulate Ammonia (%)	Ammonia (%)
Green Mountain	60.9
Muru	59.0
Bzura	50.4
Multa	44.7
Huckleberry Gold	44.7
Oct Blue x Col Rose	43.8
Laram K'anchali	43.8
NH x SPG	40.4
Lumper	39.7
Anolla	39.5
Bjorna	38.3
Iker	37.1
Sassy Lassy	37.1
X Gem	36.6
Chipitiquilla	35.7
Red Pontiac	35.0
Bolivian Blizzard	34.7
Phytophyter	34.5
Papa Amorga	34.2
Skagit Magic	34.2
Oct Blue	33.6
Dark Red Norland	33.3
Juice Valley	33.3

Studebaker x Nordic Oct	32.9
Cherry Red	32.8
Marble Gold	32.7
Fontenay	32.5
Nicola	32.3
Leona	32.3
Arma	31.9
Mich Oct x John T	31.8
Arcilla-597779	31.4
Pirola	31.2
Rose Valley	31.2
Violet Butter	30.9
Norland	30.8
Bolivian Spring	30.7
Katelidan	30.7
I 1035	30.4
I 1036	30.4
Yukon Gold	30.4
Olalla	30.0
Biddy Taro	29.8
Bareroot River	29.2
Sangre	28.9
Goldra	28.8

210

211

212

213

Garnet Chile	28.3
Asun	27.7
Monona	27.6
C97007	27.4
Charlotte	27.1
Chelan	26.6
Rush Share	26.6
Bison	26.2
Golden Anniversary	25.0
Allegany	24.1
Chella x Bulk Clover	24.0
Picasso	23.9
Purple Valley	23.9
Russet Burbank	23.7

Table 3: Amylose content (%) of 60 potato varieties and selections.

Variety	% Amylose Content	Starch Granule Ranking	Water Absorption
Green Mountain	61	3	13
Muru	59	5	10
Bzura	50	5	13
Huckleberry Gold	45	5	9
Multa	45	5	12
Oct. Blue x Col Red	44	3	9
Monona	28	2	22
Picasso	24	1	21

Purple Valley	24	2	18
---------------	----	---	----

214 **Table 4:** Compilation of traits of seven selected cultivars compared to three cultivars with high
 215 amylose content.

216 **Calculation of potential glycemic index based on amylose percentage**

217 Moreira’s study in 2012 (38) measured the blood sugar levels in subjects two hours after
 218 eating a starchy meal with a known concentration of amylose, and reported a correlation
 219 equation where x is the amylose percentage of total starch content and GI_p is the GI potential
 220 value (GI_p). Using this calculation and the amylose content of potato varieties and selections, a
 221 GI_p can be calculated (Table 5).

Variety	% amylose	GI potential	Variety	% amylose	GI potential
Green Mountain	60.9	(14.69)	Mich Oct x John T	31.8	73.28
Muru	59.0	(8.94)	Arcilla-597779	31.4	74.43
Bzura	50.4	16.93	Pirola	31.2	75.14
Huckleberry	44.7	34.18	Rose Valley	31.2	75.14
Multa	44.7	34.18	Violet Butter	30.9	75.90
Oct Blue x Col Rose	43.8	37.06	Norland	30.8	76.17
NH x SPG	40.4	47.12	Bolivian Spring	30.7	76.65
Lumper	39.7	49.35	Katelidan	30.7	76.66
Anolla	39.5	49.99	I 1035	30.4	77.31
Bjorna	38.3	53.57	Yukon Gem	30.4	77.31
Iker	37.1	57.15	I 1036	30.4	77.51
Sassy Lassy	37.1	57.22	Olalla	30.0	78.72
X gem	36.6	58.62	Biddy Taro	29.8	79.28

Chipitiquilla	35.7	61.45	Bareroot River	29.2	81.13
Red Pontiac	35.0	63.54	Sangre	28.9	81.99
Bolivian Blizzard	34.7	64.46	Goldra	28.8	82.25
Phytophyter	34.5	65.17	Garnet Chile	28.3	83.65
Iaram K'anchali	34.2	65.81	Asun	27.7	85.70
Skagit Magic	34.2	65.81	Monona	27.6	85.73
Papa Amorga	34.2	65.96	C97007	27.4	86.43
Oct Blue	33.6	67.76	Charlotte	27.1	87.37
Dark Red Norland	33.3	68.68	Chelan	26.6	88.81
Juice Valley	33.3	68.75	Rush Share	26.6	88.89
Studebaker x Nordic Oct	32.9	69.77	Bison	26.2	89.97
Cherry Red	32.8	70.12	Golden Anniversary	25.0	93.81
Marble Gold	32.7	70.58	Allegany	24.1	96.44

222

223 Table 5: Predicted glycemic index (GIP) of 52 potato varieties and selections calculated using the
 224 equation described by Moreira (38). GIp = Glycemic Index potential.

225 The GIp for Green Mountain and Muru, the two varieties with the highest percentage of
 226 amylose have negative values, which could suggest that such high amylose samples were outside
 227 the range of Moreira's equation.

228 Pearson's correlation coefficient tests were performed to study the possible relationship
229 between granular surface appearance of starch granules (GSA), granule absorption capacity
230 (GAC), and amylose content by spectrophotometric analysis in multiple potato varieties and
231 selections. There was a positive correlation between GSA scores and the amylose content of the
232 potato starch ($r^2 = 0.46$, $p < 0.05$). There was a negative correlation between GAC and amylose
233 content ($r^2 = -0.34$, $p > 0.01$), confirming the hypothesis that amylose inhibits water absorption by
234 potato starch. These results agree with observations with peas since high amylose peas are more
235 wrinkled and cause a much lower insulin response in humans than smooth peas (41). The wrinkled
236 peas also have a lower capacity to absorb water (42).

237 **Discussion**

238 Nutrition data suggest that consumption of potatoes with higher concentrations of amylose
239 would have a lower Glycemic Index and would be expected to provide a health benefit for people
240 that have prediabetes, diabetes and/or obesity. A lower sugar release during digestion is expected
241 to cause a lower sugar spike and lower release of insulin, due to differences of digestibility of
242 amylose and amylopectin (4,12,31,41,43). Furthermore, high insulin release tends to lead to an
243 undershoot in desirable blood sugar levels (hypoglycemia), stimulating hunger and potential
244 weight gain.

245 From a sample of over 60 cultivars, we have been able to identify the six-candidate
246 cultivars for further agronomic evaluation for consumer acceptability. Those samples are as
247 follows: Huckleberry Gold, Muru, Bzura, October Blue x Colorado Rose, Multa, and Green
248 Mountain. In addition to the consumer dilemma on choices of affordable healthy foods, there is
249 the Breeders' Dilemma that is the conflict between improved food nutrition that often reduces

250 agronomic yield, which usually drives the choice of varieties (40). To pursue this aim, we
251 identified patterns in properties of potato starch that could be used in rapid screening. By screening
252 of 60 potato lines A pattern was identified in the microscopic appearance of starch granules,
253 confirmed by direct starch content assays, permitting a rapid initial screening of candidate potato
254 varieties for increased amylose content and potential lower Glycemic Index. This rapid screening
255 system will allow elevation of nutrition as a priority in plant breeding.

256 This work now will be followed by glycemic index measurements and agronomic fitness (40,41)
257 on the most promising potato varieties.

258 **Acknowledgement**

259 This paper was supported by Western SARE Project SW12-108, Montana State University
260 Agricultural Experiment Station.

261 **References**

- 262 1. Sami, Waqas, et al. "Effect of diet on type 2 diabetes mellitus: A review." *International*
263 *journal of health sciences* 11.2 (2017): 65.
- 264 2. Camire ME, Kubow S, Donnelly DJ. Potatoes and human health. *Crit Rev Food Sci Nutr*.
265 2009;49(10):823–40.
- 266 3. Seidell JC. Obesity, insulin resistance and diabetes - A worldwide epidemic. In: *British*
267 *Journal of Nutrition*. 2000.
- 268 4. Diabetes Federation I. Worldwide toll of diabetes. 2019.
269 <https://www.diabetesatlas.org/en/sections/worldwide-toll-of-diabetes.html>
270
- 271 5. de Haan S, Rodriguez F, Singh J, Kaur L. Chapter 1 - Potato Origin and Production. In:
272 *Advances in Potato Chemistry and Technology*. 2016. p. 1–32.

- 273 6. Zaheer K, Akhtar MH. Recent advances in potato production, usage, and nutrition. Crit Rev
274 Food Sci Nutr. 2016;
- 275 7. Fernandes G, Velangi A, Wolever TMS. Glycemic index of potatoes commonly consumed
276 in North America. J Am Diet Assoc. 2005;105(4).
- 277 8. Ek KL, Brand-Miller J, Copeland L. Glycemic effect of potatoes. Food Chem. 2012;133(4).
- 278 9. Vamadevan V, Bertoft E. Structure-function relationships of starch components. Vol. 67,
279 Starch/Staerke. 2015.
- 280 10. Wolever TMS, DJA Jenkins, AL Jenkins and RG Josse. 1991. The glycemic index:
281 Methodology and clinical implications. Am J Clin Nutr 54:846-854
- 282 11. Tenduis M, Low-carb potatoes on the rise. Fresh Plaza. 2018; 7: 18. Available from:
283 <https://www.freshplaza.com/article/2193080/low-carb-potatoes-on-the-rise>.
- 284 12. Sarkar, D., Walker-Swaney, J., & Shetty, K. (2020). Food diversity and
285 indigenous
286 food systems to combat diet-linked chronic diseases. *Current developments in*
287 *nutrition, 4* (Supplement_1), 3-11.
- 288 13. Hofvander P, Andersson M, Larsson CT, Larsson H. Field performance and starch
289 characteristics of high-amylose potatoes obtained by antisense gene targeting of two
290 branching enzymes. Plant Biotechnol J. 2004;2(4).
- 291 14. Singh J, Kaur L. Advances in Potato Chemistry and Technology. Advances in Potato
292 Chemistry and Technology. 2009.

- 293 15. Chung HJ, Li XQ, Kalinga D, Lim ST, Yada R, Liu Q. Physicochemical properties of dry
294 matter and isolated starch from potatoes grown in different locations in Canada. *Food Res*
295 *Int.* 2014;57.
- 296 16. Fajardo, D., Jayanty, S.S., Jansky, S.H. Rapid High Throughput Amylose Determination in
297 Freeze Dried Potato Tuber Samples. *J. Vis. Exp.* (80), e50407, doi:10.3791/50407 (2013).
- 298 17. Hoover R. Composition, molecular structure, and physicochemical properties of tuber and
299 root starches: A review. *Carbohydr Polym.* 2001;45(3).
- 300 18. Karlsson ME, Leeman AM, Björck IME, Eliasson AC. Some physical and nutritional
301 characteristics of genetically modified potatoes varying in amylose/amylopectin ratios.
302 *Food Chem.* 2007;100(1).
- 303 19. Moongngarm A. Chemical compositions and resistant starch content in starchy foods. *Am*
304 *J Agric Biol Sci.* 2013;8(2).
- 305 20. Mukerjea R, Mukerjea R, Robyt JF. Starch biosynthesis: experiments on how starch
306 granules grow in vivo. *Carbohydr Res.* 2009;344(1).
- 307 21. Pietrzyk S, Fortuna T, Juszczak L, Gałkowska D, Bączkiewicz M, Łabanowska M, et al.
308 Influence of amylose content and oxidation level of potato starch on acetylation, granule
309 structure and radicals' formation. *Int J Biol Macromol.* 2018;106.
- 310 22. Smith AM. The biosynthesis of starch granules. Vol. 2, *Biomacromolecules.* 2001.
- 311 23. Man J, Yang Y, Zhang C, Zhou X, Dong Y, Zhang F, et al. Structural changes of high-
312 amylose rice starch residues following in vitro and in vivo digestion. *J Agric Food Chem.*
313 2012;60(36):9332–41.

- 314 24. Shu X, Backes G. in a Barley Variety Collection. 2012;
- 315 25. Tester RF, Karkalas J, Qi X. Starch - Composition, fine structure and architecture. Journal
316 of Cereal Science. 2004.
- 317 26. Kaur L, Singh J, McCarthy OJ, Singh H. Physico-chemical, rheological and structural
318 properties of fractionated potato starches. J Food Eng. 2007;82(3).
- 319 27. Singh N, Sandhu KS, Kaur M. Characterization of starches separated from Indian chickpea
320 (*Cicer arietinum* L.) cultivars. J Food Eng. 2004;
- 321 28. Zheng Y, Wang Q, Li B, Lin L, Tundis R, Loizzo MR, et al. Characterization and prebiotic
322 effect of the resistant starch from purple sweet potato. Molecules. 2016;21(7).
- 323 29. Li G, Zhu F. Amylopectin molecular structure in relation to physicochemical properties of
324 quinoa starch. Carbohydr Polym. 2017;164.
- 325 30. Vamadevan V, Bertoft E. Observations on the impact of amylopectin and amylose structure
326 on the swelling of starch granules. Food Hydrocoll. 2020;103.
- 327 31. Zhu L, Gu M, Meng X, Cheung SCK, Yu H, Huang J, et al. High-amylose rice improves
328 indices of animal health in normal and diabetic rats. Plant Biotechnol J. 2012;10(3):353–
329 62.
- 330 32. USDA.Potato seed bank. Available from: [https://wisconsinpotatoes.com/wp-](https://wisconsinpotatoes.com/wp-content/uploads/2020/03/DC-FlyIn-flyer.jpg)
331 [content/uploads/2020/03/DC-FlyIn-flyer.jpg](https://wisconsinpotatoes.com/wp-content/uploads/2020/03/DC-FlyIn-flyer.jpg)
- 332 33. Shi YC, Capitani T, Trzasko P, Jeffcoat R. Molecular structure of a low-amylopectin starch
333 and other high-amylose maize starches. J Cereal Sci. 1998;27(3).

- 334 34. Martin JM, Talbert LE, Habernicht DK, Lanning SP, Sherman JD, Carlson G, et al. Reduced
335 Amylose Effects on Bread and White Salted Noodle Quality. *Cereal Chem.* 2004;81(2).
- 336 35. Sasaki T, Matsuki J. Effect of wheat starch structure on swelling power. *Cereal Chem.*
337 1998;75(4).
- 338 36. Rundle RE. The Configuration of Starch in the Starch-Iodine Complex. V. Fourier
339 Projections from X-Ray Diagrams. *J Am Chem Soc.* 1947;69(7).
- 340 37. Jarvis CE, Walker JRL. Simultaneous, rapid, spectrophotometric determination of total
341 starch, amylose and amylopectin. *J Sci Food Agric.* 1993;63(1).
- 342 38. Moreira TS. The Role of Starch physicochemical properties in determining the Glycemic
343 Index of Novel Potato varieties. University of Toronto.; 2012.
- 344 39. Soh N, Brand-Miller J. The glycaemic index of potatoes: the effect of variety, cooking
345 method and maturity. *Eur J Clin Nutr.* 1999 Apr;53(4):249–54.
- 346 40. Sands DC, Morris CE, Dratz EA, Pilgeram AL. Elevating optimal human nutrition to a
347 central goal of plant breeding and production of plant-based foods. Vol. 177, *Plant Science.*
348 2009.
- 349 41. Megdal PA, Siemsen D, Sands D, Dratz EA, Handelman GJ. Facile fingerstick insulin
350 analysis: Application to monitoring postprandial insulin responses to snack foods. *J*
351 *Diabetes.* 2010;2(1):28–35.
- 352 42. Kosson R, Czuchajowska Z, Pomeranz Y. Smooth and Wrinkled Peas. 1. General Physical
353 and Chemical Characteristics. *J Agric Food Chem.* 1994;42(1).

- 354 43. Wong, J. M., & Jenkins, D. J. (2007). Carbohydrate digestibility and metabolic effects. *The*
355 *Journal of nutrition*, 137(11), 2539S-2546S.

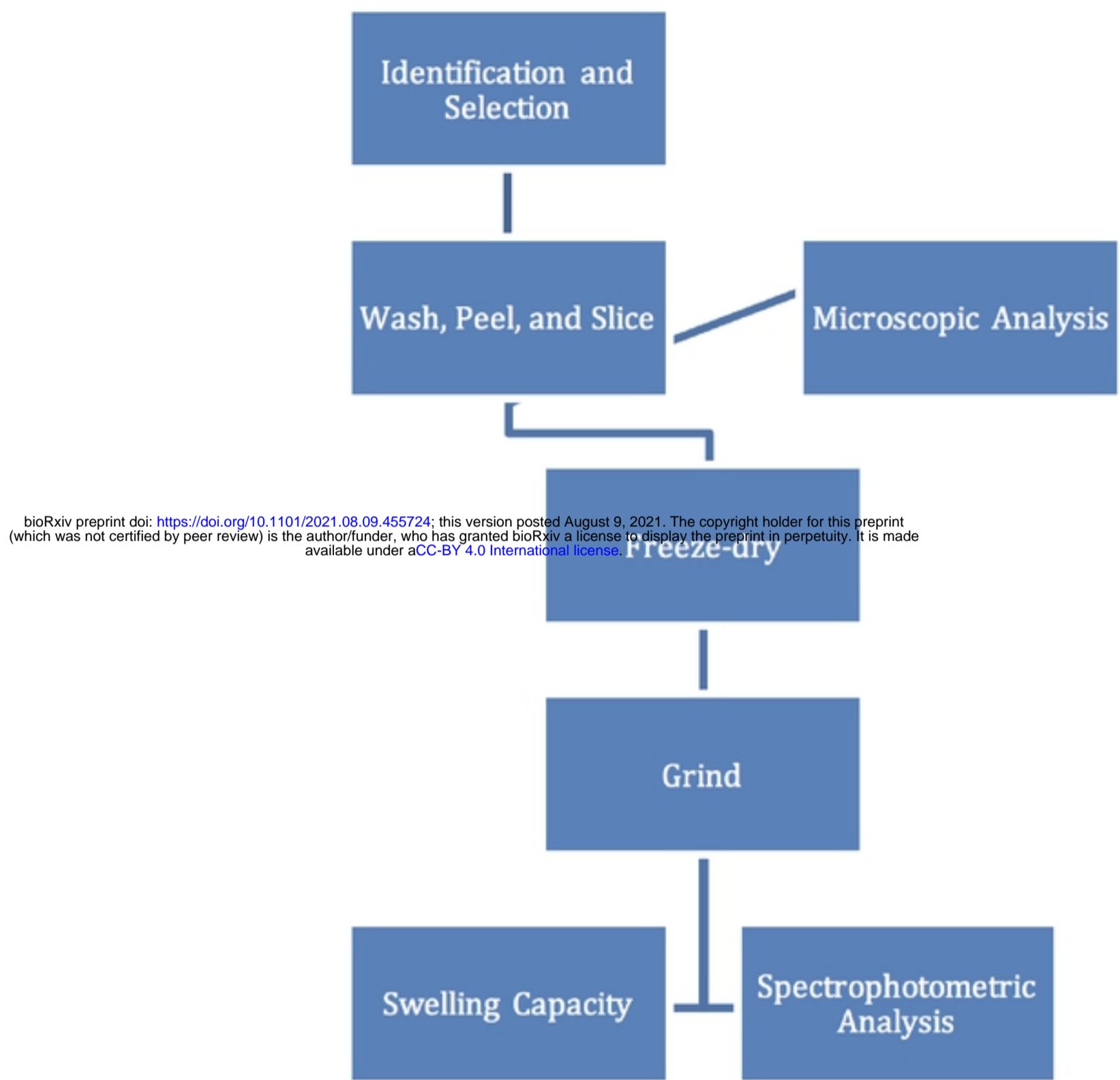


FIG 2

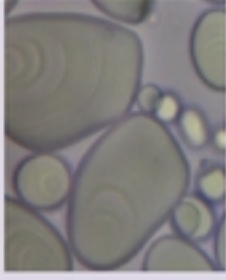
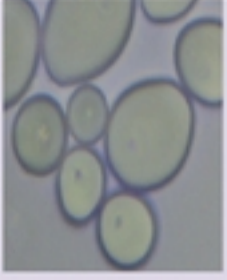




Description	Whole smooth granule, halo visible	Whole smooth granule with points	Whole smooth granule with wrinkles	Whole granule with surface alteration	Granule with fracturing	Collapsed granule
Images						
Ranking	0	1	2	3	4	5

Fig 3

bioRxiv preprint doi: <https://doi.org/10.1101/2021.08.09.455724>; this version posted August 9, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a [CC-BY 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

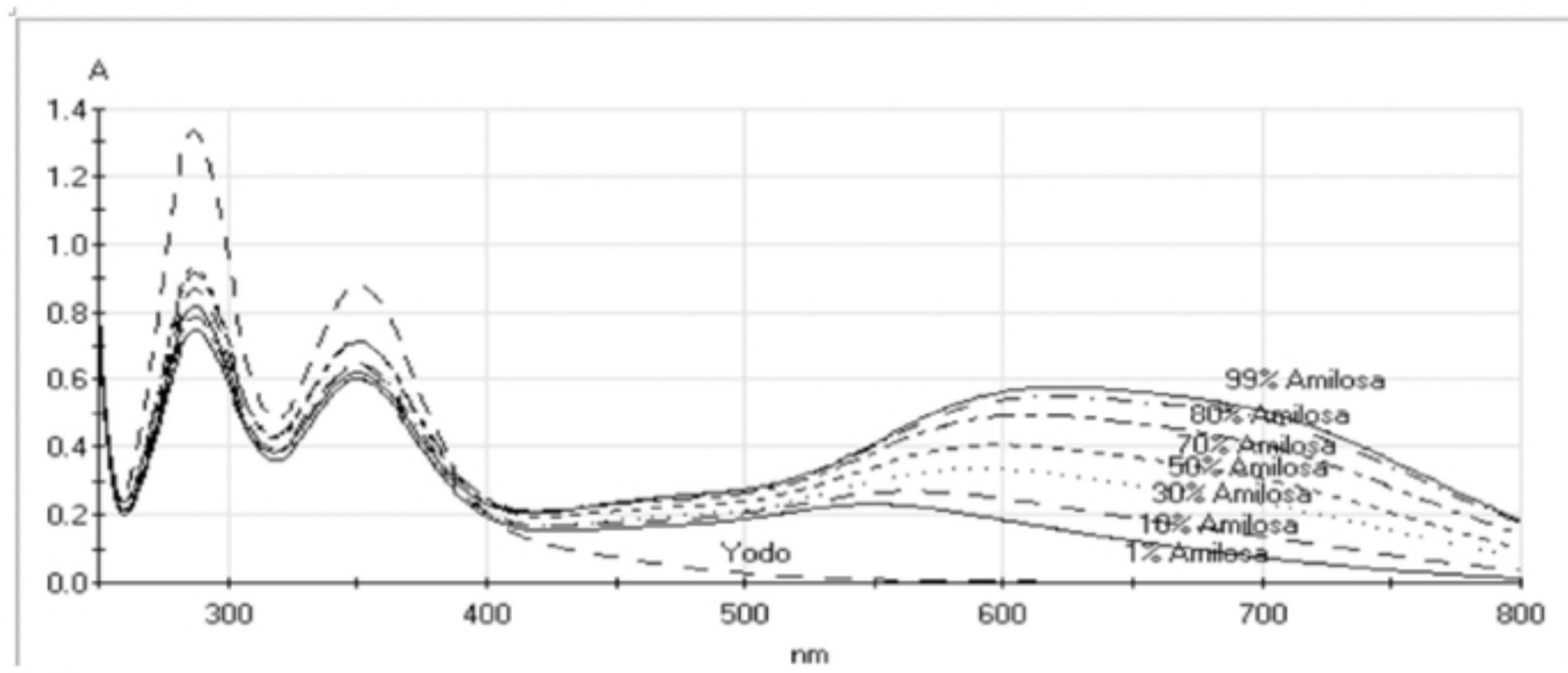


Fig 4

bioRxiv preprint doi: <https://doi.org/10.1101/2021.08.09.455724>; this version posted August 9, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a [CC-BY 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

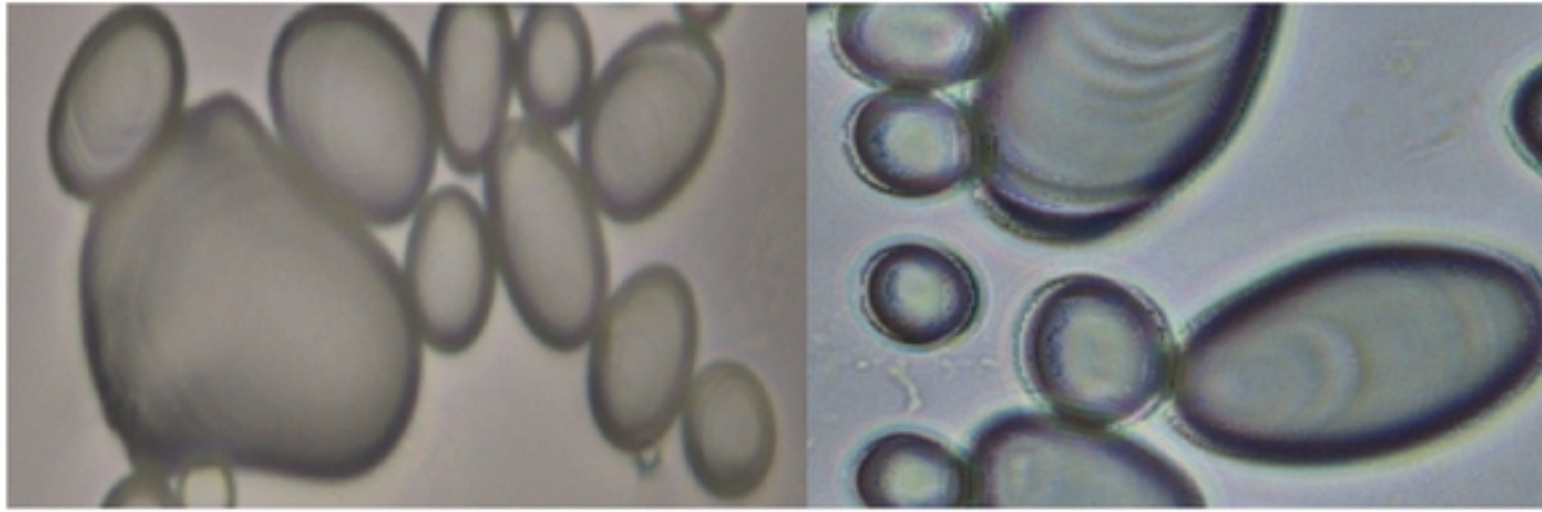
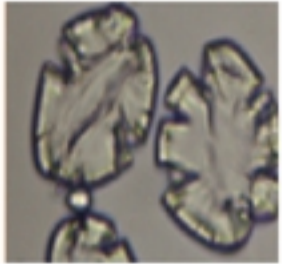
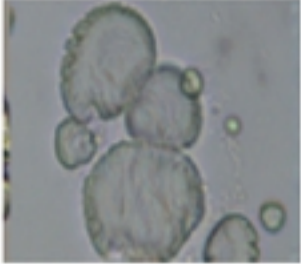

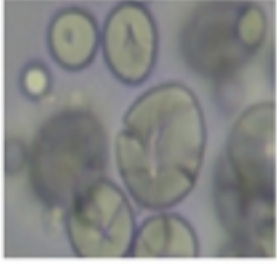
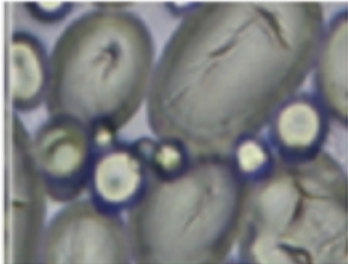
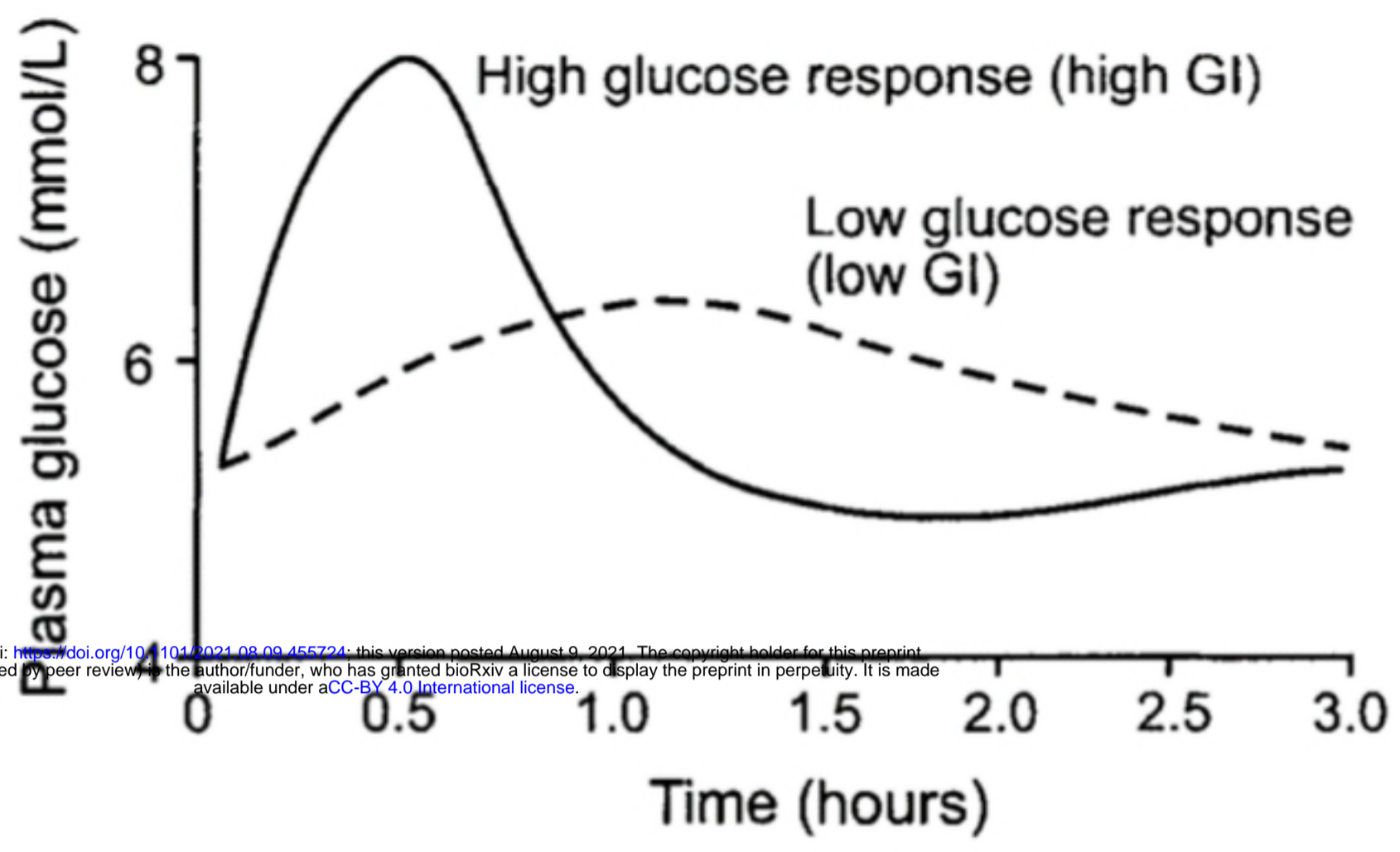


Fig 5

bioRxiv preprint doi: <https://doi.org/10.1101/2021.08.09.455724>; this version posted August 9, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a [CC-BY 4.0 International license](#).

Type	Huckleberry	Muru	Multa	Bzura	Mich. October
Image					

bioRxiv preprint doi: <https://doi.org/10.1101/2021.08.09.455724>; this version posted August 9, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a [CC-BY 4.0 International license](#).



bioRxiv preprint doi: <https://doi.org/10.1101/2021.08.09.455724>; this version posted August 9, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a [CC-BY 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

FIG 1

Fig 2

Figure 1