Balancing growth amidst salinity stress – lifestyle perspectives from the extremophyte model *Schrenkiella parvula*

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One sentence summary: *Schrenkiella parvula* salt-resilient growth is facilitated by uncompromised primary root growth, expansion of xylem vessels, maintenance of leaf water status and photosynthesis, and early flowering.

Abstract

Schrenkiella parvula, a leading extremophyte model in Brassicaceae, can grow and complete its life cycle under multiple environmental stresses, including high salinity. While foundational genomic resources have been created for *S. parvula*, a comprehensive physiological or structural characterization of its salt stress responses is absent. We aimed to identify the influential traits that lead to stress-resilient growth of this species. We examined salt-induced changes in the physiology and anatomy of *S. parvula* throughout its lifecycle across multiple tissues. We found that *S. parvula* maintains or even exhibits enhanced growth during various developmental stages at salt stress levels known to inhibit growth in Arabidopsis and most crops. The resilient growth of *S. parvula* was associated with key traits that synergistically allow continued primary root growth, expansion of xylem vessels across the root-shoot continuum, and a high capacity to maintain tissue water levels by developing larger and thicker leaves while facilitating continued photosynthesis during salt stress. These traits at the vegetative phase were followed by a successful transition to the reproductive phase via early flowering, development of larger siliques, and production of viable seeds during salt stress. Additionally, the success of self-fertilization during early flowering stages was dependent on salt-induced filament elongation in flowers that aborted in the absence of salt. Our results suggest that the maintenance of leaf water status and enhancement of selfing in early flowers to ensure reproductive success, are among the most influential traits that contribute to the extremophyte lifestyle of *S. parvula* in its natural habitat.

Introduction

Soil characteristics, water availability, and light and temperature regimes set boundaries for plant growth and dictate how plants complete their life cycles. Balancing plant growth with environmental stress responses is a constant challenge faced by all plants. This balance is

epitomized by extremophytes, which are plants adapted to thrive in extreme environmental conditions than mesophytes. Therefore, extremophytes provide insights on evolutionarily-tested lifestyle strategies that have proven successful in extreme environments (Lloyd and Oreskes, 2018; Rodell et al., 2018; Schlenker and Auffhammer, 2018; Solis et al., 2020). Knowledge transfer from extremophyte-focused research to design innovative crops that can show resilient growth under stress can be a fruitful strategy when envisioning global food security and expansion of agricultural lands to marginal lands (Oh et al., 2012; Bechtold, 2018; Kazachkova et al., 2018; Zandalinas et al., 2021). The recent genome explorations in a wider range of plants makes this an ideal time for mining representative extremophyte traits.

Salt tolerance is a complex trait that requires integration of physiological, anatomical, and metabolic responses (Cheeseman, 2013; Barros et al., 2021). These multifaceted traits cannot be achieved in stress-sensitive crops simply through the acclamatory response of a few genes (Roy et al., 2014). The complexity of these interactions is implied by the fact that breeding has had limited success in conferring salinity tolerance despite numerous attempts (Yamaguchi and Blumwald, 2005; Cheeseman, 2013; Roy et al., 2014; Barros et al., 2021). An understanding of the extent of tradeoffs between plant growth and stress tolerance is required to assess how selected traits, genes, or pathways affect short- or long-term plant growth and fitness in extremophytes. Such foundational studies are lagging for all the leading extremophyte models compared to the breadth of tools and molecular resources available to identify candidate genes in these species.

Schrenkiella parvula (Schrenk) D.A.German & Al-Shehbaz (previously known as *Thellungiella parvula* or *Eutrema parvulum*) is a leading extremophyte model in the Brassicaceae family (Zhu, 2015; Ali and Yun, 2017; Kazachkova et al., 2018; Krämer, 2018). *S. parvula* shares many traits with the model plant, *A. thaliana*, that make it an excellent model, including comparable genome size (Dassanayake et al., 2011), similar lifecycle duration, self-pollination, and prolific seed production (Table 1 and Figure 1). In the ten years following the release of its genome, *S. parvula* has been developed as a model system equipped with primary molecular tools to explore genetic mechanisms underlying plant abiotic stress tolerance (Dassanayake et al., 2011; Oh et al., 2014; Wang et al., 2018; Pantha et al., 2021; Wang et al.,

2021). *Schrenkiella parvula* is remarkably tolerant to high salinity compared to closely related Brassicaceae species (Orsini et al., 2010) and is categorized as a halophyte based on its capacity to complete its life cycle when grown at 200 mM NaCl or higher (Flowers and Colmer, 2008). In addition to high sodium, *S. parvula* is also uniquely adapted to cope with multiple edaphic factors found in its native habitat, including boron, potassium, and lithium, among other salts, that exist at levels toxic for most crops (Helvaci et al., 2004; Oh et al., 2014; Hajiboland et al., 2018; Tug et al., 2019). Its preferential distribution near saline lakes in the Irano-Turanian region makes it a robust extremophyte model to investigate multiple environmental stresses that are often found as compound stresses in many marginal agricultural landscapes.

Despite a growing body of genomic resources for *S. parvula* (Jarvis et al., 2014; Ali et al., 2018; Wang et al., 2021), a systematic study that describes its life history, and the physiological, and structural features that are associated with adaptations to environmental stresses, especially salt stress, is absent. In this study, we evaluated *S. parvula* stress adaptations at both the structural and physiological levels. We assessed adaptations that are likely to be influential in balancing growth with stress tolerance and identified inducible adaptive traits that were the most prominent features in a multivariate space across developmental stages, tissue types, and response types. We propose these inducible traits to be hallmarks of a stress-adapted lifestyle for a fast-growing annual in saline soils demonstrating the utility of *S. parvula* as an extremophyte model plant.

Results

Schrenkiella parvula adjusts root growth, structure, and form under high salinity

Schrenkiella parvula roots, in contrast to those of *A. thaliana*, maintained uninterrupted root growth in response to salt stress compared to control conditions (Figure 2A). Remarkably, *S. parvula* primary root length did not display any growth retardation even after longer durations or higher salt concentrations tested, while *A. thaliana* showed growth inhibition at 100 mM NaCl (Figure 2B). The same trend was observed for lateral root growth, although the emergence of lateral roots was delayed in *S. parvula* even at control conditions compared to *A. thaliana* (Figure 2C). The higher tolerance to salinity shown by *S. parvula* roots compared to *A.*

118 *thaliana* roots was consistent when plants were grown under a longer photoperiod and at
119 higher nutrient levels (Figure S1). Under these conditions, a 125 mM NaCl treatment caused a
120 growth stimulating effect on *S. parvula* primary root length, but inhibited root growth in *A.*
121 *thaliana* (Figure S1B). Stronger inhibition of both average lateral root length and number was
122 imposed by 175 and 225 mM NaCl, which could explain the reduced total lateral root length
123 (Figure S1C to E). Taken together, salt stress imposed by 125 mM NaCl promoted primary root
124 growth and average lateral root length, while a higher salt concentration of 175 mM NaCl
125 imposed less inhibition on main root growth and lateral root density in *S. parvula* compared to
126 *A. thaliana*. Compared to primary and lateral roots, root hair development in *S. parvula* was
127 sensitive to high salinity; root hairs showed a consistent decrease in length at higher salinities
128 or during longer exposure times to salt stress (Figures 2D and S2). The ability of *S. parvula* to
129 maintain or enhance primary root growth under high salinity remained as a consistent trait
130 shown by mature plants over longer durations and higher salinities tested using 250 mM NaCl
131 in a hydroponic growth medium (Figure S3).
132 *Arabidopsis thaliana* roots showed halotropism by growing away from salt (Figure S4)
133 (Galvan-Ampudia et al., 2013). Interestingly, under comparable growth conditions, *S. parvula*
134 showed salt-insensitive primary root growth where it grew towards higher salt concentration
135 without changing its course, further exemplifying the higher tolerance to high salinity (Figure
136 S4).
137 To investigate the potential effect of salt stress on root anatomy, we examined the
138 tissue level structural responses in roots of 8-week-old *S. parvula* plants that had been treated
139 for 4 weeks with 150 mM NaCl (Figure 3A). These traits were catalogued from root tips to
140 mature roots (Figure S5). In young roots, the length of the root tip (measured as the distance
141 from the tip to the emergence site of the first root hair) was longer under high salinity
142 conditions compared to control conditions (Figure 3B). However, we did not observe any other
143 anatomical trait adjustments to salinity in young roots of *S. parvula* (Figure 3C). Interestingly, *S.*
144 *parvula* roots develop an extra cell layer between the cortex and endodermis compared to *A.*
145 *thaliana* (Brady et al., 2007), formed early on in its root development in both control and salt-
146 treated samples (Figure S5A and C). In response to high salinity, xylem tissues in mature roots

147 significantly increased in area under high salinity (Figure 3D). However, this did not affect the
148 overall mature root area. The xylem tissue expansion was primarily caused by increase in the
149 area of vessels. The extra space taken up by the xylem tissue was compensated for by reduction
150 of the cortical air spaces (unstructured aerenchyma) in mature roots (Figure 3D).
151
152 ***Schrenkiella parvula* shoots change in form and function to response to salt stress**
153 Structural adjustments in response to long-term salt stress extended to shoot tissues in
154 *S. parvula* (Figures 4 and S6). Long-term salt stress leads to larger vessels in *S. parvula* shoots
155 (Figure 4A) to maintain the root-shoot continuum with root vessel elements that increased in
156 size when exposed to high salinity (Figure 3D). The expansion in vessel element size in shoots
157 was matched by a reduction in cambial and cortical zones that kept the average shoot area
158 constant between control and salt-treated plants (Figure 4A), while other cell and tissue layers
159 in the shoot remained unchanged (Figure S6B).
160 Leaves that developed during long-term salt treatments in *S. parvula* exhibited
161 increased succulence, as indicated by increase in leaf thickness across the leaf including the
162 midrib (Figures 4B and S7). Increased endoreplication induced by salinity can cause cell size
163 expansion and succulence in the extremophyte, *Mesembryanthemum crystallinum* (Barkla et
164 al., 2018). We therefore tested whether the increased succulence in *S. parvula* leaves in
165 response to salt coincided with endoreplication by quantifying ploidy of leaf cells. Indeed, the
166 proportion of leaf cells with higher ploidy increased with increased salinity, causing a significant
167 rise in the endoreplication index under 250 mM NaCl compared to the control condition (Figure
168 4C). *Schrenkiella parvula* leaves showed both structural and functional adjustments to salt
169 treatments that correlated with its capacity to maintain relative water content (Figure 5), but
170 the total leaf number per plant remained similar between control and salt-treated plants
171 (Figure 5A). Long-term salt treatments not only increased leaf succulence, but also leaf area
172 (Figure 5A), suggesting a growth-promoting effect at least with the long-term 150 mM NaCl
173 treatment. *Schrenkiella parvula* was photosynthetically active and maintained growth during
174 long-term salt treatments at high salinities (Figure 4), but there was a tradeoff in increasing salt
175 on shoot development at salinities surpassing 150 mM (Figure S8).

176 Leaf shape, angle and phyllotaxis, stomatal distribution, and boundary layer thickness
177 caused by thick cuticles or trichomes largely contribute to efficient use of water by plants (Yoo
178 et al., 2009). *Schrenkiella parvula* leaves are amphistomatous (i.e. stomata exist on both abaxial
179 and adaxial surfaces), linear-lanceolate with a narrower base, and trichomeless (Table 1, Figure
180 5A and 5B). In *S. parvula*, stomatal density did not change in fully-mature leaves (Figure 5B).
181 However, stomatal regulation was adjusted with increasing salinities as indicated by decreased
182 stomatal conductance (Figure 5C) and decreased CO_2 assimilation, notably at much higher
183 salinities for *S. parvula* than observed for *A. thaliana* (Figure 5D). While 50 mM salt treatment
184 was sufficient to cause a decrease in stomatal conductance and photosynthesis in *A. thaliana*, *S.*
185 *parvula* treated with 100 mM NaCl was able to maintain stomatal conductance and CO_2
186 assimilation at levels indistinguishable from those measured in plants grown under control
187 conditions. Concomitantly, *S. parvula* maintained lower average leaf temperatures than *A.*
188 *thaliana* (Figure 5E). Remarkably, *S. parvula* maintained relative leaf water content at all tested
189 salinities up to 250 mM NaCl (Figure 5F).
190 Utilizing the Qubit/PSI PlantScreen^{TM} Compact high-throughput phenomics system, we
191 undertook a comparative analysis of *A. thaliana* and *S. parvula* shoot morphometrics (RGB
192 digital camera), leaf water content (infra-red camera), and photochemistry (PAM chlorophyll
193 fluorescence). Figure S9A demonstrates that the phenomics system was able to simultaneously
194 capture morphological characteristics specific to each species. *A. thaliana* has a rosette
195 structure (i.e. less protrusions from a perfect circle), and therefore exhibited higher
196 compactness). In contrast *S. parvula* leaves on a spiral phyllotaxy (Figure 1 and Table 1) that
197 increasingly protrude as the plant grows, reflected a lower compactness in older plants
198 regardless of salt concentrations. Salt stress led to a statistically significant reduction in *A.*
199 *thaliana* leaf area but had no effect on *S. parvula* leaves (Figure S9B). *S. parvula* was also able to
200 maintain leaf water content and maximum quantum efficiency of PSII under salt stress (Figure
201 S9C and D, respectively) whereas *A. thaliana* exhibited a salt stress-mediated reduction in both
202 parameters. Non-Photochemical Quenching (NPQ), a mechanism for dissipating excess light
203 energy (Müller et al., 2001), increased in *A. thaliana* under salt stress suggesting that less
204 energy is being utilized for productive purposes (Figure S9E). On the other hand, salt stress had

205 no effect on NPQ in *S. parvula*. The resilience of *S. parvula* photochemistry under salt stress is
206 consistent with this species' ability to maintain CO_2 assimilation in saline conditions (Figure 5D).
207 Taken together, the phenomics data confirm similar growth and physiological tolerance of *S.*
208 *parvula* to salt stress regardless of growth medium.

209 Minimizing non-stomatal transpiration by enhanced epidermal boundary layer
210 resistance is equally important as stomatal regulation in preventing water loss under water
211 stress (Blum, 2009). Indeed, previous findings have demonstrated that *S. parvula* leaves possess
212 a significantly thicker leaf cuticle supported by a higher level of total wax content compared to
213 *A. thaliana* leaves (Teusink et al., 2002) (Figure S10A). Interestingly, when we compared the
214 basal expression of key genes in the wax biosynthesis pathway (Bernard and Joubès, 2013) from
215 comparable shoot tissues grown under control conditions, we observed consistently higher
216 constitutive expression in *S. parvula* compared to expression of their *A. thaliana* orthologs
217 (Figure S10B).

218 Changes to leaf shape and arrangement is not a salt-dependent, inducible trait in *S.*
219 *parvula*. However, its narrow leaves with elongated petioles arranged spirally on erect stems
220 with elongated internodes (Figure S11A) are ideally suited to rapid growth in saline habitats
221 with warm temperatures where efficient transpirational cooling imparts selective advantages
222 (Lin et al., 2017). Additionally, an erect growth habit is more desirable than a rosette to
223 minimize leaf contact with soil, given that *S. parvula* is found near saline lakes with topsoils
224 often enriched in salt crystals (Hajiboland et al., 2018; Tug et al., 2019). Key traits resulting in
225 narrow leaves, elongated petioles and internodes are influenced by the phytochrome family of
226 genes (Somers et al., 1991; Li et al., 2011), and we noted that *S. parvula* plants closely resemble
227 the morphology of *A. thaliana phyB phyD* double mutants (Devlin et al., 1996; Li et al., 2011).
228 This morphological similarity led us to search for genome-level cues for loss-of-function or
229 altered function in *S. parvula PHY* genes that may support such a phenotype being selected as
230 the only documented growth form of *S. parvula* (Figure 1 and Table 1) (German and Al-Shehbaz,
231 2010). We observed that *PHYB* was conserved as a single copy gene in six closely-related
232 Brassicaceae genomes that we examined, with a translocation event between lineage I and II

233 species. However, *PHYD* appeared to have undergone a gene loss specifically in *S. parvula*
234 (Figure S11B).

235

236 **Salt stress induces early flowering and silique formation in *Schrenkiella parvula***

237 We examined long-term salt-stress effects on reproductive traits to investigate how
238 excess Na^+ affected *S. parvula* fitness (Figure 6). Salt treatment induced early flowering
239 irrespective of whether flowering time was measured as the number of days after planting to
240 detection of the first floral buds or the number of leaves developed before flowering (Figure 6A
241 to C). Additionally, early flowering induced by salt was observed under both long-day and short-
242 day photoperiods (Figure 6C). The initial ~10 flowers produced under control conditions
243 aborted without developing into siliques, while these flowers produced mature siliques in salt-
244 treated plants (Figure 6A and B). *Schrenkiella parvula* flowers are self-fertilizing, a process
245 facilitated by the elongation of filaments to the level of the stigma. However, early flowers
246 under control conditions developed shorter filaments than in the salt-treated plants (Figure
247 6D). In control plants, these shorter filaments appeared to hinder successful fertilization and
248 delayed subsequent silique formation (Figure 6A and E). Notably, the salt-induced early
249 flowering and salt-dependent elongation of filaments were not specific to NaCl treatment and
250 were also initiated by KCl at similar concentrations (Figure S12). At later reproductive stages,
251 the continuous growth of multiple floral meristems led to fertile flowers under control
252 conditions. Despite the early success of silique formation in salt-treated plants, the cumulative
253 number of flowers and siliques produced by control plants surpassed that of salt-treated plants
254 if allowed to grow past two months. For example, at 74 days after planting, the control plants
255 had a greater number of flowers per plant than salt-treated plants (Figure 6E). The experiment
256 was discontinued when salt-treated plants stopped producing new inflorescences and were
257 senescing (approximately 74 days after planting). Therefore, *S. parvula* does not require salt to
258 increase fitness, and salt treatments appear to only provide a transient advantage during early
259 reproductive stages. Even if fitness is higher for control plants compared to salt-treated plants
260 when evaluated based solely on the total number of siliques produced per plant within a
261 longest possible growing season, there may be additional benefits when seeds develop on

262 plants exposed to long-term high salinity, especially if seasonal changes in its natural habitats
263 reward a strategy of salt-accelerated reproduction. The average size of siliques and seeds of
264 salt-treated plants were significantly greater than in control plants (Figure 6G, H, and I). This
265 suggested that more resources were available for seeds when produced by salt-treated plants
266 compared to control plants. The number of seeds per silique was not different between control
267 and treated plants and the seeds from both groups were indistinguishable in the rate of
268 germination in the subsequent generation when germinated on media without added NaCl.
269
270 ***Schrenkiella parvula* seed germination is delayed by high salinity and is sensitive to specific**
271 **salts**
272 Many plants including salt-adapted plants delay germination in saline media even if
273 their seedling stages can tolerate high salinity (Ungar, 1978; Kazachkova et al., 2016). Both *S.*
274 *parvula* and *A. thaliana* exhibited decreased seed germination as NaCl concentrations increased
275 compared to control conditions (Figure 7A). However, unlike the more salt-tolerant traits *S.*
276 *parvula* displayed compared to *A. thaliana* during seedling and mature developmental stages
277 (Figures 2, 4, and S9), *S. parvula* seed germination was more sensitive to NaCl than *A. thaliana*
278 seed germination (Figure 7A). When we sowed *S. parvula* seeds directly onto salt plates prior to
279 stratification, we observed more severe salt-mediated inhibition of germination. This was
280 demonstrated via detailed germination curves that revealed a progressive reduction in both
281 germination rate and final percent germination with increasing NaCl concentrations (Figure 7B).
282 This salt-influenced trait did not lead to a binary outcome between germinated vs
283 ungerminated seeds. The smaller fraction of *S. parvula* seeds that formed a radicle compared to
284 *A. thaliana* continued to develop in a high salinity medium similarly to those seeds germinated
285 under control conditions (Figure S13A). In contrast, *A. thaliana* seeds that formed radicles in a
286 high saline medium did not survive to the seedling stage. Up to 98% of *S. parvula* seeds that
287 failed to germinate (i.e. no visible radicle emergence) on high-salt growth medium germinated
288 and developed into seedlings when transferred to control medium without added NaCl (Figures
289 7C and S13B). Moreover, seeds that were incubated for a month on saline medium still
290 exhibited 98% germination (Figure 7C). Interestingly, adding salt and increasing the duration of

291 salt-treatment, led to faster germination when salt-inhibited seeds were transferred back to the
292 control medium. These results suggest that NaCl introduces a reversible barrier to *S. parvula*
293 seed germination.

294 We also tested whether other salts, including K, Li, and B, found in the native soils of *S.*
295 *parvula* ecotype Lake Tuz caused a reversible inhibition of *S. parvula* seed germination. KCl
296 induced a similar inhibitory response as observed for NaCl (Figures 7D and S13), but neither LiCl
297 nor H_3BO_3 led to inhibition of *S. parvula* seed germination (Figure 7E and F). In contrast, *A.*
298 *thaliana* seed germination was highly sensitive to high LiCl and H_3BO_3, and KCl at 150 or 250
299 mM was lethal to *A. thaliana* seeds that formed a radicle (Figures 7D, E, F, and S13A). These
300 data suggest that *S. parvula* is adapted to sense the soil salt level as well as the salt type before
301 radicle emergence, and if seeds germinate, they are more likely to continue development
302 despite toxic levels of salts in the growth medium that are lethal to *A. thaliana*.

303 Salt-dependent germination of *S. parvula* suggests that it is an adaptive trait to time
304 seed germination for when topsoil salinity decreases during the summer months from June to
305 September. This finding was supported by field observations made in the Lake Tuz region where
306 *S. parvula* grows as a seasonal annual during the summer months that receive the highest
307 rainfall and temperature with the longest day length (Figures S14A to C). The saline soils
308 surrounding Lake Tuz are often covered with a salt crust towards fall, which becomes a
309 dominant feature in the landscape extending to spring of the following year despite having a
310 high saline water table throughout the year (Tug et al., 2019). Seed germination may range
311 widely within the expected growing season and seedlings are more vulnerable to extreme
312 environmental conditions than mature plants. Finally, we compared *S. parvula* and *A. thaliana*
313 seedlings for their tolerance to heat, chilling, and freezing stresses to assess how *S. parvula* may
314 fit within the expected temperature tolerance range required to survive in its native habitat
315 (Figures S14D to F). *Schrenkiella parvula* seedlings displayed higher tolerance to heat stress at
316 38 °C than *A. thaliana* (Figure S14F), while chilling and freezing tolerance in both species was
317 indistinguishable (Figure S14D and E).
318

319 **Discussion**

320 Plants are known for their remarkable capacity to tolerate environmental stresses and
321 ability to modulate growth even to the extent of pausing growth entirely. This plasticity in
322 response to various environmental stresses is observed in all plants, but as shown in the current
323 study, extremophytes such as *S. parvula* exhibit a much higher tolerance to stress than
324 mesophytes, which include most crops and *A. thaliana* (Flowers and Colmer, 2008; Kazachkova
325 et al., 2018). Halophytes represent only about 0.4% of all flowering plants and about 40% of all
326 halophytes including *S. parvula* can withstand salt stress at concentrations similar to seawater
327 (Kotula et al., 2020). At high salinities, most plants respond to salt stress by inhibiting growth to
328 prioritize survival as a tradeoff (Santiago-Rosario et al., 2021). Nevertheless, the extremophyte
329 model *S. parvula* provides a genetic system to discover adaptive traits that do not show a
330 growth compromise at salt concentrations that are known to be lethal to most crops and *A.*
331 *thaliana* (Oh et al., 2014). The defining traits of *S. parvula* as an extremophyte should reflect
332 the environmental constraints that have shaped its phenotype and collectively made it distinct
333 from other more stress-sensitive annuals in Brassicaceae, including *A. thaliana*. When we
334 summarized *S. parvula* physiological and structural traits that changed at a developmental
335 stage-specific and tissue-specific scale in response to salt stress, several traits emerged as
336 significant features induced under salt stress compared to their non-stressed control group
337 (Figure 8A). Among them, the higher number of siliques produced during salt-induced early
338 flowering, expansion of xylem vessel elements, and increased leaf thickness provide the
339 greatest contribution to the total plastic or inducible trait space (Figure 8B and C).
340

341 **Salt stress accelerates reproduction thereby contributing to maximizing fitness of *S. parvula***

342 Salinity delays flowering in most plants (Blits and Gallagher, 1991; Lutts et al., 1995;
343 Apse et al., 1999; Moriuchi et al., 2016; Cho et al., 2017). For example, high salinity (≥100 mM
344 NaCl) delays or inhibits the transition from vegetative to reproductive growth in *A. thaliana*
345 (Achard et al., 2006; Li et al., 2007; Ryu et al., 2014). Delayed flowering ensures survival under
346 salt stress supported by multiple genetic mechanisms. These pathways include transcription
347 factors that inhibit flowering and are also induced by salt (Cho et al., 2017). However, a few
348 plants possess alternative strategies whereby flowering is accelerated in response to salt stress

349 (Adams et al., 1998; Ventura et al., 2014). Additionally, molecular pathways operating
350 antagonistically to salt-induced delayed flowering have been described for *A. thaliana* (Yu et al.,
351 2018). This apparent strategy would facilitate reproduction and improve fitness by allowing
352 escape from harsh environments, especially for ruderal or annual plants growing in habitats
353 that become warmer, drier, and more saline (due to increased surface evaporation) towards
354 the end of their growing season. Salt-induced flowering observed for *S. parvula* suggests that
355 such an alternative strategy was selected as the dominant trait to maximize its fitness in its
356 native habitat exemplified by the Lake Tuz ecotype used in our current study (Figures 6 and
357 S12) over the more common trait of salt-induced delay in flowering observed for many other
358 plants (Apse et al., 1999; Cho et al., 2017).

359 Compared to favorable growth conditions, environmental stresses increase the
360 opportunity for selection. Previous studies have demonstrated that phenotypic selection favors
361 stress-avoidance traits, including earlier flowering, in addition to stress-tolerance traits (Eshel et
362 al., 2021). This results in an evolutionary shift toward earlier flowering despite the prevalent
363 trait observed for model and crop plants to delay flowering when exposed to salt (Stanton et
364 al., 2000; Ventura et al., 2014; Caño et al., 2016; Moriuchi et al., 2016). Stanton et al., (2000)
365 further showed that, among multiple environmental stresses tested for their effects on
366 evolutionary selection in wild mustard, *Sinapis arvensis* (a ruderal annual in Brassicaceae), the
367 potential for selection was greatest for high salt stress and low light. *Schrenkiella parvula* is
368 native to the variable shoreline habitats of saline lakes of the Irano-Turanian region where it is
369 often a ruderal plant growing in frequently inundated saline soils (Ozfidan-Konakci et al., 2015;
370 Hajiboland et al., 2018; Tug et al., 2019). Therefore, salt stress is the norm for *S. parvula* and
371 over a multi-generational time scale, salt stress is expected to provide a constant selection
372 pressure.

373 When we examined salt-responsive traits throughout the lifecycle of *S. parvula*, an
374 increased number of siliques produced due to salt-induced early flowering had the largest fold-
375 change among other salt-induced traits that differentiated the response between control and
376 salt-treated growth (Figure 8C). Early flowering established an earlier seed set and the

377 opportunity to produce viable seeds at an earlier developmental age in the salt treated plants
378 compared to control plants (Figure 6). However, this was a plastic trait not necessarily
379 imparting higher fitness when assessed based on overall fecundity during a longer growth
380 period . When control and salt-treated plants were grown for over 2 months, the control plants
381 displayed longer vegetative growth phases subsequently leading to higher fecundity in *S.*
382 *parvula.* These results suggest that salt-induced flowering has evolved as a mechanism of stress
383 avoidance rather than stress tolerance in a habitat where the severity of environmental
384 stresses increase as the growing season continues (Figure S14).

385 A key morphological feature contributing to the transient advantage of salt-mediated
386 early flowering in *S. parvula,* was the plastic response observed for filament elongation that
387 was enhanced by salt (Figures 6D and S12). Filament elongation is an important trait for
388 successful fertilization in self-pollinating plants such as *S. parvula* and *A. thaliana*. However,
389 whereas salt stress terminally inhibits *A. thaliana* gamete formation and leads to abortion of
390 flowers and seeds (Sun et al., 2005; Figure 6), filament elongation and production of viable
391 seeds were not inhibited by salt in *S. parvula*. This finding indicates that *S. parvula* has evolved
392 to decouple the regulatory pathways that sense salt and inhibit stamen growth.
393

394 **Salt stress-induced xylem vessel expansion across the root-shoot continuum could contribute**
395 **to the salt resilient growth of *Schrenkiella parvula***

396 Selection on vessel diameter favors narrower vessels to minimize the risk of cavitation
397 independently of ancestry or habitat for a wide range of angiosperms from rainforests to
398 deserts (Olson and Rosell, 2013). Furthermore, the effect of salt stress on xylem development
399 within a species often leads to narrower vessels (Junghans et al., 2006; Ge et al., 2017; Cruz et
400 al., 2019; Sarker and Oba, 2020). Our results suggest an alternative strategy for *S. parvula*
401 where we observed an increase in both shoot and root xylem vessel area in plants exposed to
402 long-term salt stress (given via 150 mM NaCl) (Figures 3 and 4) (Li et al., 2021). The salt-
403 mediated expansion of xylem tissue in *S. parvula* was the second-most highly induced trait that
404 responded by a higher fold change between control and salt-treated plants (Figure 8). This
405 adaptive feature likely allows a higher bulk flow through the xylem to help *S. parvula* leaves

406 maintain a cooler temperature while allowing uncompromised gas exchange compared to *A.*
407 *thaliana* grown under saline conditions (Figure 5), despite the tradeoff of a higher risk of
408 cavitation. Salt-induced increases to xylem diameter correlated to maintaining growth under
409 salt stress has been reported for a few extremophytes (e.g. *Nitraria retusa* and *Atriplex halimus*)
410 adapted to extreme salinities above seawater strength, but is not a common trait associated
411 with halophytes (Boughalleb et al., 2009; Parida et al., 2016). The expansion of xylem area
412 without the expansion of root or stem area that we observed as a salt responsive trait in *S.*
413 *parvula* (Figure 3) is an atypical adaptation among plants resilient to salt stress (Olson and
414 Rosell, 2013; Nassar et al., 2020). *S. parvula* seems to optimize effective use of water instead of
415 simply reducing water loss at high salinities (Figures 5F and S9). Prioritizing effective water use
416 has been proposed to be a better strategy for plant growth during water deficit stress than
417 maximizing water conservation at the expense of photosynthetic capacity (Blum, 2009). This
418 finding has implications for agriculture. If the availability of water can be ensured in agricultural
419 systems even if the water source is brackish, the crops that are able to maintain relative water
420 content (as observed for *S. parvula* in Figure 5), while allowing uncompromised transpiration
421 and gas exchange, will be more resilient to environmental stresses.
422
423 **Adjustments to shoot architecture modulation follows root responses to cope with salt stress**
424 *Schrenkiella parvula* root growth and development was less affected by increasing salt
425 concentrations than in *A. thaliana*. *Schrenkiella parvula* primary root length increased while
426 root fresh weight was only maintained under high salt conditions (Figures 2, S1 and S3),
427 suggesting that salt causes a reallocation of resources toward deeper roots. This is also
428 supported by the reduction in root hair length, total lateral root length and number (Figures 2
429 and S1). Given the natural environment in which the *S. parvula* ecotype Lake Tuz grows, the top
430 layers of soil will dry first, causing the soil salt concentration to form a high to low gradient from
431 topsoil to deeper layers in the rhizosphere (Tug et al., 2019). Thus, *S. parvula* can avoid
432 relatively higher salinities by growing deeper to lower salt concentrations. The growth strategy
433 exemplified by *S. parvula* is correlated with uncompromised growth dependent on modulating
434 root to shoot vasculature and seemed to be efficiently coupled with leaf traits in *S. parvula*.

435 Even though saline soils directly affect roots, its effects are also observed in shoots. If roots
436 grow continuously in saline media, salts are often deposited in leaves. As plants age, salt-
437 adapted plants either need to store salts in leaves and shed those whose capacity to hold
438 excess salt is reached, or extrude excess salt via salt glands (Jennings, 1968; Dassanayake and
439 Larkin, 2017). *Schrenkiella parvula* uses the first strategy by developing succulent leaves that
440 are equipped with larger vacuoles which support salt sequestration. This trait appears to be
441 facilitated by shifting the leaf cell population from a dominant diploid state to higher ploidy
442 levels during prolonged salt stress (Figure 4). Indeed, the increase in leaf thickness, along with
443 larger leaf area, was among the top traits with highest salt-induced fold-changes in *S. parvula*
444 (Figure 8). Developing larger cells that can store excess salts within larger vacuoles enabled by
445 endoreduplication has been reported for several other salt adapted extremophytes. It was
446 proposed to be a key mechanism for surviving environmental stress (De Rocher et al., 1990;
447 Barkla et al., 2018), while leaf succulence is one of the most common traits observed in
448 halophytes (Jennings, 1968; Flowers and Colmer, 2008).
449 To maximize storage capacity via increasing the leaf surface area to volume, leaves are
450 generally more terete in many extremophytes that have fully developed succulent leaves. This
451 trait is often accompanied by the loss of leaf abaxial identity whereby the leaves become
452 adaxialized (Ogburn and Edwards, 2013). Multiple genetic networks regulated by adaxially
453 expressed HD-ZIPIII transcription factors have been shown to control leaf polarity (Du et al.,
454 2018). *Schrenkiella parvula* leaves tend to acquire a more terete state with prolonged exposure
455 to salt (Figure S7B). This phenotypic change is coincident with *S. parvula* leaves exhibiting
456 reduced differentiation between adaxial and abaxial surfaces as leaf thickness increases with
457 higher salt concentrations and a clear palisade layer at the adaxial surface is not present.
458 However, this structural alteration does not affect the relative water content nor cause leaf
459 curling or other visibly malformed vascular structures, as seen in adaxialized leaves of *A.*
460 *thaliana* mutants (Figures 4, 5, and S7) (Du et al., 2018).
461 *Schrenkiella parvula* leaves are amphistomatous (possess stomata on the upper and
462 lower leaf surfaces), in contrast to the *A. thaliana* leaves, which have more stomata on the
463 abaxial surface (hyposomatous) (Figure 5B). Amphistomatous leaves are relatively rare in

464 angiosperms (Nadeau and Sack, 2002; Drake et al., 2019) but in such leaves, the distance
465 between stomata and the mesophyll is reduced allowing greater water-use efficiency due to
466 increased CO_2 conductance in the mesophyll thereby facilitating a higher relative
467 photosynthesis rate (de Boer et al., 2016). Therefore, amphistomatous leaves are found in
468 highly productive fast-growing herbs. However, when this trait is found in extremophytes such
469 as *S. parvula* with trichomeless leaves arranged mostly with vertical leaf angles (Fig 1), it adds a
470 tradeoff for water loss through transpiration as well as higher exposure to solar heat (Drake et
471 al., 2019). Drake et al. (2019) hypothesized that to avoid desiccation in thick amphistomatous
472 leaves, vein length per unit leaf area must be larger than in hypostomatous leaves of the same
473 thickness. This anatomy would need to be accompanied by additional traits to facilitate an
474 increase in transpirational flow via xylem development from root to shoot if leaf thickness
475 increases as a response to salt stress. Additionally, having mostly vertical leaf angles with thick
476 amphistomatous leaves is considered a key adaptive trait to reduce the effect of thermal
477 radiation around midday while maximizing photosynthesis in the morning and late afternoon
478 thereby lowering the risks of desiccation (King, 1997). Further, traits known to have evolved
479 under shade avoidance such as internode elongation seems to be exapted in *S. parvula*, most
480 likely to enhance transpirational cooling selected under environments that demand rapid
481 growth amidst extreme environmental constraints (Pierik and Testerink, 2014).
482 Moderate to high salt is a persistent edaphic factor in the natural habitat of *S. parvula*
483 (Helvaci et al., 2004; Tug et al., 2019). Such an environment would have selected for multiple
484 traits that act synergistically to create an efficient lifestyle to survive stress. However, unlike
485 many slow-growing extremophytes, *S. parvula* shows fast growth achieved with short life
486 cycles, while enduring multiple environmental stresses. Often environmental stress resilience
487 comes at the cost of yield reduction in crops and the drive for increasing yields has been
488 prioritized over the need to develop resilient crops (Pardo and VanBuren, 2021). Increasing
489 threats to global agriculture due to climate change necessitate a change in our priorities for
490 crop development (IPCC, 2021). *Schrenkiella parvula* provides an excellent genetic model
491 system that illustrates growth optimization over growth inhibition to cope with environmental

492 stresses that we can explore to find innovative genetic architectures suitable and transferable
493 to develop resilient crops.
494
495 **Materials and Methods**
496 **Plant growth conditions**
497 *Schrenkiella parvula* (Lake Tuz ecotype) and *A. thaliana* (Col-0 ecotype) were grown on
498 soil, plates or in a hydroponic system as described in Wang et al., (2019), Pantha et al.,(2021),
499 and Conn et al., (2013). Soil grown plants were used for reproductive trait assessments. Plate
500 grown plants were used for root growth assessments and thermal tolerance assays.
501 Hydroponically grown plants were used for shoot physiological assessments and anatomical
502 characterization. See supplementary method S1 for details.
503 **Root growth assays**
504 Five-day-old seedlings germinated on plates were transferred to 1/4x MS supplemented
505 with 100 and 150 mM NaCl and kept at a photoperiod of 12 hr light/12 hr dark for 13 days.
506 Additionally, 4-day-old seedlings of *A. thaliana* and *S. parvula* grown at a photoperiod of 16 hr
507 light/8hr dark and on 1/2x MS agar plates were transferred to 1/2x MS plates supplemented
508 with 125, 175, and 225 mM NaCl and monitored for 6 days. These plates were imaged for
509 further processing using ImageJ (Ferreira and Rasband, 2012) to quantify primary root length,
510 lateral root number and density, total and average lateral root length, and root hair length
511 (Figures 2 and S2). See supplementary method S1 for details.
512 **Halotropism assay**
513 Seedlings were grown on 1/2x MS medium solidified with 1.2% (w/v) Phyto agar plates
514 held at an 85° incline as described in Galvan-Ampudia et al., (2013). After 5 days, the agar
515 medium 1 cm below the root tips, was removed and replaced by 1/2x MS agar supplemented
516 with 200 mM NaCl and seedlings were allowed to grow for another 5 days (Figure S4).
517 **Germination assay**
518 Sterilized stratified seeds were germinated on 1/4x MS (Figures 7A, D, E and F) or 1/2x
519 MS (Figures 7B and C) supplemented with different concentrations of NaCl, KCl, LiCl, or H_3BO_3.
520 Final percent germination was recorded 3 days and germination curves recorded for up to 12

521 days after stratification. Seeds were counted as germinated if radicle emergence was observed.
522 To check the viability of ungerminated seeds, *S. parvula* seeds that failed to germinate on high
523 NaCl or KCl plates were transferred back to 1/4x MS or 1/2x MS and monitored for up to two
524 weeks.

525 **Thermal stress tolerance assays**

526 Five-day-old seedlings were subjected to heat stress at 38 °C for 6, 18 and 24 hr; chilling
527 stress at 4 °C for 24 hr; or freezing stress at 0 °C for 12 hr followed by a recovery time of 5 days
528 at 23 °C. Seedlings were considered to have survived if roots continued to grow without visible
529 chlorosis of the cotyledons after the recovery period monitored for another 5 days.

530 **Quantification of leaf traits**

531 Stomatal conductance and CO_2 assimilation rate were measured using the sixth or
532 seventh leaf from the shoot tip of hydroponically grown plants in LiCor 6400-40 leaf chamber
533 attached to a 6400XT gas analyzer (LiCor Inc, Lincoln Nebraska). Measurements were taken
534 after acclimation to 400 µmol photons m^{-2} s^{-1}, 400 µL L^{-1} CO_2 at 23 °C and an air flow rate of 200
535 µmol s^{-1} until a steady-state photosynthesis rate was attained. Each leaf was measured three
536 times and three plants were used for each treatment.

537 Four-week-old hydroponically grown plants subjected to 150 mM NaCl or control
538 treatments were used for the following leaf measurements. To quantify total leaf area, all
539 leaves from selected plants were scanned (Perfection V600 scanner, Epson, Suwa, Nagano,
540 Japan) and analyzed with ImageJ (Ferreira and Rasband, 2012). To obtain the relative water
541 content (RWC), newly harvested leaves were weighed before and after submerging in water for
542 24 hr (to get fresh and turgid weights respectively) followed by drying until dry weights were
543 obtained. RWC was calculated as (fresh weight - dry weight)/(turgid weight - dry weight). Leaf
544 temperature was measured on intact leaves of 4-week-old plants between ZT3 to ZT4
545 (Zeitgeber Time) every day for 7 days using thermal imaging (E6, FLIR, Wilsonville, Oregon). Leaf
546 temperatures were averaged across all leaves for each plant.

547 **High-throughput phenotyping of morphometric and physiological traits**

548 *A. thaliana* and *S. parvula* soil grown plants were phenotyped for compactness, leaf
549 water status, leaf area, and chlorophyll fluorescence using a PSI PlantScreen™ Compact system
550 (Qubit Phenomics, PSI). See supplementary method S1 for details.

551 **Anatomical and flow cytometry analyses**

552 Sections obtained from young roots (1 cm from the tip), mature roots (2 to 5 cm from
553 the root and shoot junction), stems (fourth, fifth, and sixth internodes from the shoot tip) and
554 leaves (fourth, fifth, and sixth from the shoot tip) from 8-week-old plants were used for
555 anatomical trait characterization. The fifth and sixth leaves from the shoot tip were used for
556 flow cytometry analysis as described in Galbraith et al., (1983). We counted the number of
557 stomata stained with propidium iodine on both hydroponically- and soil-grown plants and on
558 adaxial and abaxial surfaces at the same developmental stage and treatment duration. See
559 supplemental method S1 for details.

560 **Reproductive trait quantification**

561 Three-week-old soil grown plants were treated with an incremental increase of 50 mM
562 NaCl every 2 days until the final concentration was reached to 150 mM NaCl treatments and
563 maintained at 150 mM NaCl for the remainder of the experiment under either a long-day (16 hr
564 light/ 8 hr dark) or short-day (12 hr light/12 hr dark) photoperiod. Flowering time was recorded
565 when the first open flower was observed. Total numbers of flowering events, including all
566 developing, mature, and aborted flowers, as well as fully developed siliques were counted
567 separately for each plant throughout its lifetime. Fully developed siliques were counted and
568 harvested for imaging from 5-week-old plants that were given 150 mM NaCl or control
569 treatments for an additional 2 weeks.

570 **Computational analysis**

571 The data used to compare the basal expression of wax biosynthesizing genes between
572 *A. thaliana* and *S. parvula* were retrieved from NCBI-SRA database, accession# SRX877979 and
573 SRX877980, respectively (Oh et al., 2014). The expression profiles were visualized using
574 Integrated Genomics Viewer (v2.6) (Thorvaldsdóttir et al., 2013). The copy numbers and
575 transposition states of PhyB/D orthologs were determined using CLfinder-OrthNet pipeline as
576 described by Oh and Dassanayake, (2019). For principal component analysis (PCA) given in Fig 8,

577 as traits were measured in different units and scales, the measurements for each trait were first
578 normalized by either subtracting the mean (for anatomical traits) or dividing by the mean (for
579 physiological traits). The normalized data were subsequently analyzed with prcomp in R.
580
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600
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602 data as major contributors to the overall project; C.W., D-H.O., N.D., J.H., H.L., H.H., and P.A.
603 conducted additional experiments and analyzed data; undergraduate students J.C.J., R.K.,
604 M.G.M., A.C., D.T., and high school student C.C. conducted complementary experiments
605 supervised by K.T., P.P., G.W., D-H.O. and C.W.. M.D., C.T., and S.B. designed independent

606 complementary experiments; supervised students in their labs; and contributed to data
607 interpretation. D.L., Y.Z., and G.S. provided additional student supervision and assisted with
608 data generation for selected experiments. K.T., P.P., G.W., S.B., and M.D. wrote the manuscript.
609 K.T., P.P., G.W., N.K., D-H.O., C.W., P.A., M.F., J.C.L., A.S., D.L., P.F., C.T., S.B., and M.D. provided
610 critical reviews. M.D. conceptualized and supervised the overall project.
611
612 **Main figure and table captions**
613 **Figure 1.** Life cycle of *Schrenkiella parvula* from seeds to siliques. Scale bars are 1 cm unless
614 indicated in the figure.
615 **Table 1**. Morphological comparison between *Arabidopsis thaliana* and *Schrenkiella parvula*.
616 **Figure 2**. Effects of NaCl stress on root growth in *Schrenkiella parvula* and *Arabidopsis thaliana*
617 seedlings. [A] 12-day-old seedlings of *A. thaliana* and *S. parvula* were grown for 5 days on 1/4x
618 Murashige and Skoog media, and 7 days on the indicated concentrations of NaCl. Plates were
619 scanned 7 days after treatment (DAT). [B] Primary root growth and average root growth rate,
620 [C] number of lateral roots and lateral root density, and [D] average length of 10 longest root
621 hairs. Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and their
622 respective control group, determined by Student's *t*-test. Data are mean ± SD (n =3, at least 3
623 plants per replicate). Open circles are individual measurements.
624 **Figure 3**. Effects of NaCl stress on *Schrenkiella parvula* root anatomy. Eight-week-old *S. parvula*
625 plants were grown under control or 150 mM NaCl conditions for an additional 4 weeks. [A] The
626 position of transverse sections of *S. parvula* roots, [B] the root tip, [C] the young root, and [D]
627 the mature root from 12-week-old hydroponically-grown *S. parvula* plants. A minimum of 20
628 sections from 4 to 13 plants were examined for each root region to quantify each parameter for
629 each condition. Asterisks indicate significant differences ($p \leq 0.05$) between the treated samples
630 and their respective control group, determined by Student's *t*-test. Data are means ± SD. Data
631 points represent individual cross-sections and colors represent individual plants. Representative
632 cross-sections were obtained from the control plants. Scale bar represents 100 µm.
633 **Figure 4**. Effects of NaCl stress on *Schrenkiella parvula* shoot anatomy. Eight-week-old *S.*
634 *parvula* plants were grown under control or 150 mM NaCl conditions for an additional 4 weeks.

635 Transverse section of [A] stem and [B] leaf of 12-week-old hydroponically-grown *S. parvula*
636 plants. A total of 23 sections from 7 control plants and 32 sections from 9 treated plants were
637 used for leaf measurements, and a minimum of 20 sections from 5 to 11 plants were used for
638 stem measurements. Data points represent individual cross-sections and colors represent
639 individual plants. [C] Nuclear DNA content, distribution of leaf cell ploidy, and
640 endoreduplication index of leaf cells from 10th and 11th leaves from the shoot meristem in
641 control and 250 mM NaCl-treated 8-week-old *S. parvula* plants. Data are mean ± SD (n= 4).
642 Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and their
643 respective control group, determined by Student's *t*-test. Representative cross-sections were
644 obtained from the control plants Scale bars represent 100 µm.
645 **Figure 5.** Effects of NaCl stress on *Schrenkiella parvula* leaf traits. All experiments were
646 performed with 4-week-old hydroponically-grown plants that were treated for an additional 4
647 weeks with the indicated NaCl concentrations. [A] Total leaf area, [B] Stomatal density, [C]
648 Stomatal conductance, [D] Photosynthesis rate, [E] Leaf relative surface temperature, and [F]
649 Leaf relative water content. Asterisks indicate significant difference ($p \leq 0.05$) between the
650 treated samples and their respective control samples, determined by Student's *t*-test. Data are
651 means ± SD (n ≥ 3). Open circles indicate the measurement from each plant (in B and F) and
652 leaves (in C and D) for each experiment. DAT, days after treatment.
653 **Figure 6.** Effects of NaCl on reproductive traits of *Schrenkiella parvula*. [A] Plants grown under
654 control conditions generally flower at the ~17th leaf stage and the first few flowers are
655 subsequently aborted. [B] Salt treated plants flower earlier at the ~14th leaf stage and the first
656 flowers develop into mature siliques. [C] Days from planting to the first observed flower of
657 plants grown under a long day (16 hr light/8 hr dark) or short day (12 hr light/12 hr dark)
658 photoperiod with and without the indicated salt treatment. Center line - median; box -
659 interquartile range (IQR); notch - 1.58 × IQR/sqrt(n); whiskers - 1.5 × IQR. [D] Flowers obtained
660 from control and treated plants described in panel A and B. [E] Number of flowers and siliques
661 per plant under control and salt treatment. Data are mean ± SD. Open circles represent
662 individual measurements from four biological replicates. [F] Ratio between the number of
663 flowers and siliques observed for control (C) and salt-treated (T) plants described in E. [G]

664 Siliques from plants under control and NaCl treated conditions. [H] Size of siliques from G. Open
665 circles represent individual silique counts. [I] Comparison of the area of seeds from plants under
666 control and NaCl treated conditions. Data are mean ± SD (n = 50). Seeds per condition were
667 selected from the seed pool of 15 to 50 plants. Open circles represent individual seeds. For
668 panel [B], salt treatment started 21 DAP, initially at 50 mM NaCl given every other day. The salt
669 concentration was increased by 50 mM every four days until it reached 200 mM; for panels [C,
670 E, F], salt treatment was applied to 3-week-old plants until the end of the experiment; for
671 panels [G, H, I], salt treatment was applied to 5-week-old plants for an additional 2 weeks.
672 Different letters or asterisks represent significant differences (*, $p < 0.05$) compared to control,
673 determined by either one-way ANOVA followed by Tukey's post-hoc test [C] or Student's *t*-test
674 [E, H, I]. DAP, days after planting.
675 **Figure 7.** Effects of salt stress on the germination of *Arabidopsis thaliana* and *Schrenkiella*
676 *parvula* seeds. [A] Final percent germination recorded at 3 days and [B] germination curves of
677 seeds treated with different concentrations of NaCl recorded for 12 days after stratification. [C]
678 Germination curves of NaCl-treated seeds transferred to control (no NaCl) media, after 2 days
679 or 30 days on salt plates. The germination curves were performed by direct sowing of seeds
680 onto salt plates prior to stratification. Black arrows point to the day when seeds were
681 transferred to non-NaCl plates. [D, E, F] Final percent germination of seeds treated with KCl [D],
682 LiCl [E] or H_3BO_3 [F]. Data are mean ± SD (n = 3-4, each replicate contains ca. 50 seeds). Open
683 circles indicate individual measurements. Different letters indicate significant difference ($p \leq$
684 0.05) determined by one-way ANOVA with post-hoc Tukey's test across all conditions in [A, D, E,
685 F], or across treatment groups at 3 and 12, and 7 days after stratification in [B] and [C]
686 respectively.
687 **Figure 8**. Summary of salt-induced structural and physiological traits in *Schrenkiella parvula*. [**A**]
688 Salt- induced changes in traits quantified in the current study. Changes in each trait were
689 calculated as the fold change between treatment and control measurements. Traits highlighted
690 in the red boxes showed significant differences ($p < 0.05$) under salt treatments compared to
691 the control determined in previous assays (Fig 2-7). [**B**] and [**C**] PCA biplot of traits quantified
692 for *S. parvula* under control and salt-treated conditions. Arrows indicate directions of loadings

693 for each trait and are color-coded by contribution to the variations in PC1 and PC2. [B]
694 Anatomical traits quantified: LT, leaf thickness; MT, midrib thickness; XS, stem xylem to stem
695 area; SVA, average area per vessel in stems; CbS, cambium to stem area; CtS, cortex to stem
696 area; XR, root xylem to root area; RVA, average area per vessel in roots; VN, number of vessels
697 across the root diameter; AS, air space to root area; YRA, young root area; CT, cortical
698 thickness; YRD, root diameter in young roots; DRH, distance from root tip to first root hair. [C]
699 Physiological traits quantified: AbS, abaxial stomatal density; adS, Adaxial stomatal density; LA,
700 leaf area; SLA, silique area; SA, seed area; PRL, primary root length; LRD, lateral root density;
701 RWC, relative water content; CA, CO2 assimilation; SC, stomatal conductance; RHL, root hair
702 length; NF, number of flowers; NS, number of siliques. DAT, days after treatment; DAP, days
703 after planting.
704
705 **Supplement figure and table captions**
706 **Figure S1**. Effects of NaCl stress on root growth of *Schrenkiella parvula* and *Arabidopsis thaliana*
707 under long days and higher nutrient conditions. [A] Root growth of 10-day-old seedlings under
708 indicated concentrations of NaCl. Quantification of [B] primary root growth, [C] number of
709 lateral roots, [D] average lateral root length, [E] total lateral root length, and [F] lateral root
710 density. Asterisks indicate significant difference (p-adj ≤ 0.05) between the treated samples and
711 their respective control samples, determined by two-way ANOVA. Data are represented as
712 mean ± SD (n = 15 to 20). Open circles indicate the number of plants used for each experiment.
713 **Figure S2**. Effects of NaCl stress on root hair growth in *Schrenkiella parvula* and *Arabidopsis*
714 *thaliana*. White arrows indicate root tip positions when the 5-day-old seedlings were
715 transferred to the indicated salt concentrations. Note that the white arrows were absent in *A.*
716 *thaliana* and *S. parvula* control because the position of the root tip at the time of transferring
717 were out of frame when we imaged the root tip region. Scale bars are 0.5 mm. DAT, Days after
718 treatment.
719 **Figure S3**. Long term effects of NaCl stress on the root growth of *Schrenkiella parvula*. [A] Root
720 fresh weight and [B] primary root length of 12-week-old hydroponically grown *S. parvula* plants.
721 Four-week-old plants were subjected to 150 mM NaCl for an additional 4 weeks and the

722 treatment was continued or increased to 250 mM NaCl for another 4 weeks. The plants were
723 grown at 100-150 µmol m^{-2} s^{-1} photosynthetic photon flux density with a 12 hr light/12 hr dark
724 photoperiod. Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples
725 and their respective control samples, determined by Student's *t*-test. Data are represented as
726 the mean ± SD (n = 9). For each plot, an open circle indicates the measurement from each plant.
727 **Figure S4**. Halotropism assay at 200 mM NaCl for *Schrenkiella parvula* and *Arabidopsis thaliana*.
728 The blue lines indicate the position of seeds sown. The yellow line marks the separation
729 between 1/4x MS media with 0 mM (top) and 200 mM NaCl (bottom). The bent root angle is
730 indicated by curved white lines. g, gravity axis.
731 **Figure S5.** Root anatomy of *Schrenkiella parvula*. [A] Transverse section of the young root taken
732 at 1-3 cm from the root tip. [1] Young root area; [2] root diameter; [3] epidermis thickness; [4]
733 cortical layer thickness; [5] endodermis thickness. [B] Transverse section of the mature root
734 taken between 2-5 cm below the root-shoot junction. [1] Mature root area; [2] xylem/root
735 area; [3] Area per vessel; [4] # vessels/root diameter; [5] Air space/root area. [C] Longitudinal
736 section of young root tip from 7-day-old seedlings. The cortex layer is highlighted in yellow. The
737 red arrow indicates the additional cell layer.
738 **Figure S6.** Stem anatomy of *Schrenkiella parvula*. [A] Transverse section of *S. parvula* stem
739 (between 4th, 5th, and 6th internodes from the shoot meristem) with cell layers marked in
740 yellow lines for trait quantification. [B] Measurements of stem anatomical features indicated in
741 [A]. A minimum of 20 sections from 5-11 plants were used for each measurement. Asterisks
742 indicate significant difference ($p \leq 0.05$) between the treated samples and its respective control
743 samples, determined by Student's *t*-test. Data points represent individual cross-sections and
744 colors represent individual plants.
745 **Figure S7**. Leaf structures of *Schrenkiella parvula*. [A] Transverse section of *S. parvula* leaves. [1]
746 leaf thickness and [2] midrib thickness. [B] Transverse sections of *S. parvula* leaves treated with
747 indicated concentration of salts. All sections represent the fifth or sixth leaf from the root-shoot
748 junction in 8-week-old plants that were treated for 4 weeks.
749 **Figure S8.** Effects of salt on growth of 4-week-old *Arabidopsis thaliana* and *Schrenkiella parvula*
750 treated for an additional 2 weeks at 22 °C under a 12 hr light/12 hr dark photoperiod in soil.

751 **Figure S9**. Effects of NaCl stress on *Arabidopsis thaliana* and *Schrenkiella parvula* morphometric
752 and physiological traits. Soil-grown, 8-day-old *A. thaliana* and 12-day-old *S. parvula* were
753 exposed to the indicated salt concentrations at 21 °C, 16 h light/8 h dark photoperiod. The
754 Measurements were taken on day 8, 12, and 15 after the start of the salt treatments. [A]
755 Compactness (the ratio between the rosette (or leaf) area and the rosette (or leaf) convex hull
756 area); [B] Leaf area; [C] Leaf water content; [D] Maximum quantum efficiency of PSII; and [E]
757 Non-photochemical quenching were measured/estimated using the Qubit/PSI PlantScreen™
758 Compact System. For leaf area and leaf water content, values are percent of control (0 mM
759 NaCl) for each time point. Letters denote significant changes at p-adj ≤ 0.05 (one-way ANOVA
760 with post-hoc Tukey's test). Data are mean ± SD (n = 10). Open circles indicate the individual
761 measurements obtained from each plant. DAT, days after treatment.
762 **Figure S10.** Leaf surface and the basal expression of genes involved in wax biosynthesis in
763 *Arabidopsis thaliana* and *Schrenkiella parvula*. [A] Scanning electron micrographs contrasting *A.*
764 *thaliana* and *S. parvula* leaf surfaces. [B] Major wax biosynthesis pathway and [C] wax
765 biosynthesis genes that exhibited significantly different basal expression between *A. thaliana*
766 and *S. parvula*.
767 **Figure S11.** The potential influence of phytochrome family of genes on *Schrenkiella parvula* leaf
768 and growth form. [A] Elongated internode and leaf petiole in *S. parvula* compared to *A.*
769 *thaliana*. [B] *PHYD* is absent in the *S. parvula* genome as illustrated using OrthNet representing
770 evolutionary histories of orthologous gene groups derived from six Brassicaceae genomes:
771 *Arabidopsis lyrata* (Aly, version 1.0), *Arabidopsis thaliana* (Ath, v. 'TAIR10'), *Capsella rubella*
772 (Cru, v. 1.0), and *Eutrema salsugineum* (Esa, v. 1.0), *Sisymbrium irio* (Sir, v. 0.2), and *Schrenkiella*
773 *parvula* (v. 2.0). Nodes are color-coded according to the species. Edges show either co-linear
774 (cl) or transposed (tr) properties.
775 **Figure S12**. Effects of KCl on reproductive traits of *Schrenkiella parvula*. [A] Plants grown under
776 control conditions generally flower late and the first few flowers are subsequently aborted. KCl-
777 treated plants flower earlier and the first flowers develop into mature siliques. [B] Early flowers
778 in KCl treated plants develop longer filaments compared to control plants. DAP, days after
779 planting.

780 **Figure S13**. Effect of salt on *Arabidopsis thaliana* and *Schrenkiella parvula* inducing barriers for
781 seed germination and seedling establishment. [A] *S. parvula* (*Sp*) and *A. thaliana* (*At*) seed
782 germination and growth on NaCl- and KCl-supplemented MS medium. Scale bars are 2 cm. [B]
783 Growth of ungerminated *S. parvula* seeds treated with different concentrations of NaCl (as
784 shown in Figure 7) after transferring to control MS medium.
785 **Figure S14**. Survival of *Schrenkiella parvula* in the climate conditions found in Lake Tuz region.
786 [A] Lake Tuz location from Google Earth; [B] Precipitation and temperature recorded in Konya,
787 Turkey (NOAA, 2019). *S. parvula* growing season is from April/May to August/September (Tug
788 et al., 2019). Bars indicate precipitation and lines represent temperature. [C] Average day
789 length recorded in Konya per month. Red line indicates the day length used for typical
790 laboratory growth assays for *S. parvula*. [D-F] Survival rate of *A. thaliana* and *S. parvula*
791 seedlings under [D] freezing stress, [E] chilling stress, and [F] heat stress. Five-day-old seedlings
792 were subjected to temperature treatments and data were recorded 5 days after recovery.
793 Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and their
794 respective control samples (student's *t*-test). Data are mean ± SD (n = 4). Each replicate plate
795 contained 3-6 plants. Open circles indicate the biological replicates.
796
797 **References**
798 **Achard P, Cheng H, De Grauwe L, Decat J, Schoutteten H, Moritz T, Van Der Straeten D, Peng**
799 **J, Harberd NP** (2006) Integration of plant responses to environmentally activated
800 phytohormonal signals. Science (80-) **311**: 91–94
801 **Adams P, Nelson DE, Yamada S, Wendy C, Jensen RG, Bohnert HJ, Griffiths H** (1998) Tansley
802 Review No. 97 Growth and development of *Mesembryanthemum crystallinum* (Aizoaceae).
803 Genet Breed 171–190
804 **Ali A, Khan IU, Jan M, Khan HA, Hussain S, Nisar M, Chung WS, Yun D-J** (2018) The High-
805 Affinity Potassium Transporter EpHKT1;2 From the Extremophile *Eutrema parvula*
806 Mediates Salt Tolerance. Front Plant Sci. doi: 10.3389/fpls.2018.01108
807 **Ali A, Yun DJ** (2017) Salt stress tolerance; what do we learn from halophytes? J Plant Biol **60**:
808 431–439

809 **Apse MP, Aharon GS, Snedden WA, Blumwald E** (1999) Salt Tolerance Conferred by
810 Overexpression of a Vacuolar Na^+/H^+ Antiport in *Arabidopsis*. Science (80-) **285**: 1256–
811 1258
812 **Barkla BJ, Rhodes T, Tran KNT, Wijesinghege C, Larkin JC, Dassanayake M** (2018) Making
813 epidermal bladder cells bigger: Developmental-and salinity-induced endopolyploidy in a
814 model halophyte. Plant Physiol **177**: 615–632
815 **Barros NLF, Marques DN, Tadaiesky LBA, de Souza CRB** (2021) Halophytes and other molecular
816 strategies for the generation of salt-tolerant crops. Plant Physiol Biochem. doi:
817 10.1016/j.plaphy.2021.03.028
818 **Bechtold U** (2018) Plant Life in Extreme Environments : How Do You Improve Drought
819 Tolerance ? **9**: 1–8
820 **Bernard A, Joubès J** (2013) *Arabidopsis* cuticular waxes: Advances in synthesis, export and
821 regulation. Prog Lipid Res. doi: 10.1016/j.plipres.2012.10.002
822 **Blits KC, Gallagher JL** (1991) Morphological and physiological responses to increased salinity in
823 marsh and dune ecotypes of *Sporobolus virginicus* (L.) Kunth. Oecologia **87**: 330–335
824 **Blum A** (2009) Effective use of water (EUW) and not water-use efficiency (WUE) is the target of
825 crop yield improvement under drought stress. F Crop Res **112**: 119–123
826 **de Boer HJ, Drake PL, Wendt E, Price CA, Schulze ED, Turner NC, Nicolle D, Veneklaas EJ** (2016)
827 Apparent overinvestment in leaf venation relaxes leaf morphological constraints on
828 photosynthesis in arid habitats. Plant Physiol **172**: 2286–2299
829 **Boughalleb F, Denden M, Tiba B Ben** (2009) Anatomical changes induced by increasing NaCl
830 salinity in three fodder shrubs, *Nitraria retusa*, *Atriplex halimus* and *Medicago arborea*.
831 Acta Physiol Plant **31**: 947–960
832 **Brady SM, Orlando DA, Lee JY, Wang JY, Koch J, Dinneny JR, Mace D, Ohler U, Benfey PN**
833 (2007) A high-resolution root spatiotemporal map reveals dominant expression patterns.
834 Science (80-). doi: 10.1126/science.1146265
835 **Caño L, Fuertes-Mendizabal T, García-Baquero G, Herrera M, Begoña González-Moro M** (2016)
836 Plasticity to salinity and transgenerational effects in the nonnative shrub *Baccharis*
837 *halimifolia*: Insights into an estuarine invasion. Am J Bot **103**: 808–820

838 **Cheeseman JM** (2013) The integration of activity in saline environments: Problems and
839 perspectives. Funct Plant Biol. doi: 10.1071/FP12285
840 **Cho LH, Yoon J, An G** (2017) The control of flowering time by environmental factors. Plant J **90**:
841 708–719
842 **Conn SJ, Hocking B, Dayod M, Xu B, Athman A, Henderson S, Aukett L, Conn V, Shearer MK,**
843 **Fuentes S, et al** (2013) Protocol: Optimising hydroponic growth systems for nutritional and
844 physiological analysis of *Arabidopsis thaliana* and other plants. Plant Methods **9**: 1–11
845 **Cruz MV, Mori GM, Signori-Müller C, da Silva CC, Oh DH, Dassanayake M, Zucchi MI, Oliveira**
846 **RS, de Souza AP** (2019) Local adaptation of a dominant coastal tree to freshwater
847 availability and solar radiation suggested by genomic and ecophysiological approaches. Sci
848 Rep **9**: 1–15
849 **Dassanayake M, Larkin JC** (2017) Making Plants Break a Sweat: the Structure, Function, and
850 Evolution of Plant Salt Glands. Front Plant Sci **08**: 406
851 **Dassanayake M, Oh D-H, Haas JSJS, Hernandez A, Hong H, Ali S, Yun DJD-J, Bressan RARA, Zhu**
852 **JKJ-K, Bohnert HJHJ, et al** (2011) The genome of the extremophile crucifer *Thellungiella*
853 *parvula*. Nat Genet **43**: 913–918
854 **Devlin PF, Halliday KJ, Harberd NP, Whitelam GC** (1996) The rosette habit of *Arabidopsis*
855 *thaliana* is dependent upon phytochrome action: novel phytochromes control internode
856 elongation and flowering time. Plant J **10**: 1127–1134
857 **Drake PL, de Boer HJ, Schymanski SJ, Veneklaas EJ** (2019) Two sides to every leaf: water and
858 CO_2 transport in hypostomatous and amphistomatous leaves. New Phytol **222**: 1179–1187
859 **Du F, Guan C, Jiao Y** (2018) Molecular Mechanisms of Leaf Morphogenesis. Mol Plant **11**: 1117–
860 1134
861 **Eshel G, Duppen N, Wang G, Oh D-H, Kazachkova Y, Herzyk P, Amtmann A, Gordon M, Chalifa-**
862 **Caspi V, Arland Oscar M, et al** (2021) Positive Selection and Heat-Response
863 Transcriptomes Reveal Adaptive Features of the Arabidopsis Desert Relative, *Anastatica*
864 *hierochuntica*. bioRxiv
865 **Ferreira T, Rasband W** (2012) ImageJ User Guide User Guide ImageJ. Image J user Guid. doi:
866 10.1038/nmeth.2019

867 **Flowers TJ, Colmer TD** (2008) Salinity tolerance in halophytes. New Phytol 945–963
868 **Galvan-Ampudia CS, Julkowska MM, Darwish E, Gandullo J, Korver RA, Brunoud G, Haring**
869 **MA, Munnik T, Vernoux T, Testerink C** (2013) Halotropism is a response of plant roots to
870 avoid a saline environment. Curr Biol **23**: 2044–2050
871 **Ge W, Zhang Y, Sun Z, Li J, Liu G, Ma Y, Gao J** (2017) Physiological and anatomical responses of
872 *Phyllostachys vivax* and *Arundinaria fortunei* (Gramineae) under salt stress. Rev Bras Bot
873 **40**: 79–91
874 **German DA, Al-Shehbaz IA** (2010) Nomenclatural novelties in miscellaneous Asian Brassicaceae
875 (Cruciferae). Nord J Bot **28**: 646–651
876 **Hajiboland R, Bahrami-Rad S, Akhani H, Poschenrieder C, Bahrami S, Hossein R, Charlotte A**
877 (2018) Salt tolerance mechanisms in three Irano-Turanian Brassicaceae halophytes
878 relatives of *Arabidopsis thaliana*. J Plant Res **0**: 0
879 **Helvaci C, Mordogan H, Çolak M, Gündogan I** (2004) Presence and Distribution of Lithium in
880 Borate Deposits and Some Recent Lake Waters of West-Central Turkey. Int Geol Rev **46**:
881 177–190
882 **IPCC** (2021) Climate Change 2021: The Physical Science Basis. Contribution of Working Group I
883 to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change.
884 Cambridge University Press
885 **IPNI** (2021) International Plant Names Index. Published on the Internet http://www.ipni.org,
886 The Royal Botanic Gardens, Kew, Harvard University Herbaria & Libraries and Australian
887 National Botanic Gardens. https://www.ipni.org/
888 **Jarvis DE, Ryu CH, Beilstein MA, Schumaker KS** (2014) Distinct roles for SOS1 in the convergent
889 evolution of salt tolerance in *Eutrema salsugineum* and *Schrenkiella parvula*. Mol Biol Evol
890 **31**: 2094–2107
891 **Jennings DH** (1968) Halophytes, Succulence and Sodium in Plants—a Unified Theory. New
892 Phytol **67**: 899–911
893 **Junghans U, Polle A, Düchting P, Weiler E, Kuhlman B, Gruber F, Teichmann T** (2006)
894 Adaptation to high salinity in poplar involves changes in xylem anatomy and auxin
895 physiology. Plant, Cell Environ **29**: 1519–1531

896 **Kazachkova Y, Eshel G, Pantha P, Cheeseman JM, Dassanayake M** (2018) Halophytism : What
897 Have We Learnt From *Arabidopsis thaliana* Relative Model Systems ? **178**: 972–988
898 **Kazachkova Y, Khan A, Acuña T, López-Díaz I, Carrera E, Khozin-Goldberg I, Fait A, Barak S**
899 (2016) Salt Induces Features of a Dormancy-Like State in Seeds of *Eutrema* (*Thellungiella*)
900 *salsugineum*, a Halophytic Relative of *Arabidopsis*. Front Plant Sci **7**: 1–18
901 **King DA** (1997) The functional significance of leaf angle in *Eucalyptus*. Aust J Bot **45**: 619–639
902 **Kotula L, Garcia Caparros P, Zörb C, Colmer TD, Flowers TJ** (2020) Improving crop salt tolerance
903 using transgenic approaches: An update and physiological analysis. Plant Cell Environ **43**:
904 2932–2956
905 **Krämer U** (2018) Conceptualizing plant systems evolution. Curr Opin Plant Biol **42**: 66–75
906 **Li AJ, Li G, Wang H, Deng XW** (2011) Phytochrome Signaling Mechanisms. Arab B. doi:
907 10.1199/tab.0148
908 **Li H, Testerink C, Zhang Y** (2021) How roots and shoots communicate through stressful times.
909 Trends Plant Sci. doi: 10.1016/j.tplants.2021.03.005
910 **Li K, Wang Y, Han C, Zhang W, Jia H, Li X** (2007) GA signaling and CO/FT regulatory module
911 mediate salt-induced late flowering in *Arabidopsis thaliana*. Plant Growth Regul **53**: 195–
912 206
913 **Lloyd EA, Oreskes N** (2018) Climate Change Attribution: When Is It Appropriate to Accept New
914 Methods? Earth's Futur **6**: 311–325
915 **Lutts S, Kinet JM, Bouharmont J** (1995) Changes in plant response to NaCl during development
916 of rice (*Oryza sativa* L.) varieties differing in salinity resistance. J Exp Bot **46**: 1843–1852
917 **Moriuchi KS, Friesen ML, Cordeiro MA, Badri M, Vu WT, Main BJ, Aouani ME, Nuzhdin S V.,**
918 **Strauss SY, Von Wettberg EJB** (2016) Salinity adaptation and the contribution of parental
919 environmental effects in *Medicago truncatula*. PLoS One **11**: 1–19
920 **Müller P, Li XP, Niyogi KK** (2001) Non-photochemical quenching. A response to excess light
921 energy. Plant Physiol. doi: 10.1104/pp.125.4.1558
922 **Nadeau JA, Sack FD** (2002) Stomatal Development in *Arabidopsis*. Arab B **1**: e0066
923 **Nassar RMA, Kamel HA, Ghoniem AE, Alarcón JJ, Sekara A, Ulrichs C, Abdelhamid MT** (2020)
924 Physiological and anatomical mechanisms in wheat to cope with salt stress induced by

925 seawater. Plants **9**: 1–15
926 **Ogburn RM, Edwards EJ** (2013) Repeated origin of three-dimensional leaf venation releases
927 constraints on the evolution of succulence in plants. Curr Biol **23**: 722–726
928 **Oh D-H, Dassanayake M, Bohnert HJ, Cheeseman JM** (2012) Life at the extreme: lessons from
929 the genome. Genome Biol **13**: 241
930 **Oh D-HHD-H, Hong H, Lee SY, Yun D-JD-J, Bohnert HJ, Dassanayake M** (2014) Genome
931 Structures and Transcriptomes Signify Niche Adaptation for the Multiple-Ion-Tolerant
932 Extremophyte *Schrenkiella parvula*. Plant Physiol **164**: 2123–2138
933 **Oh DH, Dassanayake M** (2019) Landscape of gene transposition-duplication within the
934 Brassicaceae family. DNA Res. doi: 10.1093/dnares/dsy035
935 **Olson ME, Rosell JA** (2013) Vessel diameter-stem diameter scaling across woody angiosperms
936 and the ecological causes of xylem vessel diameter variation. New Phytol **197**: 1204–1213
937 **Orsini F, D'Urzo MP, Inan G, Serra S, Oh D-HH, Mickelbart M V., Consiglio F, Li X, Jeong JC, Yun**
938 **D-JJ, et al** (2010) A comparative study of salt tolerance parameters in 11 wild relatives of
939 *Arabidopsis thaliana*. J Exp Bot **61**: 3787–3798
940 **Ozfidan-Konakci C, Uzilday B, Ozgur R, Yildiztugay E, Sekmen AH, Turkan I** (2015) Halophytes
941 as a source of salt tolerance genes and mechanisms : a case study for the Salt Lake area ,
942 Turkey.
943 **Pantha P, Oh D-H, Longstreth D, Dassanayake M** (2021) Living with high potassium: an asset or
944 a hindrance. bioRxiv 2021.07.01.450778
945 **Pardo J, VanBuren R** (2021) Evolutionary innovations driving abiotic stress tolerance in C4
946 grasses and cereals. Plant Cell
947 **Parida AK, Veerabathini SK, Kumari A, Agarwal PK** (2016) Physiological, anatomical and
948 metabolic implications of salt tolerance in the halophyte *Salvadora persica* under
949 hydroponic culture condition. Front Plant Sci **7**: 1–18
950 **Pierik R, Testerink C** (2014) The art of being flexible: How to escape from shade, Salt, And
951 drought. Plant Physiol **166**: 5–22
952 **De Rocher EJ, Harkins KR, Galbraith DW, Bohnert HJ** (1990) Developmentally regulated
953 systemic endopolyploidy in succulents with small genomes. Science (80-) **250**: 99–101

954 **Rodell M, Famiglietti JS, Wiese DN, Reager JT, Beaudoing HK** (2018) Emerging trends in global
955 freshwater availability. doi: 10.1038/s41586-018-0123-1
956 **Roy SJ, Negrão S, Tester M** (2014) Salt resistant crop plants. Curr Opin Biotechnol **26**: 115–124
957 **Ryu JY, Lee HJ, Seo PJ, Jung JH, Ahn JH, Park CM** (2014) The *Arabidopsis* floral repressor BFT
958 delays flowering by competing with FT for FD binding under high salinity. Mol Plant **7**: 377–
959 387
960 **Santiago-Rosario L, Kyle H, Elderd B, Hart P, Dassanayake M** (2021) No Escape: The Influence
961 of Substrate Sodium on Plant Growth and Tissue Sodium Responses. Ecol Evol. doi:
962 10.22541/au.162081817.72637329/v1
963 **Sarker U, Oba S** (2020) The Response of Salinity Stress-Induced A. tricolor to Growth, Anatomy,
964 Physiology, Non-Enzymatic and Enzymatic Antioxidants. Front Plant Sci **11**: 1–14
965 **Schlenker W, Auffhammer M** (2018) The cost of a warming climate.
966 **Solis CA, Yong MT, Vinarao R, Jena K, Holford P, Hill C, Chen Z** (2020) Back to the Wild : On a
967 Quest for Donors Toward Salinity Tolerant Rice. **11**: 1–14
968 **Somers DE, Sharrock RA, Tepperman JM, Quail PH** (1991) The hy3 long hypocotyl mutant of
969 *Arabidopsis* is deficient in phytochrome B. Plant Cell **3**: 1263–1274
970 **Stanton ML, Roy BA, Thiede DA** (2000) Evolution in Stressful Environments . I . Phenotypic
971 Variability , Phenotypic Selection , and Response to Selection in Five Distinct
972 Environmental Stresses Author (s): M . L . Stanton , B . A . Roy and D . A . Thiede Published
973 by : Society for the Study. Evolution (N Y) **54**: 93–111
974 **Sun K, Cui Y, Hauser BA** (2005) Environmental stress alters genes expression and induces ovule
975 abortion: Reactive oxygen species appear as ovules commit to abort. Planta **222**: 632–642
976 **Teusink RS, Rahman M, Bressan RA, Jenks MA** (2002) Cuticular Waxes on *Arabidopsis thaliana*
977 Close Relatives *Thellungiella halophila* and *Thellungiella parvula*. Int J Plant Sci **163**: 309–
978 315
979 **Thorvaldsdóttir H, Robinson JT, Mesirov JP** (2013) Integrative Genomics Viewer (IGV): High-
980 performance genomics data visualization and exploration. Brief Bioinform. doi:
981 10.1093/bib/bbs017
982 **Tug GN, Yaprak AE, Vural M** (2019) The Floristical, Ecological, and Syntaxonomical

983 Characteristics of Salt Marshes and Salt Steppes in Turkey. Tasks Veg. Sci. pp 413–446
984 **Ungar IA** (1978) Halophyte seed germination. Bot Rev **44**: 233–264
985 **Ventura Y, Myrzabayeva M, Alikulov Z, Omarov R, Khozin-Goldberg I, Sagi M** (2014) Effects of
986 salinity on flowering, morphology, biomass accumulation and leaf metabolites in an edible
987 halophyte. AoB Plants. doi: 10.1093/aobpla/plu053
988 **Wang G, Ditusa SF, Oh D, Herrmann AD, Mendoza-cozatl DG, Neill MAO, Smith AP,**
989 **Dassanayake M** (2021) Cross species multi-omics reveals cell wall sequestration and
990 elevated global transcript abundance as mechanisms of boron tolerance in plants. 1985–
991 2000
992 **Wang G, Pantha P, Tran KN, Oh DH, Dassanayake M** (2019) Plant growth and agrobacterium-
993 mediated floral-dip transformation of the extremophyte *Schrenkiella parvula*. J Vis Exp
994 **2019**: 1–8
995 **Wang XJ, Hu QJ, Guo XY, Wang K, Ru DF, German DA, Weretilnyk EA, Abbott RJ, Lascoux M,**
996 **Liu J quan** (2018) Demographic expansion and genetic load of the halophyte model plant
997 *Eutrema salsugineum*. Mol Ecol **27**: 2943–2955
998 **Yamaguchi T, Blumwald E** (2005) Developing salt-tolerant crop plants: Challenges and
999 opportunities. Trends Plant Sci **10**: 615–620
1000 **Yoo CY, Pence HE, Hasegawa PM, Mickelbart M V.** (2009) Regulation of transpiration to
1001 improve crop water use. CRC Crit Rev Plant Sci **28**: 410–431
1002 **Yu Y, Wang L, Chen J, Liu Z, Park CM, Xiang F** (2018) WRKY71 Acts Antagonistically Against Salt-
1003 Delayed Flowering in *Arabidopsis thaliana*. Plant Cell Physiol **59**: 414–422
1004 **Zandalinas SI, Fritschi FB, Mittler R** (2021) Global Warming , Climate Change , and
1005 Environmental Pollution : Recipe for a Multifactorial Stress Combination Disaster Trends in
1006 Plant Science. Trends Plant Sci **26**: 588–599
1007 **Zhu JK** (2015) The Next Top Models. Cell **163**: 18–20

Figure and table captions

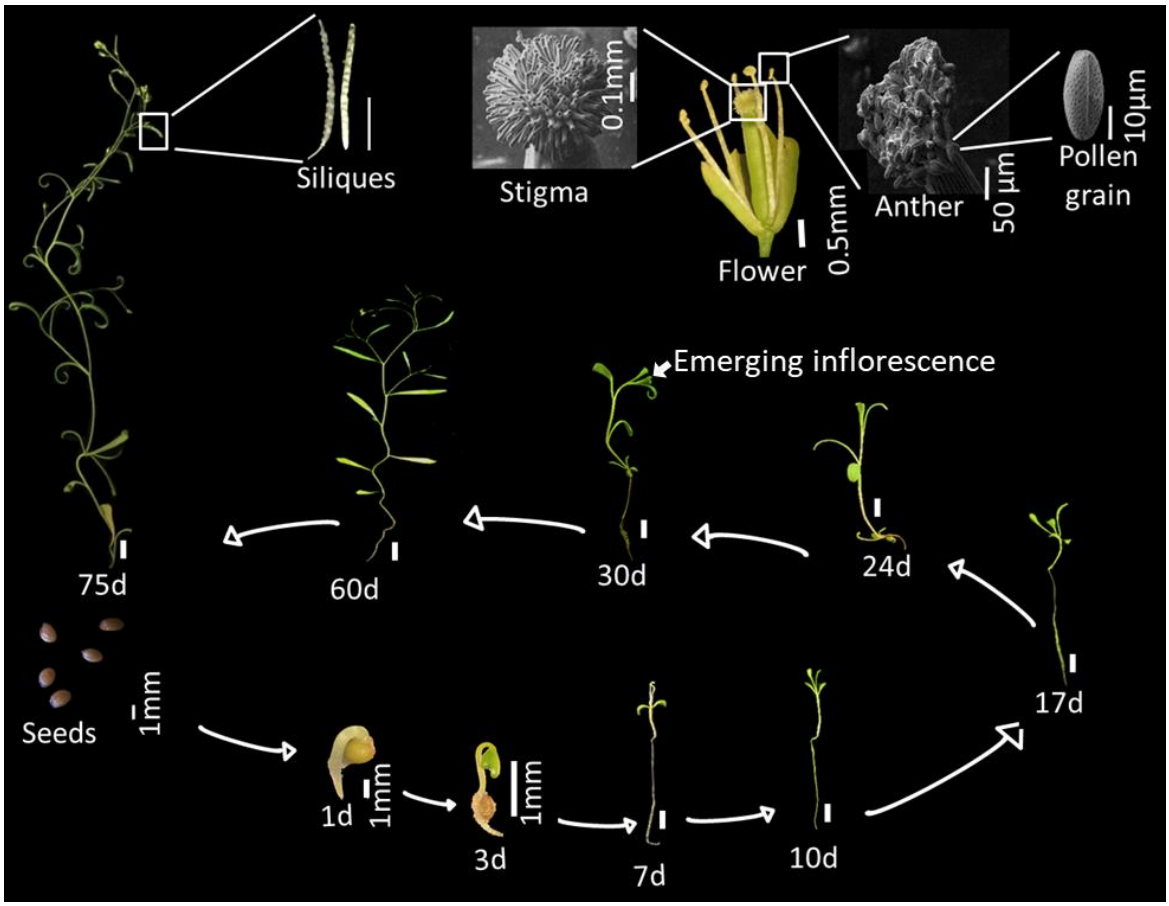

Figure 1. Life cycle of *Schrenkiella parvula* from seeds to siliques. Scale bars are 1 cm unless indicated in the figure.

Table 1. Morphological comparison between *Arabidopsis thaliana* and *Schrenkiella parvula*.

Morphology	***A. thaliana***	***S. parvula***
Growth habit	Determinate	Indeterminate
Leaf arrangement	Rosette	Spiral phyllotaxy
Leaf shape	Ovate	Linear-lanceolate
Trichomes	Present	Absent
Petals	4	Absent
Number of stamens	6 (2 short and 4 long)	6 (equal size)
Siliques/plant[a]	74.9±18.69	61.6±14.40
Silique length (cm)	1.16±0.09	1.48±0.06
Seeds/silique	23.91±13.60	21.58±5.57
Seed weight[b] (g)	0.020±0.003	0.063±0.013
Seed area (mm^2)	0.12±0.02	0.31±0.04

[a] Quantification from 10-12-week-old plants grown under 14-hr light/10-hr dark photoperiod, 22 °C – 24 °C temperature with a light intensity of 130 µmol m^{-2} s^{-1}.
[b] Average weight of 500 seeds.
Data are represented as mean of 3 independent replicates ± SD.

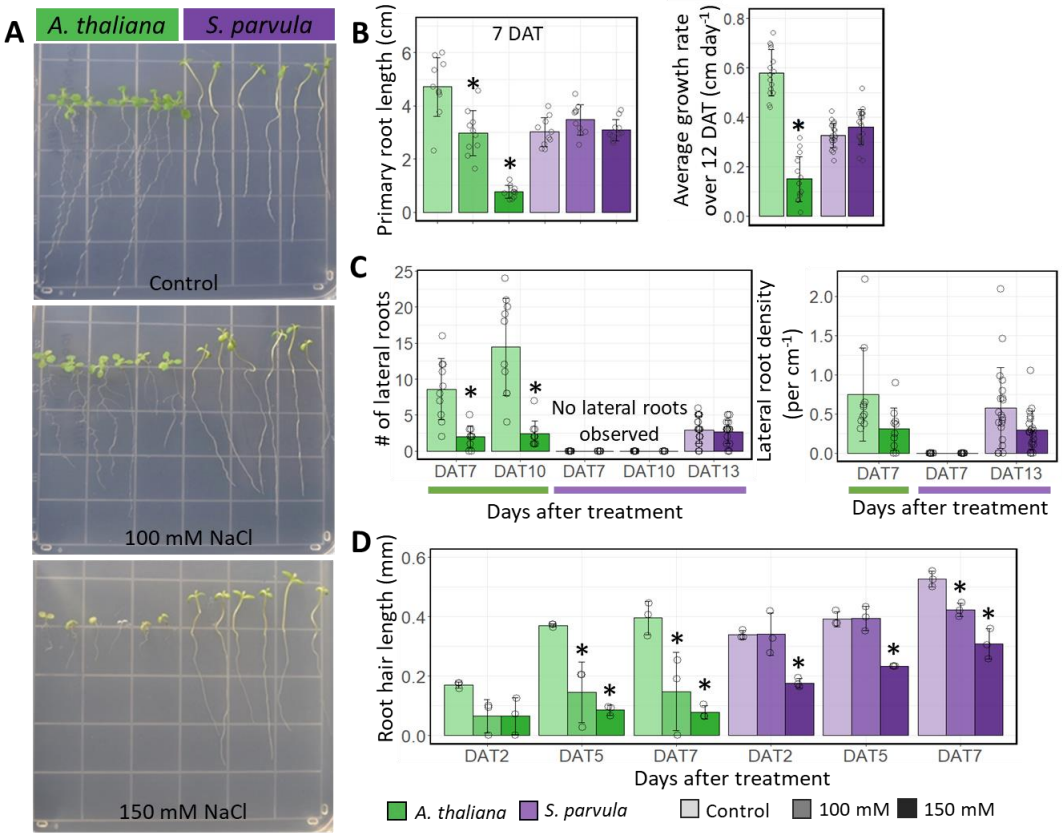

Figure 2. Effects of NaCl stress on root growth in *Schrenkiella parvula* and *Arabidopsis thaliana* seedlings. [A] 12-day-old seedlings of *A. thaliana* and *S. parvula* were grown for 5 days on 1/4x Murashige and Skoog media, and 7 days on the indicated concentrations of NaCl. Plates were scanned 7 days after treatment (DAT). [B] Primary root growth and average root growth rate, [C] number of lateral roots and lateral root density, and [D] average length of 10 longest root hairs. Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and their respective control group, determined by Student's *t*-test. Data are mean ± SD (n =3, at least 3 plants per replicate). Open circles are individual measurements.

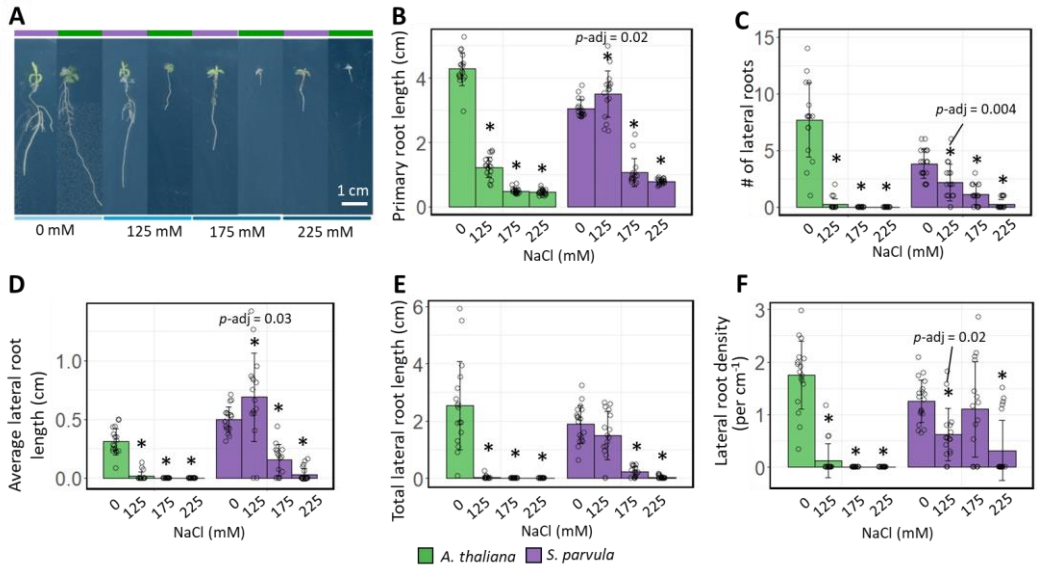

Figure S1. Effects of NaCl stress on root growth of *Schrenkiella parvula* and *Arabidopsis thaliana* under long days and higher nutrient conditions. [A] Root growth of 10-day-old seedlings under indicated concentrations of NaCl. Quantification of [B] primary root growth, [C] number of lateral roots, [D] average lateral root length, [E] total lateral root length, and [F] lateral root density. Asterisks indicate significant difference (p-adj ≤ 0.05) between the treated samples and their respective control samples, determined by two-way ANOVA. Data are represented as mean ± SD (n = 15 to 20). Open circles indicate the number of plants used for each experiment.

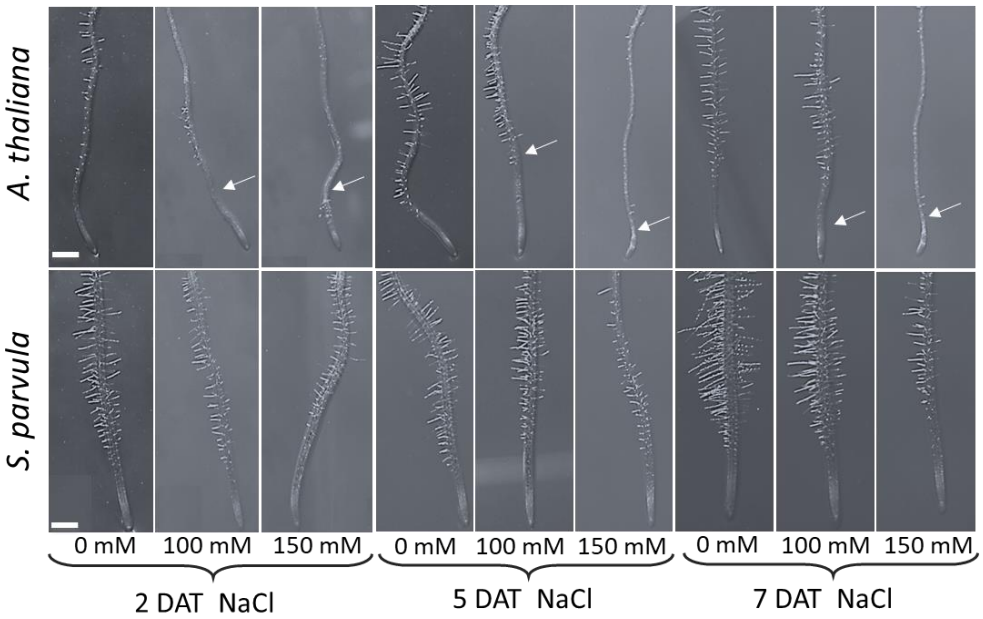

Figure S2. Effects of NaCl stress on root hair growth in *Schrenkiella parvula* and *Arabidopsis thaliana*. White arrows indicate root tip positions when the 5-day-old seedlings were transferred to the indicated salt concentrations. Note that the white arrows were absent in *A. thaliana* and *S. parvula* control because the position of the root tip at the time of transferring were out of frame when we imaged the root tip region. Scale bars are 0.5 mm. DAT, Days after treatment.

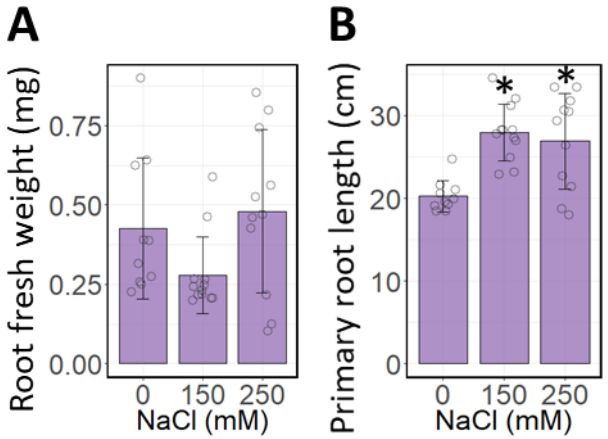

Figure S3. Long term effects of NaCl stress on the root growth of *Schrenkiella parvula*. [A] Root fresh weight and [B] primary root length of 12-week-old hydroponically grown *S. parvula* plants. Four-week-old plants were subjected to 150 mM NaCl for an additional 4 weeks and the treatment was continued or increased to 250 mM NaCl for another 4 weeks. The plants were grown at 100-150 µmol m^{-2} s^{-1} photosynthetic photon flux density with a 12 hr light/12 hr dark photoperiod. Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and their respective control samples, determined by Student's *t*-test. Data are represented as the mean ± SD (n = 9). For each plot, an open circle indicates the measurement from each plant.

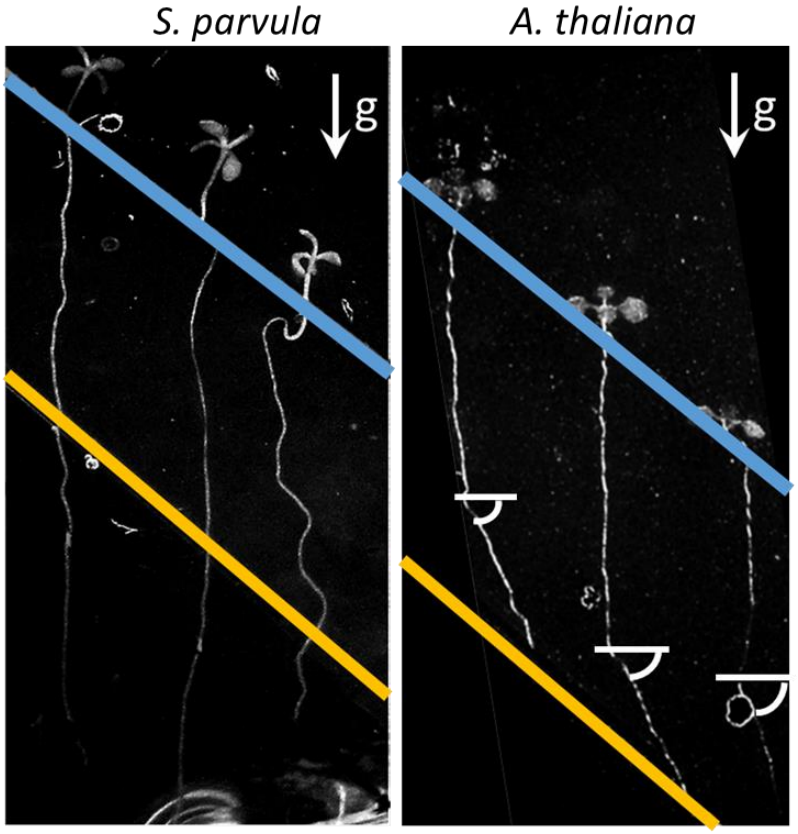

Figure S4. Halotropism assay at 200 mM NaCl for *Schrenkiella parvula* and *Arabidopsis thaliana*. The blue lines indicate the position of seeds sown. The yellow line marks the separation between 1/4x MS media with 0 mM (top) and 200 mM NaCl (bottom). The bent root angle is indicated by curved white lines. g, gravity axis.

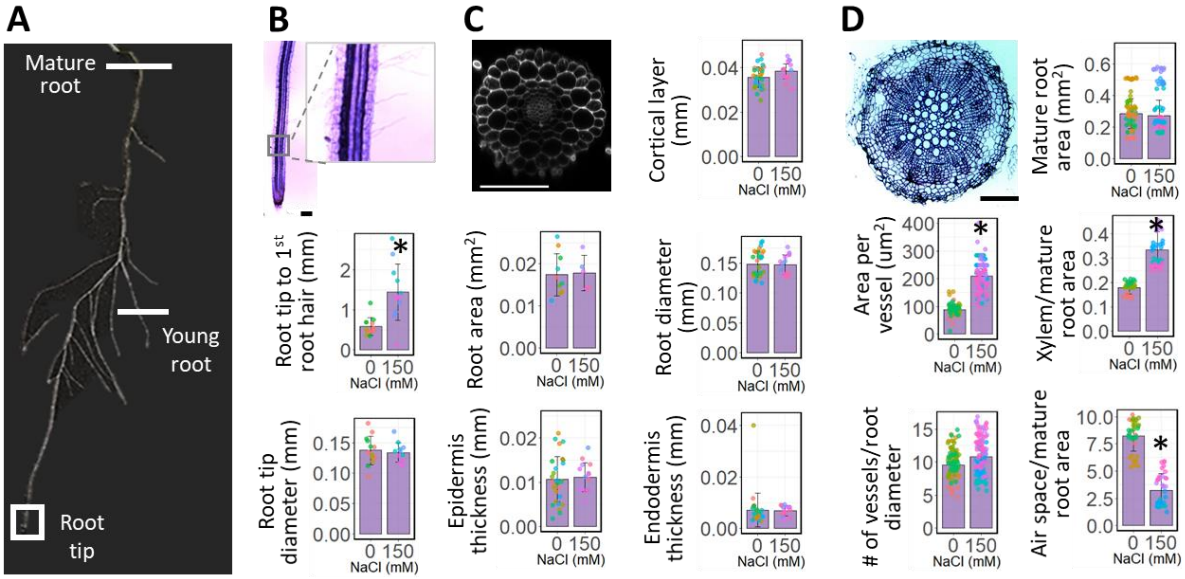

Figure 3. Effects of NaCl stress on *Schrenkiella parvula* root anatomy. Eight-week-old *S. parvula* plants were grown under control or 150 mM NaCl conditions for an additional 4 weeks. [A] The position of transverse sections of *S. parvula* roots, [B] the root tip, [C] the young root, and [D] the mature root from 12-week-old hydroponically-grown *S. parvula* plants. A minimum of 20 sections from 4 to 13 plants were examined for each root region to quantify each parameter for each condition. Asterisks indicate significant differences ($p \leq 0.05$) between the treated samples and their respective control group, determined by Student's *t*-test. Data are means ± SD. Data points represent individual cross-sections and colors represent individual plants. Representative cross-sections were obtained from the control plants. Scale bar represents 100 µm.

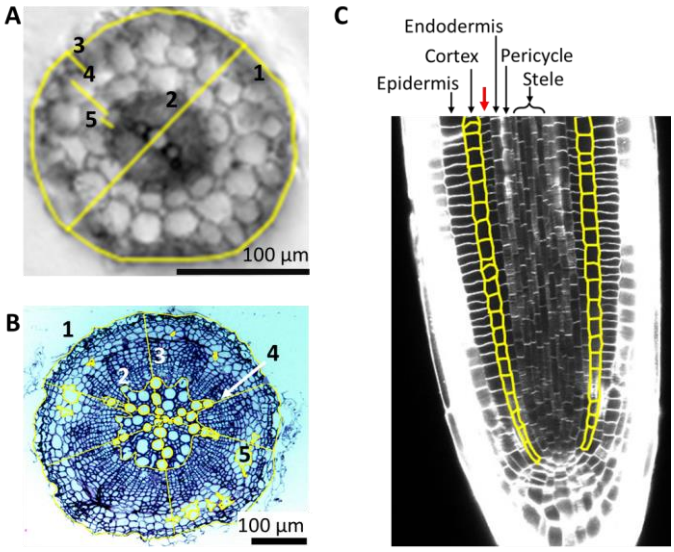

Figure S5. Root anatomy of *Schrenkiella parvula*. [A] Transverse section of the young root taken at 1-3 cm from the root tip. [1] Young root area; [2] root diameter; [3] epidermis thickness; [4] cortical layer thickness; [5] endodermis thickness. [B] Transverse section of the mature root taken between 2-5 cm below the root-shoot junction. [1] Mature root area; [2] xylem/root area; [3] Area per vessel; [4] # vessels/root diameter; [5] Air space/root area. [C] Longitudinal section of young root tip from 7-day-old seedlings. The cortex layer is highlighted in yellow. The red arrow indicates the additional cell layer.

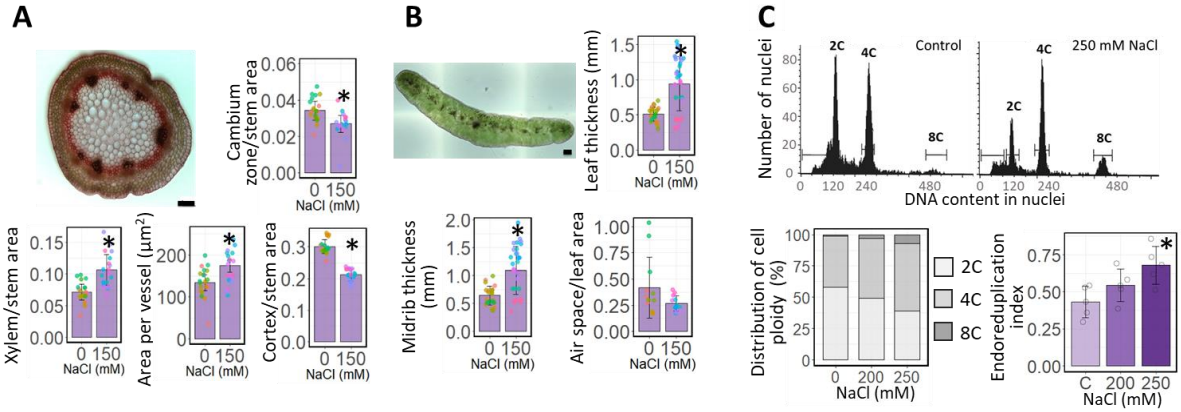

Figure 4. Effects of NaCl stress on *Schrenkiella parvula* shoot anatomy. Eight-week-old *S. parvula* plants were grown under control or 150 mM NaCl conditions for an additional 4 weeks. Transverse section of [A] stem and [B] leaf of 12-week-old hydroponically-grown *S. parvula* plants. A total of 23 sections from 7 control plants and 32 sections from 9 treated plants were used for leaf measurements, and a minimum of 20 sections from 5 to 11 plants were used for stem measurements. Data points represent individual cross-sections and colors represent individual plants. [C] Nuclear DNA content, distribution of leaf cell ploidy, and endoreduplication index of leaf cells from 10th and 11th leaves from the shoot meristem in control and 250 mM NaCl-treated 8-week-old *S. parvula* plants. Data are mean ± SD (n= 4). Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and their respective control group, determined by Student's *t*-test. Representative cross-sections were obtained from the control plants Scale bars represent 100 µm.

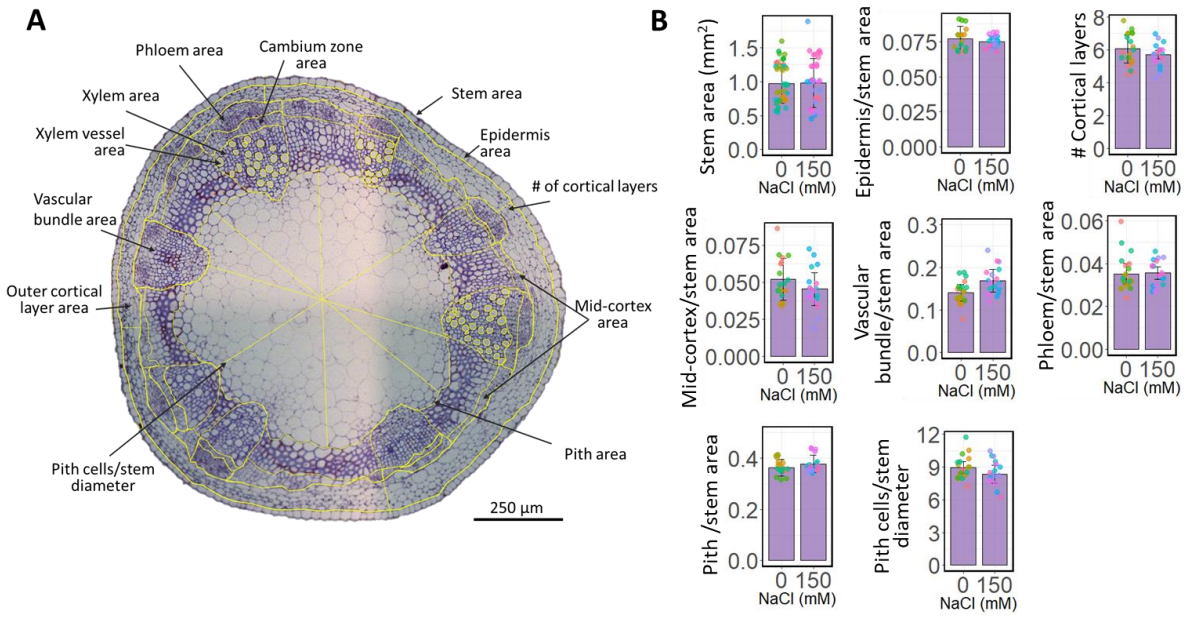

Figure S6. Stem anatomy of *Schrenkiella parvula*. [A] Transverse section of *S. parvula* stem (between 4th, 5th, and 6th internodes from the shoot meristem) with cell layers marked in yellow lines for trait quantification. [B] Measurements of stem anatomical features indicated in [A]. A minimum of 20 sections from 5-11 plants were used for each measurement. Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and its respective control samples, determined by Student's *t*-test. Data points represent individual cross-sections and colors represent individual plants.

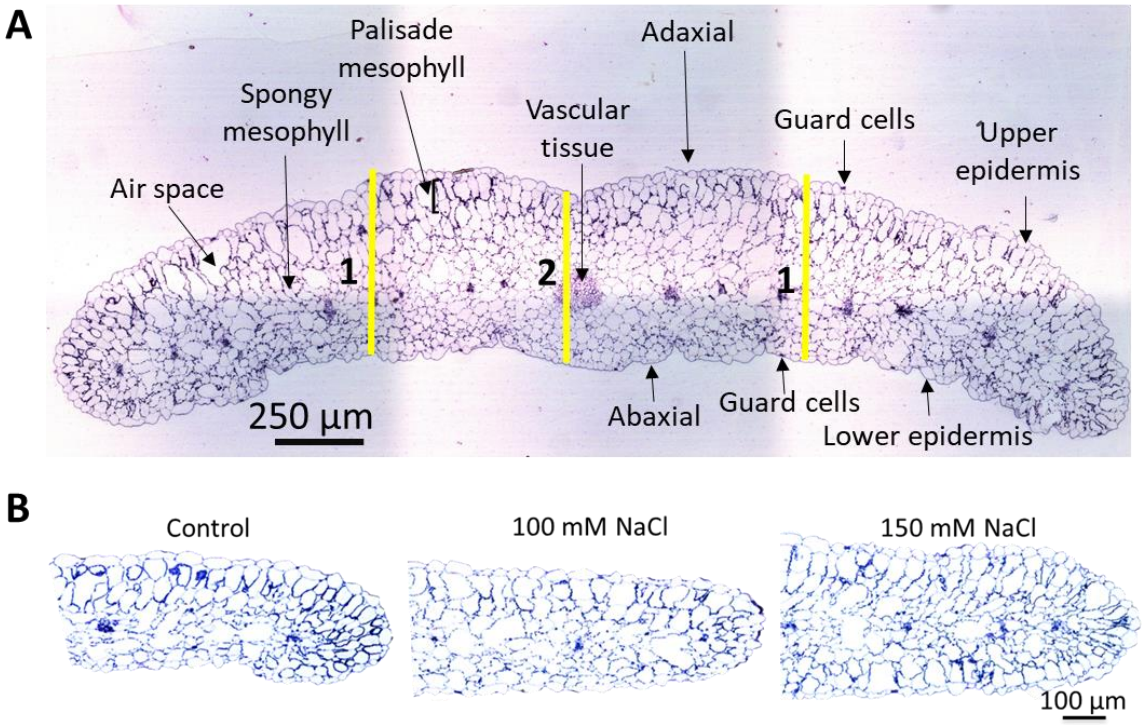

Figure S7. Leaf structures of *Schrenkiella parvula*. [A] Transverse section of *S. parvula* leaves. [1] leaf thickness and [2] midrib thickness. [B] Transverse sections of *S. parvula* leaves treated with indicated concentration of salts. All sections represent the fifth or sixth leaf from the root-shoot junction in 8-week-old plants that were treated for 4 weeks.

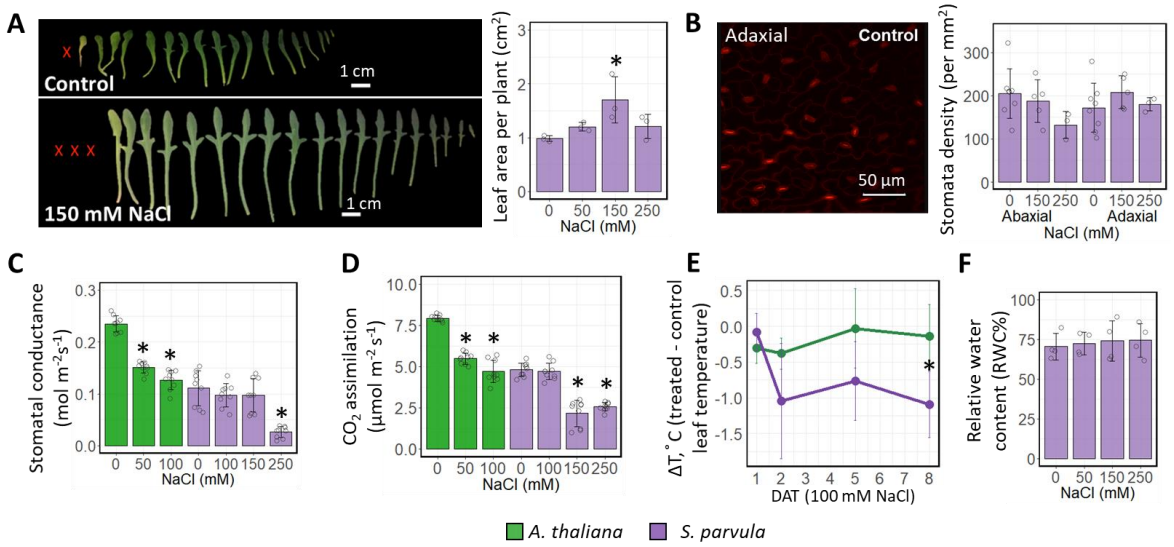

Figure 5. Effects of NaCl stress on *Schrenkiella parvula* leaf traits. All experiments were performed with 4-week-old hydroponically-grown plants that were treated for an additional 4 weeks with the indicated NaCl concentrations. [A] Total leaf area, [B] Stomatal density, [C] Stomatal conductance, [D] Photosynthesis rate, [E] Leaf relative surface temperature, and [F] Leaf relative water content. Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and their respective control samples, determined by Student's *t*-test. Data are means ± SD (n ≥ 3). Open circles indicate the measurement from each plant (in B and F) and leaves (in C and D) for each experiment. DAT, days after treatment.

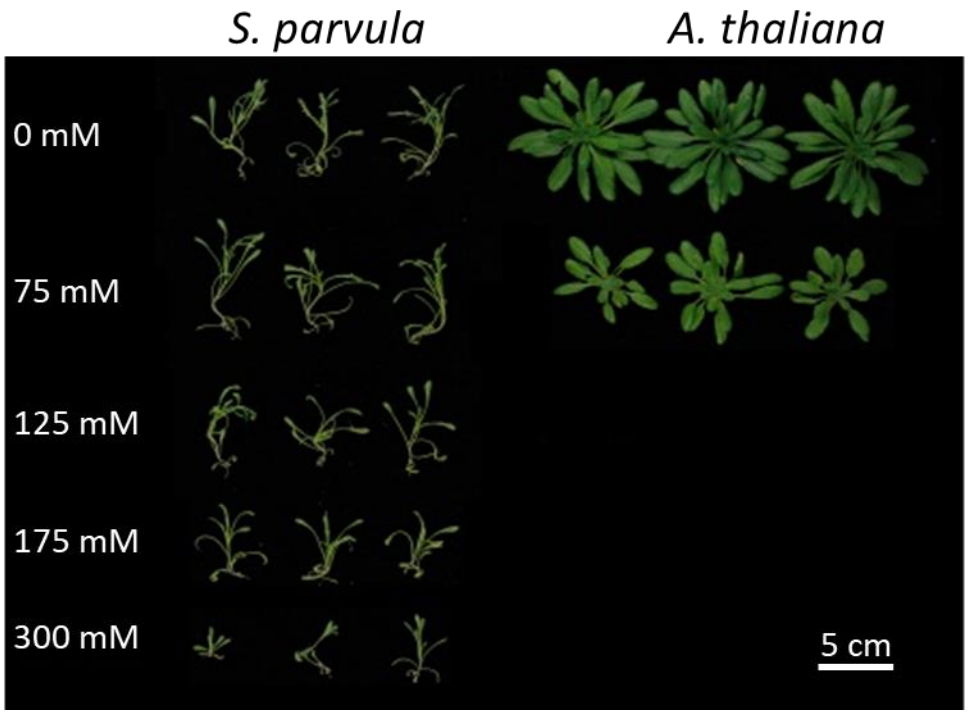

Figure S8. Effects of salt on growth of 4-week-old *Arabidopsis thaliana* and *Schrenkiella parvula* treated for an additional 2 weeks at 22 °C under a 12 hr light/12 hr dark photoperiod in soil.

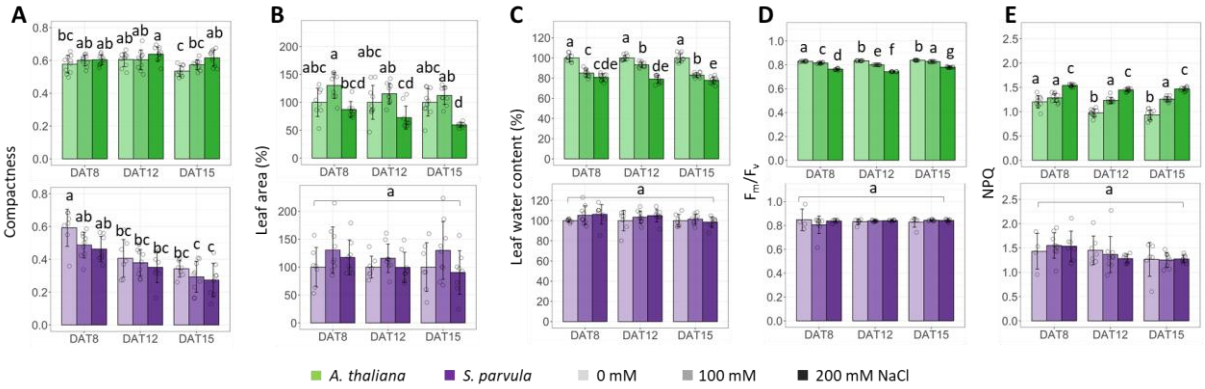

Figure S9. Effects of NaCl stress on *Arabidopsis thaliana* and *Schrenkiella parvula* morphometric and physiological traits. Soil-grown, 8-day-old *A. thaliana* and 12-day-old *S. parvula* were exposed to the indicated salt concentrations at 21 °C, 16 h light/8 h dark photoperiod. The Measurements were taken on day 8, 12, and 15 after the start of the salt treatments. [A] Compactness (the ratio between the rosette (or leaf) area and the rosette (or leaf) convex hull area); [B] Leaf area; [C] Leaf water content; [D] Maximum quantum efficiency of PSII; and [E] Non-photochemical quenching were measured/estimated using the Qubit/PSI PlantScreen™ Compact System. For leaf area and leaf water content, values are percent of control (0 mM NaCl) for each time point. Letters denote significant changes at *p*-adj ≤ 0.05 (one-way ANOVA with post-hoc Tukey's test). Data are mean ± SD (n = 10). Open circles indicate the individual measurements obtained from each plant. DAT, days after treatment.

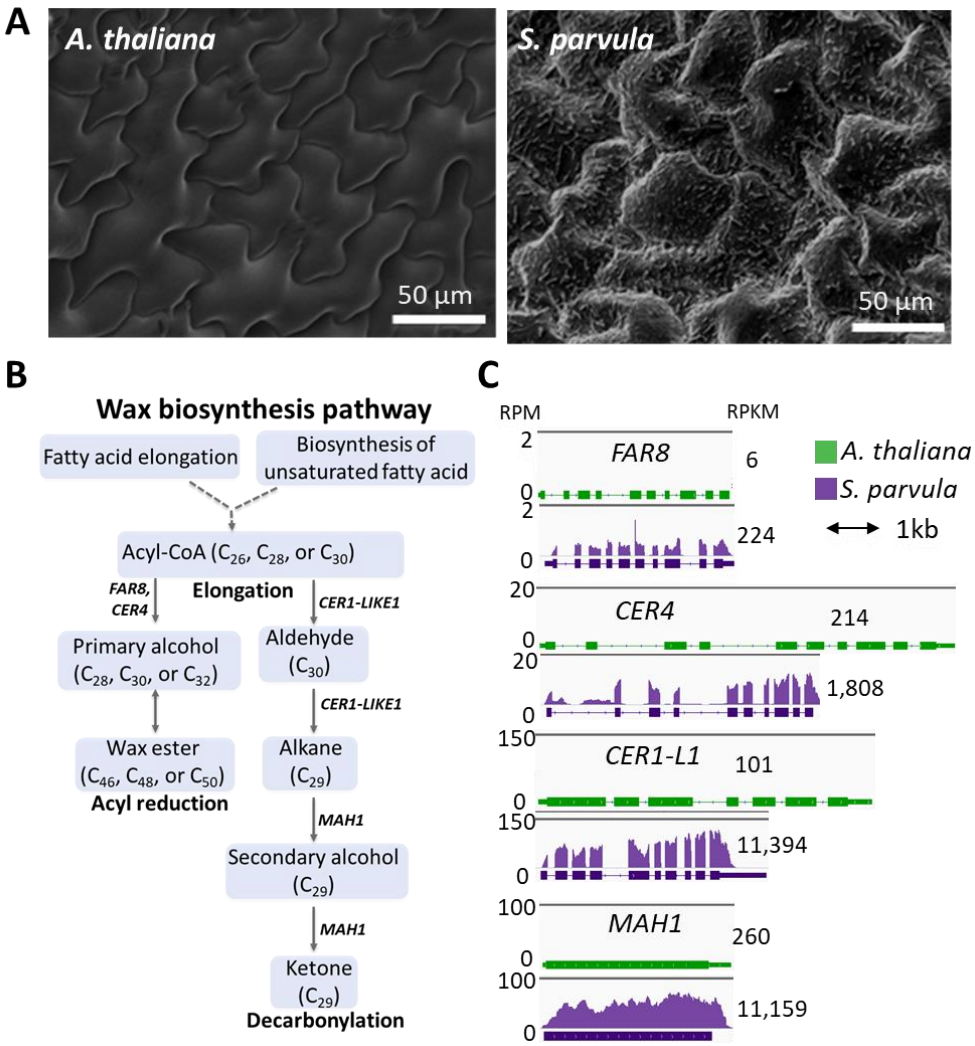

Figure S10. Leaf surface and the basal expression of genes involved in wax biosynthesis in *Arabidopsis thaliana* and *Schrenkiella parvula*. [A] Scanning electron micrographs contrasting *A. thaliana* and *S. parvula* leaf surfaces. [B] Major wax biosynthesis pathway and [C] wax biosynthesis genes that exhibited significantly different basal expression between *A. thaliana* and *S. parvula*.

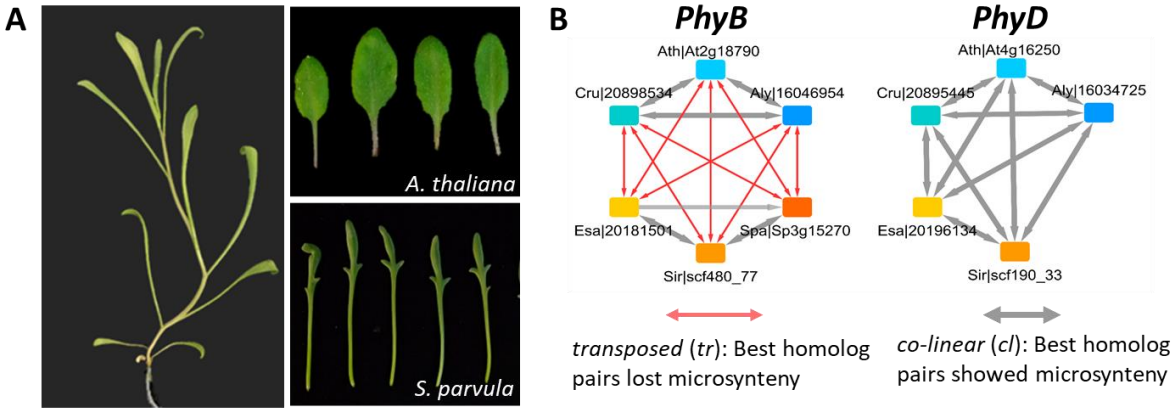

Figure S11. The potential influence of phytochrome family of genes on *Schrenkiella parvula* leaf and growth form. [A] Elongated internode and leaf petiole in *S. parvula* compared to *A. thaliana*. [B] *PHYD* is absent in the *S. parvula* genome as illustrated using OrthNet representing evolutionary histories of orthologous gene groups derived from six Brassicaceae genomes: *Arabidopsis lyrata* (Aly, version 1.0), *Arabidopsis thaliana* (Ath, v. 'TAIR10'), *Capsella rubella* (Cru, v. 1.0), and *Eutrema salsugineum* (Esa, v. 1.0), *Sisymbrium irio* (Sir, v. 0.2), and *Schrenkiella parvula* (v. 2.0). Nodes are color-coded according to the species. Edges show either co-linear (cl) or transposed (tr) properties.

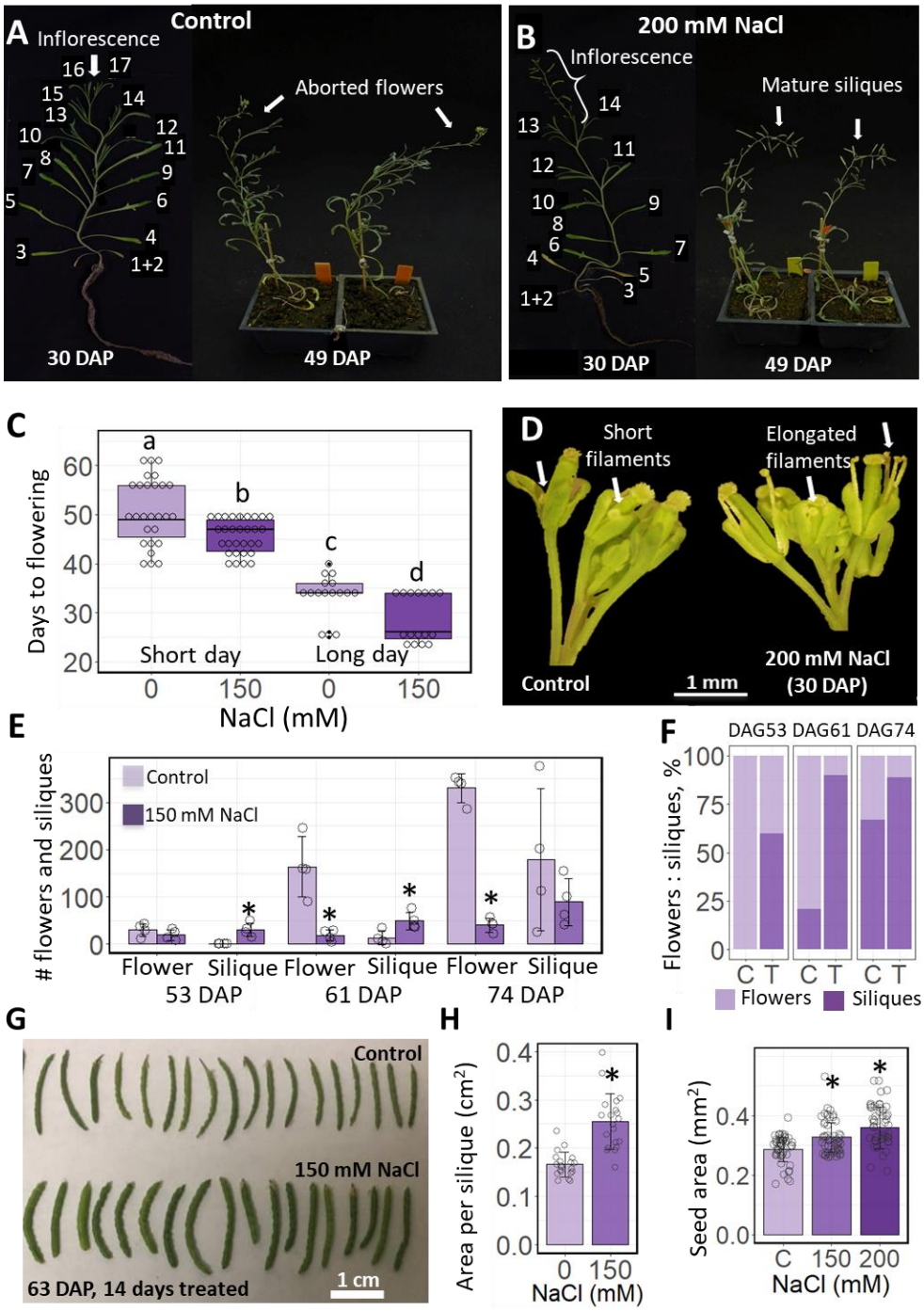
A
Control
Inflorescence
Aborted flowers
30 DAP
49 DAP
B
200 mM NaCl
Inflorescence
Mature siliques
30 DAP
49 DAP
C
Days to flowering
Short day
Long day
NaCl (mM)
D
Short filaments
Elongated filaments
Control
1 mm
200 mM NaCl (30 DAP)
E
Control
150 mM NaCl
flowers and siliques
Flower 53 DAP
Silique 53 DAP
Flower 61 DAP
Silique 61 DAP
Flower 74 DAP
Silique 74 DAP
F
DAG53 DAG61 DAG74
Flowers : siliques, %
Flowers
Siliques
G
Control
150 mM NaCl
63 DAP, 14 days treated
1 cm
H
Area per silique (cm²)
NaCl (mM)
I
Seed area (mm²)
NaCl (mM)

Figure 6.

Figure 6. Effects of NaCl on reproductive traits of *Schrenkiella parvula*. [A] Plants grown under control conditions generally flower at the ~17th leaf stage and the first few flowers are subsequently aborted. [B] Salt treated plants flower earlier at the ~14th leaf stage and the first flowers develop into mature siliques. [C] Days from planting to the first observed flower of plants grown under a long day (16 hr light/8 hr dark) or short day (12 hr light/12 hr dark) photoperiod with and without the indicated salt treatment. Center line - median; box - interquartile range (IQR); notch - 1.58 × IQR/sqrt(n); whiskers - 1.5 × IQR. [D] Flowers obtained from control and treated plants described in panel A and B. [E] Number of flowers and siliques per plant under control and salt treatment. Data are mean ± SD. Open circles represent individual measurements from four biological replicates. [F] Ratio between the number of flowers and siliques observed for control (C) and salt-treated (T) plants described in E. [G] Siliques from plants under control and NaCl treated conditions. [H] Size of siliques from G. Open circles represent individual silique counts. [I] Comparison of the area of seeds from plants under control and NaCl treated conditions. Data are mean ± SD (n = 50). Seeds per condition were selected from the seed pool of 15 to 50 plants. Open circles represent individual seeds. For panel [B], salt treatment started 21 DAP, initially at 50 mM NaCl given every other day. The salt concentration was increased by 50 mM every four days until it reached 200 mM; for panels [C, E, F], salt treatment was applied to 3-week-old plants until the end of the experiment; for panels [G, H, I], salt treatment was applied to 5-week-old plants for an additional 2 weeks. Different letters or asterisks represent significant differences (*, $p < 0.05$) compared to control, determined by either one-way ANOVA followed by Tukey's post-hoc test [C] or Student's *t*-test [E, H, I]. DAP, days after planting.

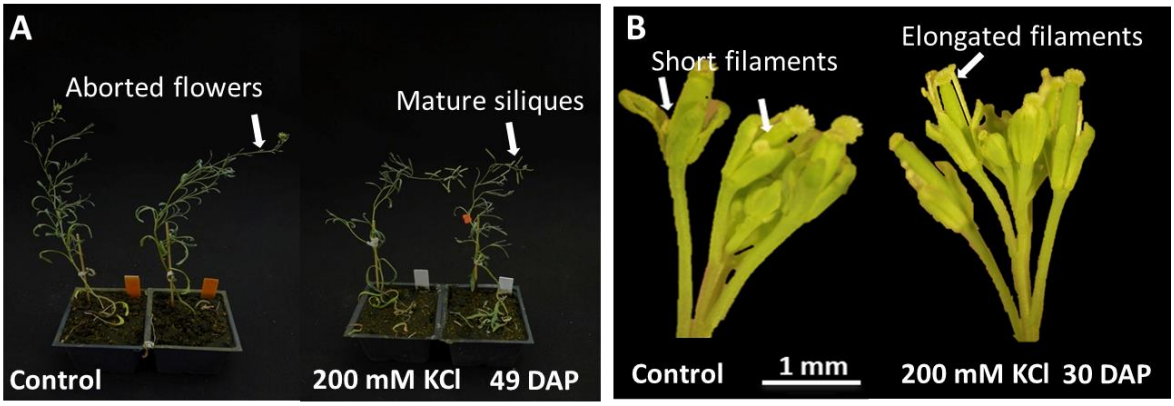

Figure S12. Effects of KCl on reproductive traits of *Schrenkiella parvula*. [A] Plants grown under control conditions generally flower late and the first few flowers are subsequently aborted. KCl-treated plants flower earlier and the first flowers develop into mature siliques. [B] Early flowers in KCl treated plants develop longer filaments compared to control plants. DAP, days after planting.

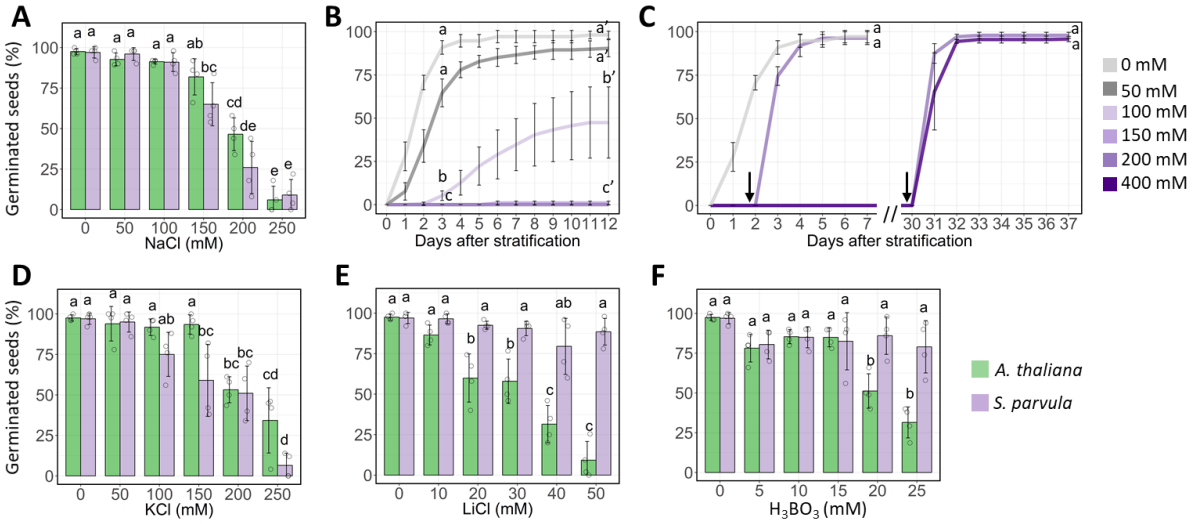

Figure 7. Effects of salt stress on the germination of *Arabidopsis thaliana* and *Schrenkiella parvula* seeds. [A] Final percent germination recorded at 3 days and [B] germination curves of seeds treated with different concentrations of NaCl recorded for 12 days after stratification. [C] Germination curves of NaCl-treated seeds transferred to control (no NaCl) media, after 2 days or 30 days on salt plates. The germination curves were performed by direct sowing of seeds onto salt plates prior to stratification. Black arrows point to the day when seeds were transferred to non-NaCl plates. [D, E, F] Final percent germination of seeds treated with KCl [D], LiCl [E] or H_3BO_3 [F]. Data are mean ± SD (n = 3-4, each replicate contains ca. 50 seeds). Open circles indicate individual measurements. Different letters indicate significant difference ($p \leq 0.05$) determined by one-way ANOVA with post-hoc Tukey's test across all conditions in [A, D, E, F], or across treatment groups at 3 and 12, and 7 days after stratification in [B] and [C] respectively.

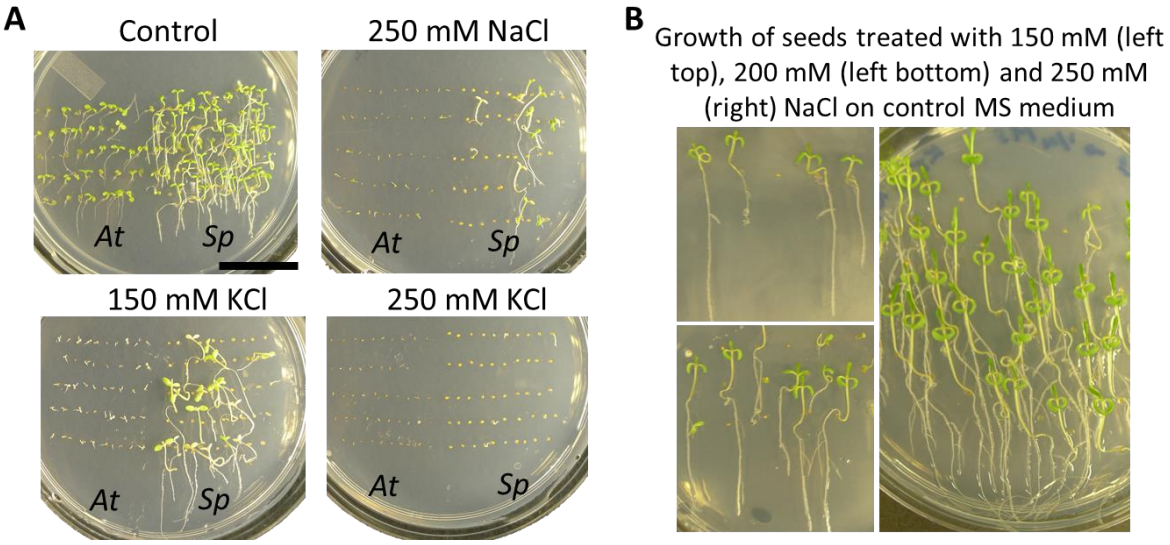

Figure S13. Effect of salt on *Arabidopsis thaliana* and *Schrenkiella parvula* inducing barriers for seed germination and seedling establishment. [A] *S. parvula* (*Sp*) and *A. thaliana* (*At*) seed germination and growth on NaCl- and KCl-supplemented MS medium. Scale bars are 2 cm. [B] Growth of ungerminated *S. parvula* seeds treated with different concentrations of NaCl (as shown in Figure 7) after transferring to control MS medium.

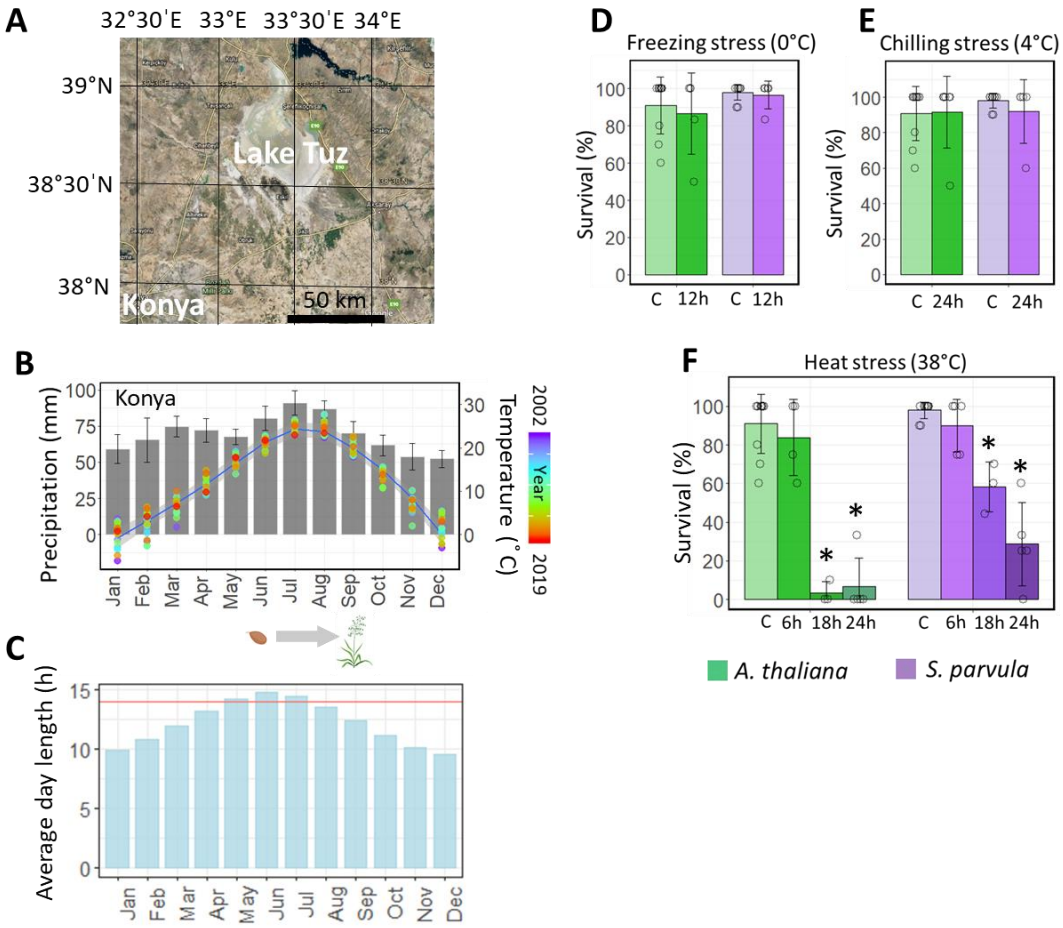

Figure S14. Survival of *Schrenkiella parvula* in the climate conditions found in Lake Tuz region. [A] Lake Tuz location from Google Earth; [B] Precipitation and temperature recorded in Konya, Turkey (NOAA, 2019). *S. parvula* growing season is from April/May to August/September (Tug et al., 2019). Bars indicate precipitation and lines represent temperature. [C] Average day length recorded in Konya per month. Red line indicates the day length used for typical laboratory growth assays for *S. parvula*. [D-F] Survival rate of *A. thaliana* and *S. parvula* seedlings under [D] freezing stress, [E] chilling stress, and [F] heat stress. Five-day-old seedlings were subjected to temperature treatments and data were recorded 5 days after recovery. Asterisks indicate significant difference ($p \leq 0.05$) between the treated samples and their respective control samples (student's *t*-test). Data are mean ± SD (n = 4). Each replicate plate contained 3-6 plants. Open circles indicate the biological replicates.

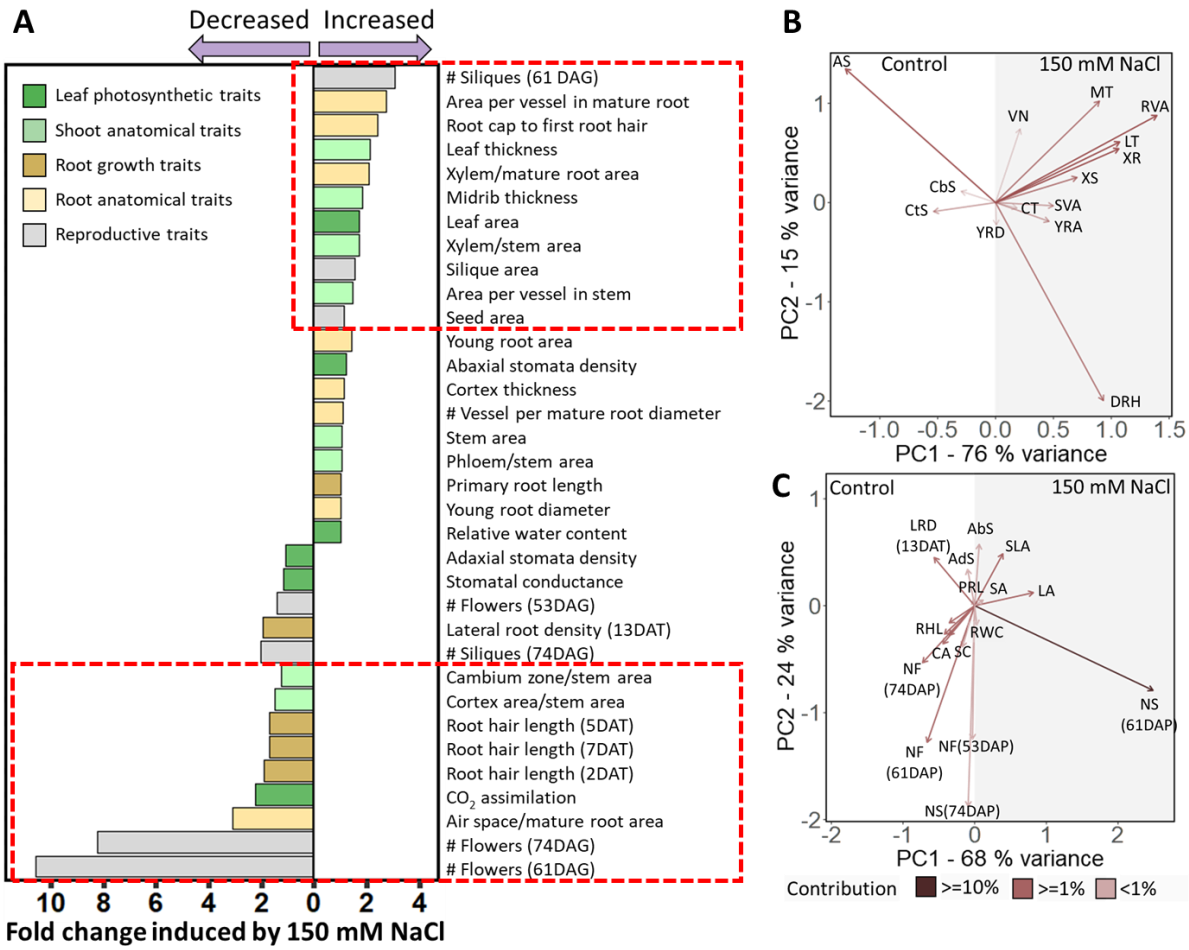

Figure 8. Summary of salt-induced structural and physiological traits in *Schrenkiella parvula*. [**A**] Salt- induced changes in traits quantified in the current study. Changes in each trait were calculated as the fold change between treatment and control measurements. Traits highlighted in the red boxes showed significant differences ($p < 0.05$) under salt treatments compared to the control determined in previous assays (Fig 2-7). [**B**] and [**C**] PCA biplot of traits quantified for *S. parvula* under control and salt-treated conditions. Arrows indicate directions of loadings for each trait and are color-coded by contribution to the variations in PC1 and PC2. [B] Anatomical traits quantified: LT, leaf thickness; MT, midrib thickness; XS, stem xylem to stem area; SVA, average area per vessel in stems; CbS, cambium to stem area; CtS, cortex to stem area; XR, root xylem to root area; RVA, average area per vessel in roots; VN, number of vessels across the root diameter; AS, air space to root area; YRA, young root area; CT, cortical thickness; YRD, root diameter in young roots; DRH, distance from root tip to first root hair. [C] Physiological traits quantified: AbS, abaxial stomatal density; adS, Adaxial stomatal density; LA, leaf area; SLA, silique area; SA, seed area; PRL, primary root length; LRD, lateral root density; RWC, relative water content; CA, CO2 assimilation; SC, stomatal conductance; RHL, root hair length; NF, number of flowers; NS, number of siliques. DAT, days after treatment; DAP, days after planting.